Getting to fit models in ${\bf R}$

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Getting started with **R**

- Introduction
- Linear Models
- Generalised Linear Models
- Mixed Models
- 5 General Additive (Mixed) Models

A statistical model represents, often in considerably idealized form, the data-generating process (https://en.wikipedia.org/wiki/Statistical_model).

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R allows to fit efficiently and easily all main kinds of linear models:

- classical linear models (t-test, correlation, linear regression, ANOVA, ANCOVA): LM
- generalized linear models (logistic regression, Poisson regression...): GLM
- linear mixed-effects models: LMM
- generalized linear mixed-effects models: GLMM
- general additive models & general additive mixed models: GAM & GAMM

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- classical linear models (t-test, correlation, linear regression, ANOVA, ANCOVA): LM
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- linear mixed-effects models: LMM
- generalized linear mixed-effects models: GLMM
- general additive models & general additive mixed models: GAM & GAMM

Note: I have a 100 hours course on the topic (https://github.com/courtiol/LM2GLMM) but it may be a bit terse without the bla bla...

Linear models in R

R is very rich in terms of capabilities to fit linear models due to an increasing number of dedicated packages!

For now, no other software seems to be remotely as good (prognostic: only Julia or Python may change that within a decade but I find it unlikely).

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Models	Packages for fitting	Helper packages
LM	none; spaMM	car; lmtest; visreg
GLM	none; spaMM	car; DHARMa; visreg
LMM	lme4; spaMM; glmmTMB	DHARMa; pbkrtest
GLMM	lme4; spaMM; glmmTMB	DHARMa; pbkrtest
GAM	mgcv	visreg
GAMM	mgcv	

Note: those are my personal favorite ones, but they are plenty more out there.

Good books dealing with linear models in ${\bf R}$



Note: it is also useful to look at books not focussed on R!

Preparing data for (G)LM(M) & GA(M)

To maximize the chances of success prepare your data as follow:

- one row = one observation (if repeated measures, use several rows!)
- qualitative variables of class factor (check the levels, drop unused ones, set the reference properly)
- no NA (models can somewhat deal with them but it is a major source of headackes)

Preparing data for (G)LM(M) & GA(M)

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- qualitative variables of class factor (check the levels, drop unused ones, set the reference properly)
- no NA (models can somewhat deal with them but it is a major source of headackes)

Example of a good dataset:

```
head(iris)
    Sepal.Length Sepal.Width Petal.Length Petal.Width Species
## 1
             5.1
                         3.5
                                      1.4
                                                  0.2 setosa
## 2
             4.9
                         3.0
                                      1.4
                                                  0.2 setosa
             4.7
                         3.2
## 3
                                      1.3
                                                  0.2 setosa
## 4
             4.6
                         3.1
                                     1.5
                                                  0.2 setosa
             5.0
                         3.6
                                      1.4
## 5
                                                  0.2 setosa
## 6
             5.4
                         3.9
                                      1.7
                                                  0.4 setosa
str(iris)
## 'data frame': 150 obs. of 5 variables:
   $ Sepal.Length: num 5.1 4.9 4.7 4.6 5 5.4 4.6 5 4.4 4.9 ...
## $ Sepal.Width : num 3.5 3 3.2 3.1 3.6 3.9 3.4 3.4 2.9 3.1 ...
## $ Petal.Length: num 1.4 1.4 1.3 1.5 1.4 1.7 1.4 1.5 1.4 1.5 ...
## $ Petal.Width : num 0.2 0.2 0.2 0.2 0.4 0.3 0.2 0.2 0.1 ...
## $ Species : Factor w/ 3 levels "setosa", "versicolor", ...: 1 1 1 1 1 1 1 1 1 1 ...
anv(is.na(iris))
## [1] FALSE
```

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Getting started with R

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 - assumptions

Generalised Linear Models

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- outputs
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- predictions
- assumptions
- Mixed Models
 - introduction & inputs
 - outputs
 - tests
 - predictions
 - assumptions
 - advanced MM
- General Additive (Mixed) Models

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Simple notation:

$$y_i = \hat{\beta}_0 + \hat{\beta}_1 \times x_{1,i} + \hat{\beta}_2 \times x_{2,i} + \dots + \hat{\beta}_p \times x_{p,i} + \varepsilon_i$$

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Simple notation:

$$y_i = \hat{\beta}_0 + \hat{\beta}_1 \times x_{1,i} + \hat{\beta}_2 \times x_{2,i} + \dots + \hat{\beta}_p \times x_{p,i} + \varepsilon_i$$

$$y_i = \hat{y}_i + \varepsilon_i$$

Simple notation:

$$y_i = \hat{\beta}_0 + \hat{\beta}_1 \times x_{1,i} + \hat{\beta}_2 \times x_{2,i} + \dots + \hat{\beta}_p \times x_{p,i} + \varepsilon_i$$

$$\mathbf{y}_i = \hat{\mathbf{y}}_i + \varepsilon_i$$

- y_i = the observations to explain / response variable / dependent variable
- \hat{y}_i = the fitted values
- $x_{i,i}$ = constants derived from the predictors / explanatory variables / independent variables
- $\hat{\beta}_i$ = the (model parameter / regression coefficient) estimates
- ε_i = the residuals (i.e. the estimates for the error which is Gaussian with constant variance)

Matrix notation:

$$\begin{bmatrix} y_1 \\ y_2 \\ y_3 \\ \dots \\ y_n \end{bmatrix} = \begin{bmatrix} 1 & x_{1,1} & x_{1,2} & \dots & x_{1,p} \\ 1 & x_{2,1} & x_{2,2} & \dots & x_{2,p} \\ 1 & x_{3,1} & x_{3,2} & \dots & x_{3,p} \\ \dots & \dots & \dots & \dots & \dots \\ 1 & x_{n,1} & x_{n,2} & \dots & x_{n,p} \end{bmatrix} \begin{bmatrix} \hat{\beta}_0 \\ \hat{\beta}_1 \\ \hat{\beta}_2 \\ \dots \\ \hat{\beta}_p \end{bmatrix} + \begin{bmatrix} \varepsilon_1 \\ \varepsilon_2 \\ \varepsilon_3 \\ \dots \\ \varepsilon_n \end{bmatrix}$$

Matrix notation:

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$$Y = X\widehat{\beta} + \varepsilon = \widehat{Y} + \varepsilon$$

Matrix notation:

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$$Y = X\widehat{\beta} + \varepsilon = \widehat{Y} + \varepsilon$$

$$\widehat{S} = Y + \widehat{Y}$$

- Y = the vector of observations
- Y = the vector of fitted values
- X = a matrix called the design matrix (or the model matrix)
- ε = the vector of residuals

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R formula notation:

mod <- lm(Petal.Length ~ Petal.Width, data = iris)</pre>

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R formula notation:

```
mod <- lm(Petal.Length ~ Petal.Width, data = iris)</pre>
```

```
formula(mod) ## the formula
## Petal.Length ~ Petal.Width
```

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R formula notation:

1.7

6

0.4

```
mod <- lm(Petal.Length ~ Petal.Width, data = iris)</pre>
formula(mod) ## the formula
## Petal.Length ~ Petal.Width
head(model_frame <- model.frame(mod)) ## the data used for the fit</pre>
    Petal.Length Petal.Width
            1.4
                        0.2
            1.4
                       0.2
       1.3
                  0.2
        1.5
                   0.2
## 5
           1.4
                        0.2
```

R formula notation:

1.4 1.4 1.3 1.5 1.4 1.7

```
mod <- lm(Petal.Length ~ Petal.Width, data = iris)</pre>
formula(mod) ## the formula
## Petal.Length ~ Petal.Width
head(model_frame <- model.frame(mod)) ## the data used for the fit</pre>
    Petal.Length Petal.Width
           1.4
                      0.2
           1.4
                 0.2
      1.3
                 0.2
       1.5
                 0.2
        1.4
                   0.2
## 5
           1.7
                      0.4
## 6
head(model.response(model_frame)) ## the response variable
       2 3 4 5 6
```

R formula notation:

```
mod <- lm(Petal.Length ~ Petal.Width, data = iris)
formula(mod) ## the formula
## Petal.Length ~ Petal.Width
head(model_frame <- model.frame(mod)) ## the data used for the fit</pre>
    Petal.Length Petal.Width
           1.4
                       0.2
           1.4
                 0.2
      1.3
                 0.2
      1.5
                 0.2
       1.4
                   0.2
## 5
           1.7
## 6
                       0.4
head(model.response(model_frame)) ## the response variable
       2 3 4 5 6
## 1.4 1.4 1.3 1.5 1.4 1.7
head(model.matrix(mod)) ## the model matrix
    (Intercept) Petal.Width
## 1
                      0.2
                      0.2
## 3
                      0.2
                      0.2
                      0.2
## 6
                      0.4
```

R formula notation:

```
mod <- lm(Petal.Length ~ Species, data = iris)
formula(mod) ## the formula
## Petal.Length ~ Species
head(model_frame <- model.frame(mod)) ## the data used for the fit</pre>
    Petal.Length Species
          1.4 setosa
           1.4 setosa
     1.3 setosa
     1.5 setosa
     1.4 setosa
## 5
## 6
          1.7 setosa
head(model.response(model_frame)) ## the response variable
        2 3 4 5 6
## 1.4 1.4 1.3 1.5 1.4 1.7
head(model.matrix(mod)) ## the model matrix
    (Intercept) Speciesversicolor Speciesvirginica
## 1
                              0
                                              0
## 3
## 6
```

R formula notation:

```
mod <- lm(Petal.Length ~ Petal.Width + Species, data = iris)
formula(mod) ## the formula
## Petal.Length ~ Petal.Width + Species
head(model_frame <- model.frame(mod)) ## the data used for the fit</pre>
    Petal.Length Petal.Width Species
          1.4
                      0.2 setosa
          1.4 0.2 setosa
     1.3 0.2 setosa
     1.5
                 0.2 setosa
## 5
     1.4 0.2 setosa
          1.7
## 6
                    0.4 setosa
head(model.response(model_frame)) ## the response variable
       2 3 4 5 6
## 1.4 1.4 1.3 1.5 1.4 1.7
head(model.matrix(mod)) ## the model matrix
    (Intercept) Petal. Width Speciesversicolor Speciesvirginica
                      0.2
## 1
                                        0
                                                       0
                      0.2
## 3
                      0.2
                     0.2
                      0.2
                                                       0
## 6
                      0.4
```

R formula notation:

```
mod <- lm(Petal.Length ~ Petal.Width + Species + Petal.Width:Species, data = iris)
formula(mod) ## the formula
## Petal.Length ~ Petal.Width + Species + Petal.Width:Species
head(model_frame <- model.frame(mod)) ## the data used for the fit</pre>
    Petal.Length Petal.Width Species
           1.4
                       0.2 setosa
           1.4 0.2 setosa
     1.3 0.2 setosa
      1.5
                 0.2 setosa
## 5
     1.4 0.2 setosa
           1.7
## 6
                    0.4 setosa
head(model.response(model_frame)) ## the response variable
       2 3 4
## 1.4 1.4 1.3 1.5 1.4 1.7
head(model.matrix(mod)) ## the model matrix
    (Intercept) Petal.Width Speciesversicolor Speciesvirginica Petal.Width:Speciesversicolor Petal.Width:Speciesvirginica
                      0.2
## 1
                                         0
                                                        0
                                                                                    0
## 2
                      0.2
## 3
                      0.2
                      0.2
                      0.2
                      0.4
## 6
```

R formula notation:

```
mod <- lm(Petal.Length ~ Petal.Width*Species, data = iris)
formula(mod) ## the formula
## Petal.Length ~ Petal.Width * Species
head(model_frame <- model.frame(mod)) ## the data used for the fit</pre>
    Petal.Length Petal.Width Species
           1.4
                      0.2 setosa
           1.4 0.2 setosa
     1.3 0.2 setosa
     1.5
                 0.2 setosa
## 5
     1.4 0.2 setosa
           1.7
## 6
                    0.4 setosa
head(model.response(model_frame)) ## the response variable
       2 3 4 5 6
## 1.4 1.4 1.3 1.5 1.4 1.7
head(model.matrix(mod)) ## the model matrix
    (Intercept) Petal.Width Speciesversicolor Speciesvirginica Petal.Width:Speciesversicolor Petal.Width:Speciesvirginica
                      0.2
## 1
                                        0
                                                       0
                                                                                   0
                      0.2
## 3
                      0.2
                      0.2
                      0.2
                      0.4
## 6
```

R formula notation:

```
mod <- lm(Petal.Length ~ Petal.Width/Species, data = iris) ## dangerous
formula(mod) ## the formula
## Petal.Length ~ Petal.Width/Species
head(model_frame <- model.frame(mod)) ## the data used for the fit</pre>
    Petal.Length Petal.Width Species
          1.4
                      0.2 setosa
          1.4 0.2 setosa
     1.3 0.2 setosa
     1.5
                 0.2 setosa
## 5
     1.4 0.2 setosa
          1.7
## 6
                    0.4 setosa
head(model.response(model_frame)) ## the response variable
       2 3 4 5 6
## 1.4 1.4 1.3 1.5 1.4 1.7
head(model.matrix(mod)) ## the model matrix
    (Intercept) Petal.Width Petal.Width:Speciesversicolor Petal.Width:Speciesvirginica
                      0.2
## 1
                                                                            0
                      0.2
## 3
                      0.2
                     0.2
                      0.2
## 6
                      0.4
```

R formula notation:

```
mod <- lm(Petal.Length ~ 1, data = iris)</pre>
formula(mod) ## the formula
## Petal.Length ~ 1
head(model_frame <- model.frame(mod)) ## the data used for the fit</pre>
     Petal.Length
            1.4
            1.4
       1.3
        1.5
         1.4
## 5
            1.7
## 6
head(model.response(model_frame)) ## the response variable
        2 3 4 5 6
## 1.4 1.4 1.3 1.5 1.4 1.7
head(model.matrix(mod)) ## the model matrix
     (Intercept)
## 1
## 3
## 6
```

R formula notation:

```
mod <- lm(Petal.Length ~ ., data = iris)
formula(mod) ## the formula
## Petal.Length ~ Sepal.Length + Sepal.Width + Petal.Width + Species
head(model_frame <- model.frame(mod)) ## the data used for the fit</pre>
    Petal.Length Sepal.Length Sepal.Width Petal.Width Species
           1.4
                       5.1
                                  3.5
                                             0.2 setosa
## 1
## 2
           1.4
                       4.9
                                  3.0
                                             0.2 setosa
          1.3
                 4.7
                                  3.2
                                             0.2 setosa
## 3
       1.5
                 4.6
                                  3.1
                                             0.2 setosa
        1.4
## 5
                       5.0
                                  3.6
                                            0.2 setosa
           1.7
## 6
                       5.4
                                  3.9
                                             0.4 setosa
head(model.response(model_frame)) ## the response variable
       2 3 4
## 1.4 1.4 1.3 1.5 1.4 1.7
head(model.matrix(mod)) ## the model matrix
    (Intercept) Sepal.Length Sepal.Width Petal.Width Speciesversicolor Speciesvirginica
                      5.1
                                 3.5
                                            0.2
## 1
                                                              0
## 2
                      4.9
                                 3.0
                                            0.2
## 3
                      4.7
                                3.2
                                            0.2
                      4.6
                                 3.1
                                            0.2
                      5.0
                                 3.6
                                            0.2
## 6
                       5.4
                                 3.9
                                            0.4
```

It is most important to understand the design matrix because the interpretation of the parameters depend on it!

The design matrix depends on:

- your formula
- your data
- the contrasts (for qualitative variables)

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But other several alternative exist; e.g.:

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Note 1: default contrats ("contr.treatment") are easy to interpret!

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The design matrix depends on:

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- your data
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By default, each category is compared to the first one:

But other several alternative exist; e.g.:

Note 1: default contrats ("contr.treatment") are easy to interpret!

Note 2: contrasts do not alter predicted values and thus likelihood, AIC...

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The linear model: understanding the design matrix

Challenge: find out whether these different representations of gender are equivalent or not?

- "boy" vs "girl"
- "male" vs "female"
- 0 vs 1
- 1 vs 2
- TRUE vs FALSE

Note: no need to fit a model, use the function model.matrix() with a formula!

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```
mod <- lm(Petal.Length ~ Petal.Width + Species, data = iris)
```

Simply printing the object provides you with the parameter estimates:

```
mod

## Call:

## Call:

## Im(formula = Petal.Length ~ Petal.Width + Species, data = iris)

##

## Coefficients:

## (Intercept) Petal.Width Speciesversicolor Speciesvirginica

## 1.211 1.019 1.698 2.277
```

outputs

Parameter estimates

mod

```
mod <- lm(Petal.Length ~ Petal.Width + Species, data = iris)
```

Simply printing the object provides you with the parameter estimates:

```
##
## Call:
## lm(formula = Petal.Length ~ Petal.Width + Species, data = iris)
##
## Coefficients:
        (Intercept)
                           Petal.Width Speciesversicolor Speciesvirginica
                                                    1.698
##
              1.211
                                 1.019
                                                                        2.277
```

If you need to work with them, use the specific extractor instead:

```
coefficients(mod) ## or coef(mod)
         (Intercept)
                           Petal.Width Speciesversicolor Speciesvirginica
##
            1.211397
                              1.018712
                                                1.697791
                                                                  2 276693
```

outputs

Parameter estimates

```
mod <- lm(Petal.Length ~ Petal.Width + Species, data = iris)
```

You can also easily extract the covariance matrix of the estimates:

```
vcov(mod)
                    (Intercept) Petal.Width Speciesversicolor Speciesvirginica
## (Intercept)
                 0.004256508 -0.005701674
                                                 0.003303912
                                                                 0.007295083
## Petal.Width
                   -0.005701674 0.023177537
                                                -0.025031740
                                                                -0.041256016
## Speciesversicolor 0.003303912 -0.025031740
                                                 0.032742072
                                                                 0.047410394
## Speciesvirginica 0.007295083 -0.041256016
                                                 0.047410394
                                                                 0.079143501
```

```
mod <- lm(Petal.Length ~ Petal.Width + Species, data = iris)
```

You can also easily extract the covariance matrix of the estimates:

```
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                     (Intercept) Petal.Width Speciesversicolor Speciesvirginica
## (Intercept)
                     0.004256508 -0.005701674
                                                   0.003303912
                                                                    0.007295083
## Petal.Width
                    -0.005701674 0.023177537
                                                  -0.025031740
                                                                   -0.041256016
## Speciesversicolor 0.003303912 -0.025031740
                                                   0.032742072
                                                                    0.047410394
## Speciesvirginica 0.007295083 -0.041256016
                                                   0.047410394
                                                                    0.079143501
```

And thus the standard errors:

```
mod <- lm(Petal.Length ~ Petal.Width + Species, data = iris)
```

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```
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                     (Intercept) Petal.Width Speciesversicolor Speciesvirginica
                     0.004256508 -0.005701674
                                                   0.003303912
                                                                    0.007295083
## (Intercept)
## Petal.Width
                    -0.005701674 0.023177537
                                                   -0.025031740
                                                                   -0.041256016
## Speciesversicolor 0.003303912 -0.025031740
                                                   0.032742072
                                                                    0.047410394
## Speciesvirginica 0.007295083 -0.041256016
                                                   0.047410394
                                                                    0.079143501
```

And thus the standard errors:

You can also get confidence intervals:

```
mod <- lm(Petal.Length ~ Petal.Width + Species, data = iris)
```

You can also easily extract the covariance matrix of the estimates:

```
vcov(mod)
                     (Intercept) Petal.Width Speciesversicolor Speciesvirginica
                    0.004256508 -0.005701674
                                                   0.003303912
                                                                    0.007295083
## (Intercept)
## Petal.Width
                    -0.005701674 0.023177537
                                                   -0.025031740
                                                                    -0.041256016
## Speciesversicolor 0.003303912 -0.025031740
                                                   0.032742072
                                                                    0.047410394
## Speciesvirginica 0.007295083 -0.041256016
                                                   0.047410394
                                                                    0.079143501
```

And thus the standard errors:

You can also get confidence intervals:

```
confint(mod)

## 2.5 % 97.5 %

## (Intercept) 1.0824564 1.340338

## Petal.Width 0.7178294 1.319594

## Speciesversicolor 1.3401762 2.055407

## Speciesvirginica 1.7206988 2.832688
```

Note: that reveals that there are much more information in the object mod than it is being printed!

The model object

The fitted model object is in fact a big list of class "lm":

```
class(mod)
## [1] "lm"

typeof(mod)
## [1] "list"

names(mod)
## [1] "coefficients" "residuals" "effects" "rank" "fitted.values" "assign" "qr"
## [8] "df.residual" "contrasts" "xlevels" "call" "terms" "model"
```

So you can extract information from it; e.g.:

```
mod$df.residual
## [1] 146
```

but it is safer to use extractors if they are available!

Example of other outputs

There are quite a few extractors out there:

```
logLik(mod)
## 'log Lik.' -64.7851 (df=5)
AIC(mod)
## [1] 139.5702
```

Example of other outputs

There are quite a few extractors out there:

```
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## 'log Lik.' -64.7851 (df=5)

AIC(mod)

## [1] 139.5702
```

Here is how you can get the list of S3 methods for the class "lm":

```
methods(class = "lm")
   [1] add1
                       alias
                                                                                  confint
                                                                                                  cooks.distance
                                     anova
                                                     case.names
                                                                   coerce
    [8] deviance
                      dfbeta
                                                    drop1
                                                                   dummy.coef
                                                                                  effects
                                                                                                  extractAIC
                                     dfbetas
## [15] family
                    formula
                                    hatvalues
                                                    influence
                                                                   initialize
                                                                                                 lahels
                                                                                  kappa
## [22] logLik
                      model.frame model.matrix
                                                    nobs
                                                                   plot
                                                                                  predict
                                                                                                 print
## [29] proj
                                     residuals
                                                    rstandard
                                                                   rstudent
                                                                                  show
                                                                                                  simulate
## [36] slotsFromS3
                      summarv
                                     variable.names vcov
## see '?methods' for accessing help and source code
```

Note: the list will change depending on the packages that are attached to the R session!

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tests

Testing coefficients

For LM, simply use summary():

```
summary(mod)
##
## Call:
## lm(formula = Petal.Length ~ Petal.Width + Species, data = iris)
## Residuals:
       Min
                 10 Median
                                  30
                                          Max
## -1.02977 -0.22241 -0.01514 0.18180 1.17449
## Coefficients:
##
                   Estimate Std. Error t value Pr(>|t|)
## (Intercept)
                  1.21140
                               0.06524 18.568 < 2e-16 ***
## Petal.Width
                   1.01871
                              0.15224 6.691 4.41e-10 ***
## Speciesversicolor 1.69779
                              0.18095 9.383 < 2e-16 ***
## Speciesvirginica 2.27669
                               0.28132 8.093 2.08e-13 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## Residual standard error: 0.3777 on 146 degrees of freedom
## Multiple R-squared: 0.9551, Adjusted R-squared: 0.9542
## F-statistic: 1036 on 3 and 146 DF, p-value: < 2.2e-16
```

Testing predictors

Don't use the default anova() function which perform type-I analysis-of-variance:

```
anova (mod)
## Analysis of Variance Table
## Response: Petal.Length
              Df Sum Sq Mean Sq F value Pr(>F)
## Petal.Width 1 430.48 430.48 3016.792 < 2.2e-16 ***
## Species 2 13.01 6.51
                               45.591 4.137e-16 ***
## Residuals 146 20.83
                        0.14
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
```

Instead use the better function Anova() from the package car which perfroms type-II analysis-of-variance:

```
library(car)
Anova (mod)
## Anova Table (Type II tests)
##
## Response: Petal.Length
               Sum Sq Df F value
                                    Pr(>F)
## Petal.Width 6.3892
                      1 44.775 4.409e-10 ***
## Species 13.0113
                      2 45.591 4.137e-16 ***
## Residuals 20.8334 146
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Testing predictors

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Anova(mod)

## Anova Table (Type II tests)

##

## Response: Petal.Length

## Sum Sq Df F value Pr(>F)

## Petal.Width 6.3892 1 44.775 4.409e-10 ***

## Species 13.0113 2 45.591 4.137e-16 ***

## Residuals 20.8334 146

## ---

## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
```

Note: p-values are the same no matter the order of the predictors in the formula for type-II (but not for type-I!).

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Testing the overall model

Before looking at significance for estimates or predictor, always start by checking that your model fits the data better than a null model:

```
mod <- lm(Petal.Length ~ Petal.Width + Species, data = iris)
mod_null <- lm(Petal.Length ~ 1, data = iris)
anova(mod, mod_null)
## Analysis of Variance Table
## Model 1: Petal.Length ~ Petal.Width + Species
## Model 2: Petal.Length ~ 1
  Res.Df
              RSS Df Sum of Sq F Pr(>F)
       146 20.83
       149 464.33 -3 -443.49 1036 < 2.2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Note 1: this was also given at the bottom of the summary table!

Note 2: here using anova() is perfectly fine!

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predictions

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Fitted values

You can easily obtain the prediction for your observation (i.e. fitted values):

```
fitted(mod)[1:39]
                                            5
                                               6
                                                        7
                                                                                       10
## 1.415139 1.415139 1.415139 1.415139 1.415139 1.618882 1.517010 1.415139 1.415139 1.313268 1.415139 1.415139 1.313268
                                   17
                                           18
                                                    19
                                                             20
                                                                      21
## 1.313268 1.415139 1.618882 1.618882 1.517010 1.517010 1.517010 1.415139 1.618882 1.415139 1.720753 1.415139 1.415139
                                           31
                                                    32
                                                             33
                                                                      34
## 1.618882 1.415139 1.415139 1.415139 1.415139 1.618882 1.313268 1.415139 1.415139 1.415139 1.415139 1.313268 1.415139
```

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```
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                                             5
                                                      6
                                                               7
                                                                                         10
                                                                                                  11
## 1.415139 1.415139 1.415139 1.415139 1.415139 1.618882 1.517010 1.415139 1.415139 1.313268 1.415139 1.415139 1.313268
                                            18
                                                     19
                                                              20
                                                                       21
  1.313268 1.415139 1.618882 1.618882 1.517010 1.517010 1.517010 1.415139 1.618882 1.415139 1.720753 1.415139 1.415139
                                                     32
                                                              33
                                                                        34
                                                                                                  37
## 1.618882 1.415139 1.415139 1.415139 1.415139 1.618882 1.313268 1.415139 1.415139 1.415139 1.415139 1.415139
```

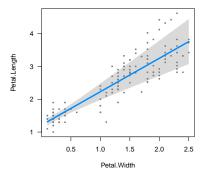
As expected, observations are equal to the fitted values + residuals:

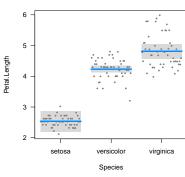
```
head(cbind("response" = model.response(model.frame(mod)),
           "fitted" = fitted(mod).
           "resid" = residuals(mod).
           "fitted + resid" = fitted(mod) + residuals(mod)))
              fitted
                             resid fitted + resid
    response
          1.4 1.415139 -0.01513927
                                              1.4
## 1
         1.4 1.415139 -0.01513927
                                              1.4
         1.3 1.415139 -0.11513927
                                              1.3
          1.5 1.415139 0.08486073
                                              1.5
## 4
                                              1.4
## 5
          1.4 1.415139 -0.01513927
         1.7 1.618882 0.08111841
                                              1.7
## 6
```

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Plotting predictions: fast & dirty

```
library(visreg)
par(mfrow = c(1, 2))
visreg(mod)
```

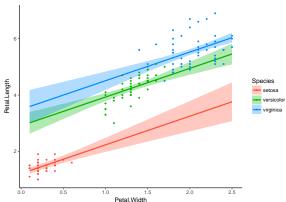




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Plotting predictions: fast & less dirty

```
library(visreg)
library(ggplot2)
visreg(fit = mod, xvar = "Petal.Width", by = "Species", overlay = TRUE, gg = TRUE) +
theme_classic()
```



Note: if you have different quantitative predictors you can specify the value for the non focal predictor using the argument "cond".

Prediction by "hand"

The most difficult step is to create the data frame defining the predictor values:

```
head(data_for_predictions)

## Species Petal.Width

## 1 setosa 0.1000000

## 2 setosa 0.1172414

## 3 setosa 0.1344828

## 4 setosa 0.1517241

## 5 setosa 0.1689655

## 6 setosa 0.1862069
```

```
tail(data_for_predictions)
## Species Petal.Width
## 85 virginica 2.310345
## 86 virginica 2.348276
## 87 virginica 2.386207
## 88 virginica 2.424138
## 89 virginica 2.462069
## 90 virginica 2.500000
```

Prediction by "hand"

The most difficult step is to create the data frame defining the predictor values:

```
library(dplyr)
data for predictions <- iris %>%
                           group by (Species) %>%
                           do(tibble(Petal.Width = seq(min(.$Petal.Width), max(.$Petal.Width), length.out = 30))) %>%
                           as.data.frame()
                                                                        tail(data_for_predictions)
        head(data_for_predictions)
            Species Petal.Width
                                                                               Species Petal.Width
                                                                        ## 85 virginica
                                                                                          2.310345
        ## 1 setosa 0.1000000
                                                                        ## 86 virginica
                                                                                         2.348276
        ## 2 setosa 0.1172414
                                                                        ## 87 virginica
                                                                                         2.386207
        ## 3 setosa 0.1344828
                                                                        ## 88 virginica
                                                                                          2.424138
        ## 4 setosa 0.1517241
                                                                        ## 89 virginica
                                                                                          2.462069
        ## 5 setosa 0.1689655
        ## 6 setosa 0.1862069
                                                                        ## 90 virginica
                                                                                          2.500000
```

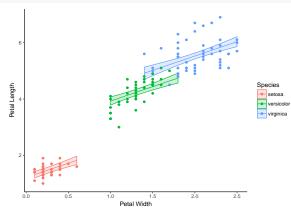
Then, it is easy:

```
pred_mod <- predict(object = mod, newdata = data_for_predictions, interval = "confidence") ## prediction intervals are also possible!
head(pred_mod)

## fit lwr upr
## 1 1.313268 1.198914 1.427622
## 2 1.330832 1.218369 1.443296
## 3 1.348396 1.237613 1.459180
## 4 1.365960 1.256636 1.475284
## 5 1.383524 1.275430 1.491618
## 6 1.401088 1.293986 1.5508190</pre>
```

Prediction by "hand"

```
data_for_plot <- cbind(pred_mod, data_for_predictions)</pre>
ggplot(data = data_for_plot, mapping = aes(x = Petal.Width, y = fit, colour = Species)) +
  geom line() +
  geom_ribbon(mapping = aes(ymin = lwr, ymax = upr, fill = Species), alpha = 0.2) +
  geom_point(data = iris, mapping = aes(y = Petal.Length, x = Petal.Width, colour = Species)) +
  labs(x = "Petal Width", y = "Petal Length") +
  theme_classic()
```



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Assumptions behind linear models

Model structure:

- linearity
- lack of perfect multicollinearity (design matrix of full rank)
- predictor variables have fixed values

Assumptions behind linear models

Model structure:

- linearity
- lack of perfect multicollinearity (design matrix of full rank)
- predictor variables have fixed values

Errors:

- independence (no serial autocorrelation)
- constant variance (homoscedasticity)
- normality

Departure from linearity can originate from a multitude of reasons and can create all kinds of problems.

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Diagnostics:

- thinking
- other assumptions violated

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Solutions:

- ullet different model structure o change the formula
- transform one or several predictors (e.g. polynomials) \rightarrow function poly()
- transform the response (e.g. log and power transformation) → function powerTransform() in car (see later)

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Alternatives:

- non-linear models → function nls or dedicated package (e.g. nlme)
- general additive models → package mgcv

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- ullet general additive models o package mgcv

Quiz: can you express the following models as LM?

•
$$y_i = \hat{\alpha} + \varepsilon_i$$

•
$$y_i = x_i^{\hat{\beta}} + \varepsilon_i$$

•
$$y_i = \hat{\alpha} + \hat{\beta}_1 x_i + \hat{\beta}_2 x_i^2 + \hat{\beta}_3 x_i^3 + \varepsilon_i$$

The number of parameters to be estimated must be equal to the rank of the design matrix.

Caused by having less data than parameters or when there is linear dependence between the column vectors of the design matrix. In such case, some parameters cannot be computed.

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- plot the predictors against each other → function pairs()
- findLinearCombos from the package caret

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- ullet change design matrix (change parameterization or drop redundant effects) o argument formula
- change the experimental design
- collect more data

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none

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Assumptions: lack of perfect multicollinearity

The number of parameters to be estimated must be equal to the rank of the design matrix.

Caused by having less data than parameters or when there is linear dependence between the column vectors of the design matrix. In such case, some parameters cannot be computed.

Diagnostics:

- plot the predictors against each other → function pairs()
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Solutions:

- change design matrix (change parameterization or drop redundant effects) → argument formula
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- collect more data

Alternatives:

none

Note: strong albeit imperfect collinearity is not great either; possible check correlation between estimates (\rightarrow cov2cor(vcov(mod))) and variance inflation factors (\rightarrow vif(mod)).

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The dependent variable are represented by fixed values.

The presence of measurement errors is the main cause of violation. Violation can trigger both estimates and tests to be biased.

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Diagnostics:

thinking & replication

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- often ignored in practice
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Diagnostics:

thinking & replication

Solutions:

- often ignored in practice
- better measurements

Alternatives:

- multipurpose numerical approaches → function optim() or dedicated packages (e.g. nloptr, rjags, nimble, rstan)
- errors-in-variables models → not much directly but any procedure allowing for latent variables can handle that; packages (e.g. sem, lavaan, OpenMX)
- reduced major axis regression → dedicated packages (e.g. lmodel2)

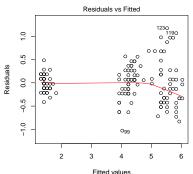
Alexandre Courtiol (IZW) Getting to fit models in R June 2018

A lack of independence (serial autocorrelation) in the residuals can appear if there is a departure from linearity, if data have been sampled non-randomly (e.g. spatial or temporal series), or if there is an overarching structure (e.g. repeated measures within individuals, families, species, ...). Lack of independence increases the risk of false positive (sometimes a lot).

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Diagnostic by eye:

plot(mod, which = 1)



Im(Petal.Length ~ Petal.Width + Species)

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Diagnostic by Durbin-Watson test:

```
durbinWatsonTest(mod) ## from package car (DW varies between 0 & 4, 2 is best, you wish for non-significant p-value)
## lag Autocorrelation D-W Statistic p-value
## 1 0.1313867 1.734855 0.076
## Alternative hypothesis: rho != 0
```

Note: the alternative from the package Imtest offer to rank the residuals according to a variable.

Alexandre Courtiol (IZW) Getting to fit models in R June 2018

A lack of independence (serial autocorrelation) in the residuals can appear if there is a departure from linearity, if data have been sampled non-randomly (e.g. spatial or temporal series), or if there is an overarching structure (e.g. repeated measures within individuals, families, species, ...). Lack of independence increases the risk of false positive (sometimes a lot).

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## 1 0.1313867 1.734855 0.076
## Alternative hypothesis: rho != 0
```

Note: the alternative from the package 1mtest offer to rank the residuals according to a variable.

Solutions:

- transformation or different model structure (see linearity)
- aggregation or sub-sampling

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```
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## lag Autocorrelation D-W Statistic p-value
## 1 0.1313867 1.734855 0.076
## Alternative hypothesis: rho != 0
```

Note: the alternative from the package lmtest offer to rank the residuals according to a variable.

Solutions:

- transformation or different model structure (see linearity)
- aggregation or sub-sampling

Alternatives:

- general additive models (GAM and GAMM) → dedicated package mgcv
- mixed models (LMM and GLMM) → dedicated packages (e.g. spaMM, lme4)

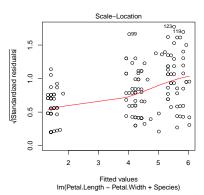
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Heteros(c/k)edasticity can emerge when there is a mean - variance relationship, when there is non independence between observations, when reaction norm changes acording to the treatement. It can create both false positives and false negative.

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Diagnostic by eye:

plot(mod, which = 3)



Heteros(c/k) edasticity can emerge when there is a mean - variance relationship, when there is non independence between observations, when reaction norm changes according to the treatement. It can create both false positives and false negative.

Diagnostic by Breusch-Pagan test:

```
library(lmtest)
bptest(mod) ## BP = df is best, you wish for non-significant p-value
##
## studentized Breusch-Pagan test
##
## data: mod
## BP = 28.571, df = 3, p-value = 2.755e-06
```

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    studentized Breusch-Pagan test
## data: mod
## BP = 28.571, df = 3, p-value = 2.755e-06
```

Solutions: modeling the heteroscedasticity

```
library(spaMM)
mod_heter_spaMM <- fitme(Petal.Length ~ Petal.Width + Species,
                         resid.model = ~ Species.
                         data = iris)
AIC (mod)
## [1] 139.5702
print(AIC(mod_heter_spaMM)) ## much better fit!
          marginal AIC:
               87.84896
```

Alexandre Courtiol (IZW) Getting to fit models in R June 2018

Heteros(c/k)edasticity can emerge when there is a mean - variance relationship, when there is non independence between observations, when reaction norm changes acording to the treatement. It can create both false positives and false negative.

Diagnostic by Breusch-Pagan test:

```
library(lmtest)
bptest(mod) ## BP = df is best, you wish for non-significant p-value

##

## studentized Breusch-Pagan test
##

## data: mod
## BP = 28.571, df = 3, p-value = 2.755e-06
```

Solutions: modeling the heteroscedasticity

Alternatives:

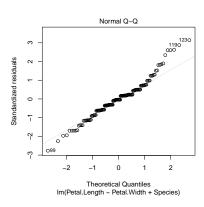
GLM (if stemming from an expected relationship between mean and variance) → function glm

The distribution of residuals can be skewed, this is often caused by the presence of outliers, and/or when the process generating the data is very different from normal (e.g. Poisson, Binomial...).

The distribution of residuals can be skewed, this is often caused by the presence of outliers, and/or when the process generating the data is very different from normal (e.g. Poisson, Binomial...).

Diagnostic by eye:

plot(mod, which = 2)



Alexandre Courtiol (IZW)

June 2018

The distribution of residuals can be skewed, this is often caused by the presence of outliers, and/or when the process generating the data is very different from normal (e.g. Poisson, Binomial...).

Diagnostic by test (many test are possible):

```
shapiro.test(mod$residuals) ## stat = 1 when normal, you wish for non-significant p-value
##
## Shapiro-Wilk normality test
##
## data: mod$residuals
## W = 0.96925, p-value = 0.001924
```

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Solutions:

- transformation or different model structure (see linearity)
- taking outliers out (mindfully!)

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##
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##
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```

Solutions:

- transformation or different model structure (see linearity)
- taking outliers out (mindfully!)

Alternatives:

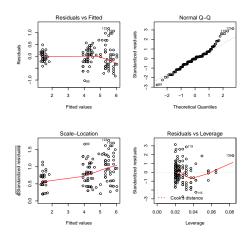
ullet GLM (if stemming from the data generating process) o function glm

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Assumptions: simple glimpse

You can check all assumptions about the erros at once:

```
par(mfrow = c(2, 2))
plot(mod)
```



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There is a powerful function in \mathbf{R} :

```
influence.measures(mod)
## Influence measures of
    lm(formula = Petal.Length ~ Petal.Width + Species, data = iris) :
##
         dfb.1 dfb.Pt.W dfb.Spcsvrs dfb.Spcsvrg
                                                      dffit cov.r cook.d
                                                                              hat inf
## 1
      -0.005155 0.000756
                               0.00102
                                          0.000367 -0.00582 1.049 8.51e-06 0.0203
## 2
      -0.005155 0.000756
                               0.00102
                                          0.000367 -0.00582 1.049 8.51e-06 0.0203
      -0.039218 0.005750
                               0.00773
                                          0.002791 -0.04424 1.046 4.92e-04 0.0203
## 3
       0.028900 -0.004237
                              -0.00569
                                         -0.002056 0.03260 1.048 2.67e-04 0.0203
## 4
                               0.00102
                                          0.000367 -0.00582 1.049 8.51e-06 0.0203
## 5
      -0.005155 0.000756
                              -0.02152
                                         -0.018998 0.03386 1.052 2.89e-04 0.0239
## 6
       0.017579 0.013609
      -0.032568 -0.006861
                               0.01940
                                          0.015075 -0.04511 1.047 5.12e-04 0.0205
## 7
## 8
       0.028900 -0.004237
                              -0.00569
                                         -0.002056 0.03260 1.048 2.67e-04 0.0203
## 9
      -0.005155 0.000756
                               0.00102
                                          0.000367 -0.00582 1.049 8.51e-06 0.0203
## 10
       0.075521 -0.029709
                                          0.015059 0.07734 1.045 1.50e-03 0.0235
                               0.00591
## 11
       0.028900 -0.004237
                              -0.00569
                                         -0.002056 0.03260 1.048 2.67e-04 0.0203
## 12
       0.062998 -0.009237
                              -0.01241
                                         -0.004483 0.07106 1.042 1.27e-03 0.0203
       0.035054 -0.013790
                               0.00275
## 13
                                          0.006990 0.03590 1.051 3.24e-04 0.0235
      -0.086276 0.033940
                              -0.00676
                                         -0.017203 -0.08835 1.043 1.96e-03 0.0235
## 14
## 15
      -0.073339 0.010753
                               0.01445
                                          0.005218 -0.08273 1.040 1.72e-03 0.0203
## 16
      -0.025768 -0.019948
                               0.03155
                                          0.027847 -0.04964 1.050 6.20e-04 0.0239
      -0.069267 -0.053624
                               0.08480
                                          0.074856 -0.13343 1.032 4.46e-03 0.0239
## 17
      -0.032568 -0.006861
                                          0.015075 -0.04511 1.047 5.12e-04 0.0205
## 18
                               0.01940
       0.050956 0.010735
                              -0.03035
                                         -0.023587 0.07058 1.042 1.25e-03 0.0205
## 19
      -0.004733 -0.000997
                                          0.002191 -0.00656 1.049 1.08e-05 0.0205
## 20
                               0.00282
## 21
       0.097189 -0.014250
                              -0.01914
                                         -0.006916 0.10963 1.033 3.01e-03 0.0203
## 22
      -0.025768 -0.019948
                               0.03155
                                          0.027847 -0.04964 1.050 6.20e-04 0.0239
## 23
      -0.141957 0.020814
                               0.02796
                                          0.010101 -0.16013 1.014 6.40e-03 0.0203
      -0.003221 -0.005781
                                          0.007085 -0.00986 1.060 2.45e-05 0.0305
## 24
                               0.00761
## 25
       0.166055 -0.024347
                              -0.03271
                                         -0.011816 0.18732 1.002 8.73e-03 0.0203
        0.062998 -0.009237
## 26
                              -0.01241
                                         -0.004483
                                                    0.07106 1.042 1.27e-03 0.0203
```

Interpretation of the output from influence.measures(mod):

- \bullet dfb.1_ \to extent to which the intercept changes if a given observation is dropped
- ullet dfb.Pt.W o extent to which the slope for Petal.Width changes if a given observation is dropped
- dfb.Spcsvrs → extent to which the estimate for versicolor changes if a given observation is dropped
- ullet dfb.Spcsvrg o extent to which the estimate for virginica changes if a given observation is dropped

Note: for the df-betas, the name would change for another model as they use the abbreviated name of the estimates:

```
abbreviate(stats:::variable.names.lm(mod))
## (Intercept)    Petal.Width Speciesversicolor    Speciesvirginica
## "(In)"    "Pt.W"    "Spcsvrg"    "Spcsvrg"
```

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- dfb.Spcsvrs → extent to which the estimate for versicolor changes if a given observation is dropped
- ullet dfb.Spcsvrg o extent to which the estimate for virginica changes if a given observation is dropped
- dffit

 extent to which the predicted y-values changes if a given observation is dropped (scaled by the standard deviation of the fit at the point)

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- ullet cook.d o F statistics comparing simultaneously the changes in all estimates when the observation is dropped or not

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- hat → diagonal element of the hat matrix (the hat values); extent to which an observation is unusual in terms of X values (leverage)

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```

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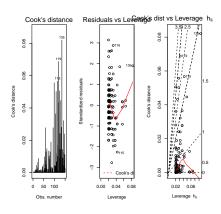
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- hat → diagonal element of the hat matrix (the hat values); extent to which an observation is unusual in terms of X values (leverage)
- ullet inf o some overal add hoc receipe to spot influential observation (not to be taken too seriously)

Note: for the df-betas, the name would change for another model as they use the abbreviated name of the estimates:

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There are also plotting possibilitites:

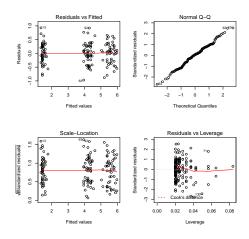
```
par(mfrow = c(1, 3))
plot(mod, which = 4:6)
```



Assumptions: simple glimpse at residuals

What would it look like if it was perfect?

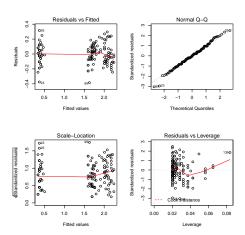
```
iris$Fake.Petal.Length <- simulate(object = mod)[, 1] ## redo it, it will change each time!
mod_perfect <- lm(Fake.Petal.Length ~ Petal.Width + Species, data = iris)
par(mfrow = c(2, 2))
plot(mod_perfect)</pre>
```



Assumptions: fixing iris

Fixing attempt:

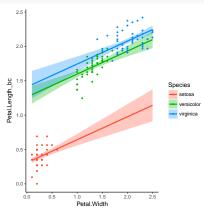
```
bc <- powerTransform(mod)
iris$Petal.Length_bc <- bcPower(iris$Petal.Length, lambda = bc$lambda)
mod_bc <- lm(Petal.Length_bc ~ Petal.Width + Species, data = iris)
par(mfrow = c(2, 2))
plot(mod_bc)</pre>
```



Assumptions: fixing iris

Plotting predictions:

```
visreg(fit = mod_bc, xvar = "Petal.Width", by = "Species", overlay = TRUE, gg = TRUE) +
theme_classic()
```

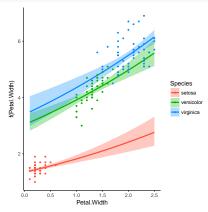


Note: that is not very useful because it is on the BoxCoxed scale!

Assumptions: fixing iris

Plotting predictions:

```
visreg(fit = mod_bc, xvar = "Petal.Width", by = "Species", overlay = TRUE, gg = TRUE,
       trans = function(x) bcnPowerInverse(x, lambda = bc$lambda, gamma = 0), partial = TRUE) +
 theme classic()
```



Note: that is not very useful because it is on the BoxCoxed scale!

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The generalised linear model: what for?

GLM are used for fitting data generating processes for which a relationship between mean and variance is expected.

The generalised linear model: what for?

GLM are used for fitting data generating processes for which a relationship between mean and variance is expected.

That includes the analysis of:

- binary events (probabilities)
- binomial events (probabilities)
- Poisson processes (counts)
- negative binomail processes (counts)
- variances (positive continuous)

The generalised linear model: specifications

Definition:

$$\mathsf{Y} = \mathsf{g}^{-1}(\widehat{\eta}) + \varepsilon = \mathsf{g}^{-1}(\mathsf{X}\widehat{\beta}) + \varepsilon$$

with:

•
$$\hat{\eta}_i = \hat{\beta}_0 + \hat{\beta}_1 \times x_{1,i} + \hat{\beta}_2 \times x_{2,i} + \dots + \hat{\beta}_p \times x_{p,i}$$

- $E(Y) = \mu = g^{-1}(\eta)$
- $Var(Y) = \phi V(\mu)$

Notation:

- ullet η the linear predictor
- g the link function (g^{-1} is sometimes called the mean function)
- V the variance function
- ullet ϕ is the dispersion parameter

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The generalised linear model: specifications

Definition:

$$Y = g^{-1}(\widehat{\eta}) + \varepsilon = g^{-1}(X\widehat{\beta}) + \varepsilon$$

with:

•
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- $E(Y) = \mu = g^{-1}(\eta)$
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Notation:

- \bullet η the linear predictor
- g the link function (g^{-1} is sometimes called the mean function)
- V the variance function
- ullet ϕ is the dispersion parameter

This is identical to the LM if:

- $\mu = g^{-1}(\eta) = \eta$, thus if g is the identity function
- ullet $\phi=\sigma^2$, thus if the dispersion parameter equals the error variance
- $V(\mu) = 1$, thus if the variance function is constant

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R formula notation:

mod <- lm(Petal.Length ~ Petal.Width, data = iris)</pre>

R formula notation:

```
mod <- lm(Petal.Length ~ Petal.Width, data = iris)</pre>
```

```
formula(mod) ## the formula
## Petal.Length ~ Petal.Width
```

0.2

0.4

R formula notation:

1.4

1.7

5

6

```
mod <- lm(Petal.Length - Petal.Width, data = iris)

formula(mod) ## the formula

## Petal.Length - Petal.Width

head(model_frame <- model.frame(mod)) ## the data used for the fit

## Petal.Length Petal.Width

## 2 1.4 0.2

## 3 1.3 0.2

## 3 1.5 0.2
```

R formula notation:

```
mod <- lm(Petal.Length ~ Petal.Width, data = iris)</pre>
formula(mod) ## the formula
## Petal.Length ~ Petal.Width
head(model_frame <- model.frame(mod)) ## the data used for the fit</pre>
    Petal.Length Petal.Width
           1.4
                       0.2
           1.4
                 0.2
      1.3
                 0.2
       1.5
                  0.2
        1.4
                   0.2
## 5
            1.7
## 6
                       0.4
head(model.response(model_frame)) ## the response variable
        2 3 4 5 6
## 1.4 1.4 1.3 1.5 1.4 1.7
```

0.4

R formula notation:

6

```
mod <- lm(Petal.Length ~ Petal.Width, data = iris)
formula(mod) ## the formula
## Petal.Length ~ Petal.Width
head(model_frame <- model.frame(mod)) ## the data used for the fit</pre>
    Petal.Length Petal.Width
           1.4
                       0.2
           1.4
                 0.2
      1.3
                 0.2
      1.5
                 0.2
       1.4
                   0.2
## 5
           1.7
## 6
                       0.4
head(model.response(model_frame)) ## the response variable
       2 3 4 5 6
## 1.4 1.4 1.3 1.5 1.4 1.7
head(model.matrix(mod)) ## the model matrix
    (Intercept) Petal.Width
## 1
                      0.2
                      0.2
## 3
                      0.2
                      0.2
                      0.2
```

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