

MIMARgof, perlgof and perlgofest - programs to test the goodness of fit of the model estimated by MIMAR

C. Becquet

December 17, 2010

This document describes how to use the Perl scripts `perlgofest` and `perlgof`, which call the program `MIMARgof` to generate samples under an “isolation-migration” model with either point estimates of the parameters provided by `MIMAR`, or sets of parameters sampled from the posterior distribution estimated by `MIMAR` (see Figure 1 and section “List of parameters and symbols”). These scripts output the distributions of summary statistics for the simulated samples (see section “The **standard output**”). These distributions allow one to perform a goodness of fit test. You can represent the distribution of a statistic graphically and consider visually whether the observed value of the statistic (i.e. computed for the data set used to perform the estimation by `MIMAR`) falls within the distribution. If the observed value of the statistic is within the range of the distribution, the estimated model fit the data for this statistic. Alternatively, you can quantify the probability of observing an observed statistic given the simulated distribution of the statistic. If this “p-value” is smaller than 0.05, the goodness of fit test is rejected: the estimated model does not fit the data for this statistic.

Below, I first describe how to run `MIMARgof`, which generates samples under the isolation-migration model. In a second step, I describe how to run `perlgofest`, which allows to test the goodness of fit for an estimated model using point estimates of the parameters. Finally, I describe how to run `perlgof`, which allows to test the goodness of fit of the estimated model by sampling sets of parameters from the posterior distribution estimated by `MIMAR`. Since this goodness of fit test takes into account the uncertainties associated with the estimates, it is equivalent to the Bayesian posterior predictive p-value (e.g., Meng, 1994).

The program and scripts are intended to run on Unix, or Unix-like operating systems, such as Linux or MacOSX. The next section describes how to download the relevant files and compile the program. The subsequent sections described how to run the scripts and perform a goodness of fit test.

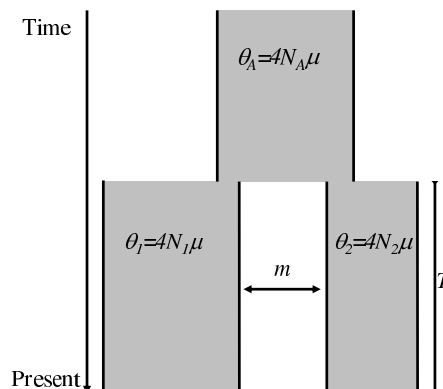


Figure 1: **The “isolation-migration” model**, in which two populations diverged T generations ago from a common ancestral population. The parameters θ_1 , θ_2 and θ_A , are the population mutation rates for populations 1, 2 and the ancestral population, respectively. μ is the mutation rate per bp and N_1 , N_2 and N_A are the diploid effective sizes of the first, second and ancestral populations, respectively. The split time in generations is T , m is the symmetrical migration rate between populations per generation such that, $M = 4N_1m$ is the expected number of individuals in population 2 replaced by migrants from population 1 each generation.

Contents

What changed since the last version?	3
Running MIMARgof	4
The input file	4
The basic command line	4
Providing the other parameters of the model	5
The standard output	5
More complex models	6
Crossing over	6
Asymmetrical migration rates	7
Other options conserved from ms. Use at your own peril!	7
Summary of command line options	7
Performing a goodness of fit test using point estimates	8
Setting perlgofest	8
The output file of perlgofest	9
Performing a goodness of fit test by sampling from the estimated posterior distribution	10
Setting perlgof	10
The output file of perlgof	11
Does the estimated model fit the data?	11
Setting "testGOF.R"	11
The output of "testGOF.R"	12
If the ancestral alleles are unknown use MIMARgof_noanc	13
Compiling MIMARgof_noanc	13
List of parameters and symbols	13
List of files in the directory "mimargofdir"	14
Downloading other programs and documentations	14
References	14

What changed since the last version?

- December 17, 2010: I changed the calculation of the p-value for S_f in “TESTgod.R” as there was an error. Thanks to Nick Levens for finding the error.
- March 18, 2010: I added objects and functions in order to specify the random seeds from the command line. The files "mimargof.c", "mimargof.h", "params.c", "rand1.c", "rand2.c", "rand1t.c" and "rand2t.c" were changed.
- February 11, 2010: I added the file "make_gametes_noanc.c". The program MIMARgof_noanc simulates polymorphism data under the isolation-migration model with parameters specified by the user and outputs the summary statistics of the polymorphism calculated assuming unknown ancestral states. A section about this option was added to "MIMARgofdoc.pdf".
- December 28, 2009: The file "params.c" of MIMARgof was changed:
 - The locus-specific recombination rates are now calculated by $\rho_y = w_y \rho(Z_y - 1)$ instead of $w_y \rho(Z_y)$.
 - I added several options to obtain the locus-specific population recombination rates.
 - I changed the instances of “calloc(1...” by “malloc(...)”.

I updated "MIMARgofdoc.pdf" accordingly and tried to clarify many points in the documentation. Note that the scaling of the recombination rate can be confusing (at least for me) so be careful when setting recombination information. Thanks to Peter Andolfatto who helped me through my confusions on getting the recombination rates right.

- November 27, 2009:
 - The file "mimargof.c" of MIMARgof was changed.: I fixed potential memory leaks.
 - The file "streec.c" of MIMARgof was changed: I fixed a bug on memory allocation that occurred when $n_{1y} + n_{2y} \gg n_{11} + n_{21}$ for any $y \in [2, Y]$. Thanks to Yongshuai Sun who mentioned the bug to me.
- March 3, 2009:
 - The file "params.c" of MIMARgof was changed: I corrected a bug on memory allocation. Thanks Susan J. Miller for mentioning this bug to me.
 - The file "mimargof.c" of MIMARgof was changed so that now when the prior on gene flow is “1 a b”, a and b can both be negative and the **summary output file** show the estimate of the gene flow rate. Thanks to Camille Roux for highlighting this problem to me.
 - The documentation was changed. I added a figure to help calculate the summary statistics.
- May 29, 2008: The file "params.c" of MIMARgof was changed: I removed several check flags to allow greater freedom to the user. Thanks Armando Geraldes for mentioning the problem to me.
- September 18, 2007: The files "params.c" and "mimargof.h" of MIMARgof were changed. In the previous version, the locus names could not start with a number. Now the locus names can be any string of up to 50 characters.

Downloading and compiling

All relevant files are included in the tar file "mimar.tar" available at <http://mplab.bsd.uchicago.edu/dataNprograms.htm>. Download this tar file to your machine then extract the files from the archive with: "tar -xvf mimar.tar". After extracting, type "cd mimargofdir/" and compile the program MIMArgof by typing:

```
gcc -o mimargof mimargof.c params.c make_gametes.c streec.c randX.c tajd.c -lm
```

(X is either 1, 2, 1t or 2t) or alternatively, by typing `make`, which contains this compilation line with optimization and `rand1.c`.

The choice of compilation depends on which pseudo-random number generator the user has available. "rand1.c" and "rand2.c" call `drand48()` and `drand()`, respectively. With "rand1.c" and "rand2.c", MIMArgof first looks for the file "seedmimar" to find the seed values for initializing the random number generator. If no "seedmimar" file is found, the generator is seeded with a default value. When the simulation is finished, the state of the random number generator is output to "seedmimar". In this way, each time MIMArgof is invoked, a new data set is produced. Note that, unlike in MIMAR and MIMARsim, the seeds are not printed in the standard output of MIMArgof. If you want to perform the same analysis, record the seeds values in "seedmimar" *before* executing MIMArgof or either of the Perl scripts, then edit "seedmimar" to those values. (The program can also be compiled with "rand1t.c" and "rand2t.c", which use the system clock for seeding the generators and does not use the file "seedmimar" at all.)

Running MIMArgof

In this section, I describe how to run MIMArgof, a program that simulates polymorphism data under the isolation-migration model with parameters specified by the user and outputs the summary statistics calculated for the simulated data set.

The input file

The input file for MIMArgof has the same format as the input file required by MIMAR. Use the switch "-lf input", to specify the input file name containing the information on the loci. See "MIMARdoc.pdf" for further details.

Below is an example for a four locus data set, in which locus1_autosom is autosomal and locus2_Xlinked, locus3_Ylinked and locus4_mtDNA are X- Y- or mtDNA-linked, respectively. The recombination scalar was set accordingly and I assumed that the mutation rate on the mtDNA was 2×10^{-7} (see the file "inputmimar"):

Name	length	x_y	v_y	w_y	n_1	n_2	S_1	S_2	S_s	S_f	//
locus1_autosom	1000	1	1	1	10	10	14	1	11	0	
locus2_Xlinked	1000	0.75	1	0.5	10	10	2	5	2	0	
locus3_Ylinked	1000	0.25	1	0	10	10	2	2	0	0	
locus4_mtDNA	1000	0.25	10	0	10	10	9	20	14	0	

The basic command line

```
mimargof  $Y$  -lf input -u  $\mu$  -t  $\hat{\theta}_1$  -ej  $\hat{T}$  [options]
```

This line shows the simplest usage of MIMArgof. There is one argument followed by the parameters (introduced by switches, such as "-t"). The argument Y , must appear first, while the switches can appear in any order.

Y	The number of loci considered.
μ	The generational mutation rate per bp.
$\widehat{\theta}_1 = \widehat{4N_1\mu}$	The estimated population mutation rate per bp for the first population. When the other population mutation rates are not specified, they are equal to $\widehat{\theta}_1$.
\widehat{T}	The estimated split time in generations, at which, backward in time, all lineages in population 2 are moved to population 1.
input	The input file name containing the information on the loci with their S statistics (see section “The input file”).
[options]	A list of any options/switches described in the next sections.

The migration rate is zero by default. The user needs to specify $\widehat{\theta}_1$ and \widehat{T} because these two parameters are the minimum information required to build the simplest isolation-migration model, in which the two populations split \widehat{T} generations ago without subsequent gene flow, and in which the ancestral and descendant populations have the same population mutation rate, $\widehat{\theta}_1$.

In the following basic command line **MIMARgof** will simulate data for the loci defined in the file "inputmimar" (see example in section “The input file”) for a model with: $\widehat{\theta}_1 = \widehat{\theta}_2 = \widehat{\theta}_A = 0.00148198$, $\widehat{T} = 7957.96$ generations and $\widehat{M} = 0$. $\widehat{\theta}_1$ and \widehat{T} are the point estimates (here I chose the modes) found in the file "exsoutput" (see section “The summary output file” of "MIMARdoc.pdf"). It will output the summary statistics of the polymorphism in the standard output (see section “The standard output”).

```
mimargof 4 -lf inputsim -u 2e-8 -t 0.00148198 -ej 7957.96
```

Providing the other parameters of the model

To provide information about the of the isolation-migration model, the user needs to use either of the following switches:

$-n \widehat{\theta}_2 \quad -N \widehat{\theta}_A \quad -M \widehat{M}$

$\widehat{\theta}_2 = \widehat{4N_2\mu}$	The estimated population mutation rate per bp for the second population.
$\widehat{\theta}_A = \widehat{4N_A\mu}$	The estimated ancestral population mutation rate per bp.
$\widehat{M} = \widehat{4N_1m}$	The estimated expected number of migrants between the two populations each generation, where $\widehat{m} = \widehat{M} \frac{\mu}{\theta_1}$ is the estimated symmetrical migration rate between the two populations. Note that M is defined in term of N_1 (see section “Spatial structure and migration:” in "msdoc.pdf" for further details).

In the following example, **MIMARgof** will proceed as before, but with the parameter values: $\widehat{\theta}_1 = \widehat{\theta}_2 = 0.00148198$, $\widehat{\theta}_A = 0.005$, $\widehat{T} = 7957.96$ generations and $\widehat{M} = 0.878097$ (i.e., the point estimates (here I chose the modes) found in the file "exsoutput"). The summary statistics of the polymorphism will be recorded in the standard output file "outgof".

```
mimargof 4 -lf inputmimar -u 2e-8 -t 0.00148198 -ej 7957.96 -n 0.00148198 -N .005 -M
0.878097 > outgof
```

The standard output

An example of a standard output from **MIMARgof** is reported below (see the file "outgof"):

```
9 13 31 1 0.443244 2.77222 3.86667 -0.817477 0.53465
```

In order the nine summaries of the simulated polymorphism are:

1. $\sum_{y=1}^Y S_{1_y}$ The number of derived polymorphisms unique to the samples from population 1, S_1 , summed over the Y loci.
2. $\sum_{y=1}^Y S_{2_y}$ The number of derived polymorphisms unique to the samples from population 2, S_2 , summed over the Y loci.
3. $\sum_{y=1}^Y S_{s_y}$ The number of polymorphisms with shared derived alleles between the two samples, S_s , summed over the Y loci.
4. $\sum_{y=1}^Y S_{f_y}$ The number of polymorphisms with fixed alleles in either sample, S_f , summed over the Y loci.
5. $\frac{\sum_{y=1}^Y F_{ST_y}}{Y}$ The mean over the Y loci of F_{st} , a measure of differentiation between the two population samples (Wright, 1931; Hudson et al., 1992).
6. $\frac{\sum_{y=1}^Y \pi_{1_y}}{Y}$ The mean over the Y loci of the mean pairwise difference for population 1, π_1 (Nei and Li, 1979).
7. $\frac{\sum_{y=1}^Y \pi_{2_y}}{Y}$ The mean over the Y loci of the mean pairwise difference for population 2, π_2 (Nei and Li, 1979).
8. $\frac{\sum_{y=1}^Y D_{1_y}}{Y}$ The mean over the Y loci of Tajima's D for population 1, D_1 (Tajima, 1989).
9. $\frac{\sum_{y=1}^Y D_{2_y}}{Y}$ The mean over the Y loci of Tajima's D for population 2, D_2 (Tajima, 1989).

More complex models

All the options and switches listed in this section of "MIMARdoc.pdf" are conserved in MIMARgof (the only exception is that the migration rates cannot be chosen from prior distributions). Read the section "More complex models" in "MIMARdoc.pdf" for details. To test the goodness of fit of a model MIMAR, one needs to all the options and switches that were used when running MIMAR. For example, if MIMAR ran with:

```
mimar 11000 1000 4 -lf inputmimar -u 2e-8 -t u .001 .01 -ej u 0 1e5 -N .005 -M 1 -2 2 -o
soutput [extra_MCMC_options] -r e 1.667 -en .5 .2 -eM .8 10 >std_output
```

The command line for MIMARgof should contain all the extra switched describing the model, in which the values $\hat{\theta}_1$, \hat{T} and \hat{M} are either the point estimates found in the summary output file from MIMAR "exsoutput" or the values from a set of parameters sampled from the posterior distribution estimated by MIMAR (i.e., from a line of the file "std_output"):

```
mimargof 4 -lf inputmimar -u 2e-8 -t -t  $\hat{\theta}_1$  -ej  $\hat{T}$  -N .005 -M  $\hat{M}$  -r e 1.667 -en .5 .2 -eM
.8 10
```

Crossing over

Fixed ρ across loci. If MIMAR ran with the command line:

```
mimar nsteps bsteps Y -lf input -u  $\mu$  -t  $\theta_1$  -ej  $T$  -r  $\rho$  -o soutput
```

MIMARgof needs to run with:

```
mimargof Y -lf input -u  $\mu$  -t  $\hat{\theta}_1$  -ej  $\hat{T}$  -r  $\rho$ 
```

$\rho = 4N_1c$ is the population cross-over rate per bp per generation and c is the probability of cross-over per generation per bp. MIMARgof calculates the locus-specific population recombination rate like MIMAR (see section "Crossing over" in "MIMARdoc.pdf").

Fixed locus-specific ρ . If the locus-specific recombination rates were fixed in the input file:

- If the switch “- r 1” was used when running MIMAR, use “- r 1” when running MIMARgof.

ATTENTION: This assumes that in the input file, the recombination scalars for the recombining loci were set to $w_y = \omega_y \widehat{\rho}_{o_y}$, the scaled sex-averaged locus-specific population recombination rate per bp, i.e., for an X-linked locus, c is the female recombination rate and $\omega_y = \frac{1}{2}$ so that $\omega_y \widehat{\rho}_{o_y} = 2\widehat{N}_1 c_y$ (see the file "inputmimar_4Nc").

- If the switch “- r 2” was used when running MIMAR, use “- r 2” when running MIMARgof.

ATTENTION: This assumes that in the input file, the recombination scalars for the recombining loci were set to $w_y = \omega_y \widehat{c}_y$, the scaled sex-averaged locus-specific recombination rate per bp, i.e., for an X-linked locus, \widehat{c}_y is the estimate of the female recombination rate so the scaled sex-averaged rate is $\frac{1}{2}\widehat{c}_y = \omega_y \widehat{c}_y$ (see the file "inputmimar_c").

Variable locus-specific ρ . The recombination rate can be allowed to vary across loci.

- If the switch “- r e λ ” was used when running MIMAR, use “- r e λ ” when running MIMARgof: the ratio $r = \frac{c}{\mu}$ is drawn from an exponential distribution prior with mean $\frac{1}{\lambda}$ for each recombining locus.
- If the switch “- r n ν σ ” was used when running MIMAR, use “- r n ν σ ” when running MIMARgof: the ratio $r = \frac{c}{\mu}$ is drawn from a normal distribution prior with mean ν and standard deviation σ for each recombining locus.

Asymmetrical migration rates

If MIMAR ran with the command line:

```
mimar nsteps bsteps Y -lf input -u  $\mu$  -t  $\theta_1$  -ej  $T$  -m  $i$   $j$   $M_{ij}$  -o soutput
```

MIMARgof needs to run with:

```
mimargof Y -lf input -u  $\mu$  -t  $\widehat{\theta}_1$  -ej  $\widehat{T}$  -m  $i$   $j$   $\widehat{M}_{ij}$ 
```

$\widehat{M}_{ij} = 4\widehat{N}_1 \widehat{m}_{ij}$, i and $j \in [1, 2]$, $i \neq j$ is the estimated expected number of individuals that migrate from population i into population j each generation, thinking forward in time, where \widehat{m}_{ij} is the estimated fraction of population j that is made up of migrant from population i every generation. Note that M_{ij} is defined in term of N_1 (see section “Spatial structure and migration:” in "msdoc.pdf" for further details).

Other options conserved from ms. Use at your own peril!

Read the section “Other options conserved from ms. Use at your own peril!” in "MIMARdoc.pdf" for details.

Summary of command line options

The following options are required

-t θ_1	Set the population mutation rate per bp to $4N_1\mu$ for population 1 (the default population).
-u μ	Set the mutation rate per bp to μ .
-lf input	Set the input file name.
-ej T	Set the time of split to T generations ago. Backward in time, all lineages in population 2 are moved to population 1 at time

The following options are not required, but should be used if they were used when running MIMAR.

<code>-n θ_2</code>	Set the population 2 mutation rate per bp to $4N_2\mu$.
<code>-N θ_A</code>	Set the ancestral population mutation rate to $4N_A\mu$.
<code>-M M</code>	Set the expected number of migrants between the two populations each generations to $4N_1m$.
<code>-m $i\ j\ M_{ij}$</code>	Set the expected number of migrants from population i into population j each generation, i and $j \in \{1, 2\}$, $i \neq j$, to $4N_1m_{ij}$.
<code>-r ρ</code>	Set the population recombination rate per bp to $4N_1c$.
<code>-r e λ</code>	Set the prior distribution of $r = \frac{c}{\mu}$ to Exponential with mean $\frac{1}{\lambda}$.
<code>-r n $\nu\ \sigma$</code>	Set the prior distribution of $r = \frac{c}{\mu}$ to Normal(ν, σ).
<code>-r 1</code>	Set the locus-specific population recombination rates per bp to the value specified with w in the input file.
<code>-r 2</code>	Set the locus-specific recombination rates per bp to the value specified w in the input file.

The following options are conserved from ms. Use at your own peril!

<code>-se seed1 seed2</code>	Specify the random seeds from the command line.
<code>seed3</code>	
<code>-f filename</code>	Read command line arguments from file <code>filename</code> .
<code>-c $f\ \lambda$</code>	Set ratio of gene conversion to recombination to f and the track length to λ .
<code>-G α</code>	Set growth parameter of all populations to α .
<code>-g $i\ \alpha_i$</code>	Set growth rate of population i to α_i .

The following options specify events occurring at time τT generations. Up to 10 such switches can be used. It is the user's responsibility to specify times that are compatible with the isolation-migration model. Note that the switch "`-ej`" can be used only once.

<code>-eG $\tau\ \alpha$</code>	Set all growth rates to α at time τT generations.
<code>-eg $\tau\ i\ \alpha_i$</code>	Set growth rate of population i to α_i at time τT generations.
<code>-eb $\tau\ x$</code>	Set all population mutation rates to $x\theta_1$ at time τT generations.
<code>-en $\tau\ i\ x$</code>	Set population i mutation rate to $x\theta_1$ at time τT generations.
<code>-eM $\tau\ x$</code>	Set the symmetrical migration rate to x at time τT generations.
<code>-em $\tau\ i\ j\ x$</code>	Set $4N_1m_{ij}$ to x at time τT generations.

Performing a goodness of fit test using point estimates

In this section, I describe how to run the Perl script `perlgofest`, which executes `MIMARgof` using the point estimates of the parameters provided by MIMAR, which can be found in the summary output file (e.g., in "exsoutput"). By simulating samples using e.g., the mode of the parameters, you can test whether the model (described by the modes of the posterior distributions) estimated by MIMAR fits the data.

Setting perlgofest

`perlgofest` runs `MIMARgof` \$`int` times with the point estimates of the parameters provided by the user. The following parameters need to be changed in the file `perlgofest` depending of the data and the user's preferences:

\$nloci Sets Y , the number of loci in the input file.
\$mu Sets μ , the generational migration rate per bp. Set μ to the same value used to run MIMAR (i.e., after the switch “-u”).
\$int Sets the number of times the program MIMARgof will run with the specified set of parameter estimates.
\$inputfile Sets the name of the input file (see section “The input file”).
\$outfile Sets the name of the output file (see “The output file of perlgofest”).
@temp Sets the parameter values to the point estimates (e.g., modes) found in the summary output file from MIMAR (e.g., in "exsoutput"):
 $\$temps[1] = \hat{\theta}_1$
 $\$temps[2] = \hat{\theta}_2$
 $\$temps[3] = \hat{T}$
 $\$temps[4] = \hat{\theta}_A$
 $\$temps[5] = \hat{M}_{12}$
 $\$temps[6] = \hat{M}_{21}$
\$cmd_line The user needs to add the extra switches used to run MIMAR in the line:
 `"/mimargof $nloci -lf $inputfile -u $mu -t $temp[1] -ej $temp[3] -N $temp[4] -n $temp[2] -m 1 2 $temp[5] -m 2 1 $temp[6] >>$outfile";`

For example, if MIMAR ran with recombination, a bottleneck $0.5T$ generations ago and a change of gene flow rate $0.8T$ generations ago:

```
mimar 11000 1000 4 -lf inputmimar -u 2e-8 -t u .001 .01 -ej u 0 1e5 -N .005 -M 1 -2 2 -o
      soutput [extra_MCMC_options] -r e 1.667 -en .5 .2 -eM .8 10
```

You need to set ($\hat{\theta}_1$, $\hat{\theta}_2$, \hat{T} and \hat{M} are the point estimates (e.g., modes) found in the summary output file "soutput"):

```
$nloci=      4;
$mu=         2e-8;
$int=        1000;
$inputfile=  "inputmimar";
$outfile=    "outputgofest";
$temps[1]=    $\hat{\theta}_1$ ;
$temps[2]=    $\hat{\theta}_2$ ;
$temps[3]=    $\hat{T}$ ;
$temps[4]=   0.005;
$temps[5]=    $\hat{M}$ ;
$temps[6]=    $\hat{M}$ ;
$cmd_line=   ./mimargof $nloci -lf $inputfile -u $mu -t $temp[1] -ej $temp[3] -N
              $temp[4] -n $temp[2] -m 1 2 $temp[5] -m 2 1 $temp[6] -m 2 1 $temp[7]
              -r e 1.667 -en .5 .2 -eM .8 10 >>$outfile";
```

Save perlgofest, in a terminal type “chmod +x perlgofest” then finally “./perlgofest” to execute the script.

By default, perlgofest uses the modes of the estimates found in the file "exsoutput", runs MIMARgof 1,000 times and print the 1,000 lines of summary statistics in the file "outputgofest".

The output file of perlgofest

Below I report the three first lines out of 1001 of the file "outputgofest", the output file of perlgofest with the default values:

```

Sim# Step# S1 S2 Ss Sf Fst pi1 pi2 D1 D2
1 NA 12 42 38 1 0.451486 2.98889 7.98889 -0.764146 0.361559
2 NA 16 19 26 0 0.215028 2.84444 3.45 -0.482247 0.0649717

```

The first line is the header of the results. For each following lines, eleven values are reported:

1. The run number of `MIMARgof` (here the two first runs).
2. “NA”, required uninformative column in order to use the script “`testGOF.R`” (see section “Does the estimated model fit the data?”).
- 3.-11. The nine summary statistics for the simulated data set output by `MIMARgof` (see section “The standard output”).

Performing a goodness of fit test by sampling from the estimated posterior distribution

In this section, I describe how to run `perlgof`, which executes `MIMARgof` with sets of parameters sampled from the posterior distribution estimated by `MIMAR`. Simulating samples using parameters chosen from the posterior distribution allows one to test whether the model described by the posterior distribution estimated by `MIMAR` fits the data by performing a Bayesian goodness of fit test (i.e., by calculating the posterior predictive p-values for the summary statistics output by `MIMARgof`, see sections “The standard output” and “Does the estimated model fit the data?”).

Setting `perlgof`

`perlgof` reads each line containing a set of parameters from the posterior distribution estimated by `MIMAR` reported in a standard output file of `MIMAR` (see section “The standard output” in “`MIMARdoc.pdf`”). `perlgof` then executes `MIMARgof` with these sets of parameters. The following parameters need to be changed in the file `perlgofest` depending of the data and the user’s preferences:

<code>\$nloci</code>	Sets Y , the number of loci in the input file.
<code>\$mu</code>	Sets μ , the generational migration rate per bp. Set μ to the same value used to run <code>MIMAR</code> (i.e., after the switch “-u”).
<code>\$int</code>	<code>\$int-1</code> sets the number of lines in the standard output file of <code>MIMAR</code> skipped between runs of <code>MIMARgof</code> . My advice: I would recommend running <code>MIMARgof</code> with about 10,000 sets of parameters sampled from the posterior distribution. This will make the process reasonably fast and provide reasonable estimates of the posterior predictive p-values. If the standard output file of <code>MIMAR</code> is larger than 10,000 lines, you should consider running <code>MIMARgof</code> only every, say, 10 recorded sets of parameters, by setting, e.g., “ <code>\$int=10</code> ”.
<code>\$addburnin</code>	Sets the number of lines of extra burnin My advice: If you find that <code>MIMAR</code> was run with too little burnin (see section “How long to run the program and how to improve the MCMC?” in “ <code>MIMARdoc.pdf</code> ”), you can choose to ignore the first few steps recorded in the file to add some extra burnin by setting “ <code>\$addburnin>0</code> ”.
<code>\$inputfile</code>	Sets the name of the input file (see section “The input file”).
<code>\$outfile</code>	Sets the name of the output file (see “The output file of <code>perlgof</code> ”).
<code>\$post_dist</code>	Sets the name of the standard output file of <code>MIMAR</code> containing the estimate of the posterior distribution.
<code>\$extra_switches</code>	The string of character with the extra switches used to run <code>MIMAR</code> .

For example, if `MIMAR` ran with recombination, a bottleneck $0.5T$ generations ago and a change of gene flow rate $0.8T$ generations ago:

```
mimar 11000 1000 4 -lf inputmimar -u 2e-8 -t u .001 .01 -ej u 0 1e5 -N .005 -M 1 -2 2 -o
      soutput [extra_MCMC_options] -r e 1.667 -en .5 .2 -eM .8 10 >std_output
```

You need to set:

```
$nloci=      4;
$mu=         2e-8;
$int=        1;
$adddburnin= 0;
$inputfile=   "inputmimar";
$outfile=     "outputgof";
$post_dist=   "std_output";
$extra_switches= "-r e 1.667 -en .5 .2 -eM .8 10";
```

Save `perlgof`, in a terminal type “`chmod +x perlgof`” then finally “`./perlgof`” to execute the script.

Note that, unlike in `perlgofest`, the user does not need to specify the parameter values since they are registered in the file “`std_output`” in this case. For example, when analyzing the standard output file of MIMAR “`outputmimar`” with `perlgof`, the first run of `MIMARgof` will be with the set of parameters: `$temp[1]=$temp[2]=0.00512989`, `$temp[4]=7179.1` `$temp[5]=0.005` and `$temp[6]=$temp[7]=5.96188`.

By default, `perlgof` uses every nine sets of parameters from the estimated posterior distribution found in the file “`outputmimar`”, runs `MIMARgof` 1,000 times with those sets of parameters and print the 1,000 lines of summary statistics in the file “`outputgof`”.

The output file of perlgof

Below I report the three first lines out of 1001 of the file “`outputgof`”, the output file of `perlgof` with the default values:

```
Sim# Step# S1 S2 Ss Sf Fst pi1 pi2 D1 D2
1 1001 30 36 16 0 0.0626264 3.37222 4.12778 0.125916 -0.509258
2 1011 18 42 27 0 0.0728636 2.93889 4.47222 0.036836 -1.14938
```

The first line is the header of the results. For each following lines, eleven values are reported:

1. The run number of `MIMARgof` (here the two first runs).
2. The step number associated with the sampled set of parameters (i.e., the first number in a recorded MCMC step in the standard output of MIMAR).
- 3.-11. The nine summary statistics for the simulated data set output by `MIMARgof` (see section “The standard output”).

Does the estimated model fit the data?

I provided the R script “`testGOF.R`” to help the user test whether the estimated model fit the data.

Setting “`testGOF.R`”

The following parameters need to be changed in the file “`testGOF.R`” depending of the data and the user’s preferences:

gof_file1 Sets the name of the output file of `perlgof` (or `perlgofest`) from one seed.
gof_file2 Sets the name of the output file of `perlgof` (or `perlgofest`) from a different seed.
true Sets the values of the nine summary statistics calculated from the original data set used to run MIMAR. Write the values in the same order as presented in the standard output of MIMARgof separated by “,”:

$$\text{true} = c(\sum_{y=1}^Y S_{1y}, \sum_{y=1}^Y S_{2y}, \sum_{y=1}^Y S_{sy}, \sum_{y=1}^Y S_{fy}, \frac{\sum_{y=1}^Y F_{STy}}{Y}, \frac{\sum_{y=1}^Y \pi_{1y}}{Y}, \frac{\sum_{y=1}^Y \pi_{2y}}{Y}, \frac{\sum_{y=1}^Y D_{1y}}{Y}, \frac{\sum_{y=1}^Y D_{2y}}{Y})$$

nbin Sets the number of bins in each histogram. Reduce this number to smooth the plots.

The output of "testGOF.R"

The execution of the functions in "testGOF.R" using R leads to four graphs displaying the average over the two `perlgof` (or `perlgofest`) runs of the expected distribution of a statistic under the estimated model and its observed value (calculated from the data set used to run MIMAR and shown as a vertical line, see Figure 2):

S statistics Displays the four expected distributions of $\sum_{y=1}^Y S_{1y}$, $\sum_{y=1}^Y S_{2y}$, $\sum_{y=1}^Y S_{sy}$ and $\sum_{y=1}^Y S_{fy}$.
F_{ST} Displays the expected distribution of $\frac{\sum_{y=1}^Y F_{STy}}{Y}$.
π Displays the two expected distributions of $\frac{\sum_{y=1}^Y \pi_{1y}}{Y}$ and $\frac{\sum_{y=1}^Y \pi_{2y}}{Y}$.
D Displays the two expected distributions of $\frac{\sum_{y=1}^Y D_{1y}}{Y}$ and $\frac{\sum_{y=1}^Y D_{2y}}{Y}$.

"testGOF.R" also output the p-values calculated for each statistic. If the p-value is small (i.e., $< .05$), it means that the estimated model does not fit this particular statistic well. Note that if the output files were generated by `perlgof`, the p-values correspond to the posterior predictive p-values.

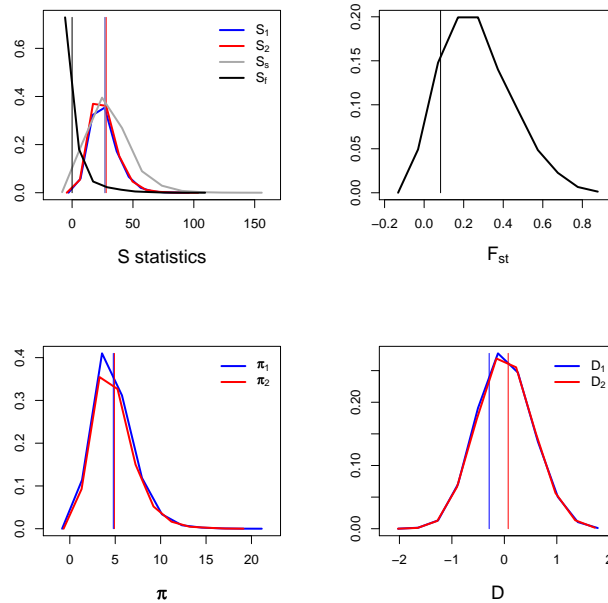


Figure 2: Example of goodness of fit graphs. Shown are the results from the files "outputgof1" and "outputgof2", the output files of `perlgof` for the standard output files of MIMAR "outputmimar1" and "outputmimar2". The observed values (vertical lines) were calculated from the data set summarized in the file "inputmimar".

If the ancestral alleles are unknown use MIMARgof_noanc

I provide the program `MIMARgof_noanc` which calculates the summary statistics of the polymorphism assuming *unknown ancestral alleles*. `MIMARgof` and `MIMARgof_noanc` are identical in all other aspects.

Use `MIMARgof_noanc` only if you used `MIMAR_noanc` to estimate the parameters of the model from data with unknown ancestral states.

Compiling MIMARgof_noanc

To compile the program type:

```
gcc -o mimargof_noanc mimargof.c params.c make_gametes_noanc.c streec.c randX.c tajd.c  
-lm
```

See section “Downloading and compiling” for more details.

List of parameters and symbols

$\theta_i = 4N_i\mu$	The population mutation rate per bp per generation for population $i \in \{1, 2, A\}$.
μ	The generational mutation rate per bp.
N_i	The diploid effective size for population $i \in \{1, 2, A\}$.
T	The split time in generations, at which, backward in time, all lineages in population 2 are moved to population 1.
$M = 4N_1m$	The expected number of individuals in population 2 replaced by migrants from population 1 each generation (in forward direction).
m	The symmetrical fraction of a population that is made up of migrant from the other population each generation.
$M_{ij} = 4N_1m_{ij}$	The expected number of individuals in population j replaced by migrants from population i (in forward direction), i and $j \in \{1, 2\}$, $i \neq j$.
m_{ij}	The fraction of population j that is made up of migrant from population i each generation.
$\rho = 4N_1c$	The population recombination rate per bp per generation.
c	The generational recombination rate per bp.
$r = \frac{c}{\mu}$	
Y	The number of loci considered.
n_i	The sample size for the locus in population $i \in \{1, 2\}$.
x	The inheritance scalar reflecting copy number differences for a locus.
v	The mutation rate variation scalar for a locus.
w	The inheritance and rate variation scalar for recombination rate for a locus.
ω	The ratio of the locus-specific population recombination rate per bp over ρ .
S_i	The number of derived polymorphisms unique to the samples from population $i \in \{1, 2\}$.
S_s	The number of polymorphisms with shared derived alleles between the two samples.
S_f	The number of polymorphisms with fixed alleles in either sample.
F_{st}	A measure of differentiation between the two population samples (Wright, 1931; Hudson et al., 1992).
π_i	The mean pairwise difference for population $i \in \{1, 2\}$, (Nei and Li, 1979).
D_i	Tajima’s D for population $i \in \{1, 2\}$ (Tajima, 1989).

List of files in the directory "mimargofdir"

Program files for MIMAR	Examples of input files	Examples of summary and standard output files from MIMAR	Examples of standard output files	Scripts to help with goodness of fit tests	Other or documentation files
make_gametes.c	inputmimar	exsoutput	outgof	perlgof	makefile
mimargof.c	inputmimar_4Nc	outputmimar	outputgof	perlgofest	make_gametes_noanc.c
mimargof.h	inputmimar_c	outputmimar1	outputgof1	TestGOF.R	MIMARgofdoc.pdf
params.c		outputmimar2	outputgof2		seedmimar
rand1.c			outputgofest		
rand1t.c					
rand2.c					
rand2t.c					
streec.c					
tajd.c					

Downloading other programs and documentations

MIMAR and "MIMARdoc.pdf" are found in "mimar.tar" available at <http://przeworski.uchicago.edu/cbecquet/download.html>.

ms and "msdoc.pdf" are available at <http://home.uchicago.edu/~rhudson1/source/mksamples.html>.

R is available at <http://www.r-project.org/>.

References

- Hudson, R. R., Slatkin, M., and Maddison, W. P., 1992. Estimation of levels of gene flow from DNA sequence data. *Genetics*, **132**:583–589.
- Meng, X. L., 1994. Posterior predictive p -values. *Ann. Stat.*, **22**:1142–1160.
- Nei, M. and Li, W. H., 1979. Mathematical model for studying genetic variation in terms of restriction endonucleases. *Proc. Natl. Acad. Sci.*, **76**:5269–5273.
- Tajima, F., 1989. Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. *Genetics*, **123**:585–595.
- Wright, S., 1931. Evolution in Mendelian Populations. *Genetics*, **16**:97–159.