

Package ‘HeritSeq’

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Title Heritability of Gene Expression for Next-Generation Sequencing

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Description Statistical framework to analyze heritability of gene expression based on next-generation sequencing data and simulating sequencing reads. Variance partition coefficients (VPC) are computed using linear mixed effects and generalized linear mixed effects models. Compound Poisson and negative binomial models are included.

Depends R (>= 3.2.3)

License GPL-2

Encoding UTF-8

LazyData true

Imports cplm, DESeq2, lme4, pbapply, tweedie, MASS,
SummarizedExperiment

Suggests glmmADMB

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NeedsCompilation no

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computeVPC.CP	<i>Calculate the compound Poisson (CP) variance partition coefficient (VPC) for one or more features.</i>
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Description

Calculate the CP VPC for one or more features following the model fitting function `fit.CP()`.

Usage

```
computeVPC.CP(para)
```

Arguments

`para` A $G \times 4$ matrix of CP fit parameters for G features, $G \geq 1$. The column order is intercept α_g , random effect σ_g^2 ($\sigma_g^2 \geq 0$), tweedie parameter p_g ($1 < p_g < 2$), and dispersion ϕ ($\phi > 0$).

Value

A $G \times 1$ matrix consisting of VPC for G features based on compound Poisson mixed models. Column name is "CP-fit"; row names are the feature names.

Examples

```
## Compute VPC for each feature under compound Poisson mixed models.
vpc.cp <- computeVPC.CP(para_cp)

## Visualize the distribution of the VPCs.
hist(vpc.cp, breaks = 50, col = "cyan")

## Plot sorted VPCs.
plot(sort(vpc.cp), ylab = "Heritability (h2)", ylim = c(0,1), main = "Sorted CP VPC scores")
abline(h = 0.9, lty = 2, col = "red")
text(50, 0.92, "h2 = 0.9", col = "red")
```

computeVPC.NB	<i>Calculate the negative binomial (NB) variance partition coefficient (VPC) for one or more features.</i>
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Description

Calculate the NB VPC for one or more features following the model fitting function fit.NB().

Usage

```
computeVPC.NB(para)
```

Arguments

para	A $G \times 3$ matrix of negative binomial fit parameters for G features, $G \geq 1$. The column order is intercept α_g , random effect σ_g^2 ($\sigma_g^2 \geq 0$), and dispersion ϕ ($\phi > 0$).
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Value

A $G \times 1$ matrix consisting of VPC for G features based on negative binomial mixed model. Column name is "NB-fit"; row names are the feature names.

Examples

```
## Compute VPC for each feature under negative binomial mixed model.
vpc.nb <- computeVPC.NB(para_nb)

## Visualize the distribution of the VPCs.
hist(vpc.nb, breaks = 50, col = "cyan")

## Plot sorted VPCs.
plot(sort(vpc.nb), ylab = "Heritability (h2)", ylim = c(0,1),
main = "Sorted NB VPC scores")
abline(h = 0.9, lty = 2, col = "red")
text(50, 0.92, "h2 = 0.9", col = "red")
```

fit.CP	<i>Fit compound Poisson mixed effect models (CPMM) for one or more features.</i>
--------	--

Description

Fit a CPMM for one or more features and output the fit parameters. It is used before the function computeVPC.CP(). This function also allows to test the presence of heritability via random effect variance of the model.

Usage

```
fit.CP(CountMatrix, Strains, test = FALSE, optimizer = "nllminb")
```

Arguments

CountMatrix	Sequencing count matrix for one or more features. Each row is for one feature, and the columns are for samples.
Strains	Strain labels for the samples.
test	TRUE or FALSE (default). Test the presence of heritability through examining the random effect variance $\sigma_g^2 = 0$.
optimizer	A character string that determines which optimization routine is to be used. Possible choices are "nlminb" (default), "L-BFGS-B", and "bobyqa".

Value

A list with two objects. The first object is a $G \times 4$ matrix indicating the fitted parameters for each feature. The columns are ordered by intercept α_g , tweedie parameter p_g , random effect variance σ_g^2 , and dispersion ϕ_g . Row names are feature names. If the argument test is set to be true, the second object of the list consists of p-values for testing the hypothesis that random effects $\sigma_a^2 = 0$; otherwise, the second object is NULL.

Examples

```
## Fit CPMM for the first two features and test the presence of
## heritability.
result.cp <- fit.CP(simData[1:2, ], strains, test = TRUE)
## Extract parameters
para.cp <- result.cp[[1]]
## Extract p-values
pval.cp <- result.cp[[2]]
```

fit.NB

*Fit negative binomial mixed models (NBMM) for one or more features.***Description**

Fit NBMM for one or more features and output the fit parameters. It is used before the function computeVPC.NB(). This function also allows to test the presence of heritability via random effect variance of the model. To fit a NBMM, the glmmADMB package is needed.

Usage

```
fit.NB(CountMatrix, Strains, test = FALSE)
```

Arguments

CountMatrix	Sequencing count matrix for a list of features. Each row is for one feature, and the columns are for samples.
Strains	Strain labels for the samples.
test	TRUE or FALSE (default). Test the presence of heritability through examining the random effect variance $\sigma_g^2 = 0$.

Value

A list with two objects. The first object is a $G \times 3$ matrix indicating the fitted parameters for each feature. The columns are ordered by $\alpha_g, \sigma_g^2, \phi_g$. Row names are feature names. If the argument `test` is set to be true, the second object of the list consists of p-values for testing the hypothesis that random effects $\sigma_a^2 = 0$; otherwise, the second object is NULL.

Examples

```
## Compute vpc for each feature under NBMM. This will take a while on the
## entire dataset. For the purpose of illustration, here we only fit on
## the first 2 features.
library(glmmADMB)
result.nb <- fit.NB(simData[1:2, ], strains)
```

fitComputeVPC.lmer	<i>Fit linear mixed models (LMM) and compute the VPC values for one or more features.</i>
--------------------	---

Description

Fit the Gaussian-like data to LMM and compute the VPC values for one or more features.

Usage

```
fitComputeVPC.lmer(CountMatrix, Strains, PriorWeights = NULL, test = FALSE,
  VPCname = "LMM")
```

Arguments

CountMatrix	Sequencing count matrix for one or more features. Each row is for one feature, and the columns are for samples.
Strains	Strain labels for the samples.
PriorWeights	Weights used in the lmer function in the package lme4. It is an optional vector used in the fitting process.
test	TRUE or FALSE (default). Test the presence of heritability through examining the random effect variance $\sigma_g^2 = 0$.
VPCname	Name of the VPC result, default = "LMM".

Value

A list with two objects. The first object is a $1 \times G$ vector indicating the variance partition coefficients (VPC). If the argument `test` is set to be true, the second object of the list consists of p-values for testing the hypothesis that random effects $\sigma_a^2 = 0$; otherwise, the second object is NULL.

Examples

```
## Compute VPC for each feature under linear mixed models for Gaussian-like datasets.

## Provide normalized data and include hypothesis testing on presence of
## heritability:
result.vst <- fitComputeVPC.lmer(simData_vst, strains, test = TRUE)
## Extract parameters
vpc.vst <- result.vst[[1]]
## Extract p-values
pval.vst <- result.vst[[2]]

## Visulize the distribution of p-values.
hist(pval.vst, breaks = 30, col = "cyan")
```

getBootCI	<i>Compute variance partition coefficient (VPC) confidence intervals (CI) for one or more features.</i>
-----------	---

Description

Compute VPC CI based on parametric bootstrap for one or more features.

Usage

```
getBootCI(CountMatrix, Strains, which.features, num.boot, method = "NB-fit",
  alpha = 0.05, optimizer = "nlminb")
```

Arguments

CountMatrix	A $G \times N$ count matrix. G is the number of features; N is the total number of samples.
Strains	A $1 \times N$ vector of strain labels corresponding to each sample.
which.features	A $1 \times k$ vector of select feature numbers for which CI is desired. $k \leq G$.
num.boot	Number of bootstraps.
method	Which method should be used, "CP-fit", "NB-fit" (default), or "VST". "VST" method bootstraps data under negative binomial mixed models.
alpha	A numerical value between 0 and 1, indicating the significance level of the CI. The CI will be $100 * (1 - \alpha)$ percent CI. Default value is 0.05.
optimizer	A character string that determines which optimization routine is to be used. It is only used for method = "CP-fit". Possible choices are "nlminb" (default), "L-BFGS-B", and "bobyqa".

Value

A list of two objects. The first object is a $k \times 2$ matrix containing the CI. The second object consists of a $k \times \text{num.boot}$ matrix of all bootstrapped VPC values.

Examples

```
## Compute CI based on 100 bootstrap samples for the first feature
## under NBMM. It takes a few minutes.

NBboot <- getBootCI(simData, strains, 1, 100)
## Extract CI
NBboot.ci <- NBboot[[1]]
## Extract vpcs
NBboot.vpc <- NBboot[[2]]

## Compute CI based on 100 bootstrap samples for the first feature
## under vst.
VSTboot <- getBootCI(simData, strains, 1, 100, method = "VST")
```

getReadMatrix.CP	<i>Simulate a read matrix from compound Poisson mixed effect models (CPMM).</i>
------------------	---

Description

Simulate a (possibly unbalanced) read matrix from CPMM. For a compound Poisson (CP) random variable Y_{gsr} with mean μ_{gs} , its variance can be expressed as $\phi_g \mu_{gs}^{p_g}$, for some $1 < p_g < 2$. Under the CPMM, with a log-link, the regression on the mean has the form:
 $\log(\mu_{gs}) = \alpha_g + b_{gs}$, $b_{gs} \sim N(0, \sigma_g^2)$.

Usage

```
getReadMatrix.CP(vec.num.rep, alphas, sigma2s, ps, phis)
```

Arguments

vec.num.rep	A vector of replicate numbers for each strain.
alphas	Intercept vector α_g 's, $1 \times \text{num.features}$.
sigma2s	Random effect variance vector σ_g^2 's, $1 \times \text{num.features}$.
ps	Tweedie parameter in CP models, p_g 's, a $1 \times \text{num.features}$ vector.
phis	Dispersion parameter in CP models, ϕ_g 's, a $1 \times \text{num.features}$ vector.

Value

A $G \times N$ matrix with CP reads. N is the total number of samples; G is the number of features. Column names are sample names of the form "Ss_r", where S stands for sample, s is the strain number, r is the replicate number within the strain. Row names are the feature names of the form "Gene g", where g is the feature index.

Examples

```
## Generate a sequencing dataset with 5 features and 6 strains.
## Assign parameter values.
rep.num <- c(3, 5, 2, 3, 4, 2)
a0s <- c(-1, 1, 2, 5, 10)
sig2s <- c(10, 0.2, 0.1, 0.03, 0.01)
ps <- rep(1.5, 5)
phis <- c(1.5, 1, 0.5, 0.1, 0.1)

set.seed(1234)
## Generate reads:
cpData <- getReadMatrix.CP(rep.num, a0s, sig2s, ps, phis)
## Generate strain names:
str <- sapply(1:length(rep.num), function(x){
  str.x <- paste0("S", x)
  return(rep(str.x, rep.num[x]))
})
str <- do.call(c, str)
```

getReadMatrix.NB	<i>Simulate a count matrix from negative binomial mixed effect models (NBMM).</i>
------------------	---

Description

Simulate a (possibly unbalanced) count matrix from NBMM. Under NBMM, an observed number of reads aligned to feature/gene g , Y_{gsr} , follows a negative binomial (NB) distribution with mean μ_{gs} and variance $\mu_{gs} + \phi_g \mu_{gs}^2$, where ϕ_g is the dispersion parameter, shared across strains. The generalized linear model uses a log-link:
 $\log(\mu_{gs}) = \alpha_g + b_{gs}$, $b_{gs} \sim N(0, \sigma_g^2)$.

Usage

```
getReadMatrix.NB(vec.num.rep, alphas, sigma2s, phis)
```

Arguments

vec.num.rep	A vector of replicate numbers for each strain.
alphas	Intercept vector α_g 's, $1 \times \text{num.features}$.
sigma2s	Random effect variance vector σ_g^2 's, $1 \times \text{num.features}$.
phis	Dispersion parameter in NB models, ϕ_g 's, a $1 \times \text{num.features}$ vector.

Value

A $G \times N$ matrix with NB reads. N is the total number of samples; G is the number of features. Column names are sample names of the form "Ss_r", where S stands for sample, s is the strain number, r is the replicate number within the strain. Row names are the feature names of the form "Gene g", where g is the feature index.

Examples

```
## Generate a sequencing dataset with 5 features and 6 strains.
## Assign parameter values.
rep.num <- c(3, 5, 2, 3, 4, 2)
a0s <- c(-1, 1, 2, 5, 10)
sig2s <- c(10, 0.2, 0.1, 0.03, 0.01)
phis <- c(0.5, 1, 0.05, 0.01, 0.1)

set.seed(1234)
## Generate reads:
nbData <- getReadMatrix.NB(rep.num, a0s, sig2s, phis)
```

para_cp

*Parameter matrix obtained from simData by fitting CPMM.***Description**

Parameter matrix obtained from simData by fitting CPMM.

Usage

```
para_cp
```

Format

An object of class `matrix` with 100 rows and 4 columns.

para_nb

*Parameter matrix obtained from simData by fitting NBMM.***Description**

Parameter matrix obtained from simData by fitting NBMM.

Usage

```
para_nb
```

Format

An object of class `matrix` with 100 rows and 3 columns.

simData	<i>A simulated sequencing dataset.</i>
---------	--

Description

A matrix containing simulated counts for 100 features (rows) and 175 samples (columns)

Usage

```
simData
```

Format

A matrix with 100 rows and 175 columns

simData_voom	<i>Voom transformed version of simData.</i>
--------------	---

Description

Voom transformed version of simData.

Usage

```
simData_voom
```

Format

An object of class `matrix` with 881 rows and 175 columns.

simData_vst	<i>Variance stabilize transformed version of simData.</i>
-------------	---

Description

Variance stabilize transformed version of simData.

Usage

```
simData_vst
```

Format

An object of class `matrix` with 881 rows and 175 columns.

strains	<i>List of strain names for the samples.</i>
---------	--

Description

List of strain names for the samples.

Usage

strains

Format

An object of class character of length 175.

weights_voom	<i>Weights used in the voom transformation.</i>
--------------	---

Description

Weights used in the voom transformation.

Usage

weights_voom

Format

An object of class matrix with 881 rows and 175 columns.

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