Microarray_Analysis_Part_1

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```
if (!requireNamespace("BiocManager", quietly = TRUE))
  install.packages("BiocManager")
BiocManager::install(c("GEOquery","oligo","pd.hugene.1.0.st.v1",
                         "hugene10sttranscriptcluster.db"))
install.packages("ggplot2")
install packages
Query Raw Data load packages - GEO Accession No: GSE50397
library(knitr)
library(ggplot2)
library(GEOquery)
extract .cell files to local machine - GEO Series records (GSExxxxx)
gse <- getGEO("GSE50397",GSEMatrix=FALSE)</pre>
view information
- names of all the GSM objects contained in the GSE
print(names(GSMList(gse)))
first GSM object on the list
class(GSMList(gse)[[1]])
head(Meta(GSMList(gse)[[1]]))
names of the GPLs represented
names(GPLList(gse))
access raw data (downloaded file paths)
file_paths = getGEOSuppFiles("GSE50397")
head(file_paths)
choose tar file
tarfile <- file.choose()</pre>
extract tar archives
untar(tarfile, exdir="RawData_Files")
list of all gz files in the directory
```

```
cel.files <- list.files("RawData_Files/", pattern = "[gz]")</pre>
length(cel.files)
list/vector/dataframe —> vector/matrix extract gz archives - gunzip
sapply(paste("RawData_Files", cel.files, sep="/"), gunzip)
Data Preprocessing load packages
library(oligo)
list of all cel files in the directory
celpath <- "~/Documents/Learning/Bioinformatics/MicroArray Analysis/Nordic Islet Analysis/Raw_Data"</pre>
celfiles_list <- list.files(celpath,pattern = ".CEL", full.names=TRUE)</pre>
length(celfiles_list)
head(celfiles_list)
import CEL files containing raw probe-level data into an R AffyBatch object
cell_files <- read.celfiles(celfiles_list)</pre>
head(cell_files)
load packages
library(pd.hugene.1.0.st.v1)
??pd.hugene.1.0.st.v1
getClass("GeneFeatureSet")
max expression
max(exprs(cell_files[1:1102489,1:89]))
replace sampleNames
filename <- sampleNames(cell_files)</pre>
pData(cell_files)$filename <- filename</pre>
pData(cell_files)$filename
sampleNames <- sub("-islet.CEL$","",filename)</pre>
sampleNames <- sub("_HTL[[:digit:]]*","",sampleNames)</pre>
sampleNames(cell_files) <- sampleNames</pre>
sampleNames(cell_files)
information on variable values/ meta-data
pData(cell_files)
boxplot before RMA normalization
boxplot(cell_files, target="probeset")
mtext(text="log2 Intensity", side=2, line=3, las=0)
histogram before RMA normalization
hist(cell_files,target="probeset")
mtext(text="Samples", side=1, line=3, las=1)
```

perform RMA normalization (Robust Multi-Array Average) - converts an AffyBatch object into an ExpressionSet object

```
normData <- rma(cell_files)</pre>
nrow(normData)
boxplot after RMA normalization
boxplot(normData, target="probeset")
mtext(text="log2 Intensity", side=2, line=3, las=0)
mtext(text="Samples", side=1, line=3, las=1)
histogram after RMA normalization
hist(normData,target="probeset")
save the expression data (output - normalized and log2 transformed)
exprs(normData)[1:3,1:5]
write.exprs(normData,file="RMA_Normalised_Original.txt")
load packages
library(Biobase)
library(hugene10sttranscriptcluster.db)
??Biobase
??hugene10sttranscriptcluster.db
gene annotation - get a list of retrievable data
keytypes(hugene10sttranscriptcluster.db)
retrieve data for selected objects (ENTREZID and SYMBOL) as a data frame
anno<- select(hugene10sttranscriptcluster.db,keys(hugene10sttranscriptcluster.db),</pre>
              c("ENTREZID", "SYMBOL"))
head(anno)
tail(anno)
optional - to keep one match per gene
anno <- anno[!duplicated(anno[,1]),]</pre>
tail(anno)
set row names to ProbeID (for convenience)
anno = anno[,-1]
row.names(anno) = keys(hugene10sttranscriptcluster.db)
tail(anno)
retrieve gene expression matrix from normData as a dataframe
expr <- data.frame(exprs(normData))</pre>
head(expr)
merge gene expression and annotation according to row names (probe IDs)
expr_anno <- merge(x=anno,y=expr,by.y=0, by.x=2,all=TRUE)
head(expr_anno)
```

save the annotated gene expression matrix to local file