



Data Analysis

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Computing Resources



Learning Objectives

In this session, we will learn:

- what High-Performance Computing (HPC) is
- how to get access to HPC systems provided by Sigma2 for Norwegian research
- how to find other local computing resources available at your university



HPC Systems

Sigma2 owns the HPC Systems Saga, Fram and Betzy and NRIS operates the systems. NRIS is a collaboration of the four BOTT universities and Sigma2 to pool competencies, resources and services.

Apply for resources: <https://www.sigma2.no/high-performance-computing>



Saga

System: HPE Apollo 2000

Number of cores: 16064

Number of nodes: 364 + 8 bigmem + 8 GPU nodes

Max floating point performance, double: 0.65 Petaflop/s

Total memory: 97.5 TiB

Total disk capacity: 5.3 PB (++)

Network: FDR IB (56 Gbit)

File System: BeeGFS, HPE 4150



Software modules

Need for an isolated environment on a shared HPC resource

(Scientific) software should not be installed in the global PATH

Need some way of bringing software in and out of your environment

Many different solutions to this:

- Lmod (modules) + EasyBuild (installation)

- Virtualenv + pip

- Conda

- Singularity containers

We use Lmod + EasyBuild for the scientific software stack on all our HPC machines (Fram, Saga and Betzy)

Users can use any of the above to manage their own software stack on our machines

EasyBuild

EasyBuild can be used as wrapper around different ways of installing software

- Targeting HPC systems

- Automation: less prone for human errors

- Reproducibility: rebuild software stack, (usually) portable to other systems

- Performance: (usually) build from source for particular hardware

- Very pedantic with versioning and dependencies

EasyBuild is tightly connected to the module system and automatically generates module files after installation

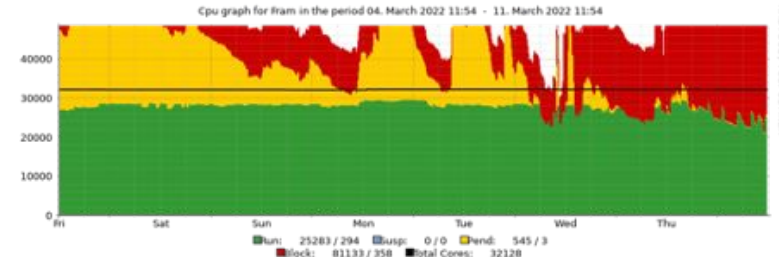
Slurm jobs optimize the usage of HPC systems

Job scripts specify:

memory: Asking for too much can mean that you block idle CPUs and get charged for them

cores: Asking for too few can lead to underused nodes or longer run time

time: Asking for too little cuts the job



General local computing resources

NREC (Norwegian Research and Education Cloud, nrec.no)

-operated by UiB and UiO, the platform is built on OpenStack, users can create virtual machines

Fox: local HPC machine at UiO

Idun: local HPC machine at NTNU



Introduction to Galaxy



Learning Objectives

At the end of this session, you will be able to:

- start exploring the Norwegian Galaxy instance: usegalaxy.no
- test 2000 analysis tools
- disseminate information to other users about this service

The screenshot displays the 'Public Galaxy Servers' dashboard. At the top, there's a navigation bar with 'Public Galaxy Servers' and a 'Zoom Out' button. Below the navigation bar, there's a section titled 'About' which explains the dashboard's purpose: to show information about public galaxy servers and their uptime. It mentions that the data is aggregated from various servers and that the uptime percentages are not perfectly accurate. It also notes that the data is freely accessible and that users can build off it (license CC0). A small 'uptime 100.00%' badge is shown as an example.

The main content area is divided into three sections:

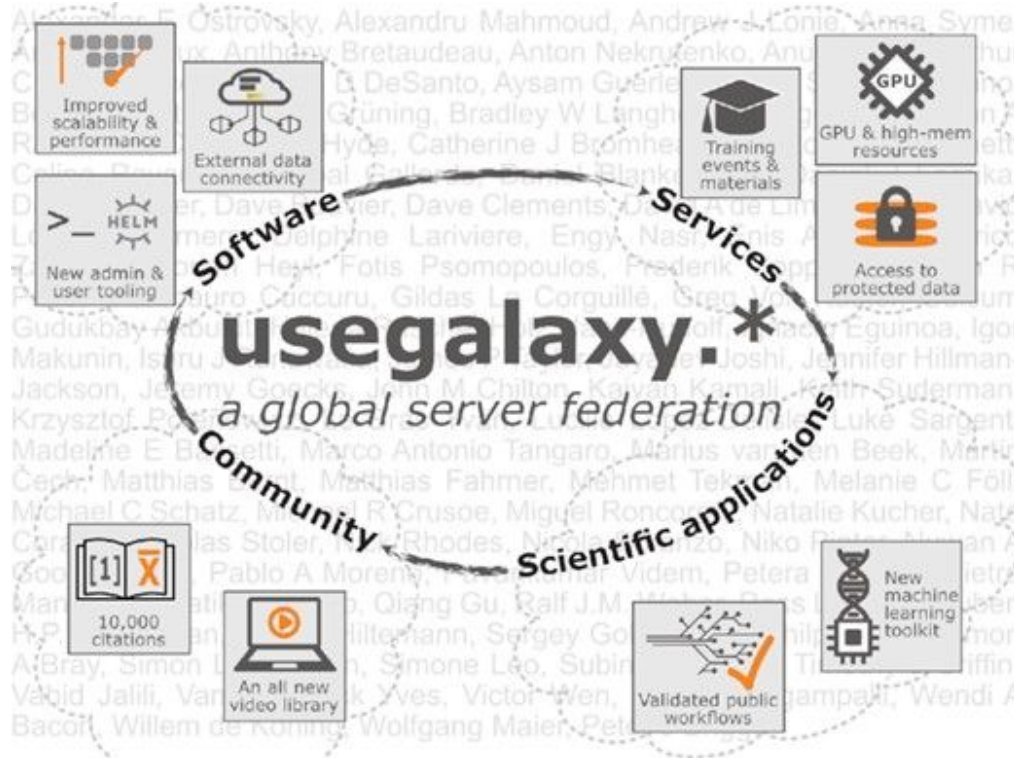
- Galaxies Across the World:** A world map showing the locations of public galaxy servers. The map is color-coded by continent: North America (blue), Europe (purple), Asia (green), Africa (orange), South America (red), and Oceania (yellow). A legend in the bottom left corner indicates the color coding: < 0 (blue), 0+ (red).
- 30 Day Uptime:** A table showing the uptime of various servers over the last 30 days. The table has two columns: 'Server' and 'Uptime'. The servers listed are: workflow4metabolomics (100.00%), BitLAB Galaxy (100.00%), deepTools2 (100.00%), MBAC Metabome Portal (100.00%), ZBIT Bioinformatics Toolbox (100.00%), Whale Shark (100.00%), GVL Melbourne (100.00%), LAPP5 Grid (100.00%), Vinther Lab (100.00%), ABIMS Tools (100.00%), VarCap Galaxy (100.00%), and Biomina (100.00%).
- Response Time:** A table showing the response time of various servers. The table has four columns: 'Server', 'Avg', 'Current', and 'Min'. The servers listed are: BISTRO Galaxy (8 ms, 8 ms, 4 ms), MiModd NacreusMap (19 ms, 19 ms, 14 m), Halogen Bonding Galaxy (40 ms, 38 ms, 32 m), and GenOuest Galaxy (43 ms, 42 ms, 39 m).

At the bottom of the screenshot, there's a URL: <https://galaxyproject.org/blog/2017-10-public-galaxy-dashboard/>.

	AGESeq @ AspenDB	Server	This site provides a user-friendly interface for AGESeq (Analysis of Genome Editing by Sequencing) in a Galaxy instance.	Tools
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usegalaxy.* – Federation of free public Galaxy servers

- Common core set of tools and reference genomes
- Open to anyone to use
- Backed by significant computational resources
- Support from multiple national infrastructure providers
- Galaxy Training Network



<https://academic.oup.com/nar/advance-article/doi/10.1093/nar/gkac247/6572001>



usegalaxy.no – The Norwegian national Galaxy server

- Provide ~2000 bioinformatic tools and workflows
- Open to all Norwegian user and collaborators
- Directly connected to the NeLS storage
- Backed by significant computational resources
- End-user training and support through ELIXIR Norway support desk

NeLS | Galaxy Norway Analyze Data Workflow Visualize Shared Data Help User Using 16%

Tools search tools

Get Data
Send Data
Collection Operations
Lift-Over
Text Manipulation
Convert Formats
Filter and Sort
Join, Subtract and Group
Fetch Alignments/Sequences
Operate on Genomic Intervals
Statistics
Graph/Display Data
Phenotype Association
Interactive Tools
Mapping
SAM/BAM
Annotation
Assembly
Imaging
ChemicalToolBox

Welcome to usegalaxy.no

ELIXIR NORWAY

Galaxy is a web-based platform for data intensive life science research that provides users with a unified, easy-to-use graphical interface to a host of different analysis tools. These tools can be run interactively, one by one, or combined into multi-step workflows that can be executed as a single analysis.

If this is your first time using Galaxy, you might want to have a look at this [Quick Start Guide](#). Additional documentation and tutorials on using Galaxy can be found [here](#).

This Galaxy server has limitations on disc usage, and you have currently used **34.0 GB** of your total quota of **200.0 GB**. To free up disc space, please move your files to the NeLS Storage after you are finished with them. If you require a larger disc quota, contact the [Help Desk](#).

Galaxy version upgrade

UseGalaxy.no has now been upgraded to version 20.09. New features include the ability to upload data directly from the tool form and support for multimedia files. Visit [this page](#) for more information.

Tweets by @eliximorway

ELIXIR Norway @eliximorway
Few spots left on the @swcarpentry course from ELIXIR Norway and @DigitaltLiv for PhD candidates & researchers korbinib.github.io/2021-02-01-DLN...

Katharina Lauer @kauerkatharina
Starting on 20 Jan, a series of 6 webinars will demonstrate that #openscience is key for responding to #COVID19 and public health crises
[bit.ly/38VpdpS](#)

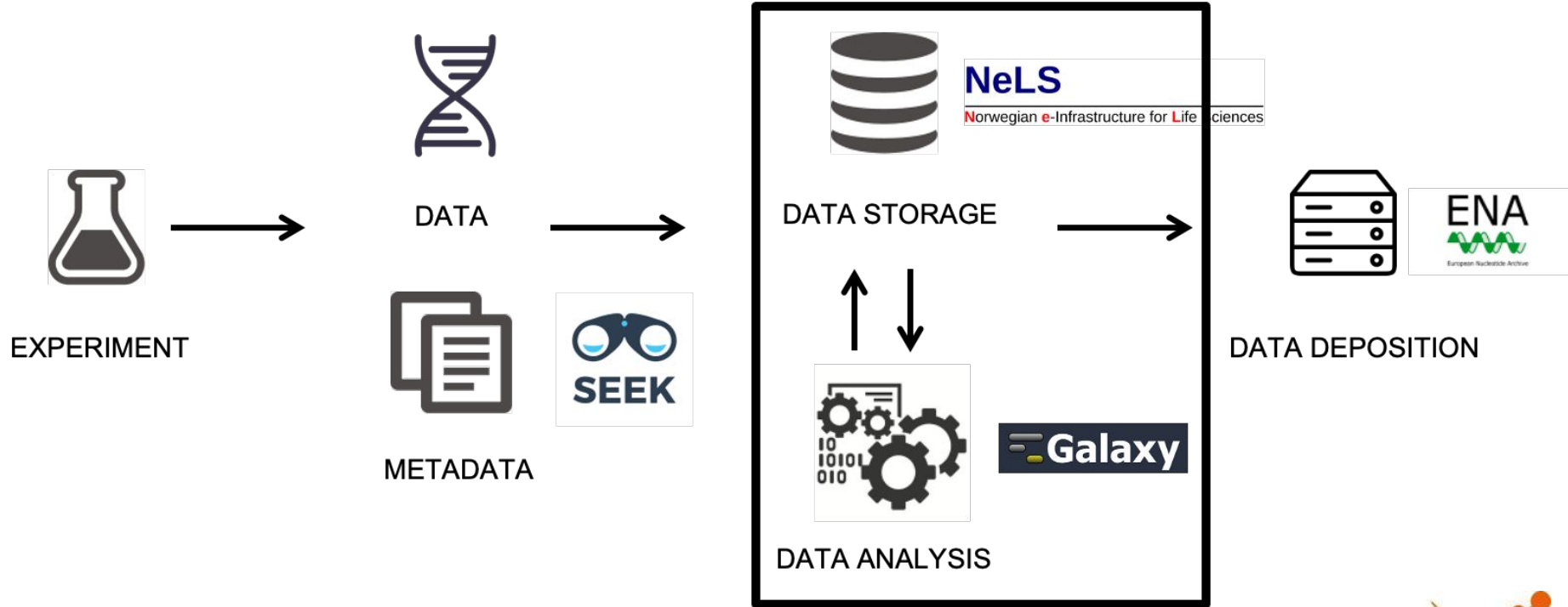
History search datasets

Unnamed history (empty)

This history is empty. You can load your own data or get data from an external source



ELIXIR Norway - analysis connected with storage



usegalaxy.no – The basics

- Important features:
- Tool menu with ~2000 tools sorted in sections
- Current disk usage (default is 200 GB total personal disk space)
- Server alerts
- Quick start guide
- Contact support
- Q&A forum

Main menu

The screenshot shows the main menu of usegalaxy.no. At the top is a navigation bar with links: NLS, Galaxy Norway, Analyze Data, Workflow, Visualize, Shared Data, Help, User, and a grid icon. A 'Using 16%' indicator is on the right. The main content area is divided into three panels. The left panel, titled 'Tools', contains a search bar and a list of tool categories: Get Data, Send Data, Collection Operations, Lift-Over, Text Manipulation, Convert Formats, Filter and Sort, Join, Subtract and Group, Fetch Alignments/Sequences, Operate on Genomic Intervals, Statistics, Graph/Display Data, Phenotype Association, Interactive Tools, Mapping, SAM/BAM, Annotation, Assembly, Imaging, and ChemicalToolBox. The middle panel, titled 'Welcome to usegalaxy.no', contains a welcome message, a link to the Quick Start Guide, and a 'Galaxy version upgrade' notice. The right panel, titled 'History', contains a search bar, a message that the history is empty, and a link to load data from an external source. A 'Tweets' section is also visible in the middle panel.

Tool menu

Main window

History



Your account and saved data

- Important features:
- Tool menu with ~2000 tools sorted in sections
- Current disk usage (default is 200 GB total personal disk space)
- Server alerts
- Quick start guide
- Contact support
- Q&A forum

The screenshot displays the Galaxy Norway web interface. The top navigation bar includes links for 'Analyze Data', 'Workflow', 'Visualize', 'Shared Data', 'Help', and a 'User' dropdown menu, which is highlighted with a red box. The 'Tools' sidebar on the left lists various categories such as 'Get Data', 'Send Data', 'Collection Operations', 'Lift-Over', 'Text Manipulation', 'Convert Formats', 'Filter and Sort', 'Join, Subtract and Group', 'Fetch Alignments/Sequences', 'Operate on Genomic Intervals', 'Statistics', 'Graph/Display Data', 'Phenotype Association', 'Interactive Tools', 'Mapping', 'SAM/BAM', 'Annotation', 'Assembly', 'Imaging', and 'ChemicalToolBox'. The main content area features a 'Welcome to usegalaxy.no' message, a 'History' tab, and a 'Tweets by @elixirnorway' section. A 'Galaxy version upgrade' notification states that the platform has been upgraded to version 20.09, with new features for uploading data and multimedia files. A user menu dropdown is open, showing the user is logged in as 'ehj000@uit.no' and listing options: 'Preferences', 'Custom Builds', 'Logout', 'Datasets', 'Histories', 'Histories shared with me', 'Pages', 'Workflow Invocations', 'Visualizations', and 'Active InteractiveTools'.



Shared data

- Data shared by other users or ELIXIR-NO with all users of usegalaxy.no
- E.g. workflows and complete histories
- You can import shared data to you user
- Instructions how to use ELIXIR-NO supported workflows are also here

The screenshot shows the Galaxy Norway web interface. The top navigation bar includes links for 'Analyze Data', 'Workflow', 'Visualize', 'Shared Data' (highlighted with a red box), 'Help', and 'User'. A left sidebar lists various tools and operations. The main content area displays a welcome message and a 'Data Libraries' panel with links to 'Data Libraries', 'Histories', 'Workflows', 'Visualizations', and 'Pages'. A right sidebar shows the 'History' section, which is currently empty.

Tools

- Get Data
- Send Data
- Collection Operations
- Lift-Over
- Text Manipulation
- Convert Formats
- Filter and Sort
- Join, Subtract and Group
- Fetch Alignments/Sequences
- Operate on Genomic Intervals
- Statistics
- Graph/Display Data
- Phenotype Association
- Interactive Tools
- Mapping
- SAM/BAM
- Annotation
- Assembly
- Imaging
- ChemicalToolBox

Welcome to usegalaxy.no

Galaxy is a web-based platform for data intensive life science research that provides users with a unified, easy-to-use graphical interface to a host of different analysis tools. These tools can be run interactively, one by one, or combined into multi-step workflows that can be executed as a single analysis.

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Data Libraries

- Histories
- Workflows
- Visualizations
- Pages

History

search datasets

Unnamed history

(empty)

This history is empty. You can load your own data or get data from an external source

Shared data: workflows

- List of all workflows that are shared with all usegalaxy.no users
- You can import shared data to your user
- By selecting any workflow you can run data analysis, import into your user or save it on another computer

The screenshot shows the Galaxy Norway interface. The top navigation bar includes 'NoLS', 'Galaxy Norway', and several menu items: 'Analyze Data', 'Workflow', 'Visualize', 'Shared Data' (highlighted with a red box), 'Help', 'User', and a grid icon. On the right of the bar is a 'Using 16%' indicator.

On the left is a 'Tools' sidebar with a search bar and a list of tool categories: Get Data, Send Data, Collection Operations, Lift-Over, Text Manipulation, Convert Formats, Filter and Sort, Join, Subtract and Group, Fetch Alignments/Sequences, Operate on Genomic Intervals, Statistics, Graph/Display Data, Phenotype Association, Interactive Tools, Mapping, SAM/BAM, Annotation, Assembly, Imaging, and ChemicalToolBox.

The main content area is titled 'Published Workflows' and contains a search bar. Below it is a table of workflows:

Name	Annotation	Owner	Community Rating	Community Tags	Last Updated
16S Workflow with Mothur program		kjetil-klepper	★★★★★		Oct 17, 2020
NGS Pipeline for Paired End Reads (R1 and R2)		kjetil-klepper	★★★★★		Oct 17, 2020
miRNA differential expression (miRBase, hg38)		kjetil-klepper	★★★★★	nels	Oct 15, 2020
miRNA differential expression (MirGeneDB, hg38)		kjetil-klepper	★★★★★	nels	Oct 14, 2020
COVID-19: PE Variation		kjetil-klepper	★★★★★	nels	Oct 13, 2020
Pre-process COVID-19 PE collections		kjetil-klepper	★★★★★	nels	Oct 13, 2020
Pre-process COVID-19 PE single sample		kjetil-klepper	★★★★★	nels	Oct 13, 2020

A context menu is open over the table, showing three options: 'Run', 'Import', and 'Save as File'.

On the right side of the interface is a 'History' panel with a search bar and a message: 'Unnamed history (empty). This history is empty. You can load your own data or get data from an external source'.

Workflows

- Your workflows. These are the imported or the workflows you have made
- You can create new workflows here

The screenshot displays the Galaxy Norway interface. The top navigation bar includes 'NoLS', 'Galaxy Norway', and several menu items: 'Analyze Data', 'Workflow' (highlighted with a red box), 'Visualize', 'Shared Data', 'Help', and 'User'. Below the navigation bar, the 'Tools' sidebar on the left lists various categories like 'Get Data', 'Send Data', 'Collection Operations', etc. The main content area shows a 'Search Workflows' bar and a table of workflows. The table has columns for 'Name', 'Tags', 'Updated', 'Sharing', and 'Bookmarked'. Two workflows are listed: 'imported: miRNA differential expression (miRBase, hg38)' and 'Taxonomic_profiling_Metaphlan2'. The right sidebar shows 'History' with a search bar and a message indicating it is empty.

Name	Tags	Updated	Sharing	Bookmarked
imported: miRNA differential expression (miRBase, hg38)		3 days ago		<input type="checkbox"/>
Taxonomic_profiling_Metaphlan2	metagenomics	2 months ago		<input type="checkbox"/>

Import data from your computer or from the web

- Drag and drop, or browse and select file
- Alternatively, paste url for data available on the web
- Specify data type if you know (e.g. Fasta)
- The two imported files will appear as two datasets in your history

The screenshot displays the Galaxy Norway web interface. The top navigation bar includes links for 'Tools', 'Analyze Data', 'Workflow', 'Visualize', 'Shared Data', 'Help', and 'User'. The left sidebar lists various tool categories such as 'Get Data', 'Send Data', 'Collection Operations', 'Lift-Over', 'Text Manipulation', 'Convert Formats', 'Filter and Sort', 'Join, Subtract and Group', 'Fetch Alignments/Sequences', 'Operate on Genomic Interactions', 'Statistics', 'Graph/Display Data', 'Phenotype Association', 'Interactive Tools', 'Mapping', 'SAM/BAM', 'Annotation', 'Assembly', 'Imaging', and 'ChemicalToolBox'. The main content area shows a 'Welcome to usegalaxy.no' message and a 'Tweets by @elixirnorway' widget. A modal window titled 'Download from web or upload from disk' is open, showing a table of files being uploaded. The table has columns for 'Name', 'Size', 'Type', 'Genome', 'Settings', and 'Status'. Two files are listed: 'test_upload.txt' (9 b, txt) and 'test_upload.fasta' (9 b, Auto-de...). A dropdown menu is open for the 'test_upload.fasta' file, showing options: 'fasta', 'csfasta', 'fasta', and 'fasta.gz'. The 'fasta' option is selected. On the right side, the 'History' panel shows 'Unnamed history' with 2 shown and 1 deleted. Two datasets are listed: '2: test_upload.fasta' and '1: test_upload.txt', both with icons for viewing, editing, and deleting.

Name	Size	Type	Genome	Settings	Status
test_upload.txt	9 b	txt	----- Additional ...	⚙	0%
test_upload.fasta	9 b	Auto-de...	----- Additional ...	⚙	0%

fasta
csfasta
fasta
fasta.gz

2: test_upload.fasta
1: test_upload.txt

Import data from NeLS

- Import data from Personal or Project folders in NeLS
- Redirect to the NeLS portal (require login)
- Files are selectable
- Imported data from NeLS will appear in your history
- Note: the yellow colour of files as this jobs are being processed (green = job complete)

The screenshot displays the ELIXIR Norway Galaxy web interface. The top navigation bar includes 'Analyze Data', 'Workflow', 'Visualize', 'Shared Data', 'Help', and 'User'. The left sidebar lists various tools, with 'Get Data' highlighted by a red box. The main content area shows a 'Welcome to usegalaxy.no' message and a 'Tweets by @elixirnorway' section. A 'NeLS' modal window is open, showing the path 'Home / Projects / ELIXIR_online_course_2021 / intro_usegalaxy' and a table with one file: 'test_NeLS_import.txt' (72 bytes). A 'Send to Galaxy' button is visible. The right sidebar shows the 'History' panel, which lists three datasets: '3: Get files' (highlighted with a red box), '2: test_upload.fasta', and '1: test_upload.txt'. The '3: Get files' entry is yellow, indicating it is being processed, while the others are green, indicating they are complete.



Use galaxy histories to organise data

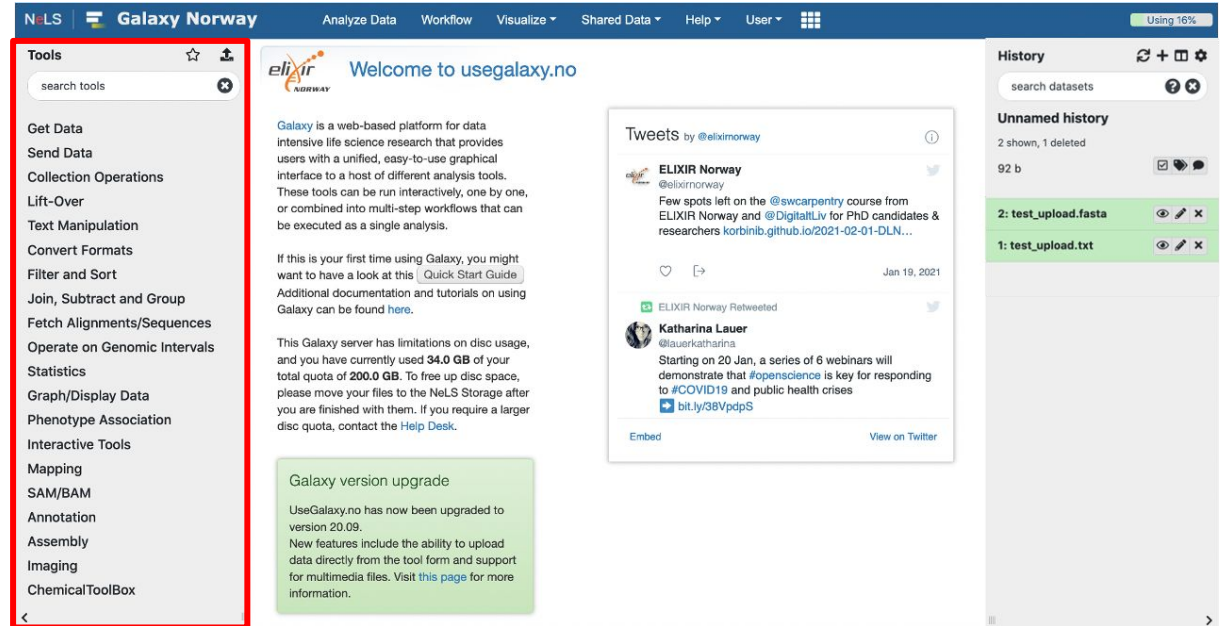
- The current history is “your current work space”
- The history panel displays datasets in the order in which they were created
- You can make as many histories as you want and switch between them
- Typically, you can have one history for each project or analysis

The screenshot displays the Galaxy Norway web interface. The top navigation bar includes links for 'Analyze Data', 'Workflow', 'Visualize', 'Shared Data', 'Help', and 'User'. The main content area is divided into three columns. The left column contains a 'Tools' panel with a search bar and a list of tools such as 'Get Data', 'EGA Download Client', 'NCBI ESummary', 'NCBI ESearch', 'NCBI EPost', 'NCBI ELink', 'NCBI EInfo', 'NCBI EQuery', 'NCBI EFetch', 'NCBI ECI', 'IEDB MHC Binding prediction', 'UniProt download', 'UniProt ID mapping', 'UniProt retrieve', and 'Download and Generate Pileup'. The middle column shows a 'Welcome to usegalaxy.no' message and a 'Galaxy version upgrade' notification. The right column features a 'History' panel, which is highlighted with a red rectangle. This panel shows a search bar and a list of datasets under the heading 'Unnamed history'. The datasets listed are '3: test_NeLS_import', '2: test_upload.fasta', and '1: test_upload.txt', each with icons for viewing, editing, and deleting. The interface also includes a 'Tweets' section and a 'Galaxy version upgrade' notification.



Galaxy tools

- Tools are available from the Tool menu
- Organised under sub-menus
- Possible to browse and search by name
- You can make your own list of favourite tools



The screenshot displays the Galaxy Norway web interface. On the left, a sidebar menu titled 'Tools' is highlighted with a red box. This menu includes sub-menus such as 'Get Data', 'Send Data', 'Collection Operations', 'Lift-Over', 'Text Manipulation', 'Convert Formats', 'Filter and Sort', 'Join, Subtract and Group', 'Fetch Alignments/Sequences', 'Operate on Genomic Intervals', 'Statistics', 'Graph/Display Data', 'Phenotype Association', 'Interactive Tools', 'Mapping', 'SAM/BAM', 'Annotation', 'Assembly', 'Imaging', and 'ChemicalToolBox'. The main content area features a 'Welcome to usegalaxy.no' message, a tweet from ELIXIR Norway about a course, and a 'Galaxy version upgrade' notice stating that the platform has been upgraded to version 20.09. On the right, a 'History' sidebar shows a list of datasets, including 'test_upload.fasta' and 'test_upload.txt'.



Galaxy tools = command line tools

- Command line tools are wrapped into Galaxy so they become accessible with a GUI
- Some Galaxy tools may have reduced the number of optional parameter settings for the tool
- Example here is the assembly tool called SPAdes

The screenshot displays the Galaxy Norway web interface. On the left, a sidebar contains a 'Tools' section with a search bar and a list of tools under categories like 'Assembly', 'Create assemblies with Unicycler', and 'Graph/Display Data'. The 'SPAdes genome assembler for regular and single-cell projects' tool is highlighted. The main panel shows the configuration for this tool, titled 'SPAdes genome assembler for regular and single-cell projects (Galaxy Version 3.12.0+galaxy1)'. It includes options for 'Single-cell?' (Yes/No), 'Run only assembly? (without read error correction)' (Yes/No), 'Careful correction?' (Yes/No), 'Automatically choose k-mer values' (Yes/No), 'K-mers to use, separated by commas' (a text input field containing '21,33,55'), 'Coverage Cutoff' (a dropdown menu set to 'Off'), and 'Libraries are IonTorrent reads?' (Yes/No). On the right, a 'History' panel shows a list of datasets, including 'test_upload.fasta' and 'test_upload.txt'.

Galaxy tools = command line tools

```
SPAdes genome assembler v3.11.1
Usage: /Users/service/tools/SPAdes/SPAdes-3.11.1-Darwin/bin/spades.py [options] -o <output_dir>

Basic options:
-o <output_dir>    directory to store all the resulting files (required)
--meta            this flag is required for MDA (single-cell) data
--rna            this flag is required for metagenomic sample data
--plasmid        runs plasmidSPAdes pipeline for plasmid detection
--iontorrent     this flag is required for IonTorrent data
--test          runs SPAdes on toy dataset
-h/-help        prints this usage message
-v/--version     prints version

Input data:
--12 <filename>    file with interleaved forward and reverse paired-end reads
--1 <filename>     file with forward paired-end reads
--2 <filename>     file with reverse paired-end reads
--s <filename>     file with unpaired reads
--pe<#>-12 <filename> file with interleaved reads for paired-end library number <#> (<#> = 1,2,...,9)
--pe<#>-1 <filename>  file with forward reads for paired-end library number <#> (<#> = 1,2,...,9)
--pe<#>-2 <filename>  file with reverse reads for paired-end library number <#> (<#> = 1,2,...,9)
--pe<#>-s <filename>  file with unpaired reads for paired-end library number <#> (<#> = 1,2,...,9)
--pe<#>-cor <#> orientation of reads for paired-end library number <#> (<#> = 1,2,...,9; <cor> = fr, rf, ff)
--s<#> <filename>     file with unpaired reads for single reads library number <#> (<#> = 1,2,...,9)
--mp<#>-12 <filename> file with interleaved reads for mate-pair library number <#> (<#> = 1,2,...,9)
--mp<#>-1 <filename>  file with forward reads for mate-pair library number <#> (<#> = 1,2,...,9)
--mp<#>-2 <filename>  file with reverse reads for mate-pair library number <#> (<#> = 1,2,...,9)
--mp<#>-s <filename>  file with unpaired reads for mate-pair library number <#> (<#> = 1,2,...,9)
--mp<#>-cor <#> orientation of reads for mate-pair library number <#> (<#> = 1,2,...,9; <cor> = fr, rf, ff)
--hqmp<#>-12 <filename> file with interleaved reads for high-quality mate-pair library number <#> (<#> = 1,2,...,9)
--hqmp<#>-1 <filename>  file with forward reads for high-quality mate-pair library number <#> (<#> = 1,2,...,9)
--hqmp<#>-2 <filename>  file with reverse reads for high-quality mate-pair library number <#> (<#> = 1,2,...,9)
--hqmp<#>-s <filename>  file with unpaired reads for high-quality mate-pair library number <#> (<#> = 1,2,...,9)
--hqmp<#>-cor <#> orientation of reads for high-quality mate-pair library number <#> (<#> = 1,2,...,9; <cor> = fr, rf, ff)
--xmate<#>-1 <filename> file with forward reads for Lucigen NxMate library number <#> (<#> = 1,2,...,9)
--xmate<#>-2 <filename> file with reverse reads for Lucigen NxMate library number <#> (<#> = 1,2,...,9)
--sanger <filename>   file with Sanger reads
--pacbio <filename>   file with PacBio reads
--nanopore <filename> file with Nanopore reads
--tslr <filename>     file with TSLR-contigs
--trusted-contigs <filename> file with trusted contigs
--untrusted-contigs <filename> file with untrusted contigs

Pipeline options:
--only-error-correction runs only read error correction (without assembling)
--only-assembler runs only assembling (without read error correction)
--careful tries to reduce number of mismatches and short indels
--continue continue run from the last available check-point
--restart-from <cp> restart run with updated options and from the specified check-point ('ec', 'as', 'k<int>', 'mc')
--disable-gzip-output forces error correction not to compress the corrected reads
--disable-rr disables repeat resolution stage of assembling

Advanced options:
--dataset <filename> file with dataset description in YAML format
--t/--threads <int> number of threads
                        [default: 16]
--m/--memory <int> RAM limit for SPAdes in Gb (terminates if exceeded)
                        [default: 250]
--tmp-dir <dirname> directory for temporary files
                        [default: <output_dir>/tmp]
--k <int,int,...> comma-separated list of k-mer sizes (must be odd and
```

Galaxy Norway

Analyze DataWorkflowVisualizeShared DataHelpUser

Tools

SPAdes genome assembler for regular and single-cell projects (Galaxy Version 3.12.0+galaxy1)

FavoriteOptions

Single-cell?

YesNo

This option is required for MDA (single-cell) data. (--sc)

Run only assembly? (without read error correction)

YesNo

(--only-assembler)

Careful correction?

YesNo

Tries to reduce number of mismatches and short indels. Also runs MismatchCorrector – a post processing tool, which uses BWA tool (comes with SPAdes). (--careful)

Automatically choose k-mer values

YesNo

k-mer choices can be chosen by SPAdes instead of being entered manually

K-mers to use, separated by commas

21,33,55

Comma-separated list of k-mer sizes to be used (all values must be odd, less than 128, listed in ascending order, and smaller than the read length). The default value is 21,33,55.

Coverage Cutoff

Off

Libraries are IonTorrent reads?

YesNo

History

search datasets

Unnamed history

2 shown, 1 deleted

92 b

2: test_upload.fasta

1: test_upload.txt



Tool output

- It is possible to preview the output (result), view it in the main window or download the dataset
- You can also copy the dataset over to another history

The screenshot displays the Galaxy Norway web interface. The top navigation bar includes links for 'Analyze Data', 'Workflow', 'Visualize', 'Shared Data', 'Help', and 'User'. The left sidebar contains a 'Tools' section with a search bar and a list of tools under 'Assembly', including 'SPAdes genome assembler for regular and single-cell projects' and 'metaSPAdes assembler for metagenomics datasets'. The main content area shows the output of a tool, with a warning message: 'This dataset is large and only the first megabyte is shown below.' Below this, a large block of text represents the output, starting with '>NODE_1_length_285604_cov_21.023695'. The right sidebar features a 'History' section with a search bar and a list of datasets. A specific dataset is highlighted in a red box, showing its details: '6: SPAdes on data 2 and data 1: scaffolds (fasta)', '7853 sequences', 'format: fasta, database: ?', and the command line used for its generation. The interface also shows a 'Download' button and a 'System' status indicator.



Galaxy workflows

- A workflow in Galaxy is basically a string of tools, where the output from one tool becomes the input for the next



Galaxy workflows

- The “nodels” indicate which output file from one acts as input for the next tool
- Each workflow has a name and version
- Additional text that describe the workflow and tags can be added

The screenshot displays the Galaxy Norway web interface. On the left is a sidebar with a search bar and a list of tool categories: Inputs, Data Managers, Get Data, Send Data, Collection Operations, Expression Tools, Lift-Over, Text Manipulation, Convert Formats, Filter and Sort, Join, Subtract and Group, Fetch Alignments/Sequences, Operate on Genomic Intervals, Statistics, Graph/Display Data, Phenotype Association, Interactive Tools, Mapping, SAM/BAM, Annotation, and Assembly. The main workspace shows a workflow titled 'Exercise II' with four tools connected in a sequence: 'Forward reads (R1)', 'Reverse reads (R1)', 'Trimmomatic', and 'Kraken'. The 'Trimmomatic' tool has multiple input/output options, and the 'Kraken' tool is configured for taxonomic classification. On the right, a panel for the workflow 'Exercise II' shows its version (1, 6 steps active), an annotation ('Taxonomic classification of metagenomes using Kraken'), and a tag ('Metagenomic'). A small thumbnail of the workflow diagram is visible at the bottom right of the main workspace.

Galaxy workflows

- New tools can be added by clicking on the tool in the Tool menu
- The tool will appear in the workflow editor
- Tool parameter settings can be pre-set or made up to the user to set when running the workflow

The screenshot displays the Galaxy Norway web interface. On the left, the 'Tools' panel is open, showing a search for 'krona'. The 'Krona pie chart from taxonomic profile' tool is highlighted with a red box. The main workspace shows a workflow titled 'Exercise II' with several steps: 'Forward reads (R1)', 'Reverse reads (R1)', 'Trimomatic', 'Kraken', and 'Convert Kraken'. A red box highlights the 'Krona pie chart' tool icon in the workflow. On the right, the configuration panel for the 'Krona pie chart from taxonomic profile' tool is shown, also with a red border. It includes fields for 'Label', 'Annotation', and 'Input file'. The 'Input file' section is expanded, showing options for 'Data input' (set to 'input') and 'Taxonomy dataset' (set to 'Classification (tabular)'). The 'show ranks from root to' checkbox is checked.

Galaxy workflows

- The output from another tool can be connected as an input for the new tool

The screenshot displays the Galaxy Norway web interface. On the left, a 'Tools' sidebar is open, showing a search for 'krona'. Under the 'Metagenomic Analysis' section, the 'Krona pie chart from taxonomic profile' tool is highlighted. The main workspace, titled 'Exercise II', shows a workflow diagram. The workflow starts with 'Forward reads (R1)' and 'Reverse reads (R2)' as inputs to 'Trimmomatic'. The output of Trimmomatic is connected to 'Kraken', which is set to 'Forward strand' and 'Reverse strand'. The output of Kraken is then connected to 'Convert Kraken', which is set to 'Choose dataset to convert'. The output of Convert Kraken is connected to 'Krona pie chart', which is set to 'Input file'. A red rectangle highlights the connection between the 'Convert Kraken' tool and the 'Krona pie chart' tool. On the right, the 'Krona pie chart' tool's configuration panel is visible, showing options for 'Label', 'Annotation', and 'Input file'.

Galaxy workflows

- Select input files
- Tools in the workflow will run successively
- You can view and download the result files

The screenshot displays the Galaxy Norway web interface. The top navigation bar includes links for 'Analyze Data', 'Workflow', 'Visualize', 'Shared Data', 'Help', and 'User'. The main content area is titled 'Workflow: Taxonomic_profiling_Metaphlan2' and features a 'Run Workflow' button. On the left, a 'Tools' sidebar lists various assembly and analysis tools. The central workflow area shows three steps: '1: Forward reads (R1)', '2: Reverse reads (R2)', and '3: FastQC (Galaxy Version 0.72+galaxy1)'. Each step has input fields and download icons. The right sidebar shows the 'History' section with a list of datasets, including 'test_run_16S' and various intermediate files like 'LCAClassifier on data 7: Taxonomic tree' and 'Predicted 16S rRNA reads from data 5'.

Reproducibility & Transparency




Dataset Information

Number	37
Name	DESeq on data 33 and data 32: PCA plot
Created	Friday Dec 4th 8:16:43 2020 UTC
Filesize	5.8 KB
Dbkey	?
Format	png
File contents	contents
History Content API ID	7ba0a550406c9f38 (9272)
History API ID	39520843fe032123 (438)
UUID	a22dce33-aaf2-4b0b-be3f-8b5241195694
Full Path	/data/part0/008/dataset_8860.dat

Dataset Information

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Full Path	/data/part0/008/dataset_8860.dat

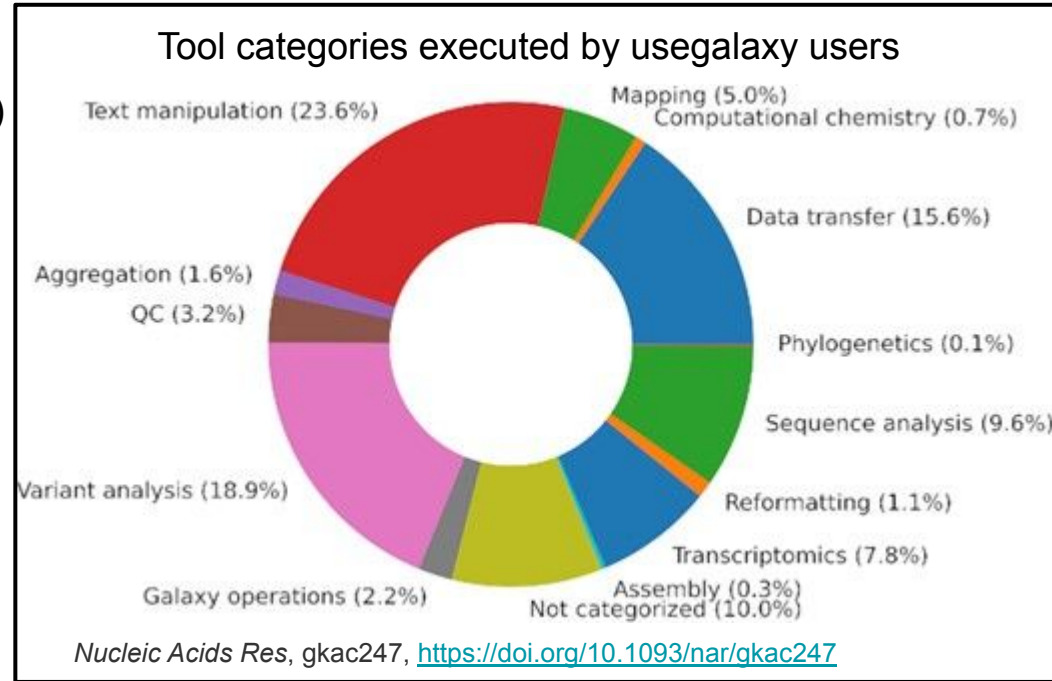
Job Information

Galaxy Tool ID:	galaxy-ntnu.bioinfo.no/toolshed/uit_deseq2_wrapper/0.0.1 
Command Line	Rscript /srv/galaxy/var/shed_tools/galaxy-ntnu.bioinfo.no/too... 
Tool Standard Output	Loading required package: S4Vectors Loading required package:... 
Tool Standard Error	empty
Tool Exit Code:	0
Job API ID:	2ade379cb271bcc4 (5499)



Why use usegalaxy.no?

- Easy to use
- Connected directly with storage (NeLS)
- Backed by significant computational resources
- Training and support
- Transparent research
- Reproducible research
- Enable easy sharing of data and tools



Quiz questions

Introduction to Galaxy:

1. usegalaxy.no is a web platform for:
 - a. Storing life science data
 - b. Analysing life science data
 - c. Developing bioinformatic tools
 - d. Generating data management plans





Useful Resources

- usegalaxy.no
- usegalaxy.eu

Workflow Management Systems



Learning Objectives

At the end of this session, you will be able to:

- Conveniently use tonnes of ready-made Nextflow pipelines to analyse your data in a FAIR manner
- Utilize auxiliary resources such as Biocontainers, Workflowhub and nf-core to find containers and workflows enabling FAIR analyses

What are workflow management systems (WfMS)?

- “Standards for describing computational data-analysis workflows”
- Streamline routine processes for optimal efficiency
- Handles input / output / execution of tools in a DAG-like manner
- Comes in various colors and flavours

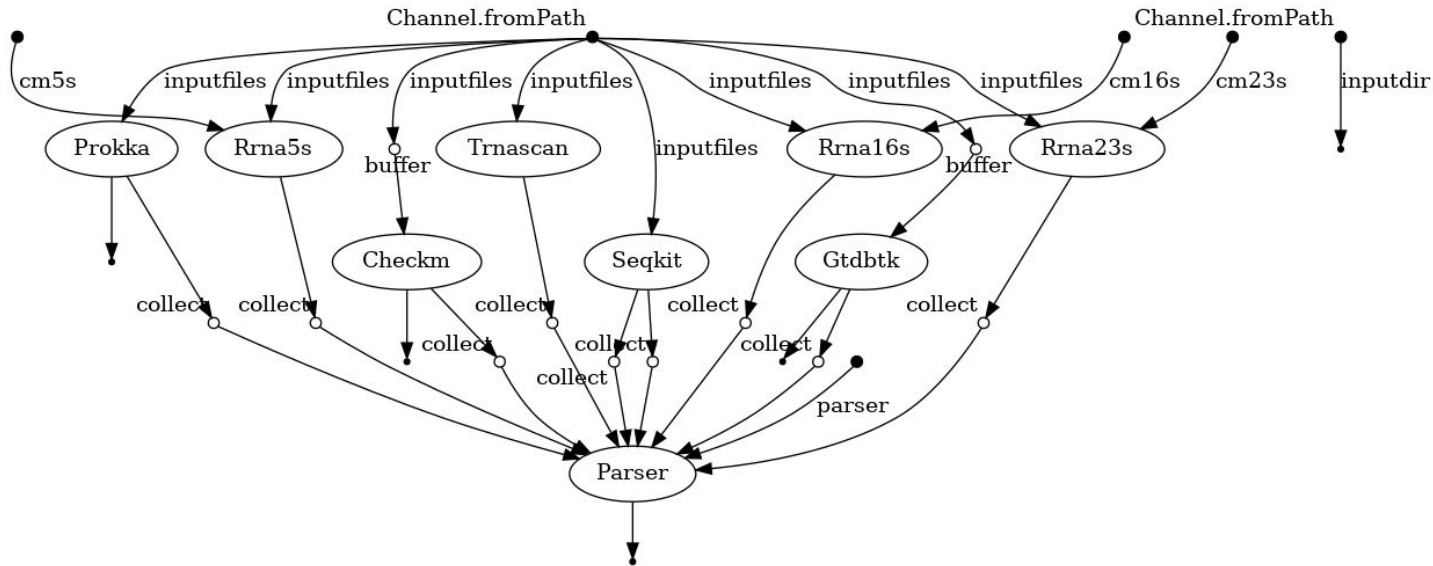
The logo for Nextflow, featuring the word "nextflow" in a sans-serif font. "next" is green and "flow" is black.

COMMON
WORKFLOW
LANGUAGE



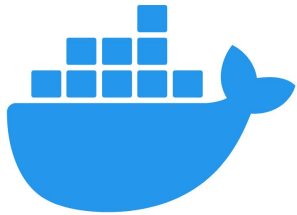
What are workflow management systems (WfMS)?

- Directed acyclic graphs (DAG) can be used to describe workflows visually



WfMS' enhances the principles of "FAIR"

- WfMS's provide an analysis description in their own right
- Enables portability and reproducibility in different computational environments
- Versioning and container integration ensures uniform data analysis



docker



podman



Most WfMS integrates with containers, which makes Biocontainers.pro very useful!

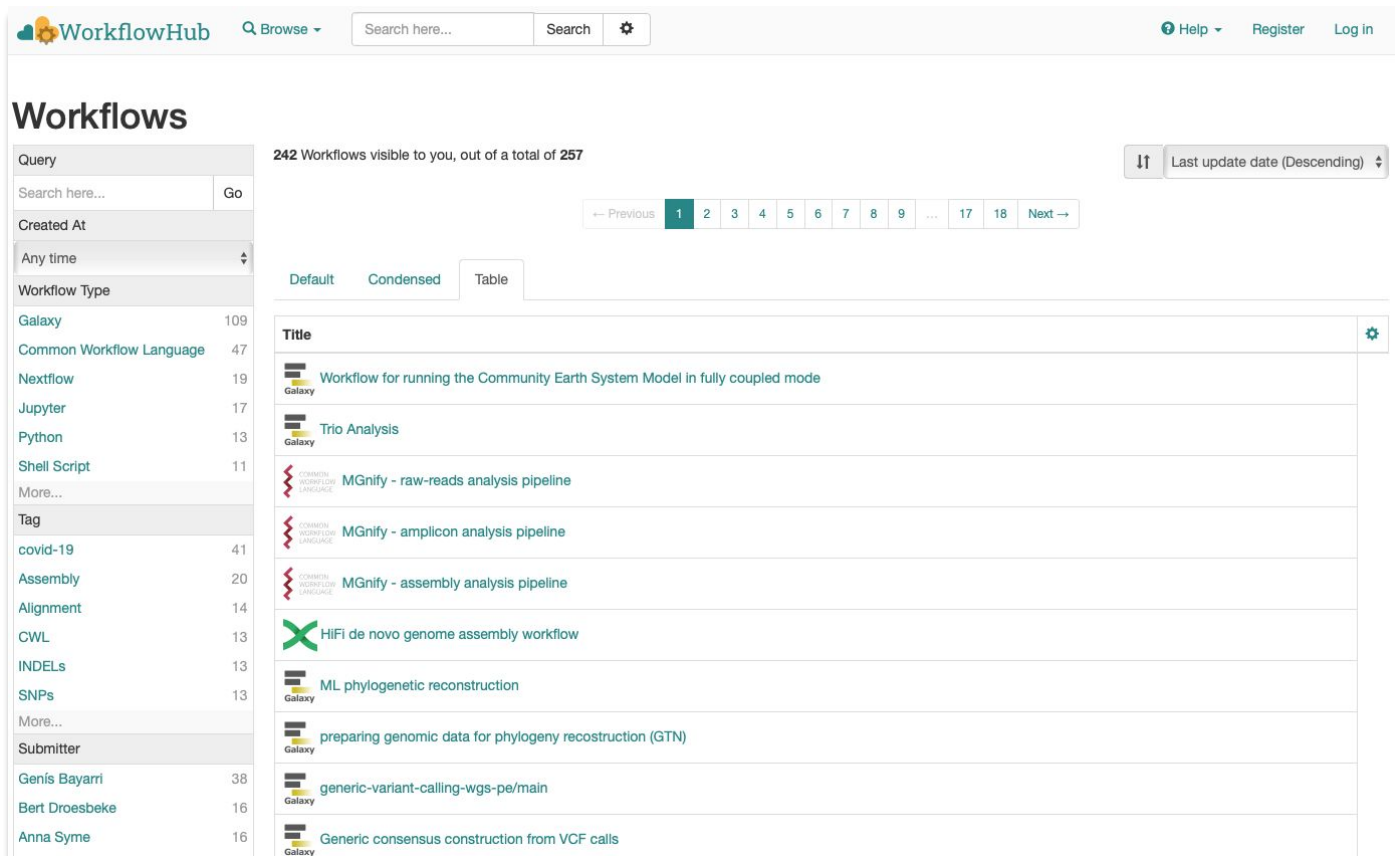
- 10.6K tools, 45.5K versions, 225.3K containers and packages
- Dockerfile recipes / Conda recipes to automatically build containers in BioContainers.
- Biocontainers Registry is a hosted registry of all BioContainers images that are ready to be used

BioContainers Flow



BIOCONDA[®]

Workflowhub covers you workflow needs and then some!



The screenshot displays the WorkflowHub interface. At the top, there is a navigation bar with the WorkflowHub logo, a 'Browse' dropdown, a search bar, and links for 'Help', 'Register', and 'Log in'. Below the navigation bar, the 'Workflows' section is active. On the left, a sidebar contains filters for 'Query', 'Created At', 'Workflow Type', and 'Tag'. The 'Workflow Type' filter is expanded, showing a list of workflow types with their respective counts. The main content area shows a list of 242 workflows visible to the user. The list is sorted by 'Last update date (Descending)'. The first few workflows are:

Workflow Type	Count
Galaxy	109
Common Workflow Language	47
Nextflow	19
Jupyter	17
Python	13
Shell Script	11
More...	
Tag	
covid-19	41
Assembly	20
Alignment	14
CWL	13
INDELs	13
SNPs	13
More...	
Submitter	
Genis Bayarri	38
Bert Dreesbeke	16
Anna Syme	16

The main list of workflows includes:

- Workflow for running the Community Earth System Model in fully coupled mode
- Trio Analysis
- MGnify - raw-reads analysis pipeline
- MGnify - amplicon analysis pipeline
- MGnify - assembly analysis pipeline
- HiFi de novo genome assembly workflow
- ML phylogenetic reconstruction
- preparing genomic data for phylogeny reconstruction (GTN)
- generic-variant-calling-wgs-pe/main
- Generic consensus construction from VCF calls

Nextflow.io

- Supports Bash, Python, R ...
- Dataflow handled by Groovy (Java super-set, “python for Java”)
- Reproducibility
- Lots of “premium” features for free
- <https://www.nextflow.io/docs/latest/>

```
nextflow.enable.dsl=2

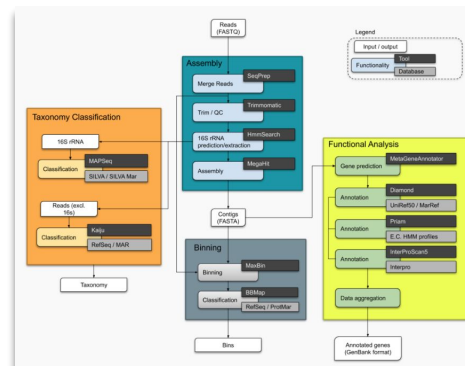
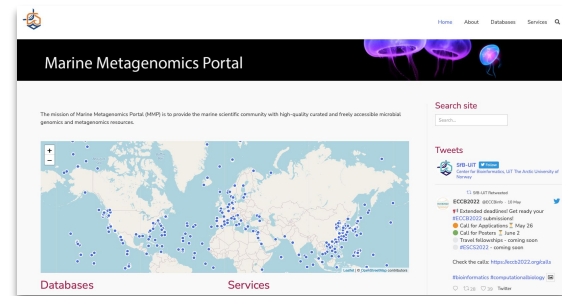
process sayHello {
    input:
        val cheers
    output:
        stdout

    """
    echo $cheers
    """
}

workflow {
    channel.of('Ciao','Hello','Hola') | sayHello | view
}
```

Some examples of Nextflow usage in ELIXIR

- Mar-databases backend processing
 - Results integrated with and available at portal pages
 - <https://gitlab.com/uit-sfb/autoget-nextflow>
- FHI-pipeline utilizing TSD
 - Desensitization pipeline for Norwegian covid19-samples affected by GDPR
 - <https://gitlab.com/uit-sfb/fhi-desensitize>
- Metapipe
 - Metagenomic analysis pipeline developed at UiT (ELIXIR)
 - <https://gitlab.com/uit-sfb/metapipe>
 - <https://mmp2.sfb.uit.no/metapipe/>



nf-core is a great resource!

- Out-of-the-box high quality, production-ready, curated analysis pipelines built using Nextflow. (66 as of Jun, 2022)
- Maintained and validated releases ensure reproducibility.
- nf-core-tools for convenience (list, launch, create templates etc.)
- Excellent documentation!

nf-core is a great resource!

Available Pipelines

Can you think of another pipeline that would fit in well? [Let us know!](#)

Search keywords

Filter:

Released 37

Under development 24

Archived 6

Sort:

Last Release

Alphabetical

Stars

Display:

Grid List

[nf-core/sarek](#) ✓

★ 169

annotation cancer gatk4 genomics germline pre-processing somatic target-panels
variant-calling whole-exome-sequencing whole-genome-sequencing

Analysis pipeline to detect germline or somatic variants (pre-processing, variant calling and annotation) from WGS / targeted sequencing

Version 2.7.2

Published 3 days ago

[nf-core/cutandrun](#) ✓

★ 27

cutandrun cutandrun-seq cutandtag cutandtag-seq

Analysis pipeline for CUT&RUN and CUT&TAG experiments that includes QC, support for spike-ins, IgG controls, peak calling and downstream analysis.

Version 2.0

Published 5 days ago

[nf-core/airflow](#) ✓

★ 19

airr b-cell immcantation immunorepertoire repseq

B-cell and T-cell Adaptive Immune Receptor Repertoire (AIRR) sequencing analysis pipeline using the Immcantation framework

Version 2.2.0

Published 1 week ago

[nf-core/circdna](#) ✓

★ 4

circle-seq circular dna ecDNA genomics

Pipeline for the identification of extrachromosomal circular DNA (ecDNA) from Circle-seq, WGS, and ATAC-seq data that were generated from cancer and other eukaryotic cells.

Version 1.0.0

Published 2 weeks ago

[nf-core/smrnaseq](#) ✓

★ 40

small-rna smrna-seq

A small-RNA sequencing analysis pipeline

Version 2.0.0

Published 2 weeks ago

[nf-core/rnaseq](#) ✓

★ 469

rna rna-seq

RNA sequencing analysis pipeline using STAR, RSEM, HISAT2 or Salmon with gene/isoform counts and extensive quality control.

Version 3.8.1

Published 2 weeks ago



Nextflow tower

- “Centralized command post”; monitoring, logging & observability of distributed workflows conveniently available in your browser
- Simplifies the deployment of pipelines on any cloud, cluster or laptop.
- Quite expensive, but some essential features are free



Enough talk! Let's play around with nextflow!

If we have time :)

- Quick tour / usage of nf-tower
- Launch nextflow pipeline -with-tower

<https://gitlab.com/uit-sfb/autoget-nextflow>

```
# Install nextflow
curl -s https://get.nextflow.io | bash
mv nextflow ~/bin/

# Launch the RNAseq pipeline
nextflow run nf-core/rnaseq \
  --input samplesheet.csv \
  --genome GRCh37 \
  -profile docker

# Install nf-core tools
pip install nf-core

# List all nf-core pipelines and show available updates
nf-core list
```

Useful Resources

- Nextflow, <https://nextflow.io>
- nf-core, <https://nf-co.re>
- nf-tower, <https://tower.nf>
- Biocontainers, <https://biocontainers.pro>
- Marine Metagenomics Portal, <https://mmp2.sfb.uit.no>
- Metapipe, <https://mmp2.sfb.uit.no/metapipe/>

Thank you!



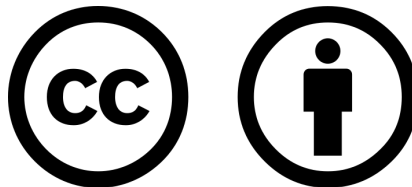
elixir-norway.org



@elixirnorway



contact@bioinfo.no



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