

# adapt\_find

**Purpose:** adapt\_find identifies adapters sequences from single end raw sequencing files in FASTQ format. To run the script on raw FASTQ files, the following dependencies are required: PYTHON pandas module, scipy, numpy, cutadapt, blast, and bowtie.

## Usage:

adapt\_find.py <sequencing platform> [-- min\_len] [-- max\_len] [-- index] [-- input\_path] [-- output\_path] [-- files]

Required arguments are indicated within <> and one of the allowed options has to be entered compulsorily. Optional arguments are indicated within square brackets. If the optional arguments are not specified by a user, then the default values will be used. Detailed information regarding required and optional arguments are indicated below.

Required arguments	
Argument	Allowed Options
Sequencing platform	1) ILLUMINA – for ILLUMINA sequencing technology 2) SOLID - for SOLID sequencing technology 3) ION_TORRENT – for Ion Torrent sequencing technology 4) 454 – for Roche 454 Sequencing technology

Optional arguments	
Argument	Default Option
--min_len	15

<code>--max_len</code>	50
<code>--index</code>	None. If the path to the bowtie genome index files is specified, the percentage of number of reads mapping to the genome after adapter trimming will be reported.
<code>--input_path</code>	Current working directory. If the FASTQ files are not in the current working directory, the path to input FASTQ files can be specified here.
<code>--output_path</code>	Current working directory. Users can also specify a different output folder path.
<code>--files</code>	None. If the files are located in different locations, the absolute path of the files can be specified. Note: if <code>--input_path</code> is specified, <code>--files</code> option cannot be specified. If <code>--files</code> argument is specified, files in the current working directory will not be analyzed, even though the output directory will be the current working directory.