PulsePAT-Nanostring

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1. Introduction

This is an Rmarkdown document which contains the R code for the Nanostring RNA-array analysis found within the manuscript "Ruiz et al. (2017)". Details about the tools and steps for processing the ileal transcriptome dataset can be found within the 'Methods' section of the respective manuscript. The files used to generate this data set is hosted on GEO under the accession number GSE98022. This workflow will use a saved RDA file from the analysis instead of hosting the raw data.

1a. Install and load the necessary libraries

```
source("scripts/geneExpress.R")
library(Biobase)
library(DESeq)
## Warning: package 'lattice' was built under R version 3.3.2
library(ggplot2)
## Warning: package 'ggplot2' was built under R version 3.3.2
library(dplyr)
library(RColorBrewer)
library(stats)
library(ggplot2)
library(pheatmap)
library(RColorBrewer)
library(org.Mm.eg.db)
## Warning: package 'AnnotationDbi' was built under R version 3.3.2
## Warning: package 'S4Vectors' was built under R version 3.3.2
1b. Import the eSet object containing the counts and metadata
```

Compare Control vs. PAT1 vs. PAT3 at day 52 of life

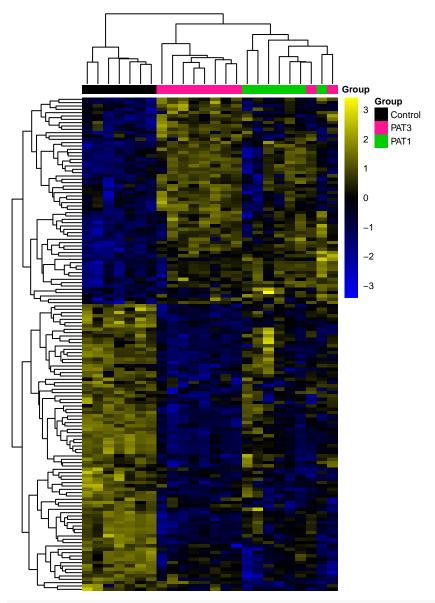
```
# Run significance testing function
## Output is an ExpressionSet, data.frame and significant-genes only data.frame
pulsePAT_results <- gene_express(expressionObject = pulsepat_eset)</pre>
```

Performing Significance Tests...

load("data/nanostring_eset.rda")

Figure 2a. Heatmap of significant genes between 3 groups

```
# Subset genes by custom significance and gather gene names
pulsePAT_results_filtered = subset(pulsePAT_results$sig_results,
                                   PAT1_Control_padj < 0.05 | PAT3_Control_padj < 0.05) %>%
 row.names()
# Subset output expresion object by only the significant genes
## make a matrix of values across all samples
mat <- pulsePAT_results$sig_expression[pulsePAT_results_filtered] %>%
  exprs()
# Choose which column variables you want to annotate the columns by.
annotation_col = data.frame(
 Group = factor(pulsePAT_results$sig_expression$Group),
 row.names = colnames(pulsePAT_results$sig_expression)
# Specify colors you want to annotate the columns by.
ann_colors = list(
 Group = c(Control = "black", PAT3 = "deeppink", PAT1 = "green3")
# Plot heatmap with annotations
pheatmap(mat = mat,
         color = colorRampPalette(c("#0000ff", "#000000", "#ffff00"))(n = 255),
         scale = "row",
         show_rownames = F,
         annotation_col = annotation_col,
         annotation_colors = ann_colors,
         fontsize = 6.5,
         cellwidth = 8,
         cellheight = 2.5,
         show_colnames = F)
```



sessionInfo()

```
## R version 3.3.1 (2016-06-21)
## Platform: x86_64-apple-darwin13.4.0 (64-bit)
## Running under: OS X 10.10.5 (Yosemite)
##
## locale:
## [1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/c/en_US.UTF-8/en_US.UTF-8
##
## attached base packages:
## [1] stats4
                parallel stats
                                   graphics grDevices utils
                                                                  datasets
## [8] methods
                base
##
## other attached packages:
## [1] org.Mm.eg.db_3.4.0
                            AnnotationDbi_1.36.2 IRanges_2.8.1
## [4] S4Vectors_0.12.1
                            pheatmap_1.0.8
                                                 RColorBrewer_1.1-2
## [7] dplyr_0.5.0
                            ggplot2_2.2.1
                                                 DESeq_1.26.0
```

```
## [10] lattice_0.20-35
                             locfit_1.5-9.1
                                                  Biobase_2.34.0
## [13] BiocGenerics_0.20.0
## loaded via a namespace (and not attached):
                           plyr_1.8.4
                                              bitops_1.0-6
## [1] Rcpp_0.12.10
## [4] tools_3.3.1
                           digest_0.6.12
                                              tibble_1.3.0
## [7] annotate_1.52.1
                           evaluate_0.10
                                              RSQLite_1.1-2
## [10] memoise_1.1.0
                           gtable_0.2.0
                                              Matrix_1.2-8
## [13] DBI_0.6-1
                           yaml_2.1.14
                                              genefilter_1.56.0
## [16] stringr_1.2.0
                           knitr_1.15.1
                                              rprojroot_1.2
## [19] grid_3.3.1
                           R6_2.2.0
                                              XML_3.98-1.6
## [22] survival_2.41-3
                           rmarkdown_1.5
                                              geneplotter_1.52.0
## [25] magrittr_1.5
                           codetools_0.2-15
                                              backports_1.0.5
                           htmltools_0.3.5
## [28] scales_0.4.1
                                              splines_3.3.1
## [31] assertthat_0.2.0
                           colorspace_1.3-2
                                              xtable_1.8-2
                                              RCurl_1.95-4.8
## [34] stringi_1.1.5
                           lazyeval_0.2.0
## [37] munsell_0.4.3
```