

# AVIDD ASAP: EVD68 & EVA71 3C protease expression and purification

PAGE22-01976

Author: Fairhead, Michael  
Date Started: 2022-Nov-21  
Experiment Started:  
Projects: Expression;Purification;ASAP  
Related Pages:  
Referenced by: PAGE23-00563  
Tags:

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D68EV3CPROA-c001/D68EV3CPROA-k001/D68EV3CPROA-e001/D68EV3CPROA-p001

MGPGFDFAQAIMKKNTVIARTEKGEFTMLGVYDRVAVIPTHASVGEIIYINDVETRVLDACALRDLTDNLEITIVKLDRNQKFR  
DIRHFLPRCEDDYNDVLSVHTSKFPNMYIPVGQVTNYGFLNLGGTPTHRILMYNFPTRAGQCQGGVTTTGKVGIVGHVGGNGA  
QGFAAMLLHSYFTDTQKHSHHHHHH

21283.3 Da

10430 mM-1cm-1

A71EV3CPROA-c001/A71EV3CPROA-k001/A71EV3CPROA-e001/A71EV3CPROA-p001

MGPSLDFALSLLRRNIRQVQTDQGHFTMLGVRDRLAVLPRHSQPGKTIWVEHKLINILDAVELVDEQGVNLELTLVTLDTNEKF  
RDITKFIPENISAASDATLVINTEHMPMSMFVPVGDVVQYGFLNLGKPTHRMMYNFPTKAGQCQGGVTSVGKVGIVGHIGGNG  
RQGFCAGLKRSYFASEQLEHHHHHHH

21331.5 Da

9970 mM-1 cm-1

From  
<https://doi.org/10.1107/S0907444913002862>  
The protein was stored in 25 mM Tris-HCl pH 8.0, 150 mM NaCl, 1 mM TCEP at 277 K.

crystallization example

The droplets were prepared by a 1:1 mix of the complex (12 mg/ml) and the reservoir solution containing 100 mM Tris-HCl pH 8.5, 25% PEG4000, and 0.8 M lithium chloride. The crystals were ready for data collection at 16 °C for about 1 week. (<https://doi.org/10.1002/jmr.2551>)

Score	Expect	Method	Identities	Positives	Gaps
214 bits(544)	4e-76	Compositional matrix adjust.	99/184(54%)	128/184(69%)	0/184(0%)
Query 1	MGPGFDFAQAIMKKNTVIARTEKGEFTMLGVYDRVAVIPTHASVGEIIYINDVETRVLDA				60
Sbjct 1	...SL...LSLLRR.IRQVQ.DQ.H.....R..L..L.R.SQP.KT.WVEHKLINI...				60

```
Query 61 CALRDLTDNLEITIVKLDNRNQKFRDIRHFLPRCEDDYNDVLSVHTSKFNPNIYPVGQV 120
Sbjct 61 VE.V.EQGV...L.L.T..T.E.....TK.I.ENISAAS..T.VIN.EHM.S.FV...D. 120

Query 121 TNYGFLNLGGTPTHRILMYNFPTRAGQCGGVVTTTGKVIIGIHVGGNGAQQGFAAMLLHSYF 180
Sbjct 121 VQ.....S.K.....TM.....K.....SV.....I.....R...C.G.KR... 180

Query 181 TDTQ 184
Sbjct 181 ASE. 184
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15 mL o/n in SOC + Kan 50

Use to inoculate 1L AIM-TB

grow 4h 37C/20h 17C 250 rpm shaking

Harvest 4000 g (grew 2L of each protease, around 100 g of cells total for each protease)

Lyse in base buffer + 1 % TX-100 and 0.5 mg/mL HEWL + 1 ug/mL benzonase + 30 mM Imidazole

Centrifuge 30,000g 1h 4C

Pour SN over 15 mL Ni Sepharose FF

Wash 3 x 100 mL Base Buffer + 30 mM Imidazole

Elute 3 x 30 mL Base Buffer + 30 mM Imidazole (protein predominantly in 1st 2 fractions)

D68EV3CPROA

Fraction 1 = 30 mL A280 of 9

Fraction 2 = 30 mL A280 of 6

Fraction 3 = 30 mL A280 of 1.3

A71EV3CPROA

Fraction 1 = 30 mL A280 of 3.4

Fraction 2 = 30 mL A280 of 3.1

Fraction 3 = 30 mL A280 of 1.2

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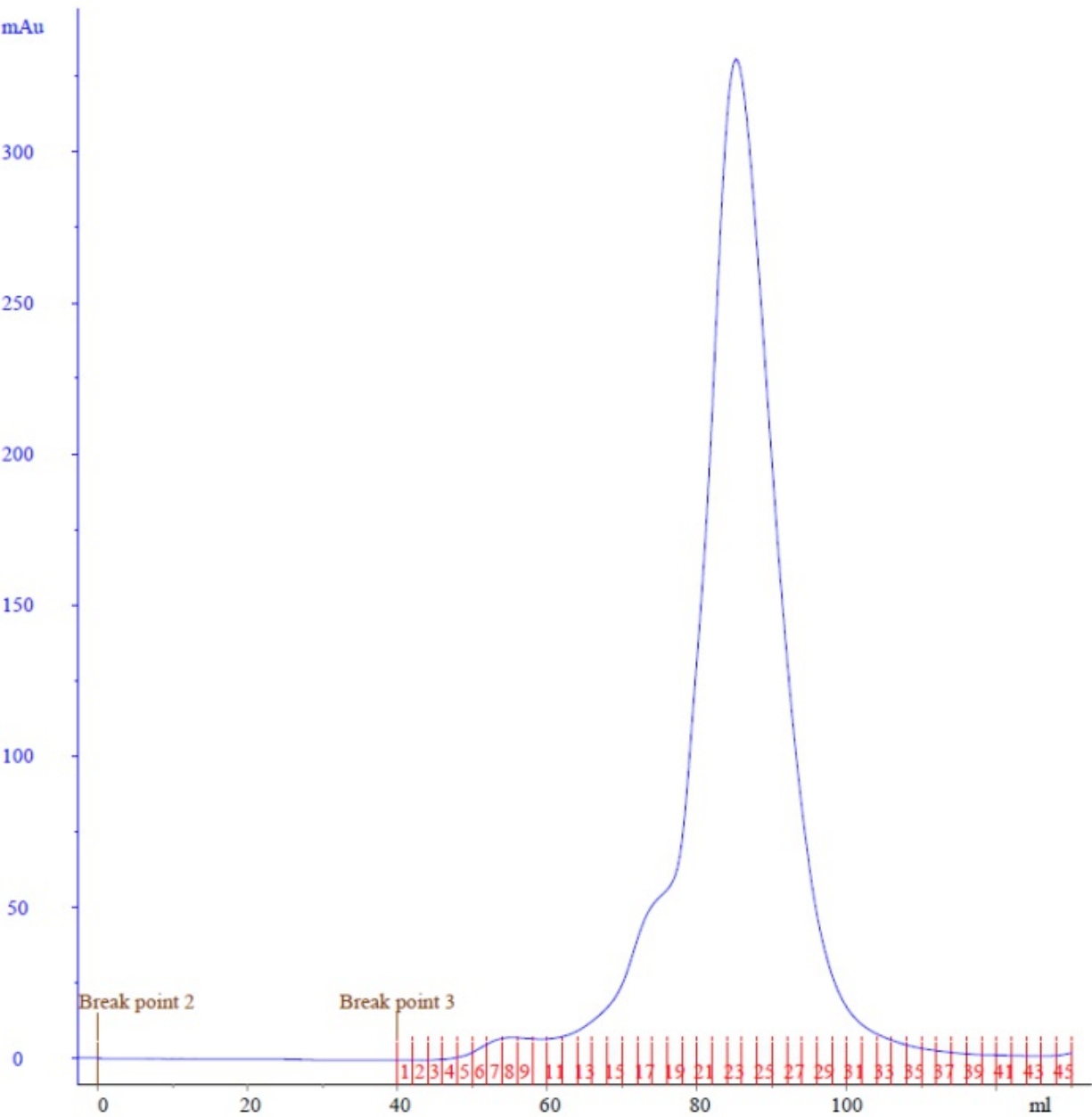
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Dialyze o/n into base buffer + 0.5 mM TCEP

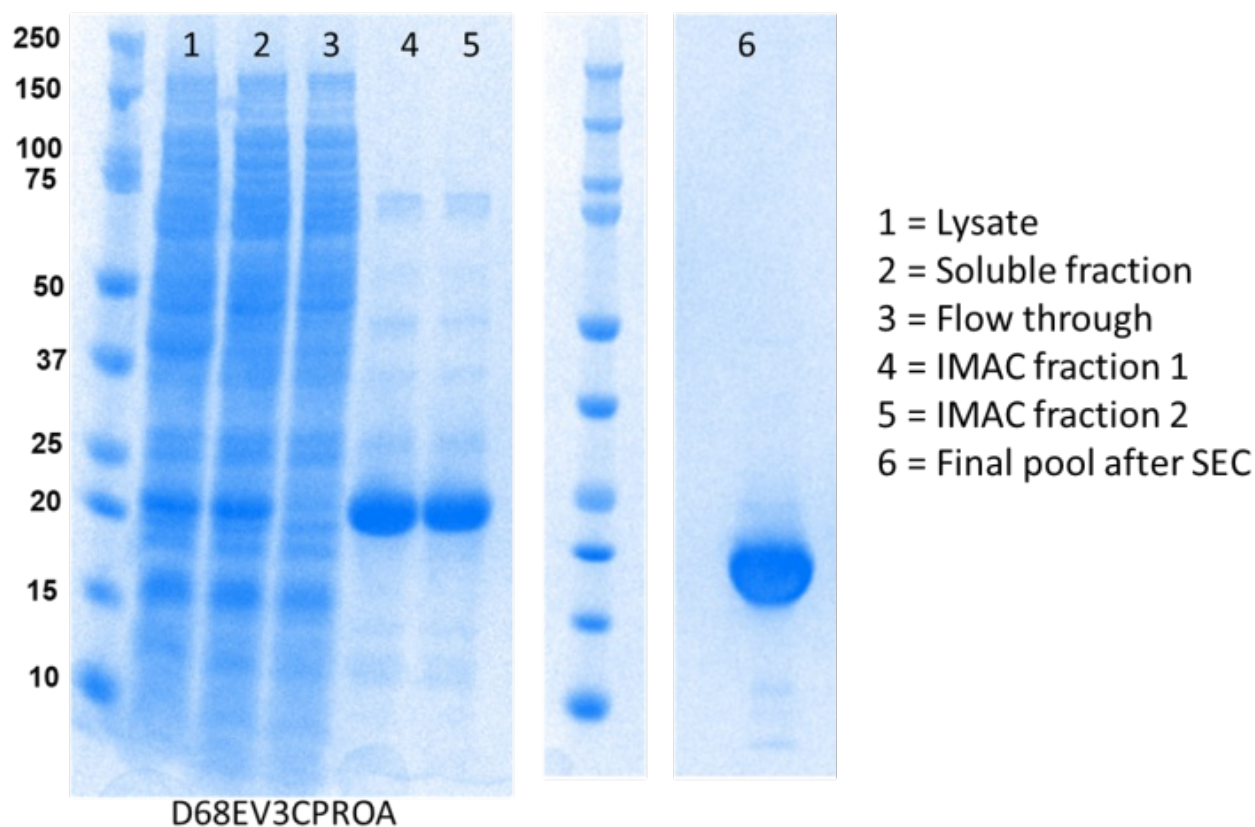
Very very slow to concentrate in both cases about 30 mL was the best I could do in both cases (about 10 mg/mL of EVD68 and 5 mg/mL for EV71)

Run 5 mL aliquots over 125 mL superose 12 pg column using base buffer + 0.5 mM TCEP as mobile phase

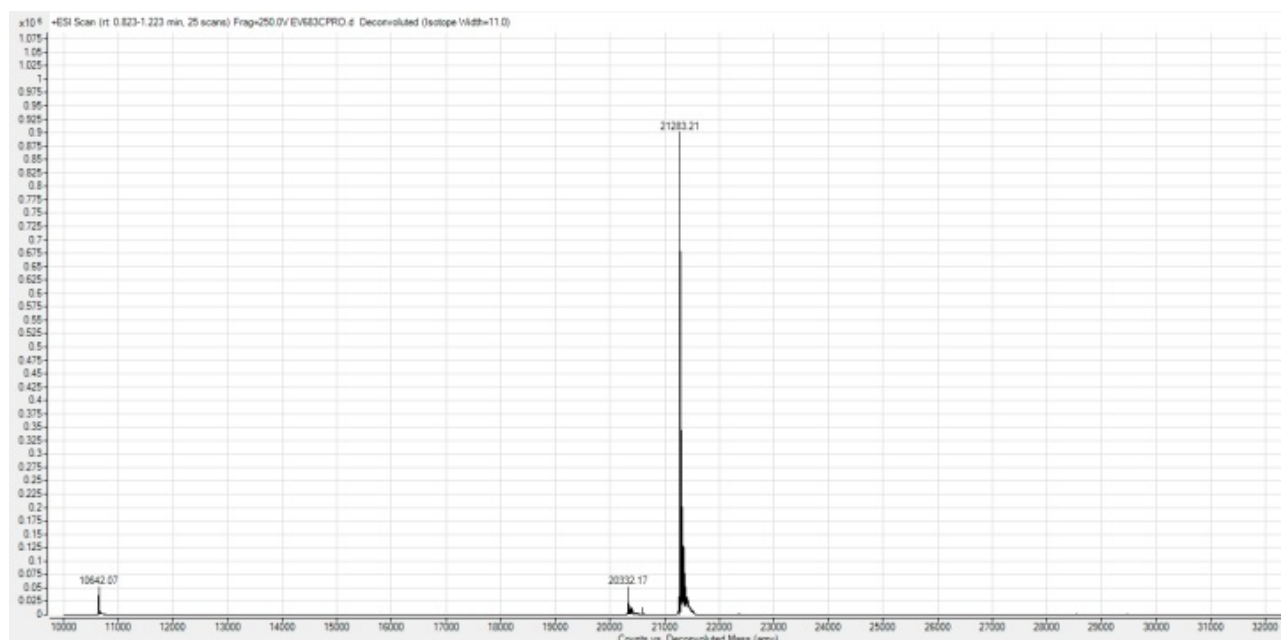
EVD68 3CL Protease superose 12 pg column



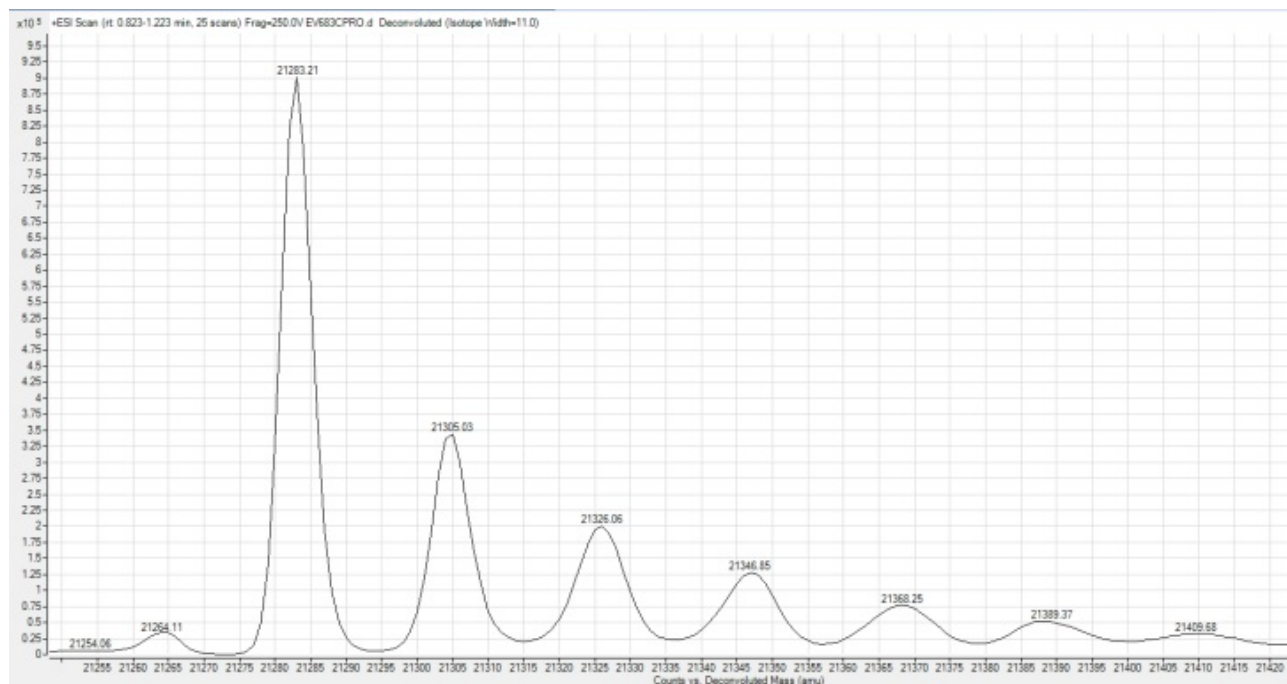
SDS PAGE D68EV3CPROA



#### D68EV3CPROA-p001 MS confirmation Z:\Agilent\_SGC\_QTOF\Rod\221124\_MS424C



#### D68EV3CPROA-p001 MS confirmation Z:\Agilent\_SGC\_QTOF\Rod\221124\_MS424C



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### EVD68 3CL Protease

EVD68 pool concentrates ok to 8.2 (786 uM) 16.7 mg mL  
just over 1.2 mL per run so about 20 mg can do 6 runs in total so about 120 mg from 2 L or 60 mg/L final  
Did three runs

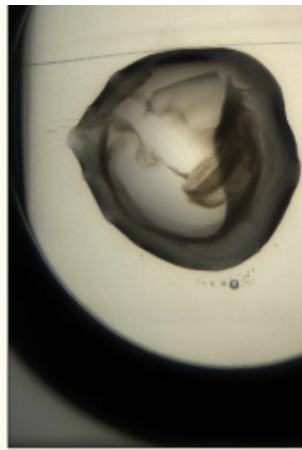
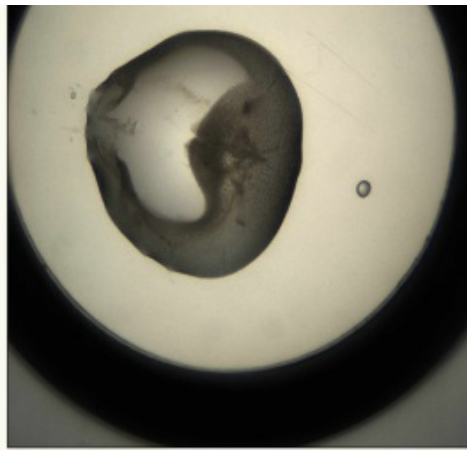
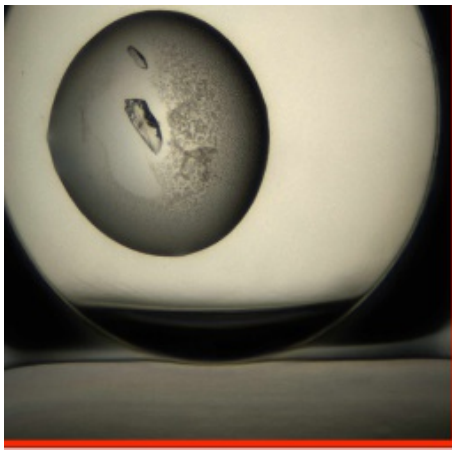
Mass-Spec shows MW 21283.21 Da observed versus 21283.3 Da expected

D68EV3CPROA-p001  
1 mM 31 x 50 uL aliquots after making plates  
300 nL drops in

CI084721 HCS  
CI084722 BCS  
CI084723 LFS  
CI084724 MORPHEUS  
CI084725 JCSG  
CI084726 HIN

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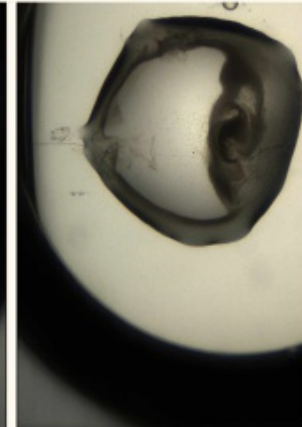
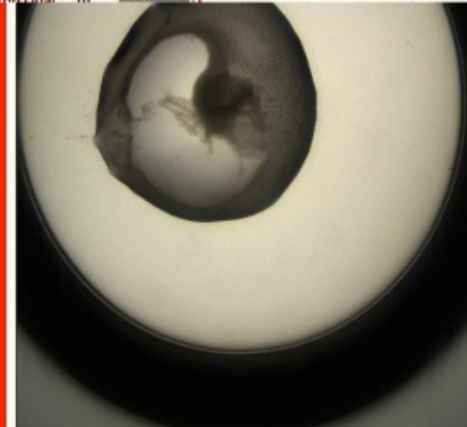
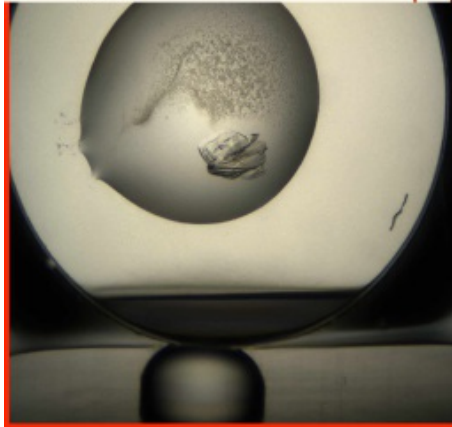
Condition: 0.2M ammonium acetate -- 25% PEG3350 -- 0.1M tris pH 8.5

Plate Stats

Drop Qual.: 10

Projects: D68EV3CPR0A

☐ Sort by last inspection date

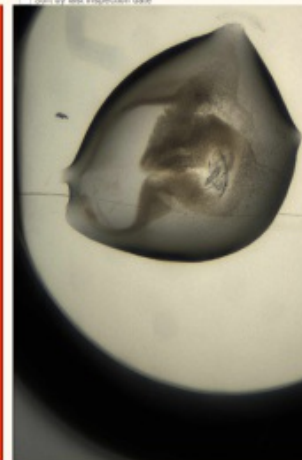
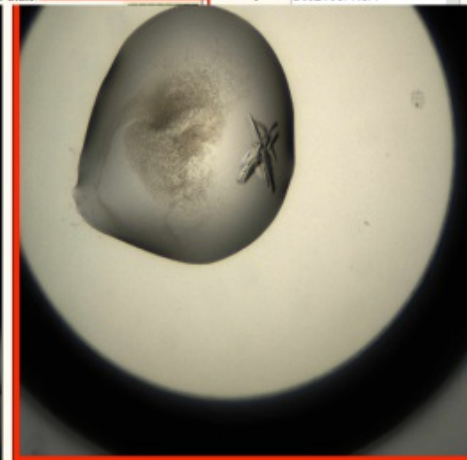
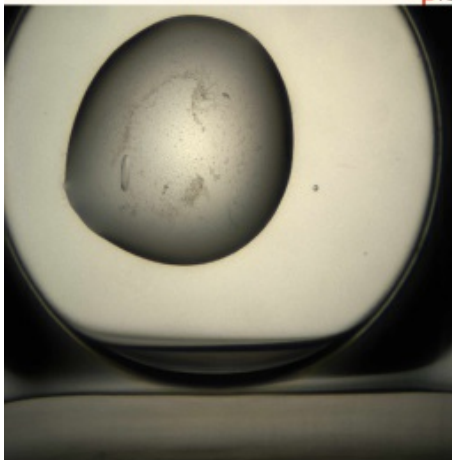


Condition: 25% PEG3350 -- 0.1M tris pH 8.5

Plate Stats

Projects: D68EV3CPR0A

☐ Sort by last inspection date



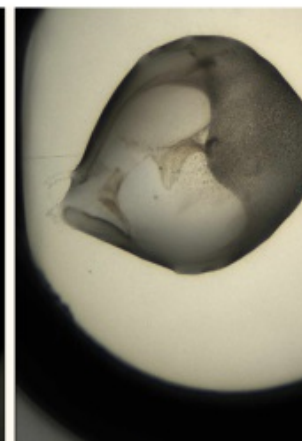
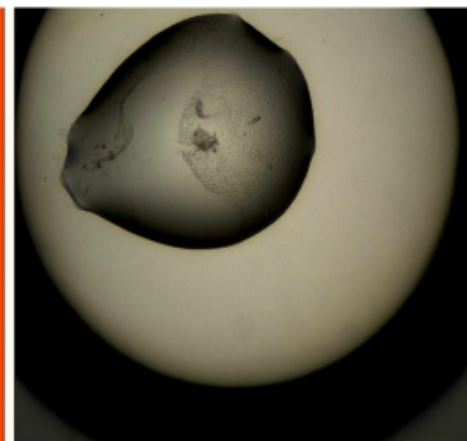
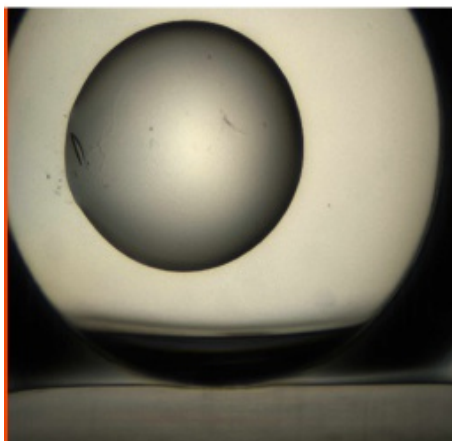
Condition: 0.2M trimethylamine N-oxide -- 20% PEG2000MME -- 0.1M tris pH 8.5

Plate Stats

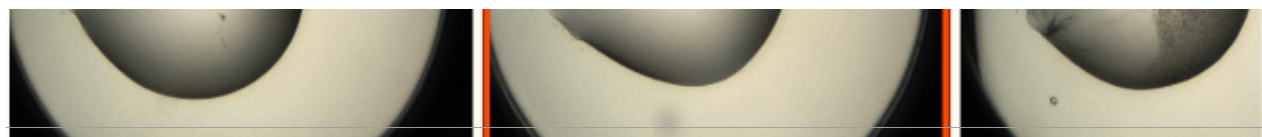
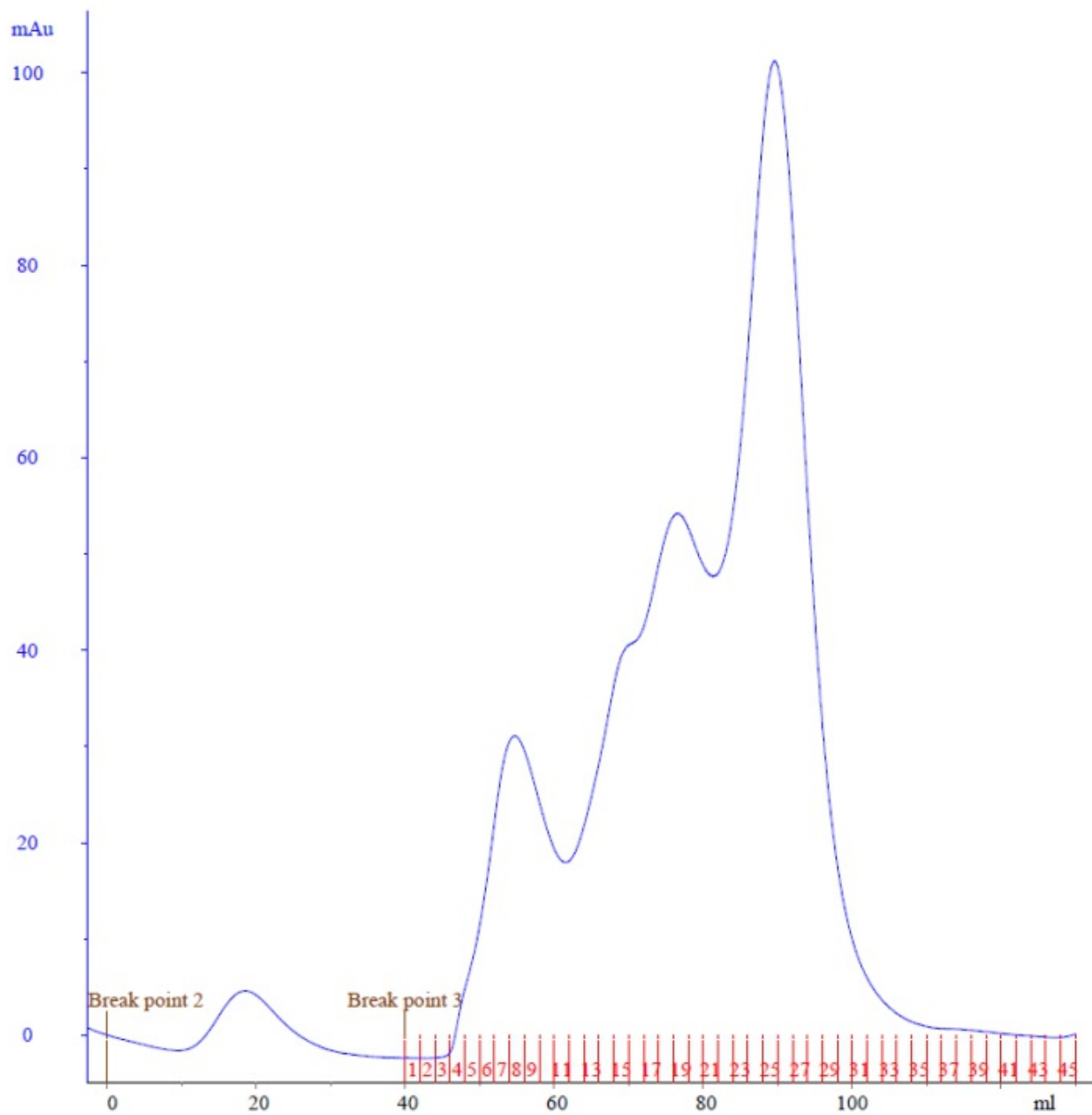
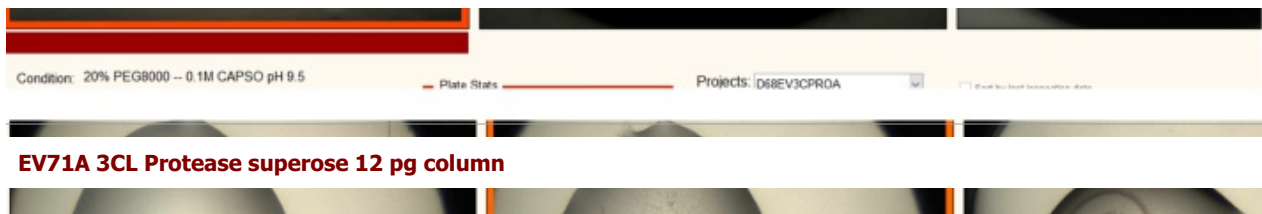
Drop Qual.: 10

Projects: D68EV3CPR0A

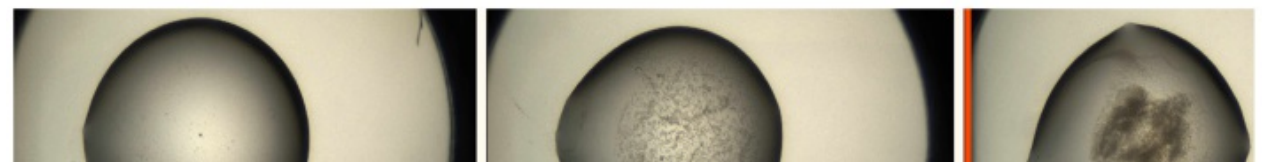
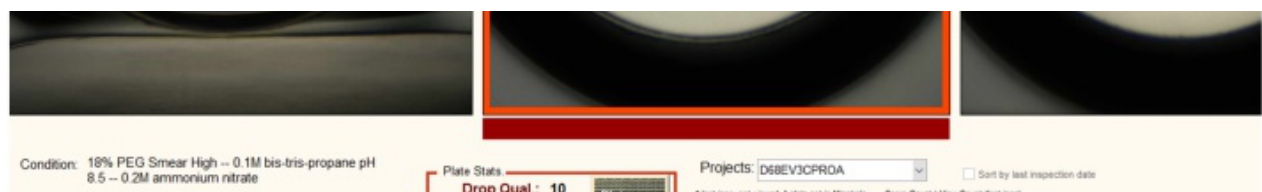
☐ Sort by last inspection date

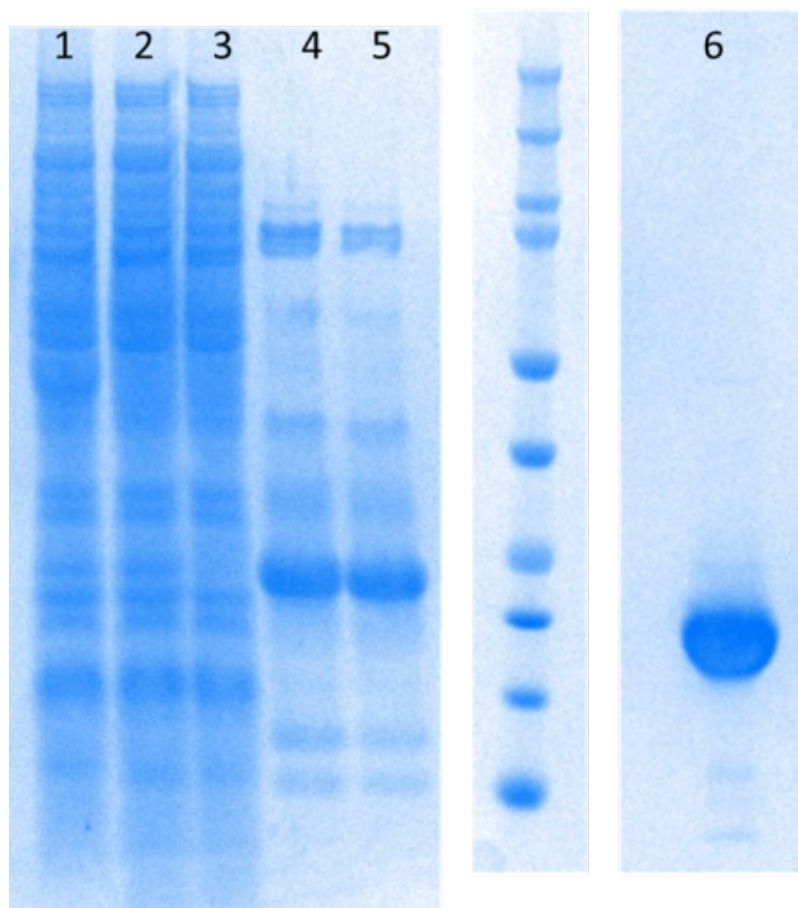






### SDS PAGE A71EV3CPROA

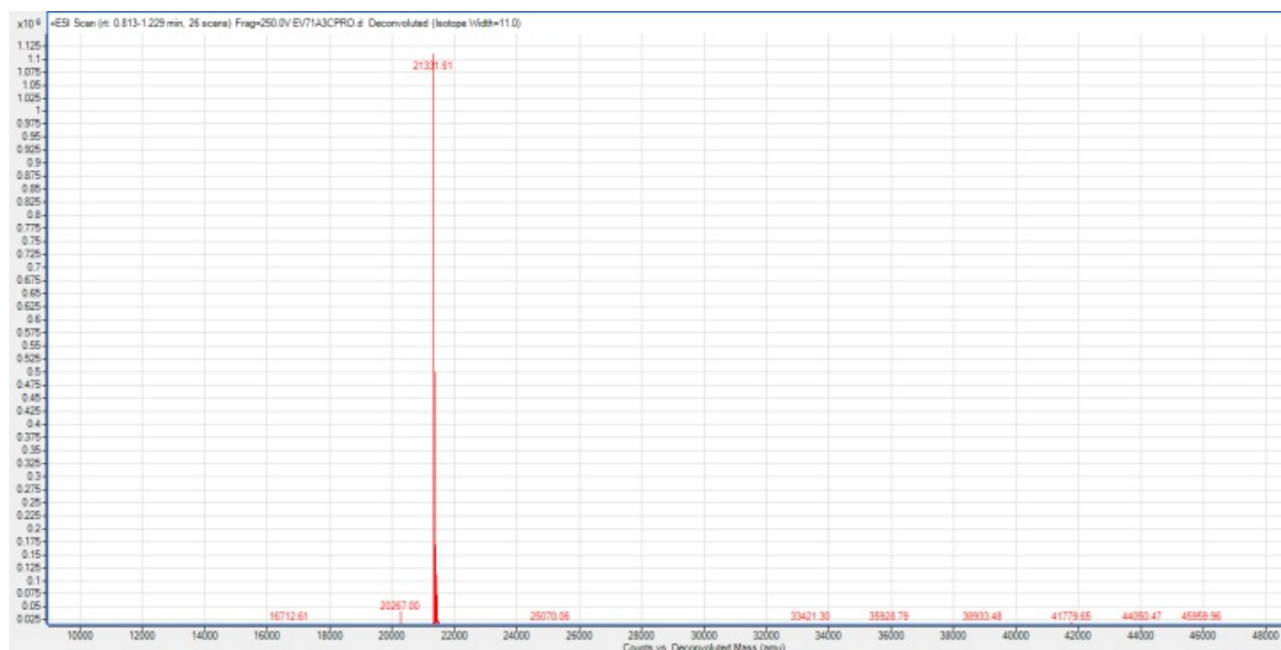




1 = Lysate  
2 = Soluble fraction  
3 = Flow through  
4 = IMAC fraction 1  
5 = IMAC fraction 2  
6 = Final pool after SEC

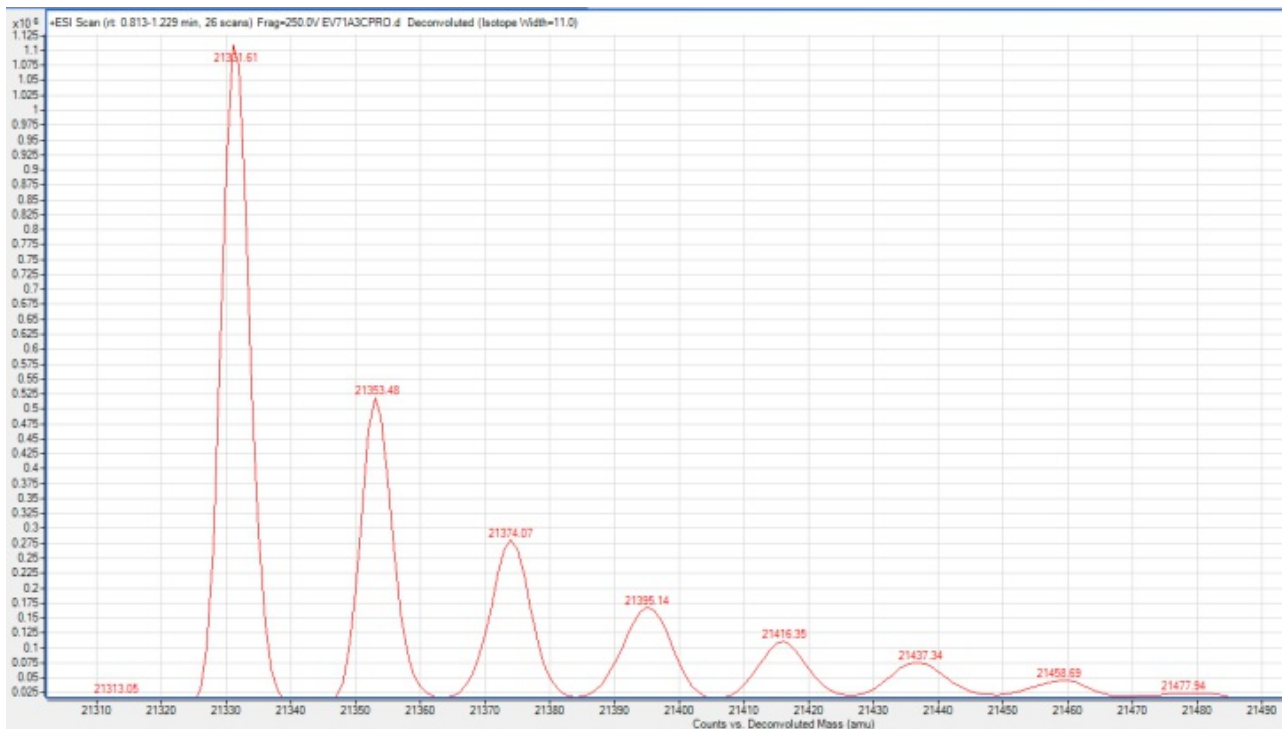
A71EV3CPROA

#### A71EV3CPROA-p001 MS confirmation Z:\Agilent\_SGC\_QTOF\Rod\221124\_MS424C



#### A71EV3CPROA-p001 MS confirmation Z:\Agilent\_SGC\_QTOF\Rod\221124\_MS424C





### EVD71A 3CL Protease

Took peak centered around 90 mL and concentrated as much as possible did a total of 4 runs  
Final yield after much concentrating is 2mL at A280 of 3.4 (341 uM) 7.3 mg/mL

Mass-Spec shows MW 21331.61 Da observed versus 21331.5 Da expected

A71EV3CPROA-p001

0.34 mM 55 x 50 uL aliquots after making plates  
300 nL drops in

CI084990 LFS6

CI084991 BCS

CI084992 MORPHEUS

CI084993 HIN3

CI084994 JCSG7

CI084995 HCS3

Difficulty in cocentrating to 1mM in base buffer so I took a 50 uL aliquot and added either  
0.5 mL of 50 mM Bicine pH 8.5, 500 mM NaCl, 5 % Glycerol, 0.5 mM TCEP  
or

0.5 mL of 50 mM MES pH 6.5, 500 mM NaCl, 5 % Glycerol, 0.5 mM TCEP

Added to 0.5 mL 10000 MWCO cocentrator and reconcentrated

MES one would not go beyond original concentration of 0.34 mM

Bicine concentrated nicely to 1 mM

Thawed several more aliquots and repeated at scale to get enough to repeat original coarse screen plates

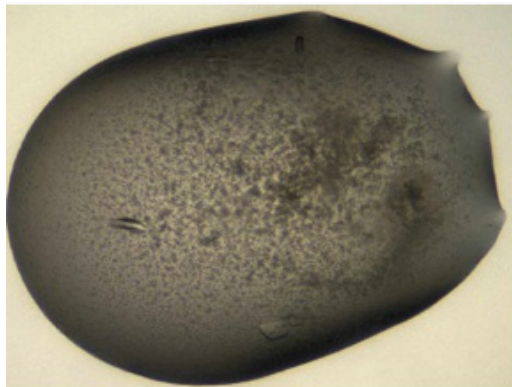
Buffer of protein now 10 mM Bicine pH 8.5, 500 mM NaCl, 5 % Glycerol, 0.5 mM TCEP and protein at 1 mM  
(21.3 mg/mL)

CI084900 BCS  
CI084901 LFS6  
CI084902 HIN3  
CI084903 JCSG  
CI084904 HCS3

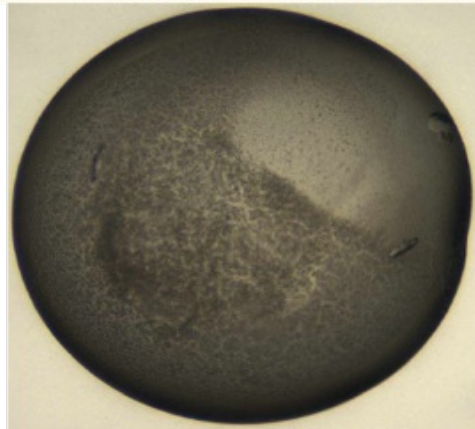
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**After 4 days clear xtals**

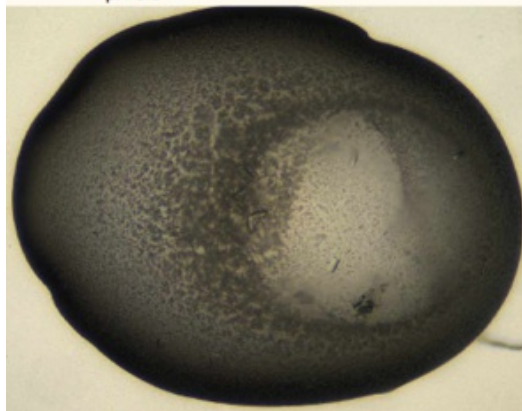
Condition: 30% PEG Smear Low -- 0.1M citrate/phosphate pH 5.5



Condition: 22.5% PEG Smear Medium -- 0.1M PIPES pH 7.0 -- 0.1M calcium chloride -- 0.1M magnesium chloride



Condition: 25% PEG Smear Medium -- 0.1M citrate/phosphate pH 5.5



Condition: 30% PEG4000 -- 0.2M magnesium chloride -- 0.1M tris pH 8.5

