

3 Enrichment Analysis of Hypoxia Pathway Data

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In this notebook, we will isolate the hypoxia response (defined as the set of genes that fulfill the genetic equalities $egl-9 = egl-9;vhl-1$ and $hif-1 = egl-9 hif-1$), and we will perform enrichment analysis on the hypoxia response. We will also perform enrichment analyses on each mutant transcriptomes, to try to understand how different each transcriptome actually is.

```
In [1]: # important stuff:
import os
import pandas as pd
import numpy as np

# TEA and morgan
import tissue_enrichment_analysis as tea
import morgan as morgan
import gvars
import epistasis as epi

# Graphics
import matplotlib as mpl
import matplotlib.pyplot as plt
import seaborn as sns
from matplotlib import rc
rc('text', usetex=True)
rc('text.latex', preamble=r'\usepackage{cmbright}')
rc('font', **{'family': 'sans-serif',
              'sans-serif': ['Helvetica']})

# Magic function to make matplotlib inline;
%matplotlib inline

# This enables SVG graphics inline.
%config InlineBackend.figure_formats = {'png', 'retina'}

# JB's favorite Seaborn settings for notebooks
```

```
rc = {'lines.linewidth': 2,
      'axes.labelsize': 18,
      'axes.titlesize': 18,
      'axes.facecolor': 'DFDFE5'}
sns.set_context('notebook', rc=rc)
sns.set_style("dark")

mpl.rcParams['xtick.labelsize'] = 16
mpl.rcParams['ytick.labelsize'] = 16
mpl.rcParams['legend.fontsize'] = 14
```

```
In [2]: q = 0.1
        # this loads all the labels we need
        genvar = gvars.genvars()

        tissue_df = tea.fetch_dictionary()
        phenotype_df = tea.fetch_dictionary('phenotype')
        go_df = tea.fetch_dictionary('go')
        respiratory_complexes = pd.read_excel('../input/respiratory_complexes.xlsx')

In [3]: tidy = pd.read_csv('../output/temp_files/DE_genes.csv')
        tidy.sort_values('target_id', inplace=True)
        tidy.dropna(subset=['ens_gene'], inplace=True)
```

2 Defining the hypoxia response

The hypoxia response can be defined in genetic terms as those genes that obey the two epistasis relationships, $egl-9 = egl-9;vhl-1$ and $hif-1 = egl-9 hif-1$.

```
In [4]: def test_equality(equal_genotypes, third_genotype, df, col='code', q=0.1, n_std=2):
        """
        A function to test epistasis equality.

        For a set of genotypes, `a`, `b`, and `ab`, suppose that we want to find those genes
        that obey the rule `a`=`ab`. To identify genes with this expression pattern, we first
        calculate the epistasis coefficient for transcripts within the STP(`a`, `ab`). Then
        we find those transcripts that are <2sigma deviations away from the line of best fit.

        Params:
        equal_genotypes: the two genotypes that we want to set equal to each other
        third_genotype: the third genotype to be considered (needed to calculate epistasis
        coeff.).
        df - dataframe to use. Must contain `target_id` column
        col - column that encodes the genotypes
        q - q-value to be used
        n_std - number of standard deviations to use as cutoff

        Output:
```

```

A list of target_ids
"""
a, ac = equal_genotypes
c = third_genotype

# make sure the dataframe only contains the desired genotypes
all_genotypes = [a, ac, c]

df = df[df[col].isin(all_genotypes)]

overlap = epi.find_overlap(equal_genotypes, df, col=col, q=q)

df = df[df.target_id.isin(overlap)]
a_df = df[df[col] == a].copy()
c_df = df[df[col] == c]
ac_df = df[df[col] == ac]

# # the code below works only if the variance is invariant to the expected values
normed_deltas = (ac_df.b.values - a_df.b.values)
normed_deltas = normed_deltas/np.std(normed_deltas)

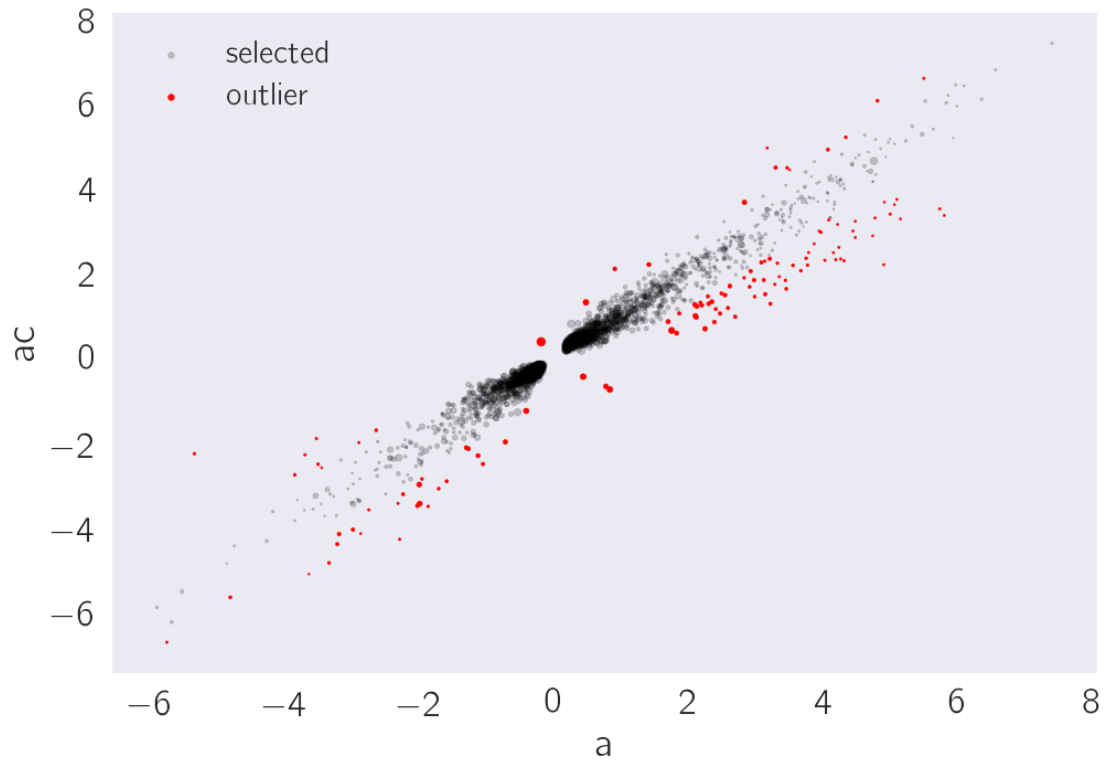
# first condition guarantees we're not too far from the line y=x
# second condition guarantees we are not on the line y=-x
inside = (np.abs(normed_deltas) < n_std) & (ac_df.b.values*a_df.b.values > 0)

# print a diagnostic plot:
plt.scatter(a_df[inside].b, ac_df[inside].b, s=1/ac_df[inside].se_b, color='black')
plt.scatter(a_df[~inside].b, ac_df[~inside].b, s=1/ac_df[~inside].se_b, color='red')
plt.xlabel('a')
plt.ylabel('ac')
plt.legend()

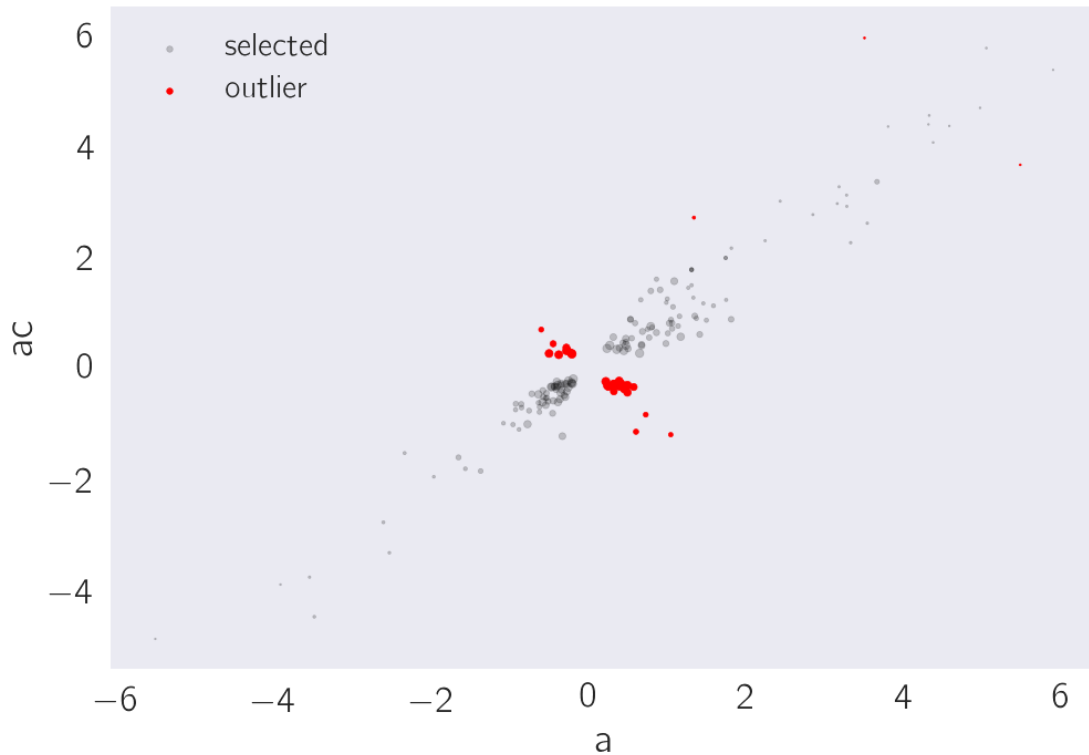
# return list of target ids that meet criteria
return a_df[inside].target_id.values

In [5]: # find the genes that obey egl-9 = egl-9;vhl-1
filtered_egl = test_equality(['b', 'a'], 'd', tidy, n_std=2)

```



```
In [6]: # find the genes that obey  $hif-1 = egl-9 hif-1$   
filtered_hif = test_equality(['c', 'f'], 'b', tidy, q=.1, n_std=2)
```



```
In [7]: # find those genes that are not DE in either hif-1 or egl-9 hif-1
not_DE_hif = (tidy.code.isin(['c', 'f'])) & (tidy.qval > q)
# a neat trick:
not_DE = epi.find_overlap(['c', 'f'], tidy[not_DE_hif], q=1)

In [8]: # genes that are DE in hif-1 and egl-9, and obey both equations:
equal_and_DE = (tidy.target_id.isin(filtered_egl)) & (tidy.target_id.isin(filtered_hif)
# genes that are DE in egl-9, but not in hif-1 genotypes and also obey both equations:
equal_no_hif = (tidy.target_id.isin(filtered_egl)) & (tidy.target_id.isin(not_DE))

# get the lists of both, then concatenate them for a hypoxia response:
# most of the genes will come from the equal_no_hif condition
de_both = tidy[(equal_and_DE)].target_id.unique()
de_one = tidy[(equal_no_hif)].target_id.unique()
overlap = list(set(np.append(de_both, de_one)))

# find the hypoxia response
hyp_response = tidy[tidy.target_id.isin(overlap)].copy()

In [9]: # annotate whether they are candidates for direct or
# indirect regulation.
def annotate(x):
    if x > 0:
```

```

        return 'candidate for direct regulation'
    else:
        return 'candidate for indirect regulation'

# annotate
hyp_response['regulation'] = hyp_response.b.apply(annotate)

In [10]: # save to file
cols = ['genotype', 'target_id', 'ens_gene', 'ext_gene', 'b', 'se_b', 'qval', 'regulation']
hyp_response[cols].to_csv('../output/temp_files/hypoxia_response.csv', index=False)

print('There are {0} genes in the predicted hypoxia response'.format(len(hyp_response)))

```

There are 1258 genes in the predicted hypoxia response

3 Enrichment Analysis of the Global HIF-1 response

Now that we have found the hypoxia response, we can perform tissue, phenotype and gene ontology enrichment analysis on this gene battery. Note that we don't show all possibilities. When a particular analysis is not present, it is because the enrichment results were empty.

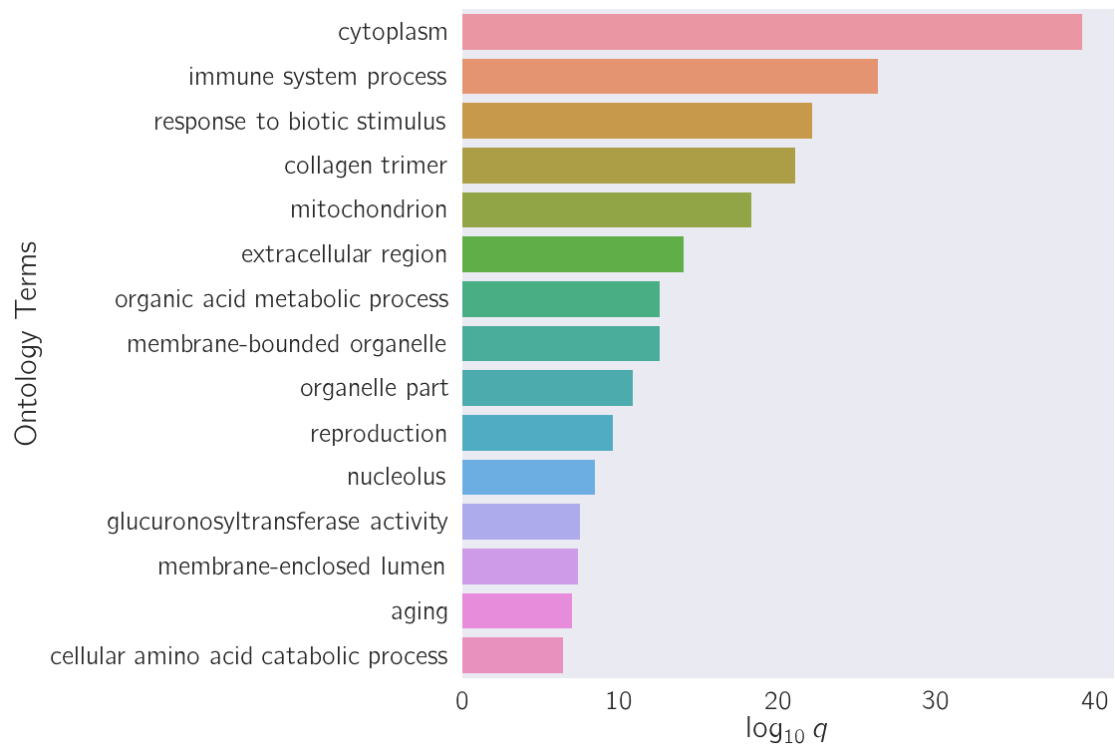
```

In [11]: teaH = tea.enrichment_analysis(hyp_response.ens_gene.unique(), tissue_df, show=False)
        peaH = tea.enrichment_analysis(hyp_response.ens_gene.unique(), phenotype_df, show=False)
        geaH = tea.enrichment_analysis(hyp_response.ens_gene.unique(), go_df, show=False)

        for df in [teaH, peaH, geaH]:
            df['logq'] = -df['Q value'].apply(np.log10)

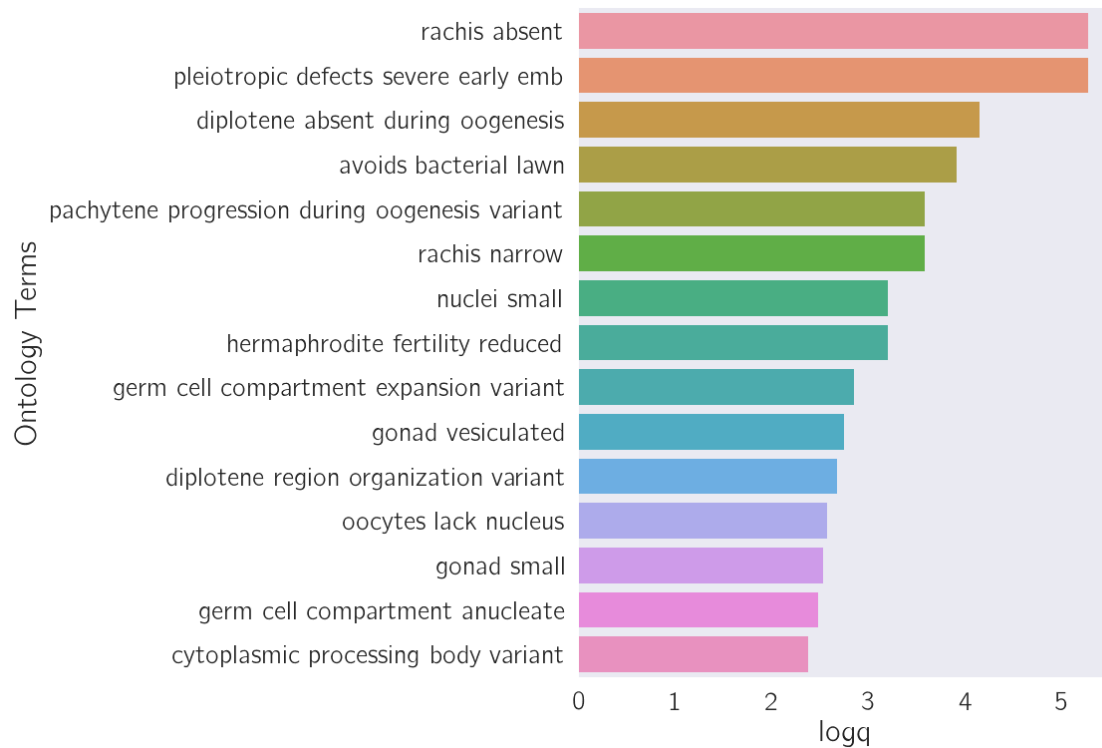
In [12]: ax = tea.plot_enrichment_results(geaH, analysis='go', y='logq')
        plt.xlabel('$-\log_{10}\{q\}$')
        plt.savefig('../output/supp_figures/supplementary_figure_3.pdf')

```

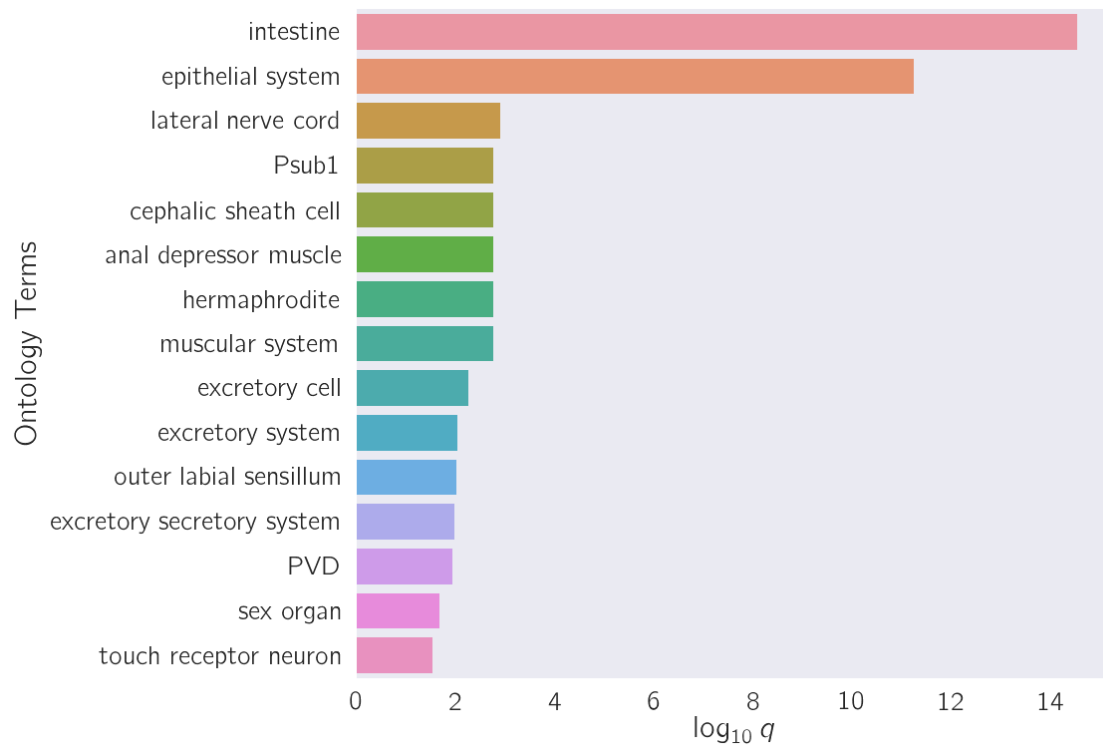


```
In [13]: tea.plot_enrichment_results(peaH, analysis='phenotype', y='logq')
```

```
Out[13]: <matplotlib.axes._subplots.AxesSubplot at 0x112ce8898>
```



```
In [15]: ax = tea.plot_enrichment_results(teaH, analysis='tissue', y='logq')
plt.xlabel('$\log_{10}\{q\}$')
plt.savefig('../output/supp_figures/supplementary_figure_4.pdf', bbox_inches='tight')
```

In []: