

Reproducible, flexible and high throughput data  
extraction from primary literature: The **metaDigitise**  
R package

Supplementary Materials

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# S1 Tutorial

## S1.1 Extracting Data From Plots

We will now demonstrate how `metaDigitise()` works using figures generated from the well known iris data set. Users can install the **metaDigitise** package from GitHub as follows:

```
R> install.packages("devtools")
R> devtools::install_github("danielnoble/metaDigitise")
R> library(metaDigitise)
```

Assume that the user would like to extract descriptive statistics from studies measuring sepal length or width in iris species for a fictitious project. There are a few studies that only present these data in figures. As the user reads papers found from a systematic search, they add figures with relevant data to a "FiguresToExtract" folder as follows

```
*FiguresToExtract/
+ 001_Anderson_1935_Fig1.png
```

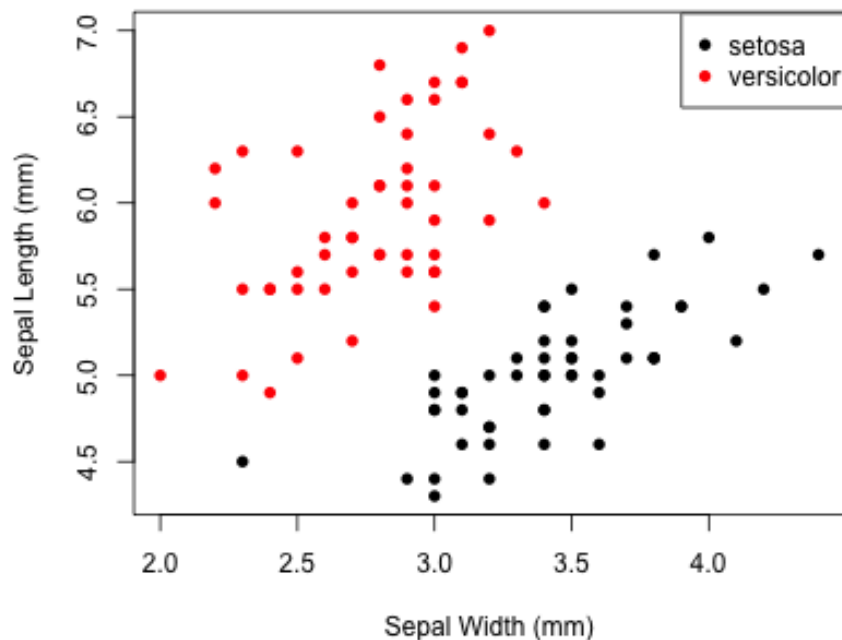


Figure 1: Example scatterplot (001\_Anderson\_1935\_Fig1.png) of sepal length and width for two species of iris (setosa and versicolor)

Here, the naming of the files placed in the folder will contain the paper number, first author and the figure number to keep data uniquely associated with figures. At first there is one figure in the folder, shown in Figure 1. Running `metaDigitise()` brings up a series of prompts for the user using a main menu that provides access to a number of its features ("..." here represents the user's path to the project directory):

```
R> digitised_data <- metaDigitise("../FiguresToExtract", summary = TRUE)
```

```

Do you want to...
1: Process new images
2: Import existing data
3: Edit existing data
Selection:
```

The user simply enters in the numeric value that corresponds to what they would like to do. In this case they want to "Process new images". The user is then asked whether there are different types of plot(s) in the folder. This question is most relevant when there are lots of different figures in the folder because it will then ask the user for the type of figure as they are cycled through.

```
Are all plot types Different or the Same? (d/s)
```

`metaDigitise()` then asks the user whether the figure needs to be rotated or flipped. This can be needed when box plots and mean and error plots are not orientated correctly. In some cases, older papers can give slightly off angled images which can be corrected by rotating. So, in this prompt the user has three options: `f` for "Flip", `r` for "rotate" or `c` for "continue".

```
mean_error and boxplots should be vertically orientated
```

```

-
|
I.E. o    NOT  |-o-|
|
-

```

```
If they are not then chose flip to correct this.
```

```
If figures are wonky, chose rotate.
```

```
Otherwise chose continue
```

Flip, rotate or continue (f/r/c)

*R> c*

After this, `metaDigitise()` will ask the user to specify the plot type. Depending on the figure, the user can specify that it is a figure containing the mean and error (**m**), a box plot (**b**), a scatter plot (**s**) or a histogram (**h**). If the user has specified **d** instead of **s** in response to the question about whether the plot types are the same or different, this question will pop up for each plot, but will only be asked once if plots are all the same.

Please specify the `plot_type` as either:

**m**: Mean and error

**b**: Box plot

**s**: Scatter plot

**h**: Histogram

*R> s*

After selecting the figure type a new set of prompts will come up that will ask the user first what the y and x-axis variables are. This is useful as users can keep track of the different variables across figures and papers. Here, the user can just add this information in to the R console. Once complete, details on how to calibrate the x and y-axis appear, so that the relevant statistics / data can be correctly calculated. When working with a plot of mean and standard errors, the x-axis is rather useless in terms of calibration so `metaDigitise()` just asks the user to calibrate the y-axis.

What is the y variable?

*R> Sepal Length (mm)*

What is the x variable?

*R> Sepal Width (mm)*

On the Figure, click IN ORDER:

y1, y2 , x1, x2

Step 1 ----> Click on known value on y axis - y1

|  
|

```
|
|
y1
|-----
....
```

Step 3 ----> Click on known value on x axis - x1

```
|
|
|
|
|
|_____x1_____
....
```

The user can just follow the instructions on screen step-by-step (instructions above have been truncated by ‘...’ to simplify), and in the order specified. Before moving on, the user is forced to check whether or not the calibration has been set up correctly. If **n** is chosen because something needs to be fixed then the user can re-calibrate.

What is the value of y1 ?

*R*> 4.5

What is the value of y2 ?

*R*> 7

What is the value of x1 ?

*R*> 2

What is the value of x2 ?

*R*> 4

Re-calibrate? (y/n)

*R*> n

Often, plots might contain multiple groups that the meta-analyst wants to extract from. `metaDigitise()` handles this nicely by prompting the user to enter the group first, followed by digitisation of this groups data. After digitising the first group, and having

exited, `metaDigitise()` will ask the user whether they would like to add another group. Users can continually add groups (a), delete groups (d), edit groups (e) or finish a plot and continue to the next one (f - if another plot exists). The number of groups are not really limited and users can just keep adding in groups to accommodate the different numbers that may be presented across figures (although it can get complicated with too many).

If there are multiple groups, enter unique group identifiers (otherwise press enter)  
Group identifier:

```
R> setosa
```

Click on points you want to add.

If you want to remove a point, or are finished with a group,  
exit by clicking on red box in bottom left corner, then follow prompts

To finish selecting points, the user can exit by clicking on the red button that appears when extracting points. The user is then asked if they want to add or delete points from that group.

Add or Delete points to this group, or Continue? (a/d/c)

```
R> c
```

Once we are done digitising all the groups our plot will look something like Figure 2.

When completed `metaDigitise()` will write the digitised data as a `metaDigitise` object to a RDS file in the `caldat` directory, such that our new directory structure is as follows

```
*FiguresToExtract/
+ caldat/
+ 001_Anderson_1935_Fig1
+ 001_Anderson_1935_Fig1.png
```

Users can access the `metaDigitise` object created (001\_Anderson\_1935\_Fig1) at any time using the `metaDigitise()` function. In the R console, the summarised data for the digitised figure can be printed on screen or even written to a .csv file:

```
R> digitised_data
```

filename	group_id	variable	mean	error	error_type	n	r	sd	plot_type
001_Anderson_1935_Fig1.png	setosa	Sepal width (mm)	3.42	0.40	sd	39	0.75	0.40	scatterplot
001_Anderson_1935_Fig1.png	setosa	Sepal length (mm)	5.00	0.38	sd	39	0.75	0.38	scatterplot
001_Anderson_1935_Fig1.png	versicolor	Sepal width (mm)	2.77	0.32	sd	44	0.52	0.32	scatterplot
001_Anderson_1935_Fig1.png	versicolor	Sepal length (mm)	5.95	0.53	sd	44	0.52	0.53	scatterplot

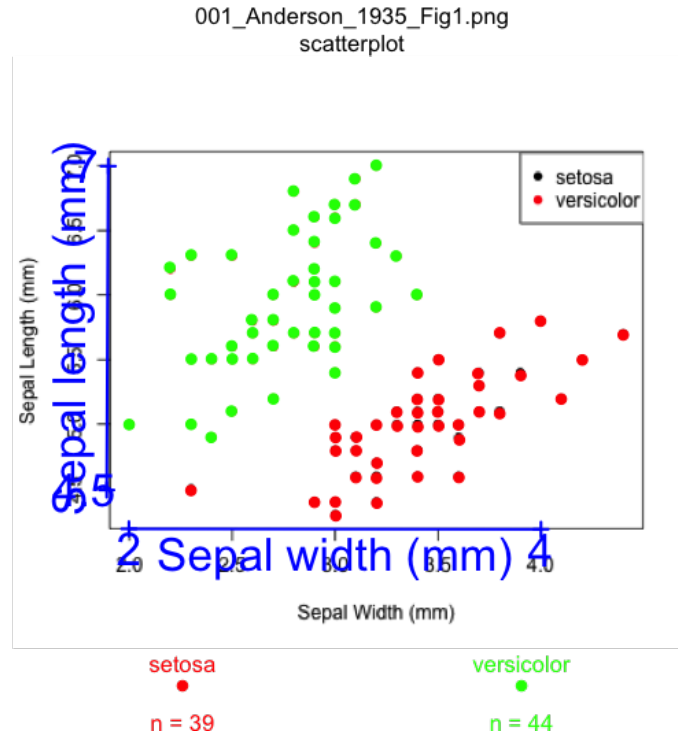


Figure 2: Digitisation of sepal length and width for two species of iris (setosa and versicolor). Names of the variables and calibration (in blue) are plotted alongside the digitised points (green = versicolor; red = setosa). The sample sizes for each group are provided on the lower part of the plot. All figures are clearly labelled at the top to remind users of the filename and plot type. This reduces errors throughout the digitisation process.

The mean for each of the two variables, along with the two species, are provided. Since this is a scatterplot, the user also gets the Person's correlation coefficient between sepal length and width for each species. These match reasonably well with the actual means of sepal length and width for each of the species in the full 'iris' dataset:

	Species	meanSL	meanSW
1	setosa	5.006	3.428
2	versicolor	5.936	2.770

One thing anyone with a familiarity with the iris dataset will notice is that the sample sizes for each of these species (which are  $n = 50$  each) are quite a bit lower. This is an example of some of the challenges when extracting data from scatter plots. Often data points will overlap with each other making it impossible (without having the real data) to know whether this is a problem. However, a meta-analyst will probably realise that the sample sizes here conflict with what is reported in the paper. Hence, **metaDigitise** also provides the user with options to input the sample sizes directly (see Editing section below), even for scatter plots and histograms where, strictly speaking, this should not be

necessary. Nonetheless, it is important to recognise the impact that overlapping points can have on summary statistics, particularly its effects on standard deviation (SD) and standard error (SE). Here, the mean point estimates are nearly exactly the same as the true values, but the SD's are slightly over-estimated:

	Species	meanSL	meanSW
1	setosa	0.3524897	0.3790644
2	versicolor	0.5161711	0.3137983

## S1.2 Adding new figures

Users can add additional figures as new papers with relevant information are found. Each figure should be in its own file with unique naming, even if a single paper has multiple figures for extraction. For example, another paper on different populations (and one new species) of iris contained two additional figures where important data could be extracted. These figures can simply be named accordingly and added directly to the same extraction folder:

```
*FiguresToExtract/
+ caldat/
  + 001_Anderson_1935_Fig1
+ 001_Anderson_1935_Fig1.png
+ 002_Doe_2013_Fig1.png
+ 002_Doe_2013_Fig3.png
```

The user has already processed one figure (001\_Anderson\_1935\_Fig1.png). We can tell this because the caldat folder has digitised data in it (caldat/001\_Anderson\_1935\_Fig1). Now the user has two new figures that have not yet been digitised. This example will nicely demonstrate how users can easily pick up from where they left off and how all previous data gets re-integrated. It will also demonstrate how different plot types are handled. All we have to do to begin, is again, provide the directory where all the figures are located:

```
R> digitised_data <- metaDigitise("~/FiguresToExtract", summary = TRUE)
```

The user gets the same set of prompts and simply chooses option one. This will permit users to digitise new figures, and will integrate previously completed digitisations along with newly digitised data together at the end of the session, or when the user decides to quit. This time, 001\_Anderson\_1935\_Fig1.png is ignored and the new plots cycle on



screen; first 002.Doe.2013.Fig1.png and then 002.Doe.2013.Fig3.png. Since there are a few different figure types, the user answers the first question in the R console as "d":

Are all plot types Different or the Same? (d/s)

R> d

\*\*\*\* NEW PLOT \*\*\*\*

mean\_error and boxplots should be vertically orientated

```
      -  
      |  
I.E. o   NOT  |-o-|  
      |  
      -
```

If they are not then chose flip to correct this.

If figures are wonky, chose rotate.

Otherwise chose continue

Flip, rotate or continue (f/r/c)

R> c

Please specify the plot\_type as either:

m: Mean and error  
b: Box plot  
s: Scatter plot  
h: Histogram

R> m

Here, the user specifies the new plot type as m for 002.Doe.2013.Fig1.png because the user has a plot of the mean and error of sepal length for each of the three species. The user is then prompted a bit differently from our scatter plot as the x-axis is not needed for calibration:

What is the y variable?

```
R> Sepal length
```

On the Figure, click IN ORDER:

y1, y2

Step 1 ----> Click on y1

```
|
|
|
|
y1
|-----
```

Step 2 ----> Click on y2

```
|
y2
|
|
|
|-----
```

What is the value of y1 ?

```
R> 5
```

What is the value of y2 ?

```
R> 6.5
```

Re-calibrate? (y/n)

```
R> n
```

Do you know sample sizes? (y/n)

```
R> y
```

If there are multiple groups, enter unique group identifiers (otherwise press enter)

Group identifier:

```
R> setosa
```

Group sample size:

```
R> 50
```

Click on Error Bar, followed by the Mean

Add group, Edit Group, Delete group or Finish plot? (a/e/d/f)

```
R> a
```

Again, `metaDigitise()` will simply guide the user through digitising each of these figures describing to them exactly what needs to be done. At any point if mistakes are made the user can choose relevant options to edit or correct things before ending the figure. This process continues for each plot so long as the user would like to continue and after completing a single plot the user is always prompted as follows:

Do you want continue: 1 plots out of 2 plots remaining (y/n)

```
R> y
```

This continues until users have completed all non-digitised figures in the folder, at which point `metaDigitise()` concatenates the new data with previously digitised data in the object:

```
data
```

	filename	group_id	variable	mean	error	error_type	n	r	sd	plot_type
001_Anderson_1935_Fig1.png		setosa	Sepal width (mm)	3.42	0.40	sd	39	0.75	0.40	scatterplot
001_Anderson_1935_Fig1.png		setosa	Sepal length (mm)	5.00	0.38	sd	39	0.75	0.38	scatterplot
001_Anderson_1935_Fig1.png		versicolor	Sepal width (mm)	2.77	0.32	sd	44	0.52	0.32	scatterplot
001_Anderson_1935_Fig1.png		versicolor	Sepal length (mm)	5.95	0.53	sd	44	0.52	0.53	scatterplot
002_Doe_2013_Fig1.png		setosa	Sepal length	5.00	0.11	se	50	NA	0.78	mean_error
002_Doe_2013_Fig1.png		virginica	Sepal length	6.59	0.18	se	50	NA	1.26	mean_error
002_Doe_2013_Fig1.png		versicolor	Sepal length	5.94	0.14	se	50	NA	1.01	mean_error
003_Doe_2013_Fig3.png		catana	Sepal length	4.95	0.36	sd	50	NA	0.36	histogram

### S1.3 Re-importing, Editing and Plotting Previously Digitised data

A particularly useful feature of **metaDigitise** is its ability to re-import, edit and re-plot previously digitised figures. We can do this from the initial options from `metaDigitise()`

```
R> digitised_data <- metaDigitise("../FiguresToExtract")
```

Do you want to...

1: Process new images

2: Import existing data

### 3: Edit existing data

Selection:

If the user chooses "Import existing data", they have the option of either 1) importing data from all digitised images or 2) importing data from a single image that has been digitised. If 2, then a list of files are provided to the user that they can select. Editing existing data allows users to easily re-plot or edit information or digitisations that have previously be done for any plot. This is accomplished by guiding the user through a new set of options:

Choose how you want to edit files:

- 1: Cycle through images
- 2: Choose specific file to edit
- 3: Enter previously omitted sample sizes

Selection:

If the user is unsure about the name of the specific figure they need to edit or simply want to just check the digitisations of figures they can choose "Cycle through images", which will bring up each figure, one by one, overlaying the calibrations, group names (if they exist), sample sizes (if they were entered) and the selected points. The user will then be given the choice to edit individual images. Alternatively, choosing option 2, will bring up a list of the completed files in the folder and the specific file can be chosen, at which point it will be replotted. Either of these options will cycle through a number of questions asking the user what they would like to edit:

Edit rotation? If yes, then the whole extraction will be redone (y/n)

*R> n*

Change plot type? If yes, then the whole extraction will be redone (y/n)

*R> n*

Variable entered as:

*R> Sepal length*

Rename Variables (y/n)

*R> n*

Edit calibration? (y/n)

*R> n*

Re-extract data (y/n)

R> y

Change group identifier? (y/n)

R> n

Add group, Delete group or Finish plot? (a/d/f)

R> d

1: setosa

2: versicolor

3: virginica

Selection:

R> 2

Add group, Delete group or Finish plot? (a/d/f)

R> a

A whole host of information can be edited including the rotation, plot type, the variable name(s) that were provided, the calibration and even the digitisation of groups. When editing the `metaDigitise` object is re-written to the `caldat` folder and the edits are immediately integrated into the existing object once complete.

## S1.4 Obtaining Processed Data

While `metaDigitise()` provides users with the summary statistics by default, for all plot types, in many cases the user may actually be interested in obtaining the processed digitised data from scatter plots (i.e. calibrated points). This is very easy to do by changing the default `summary` argument from `TRUE` to `FALSE` in `metaDigitise()`. Instead of providing the user with summary statistics it will return a list containing four slots for each of the figure types (mean error, box plot, histogram and scatter plots). An example of a data object returned from digitising figures is as follows:

```
>R str(data)
```

```
List of 3
```

```
$ mean_error :List of 1
```

```
..$ 002_Doe_2013_Fig1.png:'data.frame': 3 obs. of 5 variables:
```

```
.. ..$ id : Factor w/ 3 levels "setosa","versicolor",...: 1 2 3
```

```

.. ..$ mean      : num [1:3] 5 5.93 6.59
.. ..$ error      : num [1:3] 0.111 0.148 0.178
.. ..$ n          : num [1:3] 50 50 50
.. ..$ variable: chr [1:3] "Sepal length" "Sepal length" "Sepal length"
$ hist           :List of 1
..$ 003_Doe_2013_Fig3.png:'data.frame': 8 obs. of 3 variables:
.. ..$ midpoints: num [1:8] 4.3 4.5 4.7 4.9 5.1 ...
.. ..$ frequency: num [1:8] 4 5 7 12 11 6 2 3
.. ..$ variable : chr [1:8] "Sepal length" "Sepal length" ...
$ scatterplot:List of 1
..$ 001_Anderson_1935_Fig1.png:'data.frame': 83 obs. of 8 variables:
.. ..$ id          : Factor w/ 2 levels "setosa","versicolor": 1 1 1 1 1 ...
.. ..$ x           : num [1:83] 2.3 2.9 3 3 3 ...
.. ..$ y           : num [1:83] 4.5 4.4 4.41 4.3 4.8 ...
.. ..$ group       : num [1:83] 1 1 1 1 1 1 1 1 1 1 ...
.. ..$ col         : Factor w/ 2 levels "red","green": 1 1 1 1 1 1 1 1 1 1 ...
.. ..$ pch         : num [1:83] 19 19 19 19 19 19 19 19 19 19 ...
.. ..$ y_variable: chr [1:83] "Sepal length (mm)" "Sepal length (mm)" ...
.. ..$ x_variable: chr [1:83] "Sepal width (mm)" "Sepal width (mm)" ...

```

Here, the user can easily access the list of processed scatter plot data by simply extracting the scatter plot slot:

```
>R scatterplot <- data$scatterplot
```

## S1.5 Adding sample sizes to previous Digitisations

In many cases important information, such as sample sizes, may not be readily available or clear when digitising figures. In these circumstances users will have answered ‘no’ to the question about whether they have sample sizes or not while digitising. To expedite finding and adding in these sample sizes to do the necessary calculations (if for example a figure presented 95% CI’s or standard errors), `metaDigitise()` has a specific edit option that allows users to enter in previously omitted sample sizes. It works by first identifying the missing sample sizes in the digitised output, re-plotting the relevant figure and then prompting the user to enter the sample sizes for the relevant groups in the figure, one by one. As an example, assume that we were missing sample sizes for two groups in 002\_Doe\_2013\_Fig1.png:

filename	group_id	variable	mean	error	error_type	n	r	sd	plot_type
002_Doe_2013_Fig1.png	setosa	Sepal length	5.00	0.11	se	NA	NA	NA	mean_error
002_Doe_2013_Fig1.png	virginica	Sepal length	6.59	0.18	se	NA	NA	NA	mean_error

Here, we can see that we are missing the sample sizes for setosa and virginica, and as a result, sd is not calculated because `metaDigitise()` needs this information to make the calculation. If the user found this information after contacting the authors for clarification then they can add these back in as follows:

```
R> digitised_data <- metaDigitise("../FiguresToExtract")
```

Do you want to...

- 1: Process new images
- 2: Import existing data
- 3: Edit existing data

Selection:

```
R> 3
```

Choose how you want to edit files:

- 1: Cycle through images
- 2: Choose specific file to edit
- 3: Enter previously omitted sample sizes

Selection:

```
>R 3
```

`metaDigitise()` will replot the figure after this and list, only the groups missing data, for which the user can then update the data. This is then re-integrated back into the data automatically and the sd calculated.

Group " setosa ": Enter sample size

```
R> 50
```

Group " virginica ": Enter sample size

```
R> 50
```

## S1.6 Figure Rotation and Adjustment

Figures may have been extracted from old publications, for example from scanned images, and so are not perfectly orientated on the image. This will make the calibration of the points in the figure from the image problematic. `metaDigitise()` allows users to rotate the image. By clicking two points on the x-axis, `metaDigitise` calculates the angle needed to rotate the image so the x-axis is horizontal, and rotates it. (Figure 3A,B)

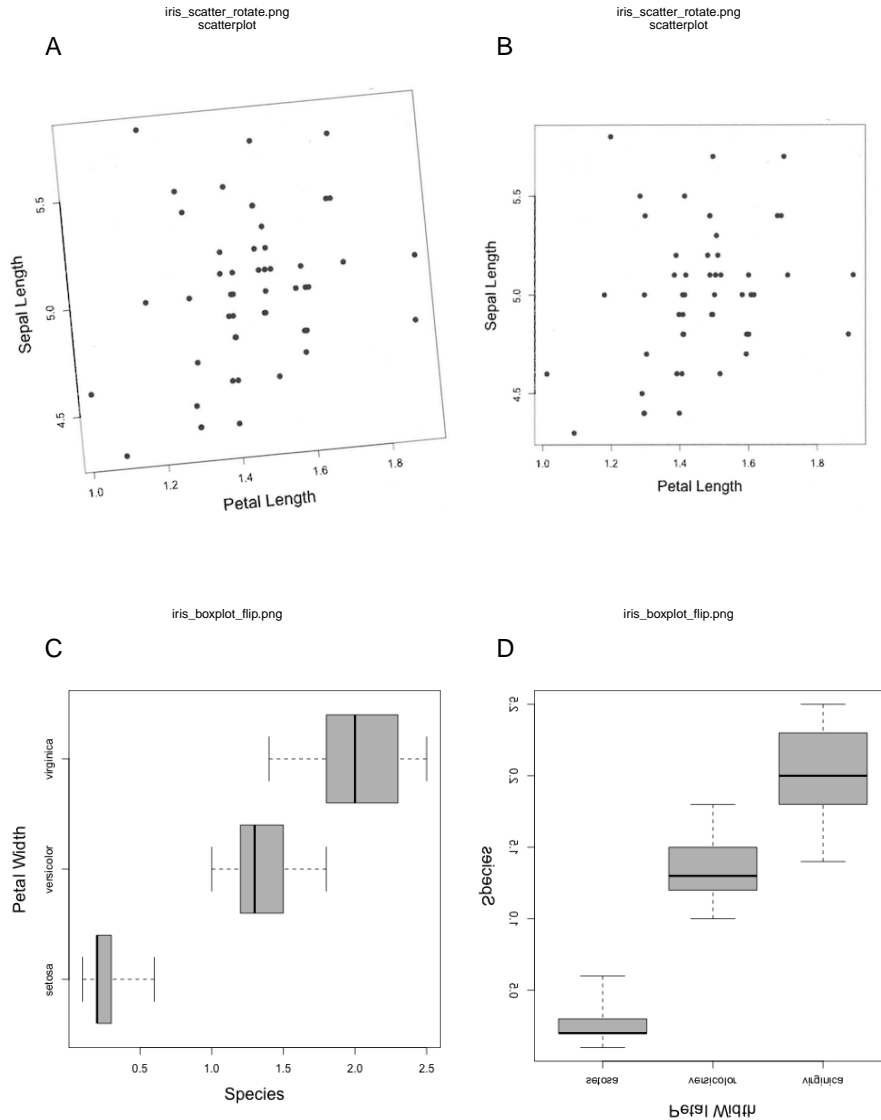


Figure 3: Figure rotation. A) and B) show how non-aligned images can be realigned through user defined rotation. C) and D) show how figures can be re-orientated so as to aid data input.



Furthermore, some figures, including mean and error, boxplots or histograms, may be presented with horizontal bars. `metaDigitise()` assumes that the bars are vertical, but allows the user to flip the image so that the bars are vertical if provided horizontally (Figure 3C,D).

## S2 Derivation of mean, standard deviation and sample size from different plot types

### S2.1 Mean/Error Plots

The standard deviation is calculated depending on the type of error presented. The user can choose from standard deviation (SD,  $\sigma$ ), standard error (SE) or 95% confidence intervals (CI95). Standard deviation is calculated from standard error as

$$\sigma = SE\sqrt{n} \quad (S1)$$

and from 95% confidence intervals as

$$\sigma = \frac{CI}{1.96}\sqrt{n} \quad (S2)$$

### S2.2 Box Plots

The mean ( $\mu$ ) and SD are calculated using the maximum ( $b$ ), upper quartile ( $q_3$ ), median ( $m$ ), lower quartile ( $q_1$ ) and minimum ( $a$ ) as

$$\mu = \frac{(n+3)(a+b) + 2(n-1)(q_1+m+q_3)}{8n} \quad (S3)$$

following Bland (2015) and

$$\sigma = \frac{b-a}{4\Phi^{-1}\left(\frac{n-0.375}{n+0.25}\right)} + \frac{q_3-q_1}{4\Phi^{-1}\left(\frac{0.75n-0.125}{n+0.25}\right)} \quad (S4)$$

where  $\Phi^{-1}(z)$  is the upper  $z$ th percentile of the standard normal distribution, following Wan et al. (2014).

### S2.3 Histograms

For each bar, the user click two point (the top of the bar). Using these points, a midpoint (m; mean x coordinates) and a frequency (f; mean y coordinates, rounded to the nearest integer) is calculated for each bar. The sample size, mean and SD are calculated as:

$$n = \sum_{i=1}^n f_i \quad (\text{S5})$$

$$\mu = \frac{\sum_{i=1}^n m_i f_i}{n} \quad (\text{S6})$$

$$\sigma = \sqrt{\frac{\sum_{i=1}^n (m_i f_i - \mu f_i)^2}{n - 1}} \quad (\text{S7})$$

## S3 Comparison with existing software

Function	metaDigitise	GraphClick <sup>1</sup>	DataThief <sup>2</sup>	DigitizeIt <sup>3</sup>	WebPlotDigitizer <sup>4</sup>	metagear <sup>5</sup>	digitize <sup>6</sup>
Scatterplots	✓	✓	✓	✓	✓	✓ <sup>7</sup>	✓
Mean/error plots	✓	✓	✓	×	×	✓ <sup>7</sup>	×
Boxplots	✓	×	×	×	×	×	×
Histograms	✓	×	×	×	✓ <sup>7</sup>	×	×
Graph rotation <sup>8</sup>	✓	✓	✓	✓	✓	×	×
Groups	✓	✓	×	✓	✓	×	×
Entry of metadata	✓	×	×	×	×	×	×
Summarising data	✓	×	×	×	×	×	×
Multiple image processing	✓	×	×	×	×	×	×
Reproducible <sup>9</sup>	✓	✓	✓	×	✓	×	×
Automated point detection	×	✓	×	✓	✓	✓	×
Line extraction	×	✓	✓	✓	✓	×	×
Zoom	×	✓	✓	✓	✓	×	×
Log axis	×	✓	✓	✓	✓	×	×
Dates	×	×	✓	×	✓	×	×
Asymmetric error bars	×	×	✓	×	×	×	×
Freeware	✓ <sup>10</sup>	✓ <sup>11</sup>	✓ <sup>11</sup>	×	✓ <sup>11</sup>	✓ <sup>10</sup>	✓ <sup>10</sup>

<sup>1</sup> Arizona-Software (2008) <sup>2</sup> Tummers (2006) <sup>3</sup> Bormann (2012) <sup>4</sup> Rohatgi (2017) <sup>5</sup> Lajeunesse (2016) <sup>6</sup> Poisot (2011)

<sup>7</sup> Only automated, no manual extraction.

<sup>8</sup> Or handles rotated graphs.

<sup>9</sup> Allows saving, re-plotting and editing of data extraction.

<sup>10</sup> R package.

<sup>11</sup> Standalone software.

Table S1: Comparison of functionality between different digitisation softwares.

## References

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