

Supplementary Information

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1. Example of the use of NMDS for reducing the dimensionality of the communicability angle matrix

In this example at table 1, we report the communicability angle matrix θ for a tree with degree sequence 3,3,2,2,1,1,1,1. Then, using the procedures
5 described in the main text of this work we obtained the communicability angles
matrices $\hat{\theta}$ for the four mapping criteria described in the main text. Below
each matrix $\hat{\theta}$ we report the RMSE and SE values for the comparison of the
corresponding matrix and θ . According to RMSE value the best fit is obtained
for the strain method $RMSE = 96.53$. However, a closer look at the matrix $\hat{\theta}$
10 obtained from this method reveals some worrying signs. In particular, there are
some angles very close to zero although in the original communicability angle
matrix no angle is below 50° . In contrast, the method based on the eigenvalues
of the communicability angle matrices reports that the best fit is obtained for
the *metricstress*, method for which such "pathologies" are not observed.

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$\begin{pmatrix} 0 & 67.0 & 88.2 & 88.2 & 83.4 & 83.4 & 44.1 & 70.9 \\ & 0 & 88.2 & 88.2 & 83.4 & 83.4 & 44.1 & 70.9 \\ & & 0 & 88.2 & 43.3 & 83.4 & 84.2 & 71.9 \\ & & & 0 & 83.6 & 43.3 & 84.2 & 71.9 \\ & & & & 0 & 71.1 & 72.4 & 47.0 \\ & & & & & 0 & 72.4 & 47.0 \\ & & & & & & 0 & 48.3 \\ & & & & & & & 0 \end{pmatrix}$							
$\begin{pmatrix} 0 & 71.8 & 142.8 & 115.7 & 120.6 & 103.6 & 35.2 & 81.0 \\ & 0 & 95.3 & 95.6 & 126.6 & 130.5 & 47.9 & 150.7 \\ & & 0 & 99.8 & 41.2 & 110.8 & 111.1 & 100.1 \\ & & & 0 & 116.8 & 40.4 & 132.6 & 106.0 \\ & & & & 0 & 98.8 & 110.0 & 59.8 \\ & & & & & 0 & 138.0 & 65.9 \\ & & & & & & 0 & 65.9 \\ & & & & & & & 0 \end{pmatrix}$							
<i>metricstress: RMSE = 102.49, SE = 86.027</i>							
$\begin{pmatrix} 0 & 66.3 & 99.5 & 99.5 & 124.1 & 124.1 & 52.8 & 150.7 \\ & 0 & 126.4 & 126.4 & 118.6 & 118.6 & 13.5 & 84.4 \\ & & 0 & 106.3 & 28.8 & 111.9 & 123.9 & 98.0 \\ & & & 0 & 111.9 & 28.8 & 124.0 & 98.0 \\ & & & & 0 & 102.6 & 124.0 & 69.3 \\ & & & & & 0 & 124.0 & 69.3 \\ & & & & & & 0 & 97.9 \\ & & & & & & & 0 \end{pmatrix}$							
<i>metricstress:RMSE = 99.07, SE = 86.387</i>							
$\begin{pmatrix} 0 & 68.9 & 98.6 & 91.5 & 138.0 & 122.3 & 54.0 & 143.6 \\ & 0 & 104.8 & 153.3 & 98.4 & 132.6 & 43.2 & 84.1 \\ & & 0 & 95.7 & 44.3 & 116.8 & 140.9 & 112.1 \\ & & & 0 & 108.2 & 44.0 & 110.7 & 104.0 \\ & & & & 0 & 96.3 & 138.7 & 67.9 \\ & & & & & 0 & 102.1 & 60.6 \\ & & & & & & 0 & 89.6 \\ & & & & & & & 0 \end{pmatrix}$							
<i>sammon:RMSE = 103.89, SE = 86.553</i>							
$\begin{pmatrix} 0 & 0.5 & 112.6 & 112.6 & 129.2 & 129.2 & 32.8 & 117.5 \\ & 0 & 112.9 & 112.9 & 129.2 & 129.2 & 32.3 & 117.0 \\ & & 0 & 101.1 & 37.8 & 113.4 & 126.7 & 105.7 \\ & & & 0 & 113.4 & 37.8 & 126.7 & 105.7 \\ & & & & 0 & 100.6 & 118.8 & 68.1 \\ & & & & & 0 & 118.8 & 68.0 \\ & & & & & & 0 & 84.7 \\ & & & & & & & 0 \end{pmatrix}$							
<i>strain:RMSE = 96.53, SE = 87.949</i>							

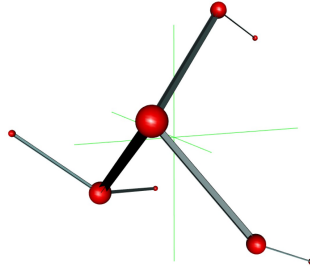


Table 1: Communicability angle matrix (top line) of a tree with degree sequence 3,3,2,2,1,1,1,1. The communicability angle matrices obtained by using NMDS and the following scaling criteria: metricstress (second line, left); metricstress (second line, right); sammon (third line, left); strain (third line, right). The values of the RMSE and SE for each scaling criterion is given below the angle matrices. The bottom line displays the embedding into 3D space using the best scaling according to *SE*, i.e., metricstress.

15 2. Nonmetric Multidimensional Scaling

The NMDS transforms θ into a matrix of squared Euclidean distances with elements $d_{pq}^2(\theta) = \sum_{r=1}^n (\theta_{ps} - \theta_{qs})^2$ which can be obtained by

$$D^{(2)} = \vec{1}\vec{\kappa}^T + \vec{\kappa}\vec{1}^T - 2\theta\theta^T, \quad (1)$$

where $\vec{\kappa} = \text{diag}(\theta)$.

The NMDS method works as follows. Starting from a completely random
 20 configuration of the n nodes of a network in a 3D space, the distances between the nodes in these random configurations are calculated and regressed using a least-squared method to those in $D^{(2)}$. Then, we calculate the goodness of fit for each configuration by the so-called *stress function* introduced by Kruskal, which is defined by

$$S(\theta, \hat{d}) = \sqrt{\frac{\sum_{p,q} (d_{pq} - \hat{d}_{pq})^2}{\sum_{p,q} d_{pq}^2}}, \quad (2)$$

25 where \hat{d}_{pq} is the distance between the corresponding nodes in the configuration model. With the goal of minimizing such stress the positions of the nodes in the ordination space are changed in the direction of the steepest descent, which is the direction in which stress changes most rapidly. The procedure is repeated until a minimum (which may be local) is reached or the total number
 30 of iterations—here set to 1,000—is reached. Once we have obtained the “optimal” distance matrix $\hat{D}^{(2)}$ we construct the corresponding estimated communicability matrix as follow. We first construct the matrix $J = I - \vec{1}\vec{1}^T / \vec{1}^T \vec{1}$ as the centering matrix and obtain

$$\theta\theta^T = -\frac{1}{2}J\hat{D}^{(2)}J. \quad (3)$$

Then, by using the spectral decomposition of $-\frac{1}{2}J\hat{D}^{(2)}J = Q\Upsilon Q^T$ we finally

35 obtain

$$\theta = Q\Upsilon^{1/2}. \quad (4)$$

3. k-Means algorithm

K-Means operates by considering the matrix $\hat{\theta}$ as the initial dataset consisting of column vectors which represents the nodes of the graph. That is, the i th column of $\hat{\theta}$ represents an n -dimensional vector representing node i . Then, K-
 40 Means generates K non-empty disjoint clusters $C = \{C_1, C_2, \dots, C_K\}$ around the centroids $c = \{c_1, c_2, \dots, c_K\}$, by iteratively minimizing the sum [29, 52]

$$W_K = \sum_{k=1}^K \sum_{i \in C_K} \sum_{\gamma=1}^n (\theta_{i\gamma} - c_{k\gamma})^2. \quad (5)$$

4. Cluster Validity Indexes

In the next paragraphs we define the CVIs that we will use in this work. Let c be the center of the network and let c_i be the center of the cluster C_i which
 45 has n_i nodes. Let x denotes any node in the network. Then, if the distance between two points p and q is given by $d(p, q)$, the *Calinski-Harabasz index* is defined by [8]

$$CH = \frac{\frac{1}{n_c} \sum_i n_i d^2(c_i, c)}{\frac{1}{n - n_c} \sum_i \sum_{x \in C_i} d^2(x, c_i)}. \quad (6)$$

Let $x \in C_i$ and let $a(x)$ represents the average dissimilarity of the node x to all other nodes in the cluster C_i . If $b(x)$ is the lowest average dissimilarity
 50 of x to any other cluster $C_j \neq C_i$, then the *Silhouette index* is defined by [49]

$$S = \frac{1}{n_c} \sum_i \left\{ \frac{1}{n_i} \sum_{x \in C_i} \frac{b(x) - a(x)}{\max[a(x), b(x)]} \right\}. \quad (7)$$

The *Davies-Bouldin index* is defined as [9]

$$DB = \frac{1}{n_c} \sum_i \max_{j, j \neq i} \left\{ \frac{\frac{1}{n_i} \sum_{x \in C_i} d(x, c_i) + \frac{1}{n_j} \sum_{x \in C_j} d(x, c_j)}{d(c_i, c_j)} \right\}. \quad (8)$$

5. Best” Approach Selection”

5.1. Normalized Mutual Information (NMI)

Here we use a normalization introduced by Strehl and Ghosh for which NMI
55 index is defined as [53]

$$NMI(x, y) = \frac{I(x, y)}{\sqrt{H(x)H(y)}}, \quad (9)$$

where $I(x, y)$ is the mutual information between x and y , and $H(x)$ is the entropy of x . In our case x is a vector which contains a label for the different communities in the ground-truth of the corresponding network and y is a similar vector containing the labels of the clusters obtained by K-means.

60 5.2. Modularity

The modularity is defined by [44]

$$Q = \frac{1}{4m} \sum_{i, j \in C_k} \left[A_{ij} - \frac{k_i k_j}{2m} \right], \quad (10)$$

where A_{ij} is the corresponding entry of the adjacency matrix, k_i is the degree of the node i , m is the number of edges and the sum is carried out over all communities C_k existing in the network.

6. Tables

N	1	2	3	4	5	6	7	8	9	10
gene	NDRG1	FGF14	NEFH	PPP2R2B	SLC25A22	GABRA1	JPH3	GJB1	UCHL1	DNM2
ref	[1]	[2]	[3]	[4]	[5]	[6]	[7]	[8]	[9]	[10]

N	11	12	13	14	15	16	17	18	19
gene	TDP1	SOD1	PARK7	LRRK2	KIF1B	HSPD1	NR4A2	Rab7	SNCAIP
ref.	[11]	[12]	[13]	[14]	[15]	[16]	[17]	[18]	[19]

Table 2: List of references for the findings of cancer activity of genes reported in cluster 1 according to the Table 4 of the main paper.

N	1	2	3	4	5	6	7	8	9	10
gene	ABCA1	ESR1	ALOX5	IL10	IL13	CIITA	PTPRC	BDNF	PLA2G7	CD36
ref	[20]	[21]	[22]	[23]	[24]	[25]	[26]	[27]	[28]	[29]

Table 3: List of references for the findings of cancer activity of genes reported in cluster 51 according to the Table 5 of the main paper.

References

- [1] T. P. Ellen, Q. Ke, P. Zhang, M. Costa, Ndr1, a growth and cancer related gene: regulation of gene expression and function in normal and disease states, *Carcinogenesis* 29 (1) (2008) 2–8.
- [2] T. P. Ellen, Q. Ke, P. Zhang, M. Costa, Ndr1, a growth and cancer related gene: regulation of gene expression and function in normal and disease states, *Carcinogenesis* 29 (1) (2008) 2–8.
- [3] M. F. Calmon, J. Jeschke, W. Zhang, M. Dhir, C. Siebenks, A. Herrera, H.-C. Tsai, H. M. O’Hagan, E. P. Pappou, C. M. Hooker, T. Fu, K. E. Schuebel, E. Gabrielson, P. Rahal, J. G. Herman, S. B. Baylin, N. Ahuja, Epigenetic silencing of neurofilament genes promotes an aggressive phenotype in breast cancer, *Epigenetics* 10 (7) (2015) 622–632.
- [4] A. Vazquez, D. Kulkarni, L. F. Grochola, G. L. Bond, N. Barnard, D. Toppmeyer, A. J. Levine, K. M. Hirshfield, A genetic variant in a pp2a regulatory

- subunit encoded by the ppp2r2b gene associates with altered breast cancer risk and recurrence, *International Journal of Cancer* 128 (10) (2011) 2335–2343.
- [5] C. C. Wong, Y. Qian, X. Li, J. Xu, W. Kang, J. H. Tong, K.-F. To, Y. Jin, W. Li, H. Chen, M. Y. Y. Go, J.-L. Wu, K. W. Cheng, S. S. M. Ng, J. J. Y. Sung, Z. Cai, J. Yu, Slc25a22 promotes proliferation and survival of colorectal cancer cells with kras mutations and xenograft tumor progression in mice via intracellular synthesis of aspartate, *Gastroenterology* 151 (5) (2016) 945–960.e6.
- [6] S. Lee, T. Oh, H. Chung, S. Rha, C. Kim, Y. Moon, Slc25a22 promotes proliferation and survival of colorectal cancer cells with kras mutations and xenograft tumor progression in mice via intracellular synthesis of aspartate, *International Journal of Oncology* 40.
- [7] X. Hu, Y. Kuang, L. Li, H. Tang, Q. Shi, X. Shu, Y. Zhang, F. K. Chan, Q. Tao, C. He, Epigenomic and functional characterization of junctophilin 3 (jph3) as a novel tumor suppressor being frequently inactivated by promoter cpG methylation in digestive cancers, *Theranostics* 7(7).
- [8] S. Sirnes, H. Honne, D. Ahmed, S. A. Danielsen, T. O. Rognum, G. I. Meling, E. Leithe, E. Rivedal, R. A. Lothe, G. E. Lind, Dna methylation analyses of the connexin gene family reveal silencing of gjc1 (connexin45) by promoter hypermethylation in colorectal cancer, *Epigenetics* 6 (5) (2011) 602–609.
- [9] E. OkochiTakada, K. Nakazawa, M. Wakabayashi, A. Mori, S. Ichimura, T. Yasugi, T. Ushijima, Silencing of the uchl1 gene in human colorectal and ovarian cancers, *International Journal of Cancer* 119 (6) (2006) 1338–1344.
- [10] H. P. Joshi, I. V. Subramanian, E. K. Schnettler, G. Ghosh, R. Rupaimoole, C. Evans, M. Saluja, Y. Jing, I. Cristina, S. Roy, Y. Zeng, V. H. Shah, A. K. Sood, S. Ramakrishnan, Dynamin 2 along with microrna-199a reciprocally

regulate hypoxia-inducible factors and ovarian cancer metastasis, *Proceedings of the National Academy of Sciences* 111 (14) (2014) 5331–5336.

- [11] C. Liu, S. Zhou, S. Begum, D. Sidransky, W. H. Westra, M. Brock, J. A. Califano, Increased expression and activity of repair genes tdp1 and xpf in non-small cell lung cancer, *Lung Cancer* 55 (3) (2007) 303–311.
- [12] L. Papa, G. Manfredi, D. Germain, Sod1, an unexpected novel target for cancer therapy, in: *Genes & cancer*, 2014.
- [13] C. M. Clements, R. S. McNally, B. J. Conti, T. W. Mak, J. P.-Y. Ting, Dj-1, a cancer- and parkinson’s disease-associated protein, stabilizes the antioxidant transcriptional master regulator nrf2, *Proceedings of the National Academy of Sciences* 103 (41) (2006) 15091–15096.
- [14] R. SaundersPullman, M. J. Barrett, K. M. Stanley, M. S. Luciano, V. Shanker, L. Severt, A. Hunt, D. Raymond, L. J. Ozelius, S. B. Bressman, Lrrk2 g2019s mutations are associated with an increased cancer risk in parkinson disease, *Movement Disorders* 25 (15) (2010) 2536–2541.
- [15] Z.-C. Wang, Q. Gao, J.-Y. Shi, L.-X. Yang, J. Zhou, X.-Y. Wang, Y.-H. Shi, A.-W. Ke, G.-M. Shi, Z.-B. Ding, Z. Dai, S.-J. Qiu, J. Fan, Genetic polymorphism of the kinesin-like protein kif1b gene and the risk of hepatocellular carcinoma, *PLOS ONE* 8 (4) (2013) e62571.
- [16] G. Li, M. Li, X. Liang, Z. Xiao, P. Zhang, M. Shao, F. Peng, Y. Chen, Y. Li, Z. Chen, Identifying dcn and hspd1 as potential biomarkers in colon cancer using 2d-lc-ms/ms combined with itraq technology, *Journal of Cancer* 8 (3) (2017) 479–489.
- [17] Y. F. Han, G. W. Cao, Role of nuclear receptor nr4a2 in gastrointestinal inflammation and cancers, *World Journal of Gastroenterology* 18 (47) (2012) 6865–6873.

- [18] W. J. Chia, B. L. Tang, Emerging roles for rab family gtpases in human cancer, *Biochimica et Biophysica Acta (BBA) - Reviews on Cancer* 1795 (2) (2009) 110–116.
- [19] A. Korshunov, F. Sahm, D. Stichel, D. Schrimpf, M. Ryzhova, O. Zheludkova, A. Golanov, P. Lichter, D. T. W. Jones, A. von Deimling, S. M. Pfister, M. Kool, Molecular characterization of medulloblastomas with extensive nodularity (mben), *Acta Neuropathologica* (2018) 1–11.
- [20] B. H. Lee, M. G. Taylor, P. Robinet, J. D. Smith, J. Schweitzer, E. Sehayek, S. M. Falzarano, C. Magi-Galluzzi, E. A. Klein, A. H. Ting, Dysregulation of cholesterol homeostasis in human prostate cancer through loss of abca1, *Cancer Research* 73 (3) (2013) 1211–1218.
- [21] D. R. Robinson, Y. M. Wu, P. Vats, F. Su, R. J. Lonigro, X. Cao, S. Kalyana-Sundaram, R. Wang, Y. Ning, L. Hodges, A. Gursky, J. Siddiqui, S. A. Tomlins, S. Roychowdhury, K. J. Pienta, S. Y. Kim, J. S. Roberts, J. M. Rae, C. H. Van Poznak, D. F. Hayes, R. Chugh, L. P. Kunju, M. Talpaz, A. F. Schott, A. M. Chinnaiyan, Activating *esr1* mutations in hormone-resistant metastatic breast cancer, *Nature Genetics* 45 (12) (2013) 1446–1451.
- [22] Y. Chen, Y. Hu, H. Zhang, C. Peng, S. Li, Loss of the *alox5* gene impairs leukemia stem cells and prevents chronic myeloid leukemia, *Nature Genetics* 41 (7) (2009) 783–792.
- [23] D. S. Michaud, S. E. Daugherty, S. I. Berndt, E. A. Platz, M. Yeager, E. D. Crawford, A. Hsing, W.-Y. Huang, R. B. Hayes, Genetic polymorphisms of interleukin-1b (*il-1b*), *il-6*, *il-8*, and *il-10* and risk of prostate cancer, *Cancer Research* 66 (8) (2006) 4525–4530.
- [24] J. Iwashita, Y. Sato, H. Sugaya, N. Takahashi, H. Sasaki, T. Abe, *mrna of muc2 is stimulated by il-4, il-13 or tnf-alpha through a mitogen-activated protein kinase pathway in human colon cancer cells*, *Immunology and cell biology* 81 4 (2003) 275–82.

- [25] C. Steidl, S. P. Shah, B. W. Woolcock, L. Rui, M. Kawahara, P. Farinha, N. A. Johnson, Y. Zhao, A. Telenius, S. B. Neriah, A. McPherson, B. Meissner, U. C. Okoye, A. Diepstra, A. Van Den Berg, M. Sun, G. Leung, S. J. Jones, J. M. Connors, D. G. Huntsman, K. J. Savage, L. M. Rimsza, D. E. Horsman, L. M. Staudt, U. Steidl, M. A. Marra, R. D. Gascoyne, Mhc class ii transactivator ciita is a recurrent gene fusion partner in lymphoid cancers, *Nature* 471 (7338) (2011) 377–383.
- [26] M. Porcu, M. Kleppe, V. Gianfelici, E. Geerdens, K. De Keersmaecker, M. Tartaglia, R. Fo, J. Soulier, B. Cauwelier, A. Uyttbroeck, E. Macintyre, P. Vandenberghe, V. Asnafi, J. Cools, Mutation of the receptor tyrosine phosphatase *jem/ptprcj/emj* (cd45) in t-cell acute lymphoblastic leukemia, *Blood* 119 (19) (2012) 4476–4479.
- [27] K. Okamura, T. Harada, S. Wang, K. Ijichi, K. Furuyama, T. Koga, T. Okamoto, K. Takayama, T. Yano, Y. Nakanishi, Expression of *trkb* and *bdnf* is associated with poor prognosis in non-small cell lung cancer, *Lung Cancer* 78 (1) (2012) 100–106.
- [28] P. Vainio, L. Lehtinen, T. Mirtti, M. Hilvo, T. Seppnen-Laakso, J. Virtanen, A. Sankila, S. Nordling, J. Lundin, A. Rannikko, M. Orei, O. Kallioniemi, K. Iljin, Phospholipase *pla2g7*, associated with aggressive prostate cancer, promotes prostate cancer cell migration and invasion and is inhibited by statins, *Oncotarget* 2 (12) (2011) 1176–1190.
- [29] J. S. Hale, B. Otvos, M. Sinyuk, A. G. Alvarado, M. Hitomi, K. Stoltz, Q. Wu, W. Flavahan, B. Levison, M. L. Johansen, D. Schmitt, J. M. Neltner, P. Huang, B. Ren, A. E. Sloan, R. L. Silverstein, C. L. Gladson, J. A. DiDonato, J. M. Brown, T. McIntyre, S. L. Hazen, C. Horbinski, J. N. Rich, J. D. Lathia, Cancer stem cell-specific scavenger receptor *cd36* drives glioblastoma progression, *STEM CELLS* 32 (7) (2014) 1746–1758.