


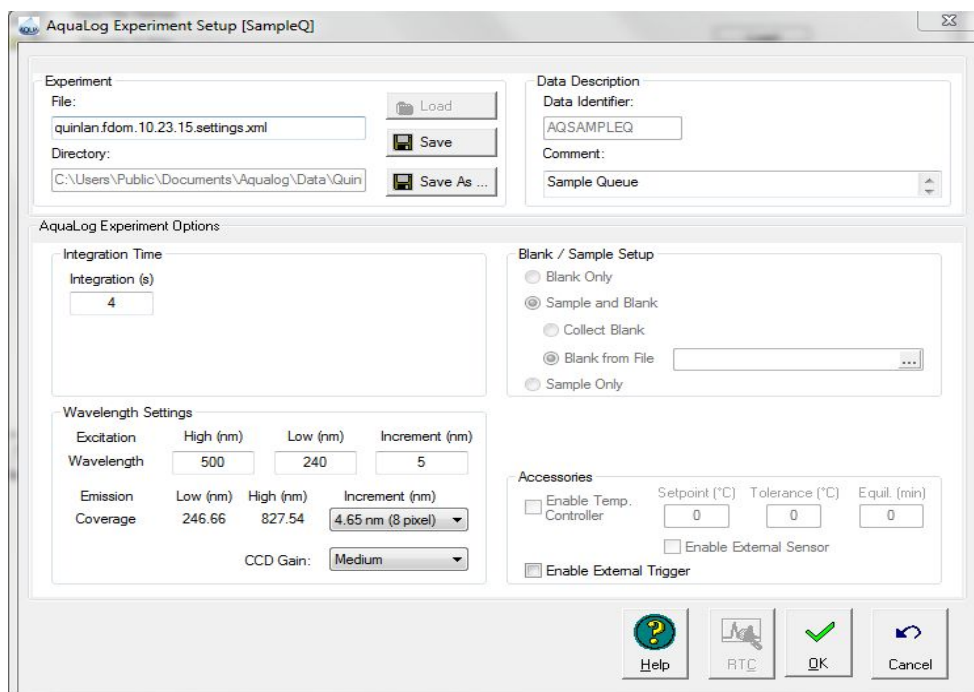
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Collection

- 1) Collect samples by filtering water through 0.2µm polyethersulfone filter into acid washed and combusted amber vials.
- 2) Store filtrate at 4°C
- 3) Analyze on aqualog spectrometer within a month

Aqualog setup

- 1) Take samples out of fridge and let warm to room temperature
- 2) Turn on the Aqualog spectrometer and let it warm up (0.5-2 hours)
- 3) Open Aqualog V3.6 software on desktop of Lenovo laptop
- 4) Make a new Q file by clicking on the Q in the Aqualog quickbar 
 - a) The Aqualog will make a series of loud noises. After the secondary screen opens. Close the entire application. (There has been some communication error where the Q needs to be activated twice otherwise the internal lazy susan won't rotate).
 - b) After re-opening and new file tool (Q) continue with set up
 - c) First create or load experiment file (settings)
 - i) New: press create and name the file
Yourname.Experiment.SampleQ.Settings.xml
 - ii) Change integration time 4-seconds
 - iii) Excitation wavelength - change to high=500 nm, Low=240 nm
 - iv) Increment - change to 5nm
 - v) Emission - select 8 pixel
 - vi) CCD gain - medium
 - vii) Blank and accessories is greyed out
 - viii) Save as - deposit in folder with your name
 - ix) Press okay



Aqualog Experiment Setup [SampleQ]

Experiment

File:

Directory:

Data Description

Data Identifier:

Comment:

Aqualog Experiment Options

Integration Time

Integration (s):

Wavelength Settings

| | High (nm) | Low (nm) | Increment (nm) |
|------------|-----------|-----------|-------------------|
| Excitation | 500 | 240 | 5 |
| Wavelength | | | |
| Emission | Low (nm) | High (nm) | Increment (nm) |
| Coverage | 246.66 | 827.54 | 4.65 nm (8 pixel) |

CCD Gain:

Blank / Sample Setup

☐ Blank Only

☒ Sample and Blank

☐ Collect Blank

☐ Blank from File

☐ Sample Only

Accessories

☐ Enable Temp. Controller

Setpoint (°C): Tolerance (°C): Equil. (min):

☐ Enable External Sensor

☐ Enable External Trigger

Help RTC OK Cancel

- d) Back at the sample Queue setup window
- e) Make a sample Q file in folder of choice (Your named folder)
 - i) Make sure output is into the same file
C:\Users\Public\Documents\Aqualog\Data\yourname\experiment
- f) Adjust sampling settings for sets and blanks
 - i) Divide your samples in sets (blank groups), usually 6-8 samples per set.
At the beginning of each set you will run another blank of Q-water
 - (1) Number of blank groups == number of sets
 - (2) Samples per blank == samples per set
 - ii) Blank group sample start # == 1
 - iii) Start Sample # == 1
- g) Do not select for any delays because it automatically runs samples in machine after delay resulting in possible reruns of samples
- h) Post processing options: uncheck all post processing options
 - i) All standardization will be done using the matlab script
- i) Export options: Uncheck "save raw data"
- j) Export ASCII: Check "save worksheets as ASCII"
 - i) Check ABS
 - ii) Check blank XYY
 - iii) Check sample XYY

Sample Queue Setup

Input File Names

Sample Q File: C:\Users\Public\Documents\Aqualog\Data\Quinlan\ModelCompounds\AqualogSampleQ.aqu [Load] [Save] [Save As]

Experiment File: C:\Users\Public\Documents\Aqualog\Data\Quinlan\fdom\quinlan.fdom.10.23.15.settings.xml [Browse...] [Open]

Comment: Sample Queue [Create]

Output File Names

Blank Group Prefix: Group Blank Base: Blank Sample Prefix: Sample Example Output: Group001Blank.ogw, Group001Sample0001.ogw, Group001Sample0001ABS.dat

Output Folder: C:\Users\Public\Documents\Aqualog\Data\Quinlan\ModelCompounds\ [Browse...]

Sample Setup

Number of Blank Groups: 6 Samples Per Blank: 6

Blank Group Start #: 1 Sample Start #: 1

Total Samples: 42

Delays

Before First Blank: 0 secs

Between Each Experiment: 0 secs

☐ Single Cell Kinetics

Post Processing Options

☐ IFE ☐ Rayleigh Masking

☐ 1st Order ☐ 2nd Order ☐ 1st Order & 2nd Order SUM of slit widths: 10

☐ Normalize

☐ Max ☐ Normalization Factor: 1

Export Options

☐ Save Raw Data in Workbook Format (*.ogw)

Export ASCII

☒ Save Worksheets as ASCII (*.dat)

☒ ABS ☒ Blank XYY ☒ Sample XYY

Sample - Blank Raw

☐ XYY ☐ XYZ

Sample - Blank Processed

☐ XYY ☐ XYZ

[Resume] [Run] [Cancel]

- 5) Press Run and the system will prompt you to insert your first two samples

Running the samples

- 1) Before proceeding with your run, make a sample list worksheet
 - a) Or fill in sample names on “matlab setup” worksheet (located in google drive)
- 2) Before pipetting samples into cuvettes, rinse the cuvettes three times and shake empty
 - a) Shake vigorously but not hard enough that you throw the cuvettes and shatter them
- 3) Fill quartz cuvettes with ~3ml of samples
 - a) First sample of set will be the first blank and then your first sample of set 1
 - b) Pop up will read
 - i) SC position 1: Blank 1 (Group001Blank)
 - ii) SC position 2: Sample 1 (Blank 1) [Group001Sample0001]
- 4) Wipe quartz cuvettes with Kimwipe to clean smudges and fingerprints
- 5) Press continue to run your samples once they are loaded and lid is shut
- 6) It takes around 8 minutes to run two samples. You can watch the progress of each samples on the “Intermediate Display” which pops up.
- 7) The sample holder will automatically advance to position 2 when position 1 has been run.
- 8) ~1minute before samples are done running pipette next samples into rinsed cuvettes and wipe down sides
 - a) Use the same pipette tip and Kimwipe(change both each blank)
- 9) Once the two samples have run, the system, will pause and a window will open listing the expected next samples for positions 1 and 2. Removed the old cuvettes and inset the new ones (note that the sample holder has shifted positions from the first two) and press continue. Keep in mind that if you are running an even number of samples per blank you will be placing the final sample of a set and the next blank in the same holder. This can be a tad confusing.
- 10) After each non-blank sample the “script window” will report that the files have been exported. Check the directory to verify they have been exported there in .dat format. Files ending in BEM are blanks, SEM are samples, and ABS are absorbance scans.

Matlab Script and runlog

- 1) On Google Drive in the Nelson Lab folder access the “fDOM basic Matlab input sheet DO NOT EDIT. COPY/PASTE ONLY”
 - a) Page one are step by step guide to editing
 - b) Page two is the actual matlab input spreadsheet
- 2) n= number of samples per blank group

| | |
|---------|---|
| Step 1. | Delete extra blank groups and samples |
| Step 2. | Delete the last (8-n) samples of each blank group. This should make it so there are n samples in between each "BLANK" |

| | |
|---------|---|
| Step 3. | In the the E column Copy and paste "C:\Users\Public\Documents\Aqualog\Data\yourfilepath\Group001Sample0001BEM.dat" into the bottom (1-n) |
| Step 4. | Put in your own sample names. These are the names that will be put into the final summary sheet. NO SPACES EVER! |
| Step 5. | "Find and replace" "yourfilepath" with your file location if it is within the .Aqualog\Data\ folder |
| Step 6. | Download as .tsv and name file something that starts with a letter (matlab is dumb). |
| Step 7. | Put name of .tsv into line 112 of .m script |
| | |

- 3) Download the latest version of the script. Found at
<https://github.com/Zquinlan/fDOMmatlab/edit/master/Script.md>



- 4) Edit the matlab script as need-be
- a) Lines 1-112 explain file
 - b) Line 113 is where you input the matlab input worksheet file name (E.G. "fDOM.tsv")
 - c) Lines 118-254 have to do with settings of ran samples
 - i) Should not have to be edited if above steps were taken

- d) Lines 255-288 are for specific peaks to acquire values and input into final spreadsheet
 - i) Edit only if you want to add new peaks (NOTE: if added need to adjust lines 335-367)
- e) Line 289-334 are for split half validation, and PARAFAC modeling
 - i) This is where you change how many PARAFAC components you would like to input. These have to be modified with each experiment because there should be a different number of components in each experiment.
- f) Lines 335-367 make the Summary Data Matrix
 - i) Edit for PARAFAC models and new peaks from above
- g) Remaining lines have to do with other spreadsheets which are created such as EEMain
 - i) These should not have to be edited