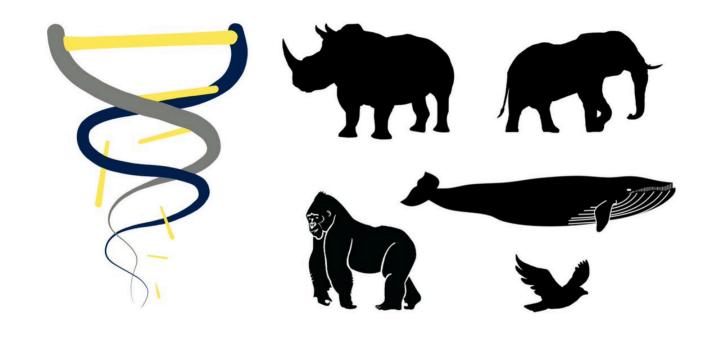
An Example Snakemake Workflow





https://github.com/NBISweden/GenErode

Kutschera et al. 2022 (BMC Bioinformatics)



 Developed in a NBIS project with Love Dalén's lab (Centre for Palaeogenetics, SU & NRM)



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- Compares population genomics statistics from historical and modern samples of endangered populations



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 Data processing from fastq files to BAM & VCF files plus downstream population genomics analyses



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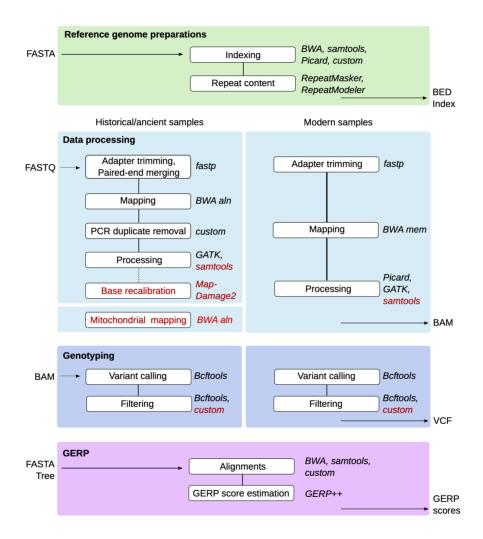
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- Historical and modern samples are processed in parallel
- Whole-genome resequencing data from historical/ancient samples needs special processing as DNA degrades over time
- Some analyses or filtering steps are run separately for modern and historical samples, or only for historical samples

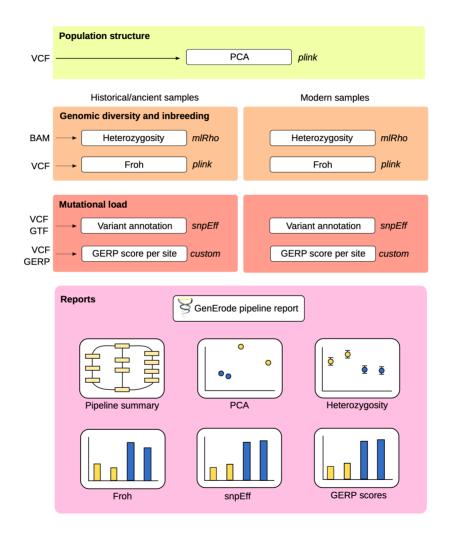


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- Snakefile
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 - all rule collecting output files produced by the different rule Snakefiles
 - Python and bash code to generate and edit the pipeline report with snakemake --report
- Cluster execution (e.g. UPPMAX) with slurm:
 - o config/cluster.yaml file to set up slurm profile



- Metadata files for historical and modern samples (separately)
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- Example historical_samples.txt file (whitespace-separated)

samplename_index_lane	readgroup_id	readgroup_platform	path_to_R1_fastq	path_to_R2_fastq
VK01_01_L2	BHYOX3ALTH.L2.01	illumina	data/S1/P01_2.R1.fq.gz	data/S1/P01_2.R2.fq.gz
VK01_02_L2	BHYOX3ALTH.L2.02	illumina	data/S1/P02_2.R1.fq.gz	data/S1/P02_2.R2.fq.gz

• **See** config/historical_samples_paths.txt **and** config/modern_samples_paths.txt



- Config file config.yaml (to be edited by users, placed in config/)
 - Paths to input data and metadata tables
 - Selection of analysis steps to be run
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Step 1: Use booleans in the config file (config/config.yaml) as on/off switches



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```
#####
# FastQC on raw reads, adapter and quality trimming (incl. read merging
# for historical samples) using fastp, FastQC on trimmed reads.
# Adapter sequences are automatically detected.
# Automatic detection of NovaSeq or NextSeq samples and activation of
# poly-G tail trimming.
fastq_processing: True

[...]
#####
# Map historical and modern reads to reference genome assembly (specified above).
mapping: False
#####
```



For many analysis steps, parameters can be specified in the config file

These parameters can be used in the workflow with the syntax

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• the <code>include</code> variable to attach the rule Snakefiles corresponding to the analysis steps that were set to <code>True</code> in the config file,

```
if config["fastq_processing"]:
    include: "workflow/rules/1.1_fastq_processing.smk"
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if config["fastq_processing"]:
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• and the rule all that takes the output files from the list all_outputs as input

```
rule all:
   input: all_outputs,
```



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• The rule Snakefile (workflow/rules/1.1_fastq_processing.smk) contains some Python code to add its final output files to the list all_outputs in the main Snakefile



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```
import os
if os.path.exists(config["historical_samples"]):
    all_outputs.append("data/raw_reads_symlinks/historical/stats/multiqc/multiqc_report.html")
    all_outputs.append("results/historical/trimming/stats/multiqc/multiqc_report.html")

if os.path.exists(config["modern_samples"]):
    all_outputs.append("data/raw_reads_symlinks/modern/stats/multiqc/multiqc_report.html")
    all_outputs.append("results/modern/trimming/stats/multiqc/multiqc_report.html")
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• config["historical_samples"] and config["modern_samples"] point to the config file where the paths to metadata files are specified:



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```

 By using the if statement to check for the presence of historical or modern metadata files, the workflow can also be run only for historical or only for modern samples



Questions?



