

Dear Author,

Here are the proofs of your article.

- You can submit your corrections **online**, via **e-mail** or by **fax**.
- For **online** submission please insert your corrections in the online correction form. Always indicate the line number to which the correction refers.
- You can also insert your corrections in the proof PDF and **email** the annotated PDF.
- For fax submission, please ensure that your corrections are clearly legible. Use a fine black pen and write the correction in the margin, not too close to the edge of the page.
- Remember to note the **journal title**, **article number**, and **your name** when sending your response via e-mail or fax.
- **Check** the metadata sheet to make sure that the header information, especially author names and the corresponding affiliations are correctly shown.
- **Check** the questions that may have arisen during copy editing and insert your answers/corrections.
- **Check** that the text is complete and that all figures, tables and their legends are included. Also check the accuracy of special characters, equations, and electronic supplementary material if applicable. If necessary refer to the *Edited manuscript*.
- The publication of inaccurate data such as dosages and units can have serious consequences. Please take particular care that all such details are correct.
- Please **do not** make changes that involve only matters of style. We have generally introduced forms that follow the journal's style. Substantial changes in content, e.g., new results, corrected values, title and authorship are not allowed without the approval of the responsible editor. In such a case, please contact the Editorial Office and return his/her consent together with the proof.
- If we do not receive your corrections **within 48 hours**, we will send you a reminder.
- Your article will be published **Online First** approximately one week after receipt of your corrected proofs. This is the **official first publication** citable with the DOI. **Further changes are, therefore, not possible.**
- The **printed version** will follow in a forthcoming issue.

#### Please note

After online publication, subscribers (personal/institutional) to this journal will have access to the complete article via the DOI using the URL: [http://dx.doi.org/\[DOI\]](http://dx.doi.org/[DOI]).

If you would like to know when your article has been published online, take advantage of our free alert service. For registration and further information go to: <http://www.link.springer.com>.

Due to the electronic nature of the procedure, the manuscript and the original figures will only be returned to you on special request. When you return your corrections, please inform us if you would like to have these documents returned.

# Metadata of the article that will be visualized in OnlineFirst

**Please note: Images will appear in color online but will be printed in black and white.**

ArticleTitle	A newt does not change its spots: using pattern mapping for the identification of individuals in large populations of newt species	
Article Sub-Title		
Article CopyRight	The Ecological Society of Japan (This will be the copyright line in the final PDF)	
Journal Name	Ecological Research	
Corresponding Author	Family Name	<b>Mettouris</b>
	Particle	
	Given Name	<b>Onoufrios</b>
	Suffix	
	Division	Department of Biology
	Organization	University of Patras
	Address	26500, Patras, Greece
	Email	omettouris@gmail.com
Author	Family Name	<b>Megremis</b>
	Particle	
	Given Name	<b>George</b>
	Suffix	
	Division	Department of Biology
	Organization	University of Patras
	Address	26500, Patras, Greece
	Email	
Author	Family Name	<b>Giokas</b>
	Particle	
	Given Name	<b>Sinos</b>
	Suffix	
	Division	Department of Biology
	Organization	University of Patras
	Address	26500, Patras, Greece
	Email	
Schedule	Received	15 January 2016
	Revised	
	Accepted	21 February 2016
Abstract	The correct identification of individuals is a requirement of capture-mark-recapture (CMR) methods, and it is commonly achieved by applying artificial marks or by mutilation of study-animals. An alternative, non-invasive method to identify individuals is to utilize the patterns of their natural body markings. However, the use of pattern mapping is not yet widespread, mainly because it is considered time consuming, particularly in large populations and/or long-term CMR studies. Here we explore the use of pattern mapping for the identification of adult individuals in the alpine ( <i>Ichthyosaura alpestris</i> ) and smooth ( <i>Lissotriton vulgaris</i> ) newts (Amphibia, Salamandridae), using the freely available, open-source software	

Wild-ID. Our photographic datasets comprised nearly 4000 captured animals' images, taken during a 3-year period. The spot patterns of individual newts of both species did not change through time, and were sufficiently varied to allow their individual identification, even in the larger datasets. The pattern-recognition algorithm of Wild-ID was highly successful in identifying individual newts in both species. Our findings indicate that pattern mapping can be successfully employed for the identification of individuals in large populations of a broad range of animals that exhibit natural markings. The significance of pattern-mapping is accentuated in CMR studies that aim in obtaining long-term information on the demography and population dynamics of species of conservation interest, such as many amphibians facing population declines.

---

Keywords (separated by '-')	Photo capture-mark-recapture - Computer-assisted photo-identification - Non-invasive individual identification - Photo-id - Wild-ID
-----------------------------	---

---

Footnote Information

---

5 Onoufriou Mettouri · George Megremis  
6 Sinos Giokas

7 **A newt does not change its spots: using pattern mapping**  
8 **for the identification of individuals in large populations**  
9 **of newt species**

10 Received: 15 January 2016 / Accepted: 21 February 2016  
11 © The Ecological Society of Japan 2016

12 **Abstract** The correct identification of individuals is a  
13 requirement of capture-mark-recapture (CMR) meth-  
14 ods, and it is commonly achieved by applying artificial  
15 marks or by mutilation of study-animals. An alternative,  
16 non-invasive method to identify individuals is to utilize  
17 the patterns of their natural body markings. However,  
18 the use of pattern mapping is not yet widespread, mainly  
19 because it is considered time consuming, particularly in  
20 large populations and/or long-term CMR studies. Here  
21 we explore the use of pattern mapping for the identifi-  
22 cation of adult individuals in the alpine (*Ichthyosaura*  
23 *alpestris*) and smooth (*Lissotriton vulgaris*) newts (Am-  
24 phibia, Salamandridae), using the freely available, open-  
25 source software Wild-ID. Our photographic datasets  
26 comprised nearly 4000 captured animals' images, taken  
27 during a 3-year period. The spot patterns of individual  
28 newts of both species did not change through time, and  
29 were sufficiently varied to allow their individual identi-  
30 fication, even in the larger datasets. The pattern-recog-  
31 nition algorithm of Wild-ID was highly successful in  
32 identifying individual newts in both species. Our findings  
33 indicate that pattern mapping can be successfully em-  
34 ployed for the identification of individuals in large  
35 populations of a broad range of animals that exhibit  
36 natural markings. The significance of pattern-mapping is  
37 accentuated in CMR studies that aim in obtaining long-  
38 term information on the demography and population  
39 dynamics of species of conservation interest, such as  
40 many amphibians facing population declines.

41 **Keywords** Photo capture-mark-recapture ·  
42 Computer-assisted photo-identification · Non-invasive  
43 individual identification · Photo-id · Wild-ID

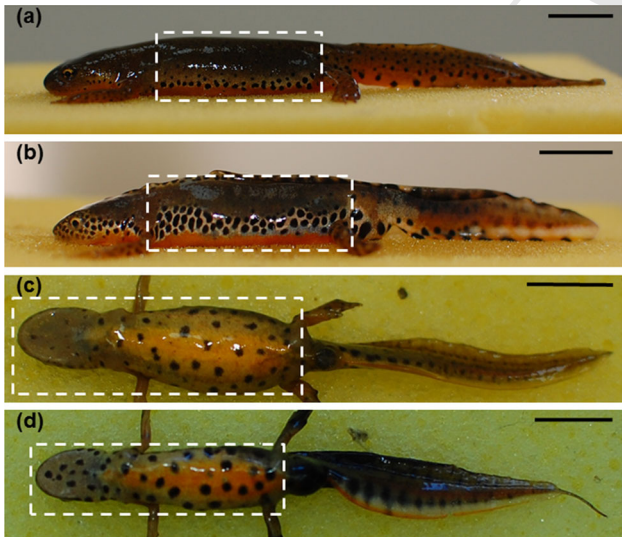
O. Mettouri (✉) · G. Megremis · S. Giokas  
Department of Biology, University of Patras, 26500 Patras, Greece  
E-mail: omettouri@gmail.com

**Introduction**

Capture-mark-recapture (CMR) methods are widely used in ecological studies to estimate population parameters (e.g., abundance, survival rates) and collect data on the demography, migration and life cycle of animals; the ability to distinguish individuals from other conspecifics in natural populations is fundamental to this process (Lebreton et al. 1992; Williams et al. 2002). Individual identification is most commonly achieved by applying artificial marks (e.g., PIT-tags, visible implant elastomers, bands, tattoos) or by removing small parts (e.g., toe or scale clipping or tipping) of animals. Some of these invasive techniques may induce stress and influence the survival and/or behaviour of the target animal (Gauthier-Clerc et al. 2004; McCarthy and Parris 2004; Langkilde and Shine 2006; Antwis et al. 2014), and are often expensive, particularly in long-term studies (Morrison et al. 2011). In addition, there have been concerns about ethical and animal welfare issues that arise from their broad use (Wilson and McMahon 2006; Parris et al. 2010; Perry et al. 2011).

An alternative, non-invasive method to identify individuals is to utilize their natural body markings, such as stripes, spots or blotches. These natural patterns are photographed and then used to distinguish between individuals. Although pattern mapping is a cheap and non-invasive method, it is considered very time-consuming (Donnelly et al. 1994), particularly in large datasets and/or long-term CMR studies. However, recent technological advances in digital photography and pattern recognition algorithms (Arzoumanian et al. 2005; Van Tienhoven et al. 2007; Gamble et al. 2008; Bolger et al. 2012) have enabled field ecologists to create and analyse large photographic databases quickly and efficiently while minimizing effort and misidentification errors (Bolger et al. 2012), thus allowing for larger sample sizes in CMR studies. In recent years pattern mapping has been employed in a wide and quite varied range of

88 animals, including beetles (Caci et al. 2013), seadragons  
 89 (Martin-Smith 2011), lizards (Sreekar et al. 2013), gir-  
 90 affes (Halloran et al. 2015) and dolphins (Martinho et al.  
 91 2014).  
 92 The global declines of many amphibian populations  
 93 (e.g. Blaustein and Wake 1990; Stuart et al. 2004, 2008)  
 94 stress the need for long-term information on their  
 95 demography and population dynamics (Blaustein et al.  
 96 1994). Long-term monitoring studies employing CMR  
 97 methods are an important tool for estimating abun-  
 98 dances and then identifying negative population-trends.  
 99 Although most amphibians exhibit some form of natural  
 100 patterns, the temporal inconsistency of these patterns  
 101 documented in some species and the fact that it is con-  
 102 sidered a time-consuming method (Donnelly et al. 1994)  
 103 prevent the wide-spread use of pattern mapping in  
 104 amphibian CMR studies. Instead, more traditional  
 105 methods such as toe-clipping and PIT-tagging are usu-  
 106 ally preferred (Ferner 2010), despite their invasive nature  
 107 and the fact that many of the target-species are of con-  
 108 servation interest. Adult newts (Amphibia, Salaman-  
 109 dridae) exhibit natural spot patterns that can be utilized  
 110 for the identification of individuals. Alpine newts, *Ich-*  
 111 *thyosaura alpestris* (Laurenti, 1768), have numerous  
 112 spots on their flanks, and smooth newts, *Lissotriton*  
 113 *vulgaris* (Linnaeus, 1758), have spots on their belly and  
 114 throat areas (Figs. 1 and 2). Hagström (1973) and  
 115 Winkler and Heunisch (1997) used these spot patterns to  
 116 successfully identify individuals of smooth and alpine  
 117 newts, albeit using a small number of animals because  
 118 the method was at the time considered impractical for  
 119 large populations (Hagström 1973).



**Fig. 1** Spot patterns of male and female alpine and smooth newts. The dashed white lines in each image indicate the cropped region that was used for the identification of individuals with Wild-ID. **a** *Ichthyosaura alpestris* female, **b** *I. alpestris* male, **c** *Lissotriton vulgaris* female, **d** *L. vulgaris* male. The black scale bars on the right represent 1 cm

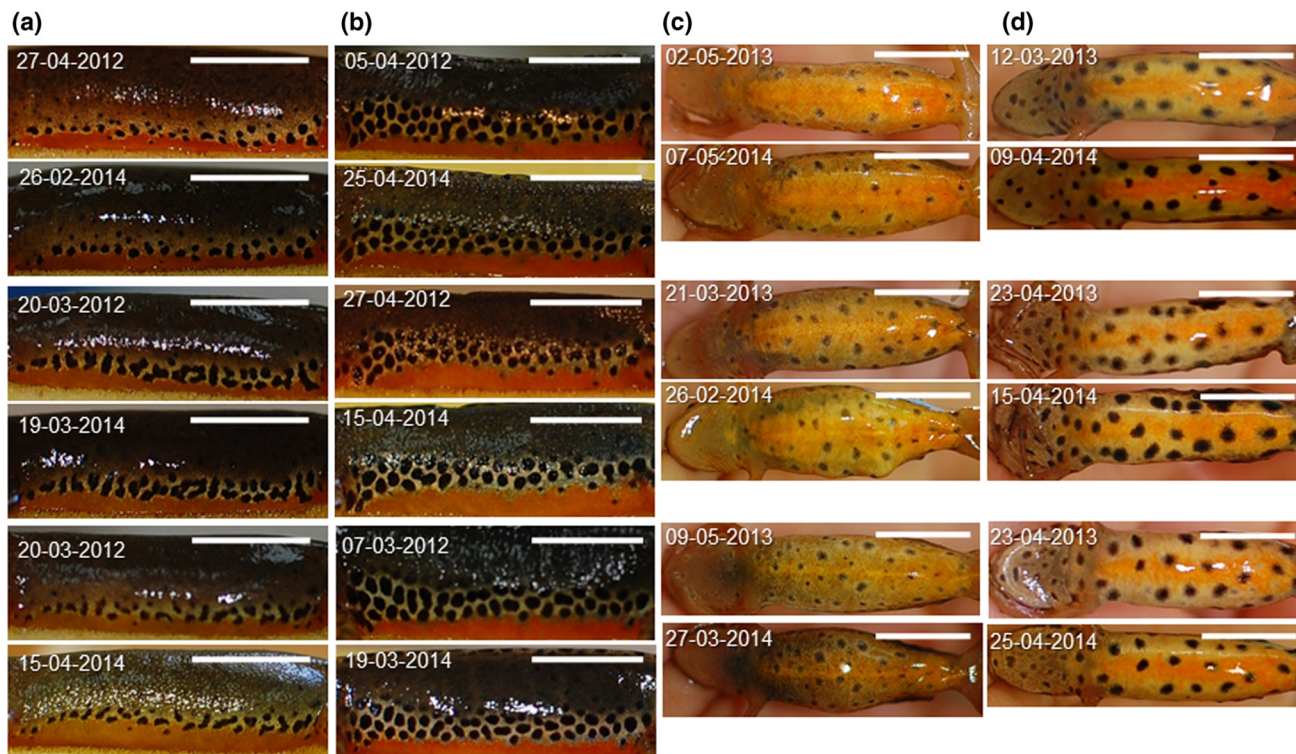
The correct identification of individuals is a basic  
 assumption of CMR methods, and violation of this  
 assumption may lead to negatively biased population-  
 parameter estimates (Morrison et al. 2011). Thus,  
 when using the natural patterns of individuals as a  
 marking method, it is vital to ensure that the patterns  
 of the target-species are both sufficiently varied and  
 distinct to allow for individual identification, and re-  
 main unchanged at least for the duration of the study  
 (Ferner 2010). Here, we explore the use of pattern  
 mapping for the identification of individuals in the  
 alpine and smooth newts over a 3-year time period  
 using the freely available, open-source software Wild-  
 ID (Bolger et al. 2012). Our aims are: (1) to assess the  
 efficacy of using the spot patterns of alpine and  
 smooth newts for their individual identification in real  
 CMR datasets varying in size, (2) to test the temporal  
 consistency of the spot patterns of these individuals  
 over the 3-year study period and (3) to evaluate the  
 performance and reliability of Wild-ID as a tool for  
 computer-assisted pattern recognition in the two tar-  
 get-species.

## Methods

### Study site and field work

Our study site is a semi-natural temporary pond in  
 northern Peloponnese, southern Greece (38°01'N,  
 22°02'E, about 800 m a.s.l.), in which adult alpine (*I.*  
*alpestris*) and smooth (*L. vulgaris*) newts gather each  
 year to breed. Sampling was carried out for three con-  
 secutive years (2012–2014), approximately twice a  
 month (2012) or once a week (2013–2014) during the  
 newt's breeding season, using dip-nets to capture adults  
 in the water. Because accurate morphometric data (not  
 shown here) on all captured newts were required, it was  
 deemed necessary to anaesthetize them. We note, how-  
 ever, that anaesthesia is not strictly required for taking  
 photographs, and other methods for immobilizing cap-  
 tured newts for a few seconds can be applied (e.g., Baker  
 and Gent 1998). We anaesthetised all captured individ-  
 uals in a solution of 0.25 g MS222 (Sigma–Aldrich) in  
 1L water, in which 0.25 g of baking soda (sodium  
 bicarbonate) was added to neutralize the pH level. Each  
 newt was placed on a wet sponge and, after measure-  
 ment, photographs were taken of the lateral (left) side of  
 alpine newts and the ventral side of smooth newts. All  
 pictures were taken using a hand-held Nikon D40 DSLR  
 camera with a 35 mm f/1.8 Nikkor lens and using the  
 built-in flash when required. The handling time for each  
 anaesthetised individual was approximately five to ten  
 seconds. Depending on the number of captured newts  
 the whole process usually lasted for a few hours. After  
 making sure that all newts recovered successfully from  
 anaesthesia, they were released at the site of capture  
 within the day of capture.





**Fig. 2** Example images of alpine and smooth newts recaptured in different years that show virtually no change in their spot patterns. Three individuals per group (a–d) are shown. Each pair of images in the different columns (a–d) corresponds to the same individual,

captured in different years. The date of capture is shown at the upper left corner of each image. **a** *Ichthyosaura alpestris* females, **b** *I. alpestris* males, **c** *Lissotriton vulgaris* females, **d** *L. vulgaris* males. The white scale bars on the right represent 1 cm

## 174 Manual identification and Wild-ID evaluation

175 Breeding adult males and females can be easily distin-  
176 guished from each other in both species (Fig. 1), thus  
177 males and females of each species were treated as a  
178 different group. After the 2012 sampling was concluded  
179 we constructed four photographic datasets, one for each  
180 group. These datasets consisted of 298 alpine (162 fe-  
181 males, 136 males) and 49 smooth (36 females, 13 males)  
182 newts. To establish the identity (i.e., if they are new  
183 captures or recaptures) of all newts in the 2012 datasets  
184 we followed a manual identification procedure, in which  
185 by eye we compared the image of each captured newt to  
186 all other images in the same group; individuals captured  
187 at a given sampling session were compared to all indi-  
188 viduals from all previous sessions. We were thus able to  
189 unambiguously identify all captured animals and assign  
190 them either as recaptures or new captures (see “Re-  
191 sults”).

192 For the computer-assisted individual identification  
193 we used the pattern-recognition software Wild-ID  
194 (Bolger et al. 2012; available at <http://envs.dartmouth.edu/people/douglas-thomas-bolger>). Wild-ID processes  
195 the images in each dataset sequentially and calculates a  
196 similarity score for each pair of images. After all simi-  
197 larity scores are computed, Wild-ID presents the top-  
198 20 ranking images for each image for visual confirma-  
199

tion through the user interface. Consequently the final  
decision for accepting or rejecting a match is made by  
the user. We analysed the 2012 datasets with Wild-ID,  
using the results of the manual identification procedure  
as the known (i.e., correct) matches. The pre-processing  
stage consisted of the removal of unnecessary back-  
ground information (Fig. 1). The user of Wild-ID was  
blind as to the true identity of all individuals and in-  
spected all top-20 images presented for each unknown  
image by Wild-ID. To evaluate the performance of  
Wild-ID we estimated the total number of recaptures  
that were not placed within the top-20 ranking images,  
as well as the number of images wrongly assigned as  
recaptures.

## Within- and between-years identification

The encouraging results of Wild-ID for the first years  
(2012) datasets (see “Results”) prompted us to use it for  
the following 2 years datasets. During these years the  
total number of captured animals ( $N$ ) was much greater  
than the first year (2012:  $N = 347$ , 2013:  $N = 2462$ ,  
2014:  $N = 1190$ ). Although Wild-ID ranked all recap-  
tures of the first year well within the top-10 positions  
(see “Results”), we chose to inspect all top-20 ranking  
images in these much larger datasets to avoid missing

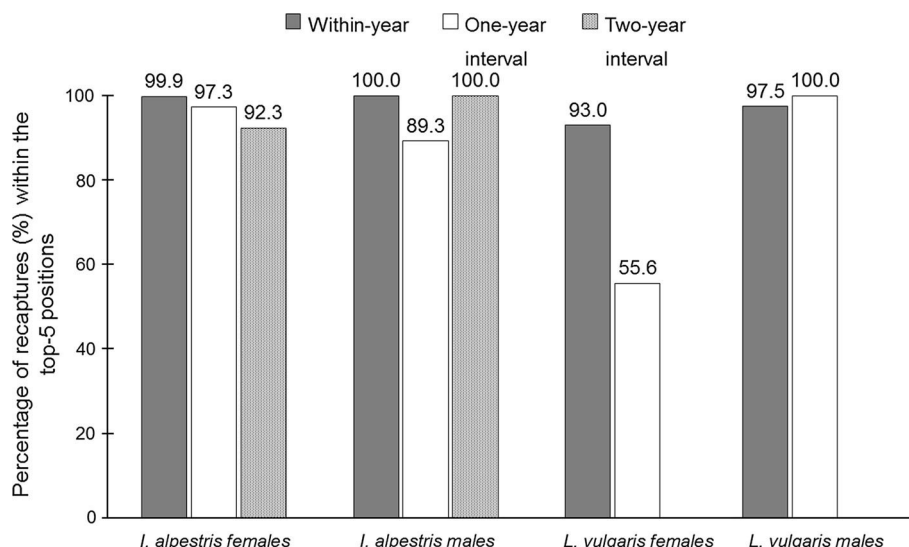
224	any recaptures and to assess the performance of Wild-ID	by the very high ranking-positions of most recaptures,	272
225	in large datasets. Again, the images of each years cap-	shown in Table 1 and Fig. 3.	273
226	tured newts were partitioned in four groups according to		
227	species and sex and were pre-processed to remove		
228	background information. We constructed one dataset	Within-year identification	274
229	for each group of each year (i.e., eight within-year da-		
230	taset), and each dataset was analysed separately with	In the alpine newt, 710 out of 722 (98.3 %) female and	275
231	Wild-ID.	514 out of 517 (99.4 %) male recaptures ranked first.	276
232	To assess whether an individual newt's spot pattern	Only four female and one male alpine newt images were	277
233	changed over time and to assess the ability of Wild-ID to	positioned below the top-3 ranking images. In the	278
234	identify individual newts captured across years, we cre-	smooth newt, 105 out of 129 (81.4 %) female and 73 out	279
235	ated three between-years datasets for each group: two	of 79 (92.4 %) male recaptures ranked first. Eleven fe-	280
236	datasets for the 1-year time interval (i.e., 2012–2013 and	male and two male smooth newt images ranked below	281
237	2013–2014) and one dataset for the 2-year time interval	the top-3 positions.	282
238	(2012–2014). In each of these datasets we searched for		
239	individuals that were last captured in the former year	Between-years identification	283
240	and recaptured in the latter. Again, we inspected all top-		
241	20 ranking images presented for each unknown image by	In both sexes of the alpine newt over 98 % of the	284
242	Wild-ID.	recaptured individuals in the 1-year interval dataset	285
243	<b>Results</b>	ranked within the top-10 positions. In the 2-year interval	286
244	Field work and temporal consistency of spot patterns	dataset, two out of 13 females and two out of eight	287
245	A total of 3333 (1902 females, 1431 males) alpine and	males ranked below the top-3 positions. In the smooth	288
246	666 (422 females, 244 males) smooth newts were cap-	newt all eight males ranked within the top-3 positions,	289
247	tured during the 3-year period (Table 1). The visual	while two females ranked below the 15 top-ranking	290
248	inspection of all within-year and between-years recap-	images in the 1-year interval dataset. Of the few smooth	291
249	tures revealed that the spot patterns of individual newts	newts captured in 2012 (36 females, 13 males), none were	292
250	of both species do not change over time. In Fig. 2 we	recaptured in 2014 (i.e., in the 2-year interval).	293
251	present some example images of newts of both species		
252	that were recaptured across years.	<b>Discussion</b>	294
253	Manual identification and Wild-ID evaluation (2012		
254	dataset)	In this study we assess the efficacy of using the spot	295
255	The spot patterns of individual newts in all four datasets	patterns of alpine and smooth newts for their individual	296
256	of 2012 were sufficiently varied to allow for the unam-	identification. Our inspection of nearly 4000 captured	297
257	biguous identification of all individuals using the manual	animals' images, taken during a 3-year time period, re-	298
258	identification procedure. We were thus able to assign	veals that the spot patterns of alpine and smooth newts	299
259	with certainty all captured animals as either recaptures	are sufficiently varied to allow their individual identifi-	300
260	or new captures.	cation. Even in the larger datasets, containing hundreds	301
261	Wild-ID successfully identified all recaptures in all	or thousands of images, there was virtually no difficulty	302
262	groups and placed them in the top-20 ranking images.	in assigning unknown individuals as recaptures or new	303
263	Additionally, none of the images were wrongly assigned	captures based on their spot patterns. However, with the	304
264	as recaptures in any group. Of the 25 recaptures, 22	number of images growing, manual inspection of a	305
265	(88 %) ranked first, 24 (96 %) ranked within the top-3	photographic dataset becomes increasingly harder and	306
266	and all 25 recaptures ranked within the top-10 posi-	potentially more error-prone, and it demands consider-	307
267	tions.	ably more effort. The employment of pattern mapping in	308
268	Within-year and between-years identification with Wild-	such large datasets without the aid of pattern-recogni-	309
269	ID	tion software would be forbiddingly time-consuming.	310
270	The pattern-recognition algorithm of Wild-ID was	Inspection of the four 2012 datasets with Wild-ID took	311
271	highly successful in all datasets and groups, as indicated	only a fraction of the time required for their manual	312
		inspection and yielded the same results, with no	313
		misidentification errors. The following 2 years' datasets,	314
		although much larger than the first year's, were not	315
		harder to inspect, as we only had to visually go through	316
		the top-20 closest-matching images presented by Wild-	317
		ID. However, as the vast majority of recaptures were	318
		ranked within the top-5 positions in all datasets (Fig. 3),	319
		Wild-ID greatly reduced the time needed for visual	320

**Table 1** Ranking positions of recaptures in Wild-ID for each group (i.e., species and sex) and each dataset

Group	Dataset	Total recaptures	Wild-ID ranking position									
			Top 1	Top 2	Top 3	Top 4	Top 5	Top 10	Top 15	Top 20		
<i>Ichthyosaura alpestris</i> females ( <i>N</i> = 1902)	Within-year	<i>N<sub>r</sub></i> %	710	714	718	720	721	721	721	722		
	1-year interval	<i>N<sub>r</sub></i> %	98.34	98.89	99.45	99.72	99.86	99.86	99.86	100		
	2-year interval	<i>N<sub>r</sub></i> %	166	173	175	178	179	182	184	184		
<i>I. alpestris</i> males ( <i>N</i> = 1431)	Within-year	<i>N<sub>r</sub></i> %	90.22	94.02	95.11	96.74	97.28	98.91	100	100		
	1-year interval	<i>N<sub>r</sub></i> %	8	9	11	12	12	12	12	13		
	2-year interval	<i>N<sub>r</sub></i> %	61.54	69.23	84.62	92.31	92.31	92.31	92.31	100		
<i>Lissonotriton vulgaris</i> females ( <i>N</i> = 422)	Within-year	<i>N<sub>r</sub></i> %	514	514	516	517	517	517	517	517		
	1-year interval	<i>N<sub>r</sub></i> %	99.42	99.42	99.81	100	100	100	100	100		
	2-year interval	<i>N<sub>r</sub></i> %	46	47	48	48	50	55	56	56		
<i>L. vulgaris</i> males ( <i>N</i> = 244)	Within-year	<i>N<sub>r</sub></i> %	82.14	83.93	85.71	85.71	89.29	98.21	100	100		
	1-year interval	<i>N<sub>r</sub></i> %	4	6	6	8	8	8	8	8		
	2-year interval	<i>N<sub>r</sub></i> %	50	75	75	100	100	100	100	100		
<i>L. vulgaris</i> males ( <i>N</i> = 244)	Within-year	<i>N<sub>r</sub></i> %	105	115	118	119	120	125	128	129		
	1-year interval	<i>N<sub>r</sub></i> %	81.40	89.15	91.47	92.25	93.02	96.90	99.22	100		
	2-year interval	<i>N<sub>r</sub></i> %	2	3	3	4	5	5	7	9		
<i>L. vulgaris</i> males ( <i>N</i> = 244)	Within-year	<i>N<sub>r</sub></i> %	22.22	33.33	33.33	44.44	55.56	55.56	77.78	100		
	1-year interval	<i>N<sub>r</sub></i> %	73	75	77	77	77	77	77	79		
	2-year interval	<i>N<sub>r</sub></i> %	92.41	94.94	97.47	97.47	97.47	97.47	97.47	100		
<i>L. vulgaris</i> males ( <i>N</i> = 244)	Within-year	<i>N<sub>r</sub></i> %	5	6	8	8	8	8	8	8		
	1-year interval	<i>N<sub>r</sub></i> %	62.50	75	100	100	100	100	100	100		
	2-year interval	<i>N<sub>r</sub></i> %	4	6	6	8	8	8	8	8		

Within-year: animals recaptured in at least 1 year's sampling, pooled for all years (2012, 2013, 2014). One-year interval: animals recaptured 1 year after last been captured, pooled for both intervals (2012–2013 and 2013–2014). Two-year interval: animals recaptured 2 years after last been captured; only *Ichthyosaura alpestris* individuals were recaptured in the 2-year interval. *N* number of animals captured over the 3-year study period, *N<sub>r</sub>* number of recaptures ranked within each ranking position, % percentage of recaptures ranked within each position over the total number of recaptures





**Fig. 3** Percentage of recaptures (%) that were ranked by Wild-ID within the top-5 positions for each group and each dataset

confirmation. Because of the high-ranking success of Wild-ID across all datasets, the likelihood of having missed any recaptures is assumed to be negligible. Up until recently, and compared to other marking methods, pattern mapping was considered appropriate only for short studies of small populations (Hagström 1973; Donnelly et al. 1994; Arntzen et al. 2004); this study clearly shows that this is not the case anymore.

In CMR studies employing pattern mapping for individual identification, it is crucial that the spot patterns of individuals are identifiable throughout the duration of the study (Ferner 2010). The visual comparison of images of individuals recaptured across years (Fig. 2) reveals that the spot patterns of individual newts of both species do not change through time. This is most clearly demonstrated in the case of newts that were recaptured in both the first and last year of the study. Although only alpine newts were recaptured across the 2-year interval, a close inspection of smooth newt images in the 1-year interval datasets confirms the temporal consistency of spot patterns in both newt species.

In the within-year datasets, alpine newts ranked higher than smooth newts, and male newts in both species ranked higher than female newts (Table 1). Alpine newts have a greater number of spots than smooth newts; in addition, male alpine newts have more spots than female alpine newts, while male smooth newts have bigger spots than female smooth newts (Fig. 1 and 2). The larger number and bigger size of spots enabled Wild-ID to distinguish more efficiently among individuals and thus rank them in higher positions. Nonetheless, the high ranking positions in all groups indicate that the performance of Wild-ID is equally satisfactory with spots of different numbers and sizes. In the between-years datasets, the ranking positions were high, although lower than those of the within-year datasets (Table 1). An inspection of the spot patterns of individuals that were recaptured between years (Fig. 2) indicates that the lower ranking positions are not a result

of these patterns changing through time. Rather, they may be a consequence of small changes in body shape as a result of weight change. Newts recaptured between years are very unlikely to have the exact same weight. This natural variation in weight, which is likely to be greater between than within years, may lead to slight differences in body shape and size, giving the individual an overall different appearance between years. However, the high ranking positions of the vast majority of between-year images by Wild-ID indicate that these small changes in body shape do not hinder the algorithm's ability to identify individuals recaptured across years.

Wild-ID has already been used with great success in various animal taxa (Morrison and Bolger 2012; Bendik et al. 2013; Cross et al. 2014; Elgue et al. 2014; Halloran et al. 2015). This lack of species-specificity, and the encouraging results reported here and in these previous studies, indicate that Wild-ID can be successfully applied in a wide range of animals that exhibit natural markings, such as many amphibian species. In our study Wild-ID performed equally satisfactory in both newt species, demonstrating that it is capable of handling both ventral/dorsal and lateral patterns. Thus, computer-assisted pattern mapping using pattern-recognition algorithms such as Wild-ID has the potential to become a widely-applied method for individual identification in a broad range of animals. The significance of pattern mapping is further accentuated in CMR studies that aim in obtaining long-term information on the demography and population dynamics of species of conservation interest, such as many amphibians facing population declines.

**Acknowledgments** We would like to thank Eva Pitta and Christos Papathanasiou for their valuable help during field work, the staff of the Management Body of Chelmos - Vouraikos National Park for providing their facilities and assistance, Kostas Sotiropoulos for helpful discussions and two anonymous reviewers for helpful comments and suggestions.

398 **Compliance with ethical standards** This study complies with the  
399 current laws of Greece.

400 **Conflict of interest** The authors declare that they have no conflict  
401 of interest.

## 402 References

403 Antwis RE, Garcia G, Fidgett AL, Preziosi RF (2014) Tagging  
404 frogs with passive integrated transponders causes disruption of  
405 the cutaneous bacterial community and proliferation of  
406 opportunistic fungi. *Appl Environ Microbiol* 80:4779–4784.  
407 doi:10.1128/AEM.01175-14

408 Arntzen JW, Goudie IJB, Halley J, Jehle R (2004) Cost comparison  
409 of marking techniques in long-term population studies: PIT-  
410 tags versus pattern maps. *Amphibia-Reptilia* 25:305–315. doi:  
411 10.1163/1568538041975116

412 Arzoumanian Z, Holmberg J, Norman B (2005) An astronomical  
413 pattern-matching algorithm for computer-aided identification  
414 of whale sharks *Rhincodon typus*. *J Appl Ecol* 42:999–1011. doi:  
415 10.1111/j.1365-2664.2005.01117.x

416 Baker J, Gent T (1998) Marking and recognition of animals. In:  
417 Gent T, Gibson S (eds) *Herpetofauna workers' manual*. Joint  
418 Nature Conservation Committee, Peterborough, pp 45–54

419 Bendik NF, Morrison TA, Gluesenkamp AG, Sanders MS,  
420 O'Donnell LJ (2013) Computer-assisted photo identification  
421 outperforms visible implant elastomers in an endangered sala-  
422 mander, *Eurycea tonkawae*. *PLoS One* 8:e59424. doi:  
423 10.1371/journal.pone.0059424

424 Blaustein AR, Wake DB (1990) Declining amphibian populations:  
425 a global phenomenon. *Trends Ecol Evol* 5:203–204. doi:  
426 10.1016/0169-5347(90)90129-2

427 Blaustein AR, Wake DB, Sousa WP (1994) Amphibian declines:  
428 judging stability, persistence, and susceptibility of populations  
429 to local and global extinctions. *Conserv Biol* 8:60–71. doi:  
430 10.1046/j.1523-1739.1994.08010060.x

431 Bolger DT, Morrison TA, Vance B, Lee D, Farid H (2012) A  
432 computer-assisted system for photographic mark-recapture  
433 analysis. *Methods Ecol Evol* 3:813–822. doi:10.1111/j.2041-  
434 210X.2012.00212.x

435 Caci G, Biscaccianti AB, Cistrone L, Bosso L, Garonna AP, Russo D  
436 (2013) Spotting the right spot: computer-aided individual identi-  
437 fication of the threatened cerambycid beetle *Rosalia alpina*. *J Insect*  
438 *Conserv* 17:787–795. doi:10.1007/s10841-013-9561-0

439 Cross MD, Lipps GJ Jr, Sapak JM, Tobin EJ, Root KV (2014)  
440 Pattern-recognition software as a supplemental method of  
441 identifying individual eastern box turtles (*Terrapene c. carolina*).  
442 *Herpetol Rev* 45:584–586

443 Donnelly MA, Guyer C, Juterbock JE, Alford RA (1994) Techniques  
444 for marking amphibians. In: Heyer W, Donnelly MA, McDiarmid  
445 RA, Hayek LC, Foster MS (eds) *Measuring and monitoring*  
446 *biological diversity: standard methods for amphibians*. Smith-  
447 *sonian Institution Press*, Washington, DC, pp 277–284

448 Elgue E, Pereira G, Achaval-Coppes F, Maneyro R (2014) Validity  
449 of photo-identification technique to analyze natural markings  
450 in *Melanophryniscus montevidensis* (Anura: Bufonidae). *Phyl-*  
451 *lomedusa* 13:59–66. doi:10.11606/issn.2316-9079.v13i1p59-66

452 Ferner JW (2010) Measuring and marking post-metamorphic  
453 amphibians. In: Dodd CK Jr (ed) *Amphibian ecology and*  
454 *conservation: a handbook of techniques*. Oxford University  
455 *Press*, Oxford, pp 123–141

456 Gamble L, Ravela S, McGarigal K (2008) Multi-scale features for  
457 identifying individuals in large biological databases: an appli-  
458 cation of pattern recognition technology to the marbled sala-  
459 mander *Ambystoma opacum*. *J Appl Ecol* 45:170–180. doi:  
460 10.1111/j.1365-2664.2007.01368.x

461 Gauthier-Clerc M, Gendner J-P, Ribic CA, Fraser WR, Woehler  
462 EJ, Descamps S, Gilly C, Le Bohec C, Le Maho Y (2004) Long-  
463 term effects of flipper-bands on penguins. *Proc R Soc Lond B*  
464 271:S423–S426. doi:10.1098/rsbl.2004.0201

Hagström T (1973) Identification of newt specimens (*Urodela*,  
465 *Triturus*) by recording the belly pattern and a description of  
466 photographic equipment for such registrations. *Br J Herpetol*  
467 4:321–326

468 Halloran KM, Murdoch JD, Becker MS (2015) Applying com-  
469 puter-aided photo-identification to messy datasets: a case study  
470 of Thornicroft's giraffe (*Giraffa camelopardalis thornicrofti*).  
471 *Afr J Ecol* 53:147–155. doi:10.1111/aje.12145

472 Langkilde T, Shine R (2006) How much stress do researchers inflict  
473 on their study animals? A case study using a scincid lizard,  
474 *Eulamprus heatwolei*. *J Exp Biol* 209:1035–1043. doi:  
475 10.1242/jeb.02112

476 Lebreton JD, Burnham KP, Clobert J, Anderson DR (1992)  
477 Modelling survival and testing biological hypotheses using  
478 marked animals: a unified approach with case studies. *Ecol*  
479 *Monogr* 62:67–118. doi:10.2307/2937171

480 Martinho F, Pereira A, Brito C, Gaspar R, Carvalho I (2014)  
481 Structure and abundance of bottlenose dolphins (*Tursiops*  
482 *truncatus*) in coastal Setúbal Bay, Portugal. *Mar Biol Res*  
483 11:144–156. doi:10.1080/17451000.2014.894244

484 Martin-Smith KM (2011) Photo-identification of individual weedy  
485 seadragons *Phyllopteryx taeniolatus* and its application in esti-  
486 mating population dynamics. *J Fish Biol* 78:1757–1768. doi:  
487 10.1111/j.1095-8649.2011.02966.x

488 McCarthy MA, Parris KM (2004) Clarifying the effect of toe  
489 clipping on frogs with Bayesian statistics. *J Appl Ecol*  
490 41:780–786. doi:10.1111/j.0021-8901.2004.00919.x

491 Morrison TA, Bolger DT (2012) Wet season range fidelity in a  
492 tropical migratory ungulate. *J Anim Ecol* 81:543–552. doi:  
493 10.1111/j.1365-2656.2011.01941.x

494 Morrison TA, Yoshizaki J, Nichols JD, Bolger DT (2011) Esti-  
495 mating survival in photographic capture-recapture studies:  
496 overcoming misidentification error. *Methods Ecol Evol*  
497 2:454–463. doi:10.1111/j.2041-210X.2011.00106.x

498 Parris KM, McCall SC, McCarthy MA, Minter BA, Steele K,  
499 Bekessy S, Medvecky F (2010) Assessing ethical trade-offs in  
500 ecological field studies. *J Appl Ecol* 47:227–234. doi:  
501 10.1111/j.1365-2664.2009.01755.x

502 Perry G, Wallace MC, Perry D, Curzer H, Muhlberger P (2011)  
503 Toe clipping of amphibians and reptiles: science, ethics, and the  
504 law. *J Herpetol* 45:547–555. doi:10.1670/11-037.1

505 Sreekar R, Purushotham CB, Saini K, Rao SN, Pelletier S,  
506 Chaplod S (2013) Photographic capture-recapture sampling for  
507 assessing populations of the Indian gliding lizard *Draco dus-*  
508 *sumieri*. *PLoS One* 8:e55935. doi:10.1371/journal.pone.0055935

509 Stuart SN, Chanson JS, Cox NA, Young BE, Rodrigues ASL,  
510 Fischman DL, Waller RW (2004) Status and trends of  
511 amphibian declines and extinctions worldwide. *Science*  
512 306:1783–1786. doi:10.1126/science.1103538

513 Stuart SN, Hoffmann M, Chanson JS, Cox NA, Berridge RJ,  
514 Ramani P, Young BE (2008) *Threatened Amphibians of the*  
515 *world*. Lynx Edicions, Barcelona

516 Van Tienhoven AM, Den Hartog JE, Reijns RA, Peddemors VM  
517 (2007) A computer-aided program for pattern-matching of natural  
518 marks on the spotted raggedtooth shark *Carcharias taurus*. *J Appl*  
519 *Ecol* 44:273–280. doi:10.1111/j.1365-2664.2006.01273.x

520 Williams BK, Nichols JD, Conroy MJ (2002) Analysis and man-  
521 agement of animal populations: modeling, estimation and  
522 decision-making. Academic Press, San Diego

523 Wilson RP, McMahon CR (2006) Measuring devices on wild ani-  
524 mals: what constitutes acceptable practice? *Front Ecol Environ*  
525 4:147–154. doi:10.1890/1540-9295(2006)004[0147:MDOWA  
526 W]2.0.CO;2

527 Winkler C, Heunisch G (1997) Fotografische Methoden der Indi-  
528 vidualerkennung bei Bergmolch (*Triturus alpestris*) und  
529 Fadenmolch (*T. helveticus*) (Urodela, Salamandridae).  
530 *Mertensiella* 7:71–77