

Supplementary Information

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Appendix S1. Descriptions of sites included in this analysis

Barro Colorado Nature Monument, Panama

Cores were collected within the Barro Colorado Nature Monument (BCNM), a 5,600-ha reserve established in 1923 that includes Barro Colorado Island (BCI) and the surrounding mainland peninsulas (Alfaro-Sánchez et al., 2017). BCNM contains a mix of old-growth and mature secondary tropical moist forest. Live canopy trees on the peninsulas were cored between January 2015 and April 2015. In addition, several fallen dead trees on BCI were sampled by taking discs with chain saws.

We note that the secondary forest status of much of BCNM differs from ForestGEO's 50-ha plot on BCI, which is old-growth, and shows little directional trend in woody productivity (Rutishauser et al., 2020).

Huai Kha Khaeng, Thailand

Cores were collected within a ~300 ha acre area of seasonal dry evergreen and mixed deciduous forest in the Huai Kha Khaeng Wildlife Sanctuary (HKK). The site underwent a widespread, catastrophic disturbance in the mid-1800s and several smaller, more localized disturbances in the 1910s, 1940s, and 1960s (Baker et al., 2005).

Cores were collected between December 2010 and December 2011, sampling trees >5 cm diameter at breast height (DBH) with the goal of representing a random subset of the local population of the target species. Details are given in Vlam et al. (2014).

Smithsonian Conservation Biology Institute, Virginia, USA

Cores were collected within the ForestGEO plot at the Smithsonian Conservation Biology Institute (SCBI). The forest is a secondary temperate broadleaf deciduous forest, which developed following agricultural abandonment in the mid 19th-century, dominated by the species included in our analysis (Bourg et al., 2013). Specifically, the 14 species sampled together account for 97% of woody productivity at the site (Helcoski et al., 2019).

In 2010-2011, cores were collected from a random subset of live trees ≥ 10 cm DBH. In 2016 and 2017, cores were extracted from trees found newly dead during an annual tree mortality census (Gonzalez-Akre et al., 2016). Details are given in Helcoski et al. (2019).

Lilly Dickey Woods, Indiana, USA

Lilly Dickey Woods (LDW) is a secondary temperate broadleaf deciduous forest dominated by oaks (*Quercus* spp.) and hickories (*Carya* spp.).

Cores were collected outside the ForestGEO plot, but within a contiguous patch of forest. In 2012 - 2014, cores were collected from live or dead dominant canopy trees with morphological characteristics typical of older trees (Maxwell et al., 2016). In 2020, additional cores were collected following an ecological study design, wherein nest subplots were used within a large plot to increase sample depth across age classes of the forest. The first subplot was 5.0 m in radius and all trees larger than 5 cm diameter at breast height (DBH) were cored. In the next subplot, 5-20 m from plot center, all trees >15 cm DBH were cored. In the final subplot, 20-30 m from plot center, all trees >40 cm DBH were cored.

Harvard Forest, USA

Harvard Forest (HF) is a secondary temperate mixed forest dominated by *Tsuga canadensis* (L.) Carrière and northern hardwood species including *Quercus rubra* L., *Acer rubrum* L., and *Betula alleghaniensis* Britton. Cores were collected at two plots within the ForestGEO megaplot, and in the footprint of the original HF eddy-flux tower (Finzi et al., 2020). Samples were collected following an ecological study design of nested circular plots (Dye et al., 2016), where sampling included all living or dead trees ≥ 10 cm DBH within a 13-m radius and ≥ 20 cm within a 20-m radius.

Žofín Forest Dynamics Plot, Czech Republic

The Žofín forest (ZOF) is a well-preserved natural forest remnant dominated by *Fagus sylvatica* L., with some *Picea abies* L. and rare occurrence of *Abies alba* Mill. The forest is not fully without past human direct interventions, and a limited historical land use effect is also possible (see Kozáková et al., 2011).

In total, 2210 individuals were cored either within the 25 ha ForestGEO plot or a larger, 74 ha plot in which it is embedded. The tree-ring data were collected for several studies around 2010 (Kašpar et al., 2021; Šamonil et al., 2016, 2014; Šamonil et al., 2015, 2013; Vašíčková et al., 2019, 2016). The sampling strategy was mainly connected with the disturbance history of the forest and evaluation of growth of juvenile trees in gaps. Therefore, these data consist mainly of dominant and codominant tree species and relatively low number of young trees.

Niobrara, Nebraska, USA

The Niobrara site sits at the prairie-forest ecotone of the North American Great Plains. There, forests occur in cool, moist canyons formed by spring-fed tributaries along the Niobrara River. *Betula papyrifera* Marshall occur in these canyons, in spots where there are groundwater springs that flow all year long. Niobrara is near the southern range limit of this species, and populations at this site are small and highly disjunct. Cores were taken from seven north-facing *Betula papyrifera* stands along a 27 km section of the Niobrara River, as detailed in Bumann et al. (2019). The largest healthy trees within each stand were selected for coring.

Little Tesuque, New Mexico, USA

The Little Tesuque (LT) site is in a mature dry mixed conifer forest outside of Santa Fe, NM, in the foothills of the Sangre de Cristo Mountains. The site had not undergone stand-clearing disturbance (e.g., fire) in more than one hundred years. Tree-ring records suggest that there has been no stand-clearing fire for at least 500 years. Low-severity fires burned the site regularly since at least 1542, with the last one in 1880. The growth of trees in the region, including at this site, are being severely affected by a strong drying trend under climate change (Touchan et al., 2011; Williams et al., 2013), with many missing growth rings in recent years.

In 2020, live canopy and sub-canopy trees, all >20 cm DBH, were sampled within a 1 ha plot (Little Tesuque plot 3; *lts3*).

Cedar Breaks, Utah, USA

Cores were collected within the Utah Forest Dynamics Plot ForestGEO plot on the Colorado Plateau in Cedar Breaks National Monument (CB, Furniss et al., 2017). The plot sits on the edge of cliff, which descends ~750 m to a valley below. The mature spruce-fir forest is dominated by the species analyzed here (*Abies balsamea* A. Murray, *Populus tremuloides* Michx., *Picea engelmannii* Engelm., *Pinus flexilis* E. James, *Pinus longaeva* D.K. Bailey, *Picea pungens* Engelm., and *Pseudotsuga menziesii* (Mirb.) Franco). The site is largely undisturbed. The most recent fire occurred in 1802, and many currently living trees established before then. Living *Pinus longaeva* at the site date back ~1600 years, and deadwood at the site dates back to 0 AD (Lutz et al., 2021).

In 2019, living individuals of each dominant species were cored.

Chronologies are published on ITRDB (Birch et al., 2020d, 2020a, 2020b, 2020c).

Scotty Creek, Northwest Territories, Canada

The Scotty Creek site is a black spruce (*Picea mariana* (Mill.) Britton, Sterns & Poggenb.) forest on rapidly thawing discontinuous permafrost (Baltzer et al., 2014; Dearborn et al., 2020; Sniderhan & Baltzer, 2016). Vertical permafrost thaw has been causing drought stress in the shallow-rooted trees, resulting in growth declines (Sniderhan & Baltzer, 2016) and high mortality (Dearborn et al., 2020).

A large recruitment pulse in the 1930s suggests that the site burned severely at that time, as would be consistent with a fire return interval of \sim 77 years in the Taiga Plains ecoregion (Johnstone et al., 2010; Walker et al., 2018).

In the summers of 2012 and 2013, seven forest stands within a 2 km^2 area were sampled. In five stands with clearly defined boundaries at edges of permafrost plateaus, all living and dead trees $\geq 7 \text{ cm DBH}$ were cored. In two stands without well-defined edges, cores were taken from a $\geq 7 \text{ cm DBH}$ tree nearest to every 20 m interval in a $100 \times 100 \text{ m}$ grid, totaling 36 trees per stand. Sampling details are given in Sniderhan & Baltzer (2016).

Appendix S2. Methods for reconstruction of diameter breast height (DBH)

Tree diameter at breast height (or height of coring), DBH, was reconstructed over the time span for which a core record existed. DBH was reconstructed either from the outside in or from the inside out, depending on the availability of recent DBH measurements (see next paragraph). We gave precedence to the outside-in approach; that is, when a recent DBH measurement was available, DBH was reconstructed from the outside in. In cases where DBH was not available, but when we knew that the core hit pith or could reasonably estimate how far off it was based on the curvature of the rings (Applequist, 1958; Duncan, 1989), DBH was reconstructed from the inside out.

DBH measurements were taken at the time of coring or – for some of the trees cored within ForestGEO plots – obtained from the most recent ForestGEO census prior to coring. For these trees, we applied the outside-in approach to DBH reconstruction (detailed below). For a minority of trees ($n=50$ at LDW), DBH measurements were not available, requiring use of inside-out approach to DBH reconstruction (detailed below).

The outside-in approach to DBH reconstruction started with a recent DBH measurement, DBH_m , where reference measurement year m is often but not necessarily at the time of coring. DBH for each year (y) of interest, Y , was then estimated using the following equation:

$$DBH_Y = DBH_m - 2 * \left[r_{bark,m} - r_{bark,Y} + \sum_{y=Y}^m RW_y \right]$$

Here, r_{bark} denotes bark thickness, which was estimated on a species-specific basis from allometries (Tables S2, S4). If DBH_m was taken before coring, RW values between $y = m$ and $y = Y$ were added instead of subtracted. If DBH_m was taken ≤ 10 years after the last year of measurement in the core, we used the equation above, assuming that the n missing measurements between the year of the last RW record in the core and year m were equal to the last n records in the core.

In some cases, negative values of DBH_Y were predicted for the earliest years in the core record; that is, the sum of RW from the earliest year in the core record to year m , plus the predicted change in r_{bark} , was greater than half of DBH_m . Such cases could occur due to a measurement or data error, or if the tree's growth was asymmetrical around the pith, as commonly occurs (Speer, 2010). In cases where the error was egregious – i.e., where predicted DBH_Y of the earliest year in the record divided by DBH_m was < -0.5 – we removed the tree from the analysis. We assumed that smaller errors were attributable to asymmetrical growth, and re-distributed the error evenly across years so that DBH_Y prior to the first year in the core was zero. This correction was applied only to DBH_Y , and not to any RW records.

When a measurement of DBH_m was not available, and when the core hit pith or the distance to pith could be reliably estimated, DBH was reconstructed from the inside-out. Specifically, DBH for each year (y) of interest, Y , was estimated using the following equation:

$$DBH_Y = 2 * \left[r_{bark,Y} + \sum_{y=0}^Y RW_y \right]$$

Again, r_{bark} was estimated on a species-specific basis from allometries (Tables S2, S4).

When there was more than one core for a tree, the DBH reconstructions from each core were averaged to produce a single estimate of the tree's DBH through time. When the start or end dates of the records from the cores differed, we extrapolated growth of the shorter core to match the years covered by the longer core. Specifically, to fill in years at the more recent end, we assumed that the average growth rate of the ten years prior to the missing records applied to the missing years. To fill in years at the beginning of the tree's lifespan, we likewise assumed that the ten years adjacent to the missing record applied to the missing years; however, if this yielded a negative DBH estimate for the earliest year in the reconstruction, we divided the existing minimum DBH by number of years missing and applied that value to each year. We note again that

these reconstructed growth records were used only for the reconstruction of DBH and were not included as response variables in any of our analyses.

For trees on which a measurement of DBH_m was not available and the distance to pith could be reliably estimated for any cores, DBH could not be reconstructed. These trees were excluded from all analyses including DBH as an independent variable.

Regardless of the DBH reconstruction approach (outside-in or inside-out), accurate reconstructions require accounting for r_{bark} , particularly for species that can attain very thick bark (e.g., *Pseudotsuga menziesii*, *Pinus ponderosa* P. Lawson & C. Lawson). For the majority of species, including all thick-barked species, we assigned allometries relating r_{bark} to DBH, which were constructed for the species of interest or a related species (Tables S2, S4). When raw data on bark thickness data were available, we generated species-specific lognormal scaling relationships with the intercept forced to zero (Table S4). When raw bark thickness data were not available, we applied published bark allometries from other sources (Table S4).

Appendix S3. Methods for climate data evaluation and correction

Monthly climate data for 1901-2019 were obtained from CRU v.4.04 (Harris et al., 2014, 2020), and in a few cases corrected based on more local records. For BCNM, we calculated monthly PPT and PDF from daily precipitation readings made on Barro Colorado Island starting in 1929 (Paton, 2019) and used these to replace the corresponding CRU records.

Additionally, we used 800 m resolution PRISM records (Daly et al., 2015, 2008) from 1930 to 2015 to test and correct for cases where the coarser-resolution CRU values were not locally representative. Variables in the PRISM data set included T_{max} , T_{mean} , T_{min} , and PPT. To test for substantial differences between CRU and PRISM records, each variable was compared on a monthly basis using linear regression. We considered CRU records to be inappropriate if (1) a paired t-test revealed a significant difference between CRU and PRISM records, and (2) the absolute value of the mean monthly difference between CRU and PRISM exceeded 2.5°C (temperature variables) or 10 mm per month (precipitation). Two sites met these criteria: LT and CB, both located in mountainous terrain in the western United States. For these sites, we used linear regression to characterize the relationship between the CRU and PRISM, and then used this relationship to correct CRU data outside the time frame of PRISM (i.e., pre-1930 and post-2015). CRU records were replaced with PRISM records over the time frame for which they were available. For these sites, CRU records for variables not included in the PRISM data set (PDF, PET) were considered unreliable and excluded from the analysis.

Appendix S4. Methods for comparing our approach with traditional methods

To test whether our methods gave similar results to traditional methods, we conducted qualitative comparisons of our results to previous studies based on the same cores (Table S5) and conducted a formal quantitative comparison for four species (Figs. S1-S4), as detailed below.

Qualitative comparison

For all species-site combinations, we reviewed previous studies characterizing the climate sensitivity of growth using conventional methods. In most cases, we were able to compare with previous studies from the same sites and sets of cores. When these were not available, we reviewed regional-level analyses believed to be representative of the site.

Results from previous studies were compiled alongside results from the climate-only model in this study (Table S5). Where previous studies examined numerous climate variables or seasonal windows (e.g., Helcoski et al., 2019), we focus on those most relevant to our findings.

Beyond the methodological differences, original studies based on the same sets of cores varied from this one and from one another in factors including the exact set of cores analyzed, climate data sources, time frame of analysis, approaches to identifying candidate climate variables and windows (including whether this is done on a site or species level), methods for detrending and standardizing to build chronologies, and whether the effects of temperature and precipitation are considered separately (original studies) or additively (this study). To standardize for such differences, we selected a subset of species for a standardized quantitative comparison, as detailed below.

Quantitative comparison

We also conducted a formal comparison of our approach to conventional methods using identical tree-ring and climate data for four species: *Liriodendron tulipifera* L. at SCBI, *Abies alba* at Zof, *Pseudotsuga menziesii* at CB, and *Picea mariana* at SC (Figs. S1-S4). These species were selected for analysis because they have been well-studied in the past. For each species, we compared climate sensitivities for the top precipitation- and temperature- group variables, as identified in the main analysis.

Prior to analysis, data were prepared and cleaned as described in the Methods section, resulting in an identical set of records for input into each analysis. For the approach developed here, analysis was conducted as described in the Methods section, but with the *climwin* climate variable selection process limited to just the species of interest (as opposed to all species at the site), climate variables considered individually rather than additively, analysis of only first-order linear relationships, and with start date adjusted to match the conventional method (see below). The R package *Climwin* is a useful analytical tool that automates the process of running and comparing numerous regressions, but its use does not alter results relative to what would be obtained via a more conventional approach; numerous climate drivers and seasonal windows can be compared without *climwin* (e.g., Meko et al., 2011), and *climwin* can also be used on residual or standard chronologies. Following the *climwin* analysis step, we extracted *beta* coefficients describing the slope of the relationship between climate and RW. *Beta* coefficients, along with their standard error, were obtained for each month within the analysis time frame (Table S1) and for the seasonal window identified as most influential by *climwin*.

For the analysis using conventional methods, the ring-width series from each core was standardized via ARSTAN using a 2/3rds n spline, where n is the number of years in the series (Cook, 1985; Cook & Kairiukstis, 1990). We used the adaptive power transformation (Cook & Peters, 1997) to reduce the influence of outliers and stabilize the variance over time. Next, to adjust for changes in variance as series replication decreased towards the earlier portion of each chronology, we stabilized each series using either the average correlation between raw ring-width series (rbar) method or a 1/3rds spline method (Jones et al., 1997). We applied the 1/3rds spline method when replication in the inner portion of each chronology (~30–50 yrs, depending on full chronology length) dropped below three trees. Next, we calculated a robust biweight mean chronology for each species from the ring-width indices (Cook, 1985). We used residual chronologies because the autoregressive standardization process in creating them removes much of the tree-level autocorrelation in growth, such that these chronologies would be expected to contain the most conservative information on interannual growth

drivers (Cook, 1985).

We defined chronology start dates according to the subsample signal strength (SSS), using a cutoff of SSS = 0.80 (or 80% of the population signal). Thus, for this analysis only, we defined chronology start dates as the year the SSS exceeded 0.80 or two years after the start of the climate record, whichever came later. SSS exceeded 0.80 well before the start of the 1901 start of climate records for *Pseudotsuga menziesii* (1800s), *Abies alba* (1700), and *Picea mariana* (1850s). For *Liriodendron tulipifera*, SSS reached 0.8 with 11 trees in 1919, which we used as the start date for this series. We note that these start date criteria differ from those used in the main analysis (Table S3), which had earlier start dates because the analysis was not constrained by a need to represent the full population signal. End dates were defined as the last full year prior to sampling (Table S3), or 2007 for CB.

Beta (slope) coefficients for the relationship between tree growth and the monthly climate variable were derived as in Helcoski et al. (2019). Briefly, we used the R packages *dplR* (Bunn, 2008) and *bootRes* (Zang & Biondi, 2013) to analyze relationships between RW and monthly climate following the bootstrapping methodology of (Biondi & Waikul, 2004). Pearson correlations between climate variables and tree-ring chronologies were converted to linear slopes using the method of Charney et al. (2016).

Finally, we generated plots comparing month-by-month *beta* coefficients describing climate sensitivity, and also comparing *beta* coefficients for the window identified as most influential by *climwin* Figs. S1-S4).

We note that despite designing the analyses to be as comparable as possible, one-to-one correspondence of *beta* coefficients is not necessarily expected for several reasons. First, although the analysis time frame is standardized between the two approaches, the relative influence of each year will generally vary between the two approaches. The traditional approach, which all cores into a single residual chronology with one value per year, gives equal weighting to each year. In contrast, under the approach developed here, the number of records per year can vary across the analysis time frame, generally increasing over time as the younger trees enter the analysis. Thus, particularly when many younger trees are included in the analysis, the two approaches will effectively apply different weighting schemes to the years of the analysis period. In cases where climate-sensitivity differs between old and young trees, or where the climate and/or climate response changed substantially over the analysis time frame [e.g., at Scotty Creek; Fig. S4; Sniderhan & Baltzer (2016)], this may lead to divergence of the climate sensitivities estimated by the two methods.

Second, traditional analysis methods (using ARSTAN) were primarily designed to distill population-level variation to obtain the strongest possible climate signal for the reconstruction of past climate (Cook & Kairiukstis, 1990), not to characterize climate responses on the individual level, where variation is inherently higher. While conversion of Pearson correlations to linear slopes *sensu* Charney et al. (2016) approximates climate responses, it does not provide an exact slope describing the relationship between individual-level or population mean growth and climate. This is because standardization of variance and averaging of individual-level residuals prior to the climate analysis fundamentally alters and obfuscates individual-level responses (Carrer, 2011; Pederson et al., 2020).

We suspect that both of these factors may underlie the tendency for the traditional method to estimate stronger climate sensitivity than the approach developed here for Scotty Creek (Fig. S4), a comprehensively sampled black spruce forest (i.e., including young trees) on thawing permafrost. We note, however, that there were no statistically significant differences in the *beta* coefficients of the two approaches at this site.

Appendix S5. Dealing with rapidly changing climate and tree growth

Our analysis included two sites where climate change has had pronounced effects on tree growth: Little Tesuque, New Mexico, USA (LT) and Scotty Creek, NW Territories, Canada (SC). At LT, increasingly warm drought has dramatically reduced growth (Touchan et al., 2011; Williams et al., 2013), resulting in many missing rings in recent years. At SC, rapidly rising temperatures are causing permafrost thaw, summer moisture stress, resulting in negative growth trends in basal area index (BAI) starting around 1950 and significant growth declines since 1970 in 56% of trees (Sniderhan & Baltzer, 2016).

Problemsatically, correlating tree growth residuals from which climate-driven trends had been removed against a climate signal with a strong directional trend would not necessarily identify the most relevant climate drivers (step 1 in Fig. 1). To explore whether selection of climate driver variables was strongly affected by directional trends in climate, we experimented with three different approaches to identifying the most important climate drivers at three sites: LT, SC, and SCBI (where the climate has not changed strongly). The first approach was the baseline method described in the Methods and presented in throughout the manuscript, where detrended growth residuals were correlated with raw climate data (step 1 in Fig. 1).

The second approach involved detrending the climate variables using a GAM with automated selection of the number of knots prior to the *climwin* step. Thus, this approach correlates growth anomalies to climate anomalies around a more slowly changing mean, and using it to select climate driver variables inherently assumes that the most important climate drivers of high-frequency (annual) growth variation are also the most important drivers of longer-term growth changes in response to changing climate. For PPT at LT, this approach and our baseline method identified identical seasonal windows for all growth metrics. For the temperature variable at LT, this approach and our baseline method consistently identified T_{max} as the most important variable, but selected slightly different seasonal windows with RW the response metric (May-July for baseline method, April-July when T_{max} was detrended). For the water variable at SC, the two methods identified identical climate drivers (previous May or June - previous August PPT, depending on growth metric but not the analysis method). For the temperature variable at SC, the methods identified different variables and windows, with the baseline method selecting July T_{max} and the detrending method selecting previous December T_{mean} (which makes little sense biologically). For the water variable at SCBI, this approach and our baseline method consistently identified previous June- current August PPT as the strongest climate driver, with the exception that current May-August came out as the strongest driver in the detrended analysis for ΔAGB . For the temperature variable at SCBI, results were somewhat different, with this approach identifying T_{max} as the most important variable, and the baseline approach identifying PET. Both identified seasonal windows within the current growing season when RW was the metric, but seasonal windows were not similar when BAI or ΔAGB was the metric. Given little meaningful difference between these approaches, and no clear advantage of detrending the climate variable, we preferred the baseline method.

The third approach involved splitting analyses into decades before and after 1970 (*sensu* Sniderhan & Baltzer, 2016) – an approach that has also been applied in other studies in rapidly warming climates (Driscoll et al., 2005; e.g., Lloyd & Fastie, 2002; Wilmking et al., 2004). This approach should accurately capture the most important climatic drivers of growth variation prior to the acceleration of climate change in recent decades, and should also identify the drivers of recent high-frequency growth variation. For PPT at LT, the most influential seasonal windows pre-1970 and post-1970 were similar to those for the entire time period. For the temperature variable at LT, T_{max} was consistently identified as the most important variable, but its most influential seasonal window extended back to the previous growing season pre-1970, but was limited to the current growing season post-1970 and for the analysis as a whole. For the water variable at SC, PPT was consistently selected as a better predictor than PDF, and responses pre- and post-1970 were generally similar to those over the full time period. For the temperature variable at SC, the pre- and post- 1970 time periods both identified T_{min} temperatures during a single month in the previous summer as the most influential climate driver for RW, whereas the analysis for the full time period identified T_{max} of current July (Fig. S14). For the water variable at SCBI, PPT was consistently selected as a better predictor than PDF, with similar seasonal windows pre-1970 and for the full analysis period, compared to only a weak effect over a different time period (current May) post-1970 (Fig. S14). For the temperature variable at SCBI, selected variables (PET, T_{max} , or T_{mean}) and seasonal windows were variable across the three analysis periods, with only weak relationships in the post-1970 analysis period. These results indicate potentially shifting climate

sensitivities, particularly at SC (Fig. S14), which may not be adequately captured by our baseline method. Further analysis into how climate sensitivities may be shifting would be valuable, but is beyond the scope of this analysis.

While we determined that our baseline method was suitable for the current analysis, we note that it remains problematic in that detrending of growth sequences ahead of identification of climate drivers precludes the possibility of the analysis capturing the climatic drivers of low-frequency growth changes. The ideal solution would likely be to avoid any type of detrending of growth sequences, but rather to use full GLS models (as in step 2 in Fig. 1) to assess the explanatory power of potential climate variables, but this would be computationally time-consuming and is beyond the scope of the current analysis.

Table S1. Additional information on sites included in this analysis.

site code	site name	latitude*	longitude*	elevation (m.a.s.l.)	cores within ForestGEO plot?	canopy positions	tree statuses	date range	dormant season**	months in climwin
BCNM	Barro Colorado Nature Monument	9.15430	-79.8461	<160	no	canopy	live, dead	1931-2014	Nov-Apr	pOct-cDec
HKK	Huai Kha Khaeng	15.63240	99.2170	550	no	all	live	1903-2011	Nov-Apr	pOct-cDec
SCBI	Smithsonian Conservation Biology Institute	38.89350	-78.1454	273-338	yes	all	live, dead	1903-2017	Oct-Mar	pMay-cAug
LDW	Lilly Dickey Woods	39.23590	-86.2181	230-303*	no	canopy	live, dead	1903-2019	Oct-Mar	pMay-cAug
HF	Harvard Forest	42.53880	-72.1755	340-368*	yes	all	live, dead	1903-2014	Oct-Mar	pMay-cAug
ZOF	Žofín Forest Dynamics Plot	48.66380	14.7073	736-829*	some	all	live, dead	1903-2013	Oct-Mar	pMay-cAug
NIO	Niobrara	42.78000	-100.0210	644-702*	no	canopy	live	1948-2015	Oct-Apr	pMay-cAug
LT	Little Tesuque	35.73838	-105.8382	2684 - 2702	n.a.	canopy / sub-canopy	live	1903-2018	Oct-Apr	pMay-cAug
CB	Utah Forest Dynamics Plot	37.66150	-112.8525	3020-3169	yes		live	1903-2007	Oct-Apr	pMay-cAug
SC	Scotty Creek	61.30000	-121.3000	280	no	all	live, dead	1903-2013	Sept-Apr	pMay-cAug

*Refers to ForestGEO plot (exception: LT) and is not necessarily the most accurate value for exact locations at which cores were taken. Geographic coordinates were used to extract climate data from CRU.

**Refers to approximate period during which woody growth slows or ceases (dry season in the tropics, winter for temperate and boreal sites), resulting in ring formation.

m.a.s.l. = meters above sea level

Table S2. Tree species analyzed, their characteristics, and bark allometries applied.

species code	family	latin name	sites sampled	leaf type	leaf phenology	light requirements*	bark allometry**
ABAL	Pinaceae	<i>Abies alba</i> Mill.	ZOF	conifer	evergreen	shade-tolerant	1
ABBI	Pinaceae	<i>Abies bifolia</i> A. Murray	CB	conifer	evergreen	shade-tolerant	1
ACRU	Sapindaceae	<i>Acer rubrum</i> L.	HF	broadleaf	deciduous (cold)	intermediate	2
ACSA	Sapindaceae	<i>Acer saccharum</i> Marsh.	LDW	broadleaf	deciduous (cold)	shade-tolerant	2
AFXY	Fabaceae	<i>Afzelia xylocarpa</i> (Kurz) Craib	HKK	broadleaf	deciduous (drought)	light-demanding	neglected
BEAL	Betulaceae	<i>Betula alleghaniensis</i> Britton	HF	broadleaf	deciduous (cold)	intermediate	3
BEPA	Betulaceae	<i>Betula papyrifera</i> Marshall	NIO	broadleaf	deciduous (cold)	light-demanding	4
CACO	Juglandaceae	<i>Carya cordiformis</i> (Wangenh.) K. Koch	SCBI	broadleaf	deciduous (cold)	light-demanding	5
CAGL	Juglandaceae	<i>Carya glabra</i> (Mill.) Sweet	SCBI	broadleaf	deciduous (cold)	intermediate	6
CAOVL	Juglandaceae	<i>Carya ovalis</i> (Wangenh.) Sarg.	SCBI	broadleaf	deciduous (cold)	intermediate	7
CAOV	Juglandaceae	<i>Carya ovata</i> (Mill.) K. Koch	LDW	broadleaf	deciduous (cold)	intermediate	6
CATO	Juglandaceae	<i>Carya tomentosa</i> (Poir.) Nutt.	SCBI	broadleaf	deciduous (cold)	light-demanding	8
CHTA	Meliaceae	<i>Chukrasia tabularis</i> A. Juss.	HKK	broadleaf	brevi-deciduous (drought)	intermediate	neglected
FAGR	Fagaceae	<i>Fagus grandifolia</i> Ehrh.	HF, SCBI	broadleaf	deciduous (cold)	shade-tolerant	neglected
FASY	Fagaceae	<i>Fagus sylvatica</i> L.	ZOF	broadleaf	deciduous (cold)	shade-tolerant	neglected
FRAM	Oleaceae	<i>Fraxinus americana</i> L.	LDW, SCBI	broadleaf	deciduous (cold)	intermediate	9
FRNI	Oleaceae	<i>Fraxinus nigra</i> Marshall	SCBI	broadleaf	deciduous (cold)	intermediate	9
JACO	Bignoniaceae	<i>Jacaranda copaia</i> (Aubl.) D. Don	BCNM	broadleaf	deciduous (drought)	light-demanding	10
JUNI	Juglandaceae	<i>Juglans nigra</i> L.	SCBI	broadleaf	deciduous (cold)	light-demanding	11
LITU	Magnoliaceae	<i>Liriodendron tulipifera</i> L.	LDW, SCBI	broadleaf	deciduous (cold)	light-demanding	12
MEAZ	Meliaceae	<i>Melia azedarach</i> L.	HKK	broadleaf	deciduous (drought)	light-demanding	neglected
PIAB	Pinaceae	<i>Picea abies</i> (L.) H. Karst.	HF, ZOF	conifer	evergreen	shade-tolerant	13
PIEN	Pinaceae	<i>Picea engelmannii</i> Engelm.	CB	conifer	evergreen	shade-tolerant	13
PIMA	Pinaceae	<i>Picea mariana</i> (Mill.) Britton, Sterns & Poggenb.	SC	conifer	evergreen	shade-tolerant	14
PIPU	Pinaceae	<i>Picea pungens</i> Engelm.	CB	conifer	evergreen	intermediate	13
PIFL	Pinaceae	<i>Pinus flexilis</i> E. James	CB	conifer	evergreen	light-demanding	16
PILO	Pinaceae	<i>Pinus longaeva</i> D.K. Bailey	CB	conifer	evergreen	light-demanding	neglected
PIPO	Pinaceae	<i>Pinus ponderosa</i> P. Lawson & C. Lawson	LT	conifer	evergreen	light-demanding	15
PIST3	Pinaceae	<i>Pinus strobus</i> Engelm.	LT	conifer	evergreen	light-demanding	16
PIST	Pinaceae	<i>Pinus strobus</i> L.	HF, SCBI	conifer	evergreen	intermediate	17
POTR	Salicaceae	<i>Populus tremuloides</i> Michx.	CB	broadleaf	deciduous (cold)	light-demanding	18
PSME	Pinaceae	<i>Pseudotsuga menziesii</i> (Mirb.) Franco	CB	conifer	evergreen	intermediate	19
QUAL	Fagaceae	<i>Quercus alba</i> L.	LDW, SCBI	broadleaf	deciduous (cold)	intermediate	20
QUMO	Fagaceae	<i>Quercus montana</i> Willd.	LDW, SCBI	broadleaf	deciduous (cold)	intermediate	21
QURU	Fagaceae	<i>Quercus rubra</i> L.	HF, LDW, SCBI	broadleaf	deciduous (cold)	intermediate	22
QUVE	Fagaceae	<i>Quercus velutina</i> Lam.	LDW, SCBI	broadleaf	deciduous (cold)	intermediate	23
TEPA	Burseraceae	<i>Tetragastris panamensis</i> (Engl.) Kuntze	BCNM	broadleaf	evergreen	shade-tolerant	24
TOCI	Meliaceae	<i>Toona ciliata</i> M. Roem.	HKK	broadleaf	deciduous (drought)	intermediate	neglected
TRTU	Meliaceae	<i>Trichilia tuberculata</i> (Triana & Planch.) C. DC.	BCNM	broadleaf	evergreen	shade-tolerant	25
TSCA	Pinaceae	<i>Tsuga canadensis</i> (L.) Carrière	HF	conifer	evergreen	shade-tolerant	26

*For extratropical species, light requirements are classified based on the database of Niinemets & Valladares (2006). For tropical species, categorization is based on Alfaro-Sánchez et al. (2017) for BCNM and Vlam et al. (2014) for HKK.

**Bark allometry field indicates the species and site sampled to construct the bark allometry. When neither raw data nor an allometric equation for the study species was available, we selected the most appropriate equation that could be located for similar species. Equations are given in Table S4.

Table S3. Sampling details for tree species by site.

site	species code	all		with DBH		DBH range		date range	year effect analyzed
		n trees	n cores	n trees	n cores	sampled	reconstructed*		
BCNM	JACO	12	18	11	17	30.2-63.5	2.6-56.4	1931-2014	FALSE
BCNM	TEPA	18	29	17	26	22.1-59.5	2.7-49.4	1931-2014	TRUE
BCNM	TRTU	23	37	20	31	20.7-43.6	4.8-41.5	1931-2014	TRUE
CB	ABBI	22	41	20	39	13.9-54.2	0-50.4	1903-2019	TRUE
CB	PIEN	14	23	12	20	14-54.9	2.2-43.4	1903-2019	TRUE
CB	PIFL	13	21	12	20	17.6-64.1	4.5-58.5	1903-2018	TRUE
CB	PILO	17	25	7	11	45.8-63.6	35.4-57.5	1903-2019	TRUE
CB	PIPU	16	29	15	28	22.4-50.8	8.6-50.5	1903-2019	TRUE
CB	POTR	17	27	17	27	23.6-47.6	7.7-44.5	1903-2019	TRUE
CB	PSME	11	21	11	21	10.6-64.2	2.6-63.3	1903-2019	TRUE
HF	ACRU	18	59	18	59	10.1-22.1	0.9-20.4	1903-2013	TRUE
HF	BEAL	13	44	13	44	10.2-37.9	1.6-20.5	1904-2013	TRUE
HF	QURU	74	180	73	177	19.5-53	1.1-48.3	1903-2014	TRUE
HF	TSCA	32	83	32	83	10.6-37	0.6-33.5	1923-2014	TRUE
HKK	AFXY	39	127	39	127	20.1-98.7	0.1-81.4	1903-2011	TRUE
HKK	CHTA	28	70	28	70	16-64.6	0.2-59.5	1904-2010	TRUE
HKK	MEAZ	46	130	46	130	25.6-98.1	3.8-80.3	1914-2011	FALSE
HKK	TOCI	45	143	45	143	16.6-116.4	1.7-80.5	1903-2011	TRUE
LDW	ACSA	35	66	34	64	9-64.6	0-52.4	1903-2019	FALSE
LDW	CAOV	9	18	8	16	unknown	0.6-37.4	1903-2013	FALSE
LDW	LITU	15	28	14	26	unknown	1.2-69.4	1903-2019	TRUE
LDW	QUAL	10	20	0	0	NA	NA	1903-2013	NA
LDW	QUMO	10	20	8	16	unknown	1.1-52.4	1903-2013	FALSE
LDW	QUVE	9	18	0	0	NA	NA	1903-2013	NA
LT	PIPO	10	20	10	20	23.2-52.8	14.6-48.4	1903-2018	TRUE
LT	PIST3	7	14	7	14	25.7-39.8	4.2-34.4	1903-2018	TRUE
NIO	BEPA	42	138	42	138	unknown	0.4-33.5	1948-2015	TRUE
SCBI	CACO	15	15	15	15	10.62-38.52	1.6-32.2	1903-2015	TRUE
SCBI	CAGL	39	39	36	36	10.28-52.31	1.6-49.3	1903-2015	TRUE
SCBI	CAOVL	25	25	24	24	15.11-60.32	2.6-47.2	1903-2015	TRUE
SCBI	CATO	15	15	14	14	12.86-35.95	3.7-28.4	1903-2015	TRUE
SCBI	FAGR	76	76	76	76	10.05-41.02	0.1-41.2	1920-2009	TRUE
SCBI	FRAM	66	66	63	63	6.85-94.73	0.1-84.4	1903-2016	TRUE

(continued)

site	species code	all		with DBH		DBH range		date range	year effect analyzed
		n trees	n cores	n trees	n cores	sampled	reconstructed*		
SCBI	FRNI	12	12	12	12	11.04-39.2	0.5-27.3	1903-1996	TRUE
SCBI	JUNI	30	30	29	29	20.4-76.19	4.6-59.5	1903-2010	TRUE
SCBI	LITU	106	106	105	105	10-91.42	0.1-81.1	1903-2010	TRUE
SCBI	PIST	36	36	36	36	13.92-50.96	0.5-44.3	1931-2010	FALSE
SCBI	QUAL	66	66	66	66	11.4-76.73	0.3-70.4	1903-2009	TRUE
SCBI	QUMO	67	67	67	67	10.22-84.59	0.3-69.5	1903-2017	TRUE
SCBI	QURU	70	70	70	70	11.07-87.65	2.5-79.2	1903-2016	TRUE
SCBI	QUVE	81	81	81	81	16.02-82.33	0.5-78.4	1903-2009	TRUE
SC	PIMA	443	443	395	395	7-24	0-16.4	1903-2013	TRUE
ZOF	ABAL	46	46	46	46	50-121	21.1-107.4	1903-2010	TRUE
ZOF	FASY	1369	1369	1369	1369	unknown	0.1-115.3	1903-2013	TRUE
ZOF	PIAB	644	644	642	642	unknown	0-125.4	1903-2011	TRUE

*Maximum reconstructed DBH's analyzed are less than maximum sampled DBH's because we discard size ranges with < 3 conspecific trees.

Table S4. Allometric equations for tree bark thickness.

ID	species	equation	source	n	DBH.range.cm	site	source,1
1	<i>Abies lasiocarpa</i>	$bark.mm = ((0.05 + 0.06 * (dbh.cm/2.54))/2) * 2.54$	equation	NA		North America	Miles, Patrick D.; Smith, W. Brad. 2009.
2	<i>Acer rubrum</i>	$bark.mm = 0.619 * log(dbh.cm + 1)$	data	10	8.2-39.6	SCBI	Anderson-Teixeira et al. (2015)
3	<i>Betula alleghaniensis</i>	$bark.mm = ((0.15 + 0.03 * (dbh.cm/2.54))/2) * 2.54$	equation	NA		North America	Miles, Patrick D.; Smith, W. Brad. 2009.
4	<i>Betula papyrifera</i>	$bark.mm = ((0.13 + 0.05 * (dbh.cm/2.54))/2) * 2.54$	equation	NA		North America	Miles, Patrick D.; Smith, W. Brad. 2009.
5	<i>Carya cordiformis</i>	$bark.mm = 0.793 * log(dbh.cm + 1)$	data	9	5.9-68.2	SCBI	Anderson-Teixeira et al. (2015)
6	<i>Carya glabra</i>	$bark.mm = 1.035 * log(dbh.cm + 1)$	data	8	19.1-78	SCBI	Anderson-Teixeira et al. (2015)
7	<i>Carya ovalis</i>	$bark.mm = 1.531 * log(dbh.cm + 1)$	data	8	6.4-63.1	SCBI	Anderson-Teixeira et al. (2015)
8	<i>Carya tomentosa</i>	$bark.mm = 1.105 * log(dbh.cm + 1)$	data	8	5-57.3	SCBI	Anderson-Teixeira et al. (2015)
9	<i>Fraxinus americana</i>	$bark.mm = 2.223 * log(dbh.cm + 1)$	data	9	6.1-94.2	SCBI	Anderson-Teixeira et al. (2015)
10	<i>Jacaranda copia</i>	$bark.mm = 2.993 * log(dbh.cm + 1)$	data	5	45.6-75	Panama	Raquel Alfaro-Sánchez (unpublished data)
11	<i>Juglans nigra</i>	$bark.mm = 2.107 * log(dbh.cm + 1)$	data	9	13.6-85.4	SCBI	Anderson-Teixeira et al. (2015)
12	<i>Liriodendron tulipifera</i>	$bark.mm = 1.637 * log(dbh.cm + 1)$	data	9	27.5-136.5	SCBI	Anderson-Teixeira et al. (2015)
13	<i>Picea engelmannii</i>	$bark.mm = ((0.15 + 0.04 * (dbh.cm/2.54))/2) * 2.54$	equation	NA		North America	Miles, Patrick D.; Smith, W. Brad. 2009.
14	<i>Picea mariana</i>	$bark.mm = 3.726 * log(dbh.cm + 1)$	data	12	6.9-7.9	Scoty Creek	Rajit Patankar and Jennifer Baltzer (unpublished data)
15	<i>Pinus jeffreyi</i>	$bark.mm = (1.298 * \sqrt{dbh.cm})^{0.802}^2$	equation	81	5-160	California (4 montane sites)	Zelhig-Kichas et al. (2016)
16	<i>Pinus monticola</i>	$bark.mm = (1.299 * \sqrt{dbh.cm})^{0.809}^2$	equation	29	10-130	California (3 montane sites)	Zelhig-Kichas et al. (2016)
17	<i>Pinus strobus</i>	$bark.mm = ((0.02 + 0.10 * (dbh.cm/2.54))/2) * 2.54$	equation	NA		North America	Miles, Patrick D.; Smith, W. Brad. 2009.
18	<i>Populus tremuloides</i>	$bark.mm = ((0.10 + 0.07 * (dbh.cm/2.54))/2) * 2.54$	equation	NA		North America	Miles, Patrick D.; Smith, W. Brad. 2009.
19	<i>Pseudotsuga menziesii</i>	$bark.mm = (0.785 * \sqrt{dbh.cm})^2$	equation	30	10-200	California (3 montane sites)	Zelhig-Kichas et al. (2016)
20	<i>Quercus alba</i>	$bark.mm = 1.828 * log(dbh.cm + 1)$	data	10	9.3-101.8	SCBI	Anderson-Teixeira et al. (2015)
21	<i>Quercus montana</i>	$bark.mm = 2.083 * log(dbh.cm + 1)$	data	8	5.8-99.1	SCBI	Anderson-Teixeira et al. (2015)
22	<i>Quercus rubra</i>	$bark.mm = 0.98 * log(dbh.cm + 1)$	data	10	24.1-143.2	SCBI	Anderson-Teixeira et al. (2015)
23	<i>Quercus velutina</i>	$bark.mm = 1.394 * log(dbh.cm + 1)$	data	8	16.2-110.7	SCBI	Anderson-Teixeira et al. (2015)
24	<i>Tetragastris panamensis</i>	$bark.mm = 1.672 * log(dbh.cm + 1)$	data	4	22.7-48.8	Panama	Raquel Alfaro-Sánchez (unpublished data)
25	<i>Trichilia tuberculata</i>	$bark.mm = 1.367 * log(dbh.cm + 1)$	data	12	21-40.5	Panama	Raquel Alfaro-Sánchez (unpublished data), Pete Kerby-Miller and Helene Muller-Landau (unpublished data)
26	<i>Tsuga canadensis</i>	$bark.mm = ((0.18 + 0.08 * (dbh.cm/2.54))/2) * 2.54$	equation	NA		North America	Miles, Patrick D.; Smith, W. Brad. 2009.

For assignments of species as proxies for those with out available bark allometries, see Table S2.

Table S5. Qualitative comparison of results from this study with previous studies employing conventional methods.

species	Precipitation response		Temperature response		reference
	previously observed	observed here	previously observed	observed here	
Barro Colorado Nature Monument, Panama					
JACO	pos. correlation to Apr-Dec <i>PPT</i> (strongest of the 3 species)	pos. correlation to Jan-Dec <i>PPT</i> (strongest of the 3 species)	no sig. correlation to annual T_{mean} or T_{min}	no correlation to Feb-Mar T_{min}	Alfaro-Sánchez et al. 2017
TEPA	pos. correlation to Apr-Dec <i>PPT</i> (response weaker than JACO, similar to TRTU)	pos. correlation to Jan-Dec <i>PPT</i> (response weaker than JACO, similar to TRTU)	no sig. correlation to annual T_{mean} or T_{min}	no correlation to Feb-Mar T_{min}	Alfaro-Sánchez et al. 2017
TRTU	pos. correlation to Apr-Dec <i>PPT</i> (response weaker than JACO, similar to TEPA)	pos. correlation to Jan-Dec <i>PPT</i> (response weaker than JACO, similar to TEPA)	no sig. correlation to annual T_{mean} or T_{min}	non-sig. pos. correlation to Feb-Mar T_{min}	Alfaro-Sánchez et al. 2017
Huai Kha Khaeng, Thailand					
AFXY	sig. pos. correlation with June <i>PPT</i> , otherwise n.s.	pos. correlation to p.Sep-Jun <i>PPT</i> frequency	sig. neg. correlation with T_{max} in Aug and Dec; T_{min} in p.Oct., Jul, Aug	slight pos. correlation to Apr-Oct T_{max}	Vlam et al. 2013
CHTA	sig. pos. correlation with April <i>PPT</i> , otherwise n.s.	no sig. correlation to p.Sep-Jun <i>PPT</i> frequency	sig. neg. correlation with T_{max} in May, Aug-Sept; T_{min} in Feb, May, Aug	neg. correlation to Apr-Oct T_{max}	Vlam et al. 2013
MEAZ	sig. pos. correlation with April <i>PPT</i> , otherwise n.s.	pos. correlation to p.Sep-Jun <i>PPT</i> frequency	sig. neg. correlation with T_{max} in May-Aug; T_{min} in May-Aug	neg. correlation to Apr-Oct T_{max}	Vlam et al. 2013
TOCI	sig. pos. correlation with p.Oct-p.Nov and April-May <i>PPT</i>	pos. correlation to p.Sep-Jun <i>PPT</i> frequency	sig. neg. correlation with T_{max} every month from pOct-June (excluding March); T_{min} in Jan and Mar-Aug	neg. correlation to Apr-Oct T_{max}	Vlam et al. 2013
Smithsonian Conservation Biology Institute, Virginia, USA					
CACO	pos. correlations with May-Aug <i>PPT</i> (sig. May, July)	pos. concave-down correlation to June-Aug <i>PPPT</i>	neg. correlations with May-Aug <i>PET</i> (sig. May-July)	neg. correlation to May-July <i>PET</i>	Helcoski et al. 2019
CAGL	pos. correlations with May-Aug <i>PPT</i> (sig. May)	pos. concave-down correlation to June-Aug <i>PPPT</i>	neg. correlations with May-Aug <i>PET</i> (n.s.)	neg. correlation to May-July <i>PET</i>	Helcoski et al. 2019
CAOVL	pos. correlations with May-Aug <i>PPT</i> (sig. Aug)	pos. concave-down correlation to June-Aug <i>PPPT</i>	neg. correlations with May-Aug <i>PET</i> (sig. all months)	neg. concave-down correlation to May-July <i>PET</i>	Helcoski et al. 2019
CATO	pos. correlations with May-Aug <i>PPT</i> (n.s.)	concave-down correlation to June-Aug <i>PPPT</i>	neg. correlations with May-Aug <i>PET</i> (sig. June)	neg. correlation to May-July <i>PET</i>	Helcoski et al. 2019
FAGR	pos. correlations with May-Aug <i>PPT</i> (sig. July-Aug)	pos. concave-down correlation to June-Aug <i>PPPT</i>	neg. correlations with May-Aug <i>PET</i> (sig. July-Aug)	no correlation to May-July <i>PET</i>	Helcoski et al. 2019
FRAM	pos. correlations with May-Aug <i>PPT</i> (sig. May-June)	pos. concave-down correlation to June-Aug <i>PPPT</i>	neg. correlations with May-Aug <i>PET</i> (sig. May-June)	neg. concave-down correlation to May-July <i>PET</i>	Helcoski et al. 2019
FRNI	no sig. correlations with peak growing season <i>PPT</i>	non-sig. pos. concave-down correlation to June-Aug <i>PPPT</i>	no sig. correlations with peak growing season <i>PET</i>	no correlation to May-July <i>PET</i>	Helcoski et al. 2019
JUNI	pos. correlations with May-Aug <i>PPT</i> (sig. Jun-Aug)	pos. concave-down correlation to June-Aug <i>PPPT</i>	neg. correlations with May-Aug <i>PET</i> (sig. July-Aug)	neg. concave-down correlation to May-July <i>PET</i>	Helcoski et al. 2019

S5, cont.

species	Precipitation response		Temperature response		reference
	previously observed	observed here	previously observed	observed here	
Smithsonian Conservation Biology Institute, Virginia, USA (cont.)					
LITU	pos. correlations with May-Aug <i>PPT</i> (sig. May-July)	pos. concave-down correlation to June-Aug <i>PPPT</i>	neg. correlations with May-Aug <i>PET</i> (sig. all months)	non-sig neg. concave-down correlation to May-July <i>PET</i>	Helcoski et al. 2019
PIST	pos. correlations with May-Aug <i>PPT</i> (n.s.)	pos. concave-down correlation to June-Aug <i>PPPT</i>	neg. correlations with May-Aug <i>PET</i> (n.s.)	no correlation to May-July <i>PET</i>	Helcoski et al. 2019
QUAL	pos. correlations with May-Aug <i>PPT</i> (sig. May)	pos. concave-down correlation to June-Aug <i>PPPT</i>	neg. correlations with May-Aug <i>PET</i> (sig. all months)	non-sig neg. concave-down correlation to May-July <i>PET</i>	Helcoski et al. 2019
QUMO	pos. correlations with May-Aug <i>PPT</i> (sig. May)	pos. concave-down correlation to June-Aug <i>PPPT</i>	neg. correlations with May-Aug <i>PET</i> (sig. May-June, Aug)	non-sig neg. concave-down correlation to May-July <i>PET</i>	Helcoski et al. 2019
QURU	pos. correlations with May-Aug <i>PPT</i> (n.s.)	pos. concave-down correlation to June-Aug <i>PPPT</i>	neg. correlations with May-Aug <i>PET</i> (sig. May, July-Aug)	neg. concave-down correlation to May-July <i>PET</i>	Helcoski et al. 2019
QUVE	pos. correlations with May-Aug <i>PPT</i> (sig. May-July)	pos. concave-down correlation to June-Aug <i>PPPT</i>	neg. correlations with May-Aug <i>PET</i> (sig. all months)	neg. concave-down correlation to May-July <i>PET</i>	Helcoski et al. 2019
Lilly Dickey Woods, Indiana, USA					
LITU	pos. correlations with Jun-Aug PDSI	pos. response to June <i>PPT</i>	neg. response to Jun-Aug <i>T_{max}</i>	neg. response to June <i>PET</i>	Maxwell, Harley, and Robeson 2016
QUAL	pos. correlations with Jun-Aug PDSI	pos. response to June <i>PPT</i>	neg. response to Jun-Aug <i>T_{max}</i>	neg. response to June <i>PET</i>	Maxwell, Harley, and Robeson 2016
QUMO	pos. correlations with Jun-Aug PDSI	pos. response to June <i>PPT</i>	neg. response to Jun-Aug <i>T_{max}</i>	neg. response to June <i>PET</i>	Maxwell, Harley, and Robeson 2016
QUVE	pos. correlations with Jun-Aug PDSI	pos. response to June <i>PPT</i>	neg. response to Jun-Aug <i>T_{max}</i>	neg. response to June <i>PET</i>	Maxwell, Harley, and Robeson 2016
Harvard Forest, Massachusetts, USA					
ACRU	NA		no response to Jan-April <i>T_{min}*</i>	neg. correlation to Mar <i>PET</i>	Alexander et al. 2019
BEAL	NA		no response to Jan-April <i>T_{min}*</i>	neg. correlation to Mar <i>PET</i>	Alexander et al. 2019
QURU	NA		no response to Jan-April <i>T_{min}*</i>	neg. correlation to Mar <i>PET</i>	Alexander et al. 2019
TSCA	NA		pos. response to Jan-April <i>T_{min}*</i>	pos. correlation to March <i>PET</i>	Alexander et al. 2019

S5, cont.

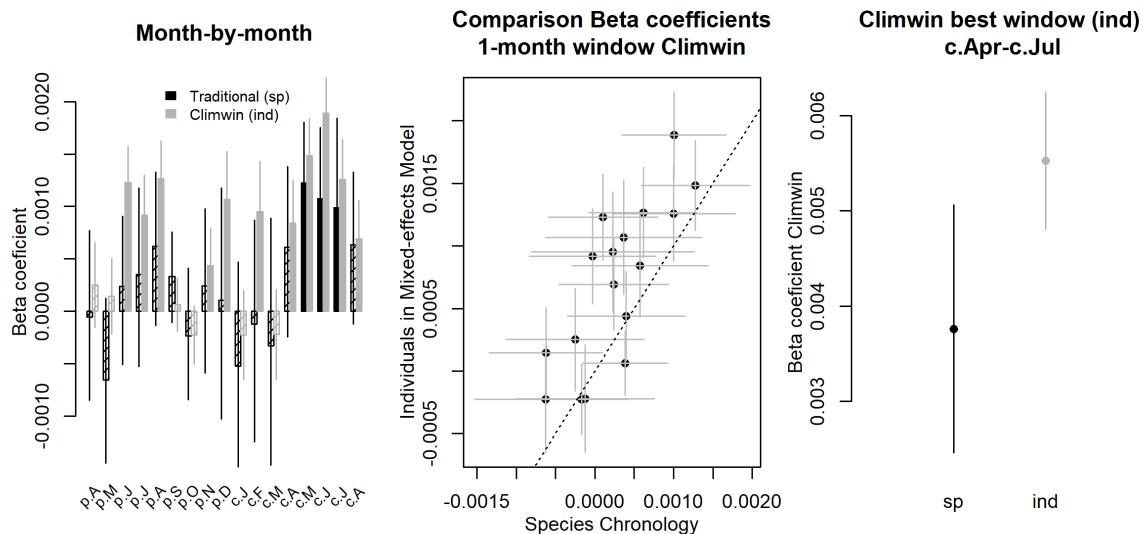
species	Precipitation response		Temperature response		reference
	previously observed	observed here	previously observed	observed here	
Žofín Forest Dynamics Plot, Czech Republic					
ABAL	no sig. correlations with June-July <i>PPT</i>	pos. correlation to p.Jun-p.July <i>PPT</i> frequency	sig. pos. correlation to April T (strongest T correlation)	pos. correlation to Jan-March T_{max}	Kašpar, Tumajer, Vašíčková, and Šamonil, in review
FASY	no sig. correlations with June-July <i>PPT</i>	pos. correlation to p.Jun-p.July <i>PPT</i> frequency	sig. pos. correlation to Jan T (strongest T correlation)	pos. correlation to Jan-March T_{max}	Kašpar, Tumajer, Vašíčková, and Šamonil, in review
PIAB	modest pos. correlations (n.s) with June-July <i>PPT</i> ≥700m elev. sites moisture limited June-Aug	non-sig. pos. correlation to p.Jun-p.July <i>PPT</i> frequency	sig. pos. correlation to March T (strongest current-year T correlation) ≥700m elev. sites temperature limited except June-Aug	pos. correlation to Jan-March T_{max}	Kašpar, Tumajer, Vašíčková, and Šamonil, in review Tumajer et al. 2017
Niobrara, Nebraska, USA					
BEPA	little relationship to <i>PPT</i> within analysis timeframe (exception: pos. correlation with pAug <i>PPT</i>); positive correlation to streamflow (<i>SF</i>) in some months (pJune, pAug, pNov, cJune); stronger relationship to PDSI	no sig. correlation to <i>PPT</i> , <i>PDF</i> , or <i>SF</i>	little relationship to T_{mean} within analysis timeframe (exception: neg. correlation with pJune and cJan T_{mean})	neg. concave-down correlation to pJune T_{mean}	Bumann et al. 2019
Little Tesuque, New Mexico, USA					
PIPO	increases with pOct-June <i>PPT</i> ** increase with cold-season <i>PPT</i> **	pos. concave-down correlation to pNov-cJuly <i>PPT</i>	NA		Touchan et al., 2011
PIST3	increases with pOct-June <i>PPT</i> **	pos. concave-down correlation to pNov-cJuly <i>PPT</i>	decrease with vapor pressure deficit of pAug-pOct and May-July** NA	neg. concave-down correlation to May-July T_{max}	Williams et al., 2013
Cedar Breaks, Utah, USA					
	NA		NA		-
Scotty Creek, NW Territories, Canada					
PIMA	predominantly pos. responses to annual <i>PPT</i>	pos. concave-down correlation to pJune-pAug <i>PPT</i>	predominantly positive responses to mean annual T_{mean} prior to 1970, shifting to predominantly negative responses after 1970	pos. correlation to July T_{max} . Temperature correlations predominantly pos. prior to 1970 (to pAug T_{min}), neg. after 1970 (to pJune T_{min})	Sniderhan and Baltzer 2016

*Indicates results from a regional study including but not limited to cores from the focal site.

**Indicates results from a regional study not including the focal site, but believed to be representative.

Figure S1. Comparison of our approach with traditional methods of identifying climate signals: *Liriodendron tulipifera* L. at the Smithsonian Conservation Biology Institute (SCBI), VA, USA.

Precipitation



Potential Evapotranspiration

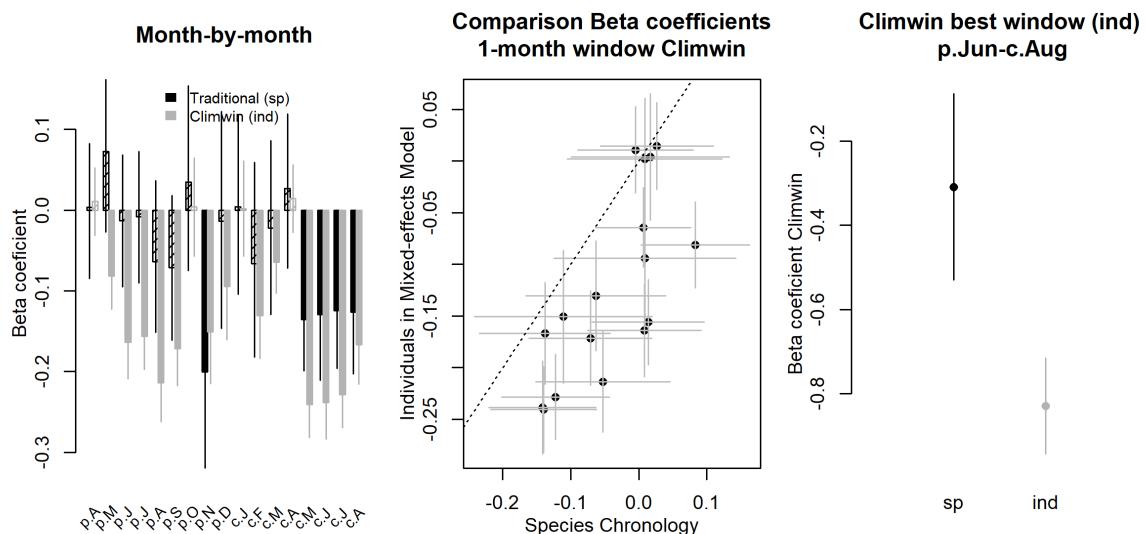
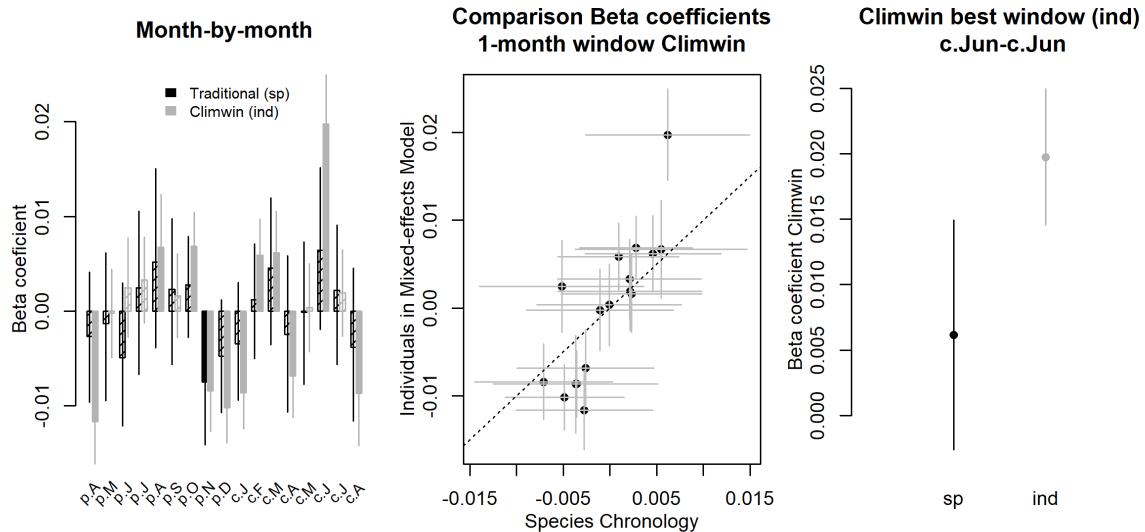


Figure S1. Comparison of our approach with traditional methods of identifying climate signals: *Liriodendron tulipifera* L. at the Smithsonian Conservation Biology Institute (SCBI), VA, USA. Shown are responses to the precipitation- and temperature-group variables selected as most influential by the *climwin* analysis. Left panels show a month-by-month comparison of *beta* (slope) coefficients for the relationship between tree growth and the monthly climate variable from species-level residual chronologies (traditional approach) and from individual-level analysis in *climwin* (approach presented here). Center panels compare the monthly *beta* coefficient estimates, with the dotted line indicating 1:1 correspondence. Finally, the right panels compare *beta* coefficients for the most influential window selected by *climwin*. Error bars indicate standard error

of slope estimates. Note that 1:1 correspondence is not necessarily expected. See Appendix 5 for analysis methods and discussion of expected correspondence.

Figure S2. Comparison of our approach with traditional methods of identifying climate signals: *Abies alba* Mill. at Zofin, Czech Republic.

Precipitation Day Frequency



Maximum temperature

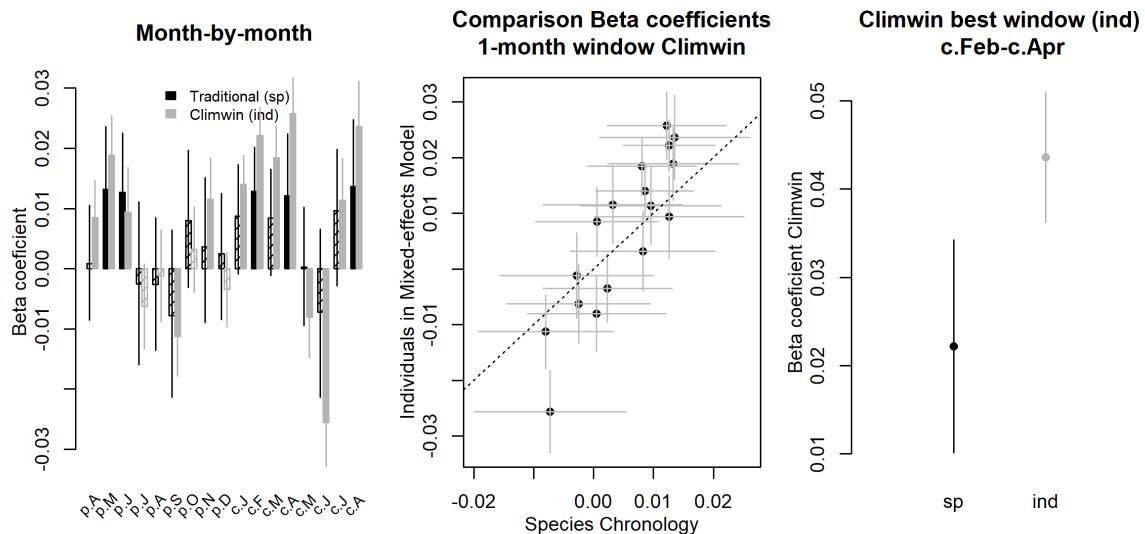
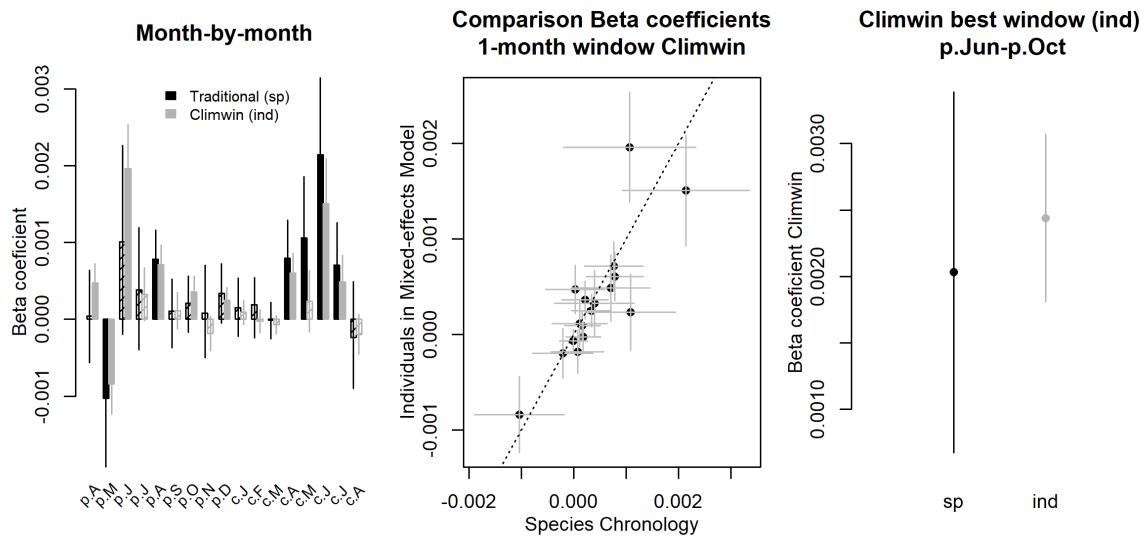


Figure S2. Comparison of our approach with traditional methods of identifying climate signals: *Abies alba* Mill. at Zofin, Czech Republic. Shown are responses to the precipitation- and temperature-group variables selected as most influential by the *climwin* analysis. Left panels show a month-by-month comparison of *beta* (slope) coefficients for the relationship between tree growth and the monthly climate variable from species-level residual chronologies (traditional approach) and from individual-level analysis in *climwin* (approach presented here). Center panels compare the monthly *beta* coefficient estimates, with the dotted line indicating 1:1 correspondence. Finally, the right panels compare *beta* coefficients for the most influential window selected by *climwin*. Error bars indicate standard error of slope estimates. Note that 1:1 correspondence is not necessarily expected. See Appendix 5 for analysis methods and discussion of expected correspondence.

Figure S3. Comparison of our approach with traditional methods of identifying climate signals: *Pseudotsuga menziesii* (Mirb.) Franco at Cedar Breaks, UT, USA.

Precipitation



Maximum temperature

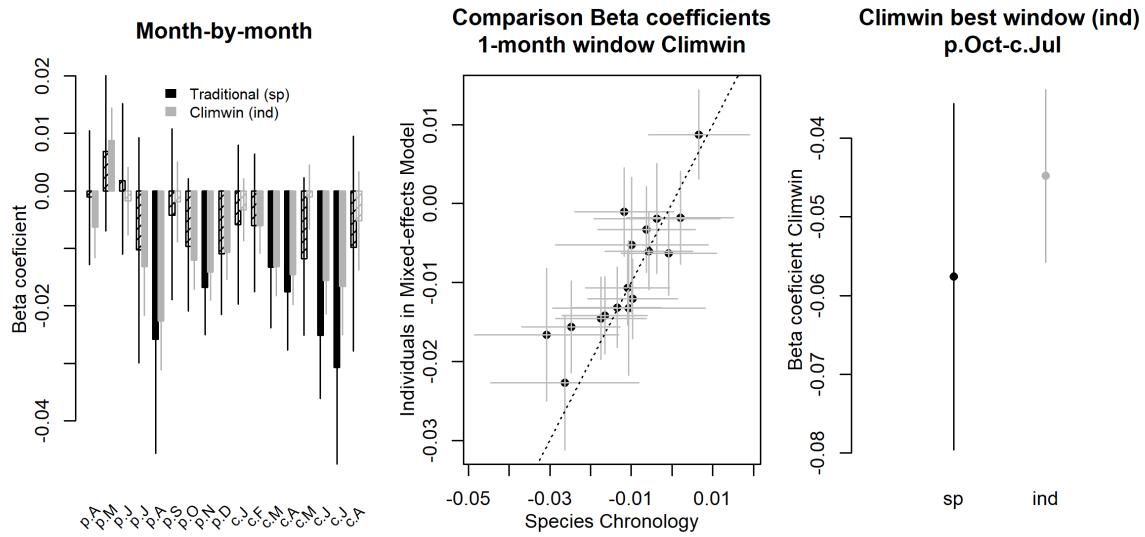
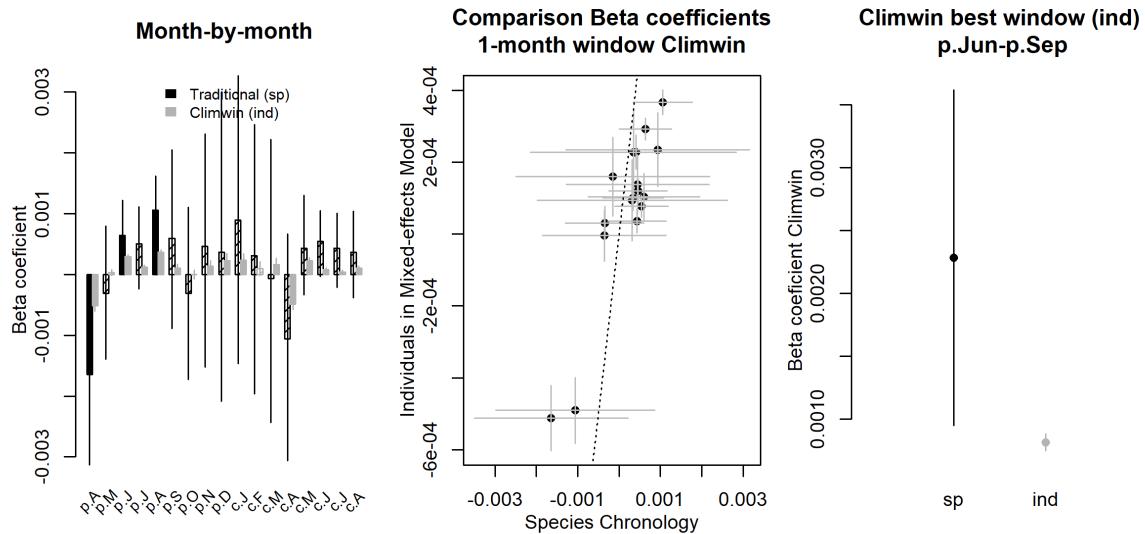


Figure S3. Comparison of our approach with traditional methods of identifying climate signals: *Pseudotsuga menziesii* (Mirb.) Franco at Cedar Breaks, UT, USA. Shown are responses to the precipitation- and temperature-group variables selected as most influential by the *climwin* analysis. Left panels show a month-by-month comparison of *beta* (slope) coefficients for the relationship between tree growth and the monthly climate variable from species-level residual chronologies (traditional approach) and from individual-level analysis in *climwin* (approach presented here). Center panels compare the monthly *beta* coefficient estimates, with the dotted line indicating 1:1 correspondence. Finally, the right panels compare *beta* coefficients for the most influential window selected by *climwin*. Error bars indicate standard error of slope estimates. Note

that 1:1 correspondence is not necessarily expected. See Appendix 5 for analysis methods and discussion of expected correspondence.

Figure S4. Comparison of our approach with traditional methods of identifying climate signals: *Picea mariana* a (Mill.) Britton, Sterns and Poggenb. at Scotty Creek, NT, Canada.

Precipitation



Maximum temperature

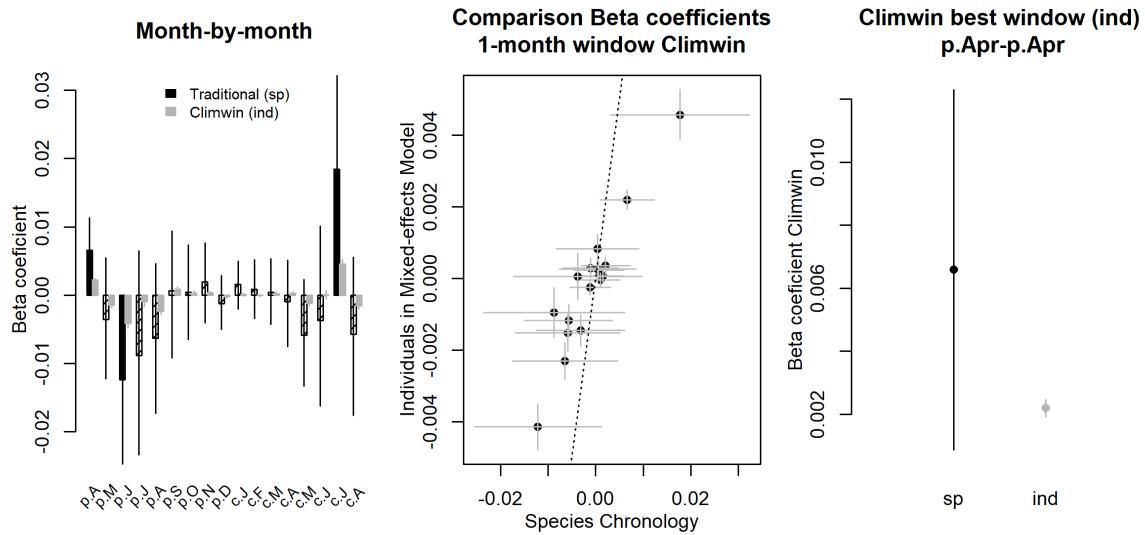


Figure S4. Comparison of our approach with traditional methods of identifying climate signals: *Picea mariana* a (Mill.) Britton, Sterns and Poggenb. at Scotty Creek, NT, Canada. Shown are responses to the precipitation- and temperature-group variables selected as most influential by the *climwin* analysis. Left panels show a month-by-month comparison of *beta* (slope) coefficients for the relationship between tree growth and the monthly climate variable from species-level residual chronologies (traditional approach) and from individual-level analysis in *climwin* (approach presented here). Center panels compare the monthly *beta* coefficient estimates, with the dotted line indicating 1:1 correspondence. Finally, the right panels compare *beta* coefficients for the most influential window selected by *climwin*. Error bars indicate standard error

of slope estimates. Note that 1:1 correspondence is not necessarily expected. See Appendix 5 for analysis methods and discussion of expected correspondence.

Figure S5. Example comparison of year effect in GLS model with independent decadal analysis for Harvard Forest, MA, USA.

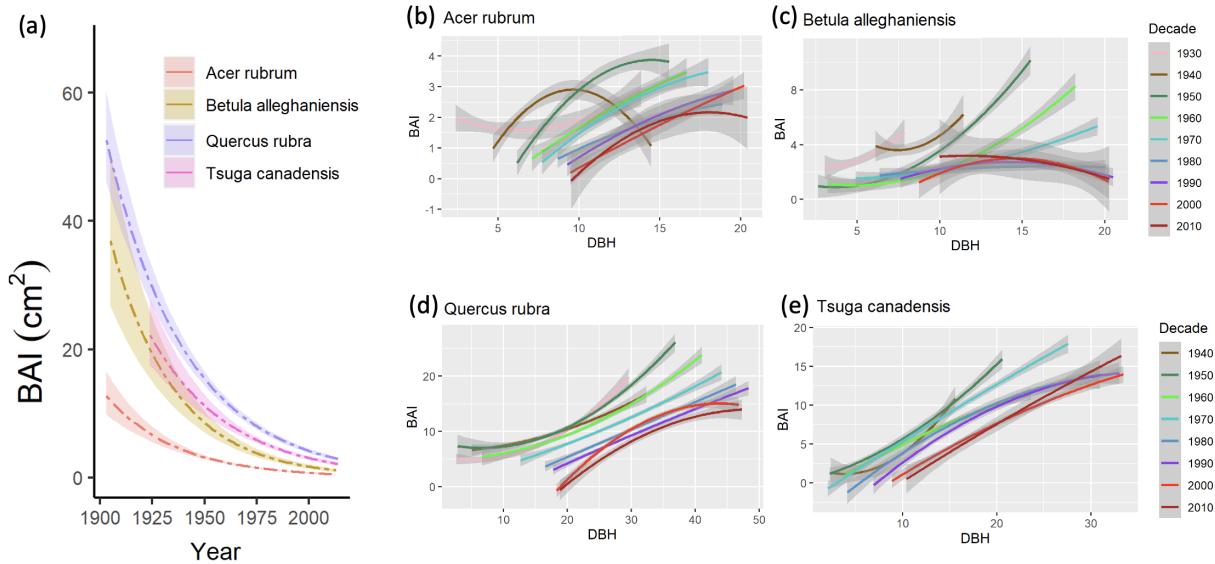


Figure S5. Example comparison of year effect in GLS model with independent decadal analysis for Harvard Forest, MA, USA.. Panel (a) shows results the effect of year in the full GLS model (all significant with t-test's p-value <0.05). Other terms in the model are held constant at their medians. Transparent ribbons indicate 95% confidence intervals. Panels (b-e) show relationships between DBH and BAI by decade for each species individually. These plots visualize the data included in the GLS model, separated by decade, using a probability density function. Transparent ribbons indicate 95% confidence intervals. Parallel plots for all sites are available in the GitHub repository for this project: https://github.com/EcoClimLab/ForestGEO-tree-rings/tree/master/doc/supplementary_figures/decadal_plots.

Figure S6. Climwin output describing the strongest predictor of interannual variation in tree growth within the temperature variable group at Huai Kha Khaeng, Thailand.

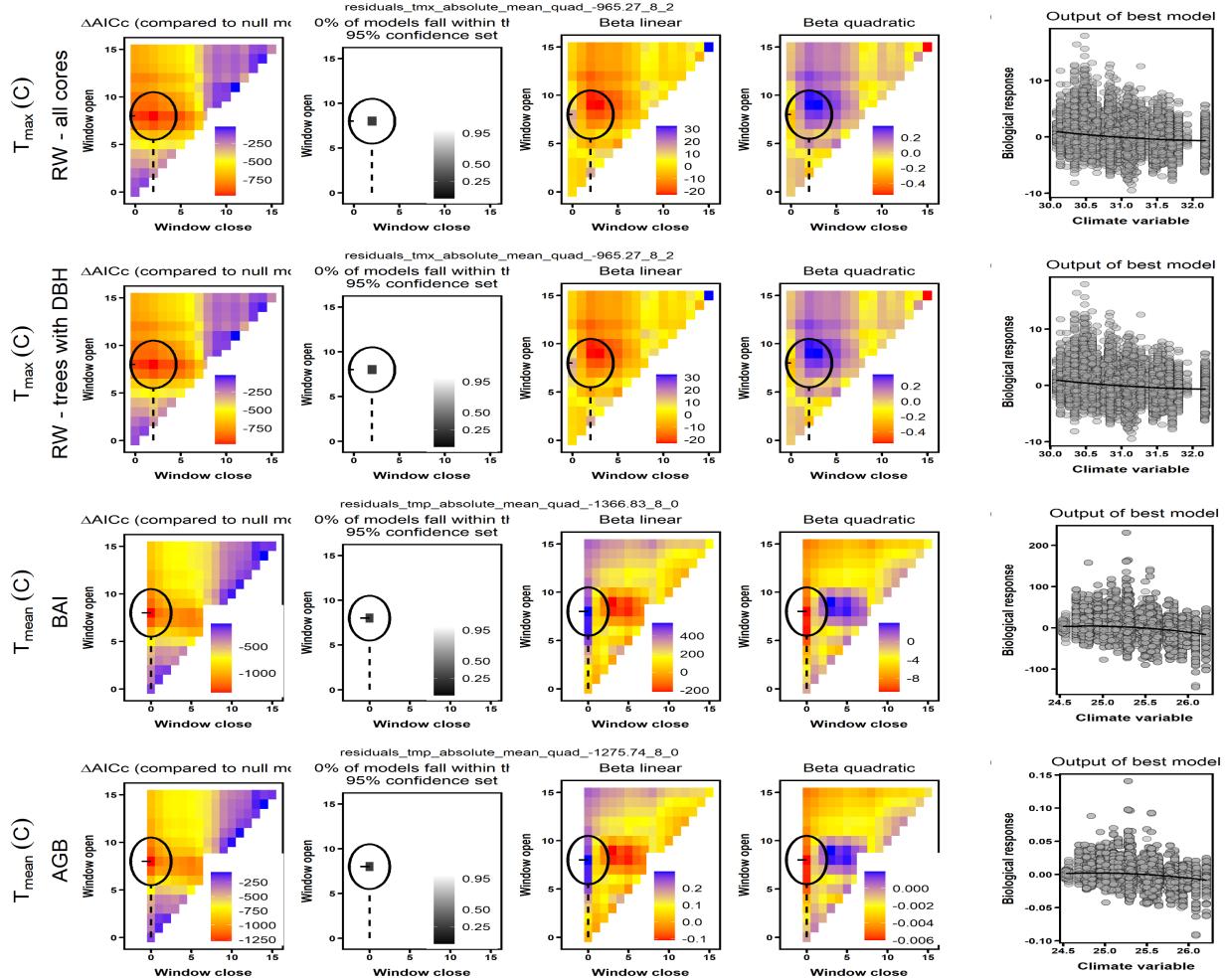


Figure S6. Climwin output describing the strongest predictor of interannual variation in tree growth within the temperature variable group at Huai Kha Khaeng, Thailand. Rows correspond to the four different models run: (1) all RW data, (2) RW for the subset of cores for which \$DBH could be reconstructed, (3) BAI, and (4) AGB growth. The first four columns give statistics for seasonal windows tested in *climwin*, where window open and close indicate months prior to the last month in the climwin analysis window for the site (Table S1). Difference in $AICc$ relative to a model with no climate term (first column) is used to select the most influential seasonal window (second column). The third and fourth columns give values of linear and quadratic terms for each seasonal window. The final column shows the correlation of individual-level residuals to the selected climate driver, with the function fit in *climwin* (not the final fit). Parallel figures for all sites may be found at https://github.com/EcoClimLab/ForestGEO-tree-rings/tree/master/doc/manuscript/tables_figures/SI_figures/climwin_plots_combined.

Figure S7. Climwin output describing the strongest predictor of interannual variation in tree growth within the water variable group at the Smithsonian Conservation Biology Institute (SCBI), VA, USA.

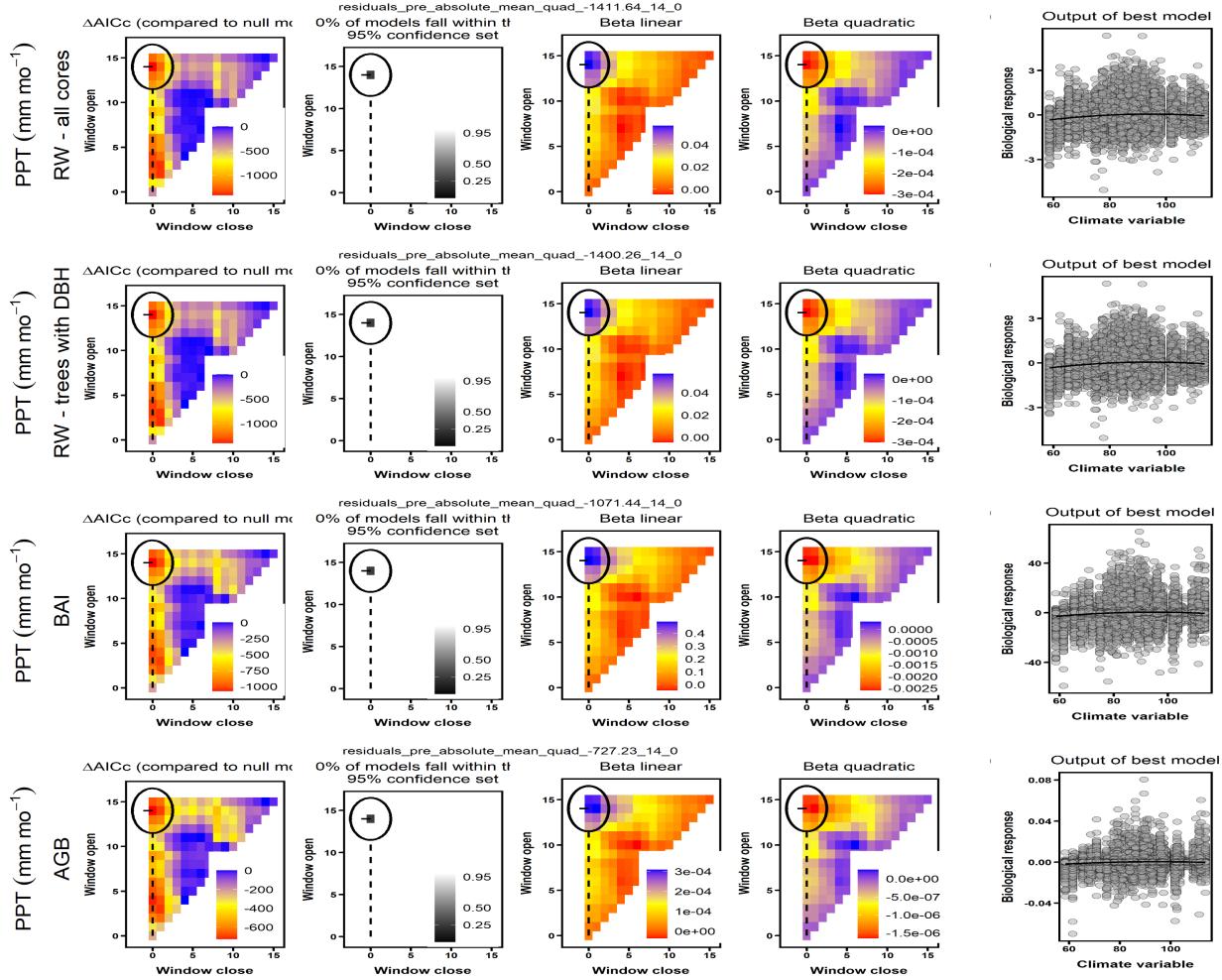


Figure S7. Climwin output describing the strongest predictor of interannual variation in tree growth within the water variable group at the Smithsonian Conservation Biology Institute (SCBI), VA, USA. Rows correspond to the four different models run: (1) all RW data, (2) RW for the subset of cores for which \$DBH could be reconstructed, (3) BAI, and (4) AGB growth. The first four columns give statistics for seasonal windows tested in *climwin*, where window open and close indicate months prior to the last month in the climwin analysis window for the site (Table S1). Difference in $AICc$ relative to a model with no climate term (first column) is used to select the most influential seasonal window (second column). The third and fourth columns give values of linear and quadratic terms for each seasonal window. The final column shows the correlation of individual-level residuals to the selected climate driver, with the function fit in *climwin* (not the final fit). Parallel figures for all sites may be found at https://github.com/EcoClimLab/ForestGEO-tree-rings/tree/master/doc/manuscript/tables_figures/SI_figures/climwin_plots_combined.

Figure S8. Climwin output describing the strongest predictor of interannual variation in tree growth within the temperature variable group at the Smithsonian Conservation Biology Institute (SCBI), VA, USA.

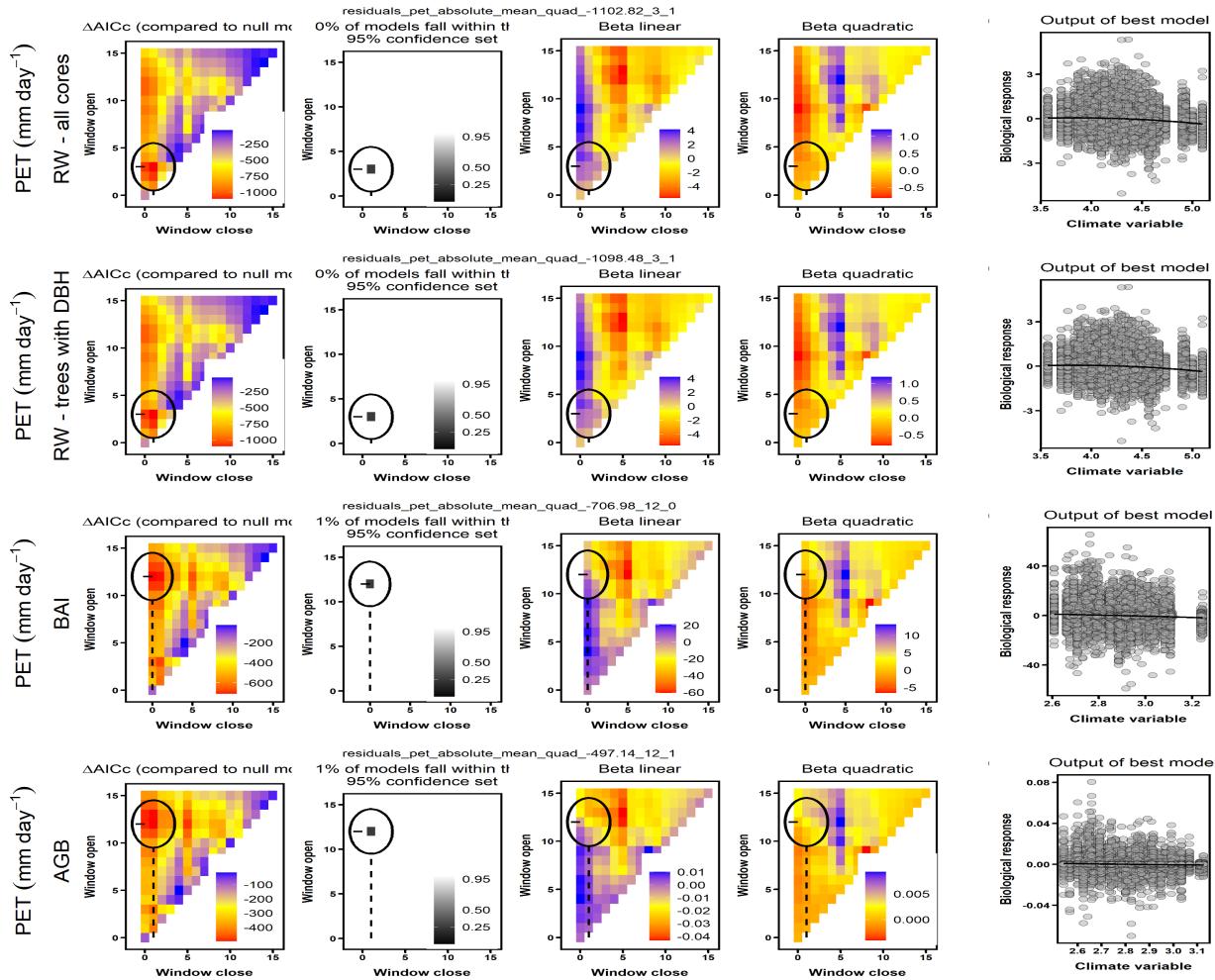


Figure S8. Climwin output describing the strongest predictor of interannual variation in tree growth within the temperature variable group at the Smithsonian Conservation Biology Institute (SCBI), VA, USA. Rows correspond to the four different models run: (1) all RW data, (2) RW for the subset of cores for which \$DBH could be reconstructed, (3) BAI, and (4) AGB growth. The first four columns give statistics for seasonal windows tested in *climwin*, where window open and close indicate months prior to the last month in the climwin analysis window for the site (Table S1). Difference in $AICc$ relative to a model with no climate term (first column) is used to select the most influential seasonal window (second column). The third and fourth columns give values of linear and quadratic terms for each seasonal window. The final column shows the correlation of individual-level residuals to the selected climate driver, with the function fit in *climwin* (not the final fit). Parallel figures for all sites may be found at https://github.com/EcoClimLab/ForestGEO-tree-rings/tree/master/doc/manuscript/tables_figures/SI_figures/climwin_plots_combined.

Figure S9. Climwin output describing the strongest predictor of interannual variation in tree growth within the water variable group at Harvard Forest, MA, USA.

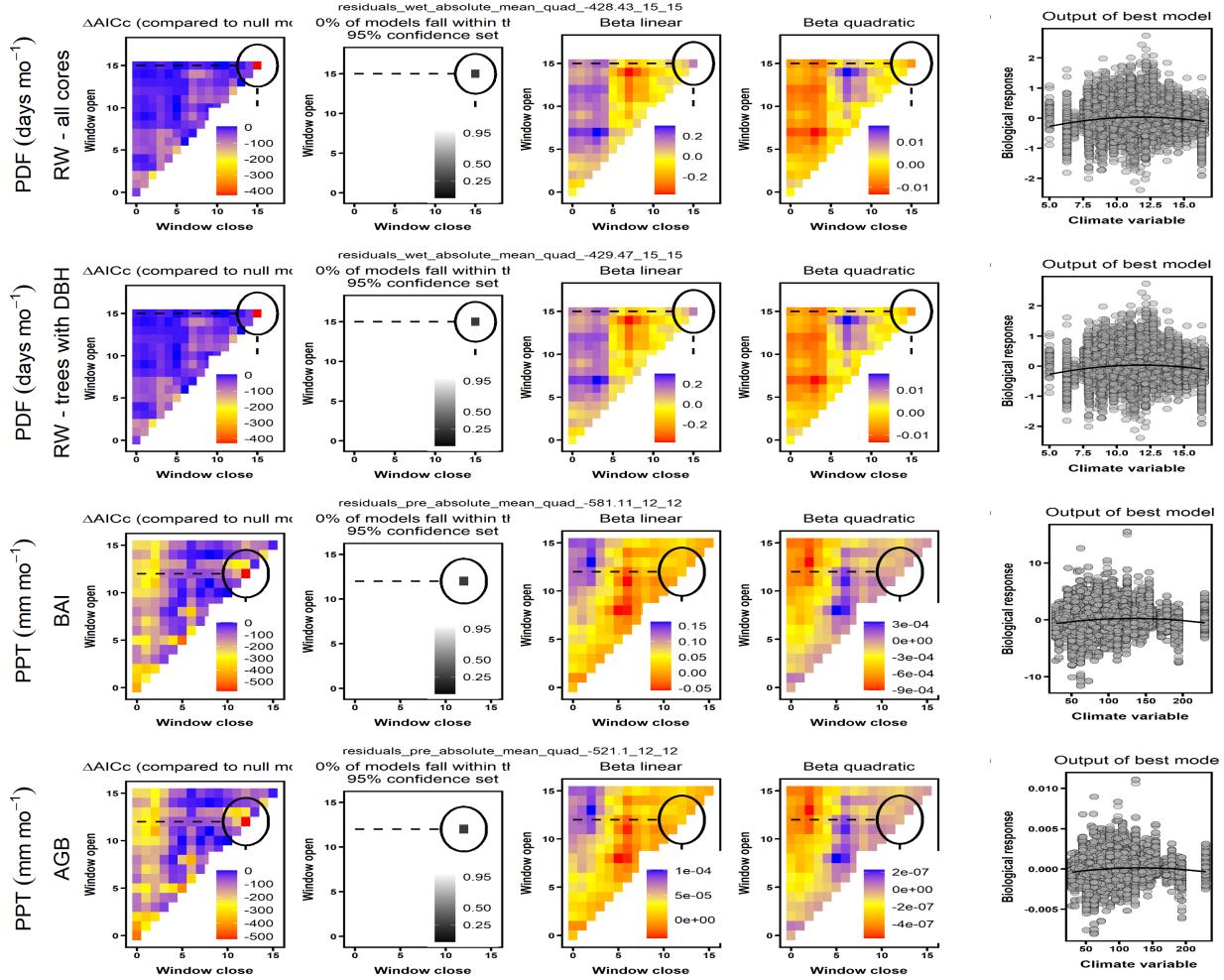


Figure S9. Climwin output describing the strongest predictor of interannual variation in tree growth within the water variable group at Harvard Forest, MA, USA. Rows correspond to the four different models run: (1) all RW data, (2) RW for the subset of cores for which \$DBH could be reconstructed, (3) BAI, and (4) AGB growth. The first four columns give statistics for seasonal windows tested in *climwin*, where window open and close indicate months prior to the last month in the *climwin* analysis window for the site (Table S1). Difference in $AICc$ relative to a model with no climate term (first column) is used to select the most influential seasonal window (second column). The third and fourth columns give values of linear and quadratic terms for each seasonal window. The final column shows the correlation of individual-level residuals to the selected climate driver, with the function fit in *climwin* (not the final fit). Parallel figures for all sites may be found at https://github.com/EcoClimLab/ForestGEO-tree-rings/tree/master/doc/manuscript/tables_SI_figures/SI_figures/climwin_plots_combined.

Figure S10. Best generalized least squares (GLS) models describing annual tree growth based on climate and diameter breast height (DBH) for Barro Colorado National Monument (BCNM), Panama.

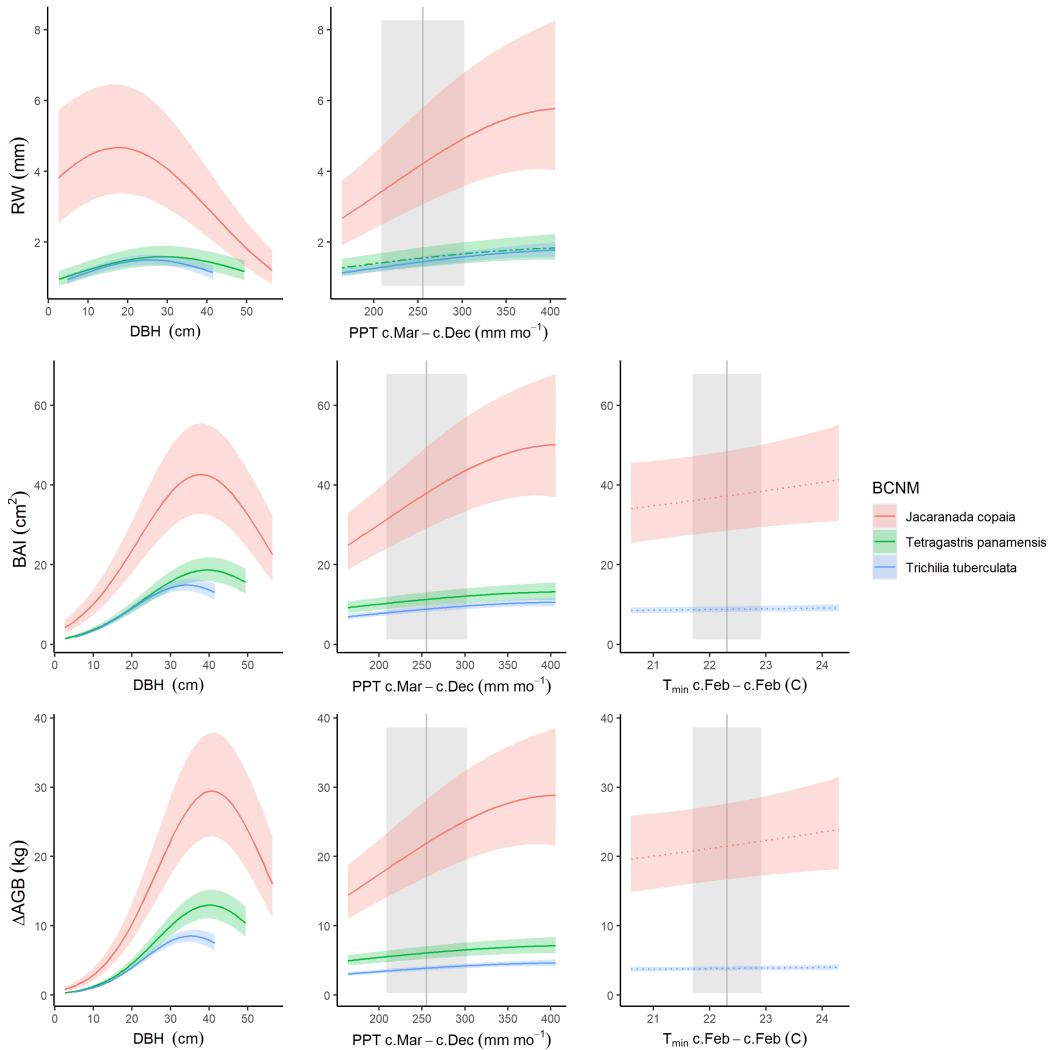


Figure S10. Best generalized least squares (GLS) models describing annual tree growth based on climate and diameter breast height (DBH) for Barro Colorado National Monument (BCNM), Panama. For all three growth metrics examined here. For each species that met the criteria for inclusion in the analysis, relationships are plotted if included in top model. For each relationship shown, other terms in the model are held constant at their medians. Best-fit polynomials are plotted with solid lines when both first- and second-order terms are significant (t-test's p-value <0.05), dash-dotted lines when only one term is significant, and dotted lines when neither is significant. Transparent ribbons indicate 95% confidence intervals. *Climwin*-selected climate drivers are coded on the x-axes as the climate variable name followed by the range of months (p=previous year, c=current year) over which it is most influential. Vertical grey lines indicate the long-term mean for the climate driver over the analysis period; shading indicates 1 SD. Parallel figures for all sites may be found at https://github.com/EcoClimLab/ForestGEO-tree-rings/tree/master/doc/manuscript/tables_figures/SI_figures/composite_plots.

Figure S11. Best generalized least squares (GLS) models describing annual tree growth based on climate and diameter breast height (DBH) for Smithsonian Conservation Biology Institute (SCBI), VA, USA.

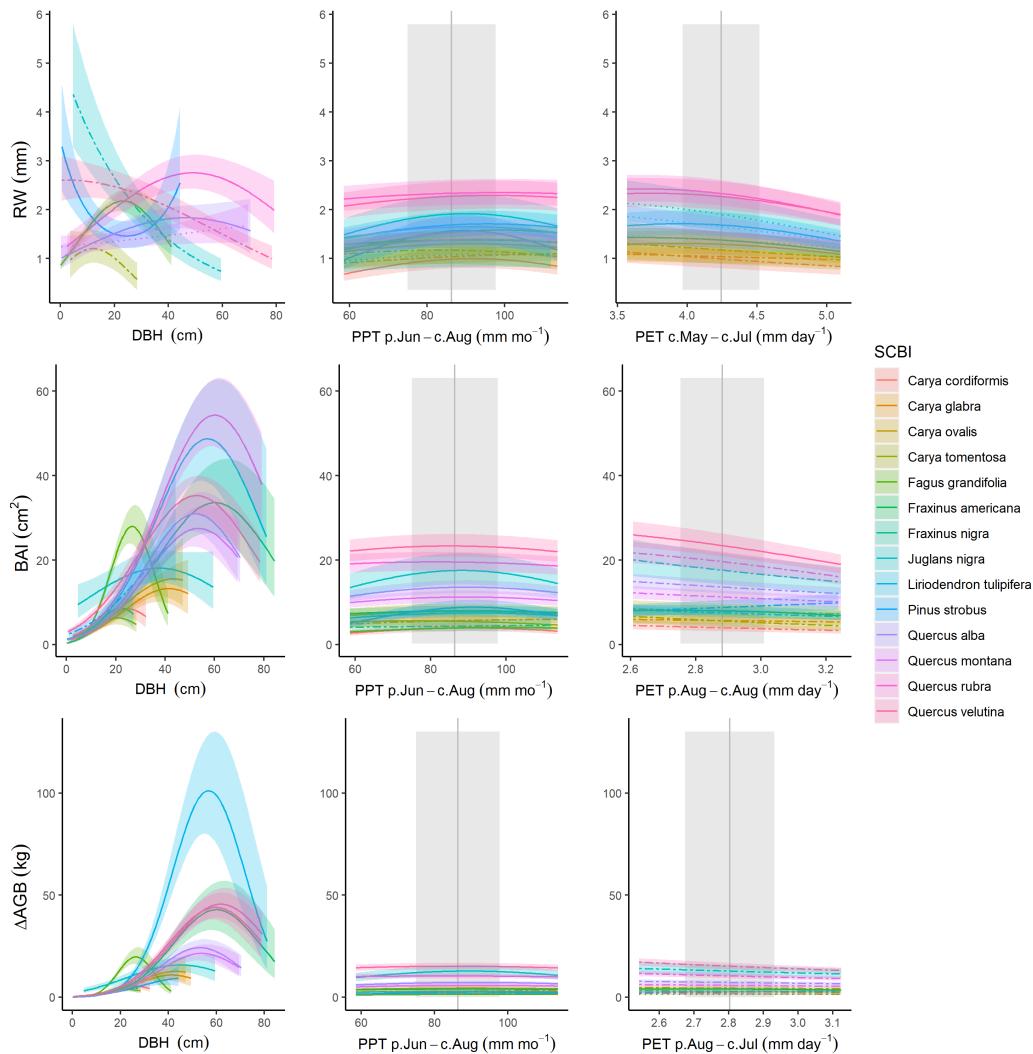


Figure S11. Best generalized least squares (GLS) models describing annual tree growth based on climate and diameter breast height (DBH) for Smithsonian Conservation Biology Institute (SCBI), VA, USA. for all three growth metrics examined here. For each species that met the criteria for inclusion in the analysis, relationships are plotted if included in top model. For each relationship shown, other terms in the model are held constant at their medians. Best-fit polynomials are plotted with solid lines when both first- and second-order terms are significant (t-test's p-value <0.05), dash-dotted lines when only one term is significant, and dotted lines when neither is significant. Transparent ribbons indicate 95% confidence intervals. *Climwin*-selected climate drivers are coded on the x-axes as the climate variable name followed by the range of months (p=previous year, c=current year) over which it is most influential. Vertical grey lines indicate the long-term mean for the climate driver over the analysis period; shading indicates 1 SD. Parallel figures for all sites may be found at https://github.com/EcoClimLab/ForestGEO-tree-rings/tree/master/doc/manuscript/tables_figures/SI_figures/composite_plots.

Figure S12. Best generalized least squares (GLS) models describing annual tree growth based on climate, diameter breast height (DBH), and year for Smithsonian Conservation Biology Institute (SCBI), VA, USA.

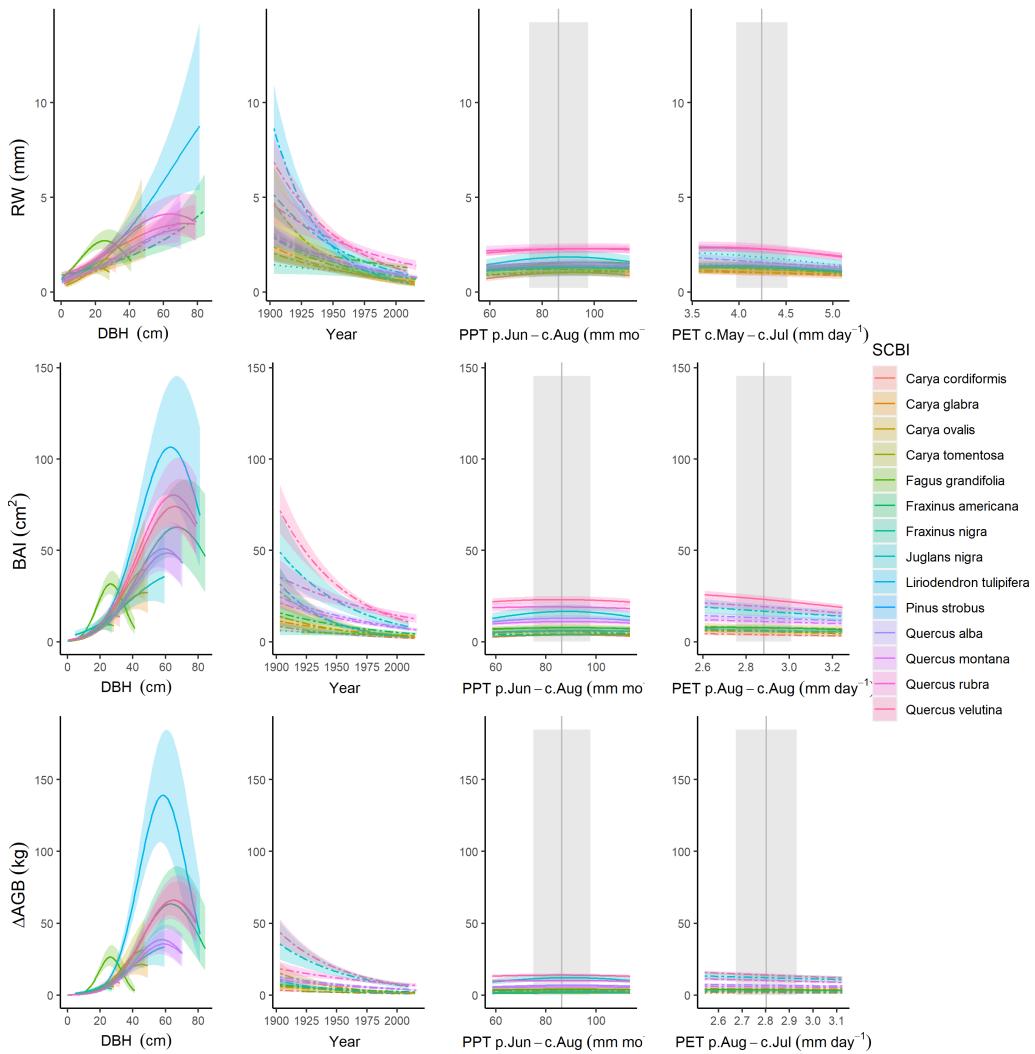


Figure S12. Best generalized least squares (GLS) models describing annual tree growth based on climate, diameter breast height (DBH), and year for Smithsonian Conservation Biology Institute (SCBI), VA, USA. for all three growth metrics examined here. For each species that met the criteria for inclusion in the analysis, relationships are plotted if included in top model. For each relationship shown, other terms in the model are held constant at their medians. Best-fit polynomials are plotted with solid lines when both first- and second-order terms are significant (*t*-test's *p*-value <0.05), dash-dotted lines when only one term is significant, and dotted lines when neither is significant. Transparent ribbons indicate 95% confidence intervals. *Climwin*-selected climate drivers are coded on the x-axes as the climate variable name followed by the range of months (p=previous year, c=current year) over which it is most influential. Vertical grey lines indicate the long-term mean for the climate driver over the analysis period; shading indicates 1 SD. Parallel figures for all sites may be found at https://github.com/EcoClimLab/ForestGEO-tree-rings/tree/master/doc/manuscript/tables_figures/SI_figures/composite_plots.

Figure S13. Best generalized least squares (GLS) models describing annual tree growth based on climate, diameter breast height (DBH), and year for Little Tesuque, NM, USA.

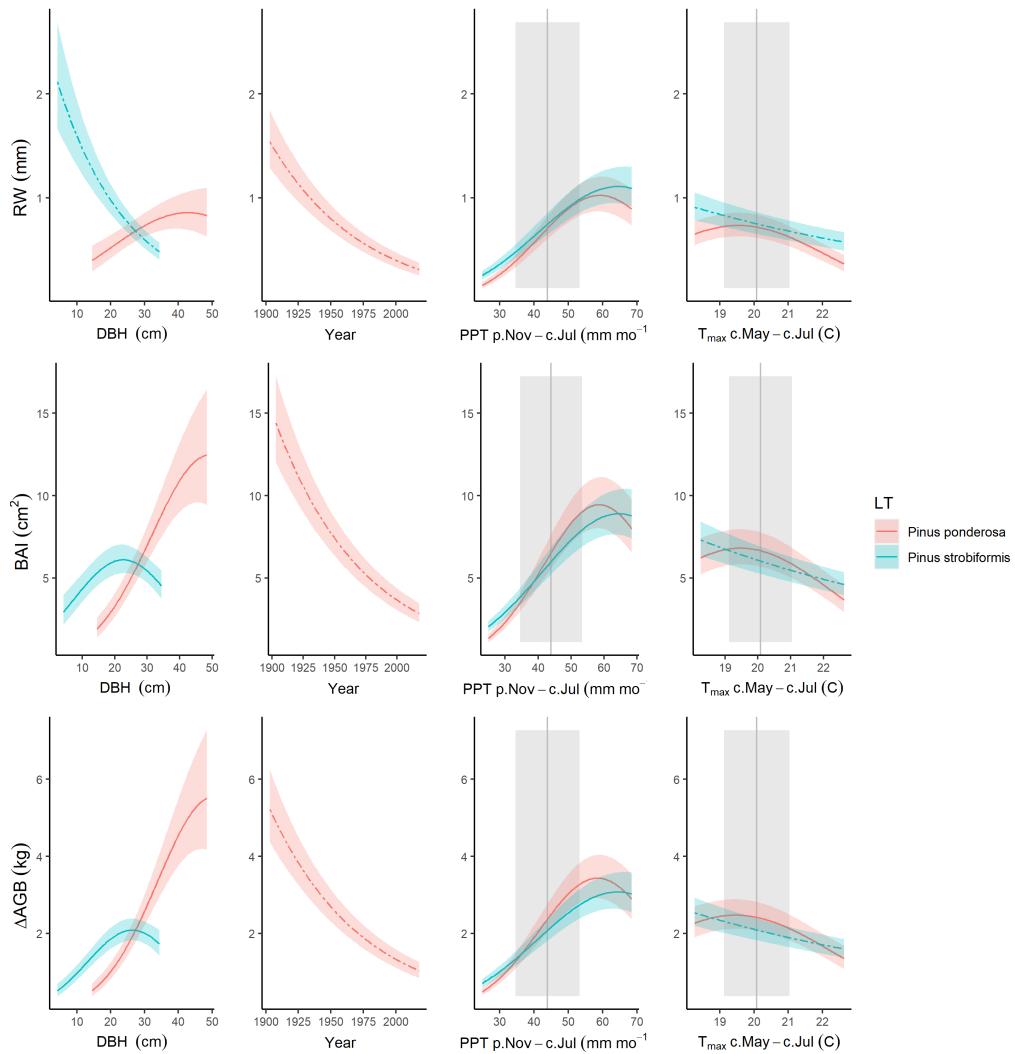
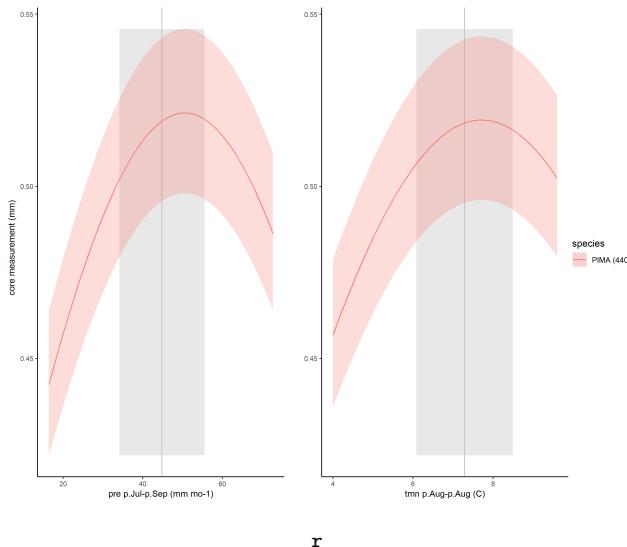


Figure S13. Best generalized least squares (GLS) models describing annual tree growth based on climate, diameter breast height (DBH), and year for Little Tesuque, NM, USA. for all three growth metrics examined here. For each species that met the criteria for inclusion in the analysis, relationships are plotted if included in top model. For each relationship shown, other terms in the model are held constant at their medians. Best-fit polynomials are plotted with solid lines when both first- and second-order terms are significant (t-test's p-value <0.05), dash-dotted lines when only one term is significant, and dotted lines when neither is significant. Transparent ribbons indicate 95% confidence intervals. Climwin-selected climate drivers are coded on the x-axes as the climate variable name followed by the range of months (p=previous year, c=current year) over which it is most influential. Vertical grey lines indicate the long-term mean for the climate driver over the analysis period; shading indicates 1 SD. Parallel figures for all sites may be found at https://github.com/EcoClimLab/ForestGEO-tree-rings/tree/master/doc/manuscript/tables_figures/SI_figures/composite_plots.

Figure S14. Interannual climate sensitivity of annual tree growth at Scotty Creek, NT, Canada before and after 1970.

(a) pre-1970



r

(b) post-1970

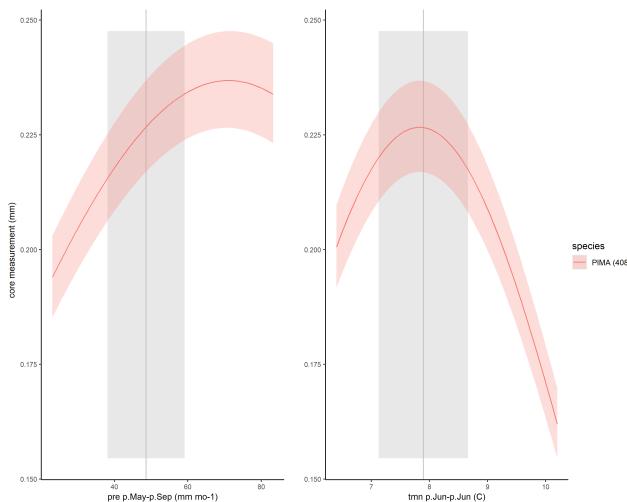


Figure S14. Interannual climate sensitivity of annual tree growth at Scotty Creek, NT, Canada before and after 1970. *Climwin*-selected climate drivers are coded on the x-axes as the climate variable name followed by the range of months (p=previous year, c=current year) over which it is most influential. For each relationship shown, other terms in the model are held constant at their medians. Best-fit polynomials are plotted with solid lines when both first- and second-order terms are significant (t-test's p-value <0.05), dash-dotted lines when only one term is significant, and dotted lines when neither is significant. Transparent ribbons indicate 95% confidence intervals. Vertical grey lines indicate the long-term mean for the climate driver over the analysis period; shading indicates 1 SD.

Figure S15. Significant climate - diameter breast height (DBH) interactions in generalized least squares (GLS) models describing interannual variation in ring width (RW) for tree species at 10 sites.

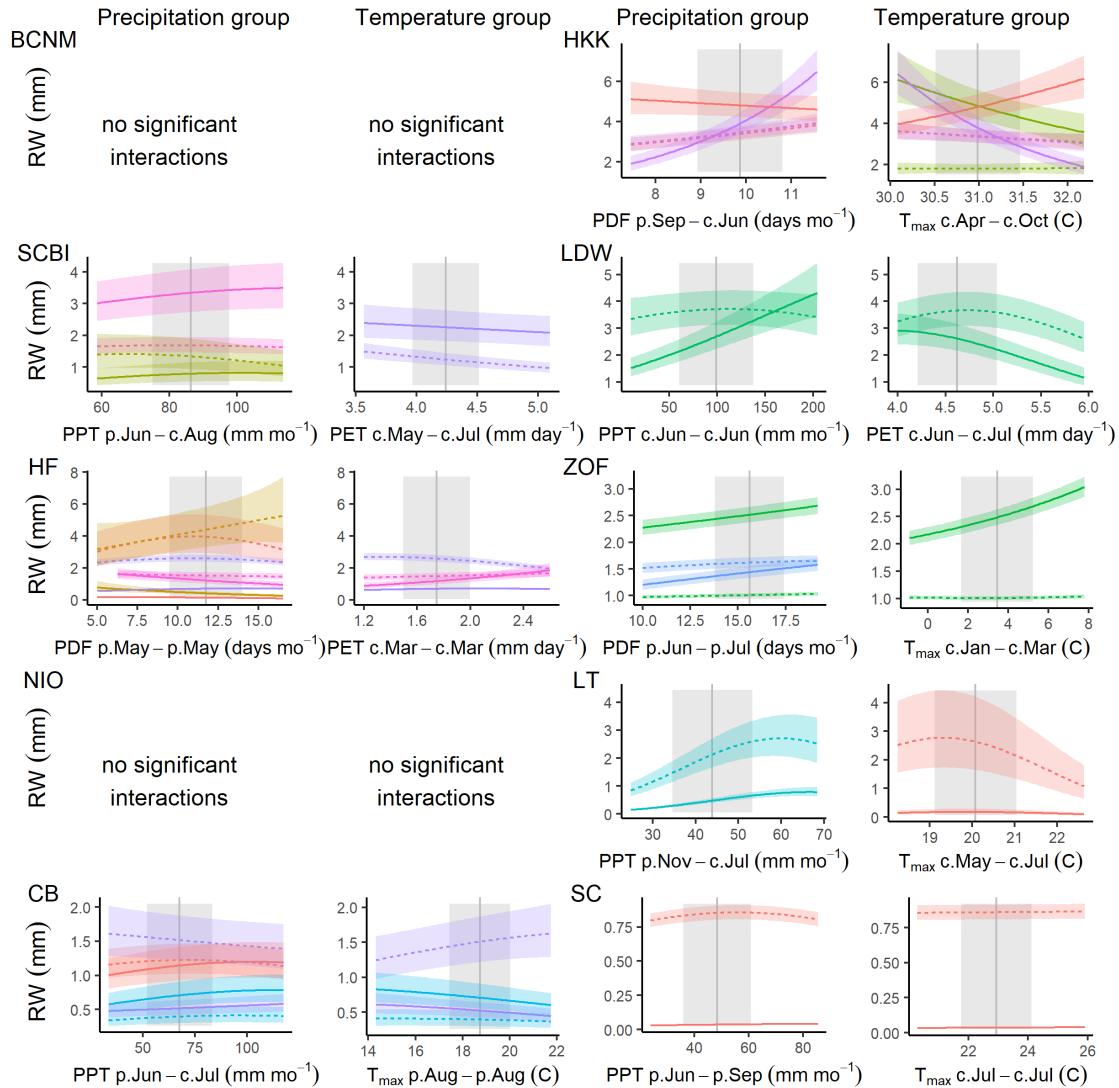


Figure S15. Significant climate - diameter breast height (DBH) interactions in generalized least squares (GLS) models describing interannual variation in ring width (RW) for tree species at 10 sites. Shown are modeled response functions for species at the minimum and maximum and maximum ends of the DBH range. Other terms in the model are held constant at their medians. Transparent ribbons indicate 95% confidence intervals. Vertical grey lines indicate the long-term mean for the climate driver over the analysis period; shading indicates 1 SD. Species are color coded as in Figure 6.

Figure S16. Responses of annual growth, expressed as ring width (RW), (b) basal area increment (BAI), or (c) aboveground biomass (AGB) increase, to calendar year for 35 species at 10 sites.

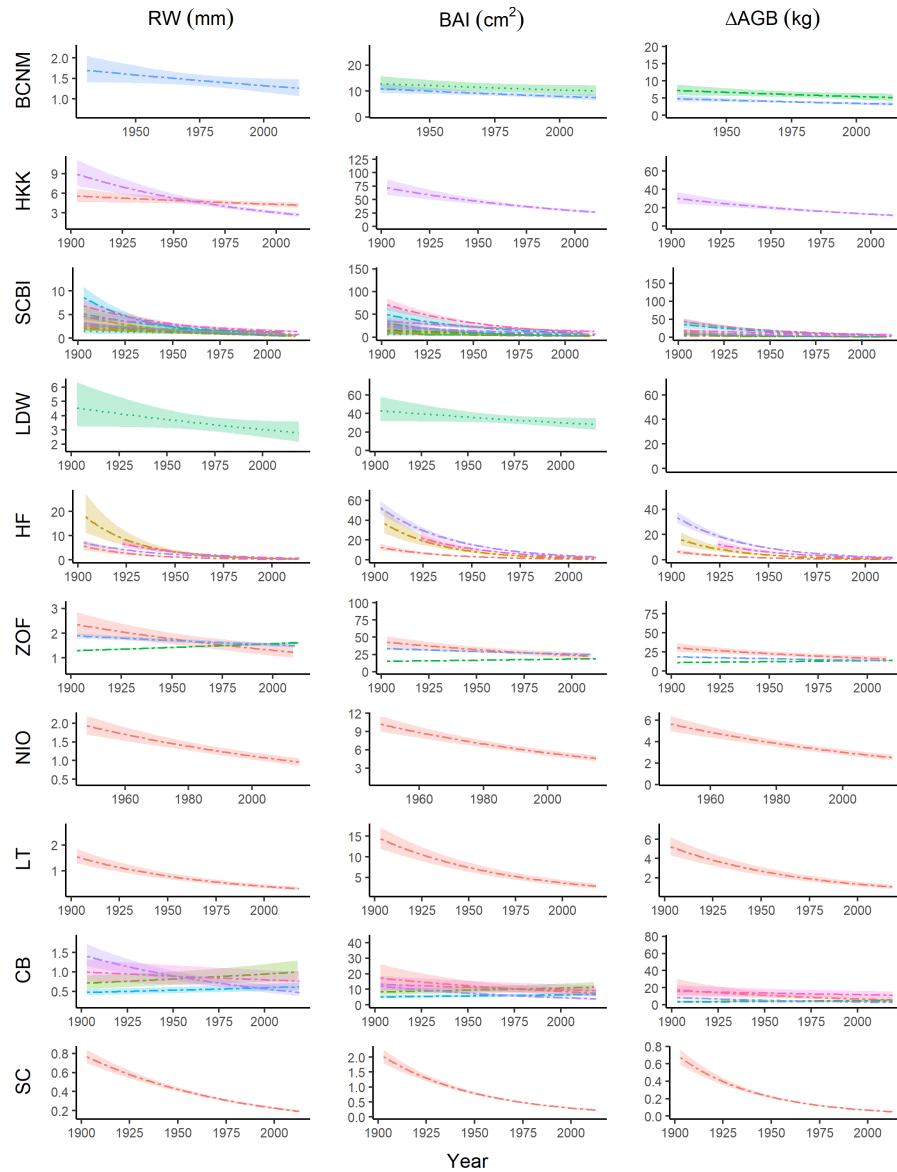


Figure S16. Responses of annual growth, expressed as ring width (RW), (b) basal area increment (BAI), or (c) aboveground biomass (AGB) increase, to calendar year for 35 species at 10 sites. For each species (color coded as in Figure 6), relationships are plotted if the year effect could be analyzed and was included in the top model. Other terms in the model are held constant at their medians. Best-fit polynomials are plotted with solid lines when both first- and second-order terms are significant (t-test's p-value < 0.05), dash-dotted lines when only one term is significant, and dotted lines when neither is significant. Transparent ribbons indicate 95% confidence intervals.

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