

Useful notes on *let-60* (*ras*)

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Analysis of dominant-negative mutations of *let-60* (*ras*)

See paper by Min Han and Sternberg¹. Briefly, Min Han found two classes of dominant negative mutations: *dn* mutations that would cause a weakly penetrant Muv phenotype when injected into wild type animals and *dn* mutations that never cause a Muv phenotype when injected into wild type animals.

From studying one allele in each class, they show: Non-Muv alleles:

1. (sy99, s101), (sy92, sy95, sy100). Parentheses signify different strains containing the same mutations.
2. Are lethal dominant. This lethality is strongly suppressed in a dosage-independent manner by *let-60(lf)*.
3. Vulval differentiation increases with decreasing *dn* dosage: $dn/dn/+ < dn/+ < dn/+ /+ = 1$.

Muv alleles (AA 119 and 16):

1. (sy93), (sy94)
2. Are recessive viable, so $+/Df$ is dead.
3. Are toxic to wild-type product, so vulval differentiation looks strange: $dn/+ < dn/dn \sim dn/+ /+ < wt < dn/dn/+$.
4. In $dn/dn/+$ animals, signaling is partially signal-independent.

sur-1/mpk-1 acts downstream of *let-60* (*ras*)

See Yan Wu and Min Han². 10% of *sur-1* (ku1) homozygotes have a P6.p non-differentiation phenotype. *sur-1* is partially dominant over *let-60* (*ras*). Null homozygotes are dead.

ksr-1 encodes a novel raf-related kinase involved in Ras-mediated signal transduction

See paper by Sundaram and Han³. *ksr-1* appears to be a parallel signal transduction pathway to *lin-45* (Raf) and MAPK. Interestingly *ksr-1* mutants do not have obvious vulval phenotypes by themselves, but loss-of-function mutants suppress *let-60(gf)* phenotypes. Double mutants of *ksr-1* and MAPK are synthetically dead.

A Ras-mediated signal transduction pathway is involved in control of sex myoblast migration in *C. elegans*

See paper by Sundaram and Han⁴. *let-60(lf)* have posteriorly positioned sex myoblasts (SM) relative to wild-type. *let-60(gf)* mutants also have posteriorly positioned SMs but the phenotype is less severe. Similarly, *ksr-1* mutants have posterior SMs. Loss of *egl-15*, an FGF receptor, turns gonad from attractive to repulsive. The *ksr-1; egl-15* double has the same phenotype as the *egl-15* single.

$egl - 15; ksr - 1 = egl - 15 < ksr - 1 < WT$

let-60(gf) causes a weak (10%) SM migration phenotype (posterior). Unpublished: *let-60(gf)* coupled with *lin-45*, *ksr-1*, *mek-2* or *sur-1* dramatically enhances SM positioning defects. Surprisingly, increasing *let-60 (ras)* function by injection of wild-type or *let-60(gf)* genes into a *+/+* background does not cause positioning defects.

Increased *let-60 (ras)* activity partially suppresses *egl-15* or *egl-17* phenotypes. n1046gf allele can't suppress, but transgenes can. n1046gf also does not suppress *sem-5* migratory defects, whereas injected wild-type transgenes do suppress (but the GF allele does NOT suppress [they argue it does, but small numbers]).

SUR-5 Negatively Regulates *let-60 (ras)* activity during Vulval induction

See Gu and Han⁵. *sur-5* does not suppress all *dn* mutants. It does not suppress *sy93* and *sy100* (no association with Muv/nonMuv classes).

Take-home: *sur-5* genetically inhibits *let-60 (ras)* and the mutations suggest there may be multiple modes of interaction between *let-60 (ras)* and effectors.

SUR-8 Positively Regulates RAS-Mediated signaling in *C. elegans*

See Sieburth and Han⁶. *sur-8*, a conserved Ras-binding protein acts to promote *let-60 (ras)* (found via suppressor screen of *let-60(gf)*), although *sur-8* by itself has no phenotype. *sur-8* is synthetically lethal with other downstream *let-60 (ras)* effectors. *sur-8(ku167); ksr-1(ku68)* has a P6.p non-induction phenotype at a 10% rate, identical to *sur-1*.

The Ras-MAPK pathway is important for olfaction in *C. elegans*

See paper by Hirotsu and Iino⁷. Note that there is an important corrigendum to this paper. In theory, *let-60 (ras)* is shown to be important for AWA- and AWC-mediated attraction to odors. Notably, gf and lf mutations are said to affect odor detection in the same direction. However, the results are weak and not satisfactory to believe in (to my mind).

SOS-1 is necessary for multiple RAS signals

See paper by Chieh-Chang and Sternberg⁸. A very confusing paper, albeit with a lot of data. Need to read again. SOS-1 is ID'ed as the GEF for *let-60 (ras)* in many situations.

EGL-15 Signaling Pathway Implicates... in FGF Signal Transduction

See paper by Schutzman and Stern⁹. *let-60(gf)* suppresses a a Clr phenotype (Soc), similarly to *egl-15(gf)*. They found a gain of function mutation (G60R). This mutant is partially Muv and has a protruding excretory pore. As hets, these animals are Clr at 25°C, though normal at 15°C.

Strangely, the canonical gf mutation, *n1046*, does not show a Clr phenotype.

Claim is that EGL-15 acts through SEM-5 in one pathway or through SOC-1/PTP-2 in another to prevent the Clr phenotype. PTP-2 and SOC-1 may talk to RAS themselves.

EOR-1 and EOR-2 regulate Ras and Wnt redundantly with SUR-2(Mediator)

See paper by Howard and Sundaram¹⁰. In this paper, the claim is that *eor-1* and *eor-2* act downstream of Ras and Wnt to modulate signal from these things. They show a number of synthetic effects, namely rod-like lethality with Ras pathway components, though evidence for involvement in Ras signaling is weak when considering Ras pathway loss-of-function (almost no Vul). However, Ras gain-of-function mutations leading to Muv phenotypes are strongly attenuated by mutations in *eor-1* and *eor-2*. This is true even for *lin-1*, suggesting these genes act downstream of Ras.

Somewhat surprisingly, *eor-1* and *eor-2* are unable to suppress even partially the synMuv phenotype of *lin-15*, although they do inhibit the 0 P11.p phenotype of *lin-15*.

Evidence for interaction with Wnt is mainly based on suppression of a 2 P11.p phenotype of *pry-1*, but no other phenotype.

lin-1 is claimed to act both positively and negatively with the Ras pathway because although *lin-1(lf)* mutants are alive, double mutants with either *eor-1* or *eor-2* are completely dead; whereas maternally rescued doubles have a double Muv/Vul phenotype reminiscent of *lin-25*. Based on these two phenotypes, the claim is that *lin-1* acts bidirectionally on the Ras pathway. Another paper, by Tiensuu and Tuck, further addresses this issue.

sur-2 functions late in *let-60* signaling

See paper by Singh and Han. *sur-2* attenuates the Muv phenotype of the *let-60(gf)* mutant. Mutations in this gene lead to Vul and Mab phenotypes.

lin-1 has positive and negative function in Ras signaling

See paper by Tiensuu¹¹. In the absence of *eor-1*, *lin-1* mutants are dead unless maternally rescued for LIN-1 protein, in which case they show a Muv/Vul dual phenotype. Larval rod-like death is typical of *let-60 (ras)* loss-of-function, whereas the Muv phenotype is reminiscent of *let-60(gf)* mutants.

This paper goes on to claim, using *egl-17::gfp* expression as a proxy, that *lin-1* has positive and negative effects on the Ras pathway. However, all of their epistatic evidence is entirely consistent with an ‘additive’ model, in which *lin-1* represses *egl-17* and none of the other genes affect *egl-17* expression. Particularly worrisome is the lack of dynamic range (almost everything is 0 or 100%).

The most interesting aspect of this paper is that in *lin-1* mutants, the excretory duct cell is duplicated (as defined by expression of a *lin-48::gfp* reporter), whereas in a *eor-1* mutant the duct cell appears wild-type. In the double mutant, however, the duct cell appears to be missing, which also happens to be the phenotype of a *let-60(gf)* mutant.

In general, a very poorly written paper with an overabundance of weak data.

A gain-of-function allele of *cbp-1* (p300) increase Histone Acetyltransferase activity and antagonizes Ras

See Eastburn and Han¹². More Mediator-associated associations with Ras.

ISWI and NURF301 antagonize Rb-like pathways in multiple cell fates

See Andersen and Horvitz. *isw-1* suppresses a number of class B phenotypes, as well as the synMuv phenotype for a number of AB doubles.

Non-cell-autonomous role of Ras in neuroblast determination

See paper by Parry and Sundaram¹³. Paper shows that the G1 cell, which helps generate the excretory pore initially, but later becomes a neuroblast, requires *let-60* (*ras*) for its maturation non-cell autonomously.

KSR forms a Multiprotein Signaling Complex and Modulates MEK localization

See paper by Stewart, Sundaram and Guan¹⁴. Nice and thorough paper showing that KSR binds MEK in an enormous 700kDA complex (KSR and MEK1/2 ;50kDA each). *C. elegans ksr* alleles cannot bind MEK1. KSR did not appear to affect MEK activity or activation, but it does appear to localize MEK1 to the membrane.

KSR is a scaffold required for MAPK activation

See paper by Roy and Therrien. Fruitfly paper. Claim is that KSR is a scaffold that facilitates phosphorylation of MEK by RAF.

PP2A positively regulates Ras-signaling

See paper by Sierburth and Han. They find that PP2A acts positively to promote Ras signaling in a common pathway with *ksr-1*, probably upstream of *raf*.

Wnt signaling bypasses Ras in vulval induction

See paper by Gleason and Eisenman. Take-home message is that *lin-39*, a Hox gene, is controlled positively by both Wnt and Ras. When Wnt is overloaded, *lin-39* activity becomes independent of Ras, as assayed by a *pry-1; let-60(n1531dn)* double mutant. At 15°C, the effect is particularly pronounced.

ksr-1 and *ksr-2* have unique and redundant functions and are required for MPK-1 ERK phosphorylation

See beautiful paper by Ohmachi and Sundaram¹⁵. This paper explains that *ksr-1* and *ksr-2* are part of a scaffold that promotes the phosphorylation of MPK-1. Whereas *ksr-1* is required for SM and dispensable for almost everything else, *ksr-2* is required for Ras signaling in the germline meiotic progression. Moreover, knocking both of these genes out results in a significant loss of activated MPK-1 and phenotype strongly reminiscent of *let-60(lf)* mutants, as assayed by vulval induction, SM migration defects and fertility.

Interestingly, KSR1/2 were both identified as synthetic interactors with *let-60(gf)* and said to be in a 'redundant' pathway. However, some of these papers suggest that they were not so much redundant pathways as concentration hubs for multiple Ras-related pathways. This is a fantastic paper.

PTEN Negatively Regulates MAPK Signaling during *C. elegans* Vulval Development

Nakdimon and Durbin¹⁶. *daf-2* promotes Ras, *daf-18* inhibits. *daf-2* allegedly talks to *sem-5* and *age-1*. AGE-1 promotes PIP₃ formation, which in turn promotes Ras signaling at an unknown location. Meanwhile, DAF-18 promotes PIP₃ degradation, thus inhibiting the Ras pathway. However, DAF-18 also appears to act in a PIP₃-independent pathway to inhibit MAPK (MPK-1) activity.

PUF-8 negatively regulates RAS/MAPK signalling to promote differentiation of *C. elegans* germ cells

Vaid and Subramaniam¹⁷. GLP-1 (Notch) promotes mitotic proliferation by blocking exit into meiosis. PUF-8 and GAP-3 (a Ras GAP protein) appear to function redundantly to block LET-60 in the mitotic and meiotic entry regions.

Phosphorylated MPK-1 is present in the transition to proximal parts of the gonad and repressed distally. However, in *puf-8;gap-3* double mutants, MPK-1 is phosphorylated throughout and leads to a constitutively mitotic germline. The authors show that RNAi knockdown of *let-60* restores meiosis to the germline. This would appear to show that *let-60* inhibits germline maturation.

In general, this paper is best understood with a view of *let-60* as a proliferative factor. It promotes mitosis by blocking entry into meiosis in the distal germline, for which reason it is repressed there. Overabundance of LET-60 thus leads to tumor formation.

References

1. Han, M. & Sternberg, P. W. Analysis of dominant-negative mutations of the *Caenorhabditis elegans* *let-60* ras gene. *Genes and Development* **5**, 2188–2198 (1991). URL <http://www.ncbi.nlm.nih.gov/pubmed/1748278><http://www.genesdev.org/cgi/doi/10.1101/gad.5.12a.2188>.
2. Wu, Y. & Han, M. Suppression of activated *let-60* ras protein defines a role of *Caenorhabditis elegans* *sur-1* MAP kinase in vulval differentiation. *Genes and Development* **8**, 147–159 (1994).
3. Sundaram, M. & Han, M. The *C. elegans* *ksr-1* gene encodes a novel raf-related kinase involved in Ras-mediated signal transduction. *Cell* **83**, 889–901 (1995). URL <http://linkinghub.elsevier.com/retrieve/pii/S0092867495902058>.
4. Sundaram, M., Yochem, J. & Han, M. A Ras-mediated signal transduction pathway is involved in the control of sex myoblast migration in *Caenorhabditis elegans*. *Development* **122** (1996). URL <http://dev.biologists.org/content/122/9/2823.long>.
5. Gu, T., Orita, S. & Han, M. *Caenorhabditis elegans* SUR-5, a novel but conserved protein, negatively regulates LET-60 Ras activity during vulval induction. *Molecular and cellular biology* **18**, 4556–64 (1998). URL <http://www.ncbi.nlm.nih.gov/pubmed/9671465><http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC109041>.
6. Sieburth, D. S., Sun, Q. & Han, M. SUR-8, a Conserved Ras-Binding Protein with Leucine-Rich Repeats, Positively Regulates Ras-Mediated Signaling in *C. elegans*. *Cell* **94**, 119–130 (1998). URL <http://linkinghub.elsevier.com/retrieve/pii/S0092867400812271>.
7. Iino, Y., Hirotsu, T., Saeki, S. & Yamamoto, M. The Ras-MAPK pathway is important for olfaction in *Caenorhabditis elegans*. *Nature* **404**, 289–293 (2000). URL <http://www.nature.com/doi/10.1038/35005101>.
8. Chang, C., Hopper, N. a. & Sternberg, P. W. *Caenorhabditis elegans* SOS-1 is necessary for multiple RAS-mediated developmental signals. *The EMBO journal* **19**, 3283–94

-
- (2000). URL <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=313952&tool=pmcentrez&rendertype=abstract>. 177
178
9. Schutzman, J. L. *et al.* The Caenorhabditis elegans EGL-15 signaling pathway implicates a DOS-like multisubstrate adaptor protein in fibroblast growth factor signal transduction. *Molecular and cellular biology* **21**, 8104–16 (2001). URL <http://www.ncbi.nlm.nih.gov/pubmed/11689700><http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC99976>. 179
180
181
182
10. Howard, R. M. & Sundaram, M. V. C. elegans EOR-1/PLZF and EOR-2 positively regulate Ras and Wnt signaling and function redundantly with LIN-25 and the SUR-2 Mediator component. *Genes & development* **16**, 1815–27 (2002). URL <http://www.ncbi.nlm.nih.gov/pubmed/12130541><http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC186391>. 183
184
185
186
11. Tiensuu, T., Larsen, M. K., Vernersson, E. & Tuck, S. lin-1 has both positive and negative functions in specifying multiple cell fates induced by Ras/MAP kinase signaling in C. elegans. *Developmental Biology* **286**, 338–351 (2005). URL <http://www.sciencedirect.com/science/article/pii/S0012160605005348>. 187
188
189
190
12. Eastburn, D. J. & Han, M. A gain-of-function allele of cbp-1, the Caenorhabditis elegans ortholog of the mammalian CBP/p300 gene, causes an increase in histone acetyltransferase activity and antagonism of activated Ras. *Molecular and cellular biology* **25**, 9427–34 (2005). URL <http://www.ncbi.nlm.nih.gov/pubmed/16227593><http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC1265831>. 191
192
193
194
195
13. Parry, J. M. & Sundaram, M. V. A non-cell-autonomous role for Ras signaling in C. elegans neuroblast delamination. *Development* **141** (2014). URL <http://dev.biologists.org/content/141/22/4279.long>. 196
197
198
14. Stewart, S. *et al.* Kinase suppressor of Ras forms a multiprotein signaling complex and modulates MEK localization. *Molecular and cellular biology* **19**, 5523–34 (1999). URL <http://www.ncbi.nlm.nih.gov/pubmed/10409742><http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC84397>. 199
200
201
202
15. Ohmachi, M. *et al.* C. elegans ksr-1 and ksr-2 Have Both Unique and Redundant Functions and Are Required for MPK-1 ERK Phosphorylation. *Current Biology* **12**, 427–433 (2002). URL <http://linkinghub.elsevier.com/retrieve/pii/S0960982202006905>. 203
204
205
16. Nakdimon, I., Walser, M., Fröhli, E., Hajnal, A. & Durbin, R. PTEN Negatively Regulates MAPK Signaling during Caenorhabditis elegans Vulval Development. *PLoS Genetics* **8**, e1002881 (2012). URL <http://dx.plos.org/10.1371/journal.pgen.1002881>. 206
207
208
17. Vaid, S., Ariz, M., Chaturbedi, A., Kumar, G. A. & Subramaniam, K. PUF-8 negatively regulates RAS/MAPK signalling to promote differentiation of C. elegans germ cells. *Development* **140** (2013). URL <http://dev.biologists.org/content/140/8/1645.long>. 209
210
211
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