

# TwoPaCo

An efficient algorithm to build the compacted de Bruijn graph  
from many complete genomes

Gaspare Ferraro

University of Pisa  
Department of Computer Science

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# Part I

## Introduction



# A pan-genomic algorithm

From the greek word *pan* (everything)

Given a series of complete genomes



# A pan-genomic algorithm

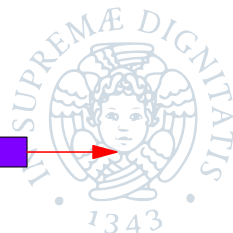
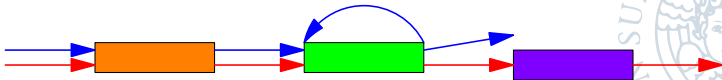
From the greek word *pan* (everything)

Given a series of complete genomes



What we want to see:

- Similarities
- Differences
- Duplications



# de Bruijn graph

Sequence #1: **TGACGTC** (2-mers: **TG GA AC CG GT TC**)



Sequence #2: **TGACTTC** (2-mers: **TG GA AC CT TT TC**)



# de Bruijn graph

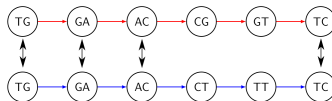
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Find common vertices



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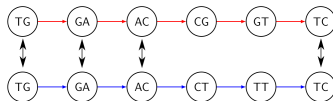
Sequence #1: **TGACGTC** (2-mers: **TG GA AC CG GT TC**)



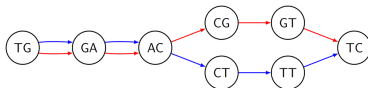
Sequence #2: **TGACTTC** (2-mers: **TG GA AC CT TT TC**)



Find common vertices



Merge them



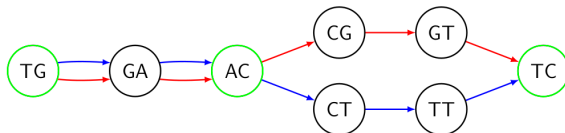
# Compacted de Bruijn graph

To compact the de Bruijn graph we need to find the junctions

A vertex is a junction if :

Is the initial or final part of some sequence

Has more than 1 in-edge or more than 1 out-edge





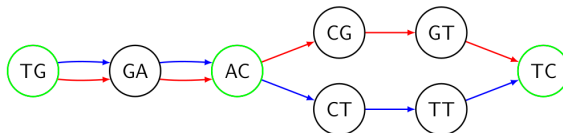
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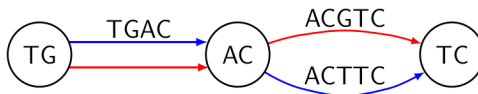
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Remove simple paths by connect junctions



# The problem

How to construct the compacted de Bruijn graph,  
from many complete genomes,  
avoiding the ordinary graph construction?



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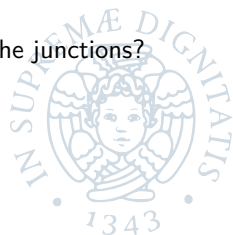
How to construct the compacted de Bruijn graph,  
from many complete genomes,  
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**Which  $k$ -mers are junctions?**



How to construct the compacted de Bruijn graph given the junctions?



# Part II

## The algorithm



# Naive algorithm

- Store all  $(k + 1)$ -mers in a hash table
- For each  $k$ -mers query the possible edges
- If only 1 in and 1 out edge, unmark as a junction

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**Algorithm 1: FILTER-JUNCTIONS**


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**Input** :  $S = \{s_1, \dots, s_m\}$  genome sequences

$k$  integer, size of  $k$ -mers

$E$  empty set data structure

$C$  Candidate set of junctions (naively all positions are marked)

**Output**: A reduce candidate set of junctions  $C$

---

```

1 foreach  $s \in S$  do
2   for  $1 \leq i < |s| - k$  do
3     if  $C[s, i] = \text{marked}$  then
4        $E \leftarrow E \cup s[i..i+k] \cup s[i-1..i+k-1]$   $\triangleright$  Store all  $(k+1)$ -mers

5 foreach  $s \in S$  do
6   for  $1 \leq i < |s| - k$  do
7     if  $C[s, i] = \text{marked}$  then
8        $(in, out) \leftarrow (0, 0)$   $\triangleright$  Count in/out edges
9       foreach  $c \in \{A, C, G, T\}$  do
10        if  $v \cdot c \in E$  then
11           $in \leftarrow in + 1$ 
12        if  $c \cdot v \in E$  then
13           $out \leftarrow out + 1$ 
14        if  $(in, out) = (1, 1)$  then
15           $C[s, i] = \text{unmarked}$   $\triangleright$  Not a junction

16 return  $C$ 

```



# The memory issue

First part of the naive algorithm:

```

foreach  $s \in S$  do
  for  $1 \leq i < |s| - k$  do
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```



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```

We don't really need and, in almost all practical cases,  
we can't store all the possible  $(k + 1)$ -mers.

Mainly because **only a little percentual** of them are  
junction in the de Bruijn graph.



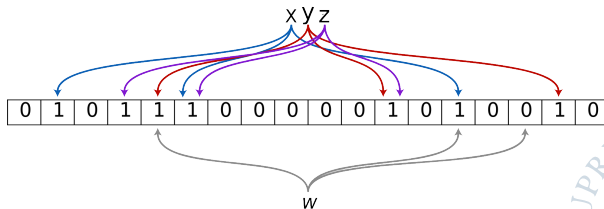
# Bloom filter

A space-efficient probabilistic hash table

Bitmap  $V$  of size  $b$ ,  $h$  hash functions  $f_0, f_1, \dots, f_{h-1} : U \rightarrow [0, b-1]$

insertion( $x$ )  $\rightarrow V[f_i(x)] = 1, \forall 0 \leq i < h$

contains( $x$ )  $\rightarrow$  **probably yes** if  $V[f_i(x)] = 1, \forall 0 \leq i < h$



Probability of false positive, after  $n$  insertion:  $p_{FP} \simeq (1 - e^{-hn/m})^h$



## Two Pass version

- First pass: Select a set of junction candidates by insert all the  $(k + 1)$ -mers in a bloom filter of choosing size
- Second pass: Filter out the false positive by storing the reduce sets of  $(k + 1)$ -mers in an hash table

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### Algorithm 2: FILTER-JUNCTIONS-TWO-PASS

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**Input** : strings  $S = \{s_1, \dots, s_m\}$  genome sequences  
 integer  $k$ , size of  $k$ -mers  
 integer  $b$ , size of bloom filter

Candidate set of junctions  $C_{in}$  (naively all positions are marked)

**Output:** A reduce candidate set of junctions  $C_{out}$

- $F \leftarrow$  empty bloom filter of size  $b$  ▷ First pass
  - $C_{temp} \leftarrow \text{FILTER-JUNCTIONS}(S, k, F, C_{in})$
  - $H \leftarrow$  empty hash table ▷ Second pass
  - $C_{out} \leftarrow \text{FILTER-JUNCTIONS}(S, k, H, C_{in})$
  - return**  $C_{out}$
-

# The memory issue<sup>2</sup>

How much memory do we use now?

- First pass: Bloom filter of size  $b$  (of our decision)
- Second pass: Hash table containing  $(k + 1)$ -mers of junction candidates

We don't know the possible size of the hash table in the second pass.  
What if the hash table is not small enough?



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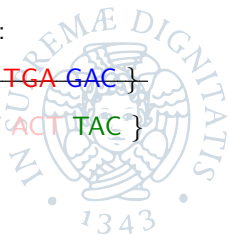
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What if the hash table is not small enough?

**Solution:** Split the input  $k$ -mers in chunks and analyze them in multiple rounds.

Split *TGACTACTGAC* in 3 chunks of 3-mers:

~~Wrong: { TGA GAC ACT }, { CTA TAC ACT }, { CTG TGA GAC }~~

Correct: { TGA TGA CTG }, { GAC GAC CTA }, { ACT ACT TAC }



# $k$ -mers splitting

- Count how many  $k$ -mers are mapped in each of the  $c$  hash value ( $c = 2^l$ )
- Iterate over all the hash value and insert them in the current chunk
- If the size of current chunks exceeds the average pass to the next chunk

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**Algorithm 3: ROUND-SPLITTING**


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**Input** : strings  $S = \{s_1, \dots, s_m\}$  genome sequences

integer  $k$ , size of  $k$ -mers

integer  $b$ , size of bloom filter

integer  $l$ , number of rounds

function  $f(x)$ , from  $k$ -mers to integers

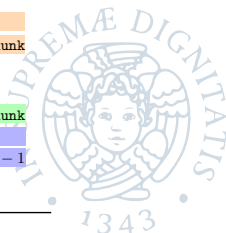
**Output:**  $(V_0, V_1, \dots, V_{l-1})$  chunks of  $k$ -mers

```

1  $V_0 \leftarrow \emptyset, V_1 \leftarrow \emptyset, \dots, V_{l-1} \leftarrow \emptyset$  ▷ Chunks, initially empty
2  $c_0 \leftarrow 0, c_1 \leftarrow 0, \dots, c_{q-1} \leftarrow 0$  ▷ Counters, for balancing  $k$ -mers
3  $c_i \leftarrow |\{ \textit{k-mer } x \mid f(x) = i \}|$ 
4  $T \leftarrow \sum_{0 \leq t < q} c_t / l$  ▷ Average size of a chunk
5  $idx \leftarrow 0$ 
6 for  $0 \leq i < q$  do
7    $V_{idx} = V_{idx} \cup \{i\}$  ▷ Add  $i$  to the current chunk
8   if  $|V_{idx}| \geq T$  then
9      $idx \leftarrow \min(idx + 1, l - 1)$  ▷ Next chunk, until reach  $l - 1$ 
10 return  $(V_0, V_1, \dots, V_{l-1})$ 

```

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# Multiple rounds: dealing with memory restrictions

- Partitionate the input with ROUND-SPLITTING
- Analyse each partition with FILTER-JUNCTIONS-TWO-PASS
- Merge the results of each rounds and output the real junctions

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## Algorithm 4: TWOpaCo

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**Input** : strings  $S = \{s_1, \dots, s_m\}$  genoma sequences

integer  $k$ , size of  $k$ -mers

integer  $b$ , size of bloom filter

integer  $l$ , number of rounds

function  $f(x)$ , from  $k$ -mers to integers

**Output:**  $C_{final}$  all the junctions in the compacted de Bruijn graph

```

1  $(V_0, V_1, \dots, V_{l-1}) \leftarrow \text{ROUND-SPLITTING}(S, k, b, l, f)$  ▷ Create chunks
2  $C_{init} \leftarrow$  Boolean array with every position unmarked
3 for  $0 \leq i < l$  ▷ For each chunks
4 do
5    $C_i \leftarrow$  mark every position of  $C_{init}$  that starts a  $k$ -mer with hash in  $V_i$ 
6    $C_i \leftarrow \text{FILTER-JUNCTIONS-TWO-PASS}(S, k, b, C_i)$  ▷ Find junctions
7  $C_{final} = \bigcup C_i$  ▷ Merge junctions of each chunks
8 return  $C_{final}$ 

```

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TwoPaCo can be easily parallelizable:

- Parallel for loops (OpenMP, TBB, ...)
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We made a clever usage of the concurrent data structs:

- Only insertion in the first part
- Only query in the second part (static table, no race conditions)





# Part III

## Results



# Source code & Dataset

Original source code by medvedev group available on:

<https://github.com/medvedevgroup/TwoPaCo>

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Personal implementation and presentation available on:

<https://github.com/GaspereG/TwoPaCo>

<https://github.com/GaspereG/TwoPaCoPresentation>

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Dataset for experiments:

- 62 Escherichia coli (~300Mb)
- 5 humans (~21Gb)
- 8 primates (~23Gb)
- 100 simulated human (~400Gb)



# Memory complexity

The memory complexity is the maximum among the first and the second pass of TwoPaCo

- First pass: insert all  $k$ -mers in a bloom filter of **size**  $b$
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- Real junction,  $J$
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Result:  $\mathcal{O}(\max\{b, (J + FP)k\})$



# Time complexity

The time complexity is the sum  
between the first and the second pass of TwoPaCo

- First pass: **insert all  $k$ -mers** in a bloom filter using  $h$  hash functions
- Second pass: iterate over all **candidate positions** and query the hash table



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$\mathcal{O}(m)$ , where  $m = \sum_{s \in S} |s|$  is the total input size



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How many candidate positions?

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- False positive induced from the bloom filter,  $FP$

Result:  $\mathcal{O}(mh + (|G_c| + FP)k)$



# Complexity comparison

State of the art for compacted de Bruijn graph construction:

- Sibelia (Minkin, Patel, Kolmogorov, Vyahhi, Pham, 2013)
- SplitMEM (Marcus, Lee, Schatz, 2014)
- bwt-based (Baier, Beller, Ohlebusch, 2015)



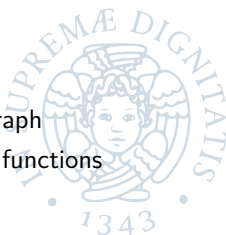
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Algorithm	Time complexity	Memory complexity
Sibelia	$\mathcal{O}(m)$	$\mathcal{O}(m)$
SplitMEM	$\mathcal{O}(m \log g)$	$\mathcal{O}( G_c  + m)$
bwt-based	$\mathcal{O}(m)$	$\mathcal{O}(m)$
TwoPaCo	$\mathcal{O}(mh + ( G_c  + FP)k)$	$\mathcal{O}(\max\{b, (J + FP)k\})$

- $m = \sum_{s \in S} |s|$ , total input size
- $g = \max_{s \in S} |s|$ , size of the biggest genoma
- $J$  and  $|G_c|$ , number of vertex and edge in the de Bruijn Graph
- $b$  and  $h$ , size of the bloom filter table and number of hash functions
- $FP$ , number of false positives in first pass



# Benchmark

What are the practical performances of TwoPaCo against the state of the art?



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Dataset	Sibelia	SplitMEM	bwt-based	TwoPaCo	
				1 thread	15 thread
62 E.coli (k=25)	10 (12.2)	70 (178.0)	8 (0.85)	4 (0.16)	2 (0.39)
62 E.coli (k=100)	8 (7.6)	67 (178.0)	8 (0.50)	4 (0.19)	2 (0.39)
7 humans (k=25)	-	-	867 (100.30)	436 (4.40)	63 (4.84)
7 humans (k=100)	-	-	807 (46.02)	317 (8.42)	57 (8.75)
8 primates (k=25)	-	-	-	914 (34.36)	111 (34.36)
8 primates (k=100)	-	-	-	756 (56.06)	101 (61.68)
50 humans (k=25)	-	-	-	-	705 (69.77)
50 humans (k=100)	-	-	-	-	927 (70.21)

Running times in minutes and memory usage in gigabytes in parenthesis

- - on Sibelia = out of time
- - on SplitMEM = out of memory
- - on bwt-based = out of memory
- - on TwoPaCo = experiment not done



# Fixed memory

In how many rounds do we need to split the input to not exceeding a given memory threshold?



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Threshold	Used memory	Bloom filter size	Running time	Rounds
10GB	8.62GB	8.59GB ( $2^{33}$ )	4h	1
8GB	6.73GB	4.29GB ( $2^{32}$ )	7h	3
6GB	5.98GB	4.29GB ( $2^{32}$ )	9h	4
4GB	3.51GB	2.14GB ( $2^{31}$ )	11h	6

Experiments on 5 simulated human genomes,  $k = 25$ , 8 threads



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Result: **we can trade-off memory for time**





# Bloom Filter false positive

Is the Bloom Filter really efficient in reducing junction candidates?



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Is the Bloom Filter really efficient in reducing junction candidates?

Dataset	k	Junction candidates		
		Initial	First pass	Second pass
62 E.coli	25	310 157 564 (100%)	24 649 489 (7.94%)	24 572 562 (7.92%)
62 E.coli	100	310 157 489 (100%)	22 848 018 (7.36%)	9 492 091 (3.06%)
7 humans	25	21 201 290 922 (100%)	3 489 946 013 (16.46%)	2 974 098 154 (14.02%)
7 humans	100	21 201 290 847 (100%)	1 374 287 870 (6.48%)	188 224 214 (0.88%)
8 primates	25	24 540 556 921 (100%)	5 423 003 377 (22.09%)	5 401 587 503 (22.01%)
8 primates	100	24 540 556 846 (100%)	1 174 160 336 (4.78%)	502 441 107 (2.04%)

- Initial: total number of  $k$ -mers in dataset
- First pass: number of junction candidates (using a bloom filter)
- Second pass: real number of junction (using an hash table)



# Conclusion

- Store in main memory only the necessary
- Lowest time/memory usage in the state of the art
- Highly scalable (best performance with multi-thread CPUs)
- Time/memory trade-off (suitable for small computer)
- Bloom filter perform very well in practice



# References

[1] Minkin, I., Pham, S., Medvedev, P. (2016).

**TwoPaCo**: An efficient algorithm to build the compacted de Bruijn graph from many complete genomes

[2] Minkin, I., Patel, A., Kolmogorov, M., Vyahhi, N., Pham, S. (2013).

**Sibelia**: a scalable and comprehensive synteny block generation tool for closely related microbial genomes.

[3] Marcus, S., Lee, H., Schatz, M. C. (2014).

**SplitMEM**: a graphical algorithm for pan-genome analysis with suffix skips.

[4] Baier, U., Beller, T., Ohlebusch, E. (2015).

Graphical pan-genome analysis with compressed suffix trees and the **Burrows-Wheeler transform**.



# Questions

Thanks for your attention!

Questions?



# My results

Source code:

<https://github.com/GaspereG/TwoPaCo/blob/master/srcgas/twopaco.cpp>

Dataset	k	Version	Time	Initial	First Pass	Second Pass
2 E.coli	25	BF+HT	18s	10.207.478	392.836	177.396
2 E.coli	25	BF	29s	10.207.478	-	177.396
4 E.coli	25	BF+HT	38s	20.301.208	1.261.345	676.686
4 E.coli	25	BF	64s	20.301.208	-	676.686
8 E.coli	25	BF+HT	87s	39.871.914	3.231.119	1.822.718
8 E.coli	25	BF	134s	39.871.914	-	1.822.718
16 E.coli	25	BF+HT	160s	77.848.026	7.076.460	4.046.561
16 E.coli	25	BF	280s	77.848.026	-	4.046.561

Running time and junctions candidate after each pass