Z-Score vs ASH

```
set.seed(1)
```

Alternative data set, generated by Poisson distribution

We simulate data such as $c_{ij} \sim Poisson(\mu_{ij})$, with μ_{ij} (expression level) like the following table.

1	2
2	1
10	20
20	10
÷	:
10^{7}	2×10^{7}
2×10^7	10^{7}

Each small block is 100×50 . In other words, for each expression level, we have 1000 genes and 50 samples for case and control each.

With the simulated count matrix and the condition vector, we could get a $\hat{\beta}$ and \hat{s} , as well as a z-score = $\hat{\beta}/\hat{s}$ for each gene by both OLS and voom + limma pipeline.

```
## effect size and standard error estimated by OLS
log_counts = log2(counts + 1)
ols_fit <- get_ols(log_counts = log_counts, condition = condition)
betahat_ols = ols_fit$betahat
sebetahat_ols = ols_fit$sebetahat
df_ols = ols_fit$df
z_ols = betahat_ols / sebetahat_ols

## effect size and measurement error estimated by voom + limma
voom_fit = voom_transform(counts, condition)
betahat_voom = voom_fit$betahat
sebetahat_voom = voom_fit$sebetahat
df_voom = voom_fit$df
z_voom = betahat_voom / sebetahat_voom</pre>
```

We group the genes to 16 bins of 1000 genes. Then we plot \hat{s} and z-score with respect to the expression level.

```
## grouping genes into K subgroups from low to high expression
group_id = rep(1:(2 * length(mu)), each = Ngene)

boxplot(abs(betahat_ols) ~ group_id)
boxplot(sebetahat_ols ~ group_id, xlab = "expression level", ylab = "sebetahat", main = "OLS")
boxplot(log(abs(z_ols)) ~ group_id, xlab = "expression level", ylab = "log |z-score|", main = "OLS")
abline(log(qnorm(0.975)), 0, lty = 3, col = "red")
```

```
boxplot(abs(betahat_voom) ~ group_id)
boxplot(sebetahat_voom ~ group_id, xlab = "expression level", ylab = "sebetahat", main = "TMM + voom boxplot(log(abs(z_voom)) ~ group_id, xlab = "expression level", ylab = "log |z-score|", main = "TMM + abline(0, 0, lty = 3, col = "red")
```

Now we use Poisson thinning to put artificial N(0, 1) effect to a count matrix.

```
pois_thinning <- function(counts, log2foldsd) {</pre>
        log2foldchanges <- rnorm(dim(counts)[1], mean = 0, sd = log2foldsd)</pre>
        foldchanges <- 2 ^ log2foldchanges
        Nsamp = dim(counts)[2]/2
        ## thin group A
        counts[log2foldchanges > 0, 1:Nsamp] <-</pre>
            matrix(rbinom(sum(log2foldchanges >
            0) * Nsamp, size = c(as.matrix(counts[log2foldchanges >
            0, 1:Nsamp])), prob = rep(1 / foldchanges[log2foldchanges > 0], Nsamp)),
            ncol = Nsamp)
        ## thin group B
        counts[log2foldchanges < 0, (Nsamp + 1):(2 * Nsamp)] <-</pre>
            matrix(rbinom(sum(log2foldchanges <</pre>
            0) * Nsamp, size = c(as.matrix(counts[log2foldchanges <</pre>
            0, (Nsamp + 1):(2 * Nsamp)])), prob = rep(foldchanges[log2foldchanges <</pre>
            O], Nsamp)), ncol = Nsamp)
    return(list(counts = counts, log2foldchanges = log2foldchanges))
}
Nsample = 20
counts = c(rpois(1000 * Nsample * 2, 1), rpois(1000 * Nsample * 2, 10), rpois(1000 * Nsample * 2, 100),
counts = matrix(counts, ncol = Nsample * 2, byrow = TRUE)
counts = pois_thinning(counts = counts, log2foldsd = 1)$counts
condition = rep(1:2, each = Nsample)
source("../code/fit method.R")
    ## effect size and standard error estimated by OLS
    log_counts = log_2(counts + 1)
    ols_fit <- get_ols(log_counts = log_counts, condition = condition)</pre>
    betahat_ols = ols_fit$betahat
    sebetahat_ols = ols_fit$sebetahat
    df_ols = ols_fit$df
    z_ols = betahat_ols / sebetahat_ols
    ## effect size and measurement error estimated by voom + limma
    voom_fit = voom_transform(counts, condition)
    betahat_voom = voom_fit$betahat
    sebetahat_voom = voom_fit$sebetahat
    df_voom = voom_fit$df
    z_voom = betahat_voom / sebetahat_voom
    ## effect size estimated by ash
    ash_fit_voom = ashr::ash(betahat_voom, sebetahat_voom, method = "shrink", mixcompdist = "normal")
```

```
beta_est_voom = ash_fit_voom$PosteriorMean
    ash_fit_ols = ashr::ash(betahat_ols, sebetahat_ols, method = "shrink", mixcompdist = "normal")
    beta_est_ols = ash_fit_ols$PosteriorMean

## grouping genes into K subgroups from low to high expression
group_id = rep(1:4, each = 1000)

boxplot(sebetahat_ols ~ group_id, xlab = "expression level", ylab = "sebetahat", main = "OLS")
boxplot(z_ols ~ group_id, xlab = "expression level", ylab = "z-score", main = "OLS")
abline(0, 0, lty = 3, col = "red")

boxplot(sebetahat_voom ~ group_id, xlab = "expression level", ylab = "sebetahat", main = "voom + limm
boxplot(z_voom ~ group_id, xlab = "expression level", ylab = "z-score", main = "voom + limma")
abline(0, 0, lty = 3, col = "red")

boxplot(beta_est_ols ~ group_id, xlab = "expression level", ylab = "estimated effect size", main = "Oboxplot(beta_est_voom ~ group_id, xlab = "expression level", ylab = "estimated effect size", main = "Oboxplot(beta_est_voom ~ group_id, xlab = "expression level", ylab = "estimated effect size", main = "Oboxplot(beta_est_voom ~ group_id, xlab = "expression level", ylab = "estimated effect size", main = "oboxplot(beta_est_voom ~ group_id, xlab = "expression level", ylab = "estimated effect size", main = "oboxplot(beta_est_voom ~ group_id, xlab = "expression level", ylab = "estimated effect size", main = "oboxplot(beta_est_voom ~ group_id, xlab = "expression level", ylab = "estimated effect size", main = "oboxplot(beta_est_voom ~ group_id, xlab = "expression level", ylab = "estimated effect size", main = "oboxplot(beta_est_voom ~ group_id, xlab = "expression level", ylab = "estimated effect size", main = "oboxplot(beta_est_voom ~ group_id, xlab = "expression level", ylab = "expression level", ylab
```