especially in genome-wide association studies

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## Introduction

#### Synopsis

Package cgmisc contains miscellaneous functions, hopefully useful for extending genome-wide association study (GWAS) analyses.

## Getting help

Like every other R function, the functions provided in this package are documented in the standard R-help (Rd) format and can be easily accessed by issuing **help**() or its shorter version, ? function. For instance, if you want to get more information on how to use the clump.markers() function, type either help(clumpmarkers) or ?clump.markers and press return/enter. To see this document from within R you type vignette('cgmisc').

#### Purpose of this document

This document aims at presenting how to use functions provided in this package in a typical GWAS data analyses workflow. It is, however, not pretending to be a GWAS tutorial as such.

#### Conventions

- All R commands are written in terminal type: myfun(foo=T, bar=54)
- In the above example: myfun is a function and both foo and bar are its arguments

# Working with cgmisc

#### Installation

In order to install cgmisc, you either use one of the R GUIs (native R GUI, RStudio etc.) or type the following command:

```
install.packages("cgmisc", repos="")
```

Functions in the cgmisc package often complement or use GenABEL package functions and data structures.GenABEL is an excellent and widely-used R package for performing genome-wide association studies and much more...

Therefore GenABEL will be loaded automagically when loading cgmisc. You can load cgmisc package by as follows:

```
require("cgmisc")

## Loading required package: cgmisc
## Warning: there is no package called 'cgmisc'
```

After having loaded the package it is time to load some data:

```
load('data/data.rda')
```

## **Association Analysis**

Some of cgmisc functions use data which are the result of GWAS analyses. Let's perform GWAS on our data to obtain GenABEL scan.gwaa-class object:

```
an0 <- qtscore(response ~ sex, data = data)
## Warning: 1 observations deleted due to missingness</pre>
```

And have a look at top 5 markers

```
summary(an0, top = 5)
## Summary for top 5 results, sorted by P1df
              Chromosome Position Strand A1 A2 N
                                                      effB se_effB
                  34 40399702 u T G 206 -0.2834 0.06954
1 15399848 u A G 189 0.5887 0.14714
## BICF2P1063345
## BICF2P682714
## BICF2G630569243
                                       u C A 206 0.3071 0.07797
                        6 80458945
                                       u C T 206 0.2628 0.06682
## BICF2S2366791
                        6 70667322
## BICF2G630450144 34 40416964
                                     u A G 206 -0.2705 0.06933
##
        chi2.1df P1df effAB effBB chi2.2df
## BICF2P1063345 16.60 4.609e-05 -0.3644 -0.5354 17.51 1.579e-04
## BICF2P682714
                  16.01 6.309e-05 0.5912 1.1515 16.01 3.339e-04
## BICF2G630569243 15.51 8.216e-05 0.2904 0.6358 15.57 4.167e-04
## BICF2S2366791
                    15.46 8.410e-05 0.4558 0.4732 20.15 4.202e-05
## BICF2G630450144 15.22 9.573e-05 -0.3339 -0.5161 15.77 3.756e-04
##
                    Pc1df
## BICF2P1063345 7.357e-05
## BICF2P682714 9.911e-05
## BICF2G630569243 1.274e-04
## BICF2S2366791 1.302e-04
## BICF2G630450144 1.472e-04
```

Once this is done, we can proceed with cgmisc functions.

#### **Functions**

#### Plot.Manhattan.LD

The plot.Manhattan.LD function allows you to visualize the LD pattern in a genome fragment on an enchanced Manhattan plot. You select one marker, typically the one with the strongest association to the analysed trait and all other markers in the region are coloured according to the degree of linkage disequilibrium with this index marker.

#### Clump.markers

clump.markers function implements clumping procedure described in PLINK documentation. Clumping is based on linkage disequilibrium. The function returns list of clumps which can be used for further analyses or plotted using plot.clumps function included in our package.

#### plot.clumps

plot.clumps function plots clumps resulting from running the clump.markers function on Manhattan Plot.

#### Compute.Fstats

Fixation index Fst is a measure of population differentiation due to genetic structure. Given a set of genotypes in two populations, the function computes fixation index (Fst) and corresponding indices: Fit and Fis. Pops is a vector of two values indicating to which population an individual belongs. Typically, one uses a vector of zeroes and ones where 0 marks an individual belonging to population 1 and 1 marks an individual belonging to population 2. Often, the vector is a result of clustering in MDS-scaled genomic kinship space.

```
fstats <- compute.Fstats(data, pops)
## Error: could not find function "compute.Fstats"</pre>
```

#### Plot.fstats

Plot results of compute. Fstats function.

```
plot.fstats(data, fstats)
## Error: could not find function "plot.fstats"
```

## create. Haploview.info

The program PHASE implements a Bayesian statistical method for reconstructing haplotypes from population genotype data. Create.Haploview.info function prepares PHASE and fastPHASE input files from GWAS data object. It can be used on a single entire chromosome or on a specified chromosomal region.

## get.adjacent.markers

This function returns the number of markers which are adjacent to the given within a distance set as parameter.

```
adj <- get.adjacent.markers(data, "BICF2P647127")
## Error: could not find function "get.adjacent.markers"</pre>
```

```
print(adj[5:10])
## Error: object 'adj' not found
```

## get.chr.midpoints

Get.chr.midpoints function might be very useful for plotting chromsome labels on x-axis. It returns a list of chromosome midpoints coordinates which are independent of coordinates used.

```
mids <- get.chr.midpoints(data)

## Error: could not find function "get.chr.midpoints"</pre>
```

## get.overlapping.windows

```
olw <- get.overlapping.windows(data, chr = 3, size = 1e+06, overlap = 2500)
## Error: could not find function "get.overlapping.windows"</pre>
```

# het.for.overlap.wind

```
het <- het.for.overlap.wind(olw)
## Error: could not find function "het.for.overlap.wind"</pre>
```

## get.window.means

## open.region.UCSC

```
open.region.UCSC(chr = 3, coords = c(40455299, 64160299), assembly = "canFam3")
```

# pop.allele.counts

```
pac <- pop.allele.counts(data, pops, progress = TRUE)
## Error: could not find function "pop.allele.counts"</pre>
```

# plot.pac

```
plot.pac(data, pac, plot.LD = FALSE)
## Error: could not find function "plot.pac"
```