

# Genome assembly strategies

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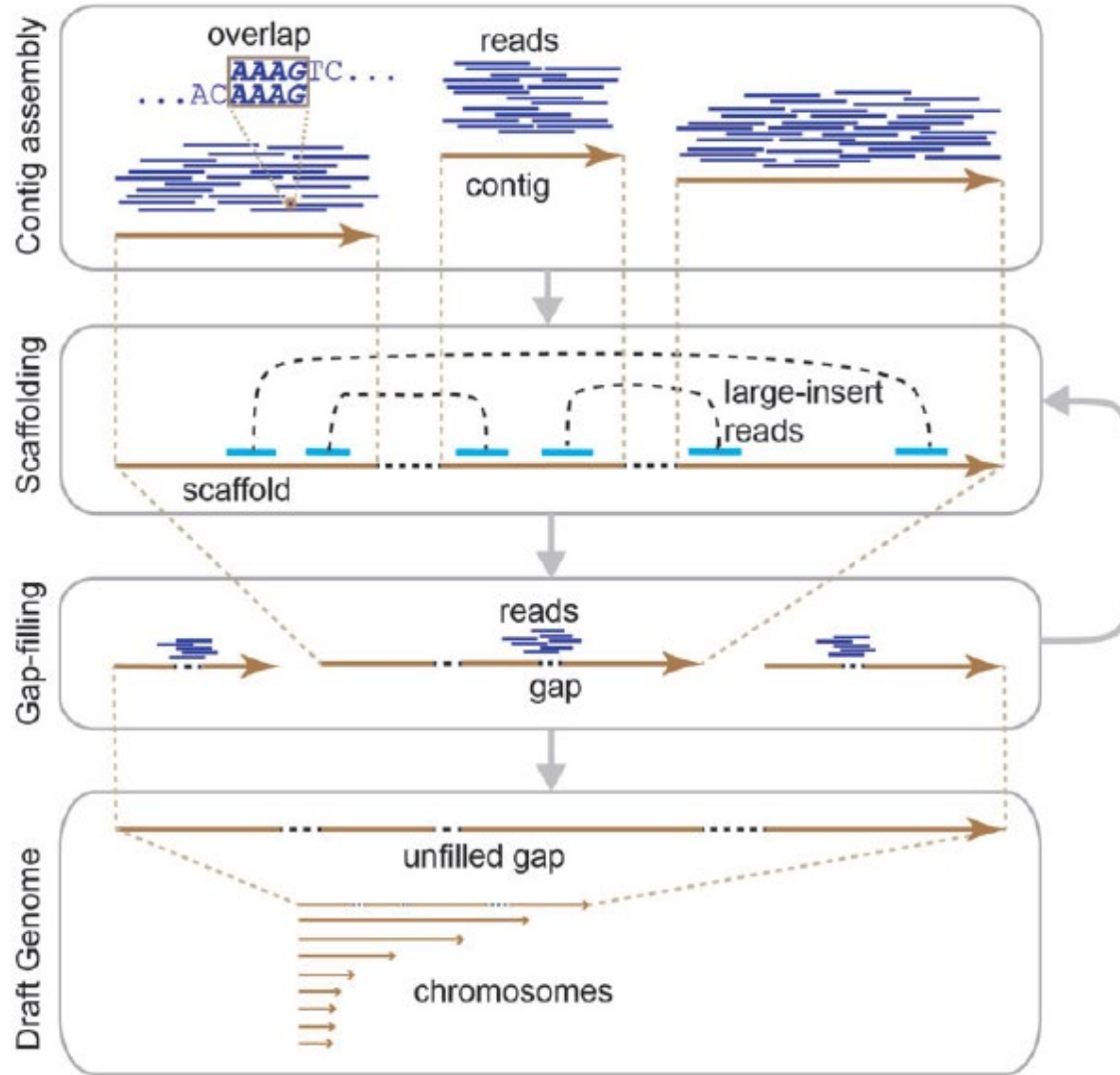
November 2021

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# Genome assembly

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## Summarizing



Reads  
(fastq)

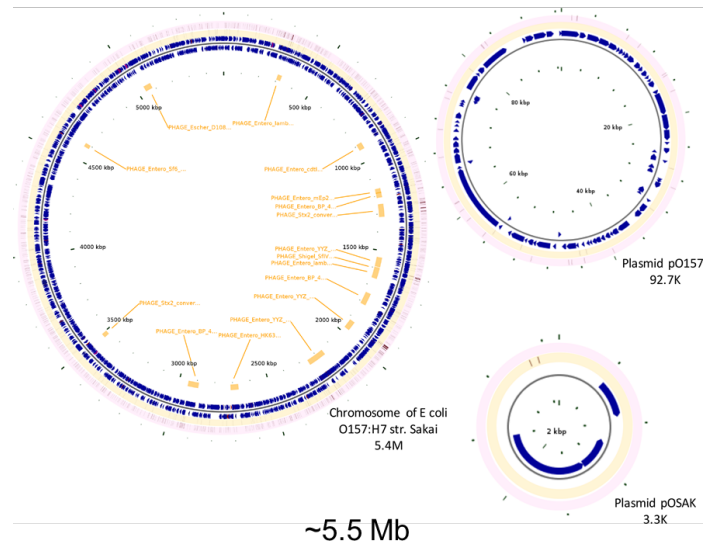
```
SSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSS.....
.....XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX..
.....IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII..
.....JJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJ...
.....JJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJ...
|_|"%%$(){}+,-./0123456789;<=>?@ABCDEFGHIJKLMNPOQRSTUVWXYZ[\`_~'abcdefghijklmnopqrstuvwxyz{|}~-|_|
33          59      64       73              104             126
0.....26.....31.....40
-5.....0.....9.....40
0.....9.....40
3.....9.....40
0.2.....26.....31.....41

S - Sanger           Phred+33, raw reads typically (0, 40)
X - Solexa           Solexa+64, raw reads typically (-5, 40)
I - Illumina 1.3+    Phred+64, raw reads typically (0, 40)
J - Illumina 1.5+    Phred+64, raw reads typically (3, 40)
with 0=unused, 1=unused, 2=Read Segment Quality Control Indicator (hold)
(Note: See discussion above).
L - Illumina 1.8+    Phred+33, raw reads typically (0, 41)
```

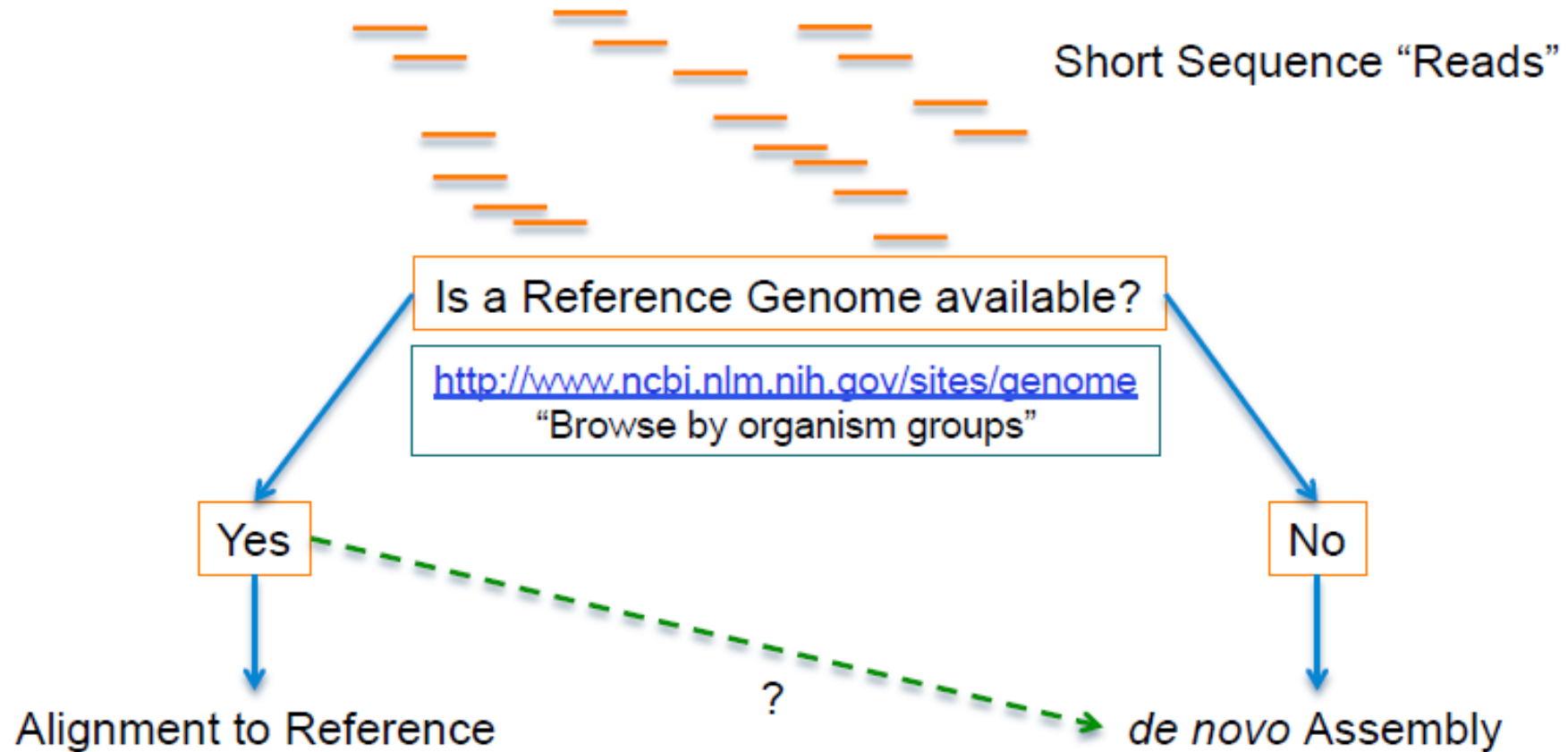
*Escherichia coli*

### Circular chromosome

## Plasmids



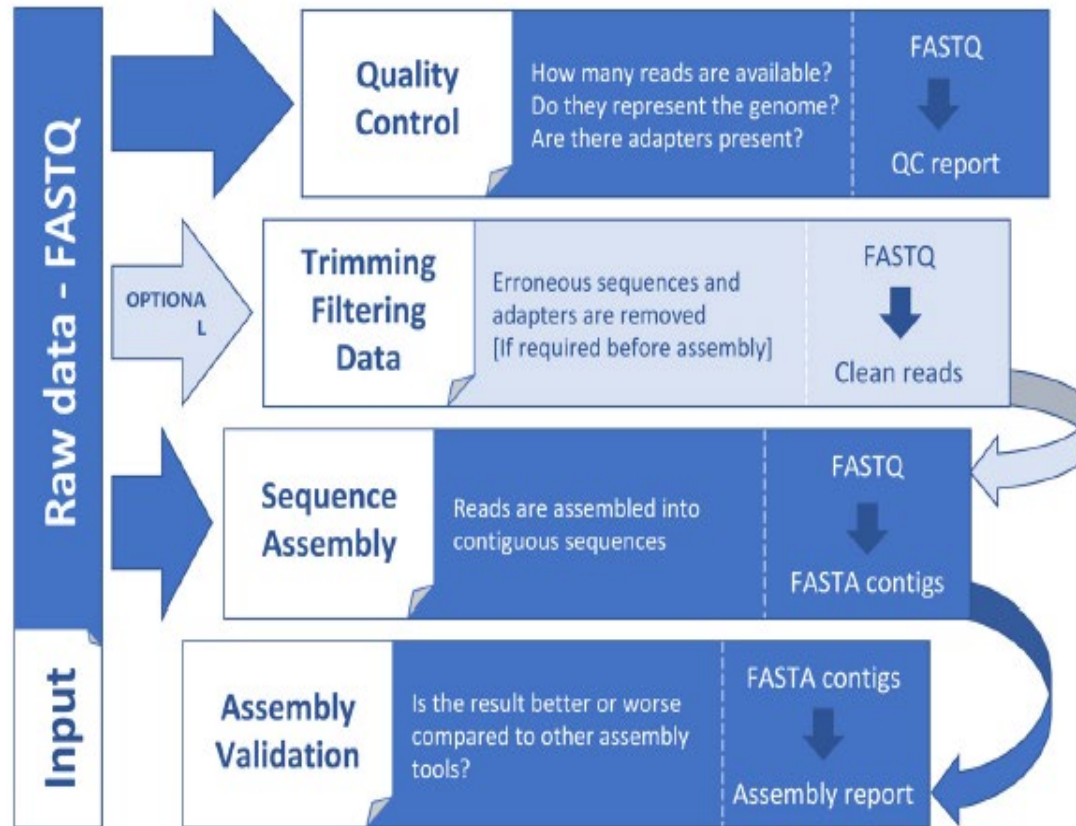
# Genome *de-novo* assembly or reference mapping



# Genome Assembly

Work flow to classic genome assembly strategies

Software used in this lecture



**fastQC**  
**NanoPlot**

**TrimGalore**

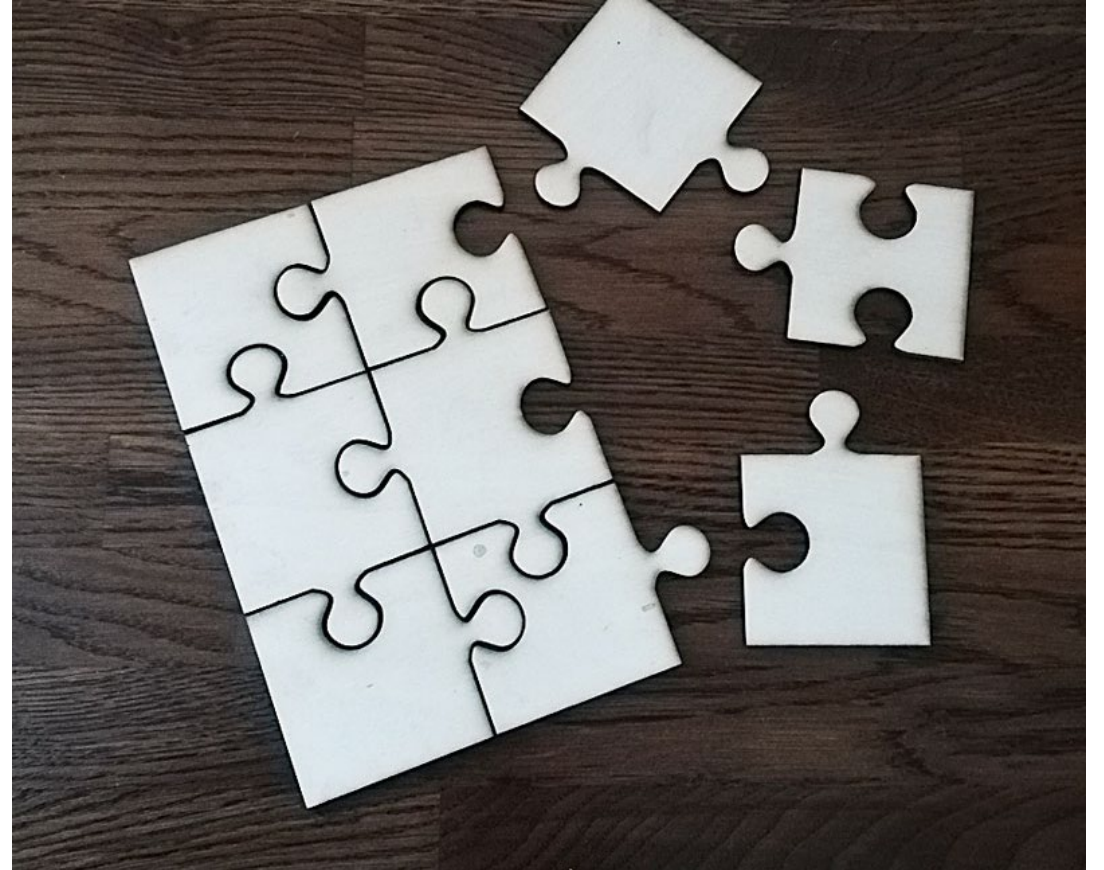
**SPADES**  
**Unicycler**

**BUSCO**

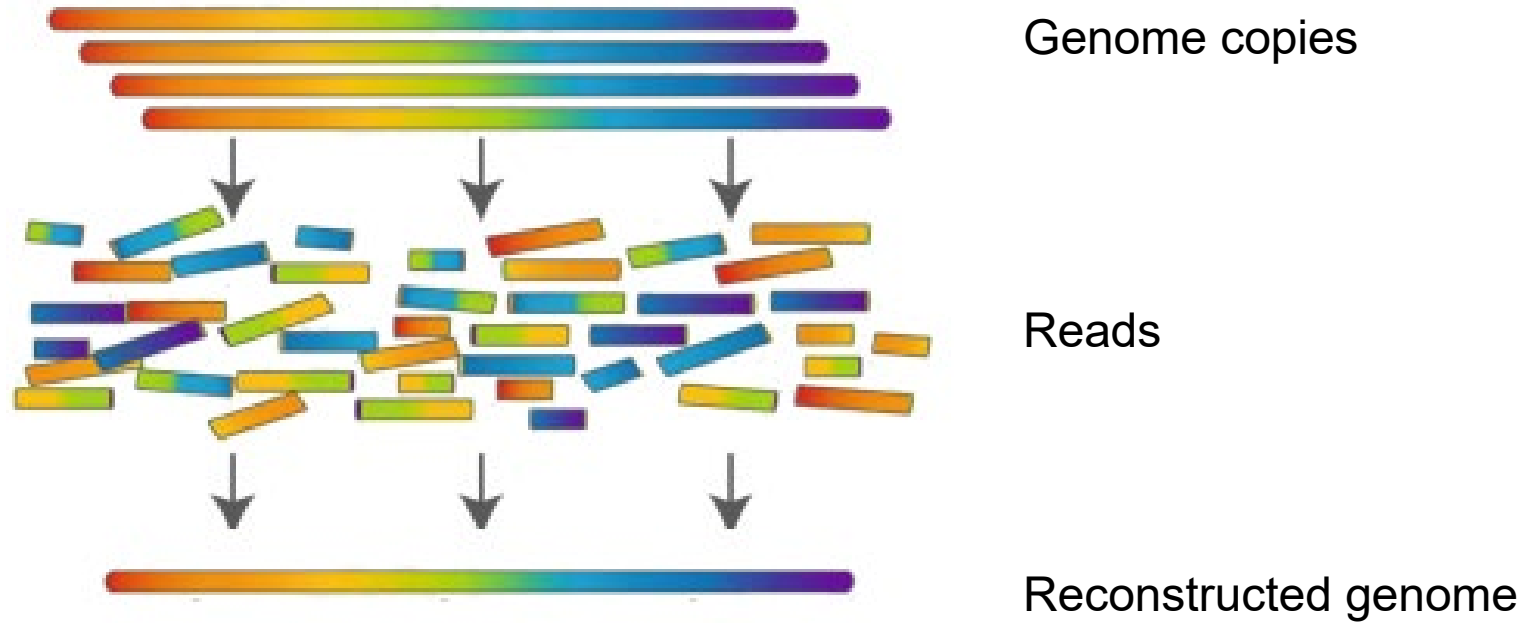
**Figure 2.** General steps in a genome assembly workflow. Input and output data are indicated for each step.

## Steps of the Assembly (recap):

1. Find all **overlaps** between reads
2. Build a **graph** (read connections)
3. **Simplify** the graph
4. Find a sensible path in the graph to generate a **consensus**



# Assembly - expectation



Picture adapted from:  
Commins, et al. (2009) Biological Procedures Online

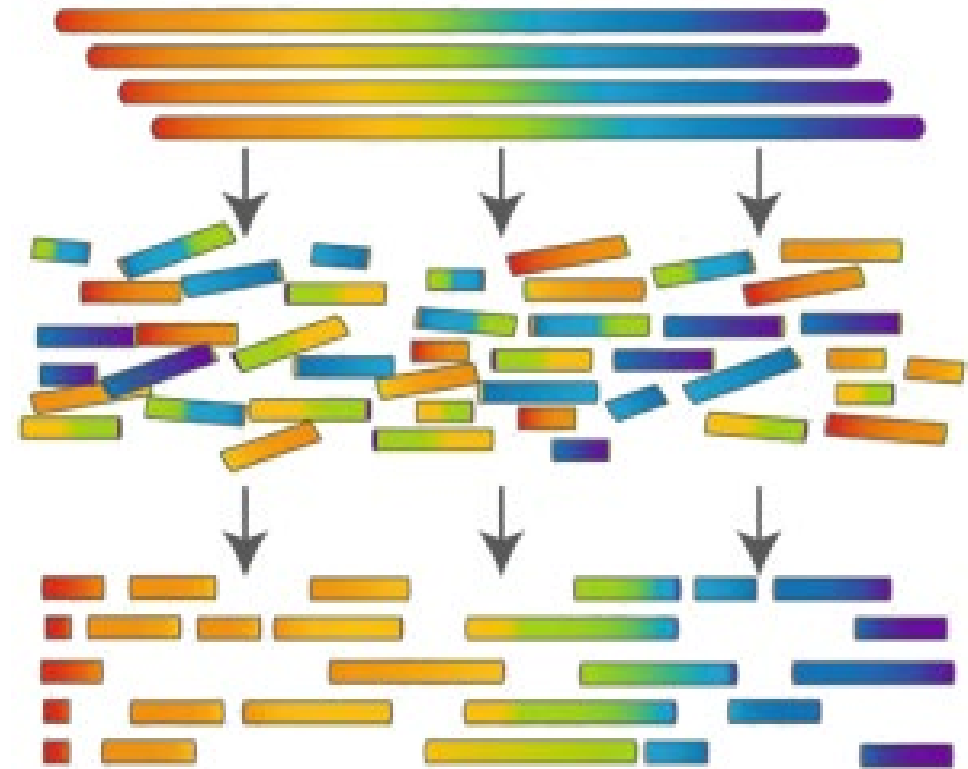
# Assembly - Reality

Especially true for short reads, such as those from Illumina sequencing (150 - 300 bp)

Genome copies

Reads

Contigs



Picture adapted from:  
Commins, et al. (2009) Biological Procedures Online



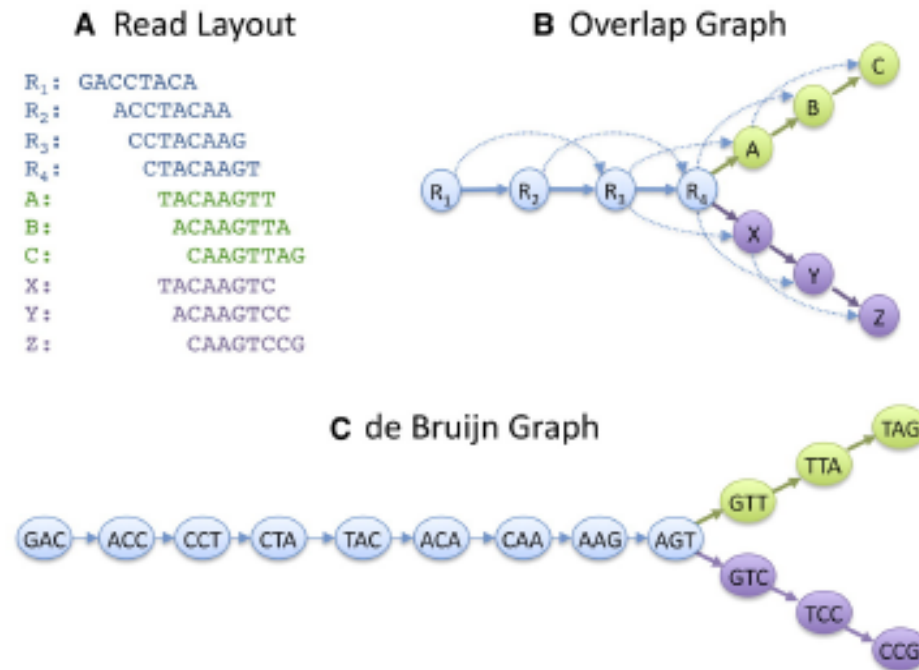
# Strategies to genome assembly

## ■ Algorithms

### 1. Greedy

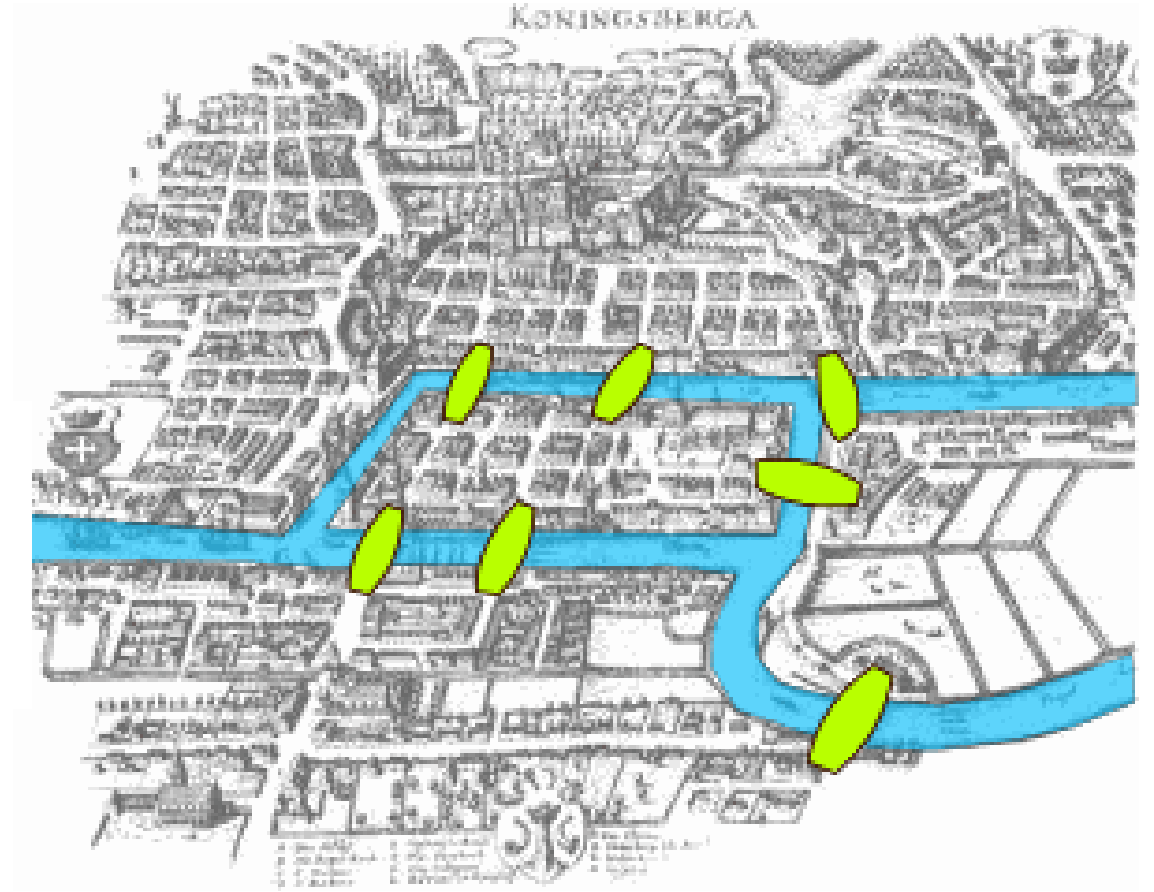
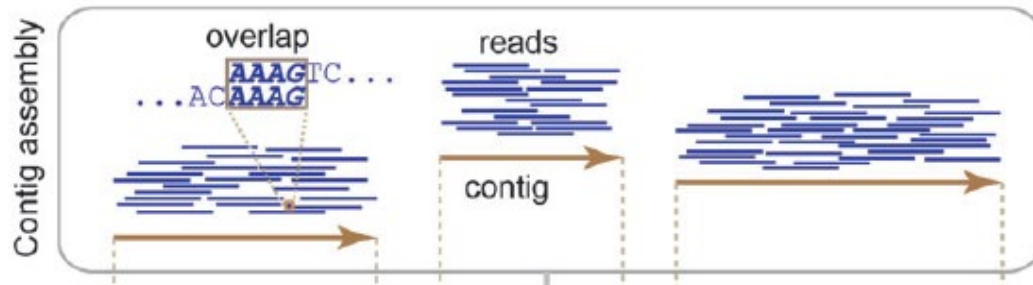
### 2. Overlap-layout-consensus (OLC)

### 3. De Bruijn Graph



# Steeps to genome assembly using De-Bruijn graph

The basic strategy for de novo assembly for short NGS reads comprises three steps: (i) contig assembly, (ii) scaffolding and (iii) gap filling.



Seven Bridges of Königsberg

# The K-mers: divide and conquer

- It breaks reads into **successive** k-mers and the graph maps the k-mers
- Each k-mer is a node and edges are drawn between each k-mer in a read.
- Repeat sequences create a fork in the graph; alternative sequences create a bubble.
- The k-mer size can only be determined by “trial and error”.
- A small value of K will create a complex graph but a large value of K may miss small overlaps. A good starting point would be a k-mer size that is  $\frac{2}{3}$  the size of the read
- Good for short reads or small genomes. With long reads and/or large genomes, may require lots of RAM (e.g., ~0.5 TB for human)

# K-mers

- k-mers are subsequences of length  $k$

For sequence **ATCG**;

$k=1$  is **A** – **T** – **C** – **G**

$k=2$  is **AT** – **TC** – **CG**

*Assemblers (and other bioinformatic tools) are often based on k-mers*

**k-mers for GTAGAGCTGT**

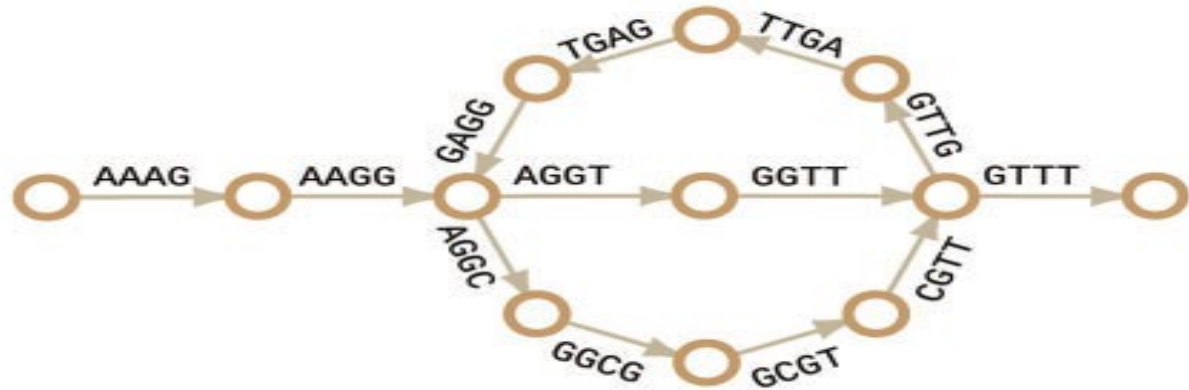
$k$	k-mers
1	G, T, A, G, A, G, C, T, G, T
2	GT, TA, AG, GA, AG, GC, CT, TG, GT
3	GTA, TAG, AGA, GAG, AGC, GCT, CTG, TGT
4	GTAG, TAGA, AGAG, GAGC, AGCT, GCTG, CTGT
5	GTAGA, TAGAG, AGAGC, GAGCT, AGCTG, GCTGT
6	GTAGAG, TAGAGC, AGAGCT, GAGCTG, AGCTGT
7	GTAGAGC, TAGAGCT, AGAGCTG, GAGCTGT
8	GTAGAGCT, TAGAGCTG, AGAGCTGT
9	GTAGAGCTG, TAGAGCTGT
10	GTAGAGCTGT

# The K-mers: divide and conquer

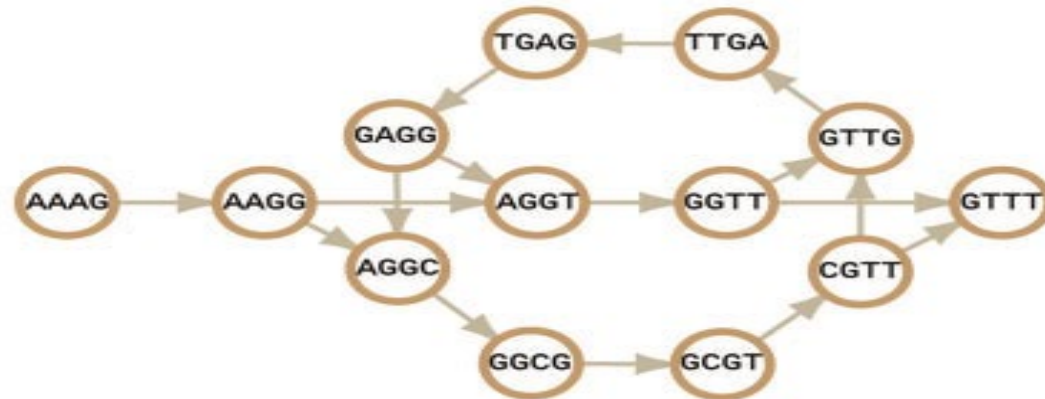
**A** Short read to  $k$ -mers ( $k=4$ )



**B** Eulerian de Bruijn graph



**C** Hamiltonian de Bruijn graph



# De Bruijn graphs – simplified

Sequence/read 1

**AAAGGCGTTGAG**

AAAG  
AAGG  
AGGC  
GGCG  
GCGT  
CGTT  
GTTG  
TTGA  
TGAG

Sequence/read 2

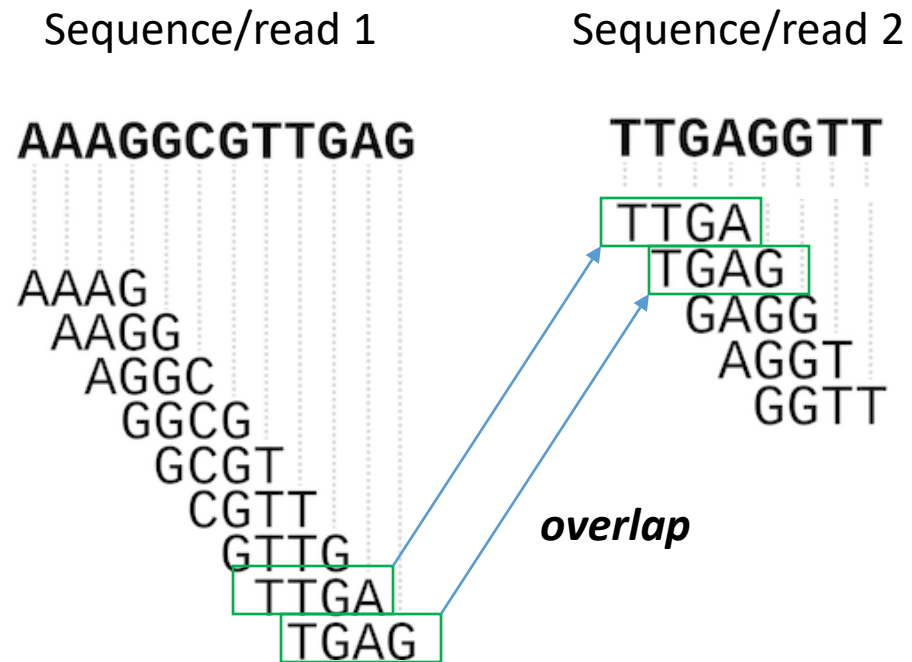
**TTGAGGTT**

TTGA  
TGAG  
GAGG  
AGGT  
GGTT

Let's choose a *k*-mer of 4  
 $K=4$

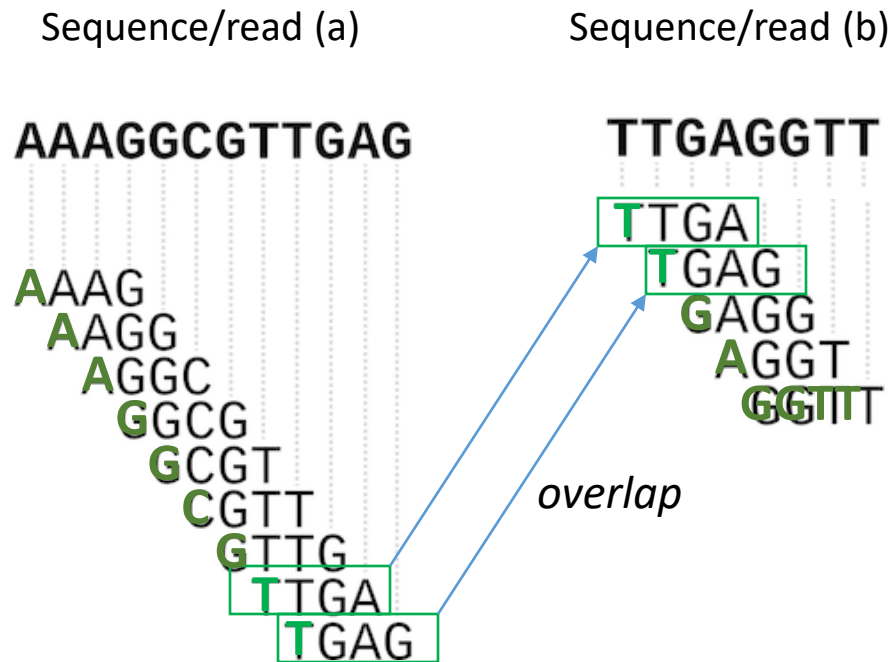
## De Bruijn graphs – simplified

$K=4$



# De Bruijn graphs – simplified

$K=4$



AAAGGCGTTGAGGTT

*Contig*

Assemblers usually use  
*k-mer values* > 31



# Evaluating the assembly

- ■ **Genome assembly results:**

- contig size and number of contigs produced
- scaffold size and number
- N50 and N90

- ■ **Coverage**

- ■ **GC Content**

- ■ **Genome annotation**

- repeats analysis and annotation
- protein-coding gene annotation (including gene structure prediction and gene function annotation)
- non-coding RNA gene annotation (including annotation of microRNA, tRNA, rRNA, and other ncRNA)
- transposon and tandem repeats annotation

- ■ **Comparative genomics and evolution (chromosome structure, conserved gene families)**

# Assembly evaluation

# Evaluating the assembly

## **Genome assembly results:**

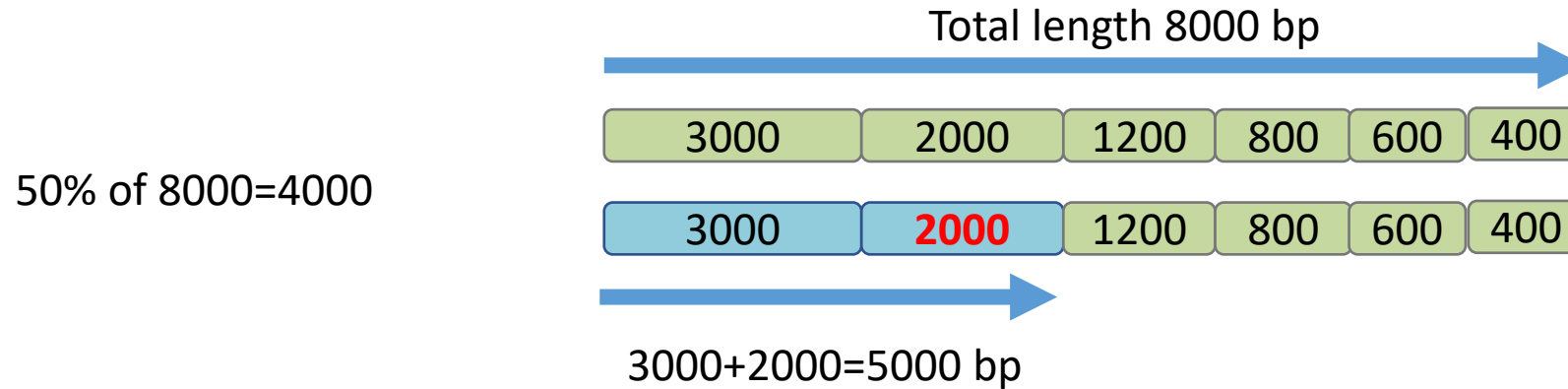
- **contig size and number of contigs produced**
- **N50 and N90**

## **Post – assembly quality check:**

- **Coverage**
- **GC Content**
- **Genome annotation** (including annotation of microRNA, tRNA, rRNA, and other ncRNA)

# Basic stats

**N50** the length of the shortest contig such that the sum of contigs of equal length or longer is at least 50% of the total length of all contigs.



**N90** = the length of the shortest contig such that the sum of contigs of equal length or longer is at least 90% of the total length of all contigs.

