

Package ‘scTSSR’

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Type Package

Title Accurate gene expression recovery for single-cell RNA sequencing via bilinear regression

Version 1.1

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Description An implementation of a regularized bilinear regression prediction and empirical Bayes method to recover the true gene expression profile in noisy and sparse single-cell RNA-seq data.

License GPL-2

Encoding UTF-8

LazyData true

RoxygenNote 6.1.1

Depends R (>= 3.1)

Imports SAVER, keras, tensorflow

Suggests knitr, rmarkdown

VignetteBuilder knitr

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baron	<i>Human pancreatic islet data</i>
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Description

This is the Human pancreatic islet dataset (GSM2230757). The raw data contains 20,125 genes and 1,937 cells. Here we use the reference and downsampled datasets generated by Huang et al (2018) which contain 2,284 genes and 1,076 cells (available at <https://github.com/mohuangx/SAVER-paper/tree/master/SAVER-data>). For details about the approach to generate the reference and downsampled datasets, please refer to Huang et al (2018). This data is an object of class list of length two. count.ref is the reference count matrix and count.samp is the downsampled count matrix.

Usage

```
baron
```

Format

An object of class `list` of length 2.

Author(s)

Ke Jin, <kej13@mails.ccnu.edu.cn>

References

Baron, Maayan, et al (2016). A single-cell transcriptomic map of the human and mouse pancreas reveals inter-and intra-cell population structure. *Cell systems*, 3(4):346-360.

Huang, M. et al. (2018). Saver: gene expression recovery for single-cell rna sequencing. *Nat Methods*, 15, 539–542.

Zhang, X. F. et al. (2019) EnImpute: imputing dropout events in single cell RNA sequencing data via ensemble learning.

Examples

```
data("baron")
```

scTSSR

use scTSSR to impute dropout values in scRNA-seq data

Description

use scTSSR to impute dropout values in scRNA-seq data

Usage

```
scTSSR(X_count, lambda1 = NULL, lambda2 = 1e+10, initA = NULL,
       initB = NULL, percent = 0.05, ncores = 1, MAX_ITER = 4,
       ABSTOL = 0.001, learning_rate = 1e-04, epochs = 100,
       batch_size = 128, run_batch = TRUE, verbose = TRUE,
       estimates.only = FALSE)
```

Arguments

X_count	An expression count matrix. The rows correspond to genes and the columns correspond to cells. Can be sparse.
lambda1	Tuning parameter to facilitate feature selection and regularization.
lambda2	Tuning parameter to penalize the diagonal elements of the parameter to eliminate the trivial solution of representing an expression level as a linear combination of itself.
initA	The initialization of A. The elements of A represent the similarities between genes.

<code>initB</code>	The initialization of B. The elements of B represent the similarities between cells.
<code>percent</code>	The expression count matrix is preprocessed by filtering out the genes expressed in at most <code>percent*100%</code> of the cells.
<code>ncores</code>	Number of cores to use. Default is 1.
<code>MAX_ITER</code>	Maximum iteration of the external circulation of scTSSR.
<code>ABSTOL</code>	Absolute tolerance of the external circulation.
<code>learning_rate</code>	A hyper-parameter that controls the speed of adjusting the weights of the network with respect to the loss gradient.
<code>epochs</code>	The number of the entire training set going through the entire network.
<code>batch_size</code>	The number of examples that are fed to the algorithm at a time.
<code>run_batch</code>	Whether to use batch or to set the number of all the samples as the value of the batch size. Default is TRUE.
<code>verbose</code>	Whether to output the value of metrics at the end of each epoch. Default is TRUE.

Value

If `'estimates.only = TRUE'`, then a matrix of scTSSR estimates.

If `'estimates.only = FALSE'`, a list with the following components

<code>estimate</code>	Recovered (normalized) expression.
<code>se</code>	Standard error of estimates.
<code>info</code>	Information about dataset.

The `info` element is a list with the following components:

<code>size.factor</code>	Size factor used for normalization.
<code>pred.time</code>	Time taken to generate predictions.
<code>posterior.time</code>	Time taken to compute the posterior distribution.
<code>total.time</code>	Total time for scTSSR estimation.

Author(s)

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Examples

```
data("baron")

baron_imputation_result = scTSSR(baron$count.samp)
```

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*Topic **datasets**

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