

Bastu - Swab's Dental Health Report





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Dental Disease 101

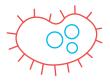
Did you know?



Dental disease affects 50-90% of cats over the age of four.

Fortunately, most dental diseases are preventable with a good dental care routine. They are also mostly treatable, if caught early. The microbes in a cat's mouth can inform us of developing dental issues, before they are advanced enough to be visible by a veterinarian during a routine checkup. The Basepaws Dental Health test looks for microbial signatures associated with three of the most common dental conditions in cats - Periodontal Disease, Tooth Resorption, Halitosis (bad breath).

The oral microbiome



Environmental factors and various food sources make the feline oral cavity a fascinating place, characterized by unique interactions between a cat's mouth and the microbes within it (the oral microbiome). The almost constant exposure to foreign microbial organisms has made the oral microbiome fiercely competitive. Once in a while, pathogenic microbes manage to colonize parts of the oral cavity which can be associated with dental problems.

The feline oral microbiome can reveal information about developing dental issues.

The oral microbiome also has implications for general health.

Can the oral microbiome change?

YES! The oral microbiome is not static.

Different factors such as diet (dry versus wet food), environment (indoor versus outdoor), supplement intake, medications (particularly antibiotics) and dental care routine can all influence the composition of the oral microbiome.

This is why testing early and testing often is key for optimal dental health!



Bastu - Swab's Dental health summary

How does this test work? We used our oral microbiome database containing healthy cats and cats suffering from periodontal disease, tooth resorption or halitosis to identify a set of predictive microbes whose compositional abundance is associated with each condition. Based on these results, we developed a 0 - 10 risk score system for each condition. The results below show Bastu - Swab's overall risk for each of the three conditions, as well as a breakdown of the predictive microbes whose compositional abundance is associated with high, medium or low risk for each dental condition. The purple line and the number next to it indicate your cat's overall risk score for each condition.

medium risk: >3.3 - 6.6 high risk: >6.6 - 10

Risk for periodontal disease

Periodontal disease affects the tissues surrounding the teeth. Initial stages are classified as gingivitis, while advanced cases are known as periodontitis.



Risk for tooth resorption

Tooth resorption is a relatively common condition characterized by progressive dentin erosion.



Risk for bad breath (halitosis)

When bad breath is a persistent problem for a cat, this could be indicative of more serious general health issues.

OVERALL RISK:

low risk: 0 - 3.3

LOW

MEDIUM

→ HIGH



What's next?

- You are strongly advised to adopt a daily dental care routine for Bastu - Swab
- Consider supplementing Bastu Swab's routine with products accepted by the <u>Veterinary Oral Health Council (VOHC)</u>
- Schedule an appointment with your veterinarian in the next month

Next recommended dental health test in: 3-6 months



What are Bastu - Swab's health implications?

Implications of having elevated periodontal disease risk:

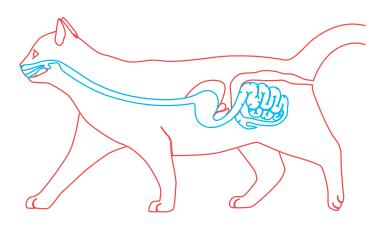
Your cat's elevated risk for periodontal disease could potentially have negative implications for their general health. Periodontal disease can sometimes be associated with a viral infection (feline leukemia virus, feline immunodeficiency virus, feline calicivirus), although it is most commonly caused by buildup of plaque and tartar on the teeth's surface - a great breeding ground for pathogenic microbes. With the progression of this dental disease, pathogenic microbes can enter the bloodstream and travel to other organs.

This is why, periodontal disease is sometimes associated with a higher incidence of chronic kidney disease, cardiovascular problems, diabetes mellitus and some autoimmune diseases.

Implications of having elevated tooth resorption risk:

Unfortunately, not much is currently known about the effect of tooth resorption (FORL) on general health. Sometimes, FORL is observed in cats who also suffer from periodontal disease.

Therefore, FORL can potentially be associated with a higher likelihood of developing diabetes, viral, renal, autoimmune and cardiovascular problems due to pathogenic microbes entering the bloodstream and traveling from the mouth to other organs.



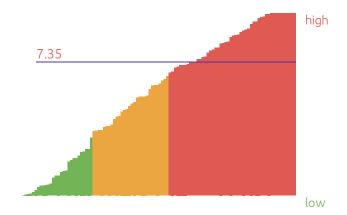
What can you do?

- It is important to regularly assess a cat's dental health in order to address any emerging issues early on and minimize chances of them negatively impacting general health.
- Adopting a thorough and consistent dental care routine at home can significantly reduce the chance of developing dental diseases. This will in turn reduce your cat's likelihood of developing more serious general health problems.



Periodontal Disease

Periodontal disease (PD) is a group of inflammatory disorders affecting the tissues surrounding the teeth. Periodontal disease is initiated by the build-up of plaque on the tooth surface resulting in the gingiva becoming inflamed (gingivitis). Without an effective oral care regime, inflammation can begin to destroy the structures that support the tooth (periodontitis). Periodontal disease affects up to 80% of the adult feline population. Below you will see how your cat's oral microbiome compares to a healthy population when it comes to microbial signatures of periodontal disease.



We analyzed Bastu - Swab's oral microbiome to establish the compositional abundance of 108 microbes predictive of periodontal disease. We ranked each microbe's abundance on a scale from 1 to 5, where 1 represents abundance levels close to a healthy control population and 5 represents abundance levels close to cats with periodontal disease. Below are Bastu - Swab's TOP 3 most significant microbes associated with high, medium, and low risk, respectively.

Currently, Bastu - Swab's compositional abundance levels for 50 out of 108 microbes are consistent with having periodontal disease (46%).

Top 3 high risk microbes

Moraxella bovoculi Corynebacterium sp. ATCC 6931 Actinomyces howellii



Top 3 medium risk microbes

Actinobacillus indolicus Bacteroides fragilis Desulfobulbus oralis



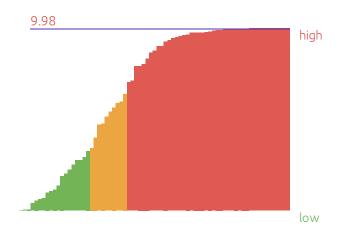
Top 3 low risk microbes

Roseburia hominis Avibacterium paragallinarum Lysobacter oculi



Tooth Resorption

Every tooth is composed of a root canal (containing nerves, blood and lymphatic vessels) and bony substances called dentin and enamel. When a cat suffers from tooth resorption, the dentin of the affected tooth starts to progressively erode. Unfortunately, tooth resorption is relatively common, affecting 20-60% of all cats and over 70% of cats over the age of five. Below, you can see how your cat compares to the healthy feline population with regards to abundance of microbes associated with tooth resorption.



We analyzed Bastu - Swab's oral microbiome to establish the compositional abundance of 74 microbes predictive of tooth resorption. We ranked each microbe's abundance on a scale from 1 to 5, where 1 represents abundance levels close to a healthy control population and 5 represents abundance levels close to cats with tooth resorption. Below are Bastu - Swab's TOP 3 most significant microbes associated with high, medium, and low risk, respectively.

Currently, Bastu - Swab's compositional abundance levels for 44 out of 74 microbes are consistent with having tooth resorption (59%).

Top 3 high risk microbes

Corynebacterium xerosis Parabacteroides distasonis Moraxella boyoculi



Top 3 medium risk microbes

Alloprevotella sp. E39 Actinobacillus indolicus Bacteroides sp. HF-5287



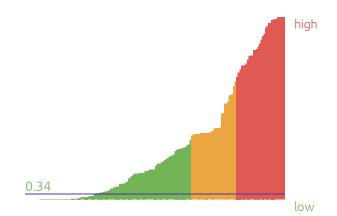
Top 3 low risk microbes

Aggregatibacter aphrophilus Gemella sp. oral taxon 928 Avibacterium paragallinarum



Bad Breath

Occasional bad breath is usually not something you should worry about. When bad breath is a persistent problem, this could be indicative of more serious issues. The most common cause of bad breath is periodontal disease. Different types of bad breath can also indicate general health problems, such as kidney disease, diabetes and some liver disorders. Here is how your cat's oral microbiome compares to the general healthy population when it comes to halitosis.



We analyzed Bastu - Swab's oral microbiome to establish the compositional abundance of 182 microbes predictive of bad breath. We ranked each microbe's abundance on a scale from 1 to 5, where 1 represents abundance levels close to a healthy control population and 5 represents abundance levels close to cats with bad breath. Below are Bastu - Swab's TOP 3 most significant microbes associated with high, medium, and low risk, respectively.

Currently, Bastu - Swab's compositional abundance levels for 32 out of 182 microbes are consistent with having bad breath (18%).

Top 3 high risk microbes

Acidovorax sp. JS42 Diaphorobacter sp. JS3050 Cardiobacterium hominis



Top 3 medium risk microbes

Neisseria bacilliformis Parvimonas micra P. propionicum



Top 3 low risk microbes

Aeromonas caviae Gemella sp. oral taxon 928 P. goodfellowii



What's next for Bastu - Swab?

At home care

To improve your cat's oral health, you are strongly advised to adopt a daily dental care routine, if you don't already have one. If you already have a routine, consider modifying it or supplementing it by implementing some of the suggestions below.

While tooth brushing is the most effective at home treatment (when done properly), we understand that every cat is unique and might have different tolerance levels for this method. We teamed up with some of the world's top veterinary dentistry professionals to provide you with support and innovative solutions on how to best approach brushing your cat's teeth and other tips and tricks for optimal dental hygiene.

Watch the video to learn how to tailor your routine to your cat's personality and comfort level.

Learn More

You can also read about some effective, off-the-beatenpath ways to maximize the effect of your cat's dental care routine in this article.

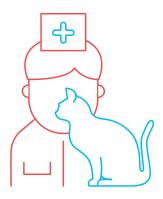
Since, your cat's report results indicate that there is a high likelihood that dental disease is already present, it is important that you learn how to routinely perform 'flip the lip' exams at home. These exams will help you identify any visible changes in your cat's teeth and gums. You should alert your veterinarian if you see any worrying signs such as ulcers, red or swollen gums, discolored teeth or anything else out of the ordinary. In addition to this, pay attention to your cat's behavior - pawing at the mouth, drooling and problems eating are strong indicators that your cat is experiencing mouth pain and needs medical attention.

Finally, you can consider incorporating some of the products recommended by the <u>Veterinary Oral Health Council (VOHC)</u> in your cat's dental care routine. These products have demonstrated efficacy in fighting plaque and tartar buildup, which are the root cause of many dental problems. However, in more advanced stages of dental disease, these products will do a good job in slowing down disease progression, but will not reverse it. In such instances, VOHC recommended products are a great complement to routine dental care provided and prescribed by your cat's veterinarian.



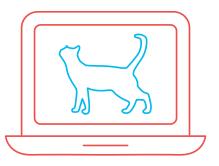
Veterinary oral exam

Have a question for Basepaws?



If you have a question about any part of this report, you can send us an email at:

meow@basepaws.com



We have identified some signs of dental disease in your cat's mouth.

We recommend scheduling an appointment with your veterinarian for an in-person general and dental health assessment within the next month.

Want to discuss your results with other cat parents? Join our facebook group!



Would you like to tell us more about your cat's dental and general health?
Contribute to feline dental health research by filling out this <u>survey</u>.

Next dental health test recommended in: 3 - 6 months



Appendix

Sequencing and analysis methodology

Most direct-to-consumer microbiome tests use a technique called '16S rRNA gene sequencing'. This technique can only provide information about the bacteria present in the microbiome.

However, it is well-known that the microbiome is composed of viruses, protozoa, fungi and archaea species, in addition to bacteria. This means that the 16S approach zooms in on just one part of the microbiome, ignoring the rest. Additionally, 16S sequencing does not provide sufficient resolution to reliably and consistently go beyond the genus level of bacterial classification. Therefore, in most cases, we don't know the exact species of bacteria in the microbiome, making analysis somewhat vague and relying on approximation.

To address these problems, Basepaws uses metagenomic sequencing instead of 16S sequencing. Our method allows us to capture organisms across all domains of life, not restricting us to just bacteria. In addition, we can reliably identify organisms to the species or even strain level, making our analysis more accurate and improving our confidence in the results. These results paint a much richer and unbiased picture of your feline companion's mouth. We used pairwise log ratio transformation to estimate the compositional abundance of microbial species and Gaussian mixture modeling to determine your cat's risk for periodontal disease, tooth resorption and bad breath.

Limitations

The Basepaws oral microbiome report is based on our ability to identify thousands of microbial species with each test.

Our large oral microbiome reference database allows us to identify a multitude of novel associations between microbes found in the mouth and a variety of diseases, as well as confirm previously reported findings. However, the field of feline oral microbiome science is extremely young and understudied, which is why we report only on conditions and microbes where previous knowledge exists and/or we see a particularly strong signal coming through in our data.

As we accumulate more data and conduct more analyses, we will aim to continuously enrich this report, providing even more helpful insights. We want to emphasize that the identification of a certain microbial signature associated with a dental disease does not constitute a diagnosis. Conversely, not detecting a particular microbial signature does not exclude the possibility of an unknown disease-causing pathogen being present or dental disease being caused by something other than pathogenic microbes. This report does not aim to substitute a diagnosis by a professional.



Amado, P.P.P., Kawamoto, D., Albuquerque-Souza, E., Franco, D.C., Saraiva, L., Casarin, R.C.V., Horliana, A.C.R.T. and Mayer, M.P.A., 2020. Oral and fecal microbiome in molar-incisor pattern periodontitis. Frontiers in cellular and infection microbiology, 10. <u>link</u>

Anesti, V., McDonald, I.R., Ramaswamy, M., Wade, W.G., Kelly, D.P. and Wood, A.P., 2005. Isolation and molecular detection of methylotrophic bacteria occurring in the human mouth. Environmental Microbiology, 7(8), pp.1227-1238. Link

Arora, N., Mishra, A. and Chugh, S., 2014. Microbial role in periodontitis: Have we reached the top? Some unsung bacteria other than red complex. Journal of Indian Society of Periodontology, 18(1), p.9. link

Awano, S., Gohara, K., Kurihara, E., Ansai, T. and Takehara, T., 2002. The relationship between the presence of periodontopathogenic bacteria in saliva and halitosis. International dental journal, 52(S5P1), pp.212-216. <u>link</u>

Binti Badlishah Sham, N.I., Lewin, S.D. and Grant, M.M., 2020. Proteomic Investigations of In Vitro and In Vivo Models of Periodontal Disease. PROTEOMICS–Clinical Applications, 14(3), p.1900043. <u>link</u>

Blum, S., Elad, D., Zukin, N., Lysnyansky, I., Weisblith, L., Perl, S., Netanel, O. and David, D., 2010. Outbreak of Streptococcus equi subsp. zooepidemicus infections in cats. Veterinary microbiology, 144(1-2), pp.236-239. link

Booij-Vrieling, H.E., van der Reijden, W.A., Houwers, D.J., De Wit, W.E.A.J., Bosch-Tijhof, C.J., Penning, L.C., Van Winkelhoff, A.J. and Hazewinkel, H.A.W., 2010. Comparison of periodontal pathogens between cats and their owners. Veterinary microbiology, 144(1-2), pp.147-152. <u>link</u>

Boyanova, L., Setchanova, L., Gergova, G., Kostyanev, T., Yordanov, D., Popova, C., Kotsilkov, K. and Mitov, I., 2009. Microbiological diagnosis of the severe chronic periodontitis. Journal of IMAB, pp.89-94. <u>link</u>

Byrne-Bailey, K.G., Weber, K.A., Bose, S., Knox, T., Spanbauer, T.L., Chertkov, O. and Coates, J.D., 2010. Completed genome sequence of the anaerobic iron-oxidizing bacterium Acidovorax ebreus strain TPSY. Journal of bacteriology, 192(5), pp.1475-1476. <u>link</u>

Chan, Y., Huo, Y.B., Yu, X., Zeng, H., Leung, W.K. and Watt, R.M., 2020. Complete Genome Sequence of Human Oral Phylogroup 1 Treponema sp. Strain OMZ 804 (ATCC 700766), Originally Isolated from Periodontitis Dental Plaque. Microbiology Resource Announcements, 9(22). link

Chan, Y., Ma, A.P., Lacap-Bugler, D.C., Huo, Y.B., Leung, W.K., Leung, F.C. and Watt, R.M., 2014. Complete genome sequence for Treponema sp. OMZ 838 (ATCC 700772, DSM 16789), isolated from a necrotizing ulcerative gingivitis lesion. Genome announcements, 2(6). <u>link</u>

Chávez de Paz, L., 2004. Gram-positive organisms in endodontic infections. Endodontic Topics, 9(1), pp.79-96. link

Cho, E., Park, S.N., Lim, Y.K., Shin, Y., Paek, J., Hwang, C.H., Chang, Y.H. and Kook, J.K., 2015. Fusobacterium hwasookii sp. nov., isolated from a human periodontitis lesion. Current microbiology, 70(2), pp.169-175. <u>link</u>

Colombo, A.P.V., Boches, S.K., Cotton, S.L., Goodson, J.M., Kent, R., Haffajee, A.D., Socransky, S.S., Hasturk, H., Van Dyke, T.E., Dewhirst, F. and Paster, B.J., 2009. Comparisons of subgingival microbial profiles of refractory periodontitis, severe periodontitis, and periodontal health using the human oral microbe identification microarray. Journal of periodontology, 80(9), pp.1421-1432. <u>link</u>

Colombo, A.P.V., Magalhães, C.B., Hartenbach, F.A.R.R., do Souto, R.M. and da Silva-Boghossian, C.M., 2016. Periodontal-disease-associated biofilm: A reservoir for pathogens of medical importance. Microbial pathogenesis, 94, pp.27-34. link

Cortelli, J.R., Barbosa, M.D.S. and Westphal, M.A., 2008. Halitosis: a review of associated factors and therapeutic approach. Brazilian oral research, 22, pp.44-54. <u>link</u>

Davis, I.J., Bull, C., Horsfall, A., Morley, I. and Harris, S., 2014. The Unculturables: targeted isolation of bacterial species associated with canine periodontal health or disease from dental plaque. BMC microbiology, 14(1), p.196. link



Davis, I.J., Bull, C., Horsfall, A., Morley, I. and Harris, S., 2014. The Unculturables: targeted isolation of bacterial species associated with canine periodontal health or disease from dental plaque. BMC microbiology, 14(1), p.196. link

Dilegge, S.K., Edgcomb, V.P. and Leadbetter, E.R., 2011. Presence of the oral bacterium Capnocytophaga canimorsus in the tooth plaque of canines. Veterinary microbiology, 149(3-4), pp.437-445. <u>link</u>

Donati, C., Zolfo, M., Albanese, D., Truong, D.T., Asnicar, F., Iebba, V., Cavalieri, D., Jousson, O., De Filippo, C., Huttenhower, C. and Segata, N., 2016. Uncovering oral Neisseria tropism and persistence using metagenomic sequencing. Nature microbiology, 1(7), p.16070. <u>link</u>

Ebrahim-Saraie, H.S., Motamedifar, M., Mansury, D., Ebrahimi, H., Pourshahidi, S., Halaji, M. and Shahraki, H.R., 2015. Association of Fusobacterium Isolation From Periodontal Pockets With Halitosis and the Related Risk Factors in Shiraz, Iran. Archives of Clinical Infectious Diseases, 10(4). <u>link</u>

Forsblom, B., Sarkiala-Kessel, E., Kanervo, A., Vaeisaenen, M.L., Helander, I.M. and Jousimies-Somer, H., 2002. Characterisation of aerobic gram-negative bacteria from subgingival sites of dogs–potential bite wound pathogens. Journal of medical microbiology, 51(3), pp.207-220. link

Galperin, M.Y., 2007. Dark matter in a deep-sea vent and in human mouth. Environmental microbiology, 9(10), p.2385. <u>link</u>

Gerner-Smidt, P., Keiser-Nielsen, H., Dorsch, M., Stackebrandt, E., Ursing, J., Blom, J., Christensen, A.C., Christensen, J.J., Frederiksen, W., Hoffmann, S. and Holten-Andersen, W., 1994. Lautropia mirabilis gen. nov., sp. nov., a Gram-negative motile coccus with unusual morphology isolated from the human mouth. Microbiology, 140(7), pp.1787-1797. link

Hampelska, K., Jaworska, M.M., Babalska, Z.Ł. and Karpiński, T.M., 2020. The role of oral microbiota in intra-oral halitosis. Journal of Clinical Medicine, 9(8), p.2484. <u>link</u>

Han, X., Lin, X., Yu, X., Lin, J., Kawai, T., LaRosa, K.B. and Taubman, M.A., 2013. Porphyromonas gingivalis infection-associated periodontal bone resorption is dependent on receptor activator of NF-κB ligand. Infection and immunity, 81(5), pp.1502-1509. link

Han, Y.W., Shi, W., Huang, G.T.J., Haake, S.K., Park, N.H., Kuramitsu, H. and Genco, R.J., 2000. Interactions between periodontal bacteria and human oral epithelial cells: Fusobacterium nucleatum adheres to and invades epithelial cells. Infection and immunity, 68(6), pp.3140-3146. link

Harris, S., Croft, J., O'Flynn, C., Deusch, O., Colyer, A., Allsopp, J., Milella, L. and Davis, I.J., 2015. A pyrosequencing investigation of differences in the feline subgingival microbiota in health, gingivitis and mild periodontitis. PLoS One, 10(11), p.e0136986. link

Holcombe, L.J., Patel, N., Colyer, A., Deusch, O., O'Flynn, C. and Harris, S., 2014. Early canine plaque biofilms: characterization of key bacterial interactions involved in initial colonization of enamel. PLoS One, 9(12), p.e113744. link

Iwamoto, T., Suzuki, N., Tanabe, K., Takeshita, T. and Hirofuji, T., 2010. Effects of probiotic Lactobacillus salivarius WB21 on halitosis and oral health: an open-label pilot trial. Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology, 110(2), pp.201-208. link

Johnson, L., Almeida-da-Silva, C.L.C., Takiya, C.M., Figliuolo, V., Rocha, G.M., Weissmüller, G., Scharfstein, J., Coutinho-Silva, R. and Ojcius, D.M., 2018. Oral infection of mice with Fusobacterium nucleatum results in macrophage recruitment to the dental pulp and bone resorption. biomedical journal, 41(3), pp.184-193. <u>link</u>

Kačírová, J., Maďar, M., Štrkolcová, G., Maďari, A. and Nemcová, R., 2019. Dental Biofilm as Etiological Agent of Canine Periodontal Disease. In Bacterial Biofilms. IntechOpen. <u>link</u>

Khazandi, M., Bird, P.S., Owens, J., Wilson, G., Meyer, J.N. and Trott, D.J., 2014. In vitro efficacy of cefovecin against anaerobic bacteria isolated from subgingival plaque of dogs and cats with periodontal disease. Anaerobe, 28, pp.104-108. <u>link</u>



Khazandi, M., Bird, P.S., Owens, J., Wilson, G., Meyer, J.N. and Trott, D.J., 2014. In vitro efficacy of cefovecin against anaerobic bacteria isolated from subgingival plaque of dogs and cats with periodontal disease. Anaerobe, 28, pp.104-108. <u>link</u>

Kushkevych, I., Coufalová, M., Vítězová, M. and Rittmann, S.K.M., 2020. Sulfate-Reducing Bacteria of the Oral Cavity and Their Relation with Periodontitis—Recent Advances. Journal of Clinical Medicine, 9(8), p.2347. <u>link</u>

Langendijk, P.S., Kulik, E.M., Sandmeier, H., Meyer, J. and Van Der Hoeven, J.S., 2001. Isolation of Desulfomicrobium orale sp. nov. and Desulfovibrio strain NY682, oral sulfate-reducing bacteria involved in human periodontal disease. International Journal of Systematic and Evolutionary Microbiology, 51(3), pp.1035-1044. link

Liu, G., Tang, C.M. and Exley, R.M., 2015. Non-pathogenic Neisseria: members of an abundant, multi-habitat, diverse genus. Microbiology, 161(7), pp.1297-1312. link

Loubinoux, J., Bisson-Boutelliez, C., Miller, N. and Le Faou, A.E., 2002. Isolation of the provisionally named Desulfovibrio fairfieldensis from human periodontal pockets. Oral microbiology and immunology, 17(5), pp.321-323. <u>link</u>

Love, D.N., Johnson, J.L., Jones, R.F. and Bailey, M.A.R.I.L.Y.N., 1985. Comparison of Bacteroides zoogleoformans strains isolated from soft tissue infections in cats with strains from periodontal disease in humans. Infection and immunity, 47(1), pp.166-168. <u>link</u>

Love, D.N., Johnson, J.L. and Moore, L.V.H., 1989. Bacteroides species from the oral cavity and oral-associated diseases of cats. Veterinary microbiology, 19(3), pp.275-281. <u>link</u>

Mallonee, D.H., Harvey, C.E., Venner, M. and Hammond, B.F., 1988. Bacteriology of periodontal disease in the cat. Archives of oral biology, 33(9), pp.677-683. link

Mineoka, T., Awano, S., Rikimaru, T., Kurata, H., Yoshida, A., Ansai, T. and Takehara, T., 2008. Site-specific development of periodontal disease is associated with increased levels of Porphyromonas gingivalis, Treponema denticola, and Tannerella forsythia in subgingival plaque. Journal of periodontology, 79(4), pp.670-676. link

Nises, J., Rosander, A., Pettersson, A. and Backhans, A., 2018. The occurrence of Treponema spp. in gingival plaque from dogs with varying degree of periodontal disease. PloS one, 13(8), p.e0201888. <u>link</u>

Nordhoff, M., Rühe, B., Kellermeier, C., Moter, A., Schmitz, R., Brunnberg, L. and Wieler, L.H., 2008. Association of Treponema spp. with canine periodontitis. Veterinary microbiology, 127(3-4), pp.334-342. link

Norris, J.M. and Love, D.N., 1999. Associations amongst three feline Porphyromonas species from the gingival margin of cats during periodontal health and disease. Veterinary microbiology, 65(3), pp.195-207. link

Özavci, V., Erbas, G., Parin, U., Yüksel, H.T. and Kirkan, Ş., 2019. Molecular detection of feline and canine periodontal pathogens. Veterinary and Animal Science, 8, p.100069. <u>link</u>

Park, S.N., Lim, Y.K., Shin, J.H., Kim, H.S., Jo, E., Lee, W.P., Shin, Y., Paek, J., Chang, Y.H., Kim, H. and Kook, J.K., 2019. Fusobacterium pseudoperiodonticum sp. nov., Isolated from the Human Oral Cavity. Current microbiology, 76(6), pp.659-665. <u>link</u>

Prates, M., Fernandes, F., Proença, F., Mussá, Y., Tavares, A. and Pereira, A., 2020. Oral Infection Caused by Stenotrophomonas maltophilia: A Rare Presentation of an Emerging Opportunistic Pathogen. Case Reports in Infectious Diseases, 2020. <u>link</u>

Richards, A.M., Abu Kwaik, Y. and Lamont, R.J., 2015. Code blue: A cinetobacter baumannii, a nosocomial pathogen with a role in the oral cavity. Molecular oral microbiology, 30(1), pp.2-15. <u>link</u>

Riggio, M.P., Lennon, A., Taylor, D.J. and Bennett, D., 2011. Molecular identification of bacteria associated with canine periodontal disease. Veterinary microbiology, 150(3-4), pp.394-400. link



Riggio, M.P., Lennon, A., Taylor, D.J. and Bennett, D., 2011. Molecular identification of bacteria associated with canine periodontal disease. Veterinary microbiology, 150(3-4), pp.394-400. link

Rodrigues, M.X., Bicalho, R.C., Fiani, N., Lima, S.F. and Peralta, S., 2019. The subgingival microbial community of feline periodontitis and gingivostomatitis: characterization and comparison between diseased and healthy cats. Scientific reports, 9(1), pp.1-10. <u>link</u>

Ruparell, A., Inui, T., Staunton, R., Wallis, C., Deusch, O. and Holcombe, L.J., 2020. The canine oral microbiome: variation in bacterial populations across different niches. BMC microbiology, 20(1), pp.1-13. <u>link</u>

Sanguansermsri, P., Nobbs, A.H., Jenkinson, H.F. and Surarit, R., 2018. Interspecies dynamics among bacteria associated with canine periodontal disease. Molecular oral microbiology, 33(1), pp.59-67. <u>link</u>

Sela, M.N., 2001. Role of Treponema denticola in periodontal diseases. Critical Reviews in Oral Biology & Medicine, 12(5), pp.399-413. <u>link</u>

Sundavist, G., 1990. Endodontic microbiology. Experimental endodontics, 6, pp.131-153.

Suzuki, M., Kimura, M., Imaoka, K. and Yamada, A., 2010. Prevalence of Capnocytophaga canimorsus and Capnocytophaga cynodegmi in dogs and cats determined by using a newly established species-specific PCR. Veterinary microbiology, 144(1-2), pp.172-176. <u>link</u>

Svartström, O., Mushtaq, M., Pringle, M. and Segerman, B., 2013. Genome-wide relatedness of Treponema pedis, from gingiva and necrotic skin lesions of pigs, with the human oral pathogen Treponema denticola. PloS one, 8(8), p.e71281. link

Sykora, S., Pieber, K., Simhofer, H., Hackl, V., Brodesser, D. and Brandt, S., 2014. Isolation of Treponema and Tannerella spp. from equine odontoclastic tooth resorption and hypercementosis related periodontal disease. Equine veterinary journal, 46(3), pp.358-363. <u>link</u>

Umeda, K., Hatakeyama, R., Abe, T., Takakura, K.I., Wada, T., Ogasawara, J., Sanada, S.I. and Hase, A., 2014. Distribution of Capnocytophaga canimorsus in dogs and cats with genetic characterization of isolates. Veterinary microbiology, 171(1-2), pp.153-159. link

Van Dyke, T.E., Offenbacher, S., Place, D., Dowell, V.R. and Jones, J., 1988. Refractory Periodontitis: Mixed Infection with Bacteroides gingivalis and Other Unusual Bacteroides Species: A Case Report. Journal of periodontology, 59(3), pp.184-189. link

Wallis, C.V., Marshall-Jones, Z.V., Deusch, O. and Hughes, K.R., 2017. 17 Canine and Feline Microbiomes. Understanding Host-Microbiome Interactions-An Omics Approach: Omics of Host-Microbiome Association, p.279. link

Wyss, C., Moter, A., Choi, B.K., Dewhirst, F.E., Xue, Y., Schüpbach, P., Göbel, U.B., Paster, B.J. and Guggenheim, B., 2004. Treponema putidum sp. nov., a medium-sized proteolytic spirochaete isolated from lesions of human periodontitis and acute necrotizing ulcerative gingivitis. International journal of systematic and evolutionary microbiology, 54(4), pp.1117-1122. link

Yasukawa, T., Ohmori, M. and Sato, S., 2010. The relationship between physiologic halitosis and periodontopathic bacteria of the tongue and gingival sulcus. Odontology, 98(1), pp.44-51. <u>link</u>

Ye, W., Zhang, Y., He, M., Zhu, C. and Feng, X.P., 2019. Relationship of tongue coating microbiome on volatile sulfur compounds in healthy and halitosis adults. Journal of breath research, 14(1), p.016005. link

Yoshino, Y., Kitazawa, T., Kamimura, M., Tatsuno, K., Ota, Y. and Yotsuyanagi, H., 2011. Pseudomonas putida bacteremia in adult patients: five case reports and a review of the literature. Journal of Infection and Chemotherapy, 17(2), pp.278-282. <u>link</u>

Zambori, C., Tirziu, E., Nichita, I., Cumpanasoiu, C., Gros, R.V., Seres, M., Mladin, B. and Mot, D., 2012. Biofilm implication in oral diseases of dogs and cats. Scientific Papers Animal Science and Biotechnologies, 45(2), pp.208-212.



Zambori, C., Tirziu, E., Nichita, I., Cumpanasoiu, C., Gros, R.V., Seres, M., Mladin, B. and Mot, D., 2012. Biofilm implication in oral diseases of dogs and cats. Scientific Papers Animal Science and Biotechnologies, 45(2), pp.208-212.

Zangenah, S., Abbasi, N., Andersson, A.F. and Bergman, P., 2016. Whole genome sequencing identifies a novel species of the genus Capnocytophaga isolated from dog and cat bite wounds in humans. Scientific reports, 6, p.22919. <u>link</u>

Zhang, Y., Zhen, M., Zhan, Y., Song, Y., Zhang, Q. and Wang, J., 2017. Population-genomic insights into variation in Prevotella intermedia and Prevotella nigrescens isolates and its association with periodontal disease. Frontiers in cellular and infection microbiology, 7, p.409. link

