

# Landsepi: List of assumptions and parameters



Stochastic, spatially-explicit, demo-genetic model simulating the spread and evolution of a plant pathogen in a heterogeneous landscape to assess resistance deployment strategies.

## Assumptions:

In orange assumptions that can be relaxed with appropriate parameterization.

1. The spatial unit is a polygon, i.e. a piece of land delimited by boundaries and possibly cultivated with a crop. Such crop may be host or non-host, and the polygon is considered a homogeneous mixture of host individuals (i.e. there is no intra-polygon structuration). An agricultural field may be composed of a single or several polygons.
2. A host ‘individual’ is an infection unit (i.e. it can be infected by one and only one pathogen propagule, there is no co-infection) and may correspond to a given amount of plant tissue (where a local infection may develop, e.g. fungal lesion) or a whole plant (e.g. systemic viral infection). In the first case, plant growth increases the amount of available plant tissue (hence the number of individuals) during the cropping season. Plant growth is deterministic (logistic growth) and only healthy individuals (state H) contribute to plant growth (castrating pathogen).
3. Host individuals are in one of these four categories: H (healthy), E (exposed and latent, i.e. infected but not infectious nor symptomatic), I (infectious and symptomatic), or R (removed, i.e. epidemiologically inactive).
4. The decreasing availability of healthy host tissues (as epidemics spread) makes pathogen infection less likely (i.e. density-dependence due to plant architecture).
5. Hosts are cultivated (i.e. sown/planted and harvested), thus there is no host reproduction, dispersal and natural death.
6. Environmental and climate conditions are constant, and host individuals of a given genotype are equally susceptible to disease from the first to the last day of every cropping season.
7. Crop yield depends on the average amount of producing host individuals during the cropping season and does not depend on the time of epidemic peak. Only healthy individuals (state H) contribute to crop yield.
8. Cultivars may be treated with chemicals which reduce the pathogen infection rate (contact treatment). Treatment efficiency decreases with host growth (i.e. new biomass is not protected by treatments) and time (i.e. pesticide degradation). Cultivars to be treated and dates of chemical applications are fixed prior to simulations, but only polygons where disease severity exceeds a given threshold (possibly 0) are treated.

9. Components of a mixture are independent each other (i.e. there is neither plant-plant interaction nor competition for space, and harvests are segregated). If one component is treated with a chemical, it does not affect other components.
10. The pathogen is haploid.
11. Initially, the pathogen is not adapted to any source of resistance, and is only present on susceptible hosts (at state I).
12. Pathogen dispersal is isotropic (i.e. equally probable in every direction).
13. Boundaries of the landscape are reflective: propagules stay in the system as if it was closed.
14. Pathogen reproduction can be purely clonal, purely sexual, or mixed (alternation of clonal and sexual reproduction).
15. If there is sexual reproduction (or gene recombination), it occurs only between parental infections located in the same polygon and the same host genotype (i.e. cultivar). At that scale, the pathogen population is panmictic (i.e. all pairs of parents have the same probability to occur). The propagule production rate of a parental pair is the sum of the propagule production rates of the parents. For a given parental pair, the genotype of each propagule is issued from random loci segregation of parental qualitative resistance genes. For each quantitative resistance gene, the value of each propagule trait is issued from a normal distribution around the average of the parental traits, following the infinitesimal model (Fisher 1919).
16. All types of propagule (i.e. either clonal or sexual) share the same pathogenicity parameters (e.g. infection rate, latent period duration, etc.) but each of them has their own dispersal and survival abilities (see after).
17. At the end of each cropping season, pathogens experience a bottleneck representing the off-season and then propagules are produced (either via clonal or sexual reproduction). The probability of survival is the same every year and in every polygon. Clonal propagules are released during the following season only, either altogether at the first day of the season, or progressively (in that case the day of release of each propagule is sampled from a uniform distribution). Sexual propagules are gradually released during several of the following seasons (between-season release). The season of release of each propagule is sampled from an exponential distribution, truncated by a maximum viability limit. Then, the day of release in a given season is sampled from a uniform distribution (within-season release).
18. Pathogenicity genes mutate independently from each other.
19. Pathogen adaptation to a given resistance gene consists in restoring the same aggressiveness component as the one targeted by the resistance gene.
20. If a fitness cost penalises pathogen adaptation to a given resistance gene, this cost is on all hosts with possibly a relative advantage on hosts carrying the resistance gene. It consists in a reduction in the same aggressiveness component as the one targeted by the resistance gene.
21. When there is a delay for activation of a given resistance gene (APR), the age of activation is the same for all hosts carrying this gene and located in the same polygon.
22. Variances of the durations of the latent and the infectious periods of the pathogen are not affected by plant resistance.

Parameter	Notation in articles	Possible values	Default value			Meaning	Remarks
			Rust	Mildew	Black sigatoka		
<b>SIMULATION PARAMETERS</b>							
<b>Nyears</b>	Y	N*	20	20	20	Number of cropping seasons (e.g. years)	
<b>nTSpY</b>	T	N*	120	120	182	Number of time steps per cropping season (e.g. days)	Duration of the epidemic season
<b>seed</b>		N	12345	12345	12345	Seed for random number generator	
<b>writeTXT</b>		{TRUE ; FALSE}	TRUE	TRUE	TRUE	Whether or not outputs must be written in text files	
<b>graphic</b>		{TRUE ; FALSE}	TRUE	TRUE	TRUE	Whether or not graphics must be generated	
<b>videoMP4</b>		{TRUE ; FALSE}	FALSE	FALSE	FALSE	Whether or not a video must be generated	Works only if graphic=TRUE and epid_outputs="audpc" (or epid_outputs="all"). Requires ffmpeg library. Up to 9 different croptypes can be properly represented.
<b>keepRawResults</b>		{TRUE ; FALSE}	FALSE	FALSE	FALSE	Whether or not raw outputs (binary files) must be kept	Keeps 6 binary files (H, Hjuv, L, I, R, P) per simulated year
<b>PATHOGEN PARAMETERS</b>							
			Basic pathogen aggressiveness components on a susceptible host for a pathogen genotype not adapted to resistance				
<b>infection_rate</b>	e <sub>max</sub>	[0 ; 1]	0.40	0.90	0.02	Maximal expected infection rate of a propagule on a healthy individual	Given that the propagule landed on the host individual
<b>propagule_prod_rate</b>	r <sub>max</sub>	ℝ <sup>+</sup>	3.125	2.0	90.9	Maximal expected effective propagule production rate per time step and per infectious individual	Number of viable propagules that disperse and land on another host individual
<b>latent_period_mean</b>	γ <sub>min</sub>	ℝ <sup>+</sup>	10	7	25.5	Minimal expected latent period duration	Parameter of the gamma distribution of the latent period (in time steps)
<b>latent_period_var</b>	γ <sub>var</sub>	ℝ <sup>+</sup>	9	8	1.5	Variance of the latent period duration	Parameter of the gamma distribution of the latent period (in time steps)
<b>infectious_period_mean</b>	Υ <sub>max</sub>	ℝ <sup>+</sup>	24	14	22	Maximal expected infectious period duration	Parameter of the gamma distribution of the infectious period (in time steps)
<b>infectious_period_var</b>	Υ <sub>var</sub>	ℝ <sup>+</sup>	105	22	14	Variance of the infectious period duration	Parameter of the gamma distribution of the infectious period (in time steps)
<b>survival_prob</b>	λ	[0 ; 1]	10 <sup>-4</sup>	10 <sup>-4</sup>	0.5	Off-season survival probability of a propagule	Determine the size of the bottleneck at the end of the cropping season.

							Vector of size Nyears*Ncroptypes to give a specific probability for every year and croptype.
repro_sex_prob		[0 ; 1]	0	0	0	Probability for an infectious individual to reproduce via sex (same value for every time step – can also be entered as a vector to give a different probability for every time step)	=0 for purely clonal reproduction =1 for purely sexual reproduction between 0 and 1 for both reproduction modes at the same time Vector of size (nTSpY+1) to simulate alternation between clonal and sexual reproduction
sex_propagule_release_mean		$\mathbb{R}^{**}$	1	1	1	Average number of cropping seasons (e.g. years) after which a sexual propagule is released	Parameter of the exponential distribution of the season of release.
sex_propagule_viability_limit		$\mathbb{N}^*$	1	5	1	Maximum number of cropping seasons (e.g. years) up to which a sexual propagule is viable	All sexual propagules are released before the viability limit (i.e. no simulated mortality).
sigmoid_kappa	$\kappa$	$\mathbb{R}^+$	5.33	5.33	5.33	Kappa parameter of the sigmoid contamination function	$\kappa=0$ and $\sigma=1$ for linear relationship
sigmoid_sigma	$\sigma$	$\mathbb{R}^+$	3	3	3	Sigma parameter of the sigmoid contamination function	$\sigma=0$ to relax density-dependence assumption
sigmoid_plateau		[0 ; 1]	1	1	1	Plateau parameter of the sigmoid contamination function	
clonal_propagule_gradual_release		{TRUE ; FALSE}	FALSE	TRUE	FALSE	Whether or not clonal propagules surviving the bottleneck are gradually released along the following cropping season.	if FALSE, propagules are released at the first time step of the following season. if TRUE, the day of release is sampled from a uniform distribution, with parameters [0, nTSpY].
<b>CROPTYPE PARAMETERS</b>						Characteristics of the landscape and of each cultivated croptype	
GPKG Landscape						Landscape in shapefile format, containing spatial coordinates of every polygon	Built-in simulated landscapes are available. Areas must be expressed in square meters.
aggreg	$\alpha$	$\mathbb{R}^+$				Level of spatial aggregation of the landscape	Units relative to the maximal distance between two polygons of the landscape: between 0 and 0.1 for fragmented landscape, 0.1 and 0.5 for balanced landscape, 0.5 and 3 for aggregated landscape, and above 3 for bloc planting
algo		{"exp"; "periodic"; "random"}				Algorithm used to control spatial aggregation in croptype allocation	Parameter “aggreg” is ignored for random allocation. Algorithm “exp” is preferable for big landscapes.
croptypeName						Name of the croptype	
prop	$\phi$	[0 ; 1]				Proportion of the landscape surface cultivated with the croptype (cropping ratio)	A minimum of 1 polygon is allocated to each croptype

<b>rotation_period</b>		N				Number of years before rotation of the landscape	There is no rotation if rotation_period=0 or rotation_period=Nyears
<b>rotation_sequence</b>		N				Indices of cultivated croptypes for each period of the rotation sequence	There is no rotation if the list contains only one element
<b>rotation_realloc</b>		{TRUE ; FALSE}	FALSE	FALSE	FALSE	Whether or not a new random allocation of croptypes is performed when the landscape is rotated.	rotation_realloc=FALSE for static allocation ; rotation_realloc=TRUE for dynamic allocation
<b>DISPERSAL PARAMETERS</b>						Probability of dispersal from any polygon to any other polygon of the landscape (total number of polygons in the landscape is $N_{poly}$ )	
<b>disp_patho_clonal</b>	$\mu$	$[0 ; 1]^{N_{poly} \times N_{poly}}$				Pathogen dispersal matrix for propagules produced by clonal reproduction.	To compute from dispersal kernel via RCALI package. Lines of the matrix can be normalised to sum to 1 (reflective boundaries); otherwise propagules dispersing outside the landscape are lost (absorbing boundaries). Dispersal matrices for rust pathogens are available for each built-in landscape. Identity matrix = no dispersal
<b>disp_patho_sex</b>		$[0 ; 1]^{N_{poly} \times N_{poly}}$				Pathogen dispersal matrix for propagules produced by sexual reproduction	To compute from dispersal kernel via RCALI package. Lines of the matrix can be normalised to sum to 1 (reflective boundaries); otherwise propagules dispersing outside the landscape are lost (absorbing boundaries). Identity matrix = no dispersal
<b>CULTIVAR PARAMETERS</b>						Characteristics of each host genotype as if cultivated in pure crop	
<b>cultivarName</b>					"Cavendish"	Name of the cultivar	Avoid using spaces in cultivar names A "cultivar index" is assigned to each cultivarName (starting from 0)
<b>initial_density</b>	$C^0$	$\mathbb{R}^+$	0.1	1	0.9	Host individuals density (in pure crop) per surface unit at the beginning of the cropping season	$C^0=0$ if the crop is not cultivated. $C^0 \leq C^{max}$
<b>max_density</b>	$C^{max}$	$\mathbb{R}^{+*}$	2	20	1.8	Maximum host individuals density (in pure crop) per surface unit at the end of the cropping season	$C^0=C^{max}$ if individual hosts are whole plants or do not grow
<b>growth_rate</b>	$\delta$	$[0 ; 1]$	0.10	0.10	0.02	Host growth rate	$\delta=0$ if individual hosts are whole plants or do not grow
<b>yield_H</b>	$y_H$	$\mathbb{R}^+$	2.5	6.7	46.8	Theoretical yield in pure crop (in weight or volume unit / ha / cropping season) associated with the sanitary status 'H'	$yield\_H=0$ if the crop is not a cash crop

<b>yield_L</b>	$y_L$	$\mathbb{R}^+$	0	6.7	46.8	Theoretical yield in pure crop (in weight or volume unit / ha / cropping season) associated with the sanitary status 'L'	$yield\_L > 0$ for non-castrating pathogen
<b>yield_I</b>	$y_I$	$\mathbb{R}^+$	0	6.7	0	Theoretical yield in pure crop (in weight or volume unit / ha / cropping season) associated with the sanitary status 'I'	$yield\_I > 0$ for tolerant cultivar
<b>yield_R</b>	$y_R$	$\mathbb{R}^+$	0	0	0	Theoretical yield in pure crop (in weight or volume unit / ha / cropping season) associated with the sanitary status 'R'	$yield\_R > 0$ for tolerant cultivar
<b>planting_cost</b>		$\mathbb{R}^+$	225	5481	0	Planting costs in pure crop (in monetary units / ha / cropping season)	
<b>market_value</b>		$\mathbb{R}^+$	200	600	0	Market value of the product (in monetary units / weight or volume unit)	
<b>GENE PARAMETERS</b>						Characteristics of each plant resistance gene and of each corresponding pathogen pathogenicity gene	
<b>geneName</b>						Name of the resistance gene	
<b>target_trait</b>	w	{"IR" ; "LAT" ; "IP" ; "PR"}	"IR"	"IR"	"IR"	Aggressiveness component targeted by the resistance gene	$IR=e_{max}$ ; $LAT=\gamma_{min}$ ; $IP=Y_{max}$ ; $PR=r_{max}$
<b>efficiency</b>	$\rho$	[0 ; 1]	1.00	1.00	1.00	Efficiency of the resistance gene (percentage of reduction of the targeted aggressiveness component: IR, 1/LAT, IP or PR)	$\rho=1$ for complete resistance, $\rho<1$ for partial resistance, $\rho=0$ for no resistance
<b>age_of_activ_mean</b>	$t_{exp}^a$	$\mathbb{R}^+$	0	0	0	Expected delay to resistance activation (for APRs)	$age\_of\_activ\_mean > 0$ for APRs
<b>age_of_activ_var</b>	$t_{var}^a$	$\mathbb{R}^+$	0	0	0	Variance of the delay to resistance activation (for APRs)	$age\_of\_activ\_var = 0$ for deterministic activation at $age\_of\_activ\_mean$
<b>mutation_prob</b>	$\tau$	[0 ; 1]	$10^{-7}$	$10^{-7}$	$10^{-7}$	Probability for a pathogenicity gene to mutate	$\tau=0$ for no evolution
<b>Nlevels_aggressiveness</b>	Q	$\mathbb{N}^*$	2	2	2	Number of adaptation levels related to each resistance gene (i.e. 1 + number of required mutations for a pathogenicity gene to fully adapt to the corresponding resistance gene)	Q=1 for no adaptation; Q=2 for adaptation via a single mutation; Q>2 for gradual adaptation to resistance. Careful, value of Q severely impacts computational time.
<b>adaptation_cost</b>	$\theta$	[0 ; 1]	0.50	0.50	0.50	Fitness penalty paid by a pathogen genotype fully adapted to the resistance gene on all hosts	$\theta = 0$ for no cost ; $\theta = 1$ for matching allele model
<b>relative_advantage</b>	$\theta$	[0 ; 1]	0.50	0.50	0.50	Fitness advantage of a pathogen genotype fully adapted to the resistance gene on hosts carrying this gene, relative to those that do not carry this gene	See Leonard KJ (1977) Selection Pressures and Plant Pathogens. Annals of the New York Academy of Sciences, 287, 207-222. <a href="https://doi.org/10.1111/j.1749-6632.1977.tb34240.x">https://doi.org/10.1111/j.1749-6632.1977.tb34240.x</a> for details



PARAMETERS							
epid_outputs			"all"	"all"	"all"	<p>Epidemiological and economic outputs to generate:</p> <ul style="list-style-type: none"> <li>▪ "audpc" : Area Under Disease Progress Curve</li> <li>▪ "audpc_rel" : relative Area Under Disease Progress Curve</li> <li>▪ "gla" : Green Leaf Area</li> <li>▪ "gla_rel" : relative Green Leaf Area</li> <li>▪ "eco_cost" : operational crop costs</li> <li>▪ "eco_yield" : total crop yield</li> <li>▪ "eco_product" : total crop products</li> <li>▪ "eco_margin" : margin (products – operational costs)</li> <li>▪ "contrib" : contribution of pathogen genotypes to LIR dynamics</li> <li>▪ "HLIR_dynamics", "H_dynamics", "IR_dynamics", etc. : epidemic dynamics related to the specified sanitary status</li> <li>▪ "all" : compute all these outputs</li> <li>▪ "" : none of these outputs will be generated.</li> </ul>	
evol_outputs			"all"	"all"	"all"	<p>Evolutionary outputs to generate :</p> <ul style="list-style-type: none"> <li>▪ "evol_patho" : evolution of pathogen genotypes</li> <li>▪ "evol_patho_bi": pathogen genotype frequencies after the off-season (after bottleneck)</li> <li>▪ "evol_aggr" : evolution of pathogen aggressiveness (i.e. phenotype)</li> <li>▪ "durability" : durability of resistance genes</li> <li>▪ "all" : compute all these outputs</li> <li>▪ "" : none of these outputs will be generated.</li> </ul>	
audpc100S		$\mathbb{R}^{**}$	0.76	8.48	0.53	AUDPC in a fully susceptible landscape	(used as reference value for graphics and video)
thres_breakdown		$\mathbb{N}^*$	50,000	50,000		Threshold (i.e. number of infections) above which a pathogen genotype is unlikely to go extinct and resistance is considered broken down	Used to characterise the time to invasion of resistant hosts (several values are computed if several thresholds are given in a vector)