



POLITECNICO
MILANO 1863

Computational Neuroscience EBRAINS – NEST Desktop



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Laboratory:
Neurorobotics and Medical Robotics Lab

Simulating Single Neuron and Network Activity

Human Brain Project infrastructure: EBRAINS

Simulating software: NEST Desktop

- | | | |
|---|---------------------------------|----------|
| 1 | LIF model simulation | HANDS-ON |
| 2 | Hodgkin-Huxley model simulation | HANDS-ON |
| 3 | Network activity | HANDS-ON |



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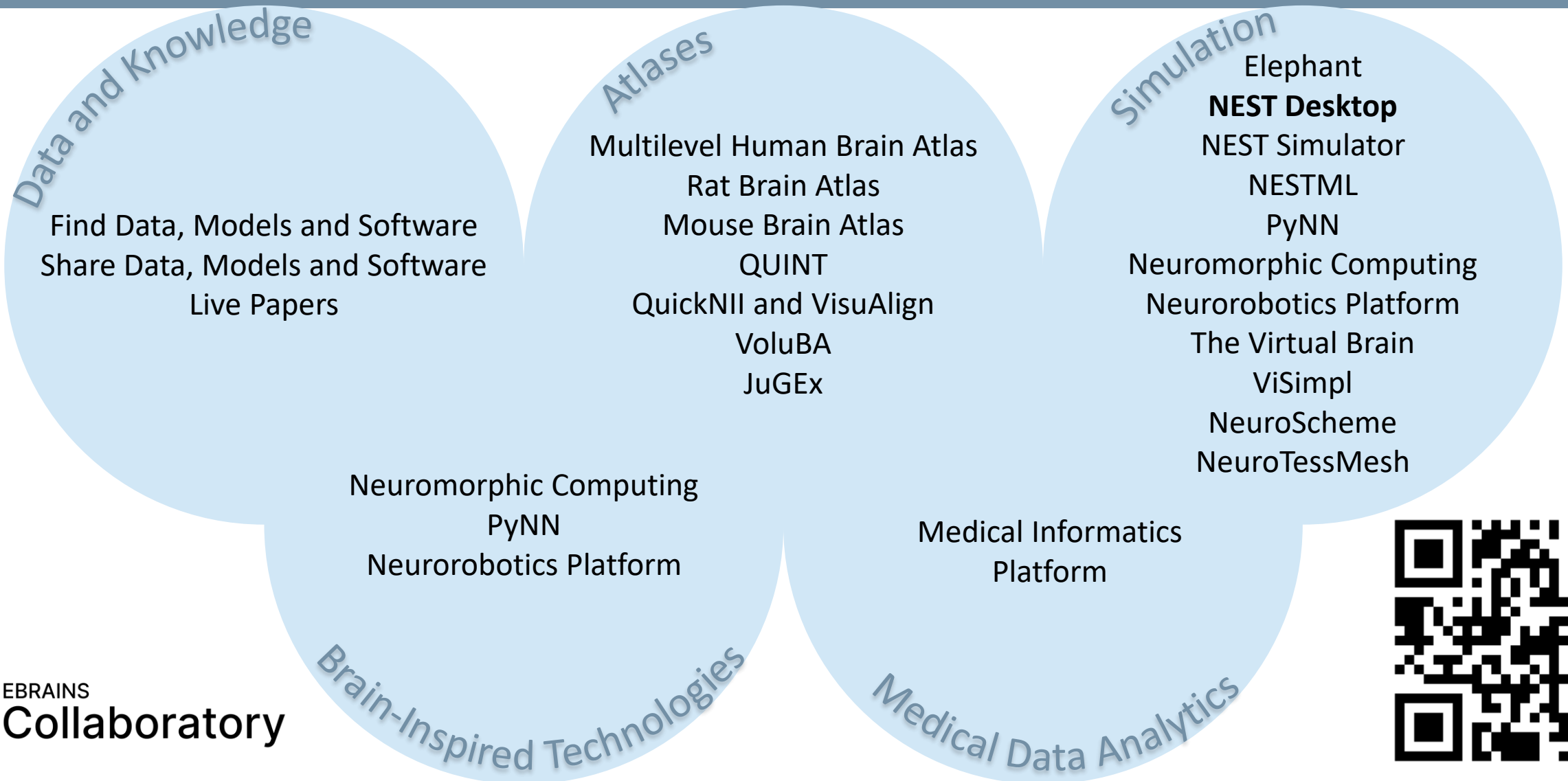


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Human Brain Project infrastructure: EBRAINS



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Code

```
import nest
import numpy

nest.ResetKernel()

# Simulation kernel
nest.SetKernelStatus({
    "local_num_threads": 1,
    "resolution": 1.0,
    "rng_seed": 1
})

# Create nodes
pg1 = nest.Create("poisson_generator", 1, params={
    "rate": 6500.0
})
n1 = nest.Create("iaf_psc_alpha", 100,
    positions=nest.spatial.free(
        nest.random.uniform(-0.5, 0.5),
        num_dimensions=2
    )
)
sr1 = nest.Create("spike_recorder")

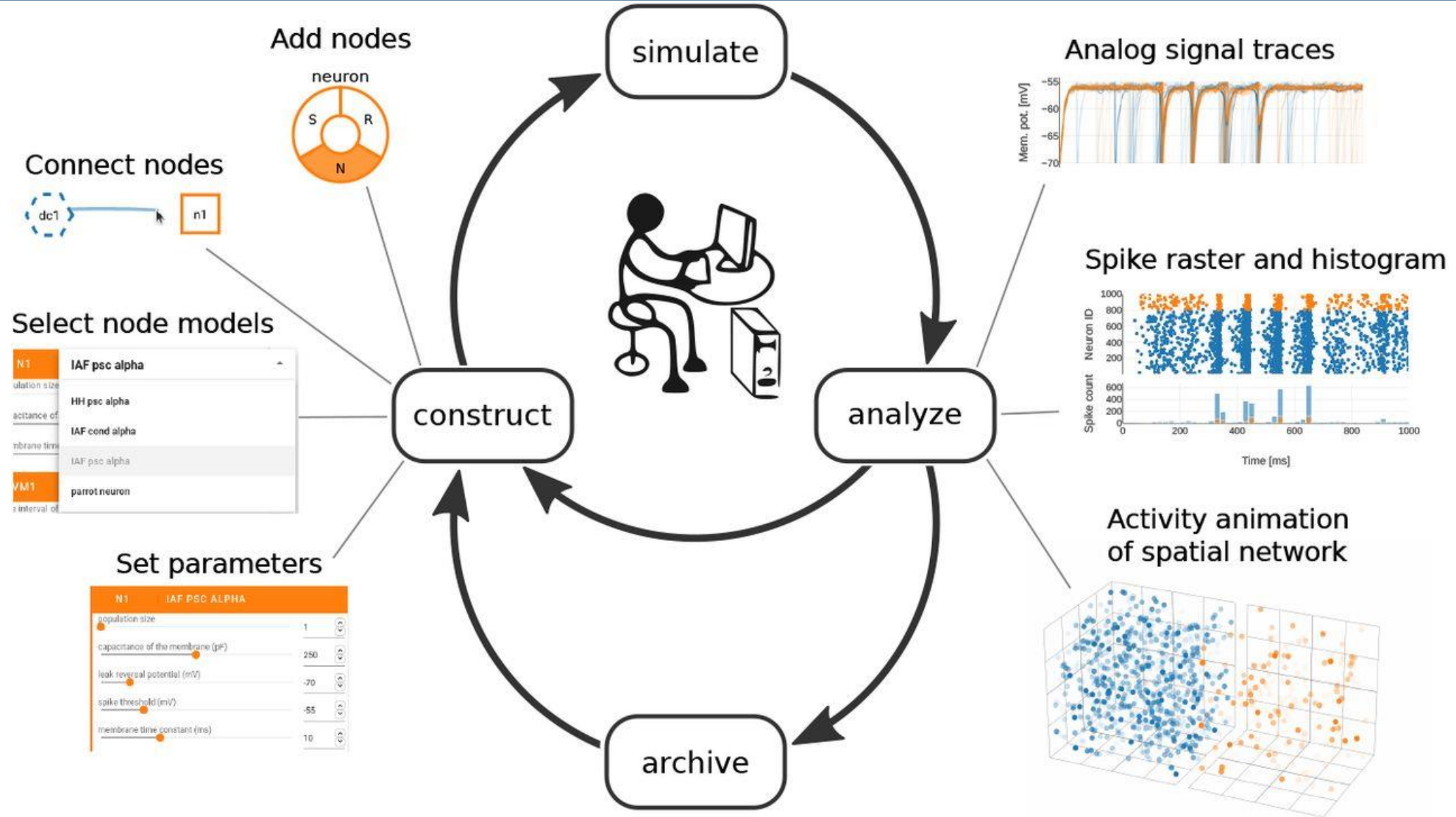
# Connect nodes
nest.Connect(pg1, n1, conn_spec={
    "rule": "all_to_all"
}, syn_spec={
    "weight": 10.0
})
nest.Connect(n1, sr1, conn_spec={
    "rule": "all_to_all"
})

# Run simulation
nest.Simulate(1000)

# Define function getting activity from the recorder
def getActivity(node):
    if node.get("model1") == "spike_recorder":
        nodeIds = nest.GetConnections(None, node).sources()
    else:
        nodeIds = nest.GetConnections(node).targets()
    return {
        "events": node.get("events"),
        "nodeIds": list(nodeIds)
    }

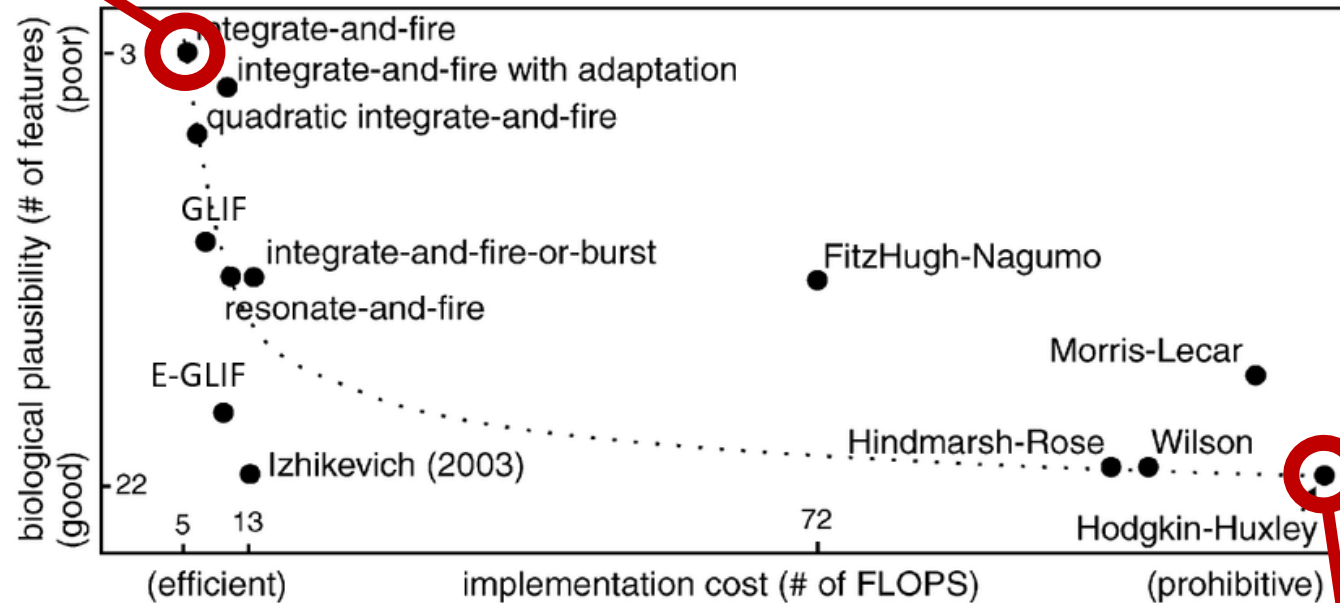
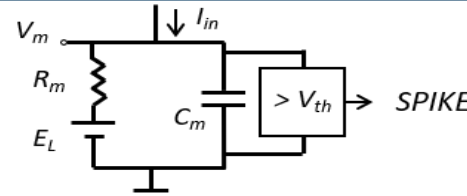
# Define function getting node positions
def getPosition(node):
    return list(zip(node.tolist(), nest.GetPosition(node)))

# Collect activities
response = {
    "kernel": {
        "biological_time": nest.GetKernelStatus("biological_time")
    },
    "activities": [
```

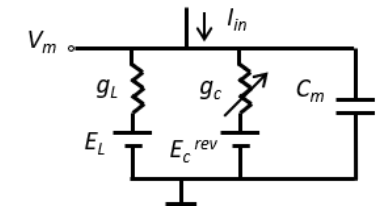


Simulating software: NEST Desktop

LIF: Only **passive** membrane properties are considered (capacitance C_m and resistance R_m). The output is a spike train, corresponding to time instants of threshold overcoming



HH: membrane potential V_m computed considering the resting potential (E_L) and the contribution of each membrane **ion channel** (represented by the conductance g_c and reversal potential E_c^{rev}).



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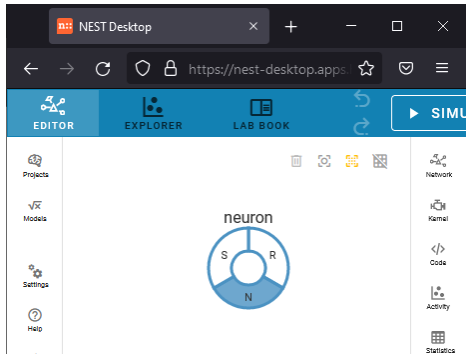
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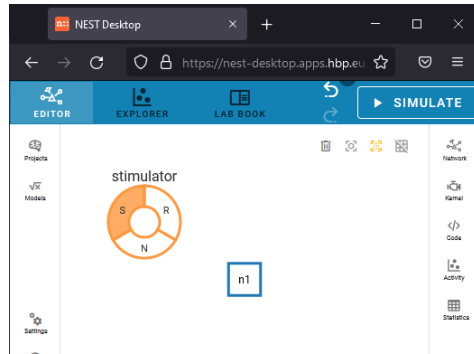
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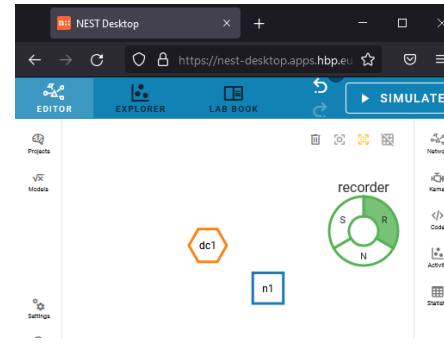
Right click → select “N”



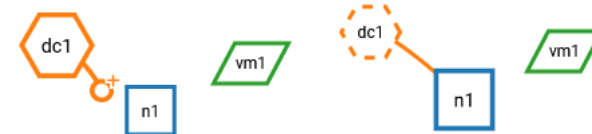
Right click → select “S”



Right click → select “R”



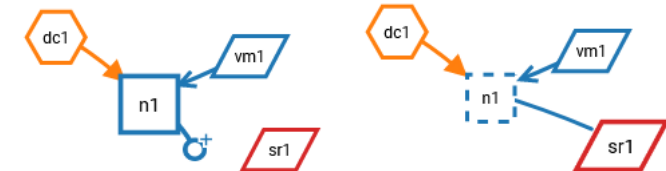
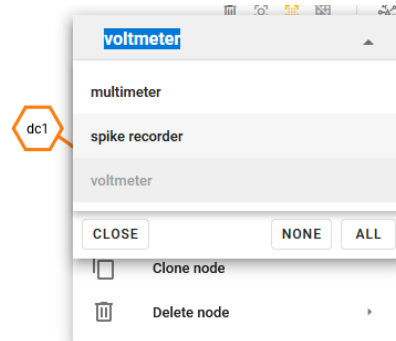
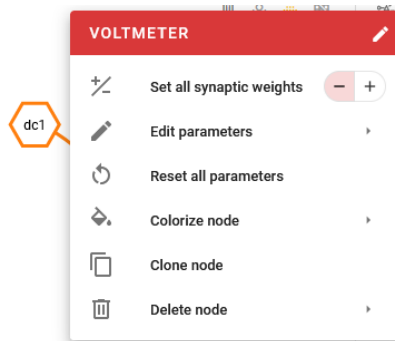
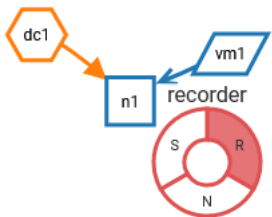
Connect the stimulator (**dc1** to **n1**)



Connect the voltmeter (**vm1** to **n1**)



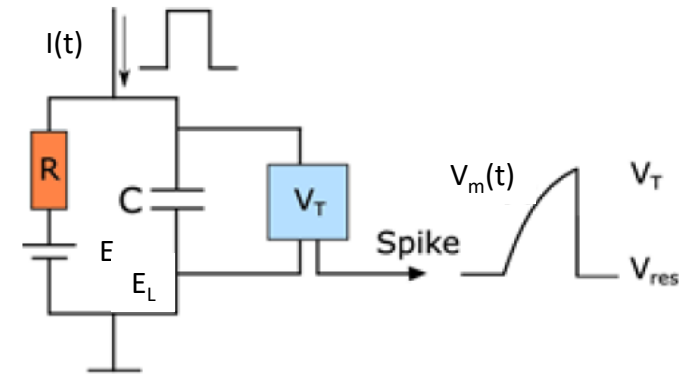
Add a spike recorder: change the default option “voltmeter” to “spike recorder”. Connect the spike recorder(**n1** to **sr1**)



Direct current injection into single neurons

1. What is the membrane potential response for both negative and positive values of the applied current? You can measure the membrane potential by performing an intracellular recording, using a voltmeter. Try different amplitudes of the applied current and describe the phenomena you observe.
2. Explore how the membrane potential response depends on the biophysical neuron parameters. In particular, describe the influence of the spike threshold, the time constant of the membrane and the absolute refractory time on the membrane potential trajectories

$$\tau_m \frac{dV_m(t)}{dt} = -[V_m(t) - E_L] + R_m I_{in}(t)$$



Poisson input into single neurons

1. Consider a LIF neuron that receives Poisson* input with a constant rate using a synapse of a specific amplitude. Analyze how the input rate influences the membrane potential and the spiking response of the neuron. The parameters of interest are the mean and the variance of the membrane potential. What happens if you change the strength of the synapse?

* Poisson input: a neuron that is firing with Poisson statistics, i.e. exponentially distributed interspike intervals. It will generate a *unique* spike train for each of its targets.

Noise current injection into single neurons

1. **EXTRA:** Play with the parameters of the noise current over a certain range of values when there is no action potential generated, and several spikes generated. What happens to the frequency of spikes and the irregularity of spike trains as mean and/or variance if the noise is increased? The irregularity of neuronal spiking can be assessed, for example, by the coefficient of variation (CV) of the interspike intervals.
2. **EXTRA:** Systematically measure two types of input-output curves of the neuron:
 - a) Keep the variance of the noise at a fixed level and systematically change the mean of the noise. What is the difference to the curve you obtained with pure DC input?
 - b) Now keep the mean of the noise at a fixed level and systematically change the variance of the noise. What is the minimal variance (“threshold”) that leads to a non-zero response rate?

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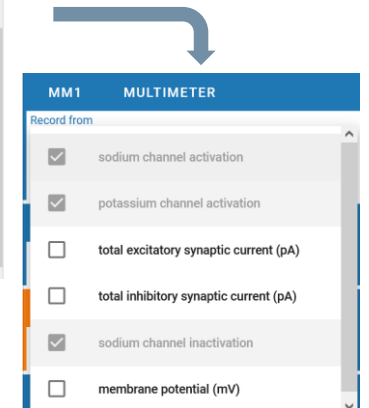
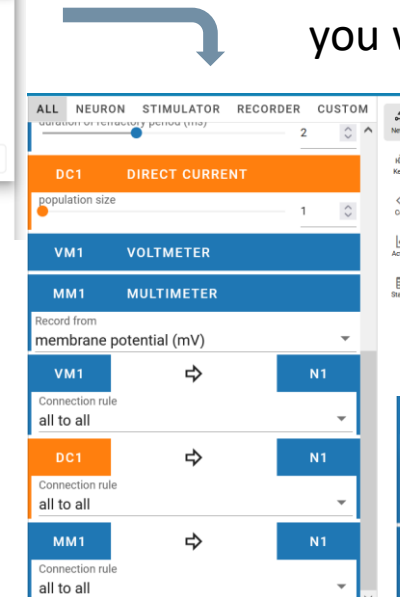
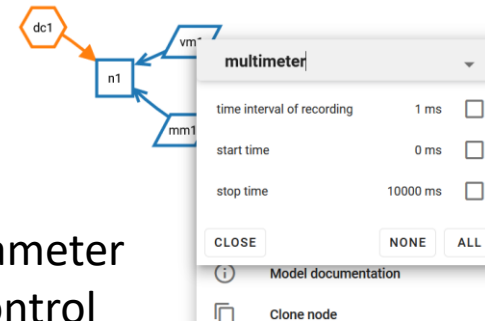
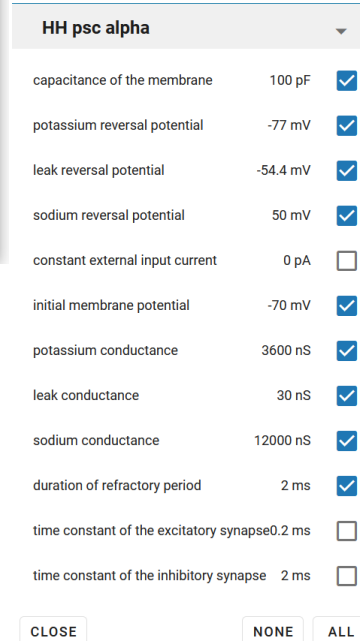
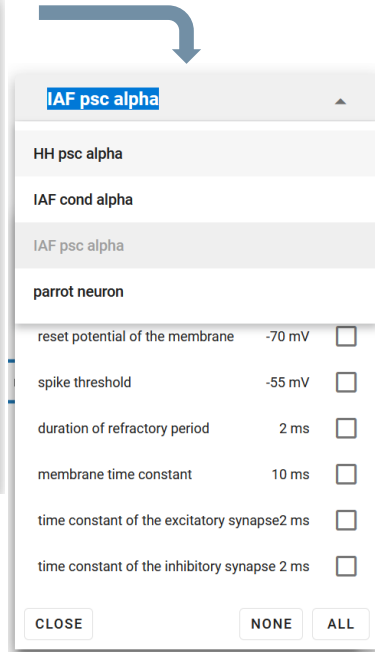
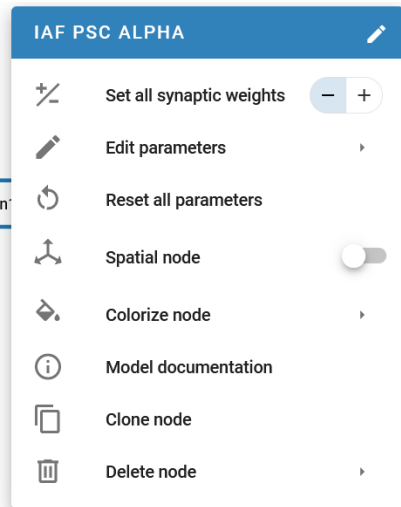
Change the neuron model to “HH model”.

Connect the multimeter recorder(**n1** to **mm1**)

Select the parameter you want to record

Select the parameter you want to control

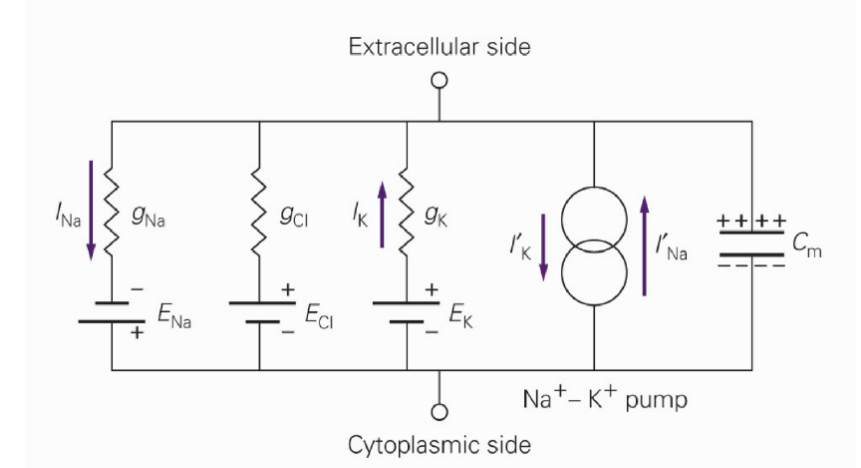
1st spike generator
2nd direct current



Sodium and potassium currents under current clamp conditions

1. Action potential generation based on voltage-dependent ion channels takes place in several stages. Use the membrane potential recording of a **single spike** to illustrate the dissection of this process.

$$C_m \frac{dV(t)}{dt} = g_{leak}[V(t) - V_{leak}] - g_{Na}(V, t)[V(t) - V_{Na}^{rev}] - g_K(V, t)[V(t) - V_K^{rev}] + I_{ext}$$

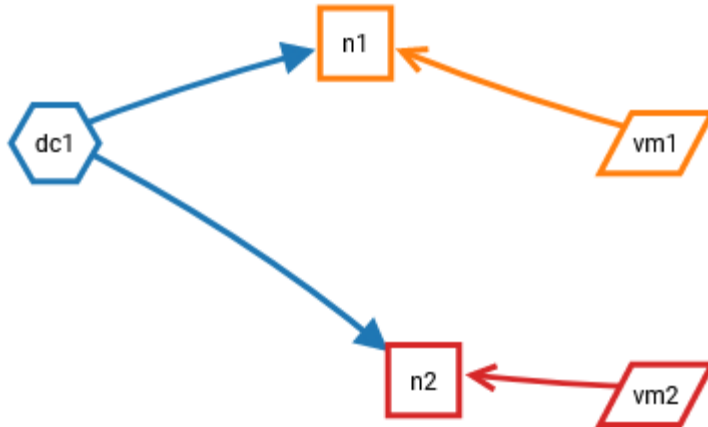


2. **EXTRA:** Explore the effect of the specific neurotoxins TTX and TEA on neuronal spiking. You can easily achieve this in your simulations by setting the peak conductances of either sodium channels or potassium channels, respectively, to zero. What happens if you only partially knock-out these channels by setting them to a non-zero, but reduced value? Formulate and explain your expectations before you perform the experiments.

Hodgkin-Huxley model simulation



► <https://nest-desktop.apps.hbp.eu/#/>



The screenshot displays the NEURON software interface for configuring a multi-compartment model. The interface is organized into several panels, each representing a different component or connection rule.

- N1 (orange):** Contains sliders for population size (1), capacitance of the membrane (250 pF), spike threshold (-30 mV), and membrane time constant (10 ms).
- IAF PSC ALPHA (orange):** Contains sliders for population size (1), potassium reversal potential (-77 mV), leak reversal potential (-68 mV), sodium reversal potential (50 mV), and initial membrane potential (-70 mV).
- DC1 (blue):** Contains sliders for population size (1), amplitude of current (2000 pA), start time (100 ms), and stop time (600 ms).
- DIRECT CURRENT (blue):** Contains connection rules for DC1 to N1 and N2, both set to "all to all".
- VM1 (orange):** Contains a slider for time interval of recording (0.1 ms).
- VOLTMETER (orange):** Contains a slider for time interval of recording (0.1 ms).
- N2 (red):** Contains sliders for population size (1), potassium reversal potential (-77 mV), leak reversal potential (-68 mV), sodium reversal potential (50 mV), and initial membrane potential (-70 mV).
- HH PSC ALPHA (red):** Contains sliders for population size (1), potassium reversal potential (-77 mV), leak reversal potential (-68 mV), sodium reversal potential (50 mV), and initial membrane potential (-70 mV).
- DC1 (blue):** Contains sliders for population size (1), amplitude of current (2000 pA), start time (100 ms), and stop time (600 ms).
- DIRECT CURRENT (blue):** Contains connection rules for DC1 to N1 and N2, both set to "all to all".
- VM2 (red):** Contains a slider for time interval of recording (0.1 ms).
- VOLTMETER (red):** Contains a slider for time interval of recording (0.1 ms).
- N1 (orange):** Contains a slider for time interval of recording (0.1 ms).
- N2 (red):** Contains a slider for time interval of recording (0.1 ms).
- SR1 (red):** Contains a slider for time interval of recording (0.1 ms).
- SR2 (orange):** Contains a slider for time interval of recording (0.1 ms).

The connection rules for the DC1 panel are:

- DC1 to N1: all to all
- DC1 to N2: all to all

The connection rules for the N1 and N2 panels are:

- N1 to SR1: all to all
- N2 to SR2: all to all

Current clamp simulation of the free-running membrane

1. Verify that the subthreshold properties of the HH neuron model are similar to the properties of the LIF neuron model. To address this issue, inject a **depolarizing or hyperpolarizing current** into a LIF neuron and into a HH neuron and perform intracellular recordings to document the membrane response in both cases. Make sure the current is weak enough to not elicit a spike.
2. For strong-enough DC input current the HH model neuron will fire a train of action potentials. Inspect the spike waveform carefully and relate it to the spikes generated in a LIF neuron. Is the spike waveform of the HH neuron really the same for different input scenarios (e.g. weak vs. strong current)?
3. **EXTRA:** Use a spike recorder to characterize the spiking response to superthreshold current input. The goal is again to characterize the neuron by a curve that depicts the firing rate response as a function of the applied current.

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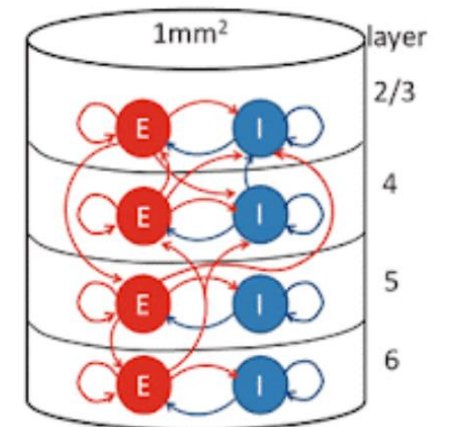
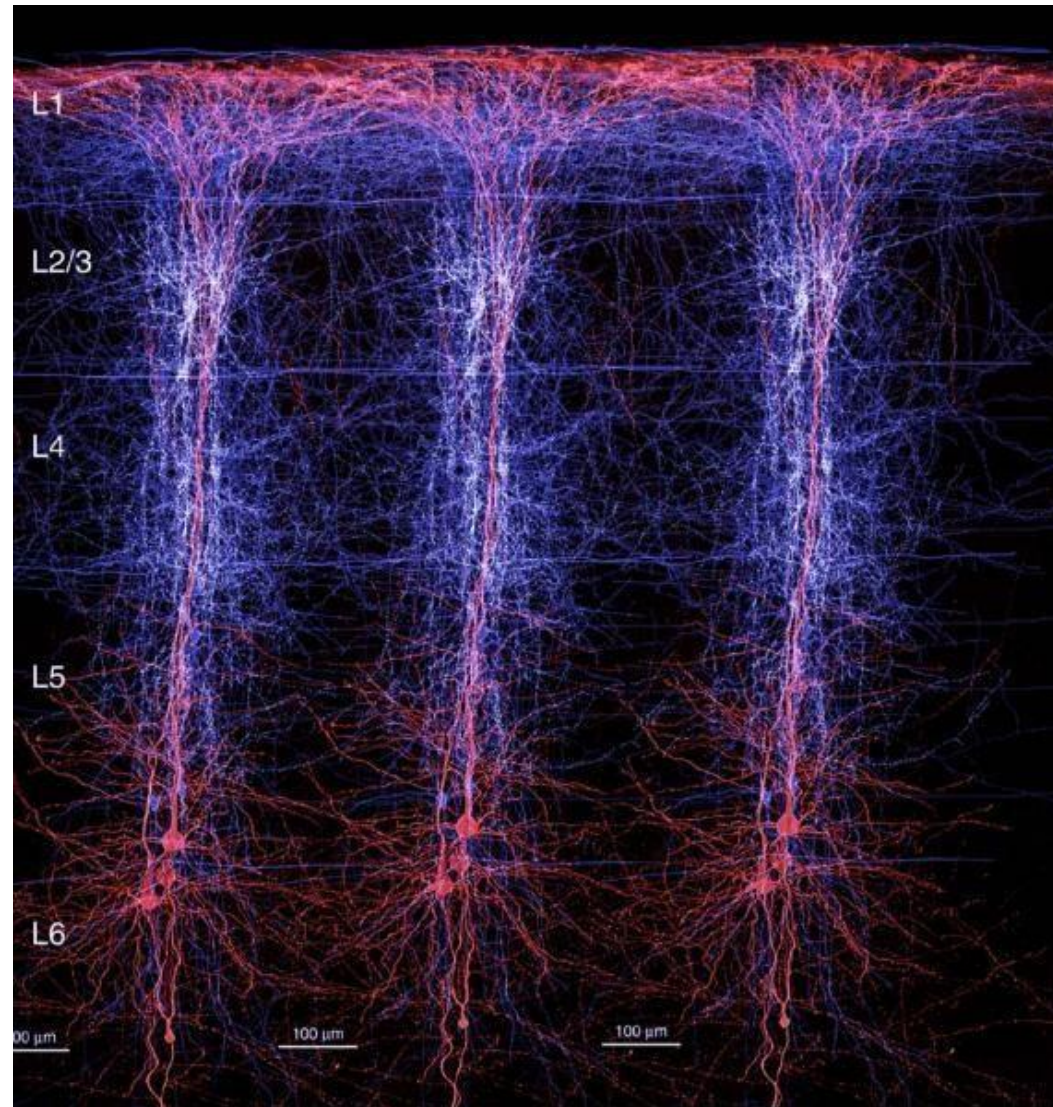


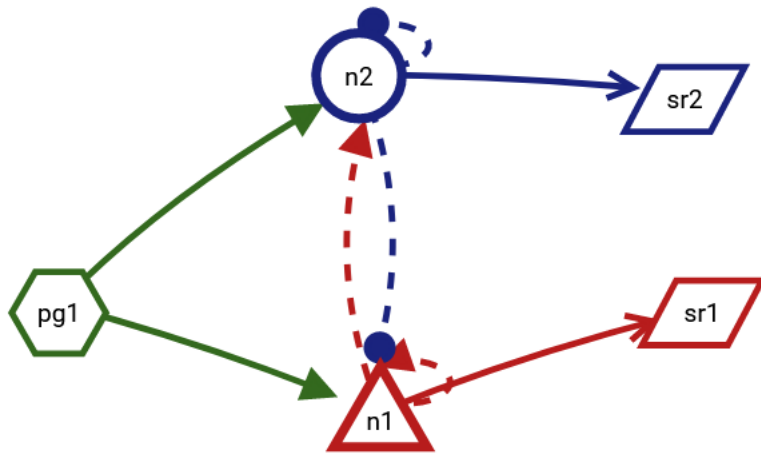
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3

Network activity

Cells in the cortex form columns. In this image the red neurons, called pyramidal cells, are revealed to be entwined by blue fibers from other, inhibitory neurons that slow their firing.





N1 IAF PSC ALPHA		N2 IAF PSC ALPHA		PG1 POISSON GENERATOR		
population size	800	population size	200	population size	1	
membrane time constant (ms)	20	membrane time constant (ms)	20	mean firing rate (Hz)	35000	
N1	⇒	SR1	N2	⇒	N2	
Connection rule	all to all	Connection rule	pairwise Bernoulli	Connection rule	all to all	
N1	⇒	N1	N2	⇒	N1	
Connection rule	pairwise Bernoulli	p	0.2	Connection rule	all to all	
p	0.2	synaptic weight (pA)	-1	PG1	⇒	N2
synaptic weight (pA)	1	synaptic delay (ms)	1	Connection rule	all to all	
synaptic delay (ms)	1	N2	⇒	N1		
N1	⇒	N2	⇒	SR2		
Connection rule	pairwise Bernoulli	Connection rule	pairwise Bernoulli			
p	0.2	p	0.2			
synaptic weight (pA)	1	synaptic weight (pA)	-1			
		synaptic delay (ms)	1			
		Connection rule	all to all			

Recurrent networks of excitatory and inhibitory neurons

Set up a large-enough population of excitatory neurons and one inhibitory, based on the standard LIF model. Establish random synaptic connections among neurons.

Explore the role of external input for the dynamics of the recurrent network. As this external input is normally provided by other neurons that are not part of the local network in question, a Poisson generator represents an adequate model for it. Fix a good value for the rate of the external drive, just above threshold.




Monitor neuronal activity in the network(membrane potentials and spike trains).

The goal should be to establish stable activity in the network, which is characterized by low firing rates, irregular (Poisson-like) spike trains, and a low degree of synchrony across neurons.



Available master thesis on Computational Neuroscience

Prof. Pedrocchi

tutors	title
	Analysis of microendoscopic calcium imaging and behavioural data recorded in-vivo in freely-moving mice
 	Diffusive plasticity model in spiking neural network