FMT_matching_analysis.Rmd



R Markdown

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When you click the **Knit** button a document will be generated that includes both content as well as the output of any embedded R code chunks within the document. You can embed an R code chunk like this:

```
Hide

setwd(dirname(rstudioapi::getActiveDocumentContext()$path))
getwd()

[1] "C:/work/fmt_enterotype/a_microbiome/analysis"

Hide

source('./pre_processing.R')

[1] 0.01
[1] 0.01
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[1] 0.01
[1] 0.01
[1] 0.01
[1] 0.01
[1] 0.01
[1] 0.01
```

```
= donor1, = before1
          CDI IBD
 failure
           22 16
 response 90
, , = donor2, = before1
          CDI IBD
            9
 failure
 response 23
, = donor1, = before2
          CDI IBD
 failure
            8 15
 response 28
              24
, = donor2, = before2
          CDI IBD
 failure
            3 16
 response
            5
```

```
meta_config_in_this <- meta_fil_config1_na</pre>
```

Hide

```
#alpha
tmp_arare <- arare
rownames(tmp_arare) <- tmp_arare$X
# tmp_arare <- tmp_arare[rownames(before_donor_L6), c('X', 'shannon')]
cbind_arare <- cbind(tmp_arare[meta_config_in_this$Donor_sra, c('X', 'shannon')], tmp_arare[met
a_config_in_this$Previous_sra, c('X', 'shannon')])#
if(unique(cbind_arare[,1] == meta_config_in_this$Donor_sra & cbind_arare[,3] == meta_config_in_
this$Previous_sra) == c(TRUE)) {
    cn_arare = cbind(meta_config_in_this$SRA_Sample, cbind_arare, meta_config_in_this$postfmt_s
ymptoms, meta_config_in_this$pre_entro, meta_config_in_this$don_entro, meta_config_in_this$PRJ,
meta_config_in_this$Diease1)
    colnames(cn_arare) <- c('names', 'Dnames', 'Darare', 'Pnames', 'Parare', 'symptom', 'pre_en
tro', 'don_entro', 'PRJ', 'Diease1')
}</pre>
```

```
DP_distance <- NULL
len <- nrow(meta_config_in_this)
L6_rela_fil_sAg_others_remove <- noise.removal(L6_rela_fil_sAg_others, percent=0.01)</pre>
```

```
for(i in 1:len) {
    x1 <- as.numeric(as.character(L6_rela_fil_sAg_others_remove[, meta_config_in_this[i, "Donor
    _sra"]]))
    x2 <- as.numeric(as.character(L6_rela_fil_sAg_others_remove[, meta_config_in_this[i, "Previ
    ous_sra"]]))
    x3 <- as.numeric(as.character(L6_rela_fil_sAg_others_remove[, meta_config_in_this[i, "SRA_S
    ample"]]))
    DP_distance <- rbind(DP_distance, c(meta_config_in_this[i, "SRA_Sample"], vegdist(rbind(x1,
    x2), method='bray'), vegdist(rbind(x1, x3), method='bray'), vegdist(rbind(x2, x3), method='bra
    y')))
}

DP_distance_dat <- as.data.frame(DP_distance, stringsAsFactors = F)
colnames(DP_distance_dat) <- c('SRA_Sample', 'distance', 'DA_distance', 'PA_distance')</pre>
```

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```
if (unique(DP_distance_dat$SRA_Sample == (cn_arare$names)) == c(TRUE)) {
    DP_distance_dat_arare_div <- cbind(DP_distance_dat, cn_arare)
}

DP_distance_dat_arare_div$distance <- as. numeric(DP_distance_dat_arare_div$distance)

DP_distance_dat_arare_div$DA_distance <- as. numeric(DP_distance_dat_arare_div$DA_distance)

DP_distance_dat_arare_div$PA_distance <- as. numeric(DP_distance_dat_arare_div$PA_distance)</pre>
```

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```
DP_distance_dat_arare_div_b1 <- DP_distance_dat_arare_div[DP_distance_dat_arare_div$pre_entro % in% c('before1'),]
DP_distance_dat_arare_div_b2 <- DP_distance_dat_arare_div[DP_distance_dat_arare_div$pre_entro % in% c('before2'),]</pre>
```

```
test_pval<-function(test, symptoms, PRJ, dataframe) {
    fc_coin <- as.data.frame(cbind(as.numeric(test), (dataframe[, symptoms]), (dataframe[, PR
J])))
    colnames(fc_coin)<- c('fc', 'group', 'prj')

prj_list <- fc_coin$prj
    list <- NULL
    for(i in unique(prj_list)) {
        if (length(prj_list[prj_list %in% i]) > 2) {
            list <- c(list, i)
        }
    }
    fc_coin_f <- fc_coin[fc_coin$prj %in% list,]
    fc_coin_f$prj <- as.factor(as.character(fc_coin_f$prj))
    fc_coin_f$group <- as.factor(as.character(fc_coin_f$group))

pval<-wilcox_test(fc ~ group | prj, fc_coin_f)
    pval
}</pre>
```

```
##PA distance ~ response test_pval(DP_distance_dat_arare_div$PA_distance, 'symptom', 'PRJ', DP_distance_dat_arare_div)
```

```
Asymptotic Wilcoxon-Mann-Whitney Test

data: fc by group (1, 2)
    stratified by prj

Z = -2.0571, p-value = 0.03968
alternative hypothesis: true mu is not equal to 0
```

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test_pval(DP_distance_dat_arare_div[DP_distance_dat_arare_div\$pre_entro %in% c('before1'),]\$PA_
distance, 'symptom', 'PRJ', DP_distance_dat_arare_div[DP_distance_dat_arare_div\$pre_entro %in%
c('before1'),])

```
Asymptotic Wilcoxon-Mann-Whitney Test

data: fc by group (1, 2)
    stratified by prj

Z = -2.3808, p-value = 0.01727
alternative hypothesis: true mu is not equal to 0
```

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test_pval(DP_distance_dat_arare_div[DP_distance_dat_arare_div\$pre_entro %in% c('before2'),]\$PA_distance, 'symptom', 'PRJ', DP_distance_dat_arare_div[DP_distance_dat_arare_div\$pre_entro %in% c('before2'),])

```
Asymptotic Wilcoxon-Mann-Whitney Test

data: fc by group (1, 2)
   stratified by prj

Z = -0.92304, p-value = 0.356
alternative hypothesis: true mu is not equal to 0
```

##plot

```
library (ggpubr)
# beeswarm(log(DA_distance+1)~symptom, DP_distance_dat_arare_div)
DP\_distance\_dat\_arare\_div\_p1 \ \leftarrow \ DP\_distance\_dat\_arare\_div\\ [DP\_distance\_dat\_arare\_div\\ [pr\_distance\_dat\_arare\_div\\ [pr\_distance\_dat\_arare\_div] \ \ \%
in% c('before1'),]
DP_distance_dat_arare_div_pl$symptom <- ifelse(DP_distance_dat_arare_div_pl$symptom == 'respons
e', 'Response', 'Failure')
DP_distance_dat_arare_div_p1$symptom <- factor(DP_distance_dat_arare_div_p1$symptom, levels=c(
'Response', 'Failure'))
ggboxplot(DP\_distance\_dat\_arare\_div\_pl, x = 'symptom', y = ('DA\_distance'), fill='symptom', colline of the context of the co
or='black', alpha=0.5, size=0.8, width=0.45)+
        scale_y\_continuous(expand = c(0, 0.1)) +
        stat_compare_means(comparisons = list(c('Response', 'Failure')), method = 'wilcox.test', la
bel = "p. signif", label.y = 1.12, size=16)+
       labs(x = c(''), title = c(''), y = c('BC Distance')) + #Donor to recipient after FMT
  scale_fill_manual(name="FMT", values=c("#4D9127", "#90908D", donor_before_after_color, "#962E
2B", "#4E86C6", "#4D9127", "#90908D", 'lightgrey'))+#'#C77CFF', '#43AFC8',
    theme(text=element_text(family = "sans", size=32), plot.title = element_text(size=34, hjust =
0.5), axis.text = element_text(size=32, color ='dimgray'), axis.title.x = element_text(size=34
), axis.title.y = element_text(size=34), axis.ticks = element_blank())+
  theme (aspect.ratio = 0.95, legend.background=element_blank(), legend.position=c(1.75, 0.6)
                , panel.background = element_rect(fill = NA, colour = "lightgrey", size = 3)
               ,axis.line=element_line(colour=NA), axis.ticks.y = element_line(size=1.5, color ='dimgra
y'), axis.ticks.length = unit(7, "pt")
                ,legend.key = element_rect(fill = NA, color = NA))+
       guides(colour = guide legend(override.aes = list(size=5)))
```

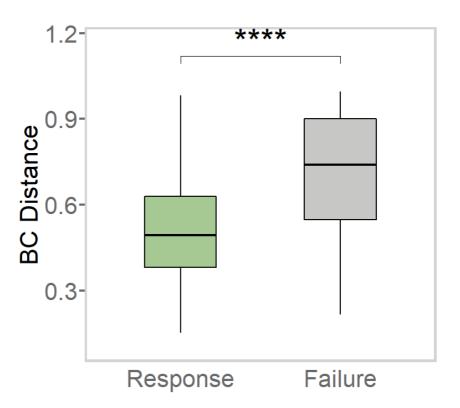


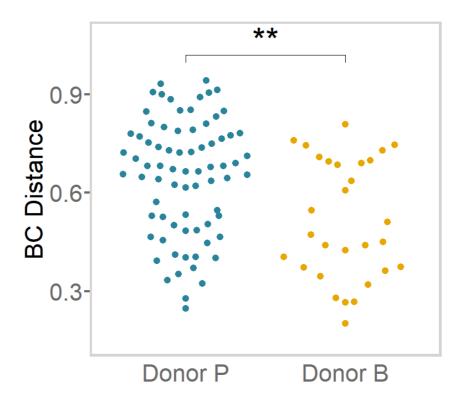
fig4i = 0
ggsave(paste("./figure4/4main_", fig4i, ".pdf", sep = ''), device = "pdf", useDingbats=FALSE)

Saving 12.9 x 8 in image

Hide

fig4i = fig4i + 1

```
##distance donor
DP_distance_dat_arare_div_p2 <- DP_distance_dat_arare_div[DP_distance_dat_arare_div$pre_entro %
in% c('before2'),]
DP distance dat arare div p2$don entro <- ifelse(DP distance dat arare div p2$don entro == 'don
orl', 'Donor P', 'Donor B')
DP_distance_dat_arare_div_p2$don_entro <- factor(DP_distance_dat_arare_div_p2$don_entro, levels
=c('Donor P', 'Donor B'))
cex=1.5
library (ggbeeswarm)
ggboxplot(DP_distance_dat_arare_div_p2, x='don_entro', y='distance', alpha = 0, color='white')+
    geom_quasirandom(aes(color=don_entro), method='smiley', size=4)+
   scale y continuous (expand = c(0, 0.1))+
   \# scale_alpha_continuous( range = c(0.6, 1))+
  stat_compare_means(comparisons = list(c('Donor P', 'Donor B')), method = 'wilcox.test', label
= "p. signif", label.y = 1.02, size=16)+
  labs(x= c(''), title=c(''), y= c('BC Distance'))+#Donor to recipient before FMT
    scale_color_manual(name="FMT", values=c('#28839B', '#E7A600', "#4D9127", "#90908D",
before_after_color, "#962E2B", "#4E86C6", "#4D9127", "#90908D", 'lightgrey'))+
  theme (text=element\_text (family = "sans", size=32), plot.title = element\_text (size=34, hjust = 1) \\
0.5), axis.text = element text(size=32, color = dimgray), axis.title.x = element text(size=34
), axis.title.y = element_text(size=34), axis.ticks = element_blank())+
 theme (aspect.ratio = 0.95, legend.background=element_blank(), legend.position=c(1.75, 0.6)
       , panel.background = element_rect(fill = NA, colour = "lightgrey", size = 3)
       ,axis.line=element_line(colour=NA), axis.ticks.y = element_line(size=1.5, color ='dimgra
y'), axis.ticks.length = unit(7, "pt")
       ,legend.key = element_rect(fill = NA, color = NA))+
   guides(colour = guide_legend(override.aes = list(size=5)))
```

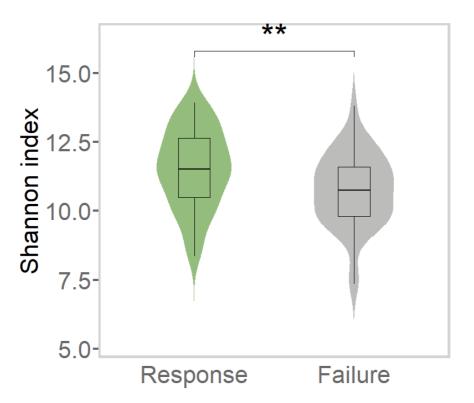


```
ggsave(paste("./figure4/4main_distance", fig4i, ".pdf", sep = ''), device = "pdf", useDingbats=
FALSE)
```

```
Saving 12.9 \times 8 in image
```

```
fig4i = fig4i + 1
```

```
DP_distance_dat_arare_div_p2 <- DP_distance_dat_arare_div[DP_distance_dat_arare_div$pre_entro %
in% c('before2'),]
DP_distance_dat_arare_div_p2$symptom <- ifelse(DP_distance_dat_arare_div_p2$symptom == 'respons
e', 'Response', 'Failure')
DP_distance_dat_arare_div_p2$symptom <- factor(DP_distance_dat_arare_div_p2$symptom, levels=c(
'Response', 'Failure'))
DP_distance_dat_arare_div_p2$c_arare <- DP_distance_dat_arare_div_p2$Darare + DP_distance_dat_a
rare div p2$Parare
cex=1.5
ggviolin(DP_distance_dat_arare_div_p2, x='symptom', y='c_arare', fill='symptom', alpha = 0.6, w
idth = 0.5, color='NA')+
    geom_boxplot(aes(x=symptom, y=c_arare), color='black', alpha=0, width=0.2, DP_distance_dat_
arare_div_p2)+
   scale_y_continuous(expand = c(0, 1)) +
   \# scale_alpha_continuous( range = c(0.6, 1))+
    stat_compare_means(comparisons = list(c('Response', 'Failure')), method = 'wilcox.test', la
bel = "p. signif", label. y = 15.8, size=16)+
scale_fill_manual(name="FMT", values=c("#4D9127", "#90908D", donor_before_after_color, "#962E
2B", "#4E86C6", "#4D9127", "#90908D", 'lightgrey'))+
  labs(x= c(''), title=c(''), y= c('Shannon index'))+#Donor + Recipient
  theme(text=element_text(family ="sans", size=32), plot.title = element_text(size=34, hjust =
0.5), axis.text = element text(size=32, color = dimgray), axis.title.x = element text(size=34
), axis.title.y = element text(size=34), axis.ticks = element blank())+
 theme (aspect.ratio = 0.95, legend.background=element_blank(), legend.position=c(1.75, 0.6)
       , panel.background = element_rect(fill = NA, colour = "lightgrey", size = 3)
       ,axis.line=element_line(colour=NA), axis.ticks.y = element_line(size=1.5, color ='dimgra
y'), axis.ticks.length = unit(7, "pt")
       , legend. key = element rect(fill = NA, color = NA))+
   guides(colour = guide_legend(override.aes = list(size=5)))
```



ggsave(paste("./figure4/4main_", fig4i, ".pdf", sep = ''), device = "pdf")

Saving 12.9 x 8 in image

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fig4i = fig4i + 1

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setwd(dirname(rstudioapi::getActiveDocumentContext()\$path))

```
classify_entero <- function(relatives_meta, a, medoids, b, c) {
    #'Previous_sra' pre_data.medoids pre_entro 'before'
    relatives_pre_data <- L6_abso[, unique(c(relatives_meta[, a], medoids))]
    sample_size <- colSums(relatives_pre_data)
    relatives_pre_data <- 1 * t(apply(relatives_pre_data, 1, function(x) {x/sample_size}))
    relatives_pre_data_remove = noise.removal(relatives_pre_data, percent=0.01)

relatives_pre_data.cluster= cluster_medoids.JSD(relatives_pre_data_remove, 0.000001, 2)

samples = dim(relatives_pre_data_remove)[2]
    relatives_meta[, b] <- paste(rep(c, length(1:(samples-2))), relatives_pre_data.cluster[1:(samples-2)], sep = '')
    relatives_meta
}</pre>
```

```
donor_engraft <- function(pre_entro) {</pre>
    # pre_entro <- "before2"</pre>
    tmp donor entro <- meta fil configl[meta fil configl$pre entro %in% c(pre entro), c('Donor
sra', 'don entro', 'PRJ')]
    tmp_donor_entro_u <- unique(tmp_donor_entro)</pre>
    L6_rela_fil_sAg_remove_simp_donor <- L6_rela_fil_sAg_remove_simp[, tmp_donor_entro_u$Donor_s
ral
    # cols <- ncol(feature_abun_dat)</pre>
    group <- as.character(tmp donor entro u$don entro)</pre>
    prj <- tmp_donor_entro_u$PRJ</pre>
    seq(0.1, 0.9, 0.05) \rightarrow quan
    tmp_pval <- apply(L6_rela_fil_sAg_remove_simp_donor, 1, function(x) {</pre>
        pt <- as.data.frame(cbind(x, group, prj))</pre>
        colnames(pt)<-c('nx', 'group', 'prj')</pre>
        # upt <- unique(pt)
        upt <- pt
        prj list <- upt$prj</pre>
        list <- NULL
         for(i in unique(prj_list)) {
             if (length(prj_list[prj_list %in% i]) > 2) {
                 list \leftarrow c(list, i)
        upt <- upt[upt$prj %in% list,]</pre>
        upt$nx <- as.numeric(as.character(upt$nx))</pre>
        tmp_test <- wilcox_test(nx ~ group | prj, upt)</pre>
        # pval a <- 1
        pval_a <- pvalue(tmp_test)</pre>
        if (is. na (pval_a)) {pval_a<-1}
        a_nx <- upt[upt$group %in% c('donor1'), 'nx']</pre>
        p_nx <- upt[upt$group %in% c('donor2'), 'nx']</pre>
        \# case \leftarrow quantile(log2(a_nx + 0.0001), quan)
        \# control \leftarrow quantile (log2(p nx+ 0.0001), quan)
        # gfc <- sum((case - control))/length(quan)
        gfc \leftarrow log2(mean(a_nx)/mean(p_nx))
        return(c(pval a, gfc))
    })
    ###select marked genus qvalue<0.05, and combine abundance
    tmp_pval_t <- data.frame(t(tmp_pval))</pre>
    colnames(tmp_pval_t) <- c('pval', 'gfc')</pre>
    # tmp_pval_t$id <- rownames(tmp_pval_t)
    # entero_diff_t$pval <- as.numeric(as.character(entero_diff_t$pval))
    tmp qvalue <- p.adjust(tmp pval t$pval, method='fdr')</pre>
    tmp_pval_adjust <- cbind(tmp_pval_t, tmp_qvalue)</pre>
    tmp_pval_adjust05 <- tmp_pval_adjust[tmp_pval_adjust$tmp_qvalue < 0.05,]#tmp_qvalue < 0.0
5,]
    tmp_L6_05 <- L6_rela_fil_sAg_remove_simp[rownames(tmp_pval_adjust05),]</pre>
    # tmp_L6_after_05 <- L6_rela_fil_sAg_remove_simp_after[rownames(tmp_pval_adjust05),]
```

```
tmp_pval_adjust05_nd \leftarrow as. data. frame(tmp_pval_adjust05_n)
    colnames(tmp_pval_adjust05_nd) <- c('genus', 'pval', 'gfc', 'qval')</pre>
    tmp_pval_adjust05_nd$genus <- factor(tmp_pval_adjust05_nd$genus, levels = rownames(tmp_pval
_adjust05_nd)[order(sign(tmp_pval_adjust05_nd$gfc)/tmp_pval_adjust05_nd$qval, decreasing = F)])
   min_y = 5
   p1 < -ggplot(tmp_pval_adjust05_nd, aes(x = genus)) + #, aes(x = genus), color = sex
      geom_linerange(data = tmp_pval_adjust05_nd, aes(ymin = ifelse(gfc > 0, -min_y, min_y), ym
ax = ifelse(gfc > 0, -min_y-gfc, min_y-gfc), color=ifelse(gfc < 0, "#E7A600", "#4E86C6")), size
= 6, alpha = 0.8)+
     # geom_linerange(data = plot_after_response[plot_after_response$response < plot_after_res
ponse$failure,], aes(ymin = min_y, ymax = ), size = 6, alpha = 0.8, color=)+ -min_y+log10(qval
ue), min_y-log10(qvalue)
     geom\ label\ (aes\ (x = genus, y = 0, label = genus, family = "sans"),
             inherit.aes = F, fontface = "italic",
            size = 6, label.padding = unit(0.0, "lines"), label.size = 0,
            label.r = unit(0.0, "lines"), fill = "NA", alpha = 0.9, color = "dimgrey")+
      scale_y = continuous(breaks = c(c(-10, -8, -6, -4, -2, 0) - min_y, c(0, 2, 4, 6, 8) + min_y),
                     labels = c("10", "8", "6", "4", "2", "0", "0", "2", "4", "6", "8")) +
     \# facet_wrap(^{\sim}genus, ncol = 2)+
      coord_flip()+
       labs(title="Marked genus in two enterotypes' donors (q<0.05)", x='', y="log2(FoldChang
e)", colour="Cluster")+
     # theme(plot.title = element_text(size=24, hjust = 0.5), axis.title.x = element_text(size
=20, hjust = 0.53),
         # title=element text(family = "sans", size=21),
           # text=element_text(family ="sans", size=21), aspect.ratio=0.95)+
      theme(text=element_text(family ="sans", size=24), plot.title = element_text(size=26, hjus
t = 0.5), axis.text = element_text(size=24, color ='dimgray'), axis.title.x = element_text(size
=0), axis.title.y = element_text(size=26), axis.ticks = element_blank())+
      theme (legend. position = c(4, .65), legend. background=element_rect(fill = NA), legend. text
= element_text(size=0))+
     scale_colour_manual(values=c("#4E86C6", "#E7A600", "#4D9127", "#90908D", "#7A1D1E", "#C47
737", "#E7A600"))+
      scale_alpha_manual(values = c(0.8)) +
      theme (panel. background = element rect(fill = NA, colour = "lightgrey", size = 3)
            , axis. line=element_line(colour="lightgrey")
           # ,axis.text.y = element_text(size=0, angle = 0)
           # ,axis.text.x = element_text(size=21, angle = 0)
            , axis. ticks = element_blank())
   sign_response <- sign(tmp_pval_adjust05$gfc)</pre>
   L6_rela_fil_sAg_remove_simp_weighted_don2 <- ifelse(sign_response<0, 1, 0) ** tmp_L6_05
   plot_marked_donor_before <- function(L6_rela_fil_sAg_remove_simp_weighted, flag){
        tmp_3column_before <- meta_fil_configl_na[meta_fil_configl_na$pre_entro %in% c(pre_entr
o) & meta_fil_configl_na$don_entro %in% ifelse(flag, 'donorl', 'donor2'), c('Previous_sra', 'SR
A_Sample', 'Donor_sra', 'postfmt_symptoms')]
       colnames(tmp_3column_before) <- c('Before_r', 'After_r', 'Donor_r', 'post')</pre>
        tmp_3column_before$Before <- ((L6_rela_fil_sAg_remove_simp_weighted[,tmp_3column_before
$Before_r]) / mean(L6_rela_fil_sAg_remove_simp_weighted[, tmp_3column_before$Donor_r]))
```

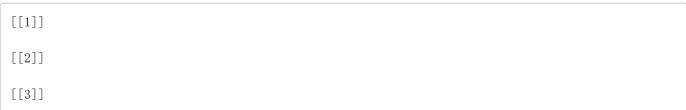
 $\label{tmp_pval_adjust05_n} $$ \leftarrow cbind(rownames(tmp_pval_adjust05), tmp_pval_adjust05)$$

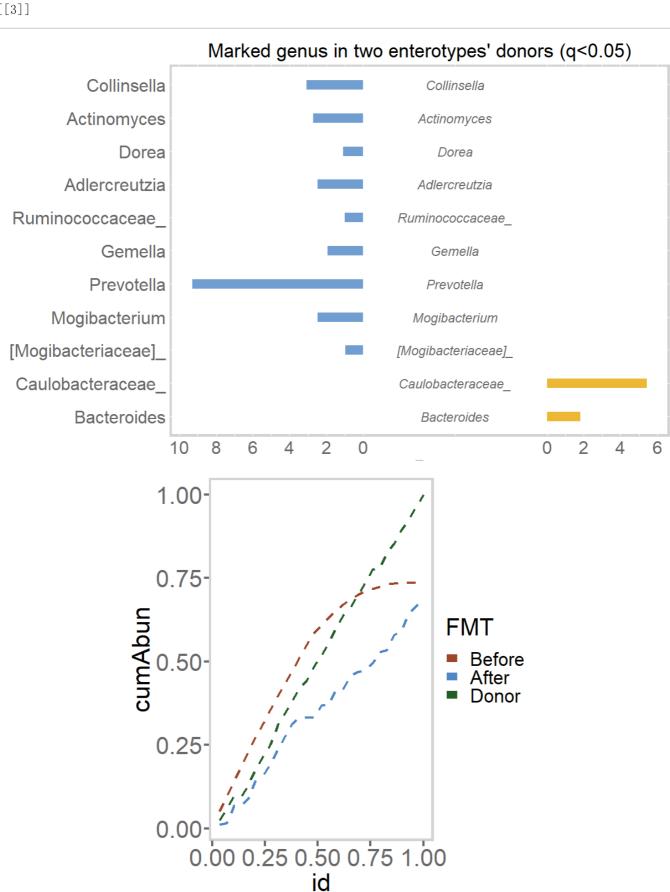
```
tmp_3column_before$After <- ((L6_rela_fil_sAg_remove_simp_weighted[,tmp_3column_before
$After_r]) / mean(L6_rela_fil_sAg_remove_simp_weighted[,tmp_3column_before$Donor_r]))
               tmp 3column before$Donor <- ((L6 rela fil sAg remove simp weighted[, tmp 3column before
$Donor_r]) / mean(L6_rela_fil_sAg_remove_simp_weighted[,tmp_3column_before$Donor_r]))
               samples num <- nrow((tmp 3column before))</pre>
               FMTstage_before <- melt(tmp_3column_before, measure.vars = c('Before', 'After', 'Donor'
))#
               FMTstage_before$variable <- factor(FMTstage_before$variable, levels = c('Before', 'Afte
r', 'Donor'))
               FMTstage_abun_before <- FMTstage_before</pre>
               FMTstage_abun_before$abun <- FMTstage_before$value#tmp_3column_before$After - tmp_3colu
mn before$Before #FMTstage before$value
               # FMTstage_abun_before <- cbind(FMTstage_before, L6_rela_fil_sAg_remove_simp_weighted[,
FMTstage_before$value])
               # colnames(FMTstage_abun_before) <- c(colnames(FMTstage_before), 'abun')</pre>
               FMTstage_abun_before$abun <- as.numeric(FMTstage_abun_before$abun/samples_num)
               FMTstage_abun_before$post <- factor(FMTstage_abun_before$post, levels = c('response',
'failure'))
               FMTstage_abun_before_1 <- FMTstage_abun_before[(order( -FMTstage_abun_before$abun)),]#F
MTstage_abun_before$post,
               order_1 <- unique(FMTstage_abun_before_1[FMTstage_abun_before_1$variable %in% c('Befor
e'), 'After_r'])
               FMTstage_abun_before_1$After_r <- factor(FMTstage_abun_before_1$After_r, levels = order
_1)
               FMTstage_abun_before_1 \( \) FmTstage_abun_before_1 \( \) FMTstage_abun_before_1 \( \) FMTstage_abun_before_1 \( \)
r),]
               # FMTstage_abun_before_1$After_r <- factor(FMTstage_abun_before_1$After_r, levels = uni
que())
               library(plyr)
               cumu <- ddply(FMTstage abun before 2, .(variable), transform, cumAbun = cumsum(abun))</pre>
               # FMTstage_abun_before$abun <- factor(FMTstage_abun_before$abun, levels=unique(FMTstage
_abun_before$abun))
               FMTstage abun before 1 <- cumu
               FMT stage\_abun\_before\_1\$id \ \leftarrow \ as.\ numeric\ (mapvalues\ (FMT stage\_abun\_before\_1\$After\_r,\ from\ 
 = order_1, to = 1:length(order_1)))/length(order_1)
               FMTstage abun before 1
           # FMTstage abun before$value <- as.numeric(FMTstage abun before$value)
       }
       p2 <- plot marked donor before (L6 rela fil sAg remove simp weighted don1, 1)
       p3 <- plot_marked_donor_before(L6_rela_fil_sAg_remove_simp_weighted_don2, 0)
       p4 < -ggp1ot() +
```

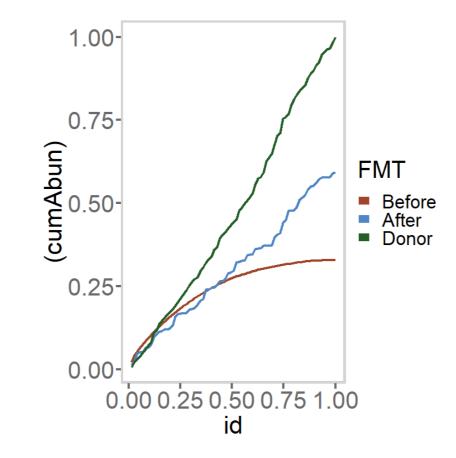
```
# geom_line(aes(x=id, y=(cumAbun), group=variable, color=variable), p2, linetype = 1, siz
e=1.5) +
      geom line(aes(x=id, y=cumAbun, group=variable, color=variable), p3, linetype = 2, size=1.
5)+
      scale_color_manual(name="FMT", values=c(donor_before_after_color, "#90908D", 'lightgrey'
))+
        theme(text=element_text(family ="sans", size=32), plot.title = element_text(size=34, hj
ust = 0.5), axis.text = element_text(size=32, color ='dimgray'), axis.title.x = element_text(si
ze=34), axis.title.y = element_text(size=34), axis.ticks = element_line(size=1.5, color ='dimgr
ay'), axis. ticks. length = unit(7, "pt"))+
 theme (aspect.ratio = 1.618, legend.background=element blank()#, legend.position=c(3.75, 0.6)
        , panel.background = element_rect(fill = NA, colour = "lightgrey", size = 3)
        , axis. line=element_line(colour="lightgrey")
        ,legend.key = element_rect(fill = NA, color = NA))+
    guides(colour = guide_legend(override.aes = list(size=5)))+theme(panel.grid.major=element_b
lank(), panel. grid. minor=element_blank());p4
   p5<-ggplot()+
      geom_line(aes(x=id, y=(cumAbun), group=variable, color=variable), p2, linetype = 1, size=
1.5) +
      # geom_line(aes(x=id, y=cumAbun, group=variable, color=variable), p3, linetype = 2, size=
1.5) +
      scale_color_manual(name="FMT", values=c(donor_before_after_color, "#90908D", 'lightgrey'
))+
        theme(text=element_text(family ="sans", size=32), plot.title = element_text(size=34, hj
ust = 0.5), axis.text = element_text(size=32, color ='dimgray'), axis.title.x = element_text(si
ze=34), axis.title.y = element_text(size=34), axis.ticks = element_line(size=1.5, color ='dimgr
ay'), axis. ticks. length = unit(7, "pt")+
  theme (aspect.ratio = 1.618, legend.background=element_blank()#, legend.position=c(3.75, 0.6)
        , panel.background = element_rect(fill = NA, colour = "lightgrey", size = 3)
        , axis. line=element_line(colour="lightgrey")
        ,legend.key = element_rect(fill = NA, color = NA))+
    guides(colour = guide_legend(override.aes = list(size=5)))+theme(panel.grid.major=element_b
lank(), panel. grid. minor=element_blank());p5
    if(dim(p1\$data)[1] > 0) \{list((p1), (p4), (p5))\}
}
```

```
# pdf('./figure4//fig4_engrament_donor_marked.pdf', width = 8, height = 8)
donor_engraft('before2')
```

The conditional covariance matrix has zero diagonal elementsThe conditional covariance matrix has zero diagonal elements







dev.off()

Hide

##enterotype change
table(meta_fil_configl\$after_entro, meta_fil_configl\$don_entro, meta_fil_configl\$postfmt_sympto
ms, meta_fil_configl\$pre_entro)

```
, , = failure, = before1
        donor1 donor2
          9
 after1
                  7
           16
 after2
 after3
           13
                  6
, , = response, = before 1
       donor1 donor2
 after1 48 7
 after2
           17
                 3
          33
 after3
                 21
, , = failure, = before2
       donor1 donor2
          7
 after1
          6
                  5
 after2
 after3 10
                 8
, , = response, = before2
        donor1 donor2
         18
 after1
 after2
          13
                  1
 after3 21
                  7
                                                                                   Hide
# table( meta_fil_configl_na$pre_entro, meta_fil_configl_na$after_entro, meta_fil_configl_na$do
n_entro)
dim(meta_fil_configl_na)
```

[1] 286 19

Hide

library("igraph") library(markovchain)

```
plot_MCC <- function(transElec, Ecolor) {</pre>
   mcPreg = new("markovchain", states = c("ET_E", "ET_B", "ET_P"), transitionMatrix = transEle
c)
   # plot(mcPreg, node.size = 10)
   netMC = markovchain:::.getNet(mcPreg, round = TRUE)
   wts = E(netMC) weight/100
    edge1 = get.edgelist(netMC)
    elcat = paste(edgel[,1], edgel[,2])
    elrev = paste(edgel[, 2], edgel[, 1])
    edge.curved = sapply(elcat, function(x) x %in% elrev)
    default.par = par(no.readonly = TRUE)
   plotMC = function(object, ...) {
      netMC = markovchain:::.getNet(object, round = TRUE)
      plot. igraph (x = netMC, ...)
   vert.sz = 40#*sapply(states(mcPreg), function(x) nrow(unique(sample_data(data_markov)[sampl
e_data(data_markov) $CST==x, "SubjectID"])))
   vert.sz = vert.sz #* 0.85
   vert.font.clrs = c("white", "white", "white", "white")
    edge. loop. angle = c(-1, 1, 0, 0, 3.14, 3.14)-1
    layout = matrix(c(0.4, 0.6, 0.4, 0.4, 0.45, 0.5), ncol=2, byrow=T) #0.6, 0.95, 0.43, 1, 0.3,
0.66
   # layout. show (n=3)
    edge. arrow. size=1.5
    edge.arrow.width= 2.5
    edge.width = ifelse(wts==1, 1, (15*wts + 0.5))
    edge. labels = as. character(round((E(netMC)$weight/100), 2))
    edge.labels[edge.labels==.98] = NA # labels only for self-loops
    plotMC (mcPreg,
           edge. arrow. size=edge. arrow. size, edge. arrow. width = edge. arrow. width,
           edge.label = edge.labels, edge.label.font=1, edge.label.cex=2, edge.label.color='bla
ck', edge.label.family = 'sans',
           ##
           edge.width=edge.width, edge.curved=edge.curved,edge.color=Ecolor,
           layout=layout, edge.loop.angle = edge.loop.angle,
           vertex. size=(vert. sz),
           vertex.label.font = 1, vertex.label.family = 'sans', vertex.label.cex = 2,
           vertex.label.color = vert.font.clrs, vertex.frame.color = NA, vertex.color = c("#7A1
D1E", "#C47737", '#4E86C6', '#E7A600'))
```

```
# donor enterotype vs after enterotype
#meta_fil_config_entro
# using distance to medoids
# don data.medoids pre data.medoids
after_enter <- (meta_fil_config1[, c('SRA_Sample', 'after_entro')])
medoids <- c(pre_data.medoids[1], don_data.medoids, pre_data.medoids[2])
a entero <- NULL
for(aj in 1:dim(after_enter)[1]) {
            a = after_enter[aj, ]
           method_dist = 'bray'
           E <- vegdist(rbind(L6_rela_fil_sAg_remove_simp[, as.character(medoids[1])], L6_rela_fil_sAg
_remove_simp[, as.character(a[1])]), method = method_dist)
            P \leftarrow vegdist(rbind(L6\_rela\_fil\_sAg\_remove\_simp[, as.character(medoids[2])], L6\_rela\_fil\_sAg_remove\_simp[, a
_remove_simp[, as.character(a[1])]), method = method_dist)
            B <- vegdist(rbind(L6_rela_fil_sAg_remove_simp[, as.character(medoids[3])], L6_rela_fil_sAg
_remove_simp[, as.character(a[1])]), method = method_dist)
           BB <- vegdist(rbind(L6_rela_fil_sAg_remove_simp[, as.character(medoids[4])], L6_rela_fil_sA
g_remove_simp[, as.character(a[1])]), method = method_dist)
            a_entero \langle -c(a_{entero}, ifelse(which.min(c(E, P, B))) == 4, 3, which.min(c(E, P, B))))
}
```

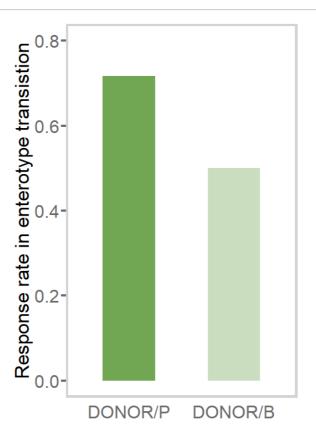
```
#1 E
#2 P
#3 B
entero_like <- cbind(a_entero, meta_fil_config1[, c('pre_entro', 'don_entro')])#, 'postfmt_sy
mptoms'
donor1_dat <- table(entero_like)[, , don_entro = 'donor1']

donor2_dat <- table(entero_like)[, , don_entro = 'donor2']
# donor2_dat[3,] = donor2_dat[3,] + donor2_dat[4,]
# donor2_dat[4,] = c(0, 0)
table(entero_like)</pre>
```

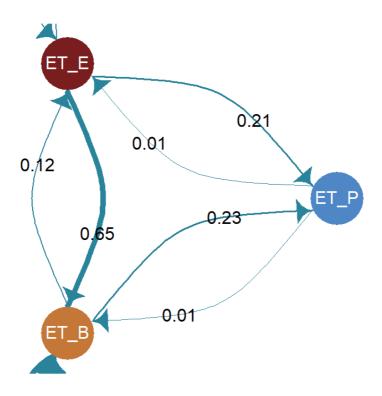
```
, , don_entro = donor1
        pre_entro
a entero beforel before2
               22
       1
                        10
       2
               98
                       56
       3
                       20
               31
, , don_entro = donor2
        pre_entro
a entero beforel before2
                8
                         2
       1
       2
               15
                       17
               30
       3
                       13
```

```
donor1\_tran \leftarrow c(t(apply(donor1\_dat, 1, function(x)\{(x / (colSums(donor1\_dat)))\})))
donor2\_tran \leftarrow c(t((apply(donor2\_dat, 1, function(x)\{(x / c(colSums(donor2\_dat)))\}))))
{\tt donor1\_tran}
[1] 0.1456954 0.6490066 0.2052980 0.1162791 0.6511628 0.2325581
                                                                                                     Hide
donor2\_tran
[1] \ \ 0.\ 1509434 \ \ 0.\ 2830189 \ \ 0.\ 5660377 \ \ 0.\ 0625000 \ \ 0.\ 5312500 \ \ 0.\ 4062500
                                                                                                     Hide
entero_like <- cbind(a_entero, meta_fil_configl[, c('don_entro','pre_entro', 'postfmt_symptom</pre>
s')])#
table(entero_like)
, , pre_entro = beforel, postfmt_symptoms = failure
        don entro
a_entero donor1 donor2
       1
            13
       2
             18
                      2
       3
             7
                      6
, , pre_entro = before2, postfmt_symptoms = failure
        don_entro
a_entero donor1 donor2
       1
       2
                     12
             13
       3
              8
                      6
, , pre_entro = before1, postfmt_symptoms = response
        don entro
a entero donorl donor2
              9
       1
                     1
       2
              71
                     10
             18
                     20
, , pre_entro = before2, postfmt_symptoms = response
        don_entro
a\_entero\ donor1\ donor2
       1
              8
                      1
       2
              33
                      3
       3
              11
                       6
```

```
plot\_change \leftarrow data.frame(don\_entro=c('DONOR/P', 'DONOR/B'), change=c(33/(33+13), 6/(6+6)))
plot_change$don_entro <- factor(plot_change$don_entro, levels = c('DONOR/P', 'DONOR/B'))</pre>
plot_change$change <- as.numeric(plot_change$change)</pre>
ggplot(plot_change, aes(x=don_entro, y=change))+#as.factor(change)
    geom_bar(stat="identity", width = 0.5, fill='#4D9127', alpha=c(0.8, 0.3))+#
        labs(x= c(''), y=c('Response rate in enterotype transistion'), title = c()+
        scale_y\_continuous(limits=c(0, 0.8))+
        scale\_fill\_manual(name="FMT", values=c("\#90908D", "\#4D9127", "\#90908D", "\#4D9127", donor\ bellemonth of the content of the c
fore_after_color, "#962E2B", "#4E86C6", "#4D9127", "#90908D", 'lightgrey'))+
  scale_color_manual(name="FMT", values=c("#90908D", "#4D9127", "#90908D", "#4D9127", donor befo
D", 'lightgrey'))+#'#C77CFF', '#43AFC8',
      # theme(text=element_text(family ="sans Neue", size=23), plot.title = element_text(size=26,
 hjust = 0.5), axis.title.x = element_text(size=24, vjust = -0.5, hjust = 0.55, color ='dimgra
y'), axis.text.x = element_text(size=0))+
    theme(text=element_text(family = "sans", size=24), plot.title = element_text(size=26, hjust =
0.5), axis.text = element_text(size=24, color = dimgray), axis.title.x = element_text(size=26
), axis.title.y = element_text(size=26), axis.ticks = element_blank())+
  theme (aspect.ratio = 1.62, legend.background=element_blank(), legend.position=c(1.75, 0.6)
              , panel.background = element_rect(fill = NA, colour = "lightgrey", size = 3)
              ,axis.line=element_line(colour=NA), axis.ticks.y = element_line(size=1.5, color ='dimgra
y'), axis.ticks.length = unit(7, "pt")
              ,legend.key = element_rect(fill = NA, color = NA), axis.ticks = element_blank(), panel.g
rid = element blank()) + #
      guides(colour = guide_legend(override.aes = list(size=5)))
```

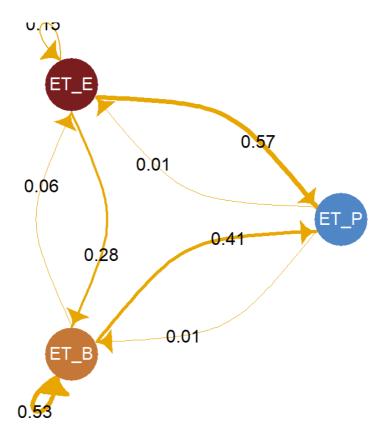


```
Saving 12.9 x 8 in image
```



Hide

plot_MCC(matrix(c(donor2_tran, 0.01, 0.01, .980), byrow = T, nrow = 3), '#E7A600')



dev.off()

Note that the echo = FALSE parameter was added to the code chunk to prevent printing of the R code that generated the plot.