

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/235689164>

# Analyzing Molecular Polar Surface Descriptors to Predict Blood–Brain Barrier Permeation.

Article in *International Journal of Computational Biology and Drug Design* · February 2013

DOI: 10.1504/IJCBDD.2013.052195 · Source: PubMed

CITATIONS

13

READS

178

4 authors:



**Sergey Shityakov**

University of Wuerzburg

49 PUBLICATIONS 112 CITATIONS

[SEE PROFILE](#)



**Winfried Neuhaus**

University of Vienna

37 PUBLICATIONS 414 CITATIONS

[SEE PROFILE](#)



**Thomas Dandekar**

University of Wuerzburg

436 PUBLICATIONS 11,471 CITATIONS

[SEE PROFILE](#)



**Carola Förster**

University of Wuerzburg

75 PUBLICATIONS 1,372 CITATIONS

[SEE PROFILE](#)

Some of the authors of this publication are also working on these related projects:



Characterization, in vivo evaluation and molecular modeling of different propofol-cyclodextrin complexes to assess their drug delivery potential at the blood-brain barrier level [View project](#)



Characterization, in vivo evaluation and molecular modeling of different propofol-cyclodextrin complexes to assess their drug delivery potential at the blood-brain barrier level [View project](#)

---

## Analysing molecular polar surface descriptors to predict blood-brain barrier permeation

---

Sergey Shityakov\*

Department of Anesthesiology and Critical Care,  
University of Würzburg,  
97080 Würzburg, Germany  
Email: E\_Shityako\_S@klinik.uni-wuerzburg.de  
\*Corresponding author

Winfried Neuhaus

Department of Anesthesiology and Critical Care,  
University of Würzburg,  
97080 Würzburg, Germany  
and  
Department of Medicinal Chemistry,  
University of Vienna,  
1090 Vienna, Austria  
Email: winfried.neuhaus@univie.ac.at

Thomas Dandekar

Department of Bioinformatics,  
University of Würzburg,  
97074 Würzburg, Germany  
Email: dandekar@biozentrum.uni-wuerzburg.de

Carola Förster

Department of Anesthesiology and Critical Care,  
University of Würzburg,  
97080 Würzburg, Germany  
Email: Foerster\_C@klinik.uni-wuerzburg.de

**Abstract:** Molecular polar surface (PS) descriptors are very useful parameters in prediction of drug transport properties. They could be also used to investigate the blood-brain barrier (BBB) permeation rate for various chemical compounds. In this study, a dataset of drugs ( $n = 19$ ) from various pharmacological groups was studied to estimate their potential properties to permeate across the BBB. Experimental logBB data were available as steady-state distribution values of the *in vivo* rat model for these molecules. Including accurate calculation of the electrostatic potential maps, polar surface descriptors, such as a two-dimensional polar surface area (2D-PSA), topological polar surface area (TPSA) and three-dimensional polar surface area or polar area (3D-PSA; PA) were measured and analysed. We report the strong correlation of these descriptors with logBB values for the prediction of BBB permeation using the linear partial least squares (PLS) fitting technique. The

3D-PSA descriptor showed the best fit to logBB values with  $R^2 = 0.92$  and  $\text{RMSD} = 0.29$  ( $p\text{-value} < 0.0001$ ). The obtained results demonstrate that all descriptors bear high predictive powers and could provide an efficient strategy to envisage the pharmacokinetic properties of chemical compounds to permeate across the BBB at an early stage of the drug development process.

**Keywords:** polar surface descriptors; blood-brain barrier; logBB; P-glycoprotein.

**Reference** to this paper should be made as follows: Shityakov, S., Neuhaus, W., Dandekar, T. and Förster, C. (2013) 'Analysing molecular polar surface descriptors to predict blood-brain barrier permeation', *Int. J. Computational Biology and Drug Design*, Vol. 6, Nos. 1/2, pp.146–156.

**Biographical notes:** Sergey Shityakov is currently a Postdoctoral Fellow in the Department of Anaesthesiology and Critical Care at Würzburg Hospital University. He received an MD in Immunology and Infectious Diseases from Novgorod State University School of Medicine in Russia, and a PhD in Molecular Biology from Würzburg University in Germany. His research interests include blood-brain barrier research, molecular docking and molecular dynamics simulation of different biological systems, such as brain transporter proteins and carbon nanostructures. His work in these areas has been published in various peer-reviewed journals and presented at scientific conferences.

Winfried Neuhaus is currently a Postdoctoral Fellow in the Department of Anaesthesiology and Critical Care at the University of Würzburg, and an Assistant in the Department of Medicinal Chemistry at the University of Vienna. He received an MS in Biotechnology and a PhD in Life Sciences from the University of Vienna in Austria. He worked both as a lecturer in Biotechnology at the University of Vienna and as a group leader of the Preclinical and Blood-Brain Barrier Research Department in the Pharmacon company. His main research interest is the blood-brain barrier, which includes establishment of *in-vitro* models and the role of the blood-brain barrier during a stroke, together with therapeutic strategies to restore blood-brain barrier integrity after brain damage.

Thomas Dandekar holds the Chair in Bioinformatics at Würzburg University Biocenter in Germany. He is a Member of the Board of Directors and currently serves as Vice Dean at the Faculty of Biology. He conducts research in the fields of functional genomics and systems biology to investigate the cell regulatory networks, which control various signaling elements, receptors, kinases, and cytoskeleton proteins including siRNA and lincRNA analysis as well as structural modeling of receptors and ligands. His research group is also developing bioinformatics tools for metabolic modeling, genome annotation, and modelling of signalling networks. Research funding includes, mainly, DFG and BMBF projects, but also IZKF, Land Bavaria and other agencies.

Carola Förster is a Professor in the Department of Anaesthesiology and Critical Care at Würzburg Hospital University. She received an MS in Biochemistry and a PhD in Molecular Biology from Hannover University in Germany. Her main research topics have included the role of the blood-brain barrier in inflammatory diseases and brain injury, effects of maternal inflammation on the blood-brain barrier integrity in fetal brain development, stem cell therapy and regenerative medicine after brain injury, characterisation and pharmacological modulation of selected transporter proteins in the brain. She has participated in numerous IZKF, BMBF, and DFG projects.

---

## 1 Introduction

Permeation of active drugs across the vascular brain endothelium into the central nervous system (CNS) is controlled by the blood-brain barrier (BBB). The BBB separates the bloodstream from the brain. Characteristics of the BBB are the restriction of paracellular substance permeation across the endothelium by intercellular tight junctions, the lack of cellular fenestrae and reduced pinocytosis (van Bree et al., 1992; Hirase et al., 1997). In contrast to several endogenous hydrophilic nutrients (glucose, amino acids, etc...), which are transported by carrier-mediated mechanisms across the brain endothelium, the BBB prevents the entry into the CNS of the majority of polar drugs (Geldenhuys et al., 2010). An important parameter for BBB penetration is the rate at which a molecule passes through a lipid membrane. Highly lipophilic chemical agents can penetrate the BBB easily by passive diffusion, whereas less lipophilic agents cannot (Hansch et al., 1987). Therefore, most of the drug compounds have a mixture of lipophilic-hydrophilic properties that allow them to partition between lipid and aqueous phases and to reach their site of action. Usually, these compounds have low molecular weight to permeate across the BBB (Fu et al., 2008).

Herewith, if a molecule is not sufficiently polar, it will not be able to leave the membrane to return to the aqueous environment. Thus, overall molecular polarity plays an obvious role: if the substance is too polar, the lipid may not be able to drag the molecule from the aqueous solvent. In this case, some molecular quantities, such as polar surface (PS) descriptors are of key interest to medicinal chemists to predict the BBB permeation fate for different drug-like chemical compounds. The descriptors commonly used to account for polarity are the so-called polar surface descriptors like two-dimensional polar surface area (2D-PSA), topological polar surface area (TPSA) and three-dimensional polar surface area or polar area (3D-PSA, PA) (Clark, 1999; Kelder et al., 1999; Ertl et al., 2000; Kim et al., 2011). The last has to be calculated with time-consuming quantum-mechanical methods usually providing better results (Dove, 2008). All of these parameters are interpretable, numerically stable and suitable for good correlation with experimental transport data, such as logBB (Kelder et al., 1999). However, one has to consider the fact that some molecular compounds are substrates for efflux transporters, such as P-glycoprotein (P-gp). Therefore, the access of these molecules to the brain is very restricted (Osterberg and Norinder, 2000; Ooms et al., 2002; Ward et al., 2004; Gunes et al., 2008; Thiel-Demby et al., 2009). Some of the molecular descriptors (2D-PSA, TPSA) were defined to be numerical properties that can be calculated from the connection table representation of a molecule (e.g. elements, formal charges and bonds, but not atomic coordinates). This type of descriptor is not dependent on the 3D conformation of a molecule and is mostly suitable for analysis of huge molecular databases and drug high-throughput screening (Ertl et al., 2000). Therefore, 2D-PSA as well as TPSA descriptors are widely used medicinal chemistry metrics for the optimisation of a drug's ability to penetrate the BBB. The calculations of 3D-PSA, for example, are time-consuming and require highly specialised software to construct and optimise the molecular surface itself.

We report the prediction of the BBB permeation by analysing polar surface descriptors of diverse CNS-active/inactive compounds to find a strong correlation of these descriptors with logBB values using the linear partial least squares (PLS) fitting technique. In the case of effective CNS-acting drugs, the understanding of their permeation mechanism through the BBB is pivotal to filter potential leads and to estimate and diminish the various neurotoxic side-effects.

## 2 Computational methods

A good *in silico* model is based on good data sets; therefore, all necessary information regarding the chemical structures and experimental logBB values for 19 diverse drugs was taken from the literature (Kelder et al., 1999). All data sets contained the blood/brain concentration ratios taken as steady-state distribution values of rat model measurements as:

$$\log BB = \log(C_{\text{brain}}/C_{\text{blood}})$$

The logBB ranged from −2.0 to 1.0 (Table 1) were either brain-penetrating (logBB > 0.5), or had moderate permeation (logBB from 0.0 to 0.5), or possessed little ability to cross the blood-brain barrier (logBB > −0.3), or demonstrated very little permeation (logBB < −0.3).

The three-dimensional structure of the corresponding molecules was built and minimised using the Spartan'10 general purpose molecular modelling tool. 2D-PSA (Spartan'10 calculated descriptor) was mainly defined as the area associated with oxygen, nitrogen and hydrogen attached to these polarising atoms (Hehre, 2003).

**Table 1** Calculated polar surface descriptors for 19 CNS-active/inactive drugs

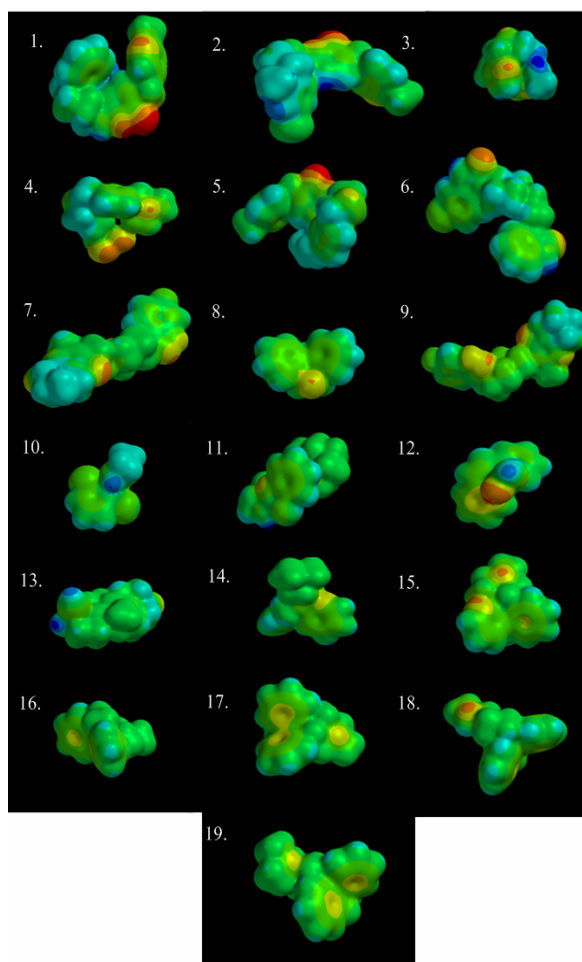
| Compound                    | MW     | logBB | PSA   | TPSA  | PA    | W2      | P-gp     |
|-----------------------------|--------|-------|-------|-------|-------|---------|----------|
| 1. icotidine                | 378.52 | −2.00 | 49.52 | 75.61 | 79.53 | 744.25  | + [11]   |
| 2. temelastine              | 442.40 | −1.88 | 47.79 | 79.27 | 91.20 | 813.25  | + [12]   |
| 3. cimetidine               | 252.39 | −1.42 | 59.14 | 88.89 | 65.34 | 632.63  | + [13]   |
| 4. ranitidine               | 413.60 | −1.23 | 77.95 | 96.50 | 71.55 | 698.38  | + [13]   |
| 5. lupitidine               | 425.96 | −1.06 | 47.55 | 82.70 | 81.89 | 803.50  | n.a      |
| 6. domperidone              | 426.54 | −0.78 | 57.98 | 67.92 | 62.71 | 893.88  | + [14]   |
| 7. 9-hydroxyrisperidone     | 426.54 | −0.67 | 63.36 | 82.17 | 56.75 | 1116.13 | + [15]   |
| 8. 10,11-epoxycarbamazepine | 252.29 | −0.33 | 48.25 | 58.86 | 45.19 | 568.75  | − [29]   |
| 9. risperidone              | 410.54 | −0.02 | 44.28 | 61.94 | 51.72 | 883.63  | + [15]   |
| 10. carbamazepine           | 236.29 | 0.00  | 36.76 | 46.33 | 34.85 | 479.88  | − [30]   |
| 11. clonidine               | 230.11 | 0.11  | 29.98 | 36.42 | 34.40 | 360.00  | − [7]    |
| 12. zolantidine             | 381.59 | 0.14  | 27.77 | 37.39 | 42.61 | 649.13  | n.a      |
| 13. tribolone               | 312.49 | 0.40  | 33.57 | 37.30 | 36.09 | 689.63  | n.a      |
| 14. pyrilamine              | 285.43 | 0.49  | 15.11 | 28.60 | 17.79 | 545.25  | − [31]   |
| 15. mirtazapine             | 265.39 | 0.53  | 10.86 | 19.37 | 15.56 | 443.75  | n.a [32] |
| 16. amitriptyline           | 277.44 | 0.98  | 2.54  | 3.24  | 11.85 | 410.00  | + [33]   |
| 17. mianserin               | 264.40 | 0.99  | 4.00  | 6.48  | 16.62 | 375.00  | n.a [34] |
| 18. desipramine             | 266.42 | 1.00  | 12.90 | 15.27 | 13.07 | 435.63  | − [35]   |
| 19. imipramine              | 280.45 | 1.00  | 3.26  | 6.40  | 15.77 | 366.00  | − [7]    |

Notes: n.a. = not available, '+' means P-gp substrate, '−' means non-P-gp substrate.

Similar to 2D-PSA, the topological polar surface area descriptor was calculated using group contributions, applying the build-in module in the MOE2009 software, to approximate the polar surface area from atomic connectivity using topological information only. TPSA of a molecule was defined as the surface sum over all polar atoms and is based simply on the summation of polar fragments representing tabulated surface contributions, i.e. bonding patterns of a molecule (Ertl et al., 2000).

The Merck Molecular Force Field (MMFF) molecular mechanics was used to refine and minimise molecular geometry. To accurately analyse the charge distribution of a molecule, a potential energy describing the electrostatic interactions between charged particles was calculated. Density functional theory (DFT), single-point energy calculations were performed with 6-31G\* basis set to build electrostatic potential maps for all the molecules (Figure 1). These maps for each molecule were used to reconstruct its three-dimensional polar surface area (3D-PSA) in order to build the electrostatic potential at locations on a particular surface, most commonly a surface of electron density corresponding to overall molecular size ([Csizmadia, 1976](#); [Petrucci et al., 2007](#); [Oxtoby et al., 2008](#)). The map was defined as an area for which the absolute value of the electrostatic potential exceeded 100 kJ/mol default parameter.

**Figure 2** Electrostatic potential maps (3D-PSA) for 19 CNS-active/inactive diverse chemical compounds. The numbers in the figure correspond to the compound numbers in Table 1. Electrostatic potential maps fit roughly to conventional space-filling models and overall sizes, and shapes were that of the electron densities. The colours indicate the value of the electrostatic potential. Red colours designate areas of negative potential (where a positive charge is most likely to be attracted), while blue colours depicted the areas of positive potential (see online version for colours)



Descriptor of hydrophilic regions, such as hydrophilic volume (W2) was computed from molecular fields using the Volsurf+ molecular modelling software. Hydrophilic W2 descriptor described the molecular envelope, which is accessible to water molecules and attractively interacts with them. The volume of this envelope varies with the level of interaction energies. For W2 descriptor, polarisability and dispersion force parameters were comprised within the range of  $-0.2$  to  $-1.0$  kcal/mol.

Finally, the root mean square difference (RMSD), an indication of the average error in the analysis; the square of the correlation coefficient ( $R^2$ ), an indication of the quality of fit of all the data to a straight line (clear correlation of descriptors versus experimental logBB values) were also calculated and compared for all the descriptors. The experimental logBB were plotted against the calculated molecular surface descriptors; linear partial least squares technique was used to perform a fit to both sets of variables.

### 3 Results and discussion

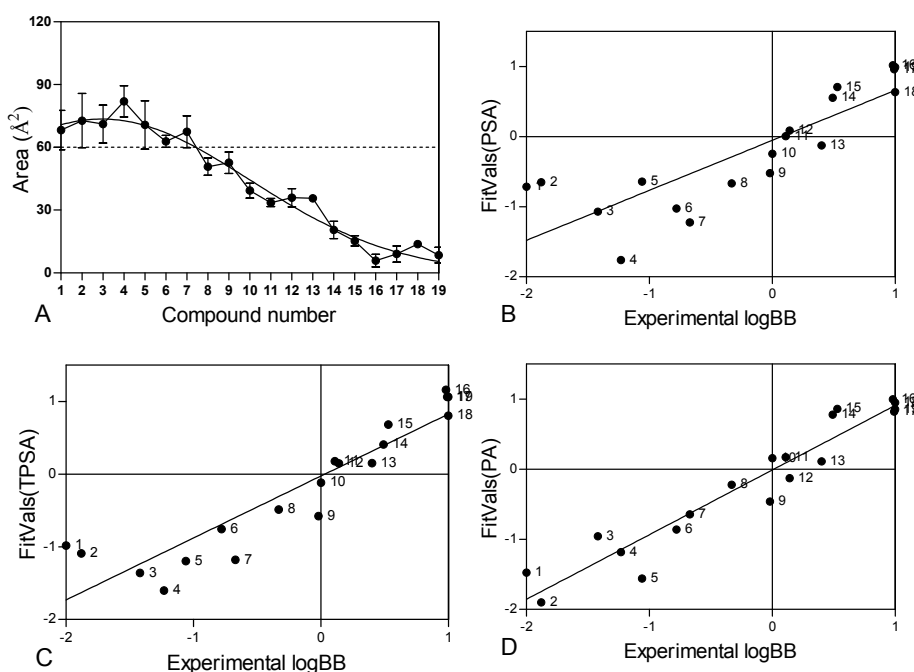
First, we did accurate calculations of the electrostatic potential maps for each molecule. Then, the PS descriptor methodology has been implemented by using published data for logBB properties. Specific descriptors were calculated by different modelling software packages for the molecular dataset ( $n = 19$ ). The results of correlation studies are summarised in Table 1, where calculated polar surface descriptors and molecular weights for drug molecules are given. Considering the role of efflux transporters, such as P-glycoprotein (P-gp) in the drug permeation of the BBB, the information about the P-gp substrate specificity was assigned for each chemical compound. The corresponding electrostatic potential maps are presented in Figure 1.

From Figure 2A it can be concluded that BBB non-penetrants (compounds 1–7) represent higher total area deviations ( $SD = 3.09$ ) for all calculated descriptors than BBB penetrants (compounds 10–19) with  $SD = 1.18$ . The polar surface area parameter was the main factor contributing to increased standard deviation rate. The serious disadvantage of using PSA as well as TPSA is the inability to distinguish the activity differences of chemical compounds differing only in their non-polar groups. Moreover, using the TPSA descriptor, it would be problematic to analyse the influence of positional charges of functional groups (Prasanna and Doerksen, 2009). In our study, we also calculated the computationally more expensive 3D descriptors (PA) to improve this mode. However, overall distribution patterns for PS descriptors remain characteristic. In other words, for molecules to penetrate the blood-brain barrier an area less than  $60 \text{ \AA}^2$  (Figure 2A, threshold depicted as a dashed line) is usually needed; and molecules with a polar surface area of greater than  $120\text{--}140 \text{ \AA}^2$  tend to be poor in permeating cell membranes (Palm et al., 1997).

It is well known that multidrug efflux transporter pumps like P-gp could limit a variety of medications from reaching their therapeutic targets thereby preventing or minimising pharmacological effects (Mizuno et al., 2003). By using the P-gp knockout mouse model, Wang and co-authors have demonstrated that some anti-psychotic drugs, such as risperidone are P-gp substrates and their entry into the brain is dramatically limited by P-gp efflux transporter in the BBB (Wang et al., 2004; Doran et al., 2005). However, anti-epileptic compound, 10,11-epoxycarbamazepine, and risperidone, both have in addition, as the calculation shows, a mean area value below the  $60 \text{ \AA}^2$  ( $50.77 \text{ \AA}^2$

and  $52.65 \text{ \AA}^2$ ) indicating BBB permeability. For the last compound, the difference in logBB distribution from its 9-hydroxy metabolite (9-hydroxyrisperidone) could be explained by the higher area value of the metabolite ( $67.43 \text{ \AA}^2$ ).

**Figure 2** Comparing the overall polar surface descriptors against each other. The numbers in the figure correspond to the compound numbers in Table 1; the descriptors are highly correlated. There are BBB– and BBB+ clusters defined by the area threshold ( $60 \text{ \AA}^2$ ) (A). Plots of experimental logBB values versus calculated fit variables (FitVals) of polar surface descriptors are shown (B–D)



Furthermore, 10,11-epoxycarbamazepine is a major metabolite of carbamazepine, which is known to be a cytochrome P4503A4 (CYP3A4) but not a P-gp substrate (Owen et al., 2001). Initially, carbamazepine was extensively catalysed by CYP3A4 via the oxidative degradation pathway, mainly in the liver (Kerr et al., 1994).

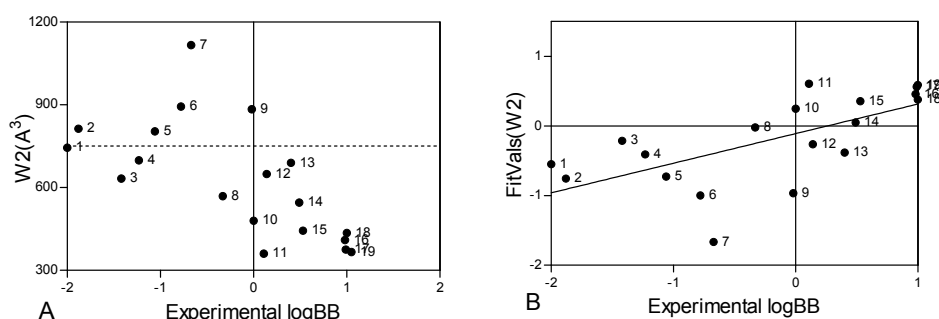
However, in spite of previously shown data, those compounds with logBB around  $-0.5$  might still be considered in vivo experiments to be BBB crossing (Crivori et al., 2000).

The linear partial least squares fitting provided high correlation coefficients of calculated descriptors vs. experimental logBB data, namely:  $R^2 = 0.71$ , RMSD = 0.54 for PSA;  $R^2 = 0.85$ , RMSD = 0.39 for TPSA and  $R^2 = 0.92$ , RMSD = 0.29 for PA descriptor with the  $p$ -value  $< 0.0001$  indicating the highly predictive properties of the introduced descriptors for drug permeation across the BBB. The RMSD values are an estimate of the absolute error prediction for a property: the smaller RMSD parameter and the closer correlation coefficient is to a unity – the better the fit (Figure 2B–D). Although the  $R^2$  deviation from calculated descriptor distribution modes indicates the different predictive power of analysed descriptors, the predictive power is also influenced by experimental difficulties, for instance, when the concentration of the drug in the brain is low.



For comparison, we calculated the hydrophilic volume descriptor W2. In general, hydrophilic volume descriptors are known to be highly correlative to logBB values (Cruciani et al., 2000; Molecular Discovery, 2002; Braiuca et al., 2007). The W2-logBB plot (Figure 3A) reports chemical substances as both BBB crossing or non-crossing compounds; and W2 vs. logBB curve fitting provided  $R^2 = 0.42$  because of too large RMSD (0.77) with the  $p$ -value = 0.0026 (Figure 3B). It is evident that the scatter observed in this plot further supports the conclusion that W2 is less reliable for BBB permeation prediction than PS descriptors.

**Figure 3** Relationships between experimental logBB and calculated hydrophilic volume (W2) descriptor (A) and its fit values (B). The correlation clearly shows that W2 increases when the brain penetration decreases. The numbers in the figure correspond to the compound numbers in Table 1



Overall, when W2 is larger than  $750 \text{ Å}^3$  (Figure 3A, dashed line), it is unlikely that compounds will penetrate the BBB. Hence, compounds with W2 exceeding this threshold are considered to be non-penetrants. This threshold shows a distinct separation of compounds into BBB penetrants (compounds 10–19) and non-penetrants (compounds 1–9) with three exceptions (two H2-histamine receptor antagonists: cimetidine and ranitidine, and 10,11-epoxycarbamazepine). In the case of H2-histamine receptor antagonists; it was already shown that their permeability is pH-dependent as well as controlled by their P-glycoprotein substrate specificity. Similarly, their brain distribution is mediated primarily by P-gp efflux mechanism (Dudley and Brown, 1996).

For more accurate predictions using PS descriptors, the basic assumption has to be that drugs penetrate across the BBB by passive diffusion. However, it is clear from the previously shown graphs that icotidine, temelastine, ranitidine, domperidone, 9-hydroxyrisperidone and risperidone were depicted as outliers because of their P-gp substrate activity (Osterberg and Norinder, 2000; Ooms et al., 2002; Ward et al., 2004; Gunes et al., 2008; Thiel-Demby et al., 2009). Surprisingly, amitriptyline was reported as P-gp substrate with a high logBB value. In this case, it could be hypothesised that amitriptyline might be transported into the brain by unknown influx transport mechanism, which exhibit higher transport rates than its efflux by P-gp at the BBB.

#### 4 Conclusion

In order to be effective as therapeutic agents, centrally acting drugs must cross the BBB, and entry into the brain is a complex phenomenon, which depends on a multiplicity of factors.

Therefore, it is important to mention that intramolecular H-bond formations in drug molecules could dramatically change the BBB permeation properties of the chemical compounds, which were not included in the calculations. The intramolecular H-bond could reduce polarity considerably and increase the lipophilicity of the chemical substance. This problem will be even more pronounced for structurally flexible molecules with great abilities to generate multiple conformational isoforms. Therefore, the BBB permeability of drugs could be improved in the future by lowering the overall hydrogen-bonding ability of a drug compound via facilitating intramolecular hydrogen bonding. This could be achieved by protecting a molecule with non-polar groups, or by synthesising less polar prodrugs (Young et al., 1988).

#### Acknowledgements

The authors are grateful to the IZKF (Interdisziplinäres Zentrum für Klinische Forschung der Universität Würzburg) and the BMBF (Bundesministerium für Bildung und Forschung) for the support of this work by providing grants (BMBF01, EO1004) to CF.

#### References

- Baltes, S., Gastens, A.M., Fedrowitz, M., Potschka, H., Kaever, V. and Löscher, W. (2007) 'Differences in the transport of the antiepileptic drugs phenytoin, levetiracetam and carbamazepine by human and mouse P-glycoprotein', *Neuropharmacology*, Vol. 52, pp.333–546.
- Besret, L., Debruyne, D., Rioux, P., Bonvalot, T., Moulin, M., Zarifian, E. and Baron, J.C. (1996) 'A comprehensive investigation of plasma and brain regional pharmacokinetics of imipramine and its metabolites during and after chronic administration in the rat', *Journal of Pharmaceutical Sciences*, Vol. 85, pp.291–295.
- Bogni, A., Monshouwer, M., Moscone, A., Hidestrand, M., Ingelman-Sundberg, M., Hartung, T. and Coecke, S. (2005) 'Substrate specific metabolism by polymorphic cytochrome P450 2D6 alleles', *Toxicology In Vitro*, Vol. 19, pp.621–629.
- Braiuca, P., Buthe, A., Ebert, C., Linda, P. and Gardossi, L. (2007) 'Volsurf computational method applied to the prediction of stability of thermostable enzymes', *Biotechnology Journal*, Vol. 2, pp.214–220.
- Clark, D.E. (1999) 'Rapid calculation of polar molecular surface area and its application to the prediction of transport phenomena. 2. Prediction of blood-brain barrier penetration', *Journal of Pharmaceutical Sciences*, Vol. 88, pp.815–821.
- Crivori, P., Cruciani, G., Carrupt, P.A. and Testa, B. (2000) 'Predicting blood-brain barrier permeation from three-dimensional molecular structure', *Journal of Medicinal Chemistry*, Vol. 43, pp.2204–2216.
- Cruciani, G., Pastor, M. and Guba, W. (2000) 'VolSurf: a new tool for the pharmacokinetic optimization of lead compounds', *European Journal of Pharmaceutical Sciences*, Vol. 11, Suppl 2, pp.S29–S39.

- Csizmadia, I.G. (1976) *Theory and Practice of MO Calculations on Organic Molecules*, Elsevier, Amsterdam.
- Doran, A., Obach, R.S., Smith, B.J., Hosea, N.A., Becker, S., Callegari, E., Chen, C., Chen, X., Choo, E., Cianfroga, J., Cox, L.M., Gibbs, J.P., Gibbs, M.A., Hatch, H., Hop, C.E., Kasman, I.N., Laperle, J., Liu, J., Liu, X., Logman, M., Maclin, D., Nedza, F.M., Nelson, F., Olson, E., Rahematpura, S., Raunig, D., Rogers, S., Schmidt, K., Spracklin, D.K., Szewc, M., Troutman, M., Tseng, E., Tu, M., Van Deusen, J.W., Venkatakrishnan, K., Walens, G., Wang, E.Q., Wong, D., Yasgar, A.S. and Zhang, C. (2005) 'The impact of P-glycoprotein on the disposition of drugs targeted for indications of the central nervous system: evaluation using the MDR1A/1B knockout mouse model', *Drug Metabolism and Disposition*, Vol. 33, pp.165–174.
- Dove, M.T. (2008) 'An introduction to atomistic simulation methods', *Seminarios de la SEM*, Vol. 4, pp.7–37.
- Dudley, A.J. and Brown, C.D. (1996) 'Mediation of cimetidine secretion by P-glycoprotein and a novel H(+)-coupled mechanism in cultured renal epithelial monolayers of LLC-PK1 cells', *British Journal of Pharmacology*, Vol. 117, pp.1139–1144.
- Ertl, P., Rohde, B. and Selzer, P. (2000) 'Fast calculation of molecular polar surface area as a sum of fragment-based contributions and its application to the prediction of drug transport properties', *Journal of Medicinal Chemistry*, Vol. 43, pp.3714–3717.
- Fu, X.C., Wang, G.P., Shan, H.L., Liang, W.Q. and Gao, J.Q. (2008) 'Predicting blood-brain barrier penetration from molecular weight and number of polar atoms', *European Journal of Pharmaceutics and Biopharmaceutics*, Vol. 70, pp.462–466.
- Geldenhuis, W.J., Allen, D.D. and Lockman, P.R. (2010) '3-D-QSAR and docking studies on the neuronal choline transporter', *Bioorganic and Medicinal Chemistry Letters*, Vol. 20, pp.4870–4877.
- Gunes, A., Spina, E., Dahl, M.L. and Scordo, M.G. (2008) 'ABCB1 polymorphisms influence steady-state plasma levels of 9-hydroxyrisperidone and risperidone active moiety', *Therapeutic Drug Monitoring*, Vol. 30, pp.628–633.
- Hansch, C., Björkroth, J.P. and Leo, A. (1987) 'Hydrophobicity and central nervous system agents: on the principle of minimal hydrophobicity in drug design', *Journal of Pharmaceutical Sciences*, Vol. 76, pp.663–687.
- Hehre, W.A. (2003) *Guide to Molecular Mechanics and Quantum Chemical Calculations*, Wavefunction, Inc. Available online at: <http://www.wavefun.com/support/AGuidetoMM.pdf>
- Hirase, T., Staddon, J.M., Saitou, M., Ando-Akatsuka, Y., Itoh, M., Furuse, M., Fujimoto, K., Tsukita, S. and Rubin, L.L. (1997) 'Occludin as a possible determinant of tight junction permeability in endothelial cells', *Journal of Cell Science*, Vol. 110, pp.1603–1613.
- Ishiwata, K., Kawamura, K., Yanai, K. and Hendrikse, N.H. (2007) 'In vivo evaluation of P-glycoprotein modulation of 8 PET radioligands used clinically', *Journal of Nuclear Medicine*, Vol. 48, pp.81–87.
- Kelder, J., Grootenhuis, P.D., Bayada, D.M., Delbressine, L.P. and Ploemen, J.P. (1999) 'Polar molecular surface as a dominating determinant for oral absorption and brain penetration of drugs', *Pharmaceutical Research*, Vol. 16, pp.1514–1519.
- Kerr, B.M., Thummel, K.E., Wurden, C.J., Klein, S.M., Kroetz, D.L., Gonzalez, F.J. and Levy, R.H. (1994) 'Human liver carbamazepine metabolism – role of CYP3A4 and CYP2C8 in 10,11-epoxide formation', *Biochemical Pharmacology*, Vol. 47, pp.1969–1979.
- Kim, H., Sulaimon, S., Menezes, S., Son, A. and Menezes, W.J. (2011) 'A comparative study of successful central nervous system drugs using molecular modeling', *Journal of Chemical Education*, Vol. 88, pp.1389–1393.
- Mizuno, N., Niwa, T., Yotsumoto, Y. and Sugiyama, Y. (2003) 'Impact of drug transporter studies on drug discovery and development', *Pharmacological Reviews*, Vol. 55, pp.425–461.
- Molecular Discovery (2002) *VolSurf+ User Manual*, Molecular Discovery Ltd. Available online at: <http://www.moldiscovery.com/docs/vsplus/> (accessed on 15 December 2011).

- Ooms, F., Weber, P., Carrupt, P.A. and Testa, B. (2002) 'A simple model to predict blood-brain barrier permeation from 3D molecular fields', *Biochimica et Biophysica Acta*, Vol. 1587, pp.118–125.
- Osterberg, T. and Norinder, U. (2000) 'Prediction of polar surface area and drug transport processes using simple parameters and PLS statistics', *Journal of Chemical Information and Computer Sciences*, Vol. 40, pp.1408–1411.
- Owen, A., Pirmohamed, M., Tettey, J.N., Morgan, F., Chadwick, D. and Park, B.K. (2001) 'Carbamazepine is not a substrate for P-glycoprotein', *British Journal of Clinical Pharmacology*, Vol. 51, pp.345–349.
- Owen, A., Pirmohamed, M., Tettey, J.N., Morgan, P., Chadwick, D. and Park, B.K. (2001) 'Carbamazepine is not a substrate for P-glycoprotein', *British Journal of Clinical Pharmacology*, Vol. 51, pp.345–349.
- Oxtoby, D.W., Gillis, H.P. and Campion, A. (2008) *Principles of Modern Chemistry*, 6th ed., Thomson Brooks/Cole: Belmont, CA.
- Palm, K., Stenberg, P., Luthman, K. and Artursson, P. (1997) 'Polar molecular surface properties predict the intestinal absorption of drugs in humans', *Pharmaceutical Research*, Vol. 14, pp.568–571.
- Petrucchi, R.H., Harwood, W.S. and Herring, F.G. (2007) *General Chemistry: Principles and Modern Applications*, 9th ed., Pearson Education, Inc., New Jersey.
- Prasanna, S. and Doerksen, R.J. (2009) 'Topological polar surface area: a useful descriptor in 2D-QSAR', *Current Medicinal Chemistry*, Vol. 16, pp.21–41.
- Störmer, E., von Moltke, L.L., Shader, R.I. and Greenblatt, D.J. (2000) 'Metabolism of the antidepressant mirtazapine in vitro: contribution of cytochromes P-450 1A2, 2D6, and 3A4', *Drug Metabolism and Disposition*, Vol. 28, pp.1168–1175.
- Thiel-Demby, V.E., Humphreys, J.E., St John Williams, L.A., Ellens, H.M., Shah, N., Ayrton, A.D. and Polli, J.W. (2009) 'Biopharmaceutics classification system: validation and learnings of an in vitro permeability assay', *Molecular Pharmaceutics*, Vol. 6, pp.11–18.
- Uhr, M., Grauer, M.T., Yassouridis, A. and Ebinger, M. (2007) 'Blood-brain barrier penetration and pharmacokinetics of amitriptyline and its metabolites in p-glycoprotein (abcblab) knock-out mice and controls', *Journal of Psychiatric Research*, Vol. 41, pp.179–188.
- van Bree, J.B., de Boer, A.G., Danhof, M. and Breimer, D.D. (1992) 'Drug transport across the blood-brain barrier: I. Anatomical and physiological aspects', *Pharmaceutisch Weekblad, Scientific Edition*, Vol. 14, No. 5, pp.305–310.
- Wang, J.S., Ruan, Y., Taylor, R.M., Donovan, J.L., Markowitz, J.S. and DeVane, C.L. (2004) 'The brain entry of risperidone and 9-hydroxyrisperidone is greatly limited by P-glycoprotein', *International Journal of Neuropsychopharmacology*, Vol. 7, pp.415–419.
- Ward, B.A., Morocho, A., Kandil, A., Galinsky, R.E., Flockhart, D.A. and Desta, Z. (2004) 'Characterization of human cytochrome P450 enzymes catalyzing domperidone N-dealkylation and hydroxylation in vitro', *British Journal of Clinical Pharmacology*, Vol. 58, pp.277–287.
- Young, R.C., Mitchell, R.C., Brown, T.H., Ganellin, C.R., Griffiths, R., Jones, M., Rana, K.K., Saunders, D., Smith, I.R., Sore, N.E. and Wilks, T.J. (1988) 'Development of a new physicochemical model for brain penetration and its application to the design of centrally acting H<sub>2</sub> receptor histamine antagonists', *Journal of Medicinal Chemistry*, Vol. 31, pp.656–671.