



NGS: technologies and challenges

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Today we will talk about



- Genomics Platforms and sequencing services at NGI, SciLifeLab
- History and current status of technologies for sequencing
- NGS applications and technologies
- NGS challenges and sample requirements
- Data analysis pipelines, R&D and strategic projects



Service areas of SciLifeLab

Bioinformatics

Bioimaging and Molecular Structure

Chemical Biology and Genome Engineering

Drug Discovery

Diagnostics

Genomics

Metabolomics

Single Cell Biology

Spatial Omics

Proteomics

Across all service areas: dedicated staff scientists that can offer support **throughout the experimental process** – from study design to data handling

SciLifeLab Genomics



Ancient DNA

We use cleanroom labs and specialized molecular genetics techniques to extract, make libraries, sequence and analyze DNA in ancient and/or degraded biological material.

[Learn More](#) - →

Clinical Genomics

Develops and provides clinical genetic tests using state-of-the-art genomic methods, such as next-generation sequencing, for translational research and healthcare.

[Learn More](#) - →

National Bioinformatics Infrastructure (NBIS)

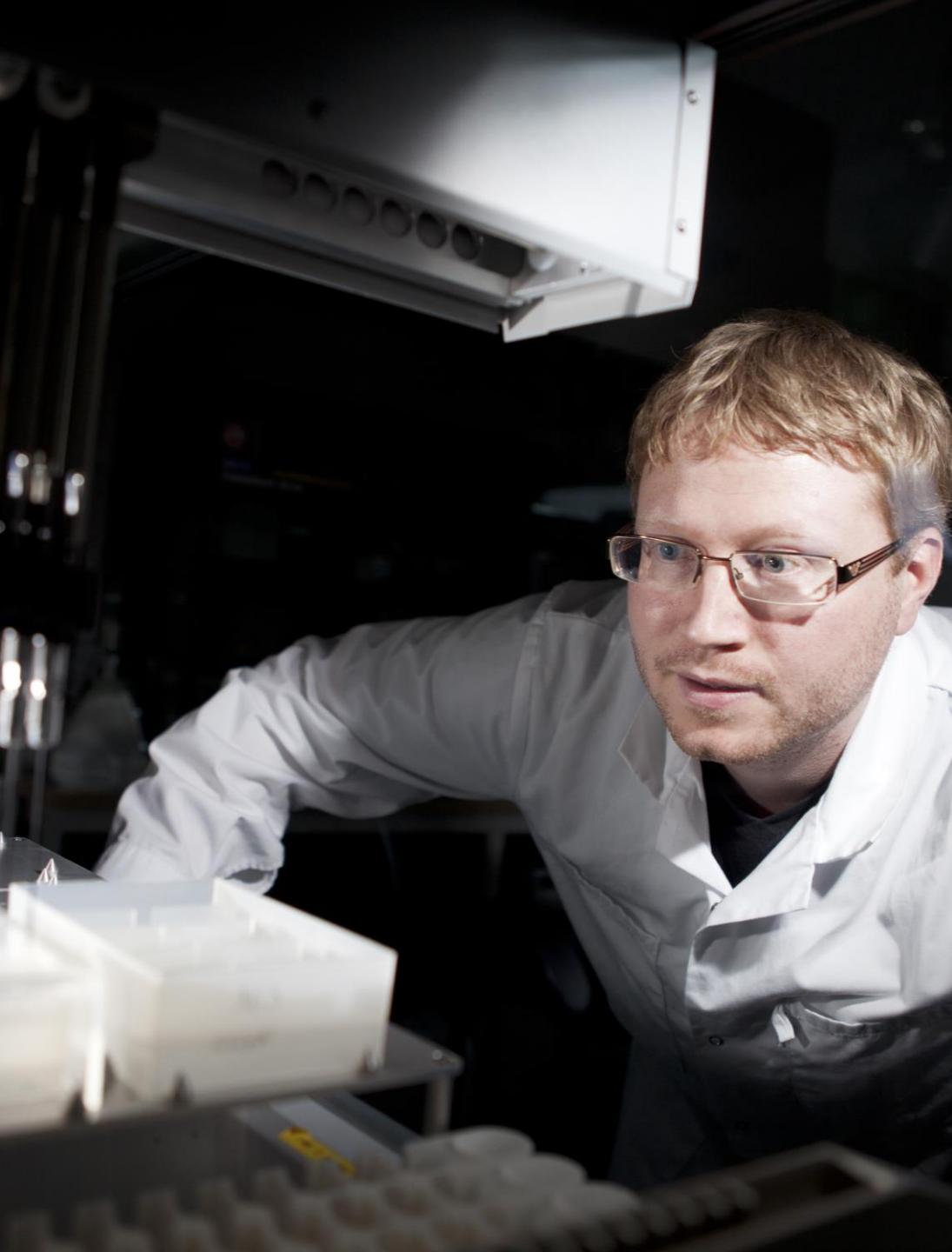
Provides custom-tailored support with analysis of genomics data generated at SciLifeLab or elsewhere, as well as tools and training.

[Learn More](#) - →

National Genomics Infrastructure (NGI)

Provides services for next generation sequencing and SNP genotyping on all scales using a comprehensive range of modern technology.

[Learn More](#) - →



What is National Genomics Infrastructure (NGI)?

NGI provides access to technology for next generation sequencing, genotyping, proteomics with NGS read out and associated bioinformatics support

NGI Platform organisation



Tuuli Lappainen
Platform Director
Professor KTH



Lars Feuk
Platform Co-Director
Professor UU

**NGI-Uppsala
SNP&SEQ
Technology
platform**

**NGI-Uppsala
Uppsala
Genome Center**

NGI-Stockholm



Projects

- Assemblies of high-quality reference genomes
- Human genome variation analyses
- Transcriptome profiling
- Single-cell sequencing and much more

Amount of sequenced base pairs

- 809 Tbp – short reads
- 24 Tbp – high quality long reads
- 10.9 B – genotypes

Technology development

- Evaluation of new protocols, applications, bioinformatics tools and sequencing methods
- Methodological developments in spatial and single-cell transcriptomics technologies

Education and Outreach

- Teaching at courses from undergraduate to PhD level
- Participating in national and international conferences
- Webinars, workshops and hackathons



Samples

- All types of sample sources: from environment, lab cultured, biobank, etc
- All types of organisms: microbes, plants, insects, mammals, ...

Support meetings

- Experimental design
- Advising on sample preparations
- Optimizing sequencing setup
- Guidelines for further data analysis

Publications

- Contribution to a number of articles in high impact journals such as Nature, Cell, Science, Nature Biotechnology, Nature Genetics, Nature Neuroscience, etc.

Users

- Unique project PIs from more than 19 different universities, institutes, healthcare and industry companies used NGI services in 2024

Communication tickets

- 42932 ticket updates
- 99% satisfaction score



NGI services

Multi-omics services

Genome Sequencing
De novo, re-seq, targeted...



Epigenomics
Methylation, chromatin state, HiC...

Pacific Biosciences
Sequel IIe
Revio

Illumina
NovaSeq X plus
NovaSeq 6000
NextSeq
MiSeq



Transcriptomics
Short-read, long-read



Oxford Nanopore
Promethion
Minion

Proteomics
Olink Explore



Proteomics
Olink Explore



SNP Genotyping

Arrays
SNPs, methylation

Source material

Tissues
Cells
Microbes
Plasma
Nucleic acids
Archaeological material
Environmental samples
Read-made libraries

E-infrastructure & pipelines
for FAIR data processing
and management



BSL-2 BSL-3



Ancient DNA



Spatial
Transcriptomics
10x Genomics
Visium

Single-cell
10x Genomics Chromium
Smart-seq

Sequencing instruments at NGI



Short read NGS

High throughput, low cost per base

NovaSeq X Plus

Illumina NovaSeq

Illumina MiSeq

Illumina NextSeq

AVITI, Element Biosciences



Long read NGS

Very long reads, lower throughput

PacBio Revio

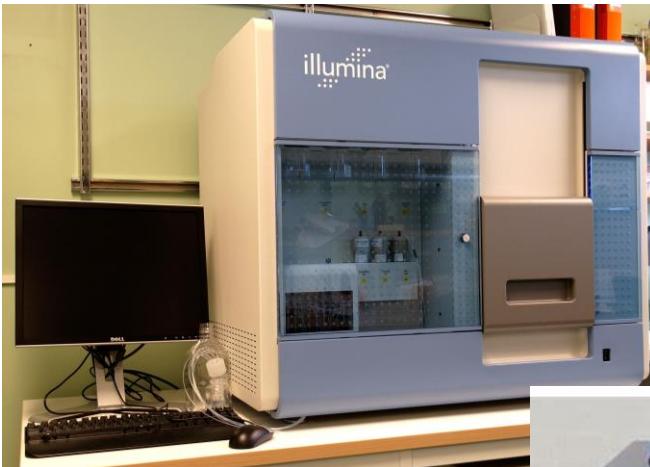
Oxford Nanopore-PromethION



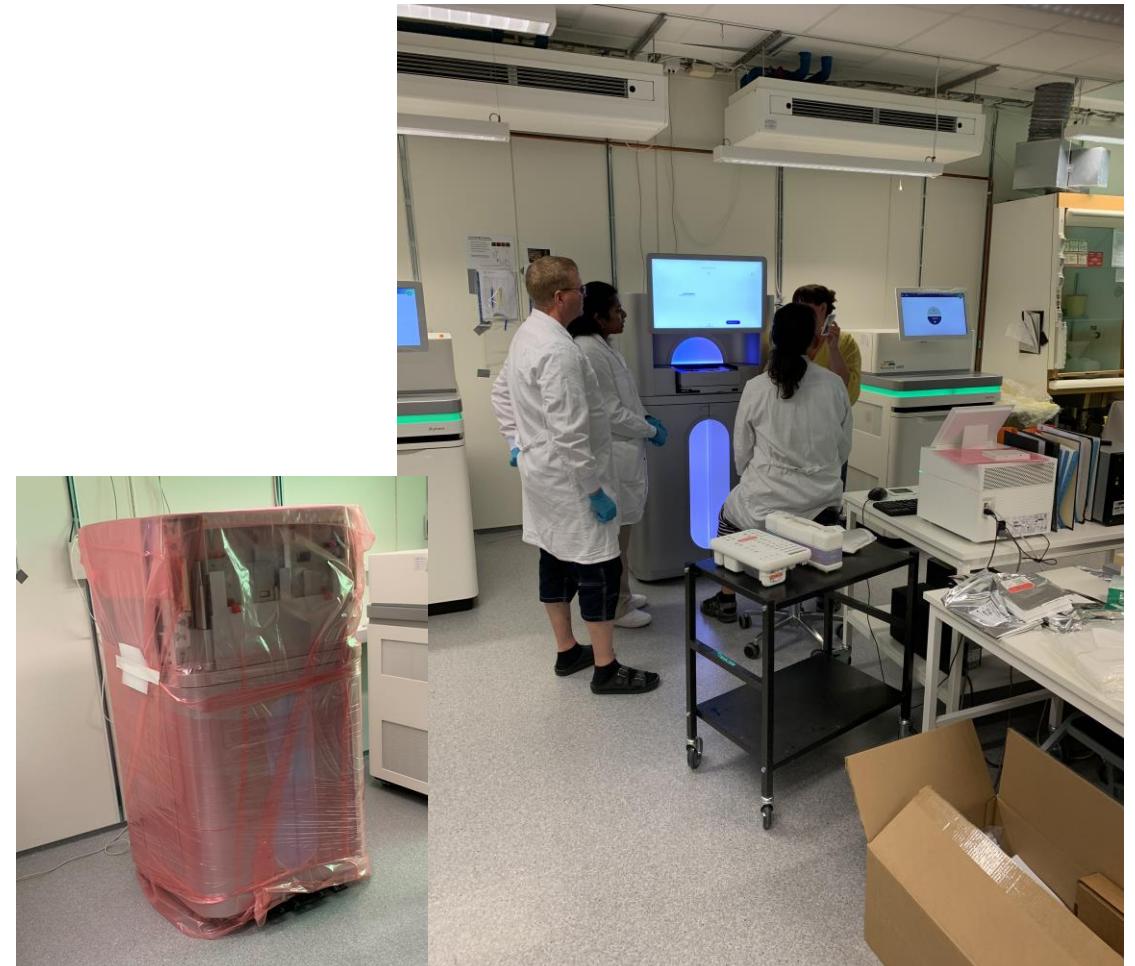
15 years of Illumina sequencing at NGI



2007: Installation of Illumina GA

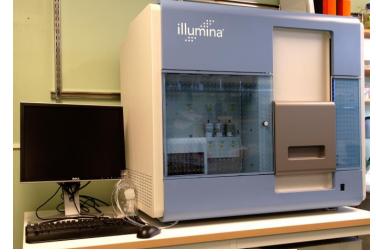


2023: Arrival of NovaSeq X Plus

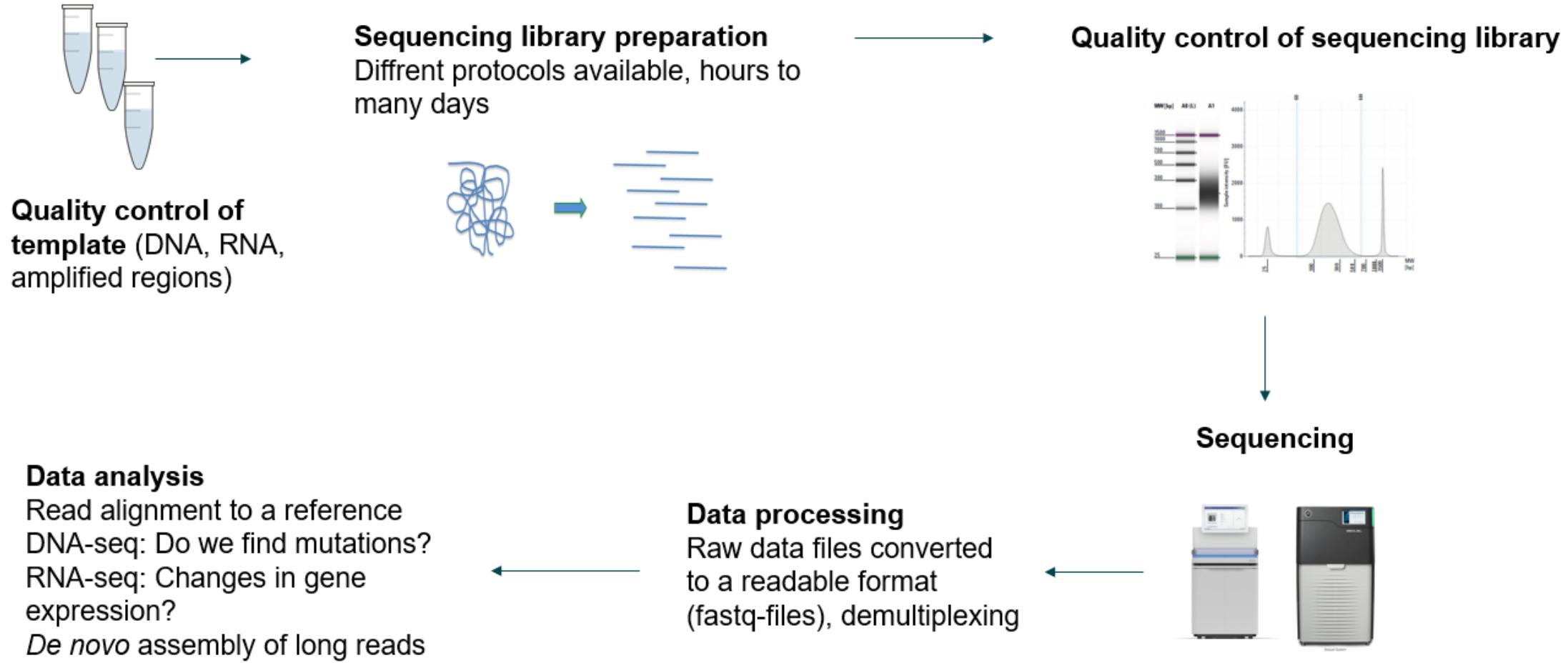


Instruments over the years



Sequencer	Launched	Runtime	Output	Cost per Base	
Illumina Genome Analyzer (GA)	2006	~3 days	~1 Gb per run	~\$0.10	
HiSeq 2000	2010	~10 days	Up to 600 Gb per run	~\$0.01	
HiSeq 2500	2012	~1 day in rapid mode	Up to 600 Gb per run	~\$0.01	
HiSeq X Ten	2014	~3 days per run	~1.8 Tb per run	<\$0.01	
NovaSeq 6000	2017	~13–44 hours	Up to 6 Tb per run	~\$0.005	
NovaSeq X / X Plus	2022	~24–48 hours	Up to 20 Tb per run	<\$0.004	 

Workflow, Illumina sequencing



Library preparation



- A sequencing library is a pool of DNA fragments with adapters attached to both ends of the fragments
- Approx. 25 protocols for Illumina library prep at NGI

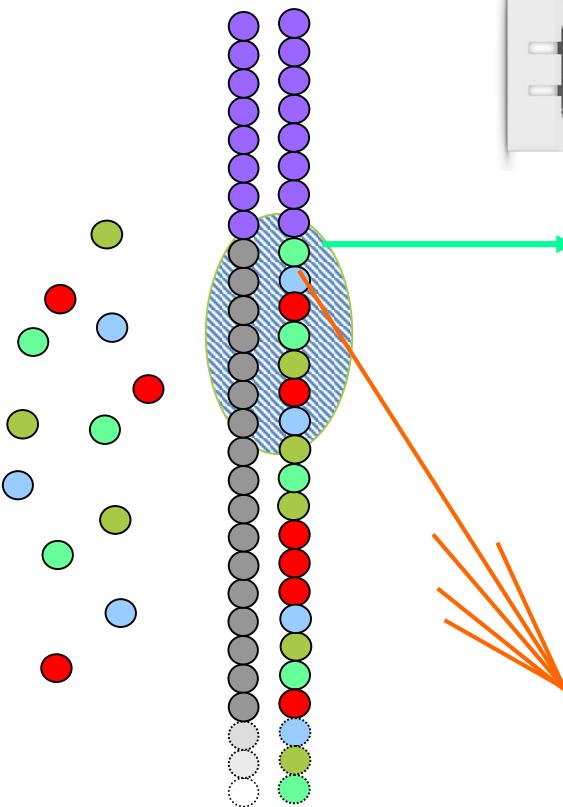
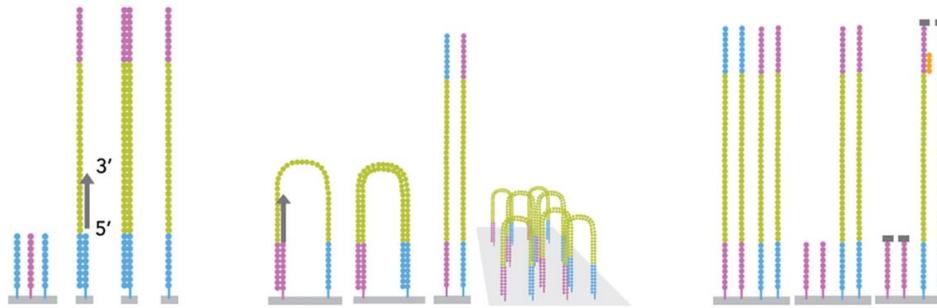
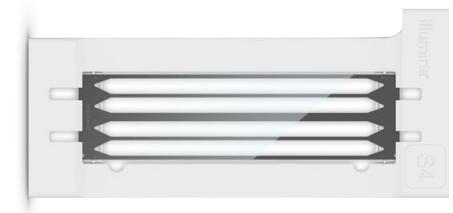
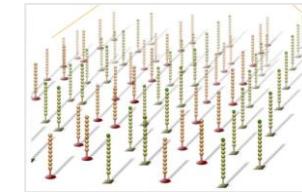


P5 Adapter -- [Index 1] -- [Read 1 Primer Site] -- [Insert DNA]
-- [Read 2 Primer Site] -- [Index 2] -- P7 Adapter

Illumina cluster generation & sequencing



- The sequencing library is hybridized to a flowcell ("cluster generation")
 - - A flowcell is a slide that is coated with oligos
- Rapid bridge amplification
- Hybridization of sequencing primers
- Sequencing by synthesis
 - fluorophore labeled nucleotides emitting light

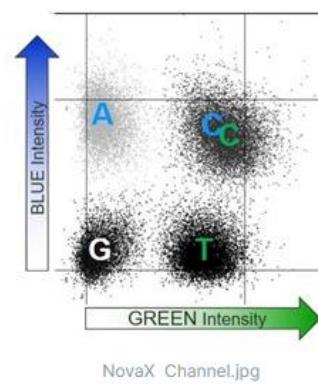


Illumina sequencing by synthesis



4-Channel Chemistry				
	A	G	T	
Image 1	●			
Image 2		●		
Image 3			●	
Image 4				
Result	A	G	T	C

2-Channel Chemistry				
	A	G	T	
Image 1	●			
Image 2		●		
Result	A	G	T	C



Youtube:
<https://www.youtube.com/watch?v=fCd6B5HRaZ8>

NovaSeq X Plus – new instrument!



Flowcell Type	1.5 B	10 B	25 B
Output per flowcell (paired end150 bp)	500 Gb	3 Tb	8 Tb
Number of human genomes per flowcell	~ 4	~ 24	~ 64
Run time (paired end150 bp)	21 h	24 h	48 h

Run ID - Lane	Mb Total Yield	M Total Clusters	% bases ≥ Q30
20230612_LH00179_0005_A2255M2LT3 - L1	295 764.0	979.4	95.4%
20230612_LH00179_0005_A2255M2LT3 - L2	323 896.8	1 072.5	95.3%
20230612_LH00179_0005_A2255M2LT3 - L3	366 557.1	1 213.8	95.6%
20230612_LH00179_0005_A2255M2LT3 - L4	383 028.6	1 268.3	95.0%
20230612_LH00179_0005_A2255M2LT3 - L5	251 454.3	832.6	97.3%
20230612_LH00179_0005_A2255M2LT3 - L6	284 351.5	941.6	97.1%
20230612_LH00179_0005_A2255M2LT3 - L7	388 065.2	1 285.0	94.0%
20230612_LH00179_0005_A2255M2LT3 - L8	363 776.7	1 204.6	95.0%

Advantages and challenges NovaSeqX

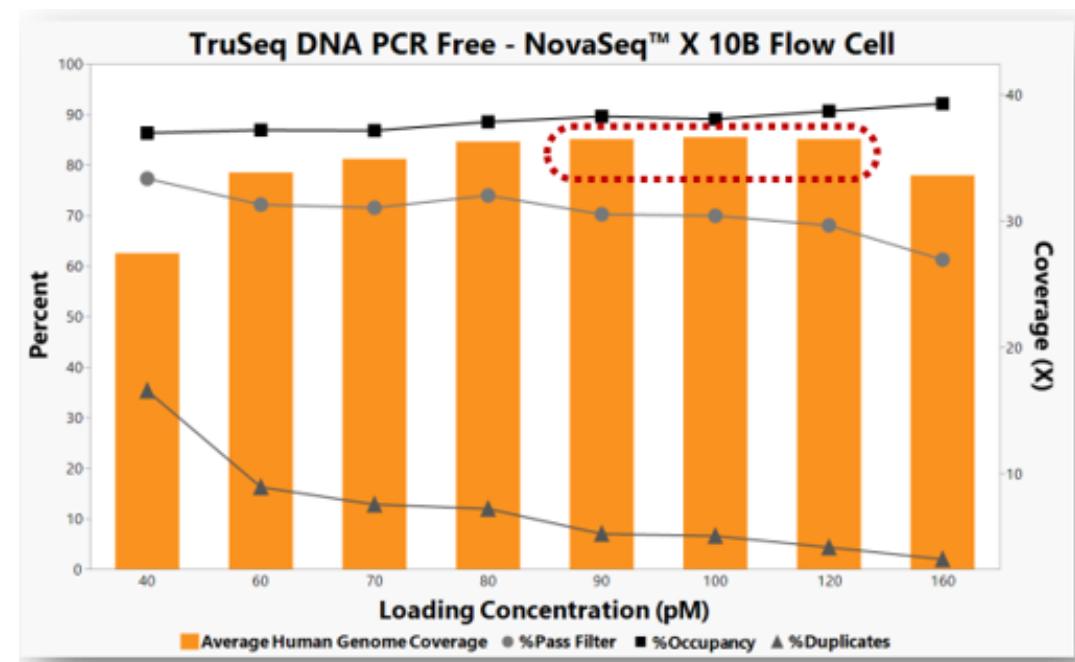


+

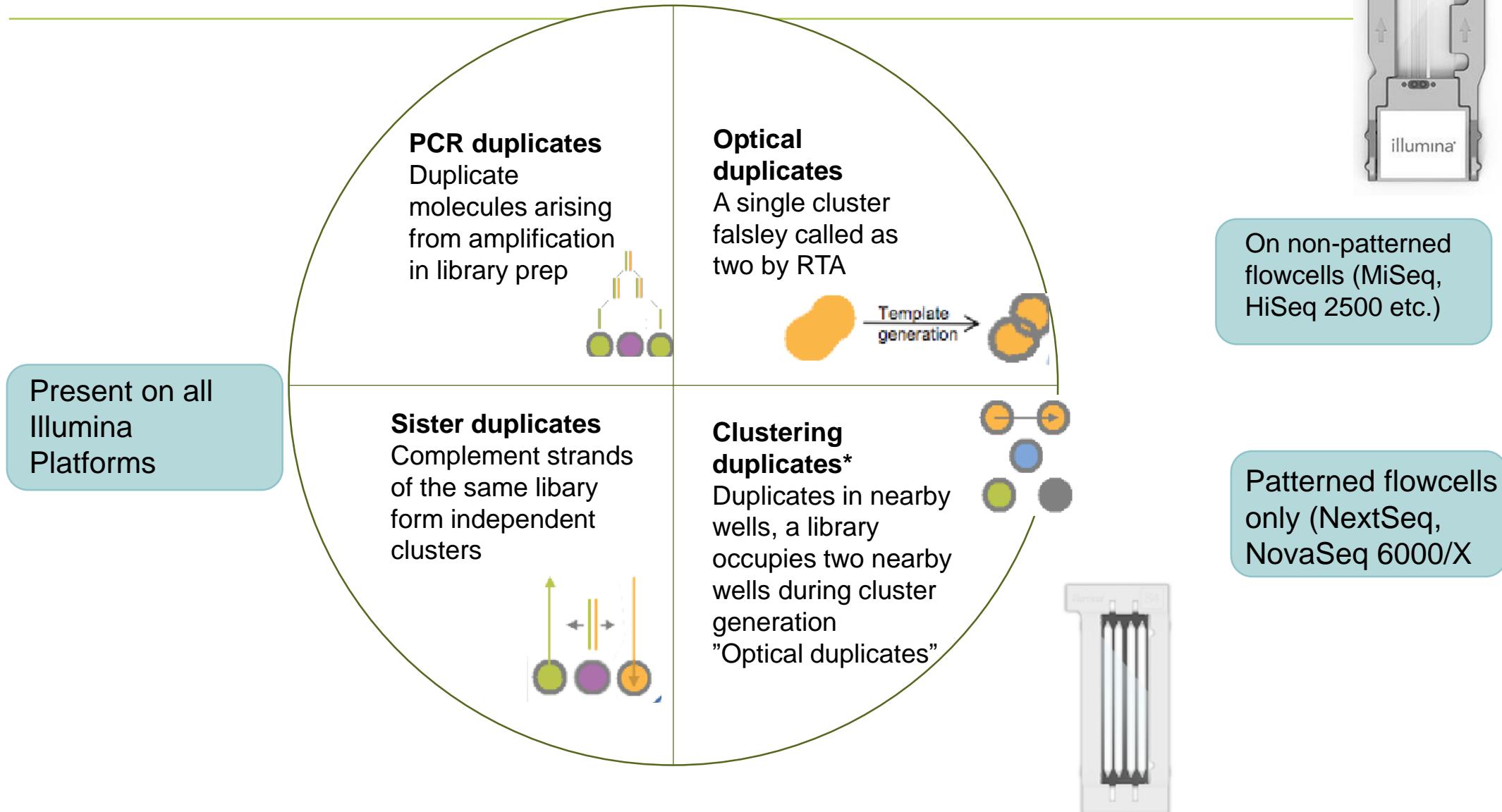
- Cost per base is low
- Quick data generation
- Easy workflow in the lab
- Reagents shipped in RT
- On-instrument analysis

-

- Yield vs duplicates
- More sensitive to challenging samples and short inserts
- Sensitive to colour balancing (C-A)



Duplicates, duplicates, duplicates....



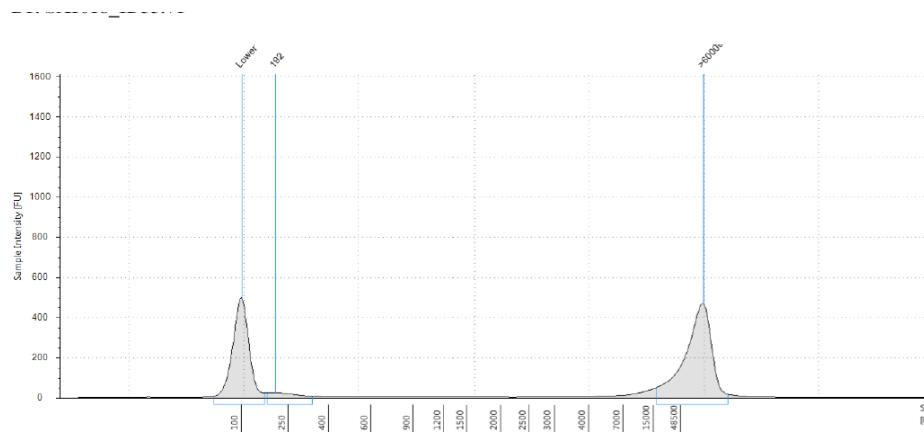
Quality control of RNA/DNA



DNA

Concentration: QuantIT

Degradation: Fragment Analyzer/TapeStation



Sample Table

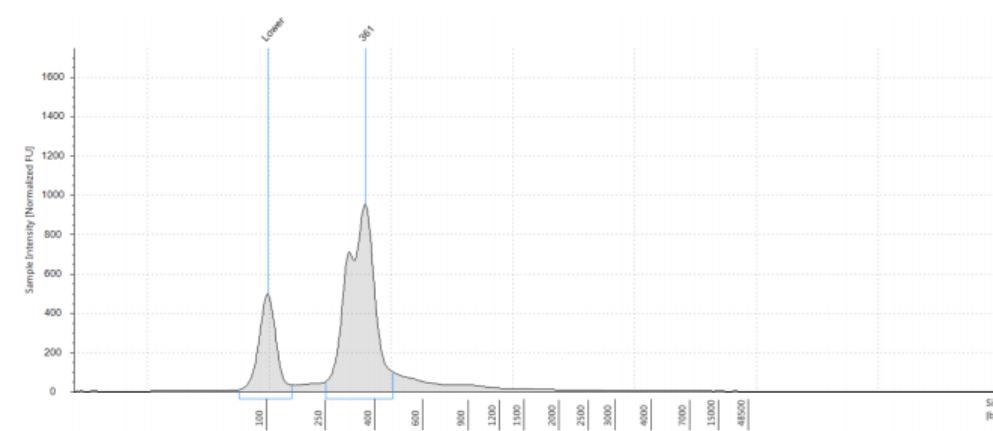
Well	DIN	Conc. [ng/ μ l]	Sample Description	Alert	Observations
B1	9.6	16.0	SXI018_ID33.v1		

High quality DNA sample

RNA

Concentration + RIN-value:

Fragment Analyzer/TapeStation



Sample Table

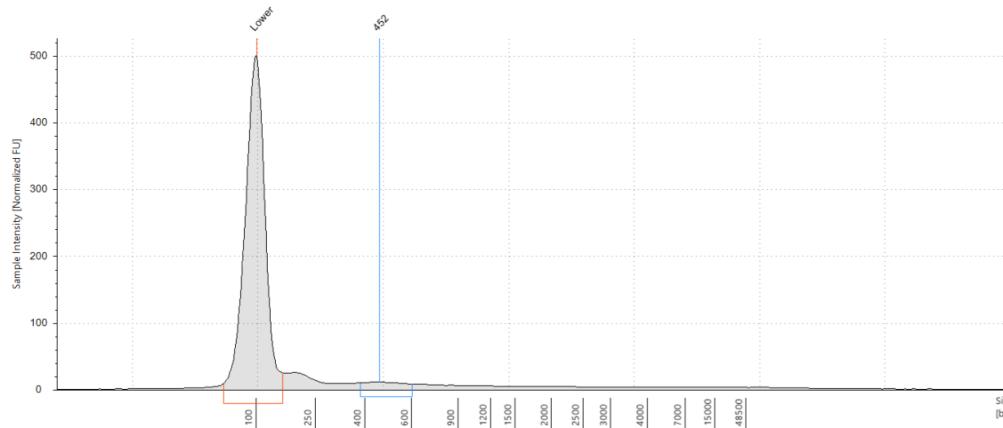
Well	DIN	Conc. [ng/ μ l]	Sample Description	Alert	Observations
E1	1.0	33.0	92-291039_RJ-1964-pool3		

Degraded DNA sample

Quality of sample/library will affect sequencing result!



DNA-sample: 2.5 ng/ul, DIN-value 0



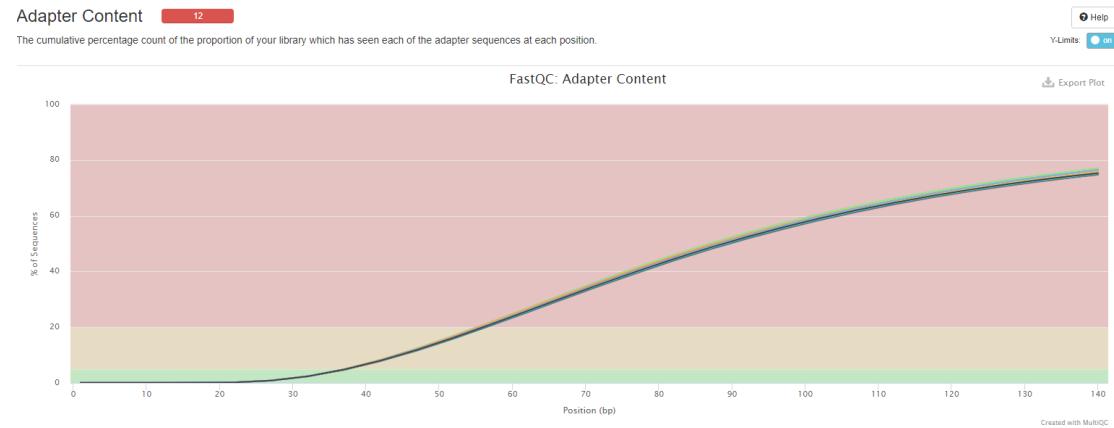
Sample Table

Well	DIN	Cone. [ng/ul]	Sample Description	Alert	Observations
A1	-	2.46	SXI162_Sl.v1	⚠	Sample concentration outside functional range for DIN

20 ng of DNA, Thurplex Low-input library prep, 3 libraries

Amount of data generated: 800 M read pairs (aiming for $\geq 60x$ coverage)

Result: 12x coverage



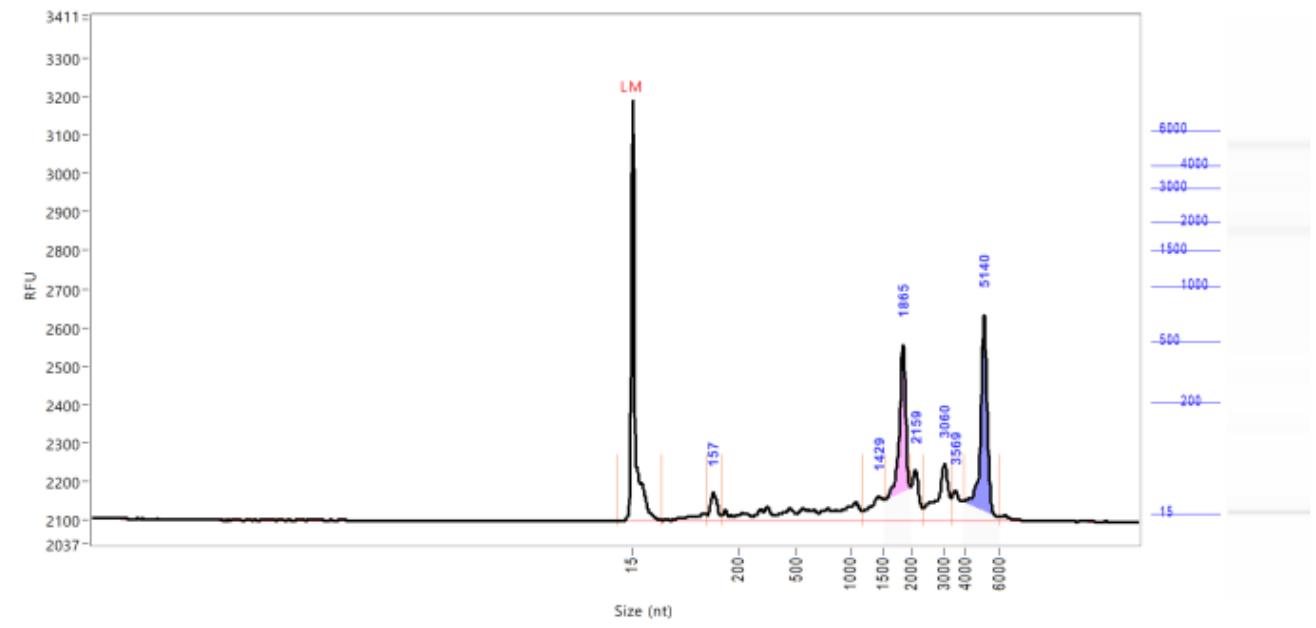
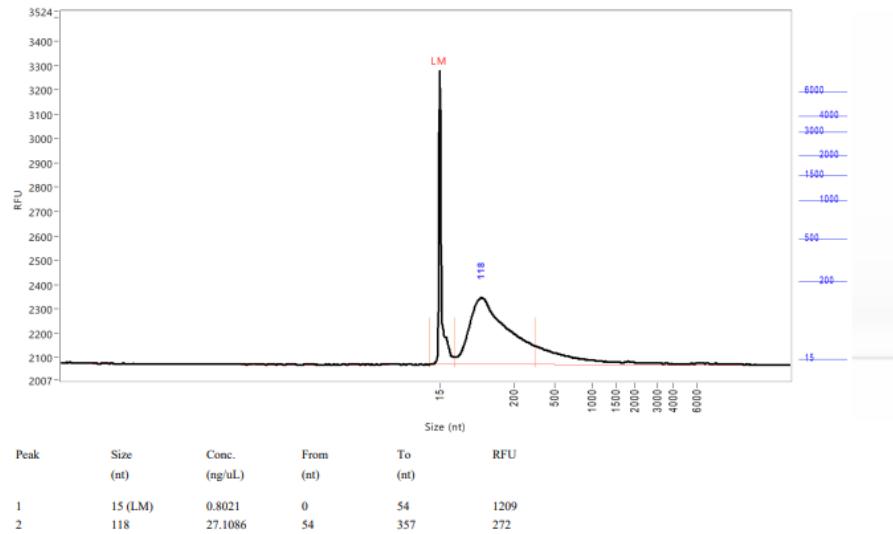
[Copy table](#) [Configure Columns](#) [Plot](#) Showing 7/7 rows and 14/23 columns.

Sample Name	% GC	Ins. size	$\geq 30X$	Coverage	% Aligned	Change rate	Ts/Tv	M Variants	TiTV ratio (known)	TiTV ratio (novel)	% Dups	% Dups	% GC	M Seqs
S1	46%	55	11.1%	2.0X	98.2%	893	1.645	3.47	2.0	1.6	76.6%	1.6	46%	1.6

Quality of sample/library will affect sequencing result!



- RNA samples, RIN-values between 1-9,6
- Library prep Illumina Ligation Ribo-Zero Plus



Results on next page...

Continued... Quality of sample/library will affect sequencing result



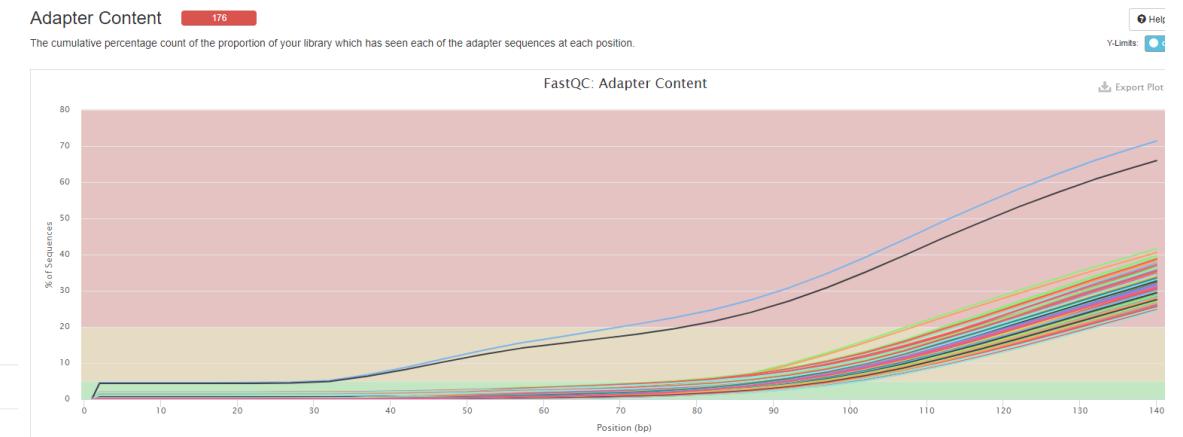
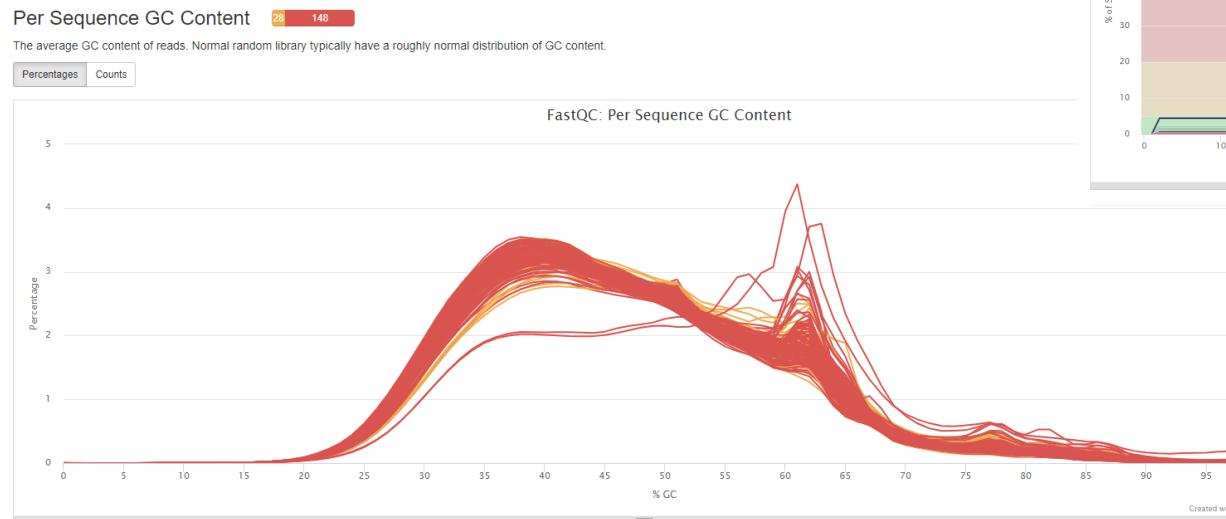
QC-results RNA-seq

Uneven amounts of data (17-100 M reads per sample)

A lot of duplicates

High rRNA content

High adapter content

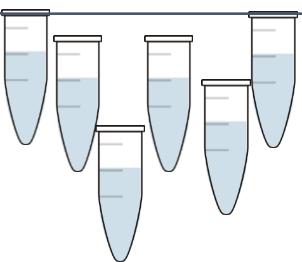


Some of the applications offered



Templates:

DNA, RNA, cells,
serum or plasma



Whole Genome Sequencing (WGS)

- *De novo* sequencing (PacBio, ONT)
- Re-sequencing (PCR-Free, low input)

Transcriptome Sequencing

- mRNA-Seq (poly-A selection)
- Total RNA-seq (ribosomal depletion)
- miRNA & small RNAs
- Full-length transcriptomes

Targeted re-sequencing

- Exome
- Gene panels
- Amplicons (including bacterial 16S for metagenomics)
- RAD-seq

Epigenetics

- Chromatin (HiC, ATAC-Seq)
- WGBS
- ChIP Sequencing

Ready-made libraries

- User-made libraries
- High throughput
- Fast turn around time

Single-cell applications

- 10x Genomics
- Dolomite Nadia
- Single-cell WGBS

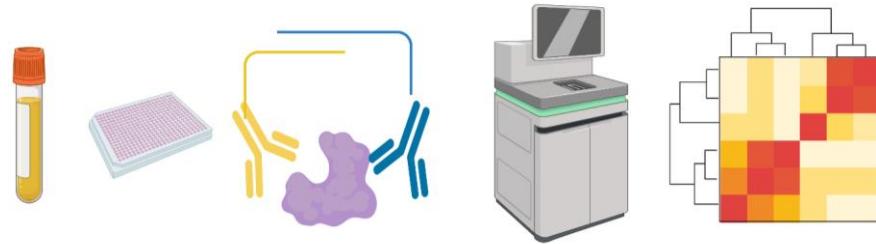
Spatial transcriptomics

- 10x Genomics Visium

Proteomics with NGS readout

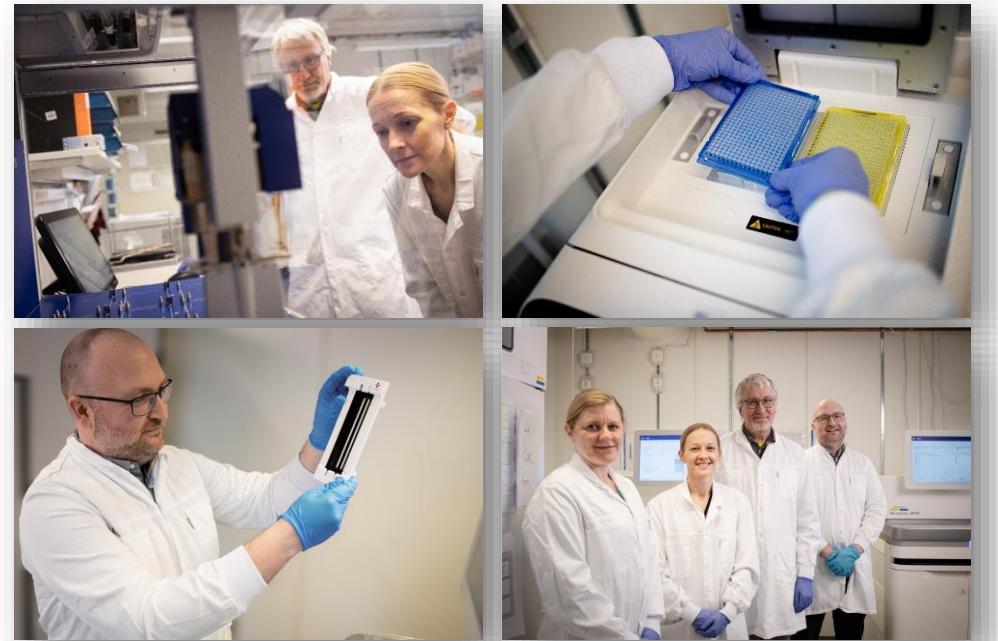
- Olink Explore 1536/3072/5300

Protein analysis, Olink Explore with NGS readout



- Highly multiplex protein biomarker analysis:
 - Olink Explore 384-5300 protein assays available
 - Cardio-metabolic
 - Inflammation
 - Neurology
 - Oncology
- Stats
 - >25 000 samples analyzed 2021-2023
 - >30 000 samples analyzed in 2024

SciLifeLab Explore Lab: NGI in collaboration with the Affinity Proteomics Uppsala unit and Olink Proteomics AB



New instrument – AVITI, Element Biosciences



Category	Specification
Technology	Sequencing by Binding (SBB)
Applications	Genomics, transcriptomics, single-cell sequencing
Read Lengths	Up to 2×150 bp (paired-end reads)
Data Output	Up to 300 Gb per run
Run Time	24-36 hours
Library Compatibility	Standard NGS library preparation protocols
Limitations	Not ideal for ultra-high-throughput projects



Examples, recent successful projects



Massively parallel analysis of single-molecule dynamics on next-generation sequencing chips

J. AGUIRRE RIVERA , G. MAO , A. SABANTSEV , M. PANFILOV , Q. HOU , M. LINDELL , C. CHANEZ , F. RITORT , M. JINEK , AND S. DEINDL

[Authors Info & Affiliations](#)

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5,195



nature genetics

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Comment | Published: 21 October 2024

Pushing the boundaries of rare disease diagnostics with the help of the first Undiagnosed Hackathon

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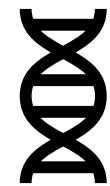
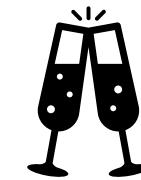
1768 Accesses | 9 Altmetric | [Metrics](#)

NGI OpenLab – opening soon!



New interactive NGS space opening at BMC in Uppsala
January 2025

Launch party at BMC Jan 16, 14.00-16.00





Long-read sequencing, data analysis pipelines, and development projects at NGI

Adam Ameur

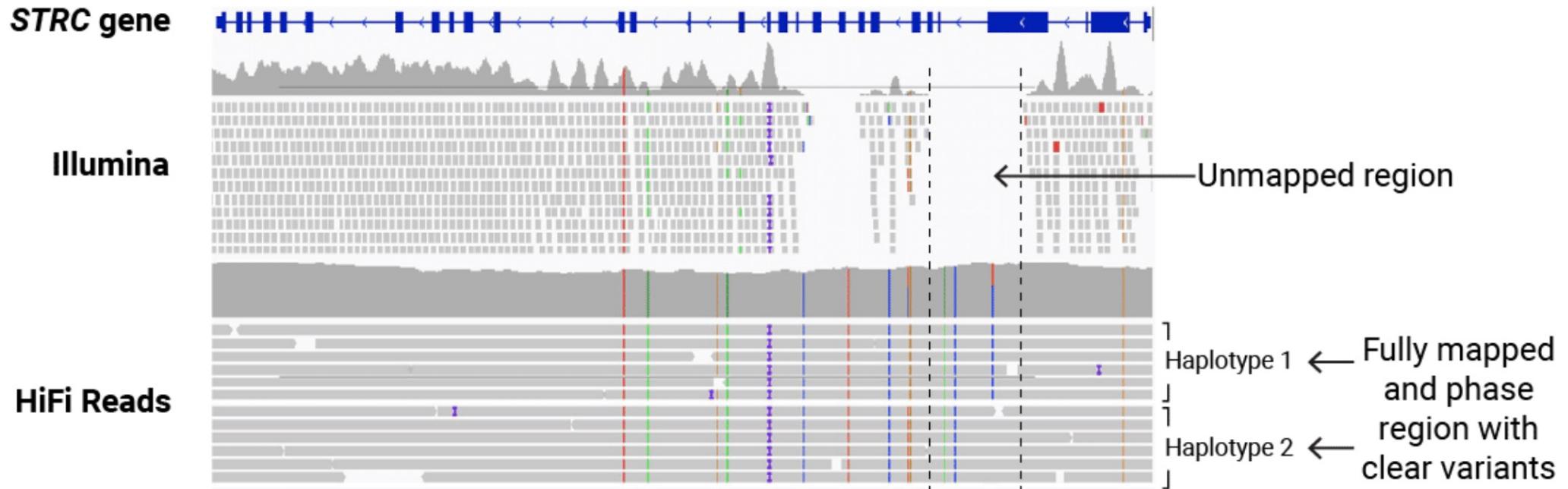
National Genomics Infrastructure, SciLifeLab, Uppsala, Sweden





Limitations with short reads

- You don't get complete genome information!



Long-read sequencing

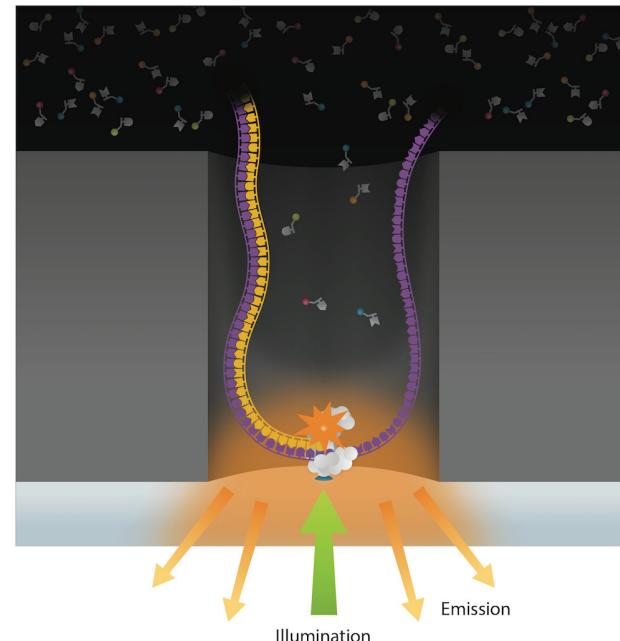
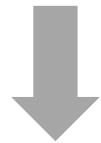
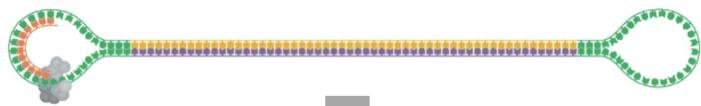


No longer a niche technology!

- Assemble complete genomes
- Find all genetic variants
- Detect epigenetic modifications
- At a “reasonable” cost



PacBio Sequencing



PacBio RSII



PacBio Sequel
(Sequel I & II)



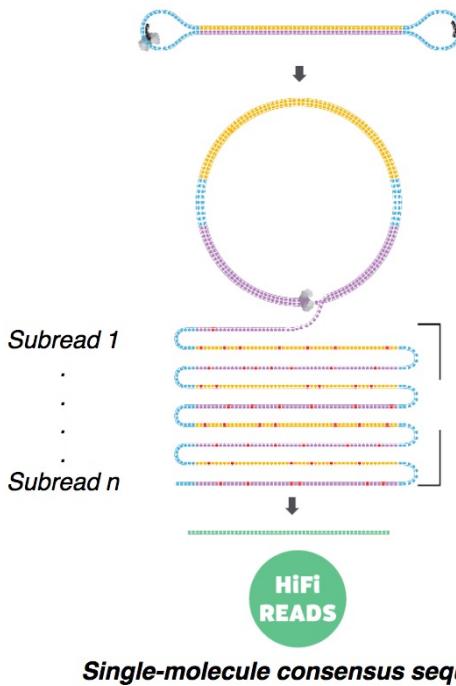
PacBio Sequencing



TWO MODES OF SMRT SEQUENCING

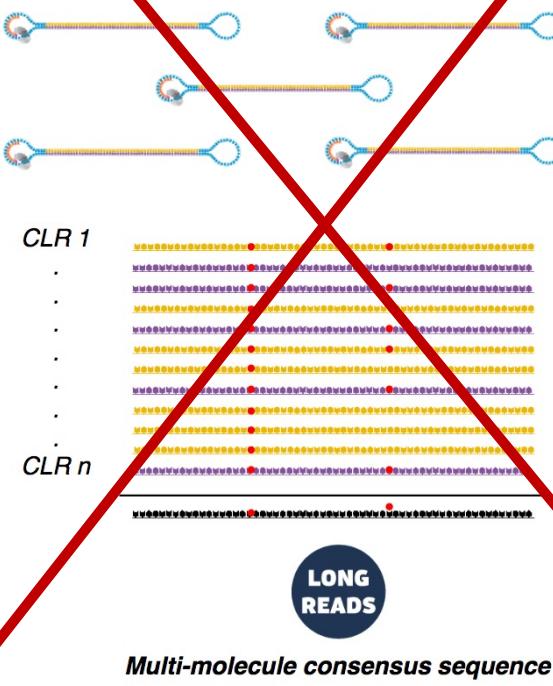
Circular Consensus Sequencing (CCS) Mode

Inserts 10-20 kb



Continuous Long Read (CLR) Sequencing Mode

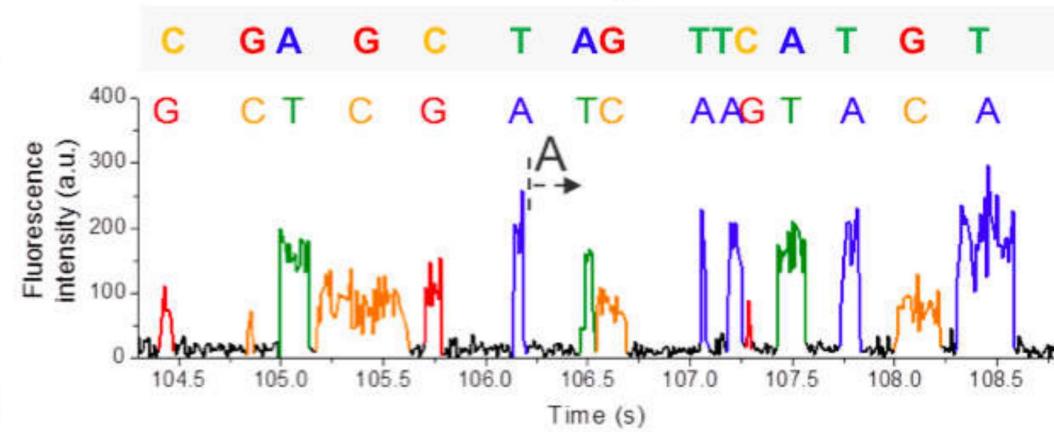
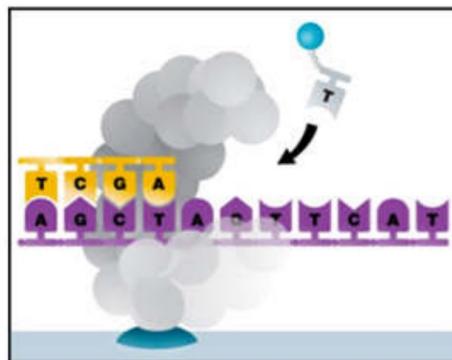
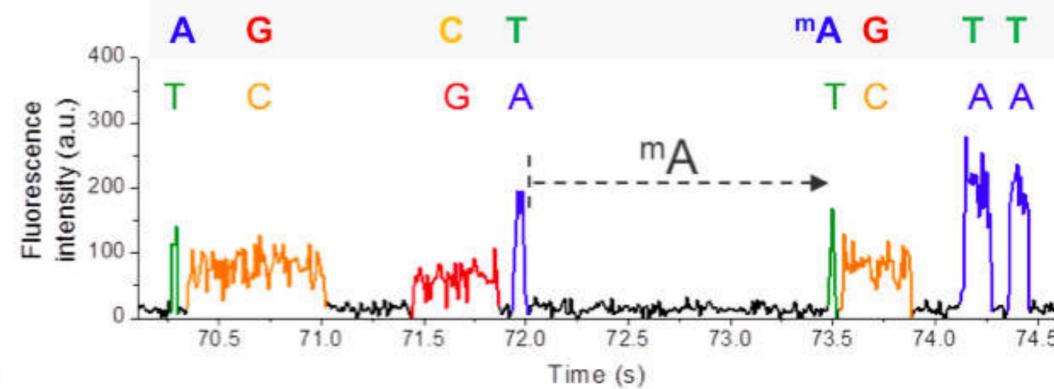
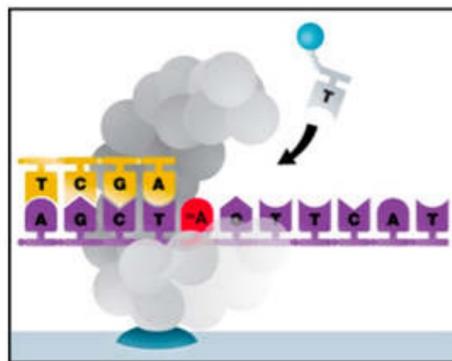
Inserts >25 kb, up to 175 kb



PacBio – Methylation detection



- Base modifications on native DNA molecules can be detected!



A decade of PacBio sequencing at NGI



2013: Installation of PacBio RSII



2023: Arrival of PacBio Revio



The PacBio Revio System



- Up to 90Gb data from one SMRT cell
- Read lengths: 15-20kb
- >QV20 quality (>99% read accuracy)
- Can run 1,300 human genomes/year!



The PacBio Revio System – Update 2025



120Gb

- Up to ~~90Gb~~ data from one SMRT cell
- Read lengths: 15-20kb
- >QV20 quality (>99% read accuracy)
- Can run ~~1,300 human genomes/year!~~
2,500 human genomes



Revio – results for our first 16 runs

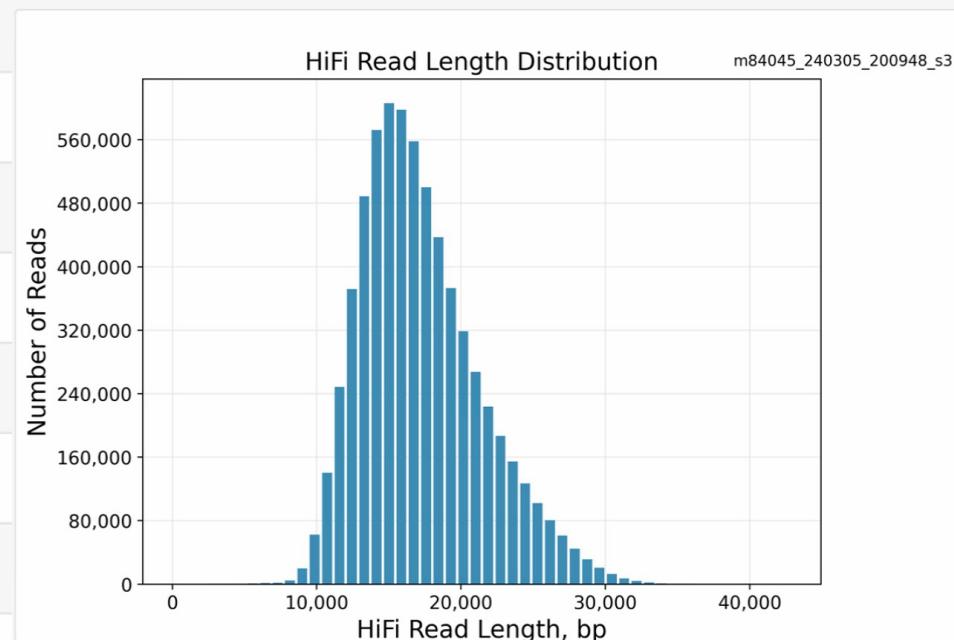


Sample/Species/Proj	Number of reads	Total yield (Gbp)	Average read length (kb)	Size selection method	Comment
Human 1_1	6,873,030	84.7	12.3	Ampure beads	Also Sequel II data
Human 1_2	6,846,419	102.2	15.0	Ampure beads	Also Sequel II data
Human 1_3	7,170,075	90.3	12.6	Ampure beads	Also Sequel II data
Human 1_4	6,015,366	67.6	11.2	Ampure beads	Also Sequel II data
Human 2_1	6,895,775	104.2	15.1	SageELF (2 fract. pooled)	
Human 2_2	5,684,755	100.3	17.6	SageELF (2 fract. pooled)	
Human 2_3	6,022,465	111.5	18.5	SageELF (2 fract. pooled)	
Human 3_1	7,544,871	72.3	9.6	Ampure beads	
Human 3_2	7,857,802	65.6	8.3	Ampure beads	
Human 3_3	7,164,744	102.3	14.3	Ampure beads	
Human 3_4	6,695,524	82.4	12.3	Ampure beads	
Human 3_5	6,541,509	80.4	12.3	Ampure beads	
Plant 1_1	7,683,014	70.1	9.1	Ampure beads	Also Sequel II data
Amphibian 1_1	2,700,447	23.5	8.7	Ampure beads	225 pM loading
Amphibian 1_1	5,219,472	42.3	8.1	Ampure beads	350 pM loading
Bird 1_1	6,812,139	90.2	13.2	Ampure beads	

Example of a good run > 114 Gb



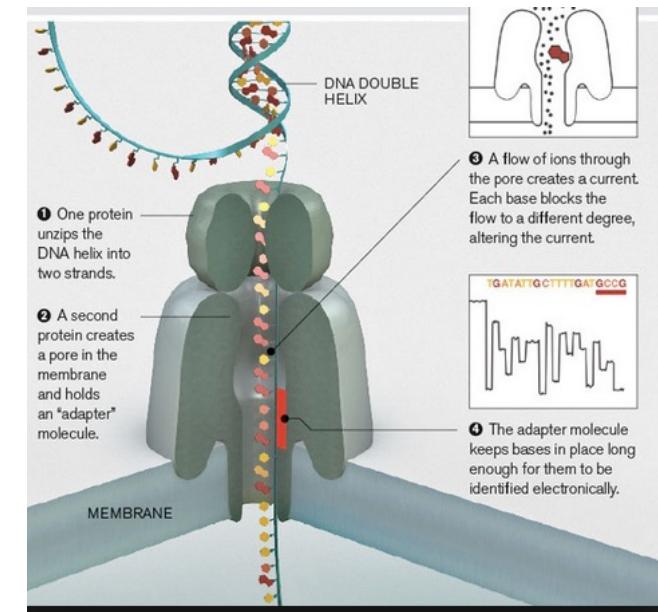
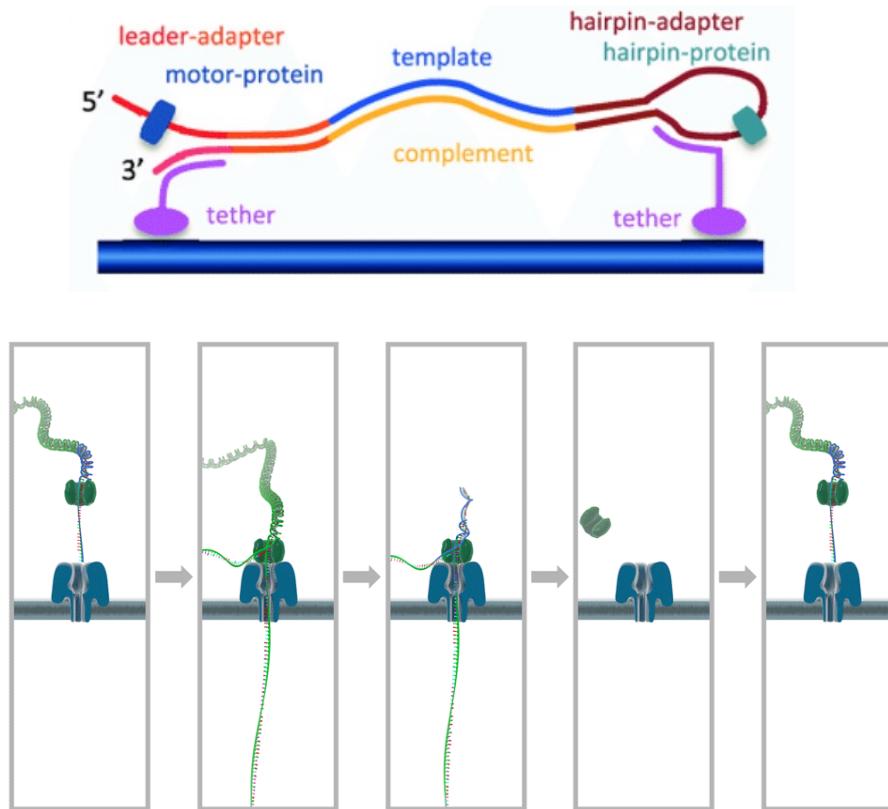
Value	Analysis Metric
6.6 M	HiFi reads
114.17 Gb	HiFi reads yield
17.21 kb	HiFi reads length (mean)
16,564	HiFi reads length (median, bp)
17,585	HiFi Read Length N50 (bp)
Q34	HiFi Read Quality (median)
92.36%	Base Quality \geq Q30 (%)
8	HiFi Number of Passes (mean)



Example: Data at a translocation site



Oxford Nanopore sequencing



Base modification info is retained

Oxford Nanopore sequencing



Instrument	Run time / FC	Output / FC	Nr of pores	Max read length
Flongle	16 hrs	1 Gb	126	1 Mb
MinION	24 hrs	2-15 Gb	512	1 Mb
GridION	24 hrs	2-15 Gb	512	1 Mb
PromethION	72 hrs	10 – 150 Gb	3 000	2 Mb

ONT - Portability



The International Space Station

In 2016, MinION was used to conduct the first ever DNA sequencing in space. MinION performance was unaffected by the flight to the International Space Station (ISS) or microgravity conditions. The team stated that '*these findings illustrate the potential for sequencing applications including disease diagnosis, environmental monitoring, and elucidating the molecular basis for how organisms respond to spaceflight!*' Further to this, in 2020, an end-to-end sample-to-sequencer workflow conducted entirely aboard the ISS resulted in off-Earth identification of microbes for the first time.

Photograph: NASA ©

[Read more >](#)



Entirely off-grid, solar-powered sequencing

In 2019, Gowers *et al.* used MinION to demonstrate '*the ability to conduct DNA sequencing in remote locations, far from civilised resources (mechanised transport, external power supply, internet connection, etc.), whilst greatly reducing the time from sample collection to data acquisition!*' The team transported their portable lab for 11 days using only skis and sledges across Europe's largest ice cap (Vatnajökull, Iceland), before carrying out a tent-based study, resulting in 24 hours of sequencing data using solar power alone.

[Read more >](#)

Uncovering cryptic transmission of Zika virus

The origin and epidemic history of Zika virus (ZIKV) in Brazil and the Americas remained poorly understood despite observed trends in reported microcephaly. Using a mobile genomics lab to conduct genomic surveillance of ZIKV, the team identified the earliest confirmed ZIKV infection in Brazil. Analysis of these genomes estimated that ZIKV is likely to have disseminated from north-east Brazil in 2014, before the first detection in 2015, indicating a period of pre-detection cryptic transmission that would not have been identified without genomic sequencing.

[Read more >](#)



Credit: Nuno R. Faria



ONT - Speed



New DNA Sequencing Tech

January 17, 2022 |

[Tweet](#) [Share 1](#) [Share](#) [Email](#)

A new ultra-rapid genome sequencing approach developed by researchers and their clinical collaborators was used to diagnose rare genetic conditions that were previously unheard of in standard clinical care.

"A few weeks is what most clinicians call 'rapid' when they receive results," said Euan Ashley, MB, professor of medicine at the University of California, San Francisco.

Genome sequencing allows scientists to see a person's entire genome, from the DNA sequence to the proteins it encodes, everything from eye color to inherited diseases. "The key is that all diseases are rooted in their DNA: Once doctors know the specific changes in a patient's genome, they can quickly figure out what's wrong and how to treat it," says Ashley.

Now, a mega-sequencing approach devised by Ashley and his team has sped up the process of diagnosis: Their fastest diagnosis was made in just 12 hours. This means that patients who need less time in critical care units, require fewer tests, and can be treated earlier.

A paper describing the researchers' work is published online in *Nature*. John Gorzynski, DVM, PhD, is the senior author of the paper. Postdoctoral scholar John Gorzynski, DVM, PhD, is the lead author.

nature

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nature > articles > article

Article | [Open access](#) | Published: 11 October 2023

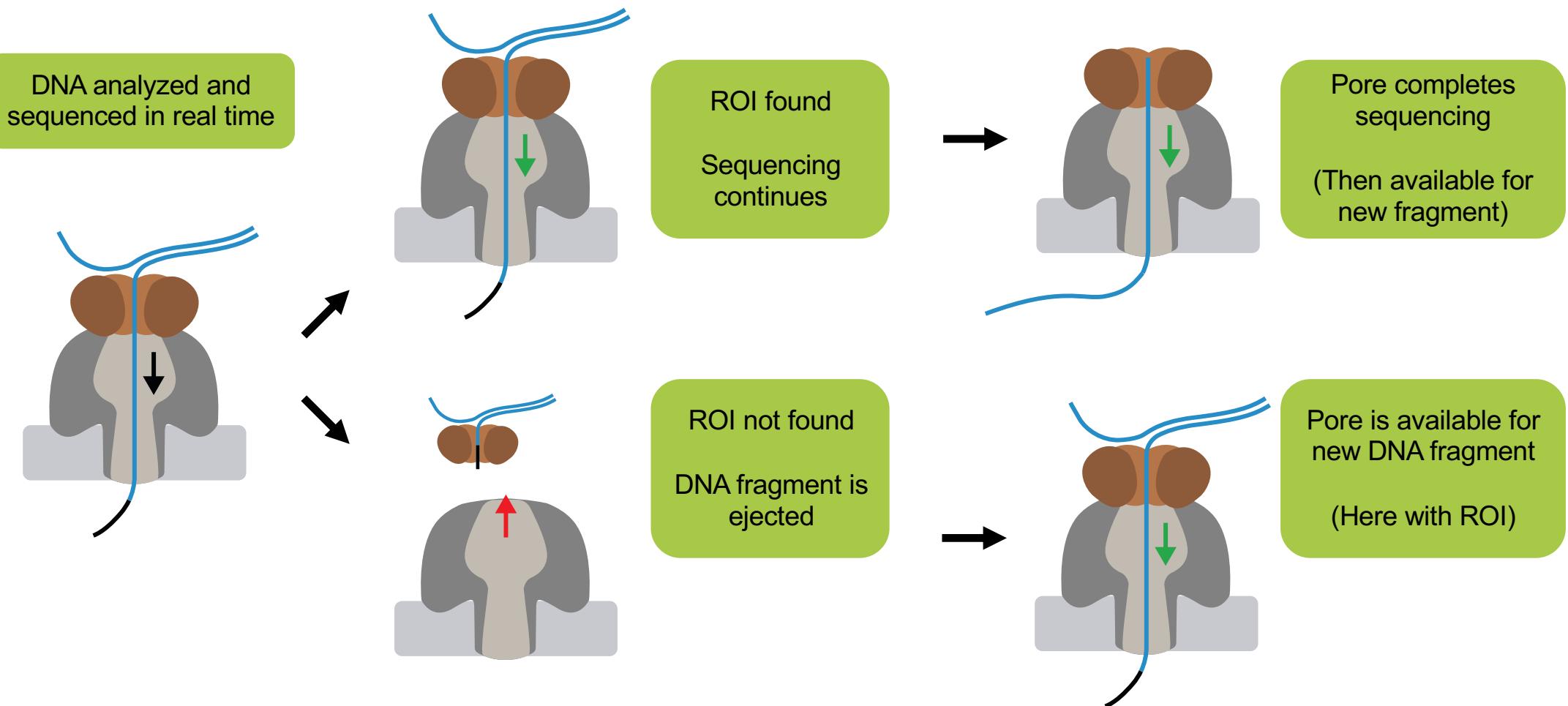
Ultra-fast deep-learned CNS tumour classification during surgery

[C. Vermeulen](#), [M. Pagès-Gallego](#), [L. Kester](#), [M. E. G. Kranendonk](#), [P. Wesseling](#), [N. Verburg](#), [P. de Witt Hamer](#), [E. J. Kooi](#), [L. Dankmeijer](#), [J. van der Lugt](#), [K. van Baarsen](#), [E. W. Hoving](#), [B. B. J. Tops](#)✉ & [J. de Ridder](#)✉

Nature 622, 842–849 (2023) | [Cite this article](#)

34k Accesses | 563 Altmetric | [Metrics](#)

ONT target sequencing - adaptive sampling

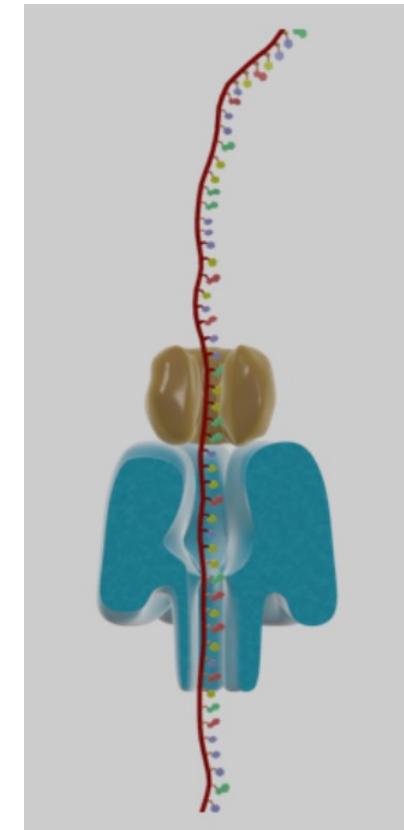


ONT direct RNA sequencing



ONT can sequence native RNA molecules!

- No bias due to cDNA conversion
- Allows to study RNA modifications
- Higher error rate
- Lower throughput



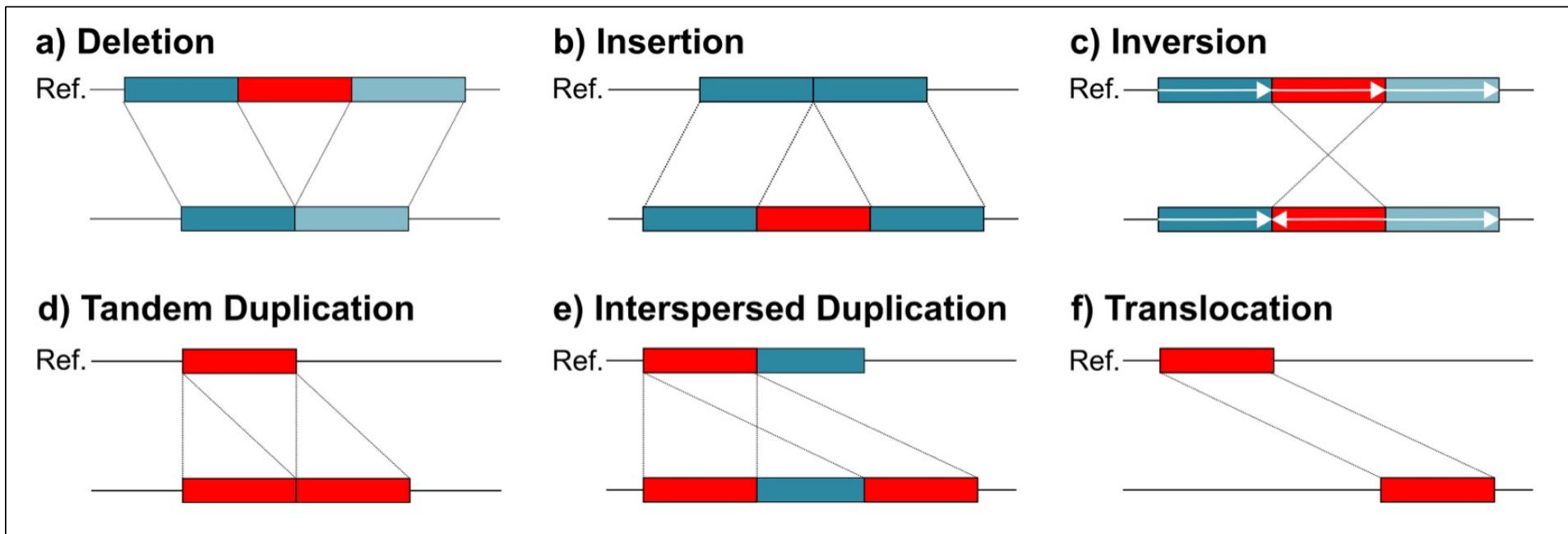
What people are using long reads for...



Example 1: Detect all genetic variants



Long-read sequencing can detect more genetic variants than with short reads:



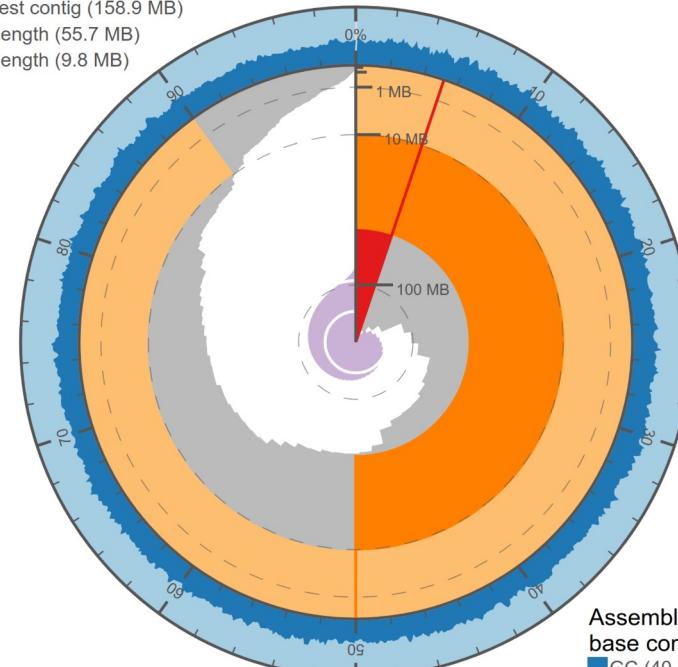
Example 2: Assemble complete genomes



It took just **3.5 h** on a **96** core compute node for *de novo* assembly of a human sample!

span (Gbp)	3.1
GC (%)	40.84
AT (%)	59.16
longest contig (Mbp)	159
contig count	373
contig N50 length (Mbp)	56
contig N50 count	17
contig N90 length (Mbp)	10
contig N90 count	59

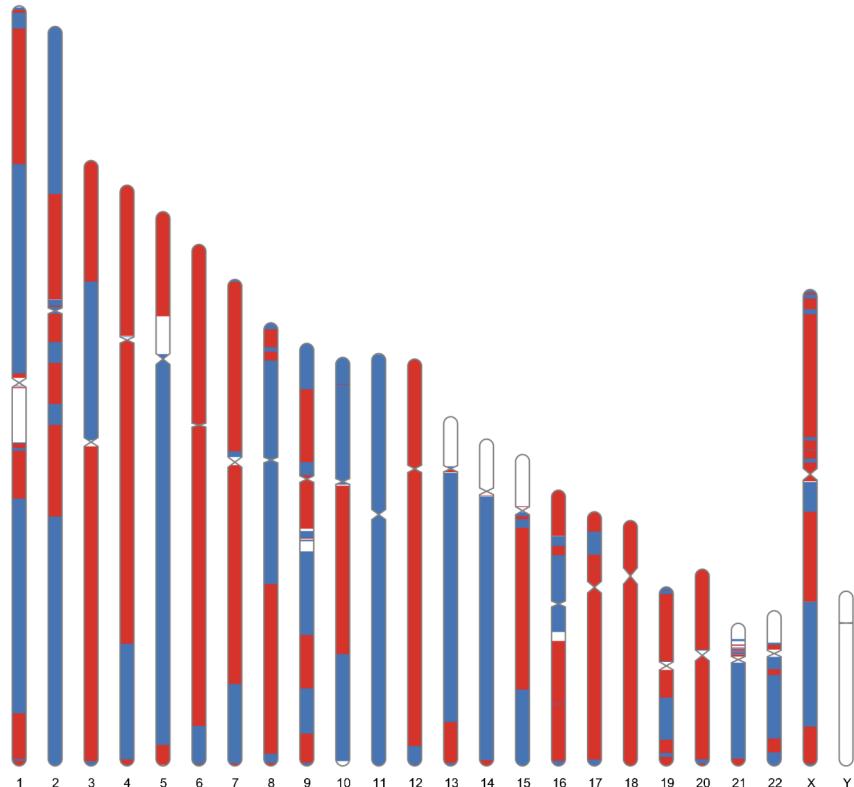
- Contig statistics
- Log₁₀ contig count (total 373)
 - Contig length (total 3 GB)
 - Longest contig (158.9 MB)
 - N50 length (55.7 MB)
 - N90 length (9.8 MB)



Assembly
base composition
GC (40.8%)
AT (59.2%)

Ignas Bunikis

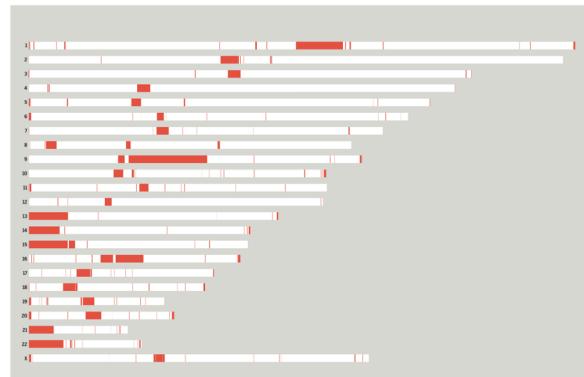
De novo assembly mapped to GRCh38



Colour change represents adjacent contigs

Chromosomes **11** and **18** were assembled in single contigs

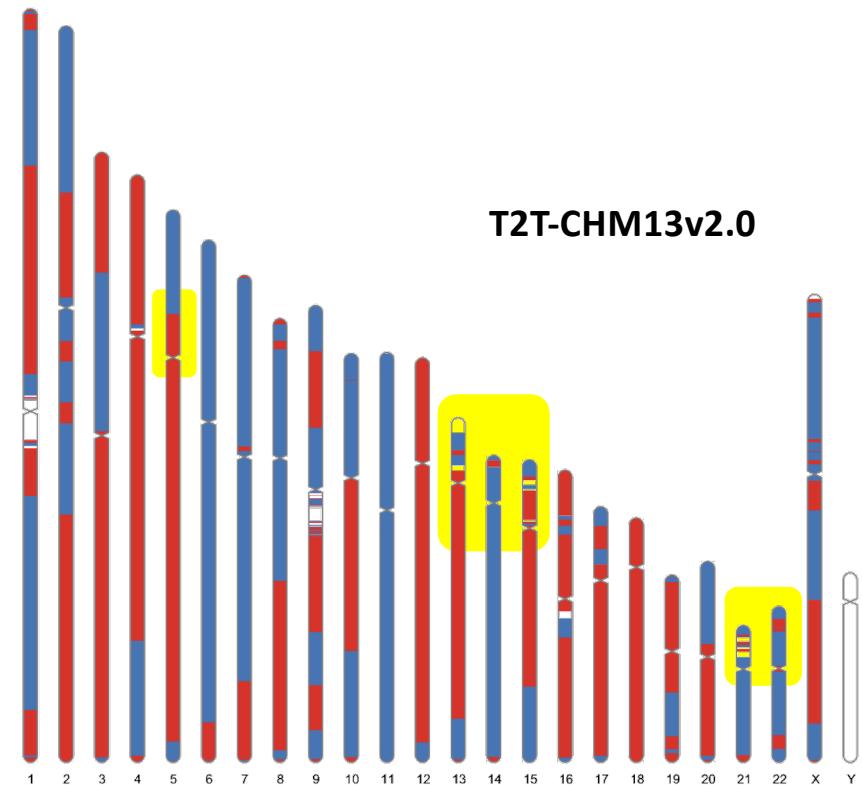
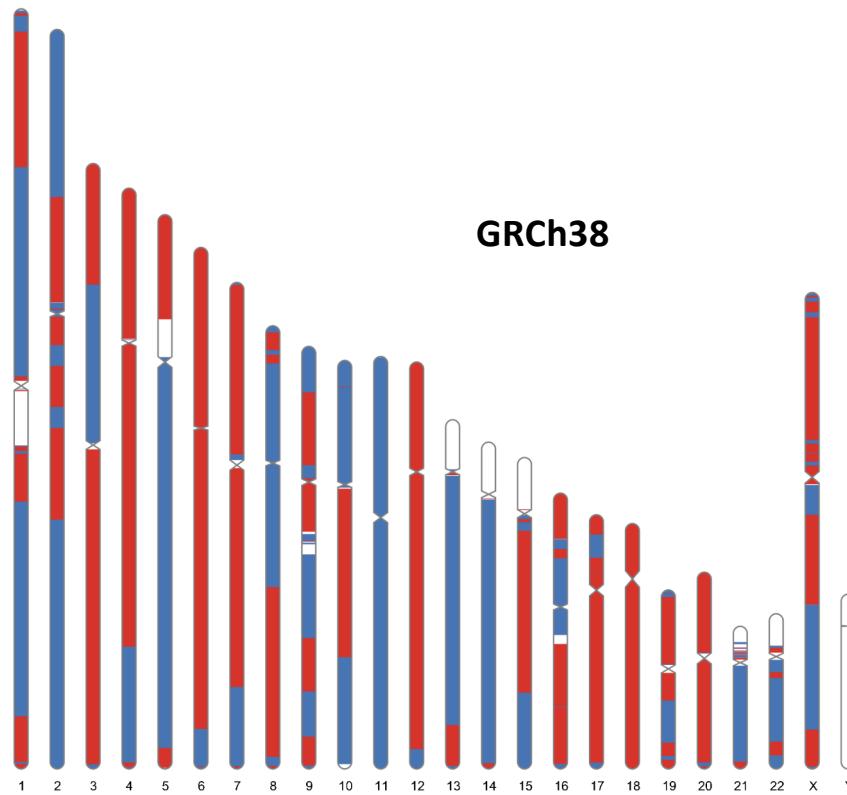
...but GRCh38 is missing ~200Mbp of genetic information...



Red segments resolved by T2T Consortium
DOI: [10.1126/science.abp8653](https://doi.org/10.1126/science.abp8653)

Ignas Bunikis

De novo assembly mapped to T2T



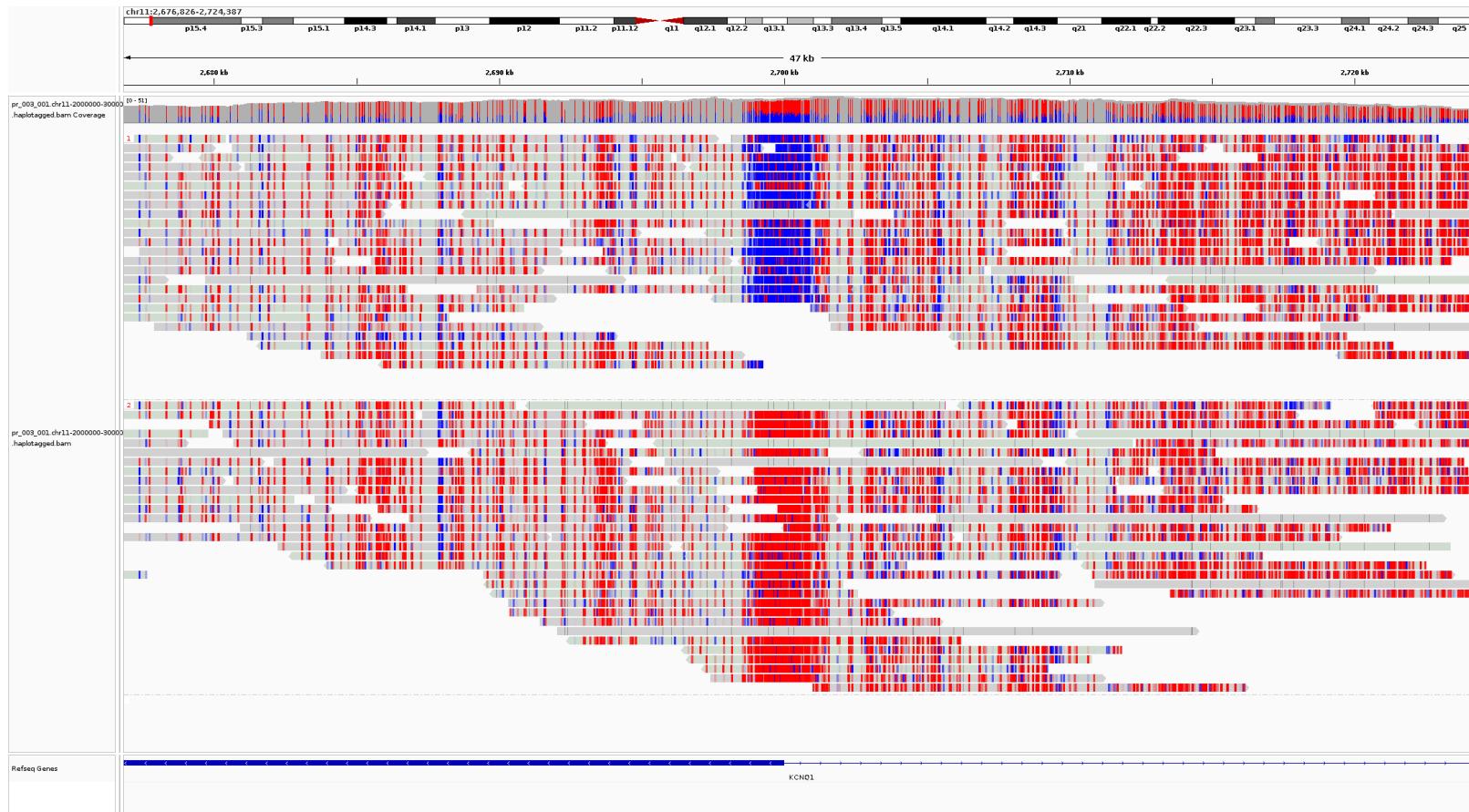
Colour change represents adjacent contigs

Ignas Bunikis

Example 3: Investigate methylation



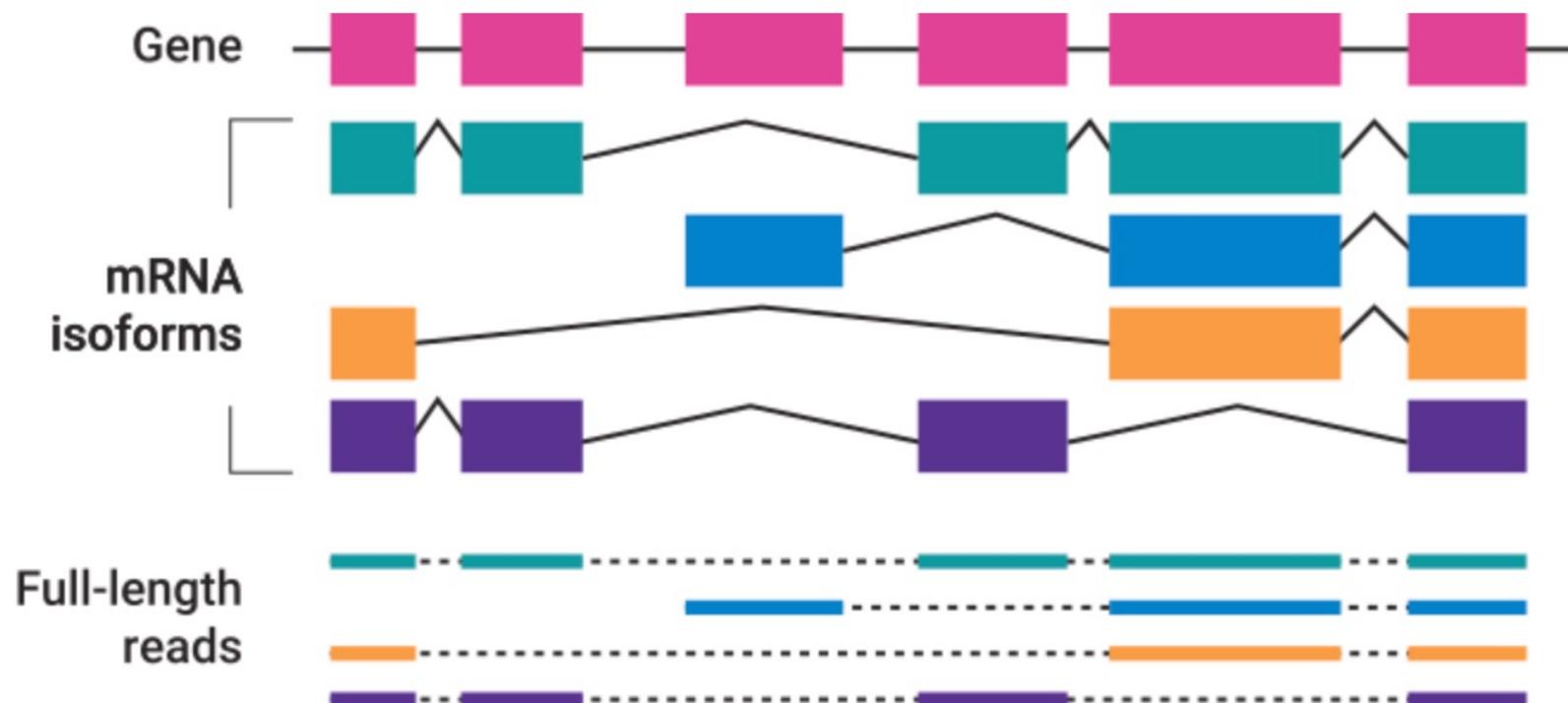
Obtain methylation patterns, phased with haplotypes (example for imprinted region)



Example 4: Full-length RNA sequencing



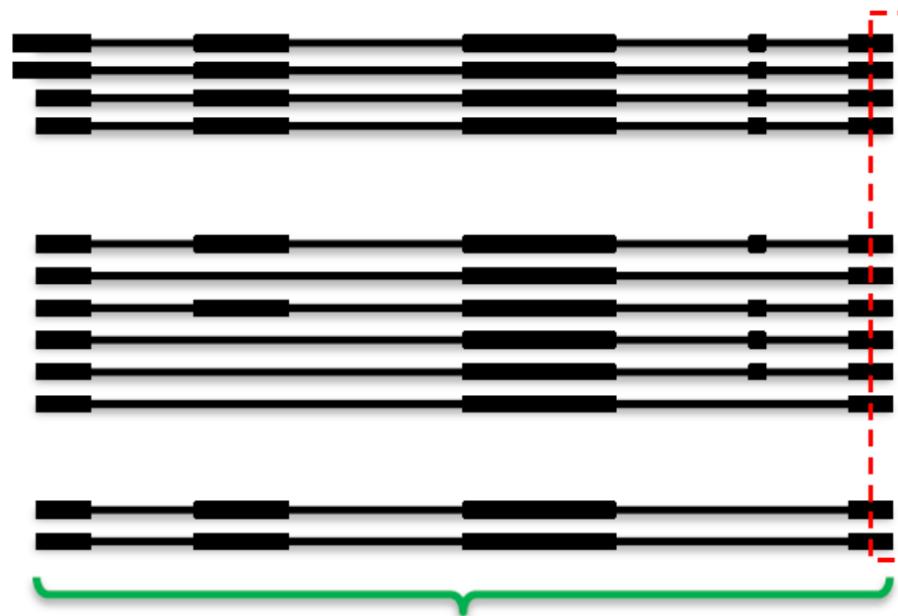
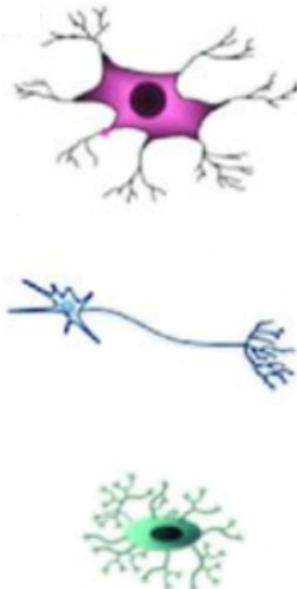
- Get complete information about RNA molecules!



Example 5: Single-cell long-read RNA



- It is possible to study RNA isoforms even in single cells!

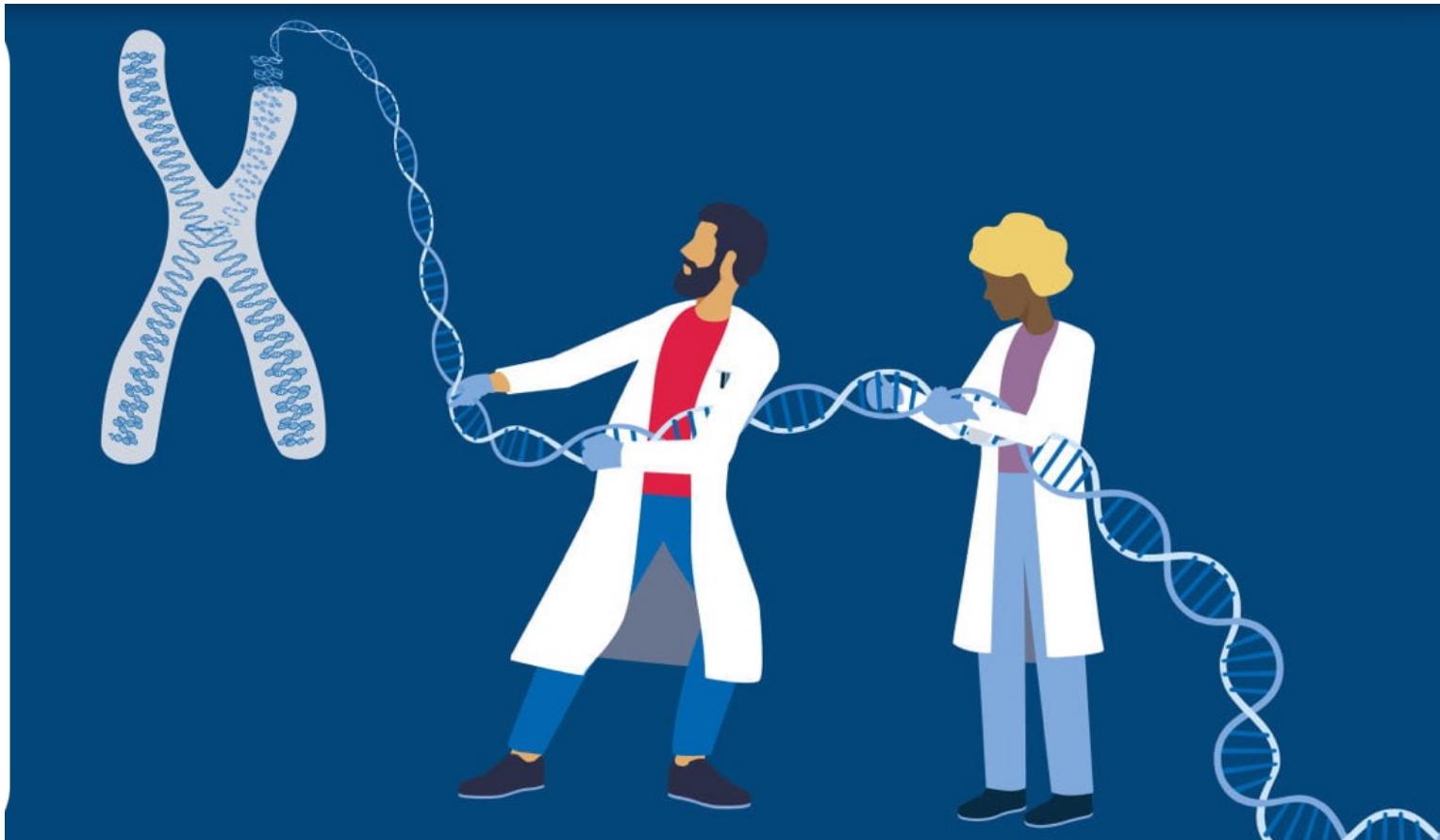


Cell type specific
mRNA splicing

Not captured with 3'-end
short-read scRNA seq

Resolved with single-cell full-length RNA seq

Challenge: good sample quality required!



<https://www.qiagen.com/ja-us/applications/molecular-biology-research/hmw-dna>

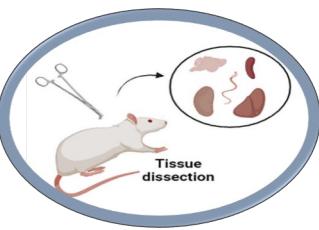
HMW-DNA Extraction – Options at NGI



Cells/Blood
 $1 \times 10^6 - 5 \times 10^6$



Tissue
25-100 mg



Insects/Mollusc/Crustaceans
25-200 mg



Plants
1-3 g



Fungi
100-600 mg



Commercial Kits

MONARCH

High input quality required
Few special protocols

Top choice for high quality samples with low amount of contaminants

NANOBIND

Lower input quality tolerated
Many special protocols
Supplemental buffers for insects

Top choice for most non-standard samples except for low input and high polysaccharide samples

Phenol/Chloroform

SDS Lysis

High polyphenol
High recovery for low input

Top choice for samples high in polyphenols without polysaccharides

CTAB Lysis

High polysaccharide
Also handles polyphenols

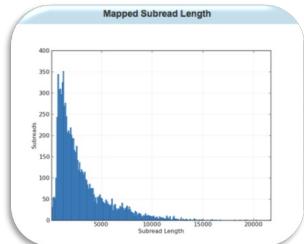
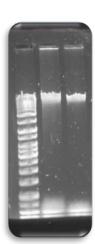
Top choice for plants, fungi, and other samples high in polysaccharides

HMW-DNA Extraction – Contaminants



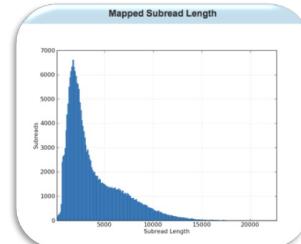
Importance of purity – even for model organisms

Impurities can originate from both host tissue and extraction chemicals.



Polished Contigs	223	Max Contig Length	36,298
N50 Contig Length	2,932	Sum of Contig Lengths	480,087

Same yeast -
different
extractions!



Polished Contigs	9	Max Contig Length	1,508,929
N50 Contig Length	1,353,702	Sum of Contig Lengths	7,813,244

We extract what we get!



Sequencing of the last supper?



VS



VS

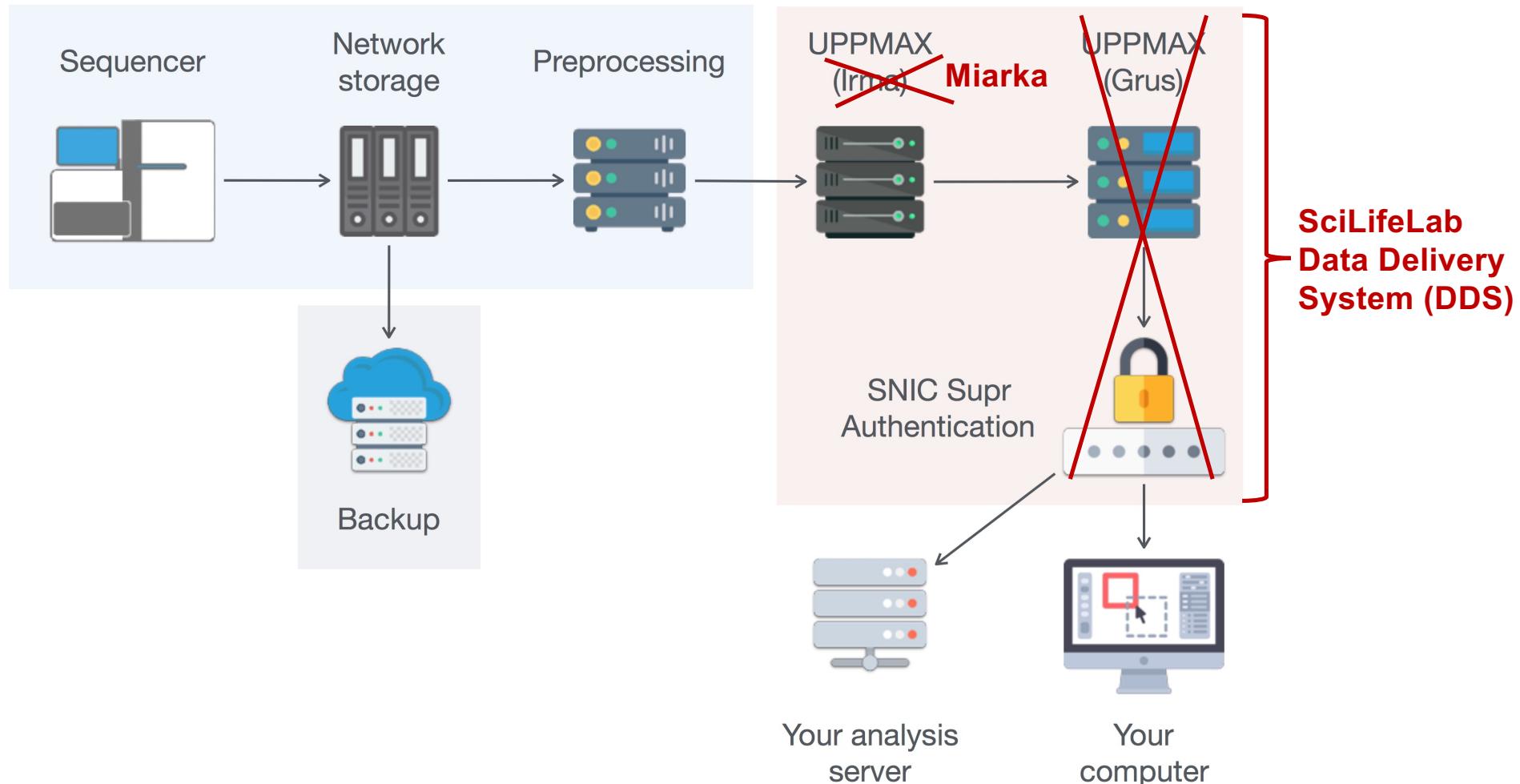


Which would you expect to have less contaminants?

NGI Data Handling and Analysis Pipelines



NGI Data Handling





Data delivery via DDS

- DDS is a system for delivery of data from SciLifeLab platforms
 - Cloud-based solution
 - Command line and web interface
 - Can handle also sensitive data
- Instruction video available on Youtube!





Quality control

- Every project has some level of quality control checks
 - Technical run performance
 - Read length distribution
 - Sequencing quality
- Analysis pipelines give application-specific QC
- Reporting done using MultiQC (Illumina projects)





Multi QC example

MultiQC v1.0

P1234: Test_NGI_Project

- General Stats
- NGI-RNaseq
- Sample Similarity
- MDS Plot
- STAR**
- Cutadapt
- FastQC
- Sequence Quality Histograms
- Per Sequence Quality Scores
- Per Base Sequence Content
- Per Sequence GC Content
- Per Base N Content
- Sequence Length Distribution
- Sequence Duplication Levels
- Overrepresented sequences
- Adapter Content

MultiQC

P1234: Test_NGI_Project

This is an example project. All identifying data has been removed.

Contact E-mail: phil.ewels@scilifelab.se
Application Type: RNA-seq
Sequencing Platform: HiSeq 2500 High Output V4
Sequencing Setup: 2x125
Reference Genome: hg19

Report generated on 2017-06-17, 18:43 based on data in:
/Users/philewels/GitHub/MultiQC_website/public_html/examples/ngi-rna/data

NGI names User supplied names

General Statistics

Copy table Configure Columns Plot Showing 22 rows and 6 columns.

Sample Name	% Aligned	M Aligned	% Trimmed	% Dups	% GC	M Seqs
P1234_1001	68.2%	22.8	10.3%	71.3%	49%	33.7
P1234_1002	67.9%	20.9	10.7%	70.1%	50%	31.1
P1234_1003	64.7%	21.7	11.0%	72.3%	50%	33.7
P1234_1004	55.2%	17.0	13.2%	73.4%	51%	31.2
P1234_1005	53.0%	17.7	15.9%	75.8%	52%	33.8
P1234_1006	52.7%	16.1	14.1%	73.8%	52%	30.8
P1234_1007	33.0%	7.0	32.0%	60.5%	52%	21.8
P1234_1008	27.5%	4.3	44.2%	79.1%	50%	16.7
P1234_1009	52.3%	10.5	20.9%	64.2%	48%	20.5



Analysis pipelines

- NGI provides data analysis for most applications
- Analysis requirements: Automated, reliable, easy to run, reproducible

The screenshot shows the homepage of the SciLifeLab National Genomics Infrastructure. At the top, there is a navigation bar with links for HOME, APPLICATIONS, TECHNOLOGIES, BIOINFORMATICS, RESOURCES, NEWS, ABOUT US, CONTACT, and NEW ORDER. A search bar is also present. The main content area features a large blue background image of laboratory pipettes. Overlaid on this image is a white callout box containing the text: "SciLifeLab National Genomics Infrastructure" and "NGI is one of the largest technical platforms at SciLifeLab. We provide access to technology for sequencing, genotyping and associated bioinformatics support to researchers based in Sweden." Below this box is a blue button labeled "Getting started at NGI". At the bottom of the page is a large blue button labeled "Get started".

nf-core: a popular pipeline system



- A community effort to collect a curated set of Nextflow analysis pipelines
- GitHub organisation to collect pipelines in one place
- No institute-specific branding
- Strict set of guideline requirements

nature biotechnology

Correspondence | Published: 13 February 2020

The nf-core framework for community-curated bioinformatics pipelines

Philip A. Ewels, Alexander Peltzer, Sven Fillinger, Harshil Patel, Johannes Alneberg, Andreas Wilm,
Maxime Ulysse Garcia, Paolo Di Tommaso & Sven Nahnsen

nf-core <https://nf-co.re>



Available pipelines at NGI

- All information available on our website: <https://ngisweden.scilifelab.se>

Amplicon-seq analysis



ATAC-seq analysis



Methylation-seq analysis



ChIP-seq analysis



Genome assemblies with HiFi data



Ion Torrent secondary analysis

Nanopore analysis



PacBio Iso-Seq Analysis

PromethION secondary analysis



Illumina QC analysis

RAD-seq analysis



RNA-fusion analysis

RNA-seq analysis



Small-RNA analysis

Spatial Transcriptomics analysis



WGS and WES germline / somatic analysis





WES and WGS analysis

WGS and WES germline / somatic analysis

Runs with illumina DNA-sequencing data, WGS or targeted sequencing e.g. WES. Aligns to the reference genome, gives QC metrics, does variant-calling and finishes with annotation.

[nf-core/sarek \(paper\)](#) is an analysis pipeline for WGS and targeted sequencing data e.g WES. Previously known as the Cancer Analysis Workflow (CAW), Sarek can handle regular samples or tumour/normal pairs, including relapse samples if required. Sarek was co-developed by NGI.

Sarek analysis can be divided into two different use cases: germline analysis and somatic analysis. These two use cases share the same main steps: mapping, variant calling and annotation.

The screenshot shows the GitHub repository page for nf-core/sarek. It features a circular icon with a green and yellow abstract shape. The repository name "nf-core/sarek" is at the top, followed by the URL "https://github.com/nf-core/sarek". A brief description states: "Analysis pipeline to detect germline or somatic variants (pre-processing, variant calling and annotation) from WGS / targeted sequencing - <https://nf-co.re/sarek>". Below the description are three statistics: 324 forks, 324 stars, and 354 issues.

When we run analysis

We routinely run Sarek germline analysis upon request for human WGS and WES projects. For the Sarek somatic analysis, the decision to run the analysis is made on a case by case basis. If you're interested, please get in touch with us and mention that you would like us to run this analysis.

The analysis currently works with the human reference genomes available in [AWS-iGenomes \(GRCh37/GRCh38\)](#). If in doubt, please ask whether we can run the pipeline for you.

Input data

Sarek can start from the unprocessed demultiplexed FastQ files from the sequencer together with a small bit of contextual data in the form of a TSV-file. For each sample, the TSV-file should denote the sex of the subject and whether the sample is tumour or normal. In most cases, this information needs to be submitted to NGI by the user.

Results

The pipeline generates BAM alignment files and variant-calling VCF files, along with numerous quality control metrics. For more information, please see the [official documentation](#).



Available pipelines at NGI

Amplicon-seq analysis



ATAC-seq analysis

Methylation-seq analysis



ChIP-seq analysis

Genome assemblies with HiFi data



Ion Torrent secondary analysis

Nanopore analysis



PacBio Iso-Seq Analysis

PromethION secondary analysis



Illumina QC analysis

RAD-seq analysis



RNA-fusion analysis

RNA-seq analysis



Small-RNA analysis

Spatial Transcriptomics analysis



WGS and WES germline / somatic analysis

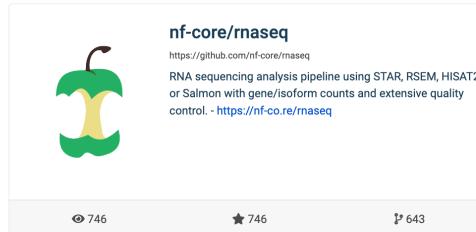


Example: RNA-seq analysis

RNA-seq analysis

Runs with illumina total RNA-sequencing data. Aligns to the reference genome, gives QC metrics and finishes with gene count matrices.

RNA-Seq is a bioinformatics analysis pipeline used for RNA sequencing data. The pipeline is built using [Nextflow](#), a workflow tool to run tasks across multiple compute infrastructures in a very portable manner. It processes raw data from FastQ inputs, aligns the reads, generates counts relative to genes or transcripts and performs extensive quality control on the results.



When we run analysis

We run this analysis routinely for all RNA-seq projects where we have prepared the sequencing library in-house. If you have prepared a library yourself and we are just sequencing, please get in touch and mention that you would like us to run this analysis.

The analysis works with any of the species that have a reference genome available in [AWS-iGenomes](#). If in doubt, please ask whether we can run the pipeline for you.

Input data

bcl2fastq demultiplexed FastQ files and a genome reference.

Results

The pipeline generates aligned BAM-files, gene count matrices and FPKM metrics for genes and transcripts, along with numerous quality control metrics. For more information, please see [https://nf-co.re/rnaseq/\[release\]/docs/output](https://nf-co.re/rnaseq/[release]/docs/output)

Last Updated: 18th October 2023



Available pipelines at NGI

Amplicon-seq analysis		ATAC-seq analysis	
Methylation-seq analysis		ChIP-seq analysis	
Genome assemblies with HiFi data		Ion Torrent secondary analysis	
Nanopore analysis		PacBio Iso-Seq Analysis	
PromethION secondary analysis		Illumina QC analysis	
RAD-seq analysis		RNA-fusion analysis	
RNA-seq analysis		Small-RNA analysis	
Spatial Transcriptomics analysis		WGS and WES germline / somatic analysis	

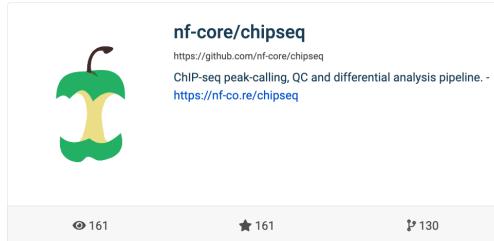


ChIP-seq analysis

ChIP-seq analysis

Runs with ChIP sequencing data. Pre-processes raw data from FastQ inputs, aligns the reads and performs peak calling and extensive quality-control on the results.

ChIP-Seq is a bioinformatics best-practice analysis pipeline used for chromatin immunoprecipitation (ChIP-seq) data analysis. The pipeline uses [Nextflow](#), a bioinformatics workflow tool. It pre-processes raw data from FastQ inputs, aligns the reads and performs peak calling and extensive quality-control on the results.



When we run analysis

We run this analysis routinely for all ChIP-seq projects where we have prepared the sequencing library in-house. If you have prepared a library yourself and we are just sequencing, please get in touch and mention that you would like us to run this analysis.

The analysis works with any of the species that have a reference genome available in [AWS-iGenomes](#). If in doubt, please ask whether we can run the pipeline for you.

Input data

bcl2fastq demultiplexed FastQ files and a genome reference.

Results

The pipeline generates aligned BAM-files, files with information about called peaks, along with numerous quality control metrics. For more information, please see <https://nf-co.re/chipseq/docs/output>.

Last Updated: 14th July 2023



Available pipelines at NGI

Amplicon-seq analysis



ATAC-seq analysis

Methylation-seq analysis



ChIP-seq analysis

Genome assemblies with HiFi data



Ion Torrent secondary analysis

Nanopore analysis



PacBio Iso-Seq Analysis

PromethION secondary analysis



Illumina QC analysis

RAD-seq analysis



RNA-fusion analysis

RNA-seq analysis



Small-RNA analysis

Spatial Transcriptomics analysis



WGS and WES germline / somatic analysis

Genome assembly with HiFi data



Genome assemblies with HiFi data

NGI can generate high quality assemblies using IPA and hifiasm assemblers

hifiasm & 
IMPROVED
PHASED
ASSEMBLY

Improved Phased Assembler (IPA) is the official PacBio software for **HiFi** genome assembly. IPA was designed to utilize the accuracy of PacBio HiFi reads to produce high-quality phased genome assemblies.

Hifiasm is a fast haplotype-resolved *de novo* assembler for PacBio HiFi reads. It emits partially phased assemblies of quality competitive with the best assemblers. Given parental short reads or Hi-C data, it produces arguably the best haplotype-resolved assemblies so far.

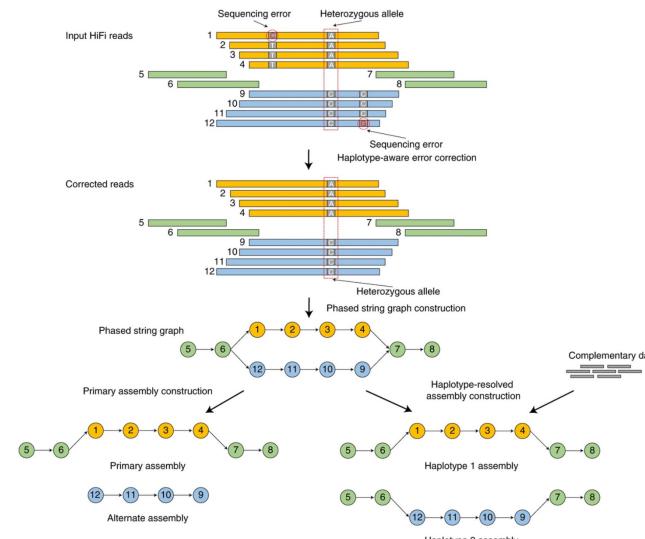


Image: Nat Methods 18, 170–175 (2021). <https://doi.org/10.1038/s41592-020-01056-5>

**Not yet implemented as a
nf-core pipeline!**



Trend: On-instrument analysis

More and more analyses being done on instrument GPUs

Illumina NovaSeqX

Mapping and variant calling (Dragen)



PacBio Revio

Onboard generation of HiFi reads



➔ Can speed up and streamline the analysis process...



You can also get help from NBIS!

[About us](#)[Services](#)[Training](#)[Contact](#)

A distributed national bioinformatics infrastructure supporting life sciences in Sweden

[Get support](#)

- All solutions are not available from NGI, but NBIS has lots of experts!

Some tips for data analysis...



Think about analysis early on – already when planning the project!

- Which tools should be used?
- Can I run the analysis myself, or do I need assistance?
- Where should the analysis be run?
- Do I have enough storage space?
- Where should the data eventually be archived?

NGI strategic projects and collaborations



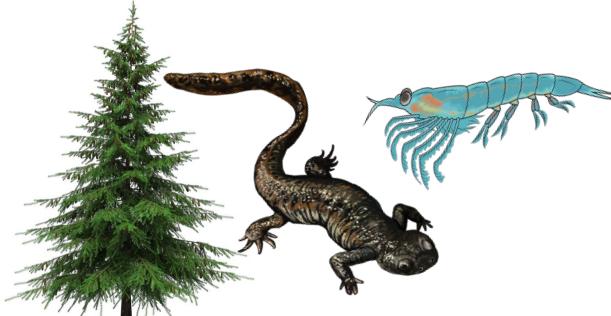
We are involved in some larger national and international projects...



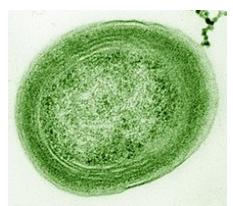


Biodiversity genomics

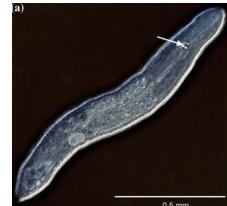
Reference genomes of **any** organism - a very challenging endeavour



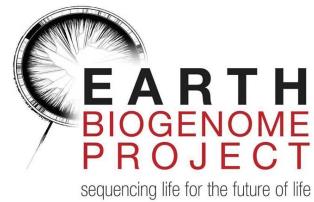
Large genomes (18-22Gb)



Tiny organisms



Tiny organisms
with large genomes



MAX-PLANCK-GESELLSCHAFT



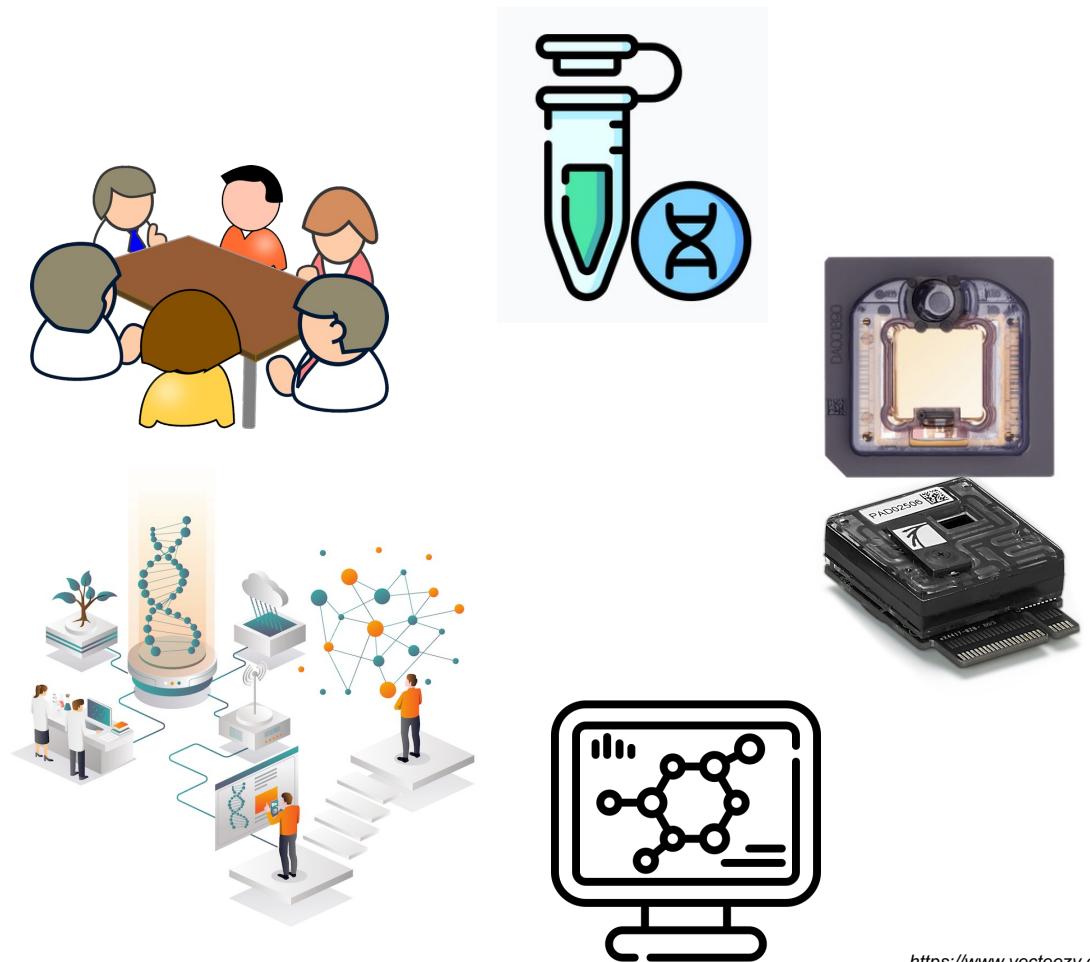
And many, many, ..., many other uncooperative organisms



Reference genome sequencing

NGI & NBIS can help out with:

- DNA/RNA extractions
- Long-read sequencing
- Hi-C Illumina sequencing
- RNA sequencing
- De novo assembly
- Genome annotation





Human genome analysis



Photo: SVT

Swedish WGS reference dataset

2017: 1,000 genomes sequenced on Illumina



2024: Time to do it again, with long reads!



Ameur et al, Eur. J. Hum. Genet. 2017

How to build a long-read reference dataset?

- Planning started 2023, with a wishlist for a new Swedish population cohort

	Description	Priority
Consent for data sharing	It must be possible to share individual-level variant information (VCF files) on national level and ideally also internationally	Crucial
Amount and quality of DNA	At least 5ug of high-quality DNA per individual, ideally from fresh samples extracted for long-read sequencing	Crucial
Phenotype information	Detailed phenotype information available, that can be used for specific research projects (after approval)	Important
A cross-section of Sweden	The individuals should not be enriched for a specific disease or phenotype, and reflect the genetic variation in Sweden (ideally including ethnic minorities)	Important
Additional OMICS data	Possibility to perform other OMICs studies (RNA, protein, etc) on samples from the same individuals	Important
Available SNP array data	Data from SNP arrays, that can be used to infer the genetic background and select representative individuals for sequencing	Beneficial
Funding and resources	Possibility to get additional local funding and resources (for re-consent, sample collection, DNA extraction, etc.)	Beneficial

How to get funding for sequencing?

“Genome of Europe” - A 40M Euro project within the 1+Million Genomes Initiative!



The Genome of Europe

Aim: Construct a european reference WGS dataset

Sweden's representatives - Adam Ameur, Anna Lindstrand, Bengt Persson, Anna Hagwall

- 100,000 individuals, from 27 countries
 - Representative of Europe's population
 - International data sharing possible
 - Swedens contribution: 2,600 WGS

What is the best sample collection for GoE?



- SCAPIS: A prospective population study for heart- and lung disease
- Over 30,000 participants, collected at 6 sites (from Lund to Umeå)
- We are planning to analyze at least 1000 individuals

Collaborations on Rare Disease



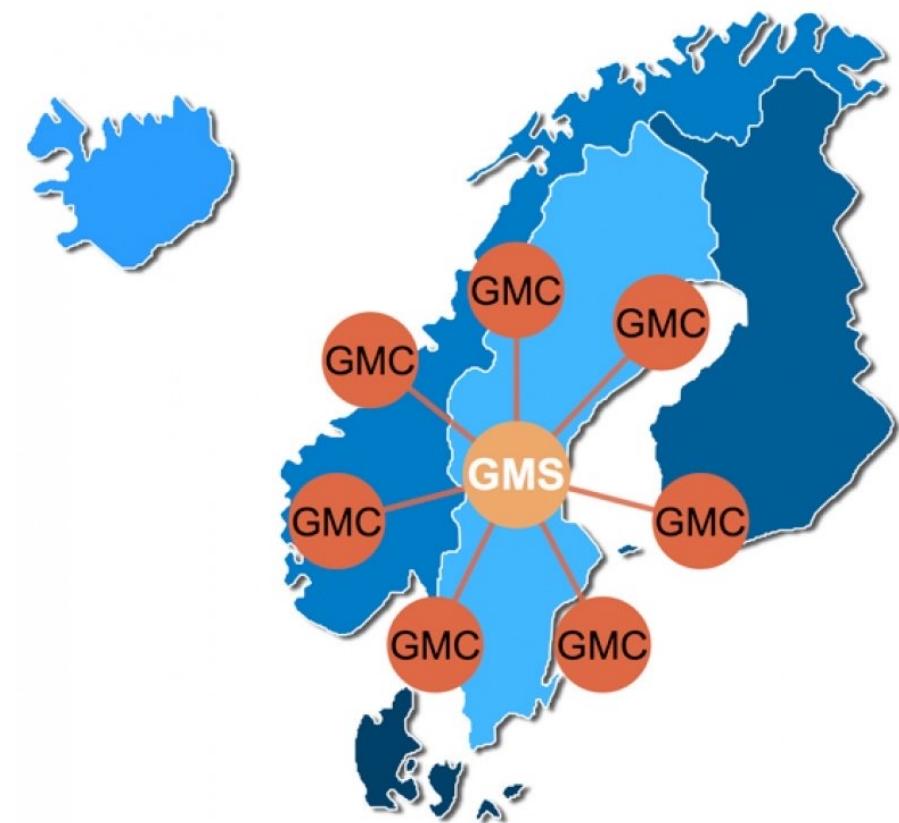
We are collaborating with Genomic Medicine Sweden - Rare Disease Group

Long-Read Whole Genome Sequencing

- Improve diagnostics of rare disease patients
- Resolve complex SVs and other variants
- PacBio Revio and ONT PromethION

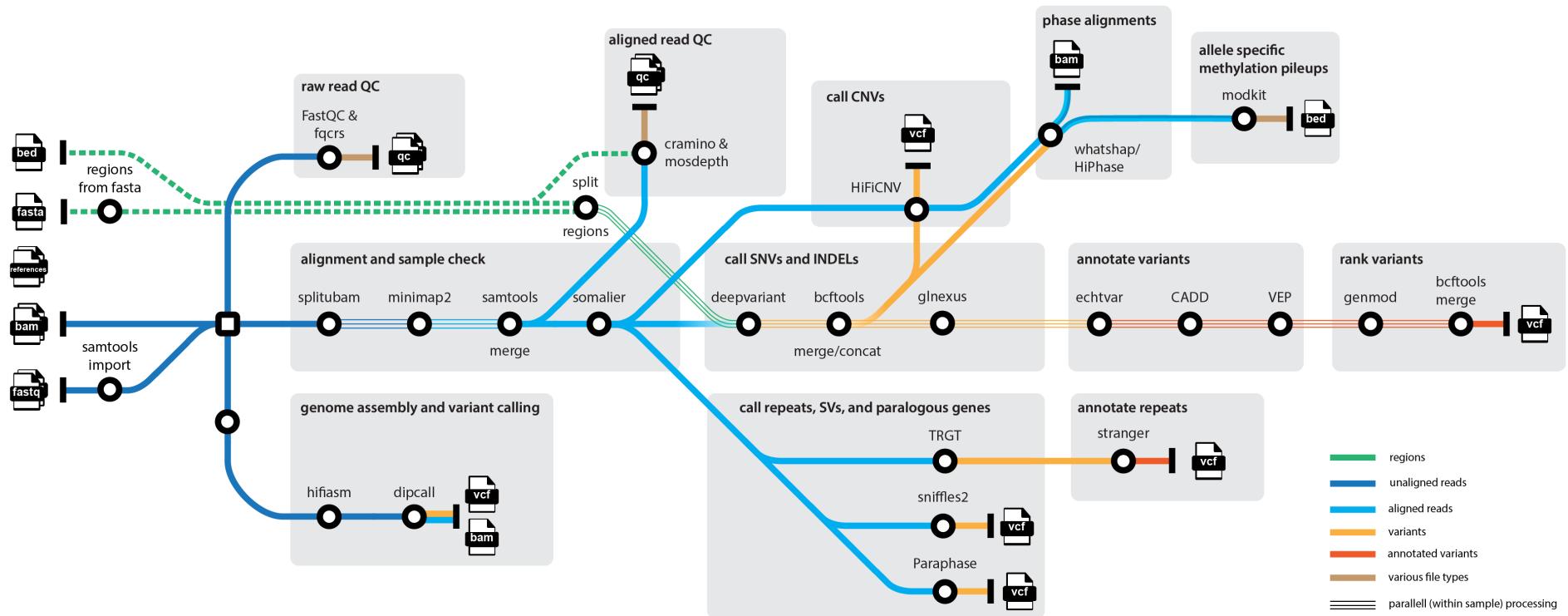
Long-Read Targeted Sequencing

- Develop clinical assays for repeat expansions
- Cas9-based capture or adaptive sampling
- Aim: implementation at different hospital nodes



How to analyze human long-read data?

Nallo: a Nextflow analysis pipeline for patients and controls



Felix Lenner, et al

Thanks for your attention!



Diabetes

Alzheimer's disease

Whole-genome sequencing

Gene therapy

Infection screen

Whole-transcriptome sequencing

Target sequencing

Cancer prognosis

Gene regulation

Crohn's disease

Genomics of ageing

Exome sequencing

Schizophrenia

Cancer diagnostics

Organ donor matching

Gut microflora

Gene fusions

RNA editing

HIV

HPV

HCV

Scoliosis

Immune response

Monogenic disorders

Sudden infant death

Cervical cancer

Lynch syndrome

Leukemia

Scoliosis

HLA typing

Dyslexia

MRSA / BRSA screen

Sudden cardiac arrest

Transcriptional regulation

Prenatal diagnostics

Muscle dystrophy

Individualised cancer therapy

and much more...