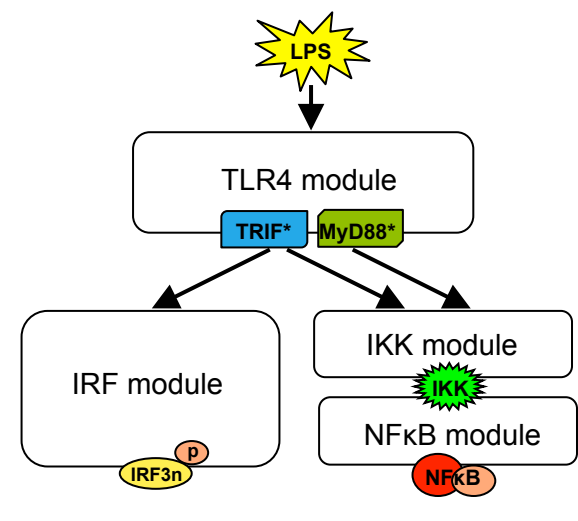
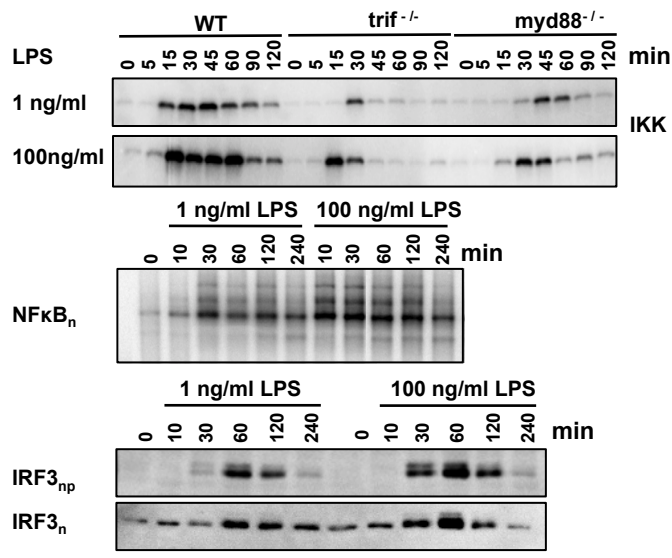
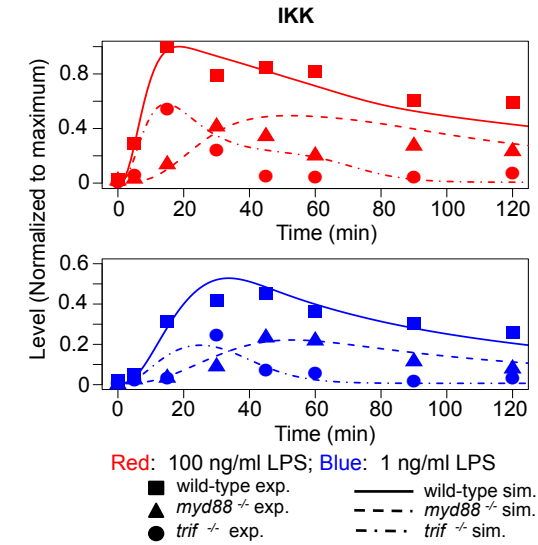
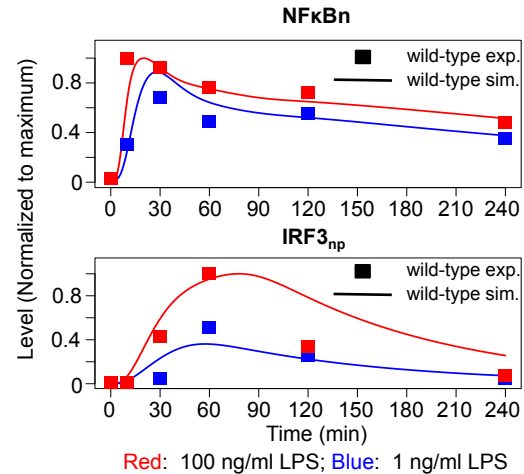
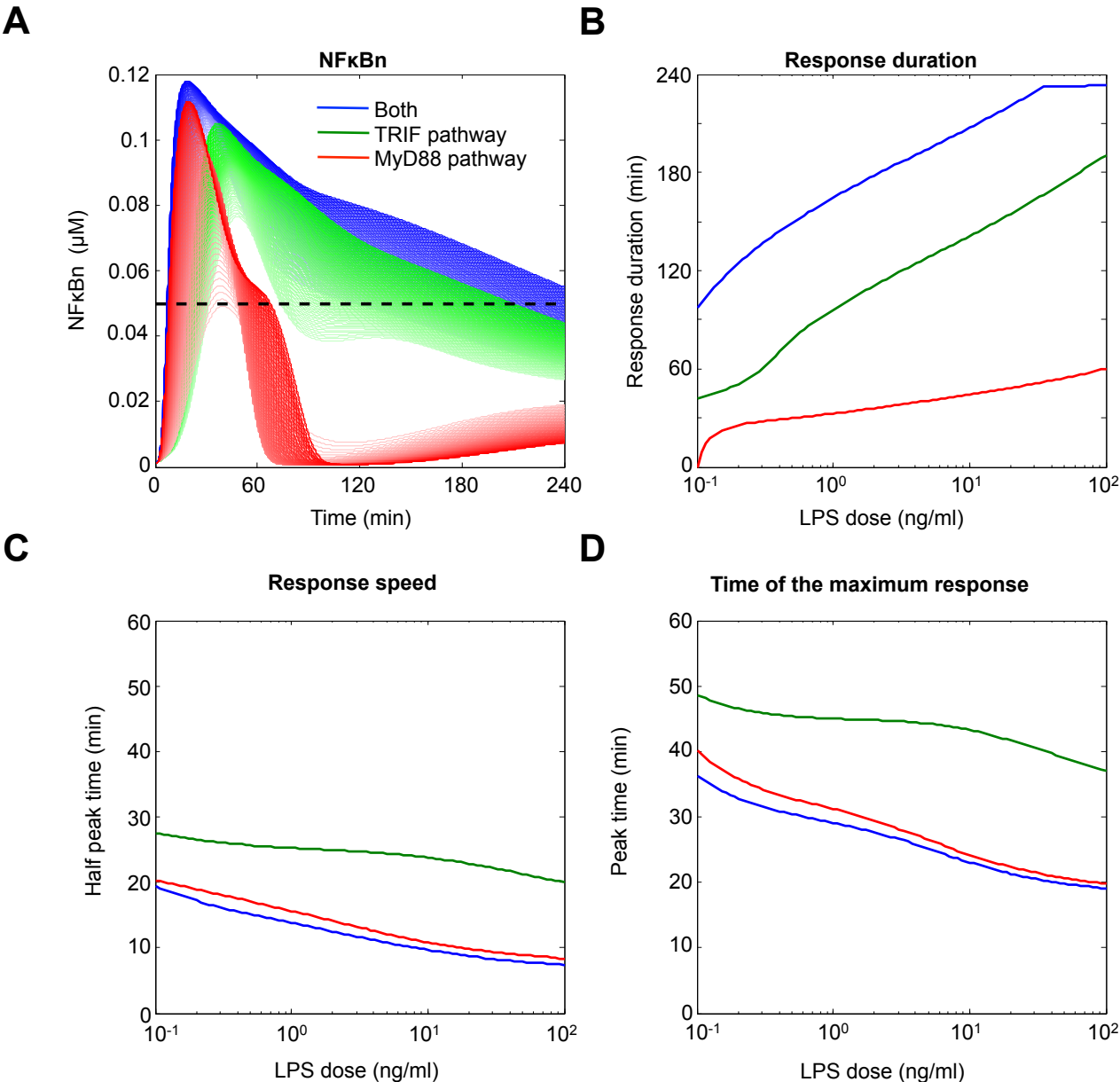
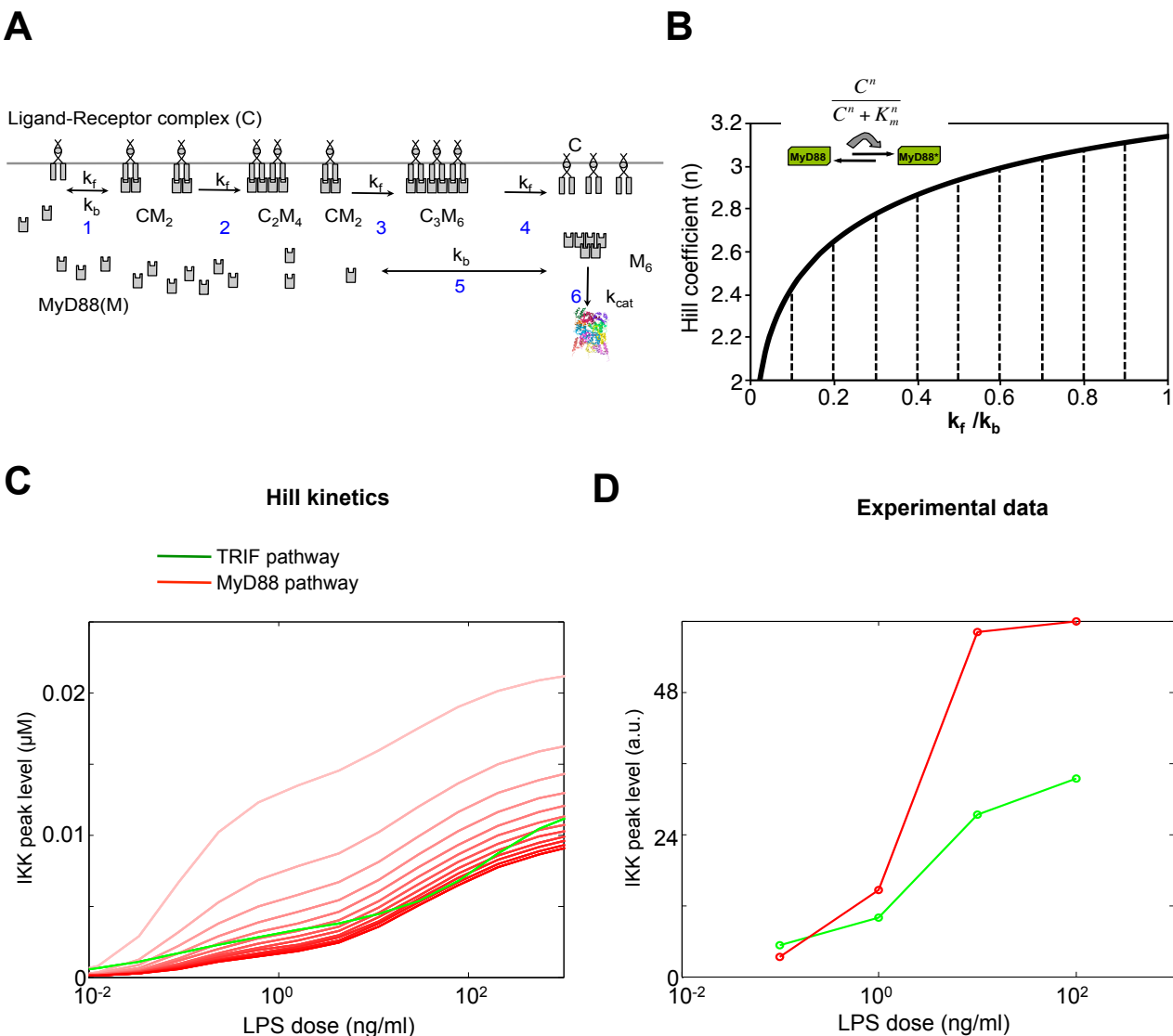


**A****B****C****D**

**Figure 1. Distinct dynamics in MyD88-dependent and TRIF-dependent pathway, in TLR4 signaling.** (A) The four modules of the model. (B) *Top*: The IKK kinase assay in 1ng/ml and 100ng/ml LPS stimulation for wt, *trif*<sup>-/-</sup> and *myd88*<sup>-/-</sup>. *Middle*: Nuclear NFκB activity measured by EMSA. *Bottom*: IRF3 activity measured by nuclear phosphorylation. (C-D) The model's simulation results against the data quantified from (B). “exp.” stands for experimental measurements; “sim.” stands for simulation result; “mko” stands for *myd88* condition; “tko” stands for *trif*<sup>-/-</sup> condition.

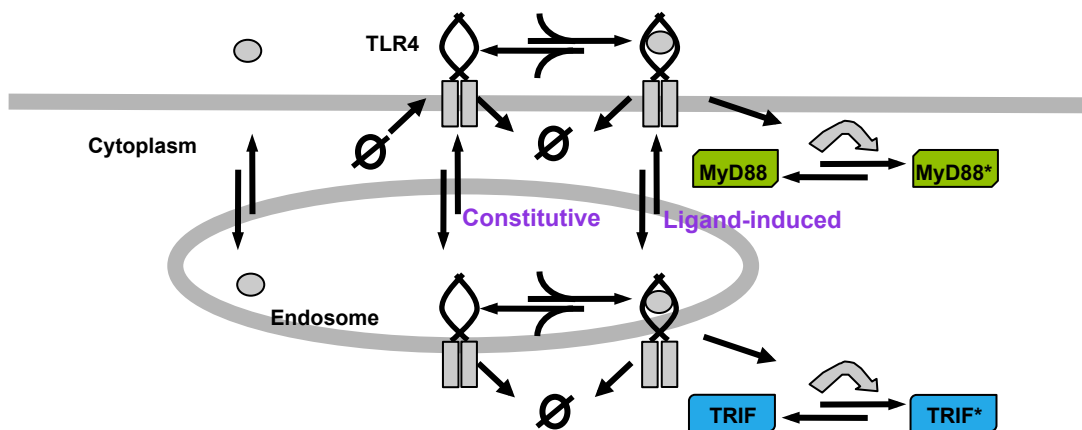
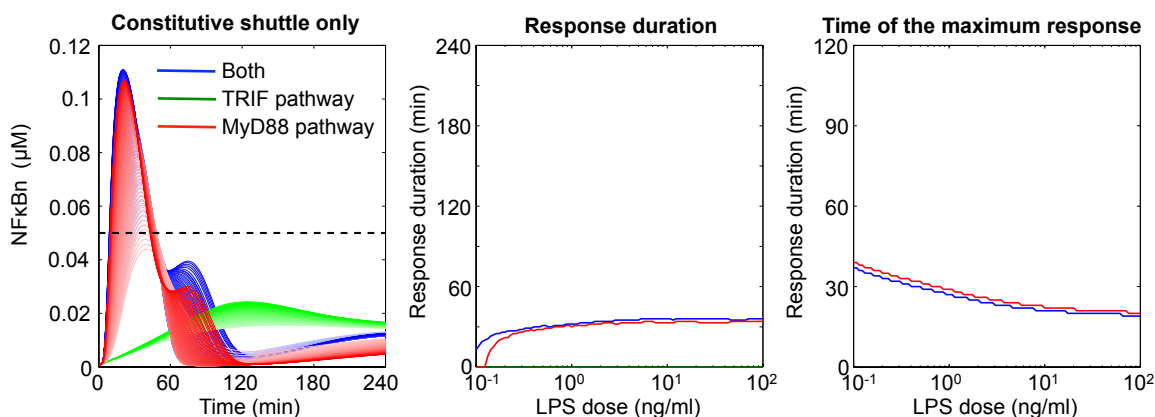
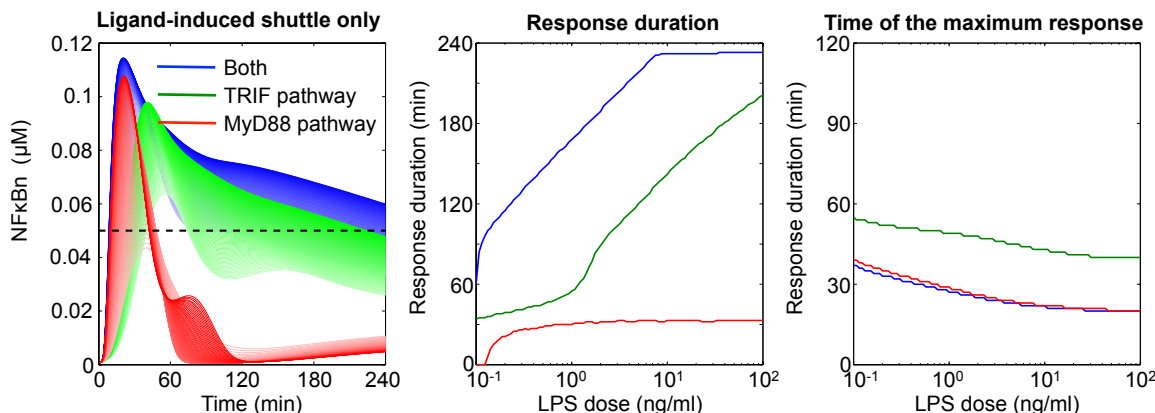


**Figure 2. Dynamics features in sub-pathways.** (A) The NF- $\kappa$ Bn time courses in wt (blue), myd88 ko (green) and trif ko (red) conditions, for LPS doses changing from 0.1 ng/ml to 100 ng/ml. (B) The NFκBn response duration (i.e. time when NFκBn > 50 nM) vs. the LPS doses. (C) The NFκBn response speed (defined by the time NFκBn level first reaches half of the peak level) vs. LPS doses. (D). The NF-κBn peak time vs. LPS doses.

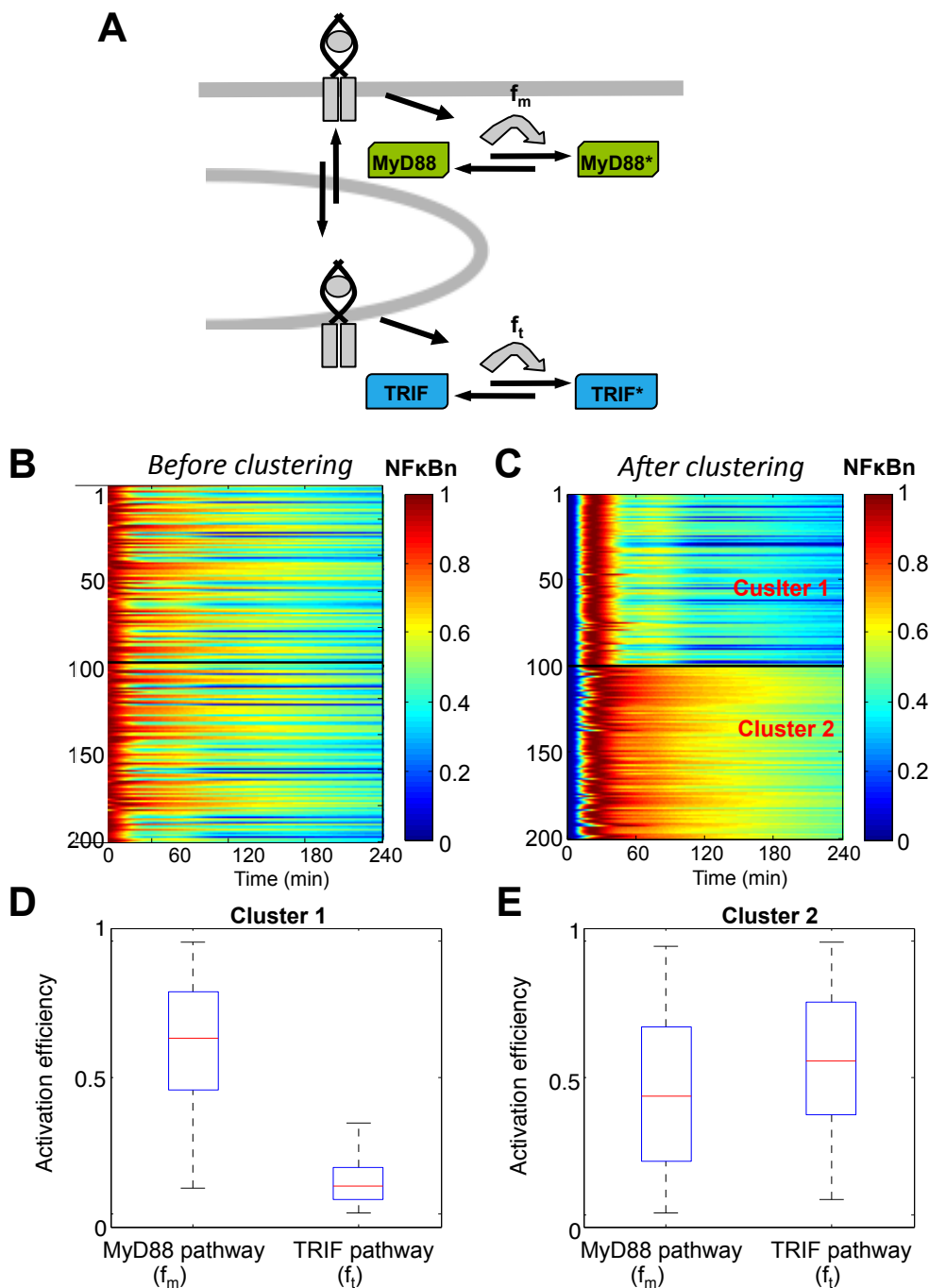


**Figure 3. Signalosome affects IKK dynamics.** (A) The MyDDosome assemble model. (B) The hill coefficient vs.  $k_f/k_b$ . (C) The IKK peak activity in TRIF and MyD88 knockouts vs. LPS concentration, predicted by model based on Hill kinetics with Hill coefficient from the range when  $k_f/k_b$  is from 0.1 to 1 in (B). (D) Quantification of the peak level from the experimental result in Fig. S4.

Figure 3

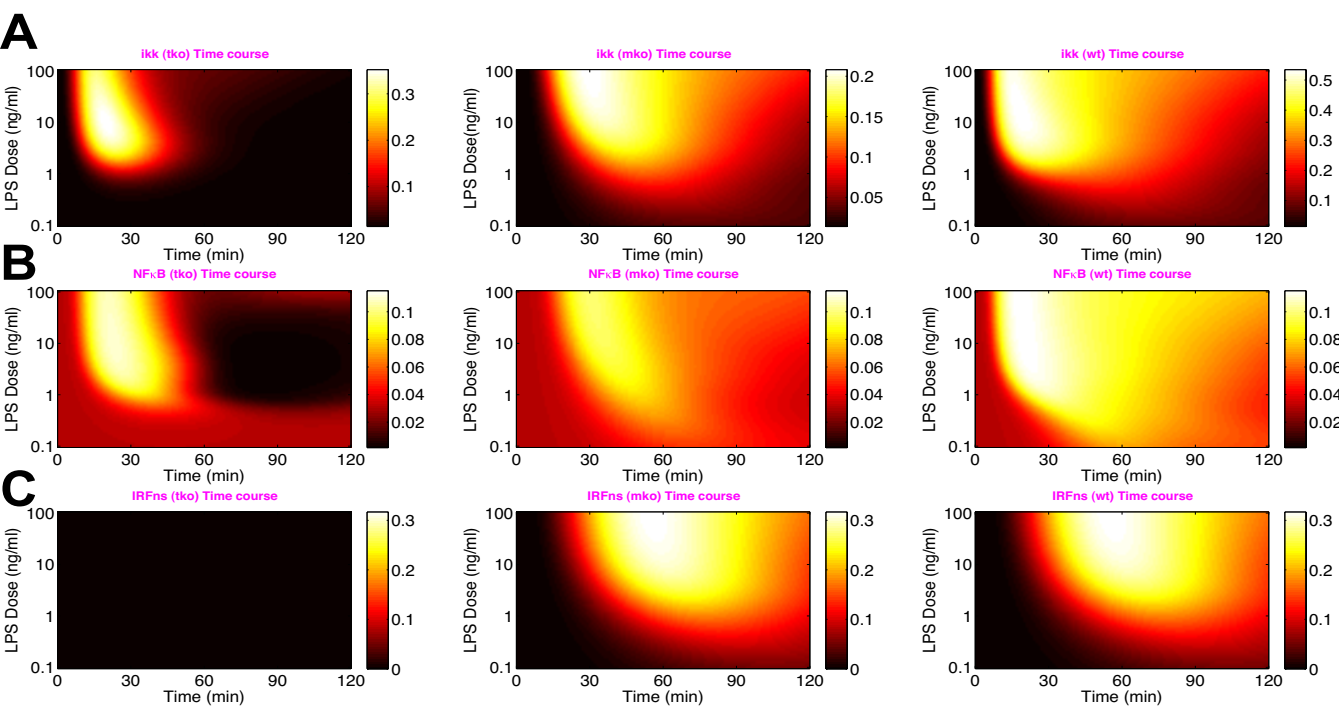
**A****B****C**

**Figure 4. The ligand induced-shuttling is responsible for the duration specificity.** (A) Two receptor shuttling processes, the constitutive shuttling and the ligand-induced shuttling, are labeled in the part of the model. (B-C) The NFκBn time courses (*left*), responses duration (middle) and peak time dose responses in constitutive shuttle only condition (B) and ligand-induced shuttle only condition (C).

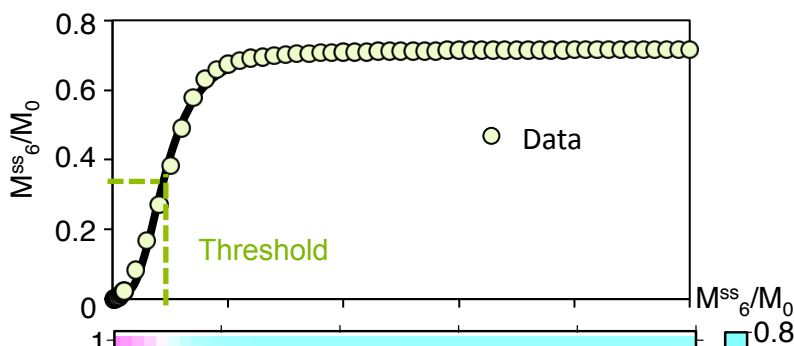
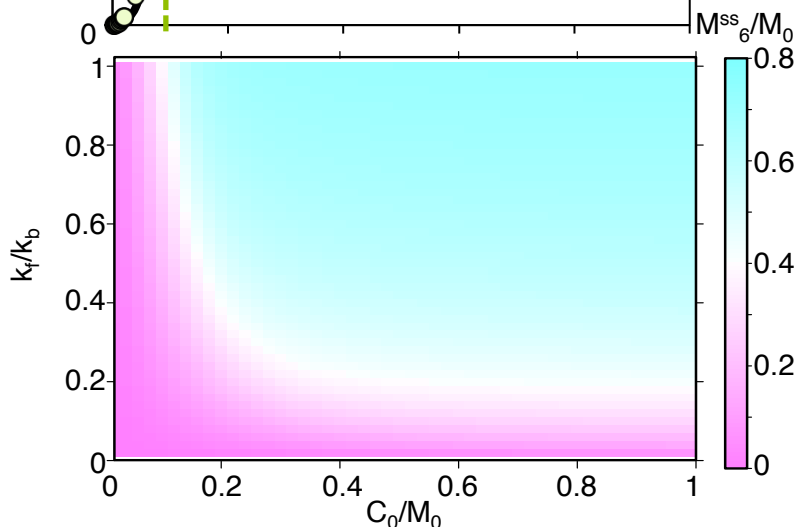


**Figure 5. Simulating two clusters of NFkB dynamics in single cell.** (A) Illustrate the two random fraction parameters in the model. 200 simulations of NF- $\kappa$ Bn dynamics before clustering (B) and after clustering (C). (D-E) The boxplot of the fraction parameters in these two clusters.



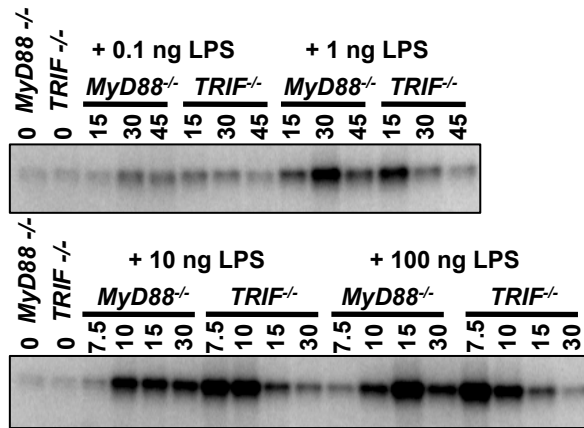


**Figure S2. Dose-responses predicted by the model for wt, *trif*<sup>-/-</sup>, and *myd88*<sup>-/-</sup>.** Simulated time-course dose response of IKK (A), nuclear NFκB (B) and IRF3 (C) activities in *trif*<sup>-/-</sup>, *myd88*<sup>-/-</sup> and wt conditions for LPS concentration ranging from 0.1 ng/ml to 100 ng/ml.

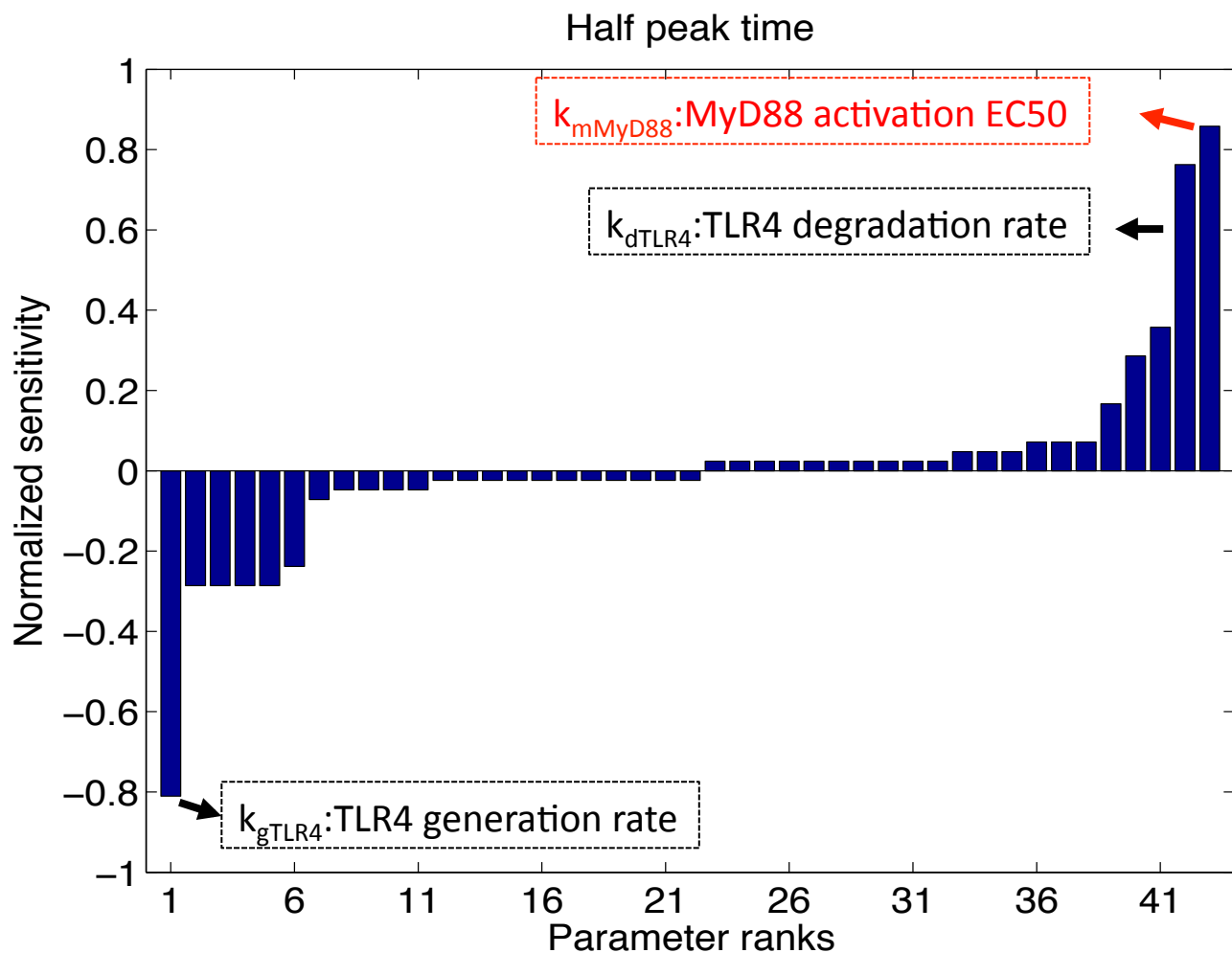
**A****B**

**Figure S3. The Myddosome formation model.** (A) The relative concentrations of  $M_6^{ss}/M_0$  versus the relative input concentration  $C_0/M_0$  in the upper left panel (dots). Parameters are  $k_f=1$ ,  $k_b=0.1$  and  $M_0=1$ . The relationship can be fitted by a Hill equation with Hill constant  $n = 3.0$  (solid line). (B) The dose-response of  $M_6^{ss}/M_0$  to  $C_0/M_0$ , by varying the fraction  $k_f/k_b$  by changing  $k_f$  only.

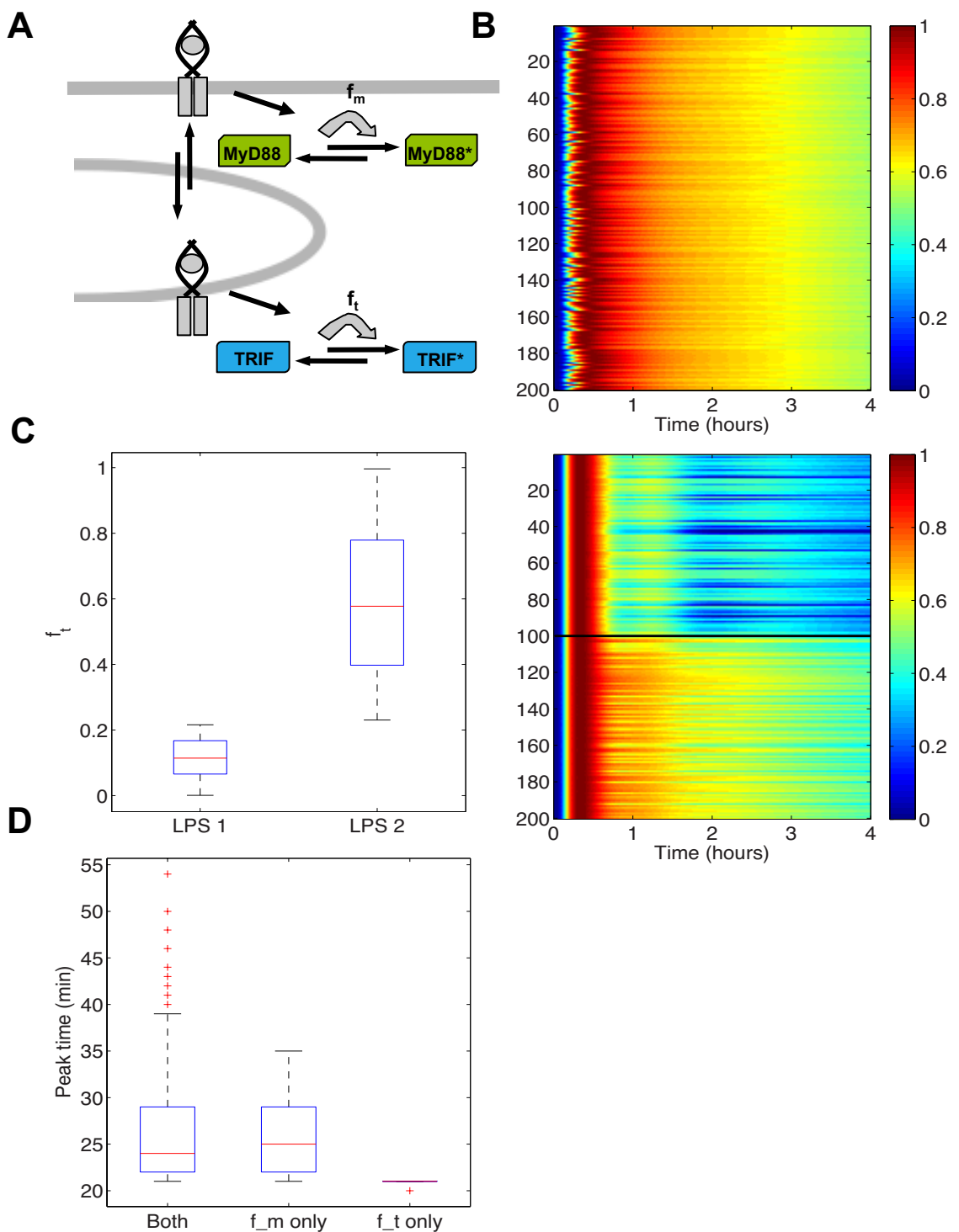




**Figure S4.** The measurements of IKK activity (*left*) and quantification of the peak level (*right*).



**Figure S5. Sensitivity analysis of the NFκB response time.** Only those parameters have non-zero sensitivity are plotted in this figure.



**Figure S6. Stochasticity in activation of TRIF-dependent pathway is responsible for the two clusters of LPS responses; Variability in MyD88-dependent pathway contributes to the heterogeneity in the peak response time.** Heat-map of the 200 simulations when randomized the fraction of activation in MyD88 activation (*B*) or TRIF activation (*C, right*). Boxplot of  $f_m$  in LPS1 and LPS2 is shown in (*C left*). (*D*) Compare the peak time distributions among the three conditions: 1) randomize both  $f_m$  and  $f_t$ , 2) randomize  $f_m$  only and 3) randomize  $f_t$  only.