An Example Snakemake Workflow





The GenErode pipeline

- Developed in a NBIS project with Love Dalén's lab (Centre for Palaeogenetics, SU & NRM)
- Compares population genomics statistics from historical and modern samples of endangered populations



Sumatran rhinoceros (Dicerorhinus sumatrensis), critically endangered

 Data processing from fastq files to BAM & VCF files plus downstream population genomics analyses





The GenErode pipeline

- Started at Snakemake version 3.10 (!), current pipeline runs with Snakemake version 5.22
- Historical and modern samples are processed in parallel
- Whole-genome resequencing data from historical/ancient samples needs special processing as DNA degrades over time
- Some analyses or filtering steps are run separately for modern and historical samples, or only for historical samples





Analysis Tracks of the Workflow

- Data processing track
 - Repeat element identification
 - Fastq file processing
 - Optional: mapping to mitochondrial genomes
 - Mapping to reference genome
 - BAM file processing
 - Optional: base quality rescaling for historical samples
 - Optional: subsampling to target depth
 - Genotyping
 - Optional: CpG site identification & removal
 - VCF file processing & merging per dataset





Analysis Tracks of the Workflow

- BAM file track
 - Optional: mlRho
- VCF file track
 - Optional: plink (PCA)
 - Optional: plink (runs of homozygosity)
 - Optional: snpEff
 - o Optional: GERP++ & relative mutational load calculation





The Workflow Structure

- Rules with the actual analyses in separate Snakefiles (in workflow/rules/)
- Snakefile
 - Python code to create sample and readgroup ID dictionaries & lists from metadata tables and a config file
 - include of rule Snakefiles
 - o all rule collecting output files produced by the different rule Snakefiles
- UPPMAX / slurm system:
 - o cluster.yaml file to set up slurm profile





The Workflow Structure

- Metadata files for historical and modern samples (separately)
 - o Sample IDs, readgroup IDs, sequencing technology, paths to fastq files
- Example historical_samples.txt file (whitespace-separated)

samplename_index_lane	readgroup_id	readgroup_platform	path_to_R1_fastq	path_to_R2_fastq
VK01_01_L2	BHYOX3ALTH.L2.01	illumina	data/S1/P01_2.R1.fq.gz	data/S1/P01_2.R2.fq.gz
VK01_02_L2	BHYOX3ALTH.L2.02	illumina	data/S1/P02_2.R1.fq.gz	data/S1/P02_2.R2.fq.gz





The Workflow Structure

- Config file config.yaml (to be edited by users, placed in config/)
 - Paths to input data and metadata tables
 - Selection of analysis steps to be run
 - Parameters for different rules
 - Lists with samples for optional analyses





Step 1: Use booleans in the config file (config/config.yaml) as on/off switches

#####
FastQC on raw reads, adapter trimming, read merging (historical samples), FastQC on trimmed reads.

[...]
#####
Map historical and modern reads to reference genome assembly (specified above).
mapping: False
#####





Step 1: Use booleans in the config file (config/config.yaml) as on/off switches

• For many analysis steps, parameters can be specified in the config file:

```
#####
# FastQC on raw reads, adapter trimming, read merging (historical samples), FastQC on trimmed reads.
fastq_processing: True

# Adapter sequences for trimming of historical samples using SeqPrep v1.1 (modified source code)
# Forward read primer/adapter sequence to trim historical samples in SeqPrep v1.1 (parameter "-A")
hist_F_adapter: "AGATCGGAAGAGCACACGTCTGAACTCCAGTCACNNNNNNNNATCTCGTATGCCGTCTTCTGCTTG" # example from Meyer & Kircher 2010
# Reverse read primer/adapter sequence to trim historical samples in SeqPrep v1.1 (parameter "-B")
# When using double indices, include full adapter length (replacing the index by "N")
hist_R_adapter: "AGATCGGAAGAGCGTCGTGTAGGGAAAGAGTGTAGATCTCGGTGGTCGCCGTATCATTT" # example from Meyer & Kircher 2010

[...]
# Minimum read length allowed after trimming.
# Historical samples (SeqPrep v1.1 with modified source code)
hist_readlength: "30" # default setting: 30 bp

# Modern samples (trim-galore)
mod_readlength: "30"
######
```

These parameters can be used in the workflow with the syntax config["parameter_name"]





Step 2: Use some Python code, include and the rule all to figure out what the workflow will do

 The main Snakefile contains an empty Python list all_outputs to collect output files from the included rule Snakefiles,

```
all_outputs = []
```

• the include variable to attach the rule Snakefiles corresponding to the analysis steps that were set to True in the config file,

```
if config["fastq_processing"]:
  include: "workflow/rules/1.1_fastq_processing.smk"
```

• and the rule all that takes the output files from the list all_outputs as input

```
rule all:
input: all_outputs
```





Step 2: Use some Python code, include and the rule all to figure out what the workflow will do

• The rule Snakefile (workflow/rules/1.1_fastq_processing.smk) contains some Python code to add its final output files to the list all_outputs in the main Snakefile

```
import os
if os.path.exists(config["historical_samples"]):
    all_outputs.append("data/raw_reads_symlinks/historical/stats/multiqc/multiqc_report.html")
    all_outputs.append("results/historical/trimmed_merged_reads/stats/multiqc/multiqc_report.html")

if os.path.exists(config["modern_samples"]):
    all_outputs.append("data/raw_reads_symlinks/modern/stats/multiqc/multiqc_report.html")
    all_outputs.append("results/modern/trimmed_reads/stats/multiqc/multiqc_report.html")
```

 By using the if statement to check for the presence of historical or modern metadata files, the workflow can also be run only for historical or only for modern samples





Questions?







