

CDM5104

Experiment in R

```
# Run this first to install packages
# install.packages(c("tidyverse", "here", "broom", "purrr"))
library(tidyverse)

## -- Attaching packages ----- tidyverse 1.3.2 --
## v ggplot2 3.4.1    v purrr   1.0.1
## v tibble  3.1.8    v dplyr   1.1.0
## v tidyr   1.3.0    v stringr 1.5.0
## v readr   2.1.4    v forcats 1.0.0
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()    masks stats::lag()
```

```
library(here)
```

```
## here() starts at /Users/lsibjb/Code/CDMS4104_Lipidomics
```

```
library(broom)
library(purrr)
```

Analysing lipid sum compositions

Import and prepare data

Read dataset, convert to long format and retrieve lipid class name from lipid name. Note: lipid class name is assumed to be anything before the first space. For ether and plasmalogen PCs (e.g. PE O- and PE P-), we change to the lipid name so that also for those the class name is before the first space.

Data from: Boretto et al., Metabolites, 2019 <https://doi.org/10.1007/s11306-019-1621-3>

```
d_orig <- readr::read_csv(here("data/DogDietStudy_PlasmaLipidomics_Diets-12_CONCENTRATIONS.csv"))

## Rows: 16 Columns: 316
## -- Column specification -----
## Delimiter: ","
## chr   (1): Diet
## dbl (315): DogID, CE 16:1, CE 18:1, CE 18:2, CE 18:3, CE 20:2, CE 20:3, CE 2...
##
## i Use 'spec()' to retrieve the full column specification for this data.
## i Specify the column types or set 'show_col_types = FALSE' to quiet this message.
```

```
# Prepare data: convert to long, get LipidClass names
d_long <- d_orig |>
  pivot_longer(-DogID:-Diet, names_to = "Lipid" , values_to = "Conc") |>
  mutate(Lipid = str_replace(Lipid, fixed(" P-"), "-P "),
         Lipid = str_replace(Lipid, fixed(" 0-"), "-0 ")) |>
  separate(col = Lipid, into = c("LipidClass", "Chain"), remove = FALSE, sep = " ")
```

Calculate sum per class per sample

```
# Calculate sums of all species per class
d_sum <- d_long |>
  group_by(DogID, Diet, LipidClass) |>
  summarise(Conc = sum(Conc))
```

'summarise()' has grouped output by 'DogID', 'Diet'. You can override using the
'.groups' argument.

```
# Save data as CSV file
write_csv(x = d_sum, file = here("output/DogDietStudy_SumCompositions.csv"))
head(d_sum)
```

```
## # A tibble: 6 x 4
## # Groups:   DogID, Diet [1]
##   DogID Diet LipidClass Conc
##   <dbl> <chr> <chr>    <dbl>
## 1     1 Diet-1 CE      8814.
## 2     1 Diet-1 Cer      5.57
## 3     1 Diet-1 DG      129.
## 4     1 Diet-1 GM3      1.28
## 5     1 Diet-1 Hex1Cer  0.547
## 6     1 Diet-1 Hex2Cer  1.32
```

Perform a Welch's t test between diet 1 and diet 2

We could also consider to log transform the data

```
# get t test results (p, FDR, and fold-change)
d_stat <- d_sum |>
  group_by(LipidClass) |>
  nest() |>
  mutate(res = purrr::map(data, \(x) broom::tidy(
    t.test(Conc ~ Diet, var.equal = FALSE, data = x)
  )) |>
  unnest(res) |>
  ungroup() |>
  mutate(FDR = p.adjust(p.value, method = "BH"),
         log2FC = log2(estimate2/estimate1)) |>
  select(LipidClass, log2FC, p.value, FDR)

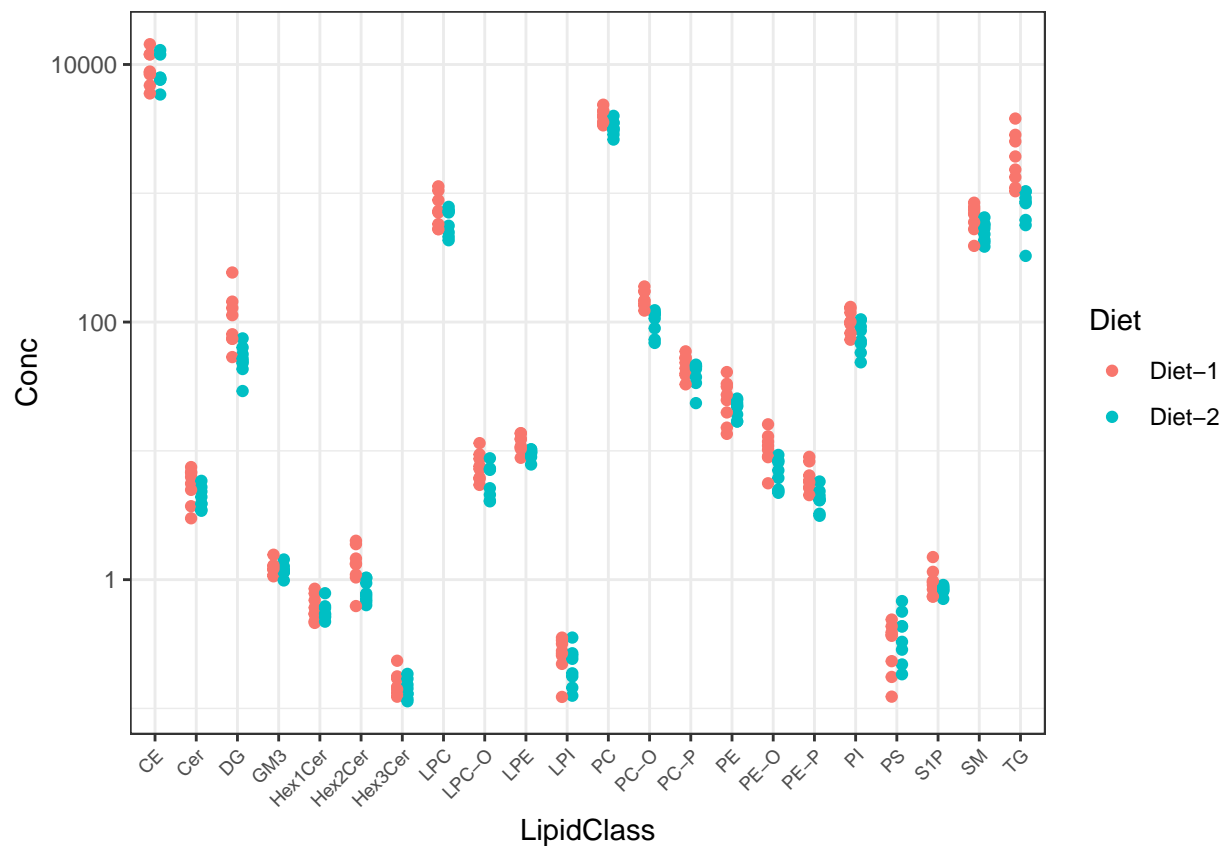
head(d_stat)
```

```
## # A tibble: 6 x 4
##   LipidClass log2FC p.value   FDR
##   <chr>      <dbl>   <dbl> <dbl>
## 1 CE        -0.148   0.491 0.495
## 2 Cer       -0.350   0.0871 0.168
## 3 DG        -1.13    0.0234 0.0643
## 4 GM3       -0.0590  0.495  0.495
## 5 Hex1Cer   -0.111   0.467  0.495
## 6 Hex2Cer   -0.718   0.0127 0.0399
```

Simple Dot plot

Note: y axis is in log scale

```
ggplot(d_sum, aes(x=LipidClass, y = Conc, group = Diet, color = Diet)) +
  geom_point(position = position_dodge(width = 0.5)) +
  scale_y_log10() +
  theme_bw() +
  theme(axis.text.x = element_text(size = 7, angle = 45, hjust = 1))
```



Simple volcano plot

```
ggplot(d_stat, aes(x=log2FC, y = -log10(p.value), label=LipidClass)) +
  geom_point() +
  geom_text(nudge_y = -0.1, nudge_x = -0.1, size=3) +
  scale_x_continuous(limits = c(-2,2)) +
  geom_vline(xintercept = 0, color = "grey50", linewidth = .4) +
  geom_hline(yintercept = -log10(0.05), color = "green") +
  theme_bw() +
  theme(aspect.ratio=1)
```

