



PowerMag[®]

Microbiome RNA/DNA Isolation Kit

(Optimized for KingFisher[®])

Catalog No.	Quantity	Total Purifications
27600-4-KF	4 x 96 Preps (Flex) or 32 x 12 Preps (Duo)	384

Instruction Manual



Please recycle

Version: 09182014

Technical Information: Toll free 1-800-606-6246, or 1-760-929-9911 Email: technical@mobio.com Website: www.mobio.com



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Introduction

The PowerMag[®] Microbiome RNA/DNA Isolation Kit is a magnetic bead based nucleic acid isolation kit optimized for use with automated platforms.

The PowerMag[®] Microbiome kit can be used for automated isolation of microbial RNA and DNA from all stool, gut, and similar sample types and other difficult samples containing high inhibitor content, such as bile, bilirubin, digested food, and can be used for soil and environmental samples high in humic acids. The kit, which can process up to 0.25 g of sample, employs patented Inhibitor Removal Technology[®] (IRT) to remove PCR inhibitors released during the extraction process. A novel, proprietary magnetic bead system is used for the isolation of nucleic acids without the binding of residual contaminants, for inhibitor-free RNA and DNA that is ready to use in the most demanding downstream applications including cDNA synthesis, RT-qPCR, PCR, qPCR and next generation sequencing.

This kit requires the use of a specialized plate shaker in order to facilitate the bead beating process in the PowerMag[®] Glass Bead Plates. We recommend the Retsch 96 Well Plate Shaker (MO BIO Catalog# 11996 in the USA only) and Adapters (MO BIO Catalog# 11990). For information outside the USA, contact technical@mobio.com.

This kit was optimized on the KingFisher[®] Flex and Duo instruments for isolation of DNA from up to 450 µl of lysate per well.

NOTE: The order of placement of components and reagents for the platform portion of the protocol will be described in the downloaded software specific to your KingFisher[®] platform.

Other open platform robots may be used with this kit however you may need to contact your local field application scientist for the manufacturer of your robot for help in adapting this protocol to your system.

Protocol Overview

Microbiome samples are added to a 96 well bead beating plate for rapid and thorough homogenization. Cell lysis occurs by a combination of mechanical and chemical methods. Inhibitory compounds are removed using Inhibitor Removal Technology[®]. Total nucleic acids are captured on specialized magnetic beads in the presence of buffers that avoid the use of chaotropic salts and ethanol. RNA and DNA are washed on the beads and then eluted using RNase-Free Water. The eluted nucleic acids are ready for qPCR, next generation sequencing and other downstream applications.

It is important to note that quantification of the DNA using PicoGreen[®] will be approximately 15% lower than the actual yield due to the presence of residual wash solution in the DNA. The wash solution does not inhibit PCR or interfere with next generation sequencing.

This kit is for research purposes only. Not for diagnostic use.

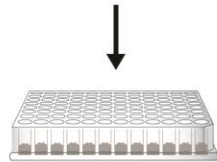
Other Related Products	Catalog No.	Quantity
96 Well Plate Shaker	11996	1 unit (120 V)
Plate Adapter Set	11990	1 set
Anti-Static Polypropylene Weighing Funnels, Small	23302-50	1 bag of 50

PowerMag® Microbiome RNA/DNA Isolation Kit

Prepare Sample



- Add PowerMag® Microbiome Lysis Solution/ β -ME to the PowerMag® Glass Bead Plate



- Add 0.25 grams of stool or biosolid to the PowerMag® Glass Bead Plate

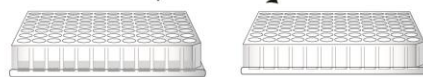
Cell Lysis



- Place PowerMag® Glass Bead Plate in the 96 Well Plate Shaker and shake

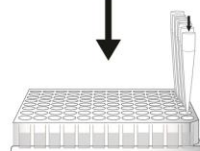


- Centrifuge



- Transfer the supernatant to a clean PowerMag® 1 ml Collection Plate

Inhibitor Removal Technology®



- Add PowerMag® Inhibitor Removal Solution and vortex briefly
- Incubate at 4° C



- Centrifuge

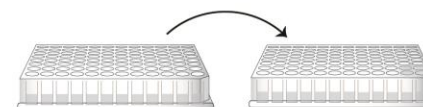


- Transfer the entire volume of supernatant to a new PowerMag® 1 ml Collection Plate



- Centrifuge

RNA/DNA Isolation



- Transfer up to 450 μ l of supernatant to a KingFisher® Deep Well 96 Plate



Place KingFisher® Deep Well 96 Plate on the KingFisher® platform and initiate the PowerMag® Microbiome protocol



Equipment and Reagents Required

- Centrifuge capable of handling two 96 Well Blocks (13 cm x 8.5 cm x 6.0 cm) at 4500 x *g*
Note: If you have a centrifuge with a maximum speed less than 4500 x *g* see the Hints and Troubleshooting Guide.
- Multi-channel Pipettor(s) (volumes of 100 μ l - 850 μ l)
Note: The KingFisher[®] Duo applications require a 12 channel pipettor if multi-channel pipetting is desired on that platform.
- Single Pipettor(s) (volumes of 5 μ l – 1000 μ l)
- Mechanical Shaker for 96 Well Blocks and Plate Adapters (MO BIO Catalog# 11996 and 11990)
- Vortex-Genie[®] 2 Vortex with 3 inch platform (MO BIO Catalog# 13111-V or 13111-V-220)
- B-mercaptoethanol
- Optional - Phenol:Chloroform:Isoamyl Alcohol (PCI) (25:24:1, pH 6.5 – 8)

Consumables not Included

- Contact your Thermo Scientific representative for the KingFisher[®] Flex and Duo consumables specific to your platform. Go to www.mobio.com/powermag for links to the necessary KingFisher[®] products on the ThermoFisher website.
- Multi-channel pipettor reagent reservoirs for 10 – 150 ml volumes.
- Appropriate pipet tips for the Multi-channel pipettors to be used in the lysate preparation steps.
Note: The tips must fit in the round wells of the 1 ml blocks (examples of these are Molecular Bioproducts ART Catalog# 2179-HR, Eppendorf Catalog# 0030 077.750 and Rainin Catalog# RT-1000F).

Kit Contents

Kit Catalog# 27600-4-KF		
Component	Catalog #	Amount
PowerMag [®] Glass Bead Plates (w/Sealing Mat)	27600-4-KF-BP	4
Bead Plate Sealing Mats	27600-4-KF-SM	4
PowerMag [®] Microbiome Lysis Solution	27600-4-KF-1	2 x 141 ml
PowerMag [®] Inhibitor Removal Solution	27600-4-KF-2	3 x 22 ml
ClearMag [®] Binding Solution	27600-4-KF-3	200 ml
ClearMag [®] Beads	27600-4-KF-4	9 ml
ClearMag [®] Wash Solution	27600-4-KF-5	640 ml
ClearMag [®] RNase-Free Water	27600-4-KF-6	44 ml
PowerMag [®] 1ml Collection Plates	27600-4-KF-1CP	8
Sealing Tape	27600-4-KF-ST	32

Kit Storage

The kit reagents and components should be stored at room temperature (15-30°C).

Precautions

Please wear gloves when using this product. Avoid all skin contact with kit reagents. In case of contact, wash thoroughly with water. Do not ingest. See Material Safety Data Sheets for emergency procedures in case of accidental ingestion or contact. All MSDS information is available upon request (760-929-9911) or at www.mobio.com. Reagents labeled flammable should be kept away from open flames and sparks.



Protocol

Warm the PowerMag[®] Microbiome Lysis Solution at 60°C for 15-20 minutes before starting to dissolve any precipitates.

Add β -mercaptoethanol (β -ME) at a ratio of 25 μ l per ml of the PowerMag[®] Microbiome Lysis Solution. You will need 64 ml of PowerMag[®] Microbiome Lysis Solution/ β -ME per 96 well plate.

1. Briefly centrifuge (1 minute at 4500 x g) the **PowerMag[®] Glass Bead Plate** to bring all of the glass beads down into the well. Carefully peel off the Sealing Mat that covers the **PowerMag[®] Glass Bead Plate** and discard.
2. Add 650 μ l of warmed **PowerMag[®] Microbiome Lysis Solution/ β -ME** to each well of the **PowerMag[®] Glass Bead Plate**.
Note: PowerMag[®] Microbiome Lysis Solution contains SDS. If it gets cold, it will precipitate. Heating at 60°C will dissolve the SDS. PowerMag[®] Microbiome Lysis Solution can be used while it is still warm.
Optional: To enhance the recovery and integrity of RNA, addition of 100 μ l of phenol:chloroform:isoamyl alcohol (PCI) (25:24:1, pH 6.5-8) to the wells of the bead plate pre-loaded with 650 μ l of PowerMag[®] Microbiome Lysis Solution/ β -ME before filling with stool samples will allow for fast nuclease inactivation during the filling process.
3. Add 0.25 grams of stool or environmental sample to each well of the **PowerMag[®] Glass Bead Plate**.
Note: This is the most time consuming step of the protocol. Care must be taken to avoid cross contamination between sample wells. Use of an Anti-Static Polypropylene Weighing Funnel (MO BIO Catalog# 23302-50) can make it easier to weigh and add some sample types to each well without spilling into adjacent wells.
4. Secure a new **Bead Plate Sealing Mat** tightly to the **PowerMag[®] Glass Bead Plate**. Vortex horizontally for 5 seconds on the vortex ensuring that the solution / sample is well mixed.
Note: A proper seal of the mat is critical to prevent loss of sample and leakage that might cause damage to your shaker.
This is an appropriate stopping point. You can store the PowerMag[®] Glass Bead Plates overnight at -20°C covered with a new Bead Plate Sealing Mat. Be sure to equilibrate to room temperature prior to proceeding with the next step.
5. Place each of the **PowerMag[®] Glass Bead Plates** (with **Sealing Mats** securely affixed) between 2 adapter plates (MO BIO Catalog# 11990) and place on the 96 Well Plate Shaker (MO BIO Catalog# 11996). Reference the protocol provided with the adapter plates for proper placement. Shake at speed 20 for 10 minutes.
6. After the first 10 minute cycle, remove the block and rotate it so that the side closest to the machine body is now furthest from the machine. Shake again at speed 20 for 10 more minutes.
Note: The block needs to be rotated to ensure that bead beating is uniform for all of the wells.
7. Centrifuge the **PowerMag[®] Glass Bead Plate** at room temperature for 6 minutes at 4500 x g.



8. Carefully and without splashing remove and discard the Sealing Mat and transfer the supernatant to a clean **PowerMag[®] 1 ml Collection Plate**.
Note: The supernatant may still contain some bio-solid particles.
9. Add 150 μ l of **PowerMag[®] Inhibitor Removal Solution** to each well and apply Sealing Tape to the **PowerMag[®] 1 ml Collection Plate**. Vortex horizontally for 5 seconds on the vortex ensuring that the solution is well mixed. Incubate at 4°C for 10 minutes.
10. Centrifuge the **PowerMag[®] 1 ml Collection Plate** at room temperature for 6 minutes at 4500 x *g*. Remove and discard Sealing Tape.
11. Avoiding the pellet, transfer the entire volume of supernatant to a new **PowerMag[®] 1 ml Collection Plate**. For the wells at the center of the plate, it may help to mark a line on the pipet tips to show how far to insert the tips without touching the pellets. Apply Sealing Tape to the **PowerMag[®] 1 ml Collection Plate**. Centrifuge again at 4500 x *g* for 6 minutes to clear any residual particulates that may have carried over.
12. Taking care to avoid any residual pellet, transfer no more than **450 μ l** of supernatant from each well to the wells on a clean KingFisher[®] Deep Well 96 Plate.
Note: Any prepared lysates at this point that cannot be processed immediately on the robot should be transferred to and stored in clean KingFisher[®] Deep Well 96 Plates at 4°C for several hours.
13. Open the appropriate protocol on your instrument specific to your platform and then proceed. For the KingFisher[®] Flex go to page 8. For the KingFisher[®] Duo go to page 9.



KingFisher[®] Flex Protocol (continued from step 13)

14. For each 96 well plate to be processed, resuspend the **ClearMag[®] Beads** by vortexing the bottle and add 2 ml of the resuspended **ClearMag[®] Beads** to 45 ml of the **ClearMag[®] Binding Solution** in an appropriate vessel (user provided) and mix well. Immediately transfer to a multi channel reservoir.

Note: As time progresses the **ClearMag[®] Beads/ClearMag[®] Binding Solution** will slowly settle. Maintain the beads in suspension for uniform distribution to each well in the next step.

15. Add 470 µl of the **ClearMag[®] Beads/ClearMag[®] Binding Solution** to each well of lysate in a KingFisher[®] Microtiter Deep Well 96 Plate.
16. Place the KingFisher[®] Microtiter Deep Well 96 Plate containing the lysate and ClearMag[®] Beads/ ClearMag[®] Binding Solution onto the robotic deck at the specified location indicated in the program.
17. Place 500 µl of **ClearMag[®] Wash Solution** into each well of three clean KingFisher[®] Microtiter Deep Well 96 plates and place on the deck at the specified locations indicated in the program.
18. Place 100 µl of **ClearMag[®] RNase-Free Water** into each well of a KingFisher[®] 96 KF plate and place on the deck at the specified location. Initiate the KingFisher[®] MO BIO PowerMag[®] Microbiome robotic program.
19. Upon completion of the robotic program, cover the wells of the KingFisher[®] 96 KF plate with an appropriate storage seal. DNA and RNA are now ready for any downstream application. No further steps are required.

We recommend storing RNA and DNA frozen (-80°C or -20°C).

Thank you for choosing the PowerMag[®] Microbiome RNA/DNA Isolation Kit.



KingFisher[®] Duo Protocol (continued from step 13)

14. Transfer lysate from up to twelve (12) wells to the first long row (A) on a clean KingFisher[®] Microtiter Deep Well 96 Plate.
15. Add 450 µl of the **ClearMag[®] Binding Solution** to each well in row (A) containing lysate.
16. Prepare the **ClearMag[®] Beads** by vortexing the bottle to resuspend. Immediately add 20 µl of the resuspended **ClearMag[®] Beads** to each well containing the lysate/ClearMag[®] Binding Solution mixture.
Note: The beads will slowly settle so it is critical to make sure the beads stay in suspension.
17. Place a KingFisher[®] Duo 12-tip comb into the second row (B) of the KingFisher[®] Microtiter Deep Well 96 Plate.
18. Place 500 µl of **ClearMag[®] Wash Solution** into each well of the next three rows (C, D & E) on the plate and place onto the deck.
19. Place 100 µl of **ClearMag[®] RNase-Free Water** into each well of a KingFisher[®] Duo Elution Strip and place the strip onto the deck.
20. Initiate the KingFisher[®] MO BIO PowerMag[®] Microbiome robotic program.
21. Upon completion of the robotic program, cover the wells of the KingFisher[®] Duo Elution Strip with an appropriate storage seal. DNA and RNA are now ready for any downstream application. No further steps are required.

We recommend storing RNA and DNA frozen (-80°C or -20°C).

Thank you for choosing the PowerMag[®] Microbiome RNA/DNA Isolation Kit.



Hints and Troubleshooting Guide

Amount of Sample to Process

This kit is designed to process 0.25 g of bio-solid or soil. For efficient 96 well homogenization, we do not recommend increasing the amount of sample.

Stabilizing Samples for Storage and During Processing

Addition of 100 μ l of phenol:chloroform:isoamyl alcohol (PCI) (25:24:1, pH 6.5-8) to the wells of the bead plate pre-loaded with 650 μ l of PowerMag[®] Microbiome Lysis Solution/ β -ME before filling with stool samples will allow for fast nuclease inactivation and sample stabilization during the filling process and storage at -20°C overnight if desired. If the use of PCI is not desired, pre-loading the wells with PowerMag[®] Microbiome Lysis Solution/ β -ME before adding the samples and then storage overnight in lysis buffer will offer additional protection during the time while samples are sitting in the block at room temperature during filling.

Centrifuge with a Maximum Speed Less Than 4500 x g

Multiply the protocol time and speed to determine the total force (or speed) required (x g). Divide the total by the maximum speed of your centrifuge (round up if necessary). This will be the number of minutes your centrifuge will need to run to achieve the appropriate overall force.

Example: 10 minutes at 4500 x g = 45000.

If your centrifuge has a maximum speed of 2500 x g, divide 45000 \div 2500 = 18 minutes of centrifugation.

If DNA does not PCR amplify

- Check RNA and DNA yield by gel electrophoresis and spectrophotometer reading. DNA template is typically added to 10 ng per reaction, although more or less may be needed depending on the reaction conditions, enzyme activity, and copy number of the target sequence.
- If DNA does not amplify after altering the amount of template in the reaction, PCR optimization (*i.e.* changing reaction conditions, validating primers, or testing a different polymerase) should be attempted.

Concentrating the DNA

The final volume of eluted RNA and DNA will be 100 μ l. Nucleic acids may be concentrated by adding 5 μ l of 5M NaCl and inverting 3-5 times to mix. Next, add 200 μ l of 100% cold ethanol and invert 3-5 times to mix. Incubate at -20°C for at least 10 minutes to overnight. Centrifuge at 13,000 x g for 15 minutes. Decant all liquid. Wash the DNA pellet with 70% cold ethanol. Centrifuge at 13,000 x g for 10 minutes to re-pellet the sample. Decant ethanol and dry in a speed vacuum, desiccator, or ambient air. Resuspend precipitated DNA in sterile water or sterile 10 mM Tris.

Note: This procedure must be done individually after transferring the eluted sample to a microcentrifuge tube.

Storing DNA

The RNA and DNA are eluted in ClearMag[®] RNase-Free Water. Store the RNA/DNA at -20°C to prevent degradation and at -80°C for long term storage. RNA and DNA can be eluted in 10 mM Tris buffer pH 7, or TE without loss, but the EDTA may inhibit downstream reactions such as PCR and automated sequencing. Prolonged storage in the microplates at 4°C will result in the loss of liquid due to evaporation.

MO BIO offers TE-4 (10 mM Tris, 0.1 mM EDTA, pH 8.0) which will allow for maximal protection of DNA during storage with no PCR inhibition (Catalog# 17320-1000).

Technical Information: Toll free 1-800-606-6246, or 1-760-929-9911 Email: technical@mobio.com Website: www.mobio.com



Contact Information

Technical Support:

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For the distributor nearest you, visit our website at www.mobio.com/distributors



Products recommended for you

For a complete list of products available from MO BIO Laboratories, Inc., visit www.mobio.com

Description	Catalog No.	Quantity
PowerMag® Microbial DNA Isolation Kit	27200-4	4 x 96 preps
PowerMag® Soil DNA Isolation Kit (Optimized for KingFisher®)	27000-4-KF	4 x 96 preps
96 Well Plate Shaker	11996	1 unit (120 V)
Plate Adapter Set	11990	1 set
PowerMicrobiome™ RNA Isolation Kit	26000-50	50 preps
PowerFecal® DNA Isolation Kit	12830-50	50 preps

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