TwoPaCo

An efficient algorithm to build the compacted de Bruijn graph from many complete genomes

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TwoPaCo



Parte I

Introduction



A pan-genomic algorithm

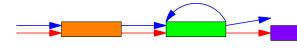
From the greek word *pan* (everything)

Given a series of complete genomes



What we want to see:

- Similarities
- Differences
- Duplications





de Bruijn graph

Sequence #1: TGACGTC (2-mers: TG GA AC CG GT TC)



Sequence #2: TGACGTC (2-mers: TG GA AC CT TT TC)



Find common vertices



Merge them

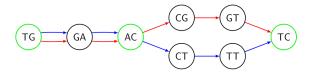




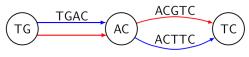
Compacted de Bruijn graph

To compact the de Bruijn graph we need to find the junctions

A vertex is a junction if : Is the initial or final part of some sequence Has more than 1 in-edge or more than 1 out-edge



Remove simple paths by connect junctions



The problem

How to construct the compacted de Bruijn graph, from many complete genomes, avoiding the ordinary graph construction?

 \downarrow

Which *k*-mers are junctions?

+

How to construct the compacted de Bruijn graph given the junctions?

Parte II

The algorithm



Naive algorithm

- Store all (k+1)-mers in a hash table
- For each k-mers query the possible edges
- If only 1 in and 1 out edge, unmark as a junction

```
Algorithm 1: FILTER-JUNCTIONS
  Input : S = \{s_1, ..., s_m\} genoma sequences
           k integer, size of k-mers
            E empty set data structure
           C Candidate set of junctions (naively all positions are marked)
  Output: A reduce candidate set of junctions C
1 foreach s \in S do
      for 1 \le i < |s| - k do
          if C[s,i] = marked then
              E \leftarrow E \cup s[i..i+k] \cup s[i-1..i+k-1] \triangleright Store all (k+1)-mers
5 foreach s \in S do
      for 1 \le i < |s| - k do
          if C[s, i] = marked then
              (in, out) \leftarrow (0, 0)
                                                               ▷ Count in/out edges
              foreach c \in \{A, C, G, T\} do
                  if v \cdot c \in E then
10
11
                  if c \cdot v \in E then
12
                      out \leftarrow out + 1
13
              if (in, out) = (1, 1) then
14
                  C[s, i] = unmarked
                                                           15
16 return C
```

The memory issue

First part of the naive algorithm:

$$\begin{array}{c|c} \text{for each } s \in S \text{ do} \\ & \text{for } 1 \leq i < |s| - k \text{ do} \\ & \text{if } C[s,i] = marked \text{ then} \\ & \text{} L \leftarrow E \cup s[i..i+k] \cup s[i-1..i+k-1] \end{array}$$

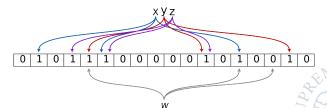
We don't really need and, in almost all pratical cases, we can't store all the possible (k+1)-mers.

Mainly because **only a little percentual** of them are junction in the de Bruijn graph.

Bloom filter

A space-efficient probabilistic hash table

Bitmap
$$V$$
 of size b , h hash functions $f_0, f_1, ..., f_{h-1}: U \to [0, b-1]$ insertion(x) $\to V[f_i(x)] = 1$, $\forall \ 0 \le i < h$ contains(x) \to probabily yes if $V[f_i(x)] == 1$, $\forall \ 0 \le i < h$



Probability of false positive, after n insertion: $p_{FP} \simeq (1-e^{-hn/m})^{\hbar}$

Two Pass version

- First pass: Select a set of junction candidates by insert all the (k+1)-mers in a bloom filter of choosing size
- Second pass: Filter out the false positive by storing the reduce sets of (k+1)-mers in an hash table

Algorithm 2: FILTER-JUNCTIONS-TWO-PASS Input : strings $S = \{s_1, ..., s_m\}$ genoma sequences integer k, size of k-mers integer b, size of bloom filter Candidate set of junctions C_{in} (naively all positions are marked) Output: A reduce candidate set of junctions C_{out} 1 $F \leftarrow$ empty bloom filter of size b ▷ First pass 2 $C_{temp} \leftarrow$ FILTER-JUNCTIONS (S, k, F, C_{in}) 3 $H \leftarrow$ empty hash table ▷ Second pass 4 $C_{out} \leftarrow$ FILTER-JUNCTIONS (S, k, H, C_{in}) 5 return C_{out}

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4 D > 4 A > 4 B > 4

The memory issue²

How much memory do we use now?

- First pass: Bloom filter of size b (of our decision)
- Second pass: Hash table containing (k+1)-mers of junction candidates

We don't know the possible size of the hash table in the second pass. What if the hash table is not small enough?

Solution: Split the input k-mers in chunks and analyze them in multiple rounds.

Split TGACTACTGAC in 3 chunks of 3-mers:

```
Wrong: { TGA GAC ACT }, { CTA TAC ACT }, { CTG TGA GAC }
```

Correct: { TGA TGA CTG }, { GAC GAC CTA }, { ACT ACT TAC

k-mers splitting

Algorithm 3: ROUND-SPLITTING

```
Input : strings S = \{s_1, ..., s_m\} genoma sequences
              integer k, size of k-mers
              integer b, size of bloom filter
              integer l, number of rounds
              function f(x), from k-mers to integers
   Output: (V_0, V_1, \dots, V_{l-1}) chunks of k-mers
1 V_0 \leftarrow \emptyset, V_1 \leftarrow \emptyset, \dots, V_{l-1} \leftarrow \emptyset
                                                                  D Chunks, initially empty
c_0 \leftarrow 0, c_1 \leftarrow 0, \dots, c_{\sigma-1} \leftarrow 0
                                                        ▷ Counters, for balancing k-mers
3 F \leftarrow \text{empty Bloom filter of size } b
4 foreach s \in S do
        for 1 \le i < |s| - k do
            if s[i..i+k-1] not in F then
          9 T \leftarrow \sum_{0 \le t \le a} c_t/l
                                                                  D Average size of a chunk
10 acc \leftarrow 0, idx \leftarrow 0
                                                          D Create nearly balanced chunks
11 for 0 \le i \le q do
      V_{idx} = V_{idx} \cup \{i\}
   acc \leftarrow acc + c_i
    if acc > T then
            acc \leftarrow 0
15
            idx \leftarrow \min(idx+1,l-1)
                                                           \triangleright Next chunk, until reach l-1
17 return (V_0, V_1, \dots, V_{l-1})
                                                                             4 □ > 4 □ > 4 ≡ > 4
```

Multiple rounds: dealing with memory restrictions

- Partitionate the input with ROUND-SPLITTING
- Analyse each partition with FILTER-JUNCTIONS-TWO-PASS
- Merge the results of each rounds and output the real junctions

```
Algorithm 4: TwoPaCo
  Input: strings S = \{s_1, ..., s_m\} genoma sequences
            integer k, size of k-mers
            integer b, size of bloom filter
            integer l, number of rounds
            function f(x), from k-mers to integers
  Output: C_{final} all the junctions in the compacted de Bruijn graph
1 (V_0, V_1, \dots, V_{l-1}) \leftarrow \text{ROUND-SPLITTING}(S, k, b, l, f)
                                                                              D Create chunks
2 C_{init} \leftarrow \text{Boolean array with every position unmarked}
3 for 0 \le i < l
                                                                           > For each chunks
   dο
       C_i \leftarrow \text{mark every position of } C_{init} \text{ that starts a } k\text{-mer with hash in } V_i
       C_i \leftarrow \text{Filter-Junctions-Two-Pass}(S, k, b, C_i)
                                                                            ▷ Find junctions
\overline{C}_{final} = \bigcup C_i
                                                      ▷ Merge junctions of each chunks
8 return C_{final}
```

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Parallelization scheme

As far we focused in reducing memory. What about the time?

TwoPaCo can be easily parallelizable:

- Parallel for loops (OpenMP, TBB, ...)
- Concurrent bloom filter (parallel insert and search)
- Concurrent hash table (parallel insert and search)

We made a clever usage of the concurrent data structs:

- Only insertion in the first part
- Only query in the second part (static table, no race conditions)

Parte III

Results



Source code & Dataset

Original source code by medvedev group available on:

https://github.com/medvedevgroup/TwoPaCo

Personal implementation and presentation available on:

https://github.com/GaspareG/TwoPaCo

https://github.com/GaspareG/TwoPaCoPresentation

Dataset for experiments:

62 Escherichia coli (~300Mb)
5 humans (~21Gb)
8 primates (~23Gb)
100 simulated human (~400Gb)



Memory complexity

The memory complexity is the maximum among the first and the second pass of TwoPaCo

- First pass: insert all k-mers in a bloom filter of size b
- Second pass: store all junction candidates in a hash table

How many junction candidates?

- Real junction, J
- False positive induced from the bloom filter, FP

Result: $\mathcal{O}(\max\{b, (J+FP)k\})$



Time complexity

The time complexity is the sum between the first and the second pass of TwoPaCo

- First pass: insert all k-mers in a bloom filter using h hash functions
- Second pass: iterate over all candidate positions and query the hash table

How many
$$k\text{-mers?}$$
 $\mathcal{O}(m),$ where $m=\Sigma_{s\in S}|s|$ is the total input size

How many candidate positions?

- Real positions, $|G_c|$
- False positive induced from the bloom filter, FP

Result: $\mathcal{O}(mh + (|G_c| + FP)k)$



Complexity comparison

State of the art for compressed de Bruijn graph construction:

- Sibelia (Minkin, Patel, Kolmogorov, Vyahhi, Pham, 2013)
- SplitMEM (Marcus, Lee, Schatz, 2014)
- bwt-based (Baier, Beller, Ohlebusch, 2015)

Algorithm	Time complexity	Memory complexity
Sibelia	$\mathcal{O}(m)$	$\mathcal{O}(m)$
SplitMEM	$\mathcal{O}(m \log g)$	$\mathcal{O}(G_c + extbf{m})$
bwt-based	$\mathcal{O}(m)$	$\mathcal{O}(m)$
TwoPaCo	$\mathcal{O}(mh + (G_c + FP)k)$	$\mathcal{O}(\max\{b, (J+FP)k\})$

- $m = \sum_{s \in S} |s|$, total input size
- $q = \max_{s \in S} |s|$, size of the biggest genoma
- J and $|G_c|$, number of vertex and edge in the de Bruijn Graph
- b and h, size of the bloom filter table and number of hash functions
- FP, number of false positives in first pass

TwoPaCo

Running time comparison

What are the practical performances of TwoPaCo against the state of the art?

Dataset	Sibelia	SplitMEM	bwt-based	TwoPaCo	
				1 thread	15 thread
62 E.coli (k=25)	0 (12.2)	70 (178.0)	8 (0.85)	4 (0.16)	2 (0.39)
62 E.coli (k=100)	8 (7.6)	67 (178.0)	8 (0.50)	4 (0.19)	2 (0.39)
7 humans (k=25)	-	-	867 (100.30)	436 (4.40)	63 (4.84)
7 humans (k=100)	-	-	807 (46.02)	317 (8.42)	57 (8.75)
8 primates (k=25)	-	-	-	914 (34.36)	111 (34.36)
8 primates (k=100)	-	-	-	756 (56.06)	101 (61.68)
50 humans (k=25)	-	-	-	-	705 (69.77)
50 humans (k=100)	-	-	-	-	927 (70.21)

Running times in minutes and memory usage in gigabytes in parenthesis

- on Sibelia = out of time
- ullet on SplitMEM = out of memory
- on bwt-based = out of memory
- on TwoPaCo = experiment not done



Fixed memory

In how many rounds do we need to split the input to not exceeding a given memory threshold?

Threshold	Used memory	Bloom filter size	Running time	Rounds
10GB	8.62GB	8.59GB (2 ³³)	4h	1
8GB	6.73GB	$4.29GB(2^{32})$	7h	3
6GB	5.98GB	4.29GB (2^{32})	9h	4
4GB	3.51GB	$2.14GB(2^{31})$	11h	6

Experiments on 5 simulated human genomes, k=25, 8 threads.

Result: we can trade-off memory for time.

Bloom Filter false positive

Is the Bloom Filter really efficient in reducing junction candidates?

		Junction candidates		
Dataset	k	Initial	First pass	Second pass
62 E.coli	25	310 157 564 (100%)	24 649 489 (7.94%)	24 572 562 (7.92%)
62 E.coli	100	310 157 489 (100%)	22 848 018 (7.36%)	9 492 091 (3.06%)
7 humans	25	21 201 290 922 (100%)	3 489 946 013 (16.46%)	2 974 098 154 (14.02%)
7 humans	100	21 201 290 847 (100%)	1 374 287 870 (6.48%)	188 224 214 (0.88%)
8 primates	25	24 540 556 921 (100%)	5 423 003 377 (22.09%)	5 401 587 503 (22.01%)
8 primates	100	24 540 556 846 (100%)	1 174 160 336 (4.78%)	502 441 107 (2.04%)

- Initial: total number of k-mers in dataset
- First pass: number of junction candidates (using a bloom filter)
- Second pass: real number of junction (using an hash table)

References

- [1] Minkin, I., Pham, S., Medvedev, P. (2016).
- **TwoPaCo**: An efficient algorithm to build the compacted de Bruijn graph from many complete genomes
- [2] Minkin, I., Patel, A., Kolmogorov, M., Vyahhi, N., Pham, S. (2013).
- **Sibelia**: a scalable and comprehensive synteny block generation tool for closely related microbial genomes.
- [3] Marcus, S., Lee, H., Schatz, M. C. (2014).
- SplitMEM: a graphical algorithm for pan-genome analysis with suffix skips.
- [4] Baier, U., Beller, T., Ohlebusch, E. (2015).
- Graphical pan-genome analysis with compressed suffix trees and the Burrows-Wheeler transform.

Conclusion

Thanks for your attention!

Questions?

