

Simulation Best Practices for Lipid Membranes : v1.0

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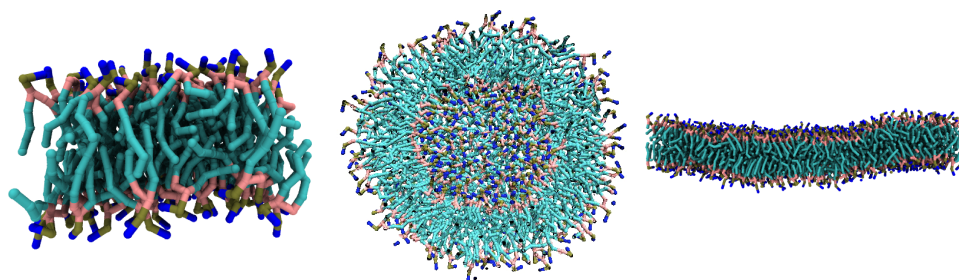
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Abstract

Here, we establish a reliable and robust standardization of settings and setup practices for practical molecular dynamics (MD) simulations of pure and mixed lipid bilayer membranes. The simulation of lipid membranes can be a daunting task, given the uniqueness of lipid membranes relative to conventional liquid-liquid and solid-liquid interfaces, the immense and complex thermodynamic and statistical mechanical theory, the diversity of multiscale lipid models, limitations of modern computing power, the difficulty and ambiguity of simulation controls, finite size effects, competitive continuum simulation alternatives, and the desired application (including vesicle experiments and biological membranes). These issues can complicate an essential understanding of the field of lipid membranes, and create major bottlenecks to simulation advancement. In this document, we hope to clarify these issues, and arrive at a consistent, prioritized, and user-friendly framework for the rational design of lipid membrane molecular dynamics simulations that bridges the gap between a first-timer and world expert.



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1 Introduction

Lipid bilayer membranes have diverse applications in soft matter physics, pharmacology, and consumer products, and are first approximants to biological membranes. Lipid bilay-

ers are quasi-two-dimensional structures consisting of two molecularly-thick layers, or leaflets, on the scale of nanometers in aqueous solvent, where the roughly cylinder-shaped lipids are oriented with their polar head groups outward

and their nonpolar tail groups inward. The dominant driving forces in their formation and stability include hydrophobic and dispersion interactions, wherein the lipid tail groups maximize their contacts with each other and minimize their contacts with water, decreasing area per lipid; and head group electrostatic and excluded-volume repulsion and tail group conformational entropy, which work to increase the area per lipid [3]. Generally, conical and inverse conical lipid amphiphiles more favorably form micelles and inverted micelles, respectively [34], but this simple view of lipids has recently been of debate (Alex...put citations).

Lamellar lipid bilayers often exist as vesicles (also known as liposomes—basically bilayer spheres) in solution, but can also exist as planar bilayers in periodic simulation systems, with experimental supports, or multilayer stacks for structural analysis. Lamellar lipid membranes are interfaces embedded in three dimensions, but are more complicated than typical liquid-liquid interfaces, due to their finite thickness and preferred area per molecule depending on its molecular neighbors and composition [21, 71]. They also differ from solid-solid interfaces, due to their negligible surface tension. Because of these complexities, simple interfacial theories of surface tension in terms of, e.g., an oil-water interface or even a fluctuating interface with capillary fluctuations are often insufficient for membranes.

Fluid lipid membranes are normally modeled as liquid-like laterally (no in-plane shear modulus), and solid-like transversely (out of plane). Another major assumption is that the lipids are strongly surface active, and therefore are not soluble in bulk (aqueous) solvent. The surface is consequently saturated; this achieved trivially through area relaxation in the case of tension control, and in area control, the bilayer buckles to maintain saturation. For lipid bilayer physics, it is almost always safe to assume volume incompressibility, as negligible fluctuations in volume cost far more than typical thermal fluctuations. Area incompressibility is a less common assumption and invalid in many cases; however, area fluctuations are often assumed to exchange with thickness (peristaltic) fluctuations through a simple equation of state (whereby area and thickness are inversely correlated) [71]. For this reason, a Gibbs monolayer (two-dimensional surface) description is a common and often reasonable theoretical approach. Perhaps the most studied fluctuations in lipid membranes, and what separates them from most conventional solid-liquid and liquid-liquid interfaces, are in mesoscopic shape, termed undulations [15, 30]. This is typically approached from the perspective of membrane curvature elasticity, where large wavelength bending modes are highly accessible via thermal fluctuations. These out-of-plane modes lead to a distinction between the projected (in-plane) area and membrane contour area; therefore, care should be taken in deconvoluting

deformations in membrane curvature (bending) from those in contour area (expansion/compression). Still, other fluctuations are accessible at smaller length scales, and typically involve local lipid orientation (“tilt,” relative to mesoscopic shape) and operations thereof, as in other liquid crystalline systems [28, 51, 86, 87].

Despite the extensive physical framework for lipid bilayer membranes from both continuum theory and experiments, detailed molecular simulations can be a tremendous asset to a further understanding. Molecular simulations were first applied to lipid membrane systems in the early 1990s, and have since become increasingly amenable to larger spatiotemporal scales and higher resolutions [78]. In general, molecular simulations work well for lipid membrane studies in the following instances:

- When nanometer resolution is required, and chemical detail is important (e.g. for heterogeneous membranes, and in the case of additional non-lipid components)
- For finite-sized systems where macroscopic continuum principles may not apply
- For a detailed view of thermal fluctuations and the role of entropy
- When the interior of the membrane is being probed, and the two-dimensional/thin film assumption is not a given
- When bilayer self-assembly and the saturation assumption are not a given
- To test continuum mechanical assumptions (e.g. volume/area compressibility, tension, bending renormalization, structure of pores, etc.)
- To parameterize continuum theory and simulations (i.e. with spatiotemporal and macroscopic properties)

In any of the above conditions are met, the researcher should consider the utility of molecular simulations.

In the recommendations that follow, we seek to establish robust guidelines for equilibrium simulations of lipid bilayer membranes. We focus on dilute lamellar bilayer membranes in water (i.e. at high hydration), particularly the membrane fluid (liquid-crystalline, L_α) phase and, where relevant, gel phases (L_β or $L_{\beta'}$). While we do not dictate the choice of MD package for simulation, we draw heavily on tools available in the GROMACS package, which has several built-in routines and add-on patches [1, 13, 47] and is the default for several multiscale lipid membrane models [2, 35, 48, 50, 64]. That said, NAMD [62], CHARMM [10], and AMBER [16] are well established for lipid membrane simulations as well.

2 Prerequisites

Some good textbooks for statistical mechanical and thermodynamic background on membranes include:

- Safran, Samuel A. "Statistical Thermodynamics of Surfaces, Interfaces, and Membranes." 2003: Westview Press.
- Nelson, David R., et al. "Statistical Mechanics of Membranes and Surfaces." 2004: World Scientific Publishing Company.
- Boal, David. "Mechanics of the Cell." 2012: Cambridge University Press, New York.

Good papers and textbooks for computational and simulation guidance on membranes include:

- Sundararajan, V. "Computational Modeling of Membrane Bilayers, Volume 60 (Current Topics in Membranes)." 2008: Academic Press.
- Tieleman, Marrink, and Berendsen. *Biochimica et Biophysica Acta*, 1997.

In this document, we also assume basic proficiency with MD simulation. Where necessary, we refer simulators to other best practices documents for introductory guides:

- MD basics https://github.com/MobleyLab/basic_simulation_training
- MD setup, biomolecular setup <https://github.com/michellab/BioMolSetupPaper>
- Transport properties <https://github.com/ejmaginn/TransportCheckList>
- Statistical error and uncertainty analysis <https://github.com/dmzuckerman/Sampling-Uncertainty>

3 Checklist

Here, we provide a checklist for the four major steps in the lipid membrane simulation process: (1) model selection, (2) pre-simulation considerations (selection of MD settings), (3) preparation of initial configurations, and (4) post-simulation considerations (property validation).

3.1 Model selection

As with other systems, model selection for lipid membranes is crucial. Lipid membrane models are relatively diverse—resolution can range from all-atom to united atom to coarse-grained and from explicit to implicit solvent—and ultimately, the lipid model may be implicated in some more complicated application (e.g. small solute transport, peptide-induced pore formation, embedded proteins). The model selection process for a given physical problem can at times be daunting, especially for an undergraduate, experimentalist, or otherwise newcomer. The main goal for a lipid membrane model study should be to correctly capture the correct structural, mechanical, thermodynamic, and/or dynamic properties (whatever is relevant) at the relevant length and timescales and

the correct equilibrium conditions (thermodynamic, temperature, pressure, etc.) and/or nonequilibrium conditions (thermal/mechanical/chemical/other gradients). However, accuracy must be optimized with efficiency. Generally speaking, the simulation time t_{sim} is a function of the (1) model, (2) system size, (3) computing resources, and (4) MD package, amongst other factors. These contributions are often overlapping, but can be deconvoluted in some simple scaling laws (cf. Section 4.1 for more details). In general, force field developers seek to first capture thermodynamic properties, then address dynamical properties. This balance can be tricky, especially with all-atom force fields, as parameters that work for thermodynamics may not accurately match dynamical properties. Rigorous models are validated via their properties through experimental comparison, but with the proper corrections and normalization (most importantly, due to finite size effects in periodic simulations) [40, 79]. Transferability is ideal, especially for all- and united-atom force fields, but not necessarily a priority for coarse-grained.

3.2 Pre-simulation considerations (selection of MD settings)

Once the model is selected, the pre-simulation considerations mainly concern the thermodynamic conditions under which the membrane simulation is ultimately going to be run. Proper control over membrane phase behavior and mechanical tension often necessitates the use of thermostats and barostats. The relevant thermodynamic ensembles for the study of lipid membranes are the canonical (NVT), isobaric/isothermal (NPT) and multiphase ($NP_z\gamma T$) ensembles. In general, now with well-tuned force fields the semi-isotropic NPT ensemble is recommended for planar bilayers with the constraint that the planar dimensions are equal ($x = y$). Alternatively, the NVT ensemble may be required to avoid cell dimension fluctuations, or the $NP_z\gamma T$ ensemble may be required to probe the effect of surface tension.

The target temperature should be guided by the experimental correspondence. For the ideal model, the simulation temperature would be set to match that of experiment. In reality, however, the imprecise energy-entropy breakdown in the membrane model may lead to shifted phase transition temperatures, and therefore the need to simulate at a higher or lower temperature, depending on the desired membrane phase. However, force fields like CHARMM36 [42] appear to be well-tuned for phase changes within 5°C. The main transition temperature of interest is the gel-to-liquid phase transition temperature T_g , above which the membrane exists in a disordered liquid crystalline L_α state and below which the membrane exists in an ordered gel L_β state. Some models may accurately capture intermediate tilted gel $L_{\beta'}$, ripple $P_{\beta'}$,

and interdigitated $L_{\beta I}$ phases whose relevance depends on the experiments you are trying to model. Most simulations approximate a cellular membrane as a fluid lipid bilayer to match biological conditions, and build chemical and mechanical heterogeneity in later.

For the multiphase and the isobaric/isothermal ensembles, pressure control will additionally be required. For membranes with the NPT ensemble, pressure control is often conducted in a semiisotropic scheme to incorporate the isotropic conditions in the bilayer plane. Since experimental and biological membranes often operate at negligible tension (as their conjugate variable, the area per lipid, is unconstrained and therefore used to minimize the free energy), tensionless membranes are currently the most common. For membranes, the definition of tension is a precarious one that might not be trivial to a newcomer. While there is an important distinction between the frame tension τ and the Laplace tension γ , conjugate to the fluctuating membrane contour area, it has been clearly shown through thermodynamic arguments that these tensions and areas are directly related, and therefore not independent [21]. The Laplace tension is defined to a first approximation as [39]:

$$\gamma = 0.5L_z(P_z - (P_x + P_y)/2) \quad (1)$$

Both tensions reduce to zero when the component pressures are set to be equal.

In general, most MD settings come with the model/force field. For other appropriate simulation settings, see Section 4.2 and https://github.com/MobleyLab/basic_simulation_training

3.3 Preparation of initial configurations

Lamellar lipid bilayers exist for a variety of concentrations of water to lipid ratios. However, all modern force fields will conform to standard bilayers if hydration levels are at full hydration (~ 30 waters per lipid) or an order higher. For a planar bilayer, the desired number of lipids or membrane size in the xy -plane can be estimated from one another if the area per lipid is roughly known:

$$N_l = 2L_{xy}^2/a_l \quad (\text{planar}) \quad (2)$$

where N_l is the total number of lipids, L_{xy} is the box length in the x and y directions, and a_l is the area per lipid. If a mixed lipid component bilayer, then the a_l is the composition-weighted average. The amount of solvent can be independently varied by changing the box z -dimension L_z . The number of solvent molecules can be estimated from the following:

$$N_{solv} = L_{xy}^2(L_z - D_B)\rho_{solv} \quad (\text{planar}) \quad (3)$$

where N_{solv} is the number of solvent molecules, D_B is the bilayer thickness, and ρ_{solv} is the estimated or known bulk density of the solvent model. Similarly, for a vesicle, the number of lipids/membrane size and amount of solvent can be estimated via:

$$N_l = 8\pi R^2/a_l \quad (\text{vesicular}) \quad (4a)$$

$$N_{solv} = (L_{xy}^2 L_z - 4\pi R^2 D_B)\rho_{solv} \quad (\text{vesicular}) \quad (4b)$$

where R is the vesicle radius defined from the center to the bilayer midplane.

For a vesicle, the total membrane area must be smaller than the smallest plane in periodic box; otherwise, the lipids will form a planar bilayer to minimize the free energy. (The cost of forming a vesicle from a planar membrane incurs at least a bending and Gaussian curvature energetic penalty.) Furthermore, simulating a membrane area smaller than the smallest plane in the periodic box does not ensure the formation of a stable vesicle; in fact, a pancake structure (bilayer patch with splayed edges) can be more stable up to a critical size on the order of a 10 nm radius, and therefore 10^3 to 10^4 total lipids [20]!

Once the number of lipids and solvent molecules and periodic box dimensions are determined, there are two main methods for putting them all together: (1) “templating” and (2) self-assembly.

3.3.1 Building the membrane by templating

In the templating method, the lipids are pre-arranged in a planar or vesicular bilayer, close to the final equilibrated structure. Due to potential core overlaps for both planar and vesicular bilayers and the desired leaflet number asymmetry for vesicles (especially at smaller radii), existing packages and routines are recommended. CHARMM-GUI (<http://www.charmm-gui.org/>) is an excellent resource, and the most common for setting up membranes in a variety of configurations and for a variety of models/force fields [18, 36, 37, 90]. Once the membrane is templated, it can then be solvated. Due to the fluctuating nature and molecular scale roughness of lipid membranes, a trial-and-error solvation routine may be preferred over appending solvent “slabs” to each side of the membrane (although judgment can be used to determine routine one will be more efficient). The GROMACS *g_solvate* routine is one such example of a trial-and-error approach, whereas CHARMM-GUI uses a slab-based approach.

3.3.2 Building the membrane by self-assembly

In the self-assembly method, the lipids are dispersed in solvent, and allowed to dynamically and spontaneously arrange into their final equilibrium structure. The proof-of-concept approach is more scientifically satisfying, but not necessarily

reasonable for higher resolution models and/or larger length scale simulations due to the long time scale of assembly. Again, care should be taken to generate a vesicle or bilayer, and not a pancake or other structure. Because the system will always proceed to minimize its total free energy, the initial box dimensions are crucial to the outcome of the self-assembly approach. For a planar bilayer, the in-plane box dimensions must be initialized near their intended end state (which can be predicted with the number of lipids and area per lipid), and for a vesicle, all box areas must be significantly larger than the corresponding planar bilayer with the same number of lipids (otherwise, the lipids will form a planar bilayer and not a vesicle). For one example of the self-assembly technique, you can visit the Martini website (<http://www.cgmartini.nl/index.php/tutorials-general-introduction/bilayers>).

3.3.3 Preparation procedure

After the building step, the system should be minimized (without dynamics, at 0 K) to remove potential bad site contacts (steric overlap) generated during the original placement of molecules. Minimization routines like steepest descent are perfectly reasonable, and are available in conventional MD packages.

Then, the system should be gradually annealed from 0 K to the target temperature in the relevant thermodynamic ensemble (*NVT* or *NPT*, depending on whether or not a barostat will ultimately be used in the production run). In general, a short 100 ps to 1 ns run should suffice. The user may want to consider using position restraints during this process to prevent major lipid fluctuations, particularly if the templating method is used to initially build the bilayer. CHARMM-GUI provides a pre-described set of equilibration scripts that have been tested on various bilayers to work well and is recommended based on their initial structure. For soft enough interactions, you may be able to skip this step.

In the case of self-assembly, a longer dynamical simulation is needed to allow for bilayer formation. For smaller (≈ 128 -lipid) membranes, this can take 10-100 ns, and can potentially take much longer for larger and vesicular ones. Visualization can be very helpful to track the assembly process. Care should still be taken after assembly is qualitatively confirmed to ensure quantitative equilibration.

Before the production run, the equilibrium structure can be further standardized for ease of analysis and comparison with external studies. Out of convention, for an open planar membrane, the membrane in-plane directions are often defined to be the *x* and *y* directions, while the out-of-plane direction is defined as *z*. All subsequent discussion assumes this directionality, but the choice of direction is otherwise arbitrary. In GROMACS, this can easily be achieved with the *editconf* command. For both open planar and closed vesicu-

lar membranes, centering the membrane in the periodic box will largely prevent the jumping of bilayer leaflets and pieces across the periodic box, assuming drift does not occur. This will make subsequent visual observation and post-simulation analysis easier. Centering can also be achieved with *editconf*.

Before going into the details of production run, it should be noted that additional equilibration beyond the initial structure is needed and timescales depend strongly on the system complexity and size. For single-component lipid membranes in the liquid crystalline state, equilibration typically only requires 5-10ns with a well educated initial setup of surface area per lipid. Membranes that have several lipids or conditions in more condensed phases can require 100+ ns of equilibration. A typical first order method to determine equilibration is to track surface area per lipid as a function of time and verify that this has reached equilibrium via block averages and autocorrelation analysis. However, all other key properties should also be verified—i.e., that they have stabilized and that reported averages are over equilibrated systems.

3.3.4 Guidelines for production run sampling

Finally, the initial estimation of the production run time is useful. You will ultimately want to incorporate an adequate amount of sampling for your phenomenon/phenomena of interest. In general, this means capturing several autocorrelation times for the relevant degrees of freedom. Lipid membranes are characterized by a hierarchy of time scales, including: bond vibrations (fs-ns), trans-gauche isomerization and rotation about chemical bonds (1-100 ps), rotation (axial diffusion) about the lipid axis and wobbling (1-100ns), lateral diffusion (1-100 μ s), and flip flop and undulations (1 ms-1 s) [43, 45, 80]. Collective order fluctuations (e.g. undulations, flip flop, lateral diffusion) are the longest timescale fluctuations in the system because they involve the coordination and motion of several lipids [80]. The time of undulations will scale as L^4 [85]. Even at equilibrium, area per lipid can fluctuate on time scale of tens of nanoseconds [63, 78]. However, if your study concerns more localized or molecular degrees of freedom (e.g. rotation about chemical bonds, trans-gauche isomerization, lipid axial diffusion, etc.) at constant membrane macroscopic shape and lipid leaflet number, then smaller sampling times may be permissible.

3.4 Post-simulation considerations (property validation)

Fluid lipid membranes are normally modeled as liquid-like laterally (no in-plane shear modulus) and solid-like transversely. Because of this, important properties include in-plane structure, elasticity, and dynamics and out-of-plane structure and elasticity.

Experimental reference data can be classified as either

direct or indirect. Direct data includes experimentally measured properties (e.g. x-ray and neutron form factors from scattering experiments), while indirect data includes properties inferred from direct experimental data based on a given theoretical model (e.g. area per lipid, bilayer thickness, NMR bond order parameter, lipid diffusion coefficient). Ideally, force field and simulation validation should be based on the comparison of simulated data with direct experimental data, but this is not always possible [63]. Therefore, we discuss both types of data comparisons.

Perhaps the most important verification of any lipid membrane model involves the continuous structural and mechanical profiles in the out-of-plane ("transverse", or z) direction. The lipid bilayer membrane's internal structure and mechanics are crucial to its physics. These metrics include (1) the local density profile and (2) the stress profile.

3.4.1 Transverse local density profile

As for general liquid-liquid interfaces, the density profile $\rho(z)$ is a crucial structural metric for lipid bilayer membranes. Various regional models have been proposed to characterize the membrane based on its component and overall densities. For atomistic models, the local density profile is explained by a four-region model. The region numbering scheme proceeds from the exterior water layer to the bilayer midplane, and the typical density hierarchy is as follows: region 2 ("interface") > region 3 ("soft polymer") > region 1 ("perturbed water") > region 4 ("decane") [76]. The local density can be calculated through most conventional MD packages, although care should be taken to ensure appropriate averaging, especially for larger, more appreciably fluctuating membranes. Be careful with programs like *g_density* in GROMACS, which appear to tabulate density averages based on absolute coordinates and not relative positions (e.g. aligning profiles each frame by the apparent membrane center). Relative positions can be valuable when there are even minor fluctuations. Failure to correct for this can result in smearing of the density profile; one possible outcome is that region 4 (local lipid density drop in center) is not properly captured when it may exist for a given model. For larger membranes with significant undulations, the order parameter for the density profile (i.e. the relative z -coordinate location) must be replaced with a more appropriate one that reflects the depth into the fluctuating interface. The local density profile can be directly compared with x-ray and neutron scattering experiments through conversion of atomic density profiles to electron and neutron density profiles, respectively, and transformation of these profiles to Fourier space. Care should be taken here as well to account for geometric differences between planar and vesicular membranes (see Section 4.1.5). For AA MD data comparisons, there are several thorough studies [8, 41]. Elec-

tron densities proceed generally in the same trends as above [42]. For more information on techniques, the following reference is instructive [63].

3.4.2 Transverse lateral stress profile

The lateral stress profile (in the transverse direction) $P_{xy}(z)$ is a more unique mechanical metric that is highly relevant for lipid bilayers because (1) it proves whether or not your model qualitatively and quantitatively captures the competing forces in lipid bilayer assembly and stability and (2) it is the gateway to calculating a host of mechanical properties. Despite its importance, however, the stress profile is often more difficult and expensive to calculate. One means of calculating is through GROMACS-LS, a customized version of the MD package GROMACS. GROMACS-LS can even calculate stress component profiles, including those arising from van der Waals and electrostatic interactions. For more information about package and theory: <http://mdstress.org/> [58]. As an important verifying metric of force field development, the stress profile can be thoroughly compared with existing atomistic simulation studies. There is no direct means of stress profile comparison with experiments, though as we will explain, properties calculated from the stress profile can be compared with experiment (Section 3.4.6).

3.4.3 Bilayer thickness

The membrane thickness is a structural metric that is consequently calculated from the density profile. Three experimentally-relevant definitions include (1) the Luzzati (total lipid) thickness D_B , (2) the head-to-head distance D_{HH} , and (3) the hydrocarbon thickness $2D_C$. The Luzzati thickness is relevant to neutron scattering, and is calculated as the distance between the two locations on each side of the bilayer where the water density drops to one half its bulk value. This thickness metric in reality is based on the spatial profile of protiated and deuterated water, and is physically indicative of the degree of water penetration into the bilayer [63]. The head-to-head distance is relevant to x-ray scattering, and calculated as the distance between the two peaks in the electron density profile. More simply, this can also be approximated as the distance between the maximal phosphate group densities in each leaflet (relevant to coarser lipid models) [63]. The hydrocarbon thickness is also an important measure when comparing with transmembrane proteins and their length of surface exposure of the hydrophobic residues. All of these thickness calculations can be indirectly compared with experiment. For comparison, the experimental results are typically converted to real space. Typical values for phospholipid bilayers in simulation and experiment are around 3 to 5 nm.

3.4.4 Area per lipid and NMR order parameter

Two important structural parameters, complementary to one another, are (1) the area per lipid (providing information about in-plane structure or lateral density) and (2) the deuterium NMR order parameter (for out-of-plane structure).

The area per lipid a_l , or in-plane area occupied by a given lipid, is a critical target in force field parameterization of lipid membranes [42, 78]. In simulation, the area per lipid is typically calculated via

$$a_l = (L_x L_y) / n \quad (5)$$

where L_x and L_y are the lateral box dimensions, and $n = N_l/2$ is the number of lipids per leaflet [63]. This equation assumes a lipid number symmetric bilayer with negligible undulations, such that the contour (membrane) and projected (periodic box) areas are roughly equal [42, 78]. However, for larger membranes, undulations will lead to significant differences between the contour and projected areas. Theoretically speaking, undulations will increase the ratio of contour area to the projected area, and this is specifically because the undulations reduce the projected areas [78]. The area per lipid is rigorously an intensive, or size-independent, quantity, so the appropriate steps should be taken to normalize for significant size effects when necessary before experimental comparison. This may warrant simulation of different membrane sizes, extrapolating results to zero system size to allow for convergence of contour area to frame area and arriving at a size-independent metric [81]. However, the area per lipid may not vary much across typical sizes of MD simulations, or even in experiments. In any event, care should be taken to ensure proper statistics. Even at equilibrium, the area per lipid can fluctuate on a time scale of 10-100 ns [63], especially for complex bilayers and for those with many components. For multicomponent membranes, the membrane should be partitioned into individual values for each lipid species (cf. Voronoi tessellation-based methods, grid-based methods using atomic van der Waals radii, etc.). Partial molar areas can be determined, but this requires that a range of concentration ratios be simulated. The area per lipid can be compared directly with other simulations; normalized size-independent metrics should be obtained where possible.

The deuterium P_2 NMR order parameter S_{CD} describes the alignment of lipid constituent bonds with the global membrane normal, and will vary along the length of the lipid tail group chains and between liquid disordered and ordered membranes. The metric is defined as:

$$S_{CD} = \frac{1}{2} \langle 3 \cos^2 \theta - 1 \rangle. \quad (6)$$

A value of 1 indicates perfect alignment of the chain with the global bilayer normal, while -0.5 indicates anti-alignment.

This metric, however, can be ambiguous. For example, a value of zero can mean either that the lipids are isotropically disordered with respect to the bilayer normal, or that the lipids are perfectly oriented at a constant angle equal to the “magic angle” of 54.74° [63]. Computing error bars is tricky, but the *order_parameters* tool in LOOS offers one approach. Since the area per lipid and NMR order parameter are tightly coupled, a similar magnitude of statistics (≈ 10 -100 ns) is required for S_{CD} as well. Calculated metrics can be compared with other simulations and indirectly with quadrupolar NMR splitting experiments. From the top of the acyl chains to the bottom, values typically rise from about 0.17-0.20, then fall from around 0.20-0.22 to 0.10 for a fluid phase phospholipid bilayer. Averages across the entire chains are therefore typically around 0.17 [78].

While the area per lipid cannot be directly compared with experiments, due to the complications of lateral and transverse structure fluctuations, it can usually be inferred from a theoretical model that sometimes involves S_{CD} . One example simply uses the lipid volume V_L and the Luzzati thickness: $a_l = 2V_L/D_B$. This assumes, however, that the lipid volume can be estimated accurately [63]. Another approach uses the deuterium NMR order parameter: $a_l/\text{chain} = 2V_{CH_2}/((1+2S)b_{cc})$, where V_{CH_2} is the volume of a CH_2 group, S is the plateau value of S_{CD} (maximum described above) and b_{cc} is the projected C-C bond length along bilayer normal [57]. Practically, these comparisons are difficult because S_{CD} contains contributions from conformational disorder, local lipid tilting, and assorted collective motions [78]. The value for double-tailed phospholipids is generally larger than single-chain hydrocarbons in systems like SAMs (60 Å²/molecule versus 30-40 Å²/molecule).

3.4.5 Lipid lateral diffusivity

The lipid lateral diffusivity D_l is used to characterize lipid mobility, to gain insight into collective lipid motion timescales, and also potentially to discriminate between liquid crystalline and gel phase lipids. The relevance of a diffusivity is predicated on the assumption of classical in-plane diffusion. As such, the diffusivity can be determined from the slope of the average lipid mean-squared displacement (MSD) plotted against lag time (equal to $4D_l$ for 2D diffusion). In the MSD calculation, the user should be careful of artifacts due to lipid molecules partially or fully jumping across periodic boxes; periodicity can be accounted for by reimaging where appropriate. For homogeneous, single-component lipids, averages are normally taken over all lipids. Results typically fall around $O(10^{-8}$ to 10^{-7} cm²/s). Simulation results can vary significantly with the force field, system size, truncation scheme for long-range interactions, time step, and non-bonded interaction pair list update frequency [63]. Venable et al. recommends that, when reporting and comparing with other simulations,

the system size in particular should be noted and accounted for. Given the hydrodynamic theory, the lateral and transverse dimensions would be even more explicit, and ideally, studies should report extrapolated infinite size values with a confidence interval [79]. Results can be indirectly compared with experiment. Experimental estimates can generally vary over three orders of magnitude, due to a number of reasons (cf. Section 4.4.1 for more details) [63].

3.4.6 Mechanical/elastic properties

Mechanical properties for membranes can be calculated from a number of techniques that can be broadly classified by (1) equilibrium fluctuations, (2) stress profile, and (3) biased/active deformations. We discuss the merits of each umbrella of approaches in general terms in Section 4.4.2. In this section, we merely attempt to recommend the best technique(s) for a given property. These mechanical properties include: (1) lateral tension, (2) area compressibility modulus, (3) bending modulus, (4) monolayer spontaneous curvature, (5) Gaussian curvature modulus, and (6) line tension.

(1) Calculation of the lateral tension γ from a membrane simulation is a way to calculate the variable conjugate to the membrane [contour] area in canonical simulations, and in tension control, a way to directly confirm the simulation controls for the intended ensemble. In model development, the tension is a parameter probed to capture the correct area per lipid (which should ideally occur at zero tension) [93]. The tension is related to the zeroth moment of the stress profile, and can be easily approximated with the Kirkwood-Buff equation Equation 1 above.

(2) The area compressibility modulus K_A determines a membrane's ability to compress and expand, and is another important property in force field parameterization [78]. The recommended way to calculate K_A is based on area fluctuations in the tensionless ensemble:

$$K_A^{app} = k_B T A_p / \sigma_{A_p}^2 \quad (7)$$

where K_A^{app} is the apparent area compressibility modulus based on the fluctuations in projected area $A_p = L_{xy}^2$. This equation is fairly simple, but involves two major nuances. The first is that the calculation may take 100 ns to converge [78], if not longer. The second is that this is an apparent value based on size-dependent projected area fluctuations, and is not the end point for characterizing the intensive, size-independent in-plane compressibility. The recommended correction to deconvolute expansion-compression modes from undulatory modes is to simulate multiple membrane sizes and extrapolate the results to zero system size, where undulations no longer exist [81]. At smaller simulated system sizes, the result of not accounting for undulatory contributions can be negligible (~ 5 -10% correction with experiment), but becomes

substantial and must be corrected for larger system sizes (where they can be $\sim 50\%$ larger than experimental results) [78, 81]. Results typically vary between 100 and 400 mN/m [78], and are often compared with micropipette aspiration experiments [67]. Results can also be compared with predictions from polymer brush theories, where K_A is sometimes related to the oil-water interfacial tension $\gamma^{o/w}$ —for example, $K_A = 6\gamma^{o/w}$ [67].

(3) The bending modulus (bending rigidity/constant) κ determines the ability of the membrane to bend. The undulation spectrum method [25, 46] is the traditional and well-established way to calculate κ . The spectrum describes the height-height correlations $|h_q|^2$ of the membrane continuum shape, and for a tensionless membrane, the large wavelength/small wavevector (q) behavior follows an inverse fourth power relation with a constant of proportionality that contains the bending modulus [25, 46]:

$$\langle |h_q|^2 \rangle = k_B T / \kappa q^4 \quad (8)$$

The undulation spectrum method, however, must be used over "mesoscopic" length scales (approximately ten times the bilayer thickness, or about 50 nm / 5000 lipids) that are out of reach for AA and most CG simulations. If simulations are too small, deviations to the undulation spectrum will result from individual lipid tilting (below ten thicknesses) and protrusions (below three thicknesses) [78]. Also, there will simply not be a large enough range of wave vector magnitudes to fit the spectrum and obtain a bending modulus estimate. For this reason, we recommend the lipid director field spectrum approach [78, 84], which analyzes thermal fluctuations of lipid orientation via a director vector field \hat{n}_q . Specifically, the longitudinal component \hat{n}_q^{\parallel} relates to the macroscopic bending modulus through an inverse second power relation in the wave vector:

$$\langle |\hat{n}_q^{\parallel}|^2 \rangle = k_B T / \kappa q^2 \quad (\text{recommended}) \quad (9)$$

The lipid director field spectrum method works well for "modestly sized" membranes, down to approximately three bilayer thicknesses (≈ 12 -15 nm); AA simulations of 648 lipids have been shown to be well converged. This general approach also provides a route to calculating bilayer tilt and twist moduli. Both the undulation and lipid director field spectrum approaches will take at least 100 ns to converge [78]. Technically speaking, the bending modulus is not a material property; it is dependent on system size, and decreases at larger wavelengths λ as $\kappa[\lambda] = \kappa_0 - 3k_B T / 4\pi \ln(\lambda a)$, where a is some constant [61]. The determination of the appropriate size renormalization is somewhat ambiguous, but the length scale of the relevant experimental system can be used as a guide. Bending modulus results typically vary between

10 and 40 $k_B T$ [78], and can be compared with a host of experimental techniques, including flicker spectroscopy and micropipette aspiration. See Section 4.4.2 for more details. Results can also be compared with polymer brush theory, which relates κ to K_A and the bilayer hydrophobic thickness h_{phob} via $\kappa = K_A h_{phob}^2 / 24$ [67].

(4) While lipid bilayers (planar and vesicular) have a net zero bilayer spontaneous curvature C_0 , their monolayers individually may have a propensity to curve. This propensity is quantified by the monolayer spontaneous curvature c_0 , and is defined to be positive for a lipid that forms micelles and negative for one that forms inverted micelles [78]. Together with the monolayer bending modulus κ_m , c_0 is related to the first moment of the stress profile, integrated from the bilayer midplane to the upper edge of the simulation cell [71]:

$$\kappa_m c_0 = \int_0^{L_z/2} z [P_{xy}(z) - P_z] dz \quad (10)$$

Thus, if κ_m is known, then c_0 can be quantified. κ_m is predicted by elastic theory to be one half the bilayer value (κ). Monolayer spontaneous curvatures are typically calculated indirectly in experiment from studies in the completely different inverted hexagonal H_{II} phase, where lipid monolayers are assembled in long, hexagonally-arranged water-filled tubes. Results are then extrapolated to the lamellar phase. This is hard to study for lipids like DPPC that are more cylindrical in shape, but easier for those like DOPE that are more like inverted truncated cones (due to acyl chain unsaturation) [34, 78].

(5) The Gaussian curvature modulus κ_G describes the propensity of the membrane to change topology, and is especially important for fission and fusion events [31]. In general, even simulation methods for calculating κ_G are controversial. The difficulty in calculating κ_G lies in controlling topology and boundary behaviors in which Gaussian curvature plays a role [31]. The stress profile approach, specifically involving the second moment, is not reliable here [31, 32]. We recommend the patch closure method, which has shown promise in preliminary work [31]. While there is virtually no means for experimental comparison, elasticity theories offer simple predictions, particularly in the approximation $\kappa_G \approx -\kappa$ [20, 31, 66].

(6) A line tension (edge energy) Γ is an energy per unit length that can describe lipid phase segregation and hole formation. Specifically for pores (hydrophilic holes, where lipids splay to connect the two leaflets), the line tension can be studied through simulation of a bilayer "strip" or "half-connected bilayer" (with exposed bilayer edges in one in-plane box dimension). The bilayer is thus periodic in one lateral dimension and exposed in the other, resulting in lipid splaying and therefore a rim on both sides. In this scenario,

the line tension can be determined from the stress profile, specifically the lateral normal stresses. If the strip is periodic in y and non-periodic in x , then:

$$\Gamma = \frac{1}{2} L_x L_y (P_{xx} - P_{yy}) \quad (11)$$

and $P_{xx} = P_{zz}$ [77]. Typical values are around 35 to 50 pN in simulation [77, 89] and 5 to 30 pN in experiment [9, 54, 95], the latter of which are typically determined through dynamical pore closure studies.

3.4.7 Leaflet-dependent properties

While thickness, area per lipid, deuterium order parameter, and lipid lateral diffusivity calculations do not necessarily require leaflet identification, other metrics do. Leaflet identification and the relevant metrics are discussed in Section 4.4.3.

3.4.8 Property validation summary

It is the responsibility of computationalists in the field of membranes and otherwise to adequately validate the physical properties resulting from their model. The method of calculation in experiment does matter to the method of calculation in simulation. For example, methods based on biased membrane perturbations may deviate significantly from those based on equilibrium fluctuations (e.g. micropipette aspiration versus flicker experiments) [78]. Wherever possible, the theoretical basis for both the experimental and simulation property calculations should be used as a guide. A final point is that experimental results are not infallible. When simulation and experiment do not agree and one is fairly confident in the simulation procedure and results, the experimental results can be questioned [78].

4 Rationale

In this section, we elaborate on items in the checklist. Where relevant, we include details on alternatives, compare different approaches, and provide physical and literature justification.

4.1 Model selection

Efficiency is a major concern in virtually any lipid membrane simulation. In the MD loop, the most expensive step involves the pairwise force evaluations, and therefore the number of particles in your system N and the system density $\rho_{model} \propto 1/a_{model}^3$, where a_{model} is the average spacing between sites in the model. Exact simulation time t_{sim} scalings depend on the force field, and can range from $N\rho_{model}$ for short-ranged/mean field types of force fields to $N \log N$ (e.g. PME) to N^2 for long-ranged/rigorous pairwise interactions. Since the number of particles can be related to the system

1. MODEL SELECTION

Goal: set up lipid membrane model to accurately capture properties of interest in an efficient manner.

- ☐ Lay out problem/phenomenon of interest, and establish whether or not molecular simulations are needed.
- ☐ If molecular simulations are needed, determine:
 - ☐ Available computing resources (cores, memory, storage, etc.)
 - ☐ Desired lamellar membrane configuration (planar vs. vesicular). See Section 4.1.5 for details.
 - ☐ Relevant length and time scales of the system of study (or, if possible, of an appropriate subsystem). See Sections 3.3.4 and 4.1.4 for a discussion.
 - ☐ Primary properties of interest (structural, mechanical, thermodynamic, and/or dynamic)
 - ☐ Required spatiotemporal resolution
- ☐ Based on the above considerations, determine the optimal model in terms of accuracy and efficiency.

2. PRE-SIMULATION CONSIDERATIONS (SELECTION OF MD SETTINGS)

Goal: establish rational and physically-intuitive settings for the desired lipid membrane simulation.

- ☐ Determine desired thermodynamic ensemble.
- ☐ Given selected model and experimental correspondence, determine target temperature.
- ☐ If relevant, determine pressure control settings.
- ☐ Determine remaining settings necessary for MD configuration file.

3. PREPARATION OF INITIAL CONFIGURATIONS

Goal: build reasonable equilibrium starting structure for a lipid bilayer in a periodic simulation box.

- ☐ From the desired membrane size and geometry, estimate the number of required lipids (Equations 2 and 4a).
- ☐ Estimate the number of solvent molecules and full periodic box dimensions (Equations 3 and 4b).
- ☐ Using the above estimates, proceed through to system setup. See Sections 3.3.1 and 3.3.2 for options.
- ☐ Minimize (no dynamics) to remove bad site contacts
- ☐ Anneal from 0 K to target temperature
- ☐ Dynamics
 - ☐ For a templated membrane, just equilibrate
 - ☐ For membrane self-assembly, assemble, then equilibrate
- ☐ Qualitatively confirm equilibrium structure and quantitatively confirm thermodynamic averages before a production run.
- ☐ Rotate/re-center system as necessary.
- ☐ Determine reasonable amount of sampling for production run. See Section 3.3.4 for details.

SIMULATION PRODUCTION RUN

4. POST-SIMULATION CONSIDERATIONS (PROPERTY VALIDATION)

Goal: comprehensively justify simulation procedure and model selection via rigorous production run analysis.

- ☐ Calculate and validate density profile via scattering form factors, and if possible, calculate stress profile.
- ☐ Calculate and validate thickness, area per lipid, deuterium NMR order parameters, and lipid lateral diffusivity.
- ☐ Calculate and validate mechanical properties as necessary. If required, set up and run additional simulations (e.g. for more time, with external biases, etc.) for certain property determinations.
- ☐ Determine if leaflet identification is necessary for other metric calculations. Proceed as necessary.

density ρ_{model} and the system length scale L (e.g. for a cubic box):

$$N = \begin{cases} \rho_{model} L^3 & (\text{explicit solvent}) \\ \rho_{model}^2 L^2 & (\text{implicit solvent}). \end{cases} \quad (12)$$

This scaling incorporates the model resolution ρ_{model} and system size L /nature of solvent, respectively. Thus, for short-ranged interactions:

$$t_{sim} \propto \begin{cases} \rho_{model}^2 L^3 & (\text{explicit solvent}) \\ \rho_{model}^2 L^2 & (\text{implicit solvent}) \end{cases} \quad (13)$$

and for long-ranged:

$$t_{sim} \propto \begin{cases} \rho_{model}^2 L^6 & (\text{explicit solvent}) \\ \rho_{model}^2 L^4 & (\text{implicit solvent}). \end{cases} \quad (14)$$

There is additionally a system size contribution (added to the geometric one) that accounts for sufficient sampling. This accounts for the largest wavelength undulations that are the slowest degree of freedom, and scale as L^4 [85]. Thus, the overall scaling in system size can be from L^6 up to L^{10} , depending on the range of interactions and presence/absence of solvent. In other words, at a minimum, an order of magnitude increase in membrane length scale leads to six order of magnitude increase in computational expense! Coarser models may contribute to higher accessible time scales in two ways: (a) by increasing the time scale of the fastest (vibrational) mode (i.e. $t_{sim} \propto 1/\Delta t_{model}$) and (b) by also inherently smoothening the free energy landscape, and therefore enhancing dynamics across it (e.g. via a simple scale factor). Computing resources and the specific MD package can be viewed as (a) affecting the range of possible parallelization schemes (described in speedup by e.g. Amdahl's Law, but convoluted with the specific MD scheme, e.g. domain decomposition) and (b) contributing to some intrinsic scale factor related to the hardware type (e.g. CPUs vs. GPUs) and software type.

4.1.1 Survey of lipid membrane models

In general, there is a very diverse range of models that can be leveraged for simulations of lipid bilayer membranes. Atomistic (all-atom, or AA) models are the “gold standard,” as is the case with MD simulations of most other systems. The quality of AA MD simulations of lipid bilayers has improved dramatically since their initial development in the early 1990s [78]. AA models can have hundreds of atomic sites per lipid molecule. United atom (UA) force fields remove hydrogen atoms for an increase in efficiency (factor of ~ 2), and are becoming competitive with AA force fields in accuracy. AA and UA can therefore easily reach the 100 ns time scale and 5 to 10 nm length scales [75], but with the appropriate resources and GPU-enabled codes, microsecond timescales are

attainable. Well-validated force fields include AMBER Lipid14 (AA) [22], CHARMM36 (AA) [42], GROMOS 54A7 (UA) [64], and Slipids (AA) [35].

Coarse-grained (CG) models have also been well-developed for efficient, large-scale simulations of lipid membranes, often where the interest is in mechanical and qualitative behaviors and less in the quantitative and chemical detail. That said, systematic CG models can still retain some level of chemical specificity (e.g. the types of lipids that they represent). CG models can easily reach 1000 ns (1 μ s) time scales and ~ 20 nm length scales [75], but can go beyond this toward the 100 μ s with the proper resources. One of the best-known CG models for membranes is the Martini force field [48, 50], which via a 4:1 heavy (non-hydrogen) atom mapping reduces to about ten pseudoatom sites per lipid. Martini is a hybrid bottom-up and top-down method of coarse-graining, combining non-bonded interactions parameterized to capture experimental partitioning free energies and bonded interactions parameterized from AA simulations [49]. Because the aqueous solvent can contribute up to 90 percent of the force evaluations [2], implicit solvent (IS) simulations can be much more efficient and potentially advantageous. The Dry Martini force field is one of many such examples, and offers a factor of between 2 and 100 speed-up relative to conventional (“wet”) Martini [2]. There are many other examples of both explicit and implicit solvent CG models for lipid membranes [6, 66]. However, CG models in general and IS CG models in particular are sometimes deficient in capturing solvent-mediated effects, entropic driving forces, and hydrodynamics. The wet Martini model in particular has paired lipids with many different types of CG water models; successive models incorporate crucial features like polarity, but at the expense of computational efficiency. With the intuition that the effective thermodynamic properties are already well captured with IS CG Martini, recent work has sought to improve dynamical properties via a hydrodynamics thermostat, with minimal computational overhead to model the solvent (about a factor of four increase in efficiency relative to the corresponding explicit solvent model) [93].

Phenomenological, generic, and toy models work in more of a top-down fashion by incorporating and tuning minimalistic interactions to obtain the desired phenomena and properties. Examples include IS CG models that have been developed with five [7] and three [19] sites, and recent efforts have sought to incorporate hydrodynamic thermostating in the latter model as well [82]. These models extend the range of accessible scales even further to 100 μ s and 100 nm [75]. These types of simulations have executed as low as two- or three-site lipid models in conjunction with a dissipative particle dynamics (DPD) thermostat (i.e. DPD simulations). Practically speaking, only two sites per lipid are needed (one hydrophilic and one hydrophobic) to capture amphiphilicity

and consequently self-assembly into lipid bilayers.

For lipid membranes, there is also an extensive subcommunity that uses continuum mechanical theory and field-theoretic simulations that are sometimes in fact the preferred approach at larger length scales (100 nm to 100 μm) due to their efficiency [12, 72, 75]. Simulations are performed on the basis of the above continuum theoretical descriptions and by parsing different deformation modes and perturbing them, accepting or rejecting perturbations based on a Metropolis criterion [5]. It is often crucial to compare with these techniques wherever possible. If some of the above conditions motivating the use of molecular simulations are not met (Section 1), it is useful to evaluate whether or not a continuum approach would be better.

It should be evident from the thermodynamic and statistical mechanical framework above that there are some crucial considerations for any lipid membrane model, regardless of resolution, including: (1) membrane composition, (2) other thermodynamic constraints, (3) model size, and (4) model geometry.

4.1.2 Membrane composition

It is generally important to consider the chemical mapping of the model to the real system, especially for multicomponent membranes. The desired heterogeneity and particular lipids may determine the model you ultimately choose. In lipid membrane simulations (for both AA/UA and CG resolutions), the canonical system is DPPC (dipalmitoylphosphatidylcholine, with two fully saturated 16-carbon chains) in water. DPPC is a lecithin, a common choice for vesicle experiments, and a major component of pulmonary surfactant. This is typically the system for which new force fields are first tested. DPPC is sometimes not preferred in experiments, due to its high melting point (from the gel to liquid-crystalline state). A more relevant lipid would be the 14-carbon chain DMPC or one that has a chain with a single double bond (unsaturation) like POPC.

The recent progression in the field is to go beyond single-component membranes toward more realistic membrane mixtures [38]. For multicomponent membranes, there is a well established body of literature. For phase coexistence studies, typical model experiments consists of a ternary mixture of cholesterol and both saturated and unsaturated lipids [4], but more biologically-relevant studies include greater than three lipid types [38]. In fact, for biologically-relevant simulations, we advise caution in the selection and relative composition of lipids in the membrane model. While there are good guidelines for the contributions of major lipid head and tail groups to biological membranes, composition can potentially vary across different domains and even between the two leaflets [33], and the best choice for a given model will

depend highly on the analogous experimental system. The development of biologically-relevant membranes is at the forefront of the field for AA and CG models [38, 52], and in probing biological processes, one must ensure a membrane of the appropriate phase and composition (e.g. proteins may function best in their native lipid environment, with certain lipid types).

4.1.3 Other thermodynamic constraints

Given a membrane's composition, the thermodynamic constraints of temperature and tension (or area) will largely determine phase behavior. As discussed above, simulations are sometimes amenable to different constraints from experiments (Section 3.2), but the appropriate experimental conditions can be achieved in a corresponding simulation ensemble. It is worth noting that certain models, particularly CG ones, sometimes experience difficulty in capturing phase transition temperatures and even entire phases (for example, subgel and ripple for Martini) [69].

4.1.4 Model size

Whether or not the membrane physically reflects the experimental setup also depends largely on the dimensions of the model. It has been shown for membranes that finite size effects can play a significant role for thermodynamic (especially mechanical) and dynamical properties [17, 78, 79, 81]. This refers not only to the in-plane dimensions, but also for the out-of-plane one; despite the quasi-two-dimensional structure of membranes and two-dimensional approximation at larger length scales, hydrodynamic theoretical models for periodic systems have shown that the thickness of the water layer(s) matter as well in convergence to macroscopic system dynamics [79]. In determining the model dimensions, one should search for the emergent length scales in the experimental system that can serve as the periodicity length scales in the simulation. For biological membranes, there is experimental evidence to show that an appropriate in-plane length scale is around 150-500 nm. This is set by the cortical cytoskeletal mesh, which pins membrane proteins and therefore constrains lipid motion (anchored protein picket model) [53, 68]. This is too large for most molecular simulations, but if a highly resolved picture of biological membranes is still desired, different subsystems can be simulated. If you're not going to simulate the experimentally relevant size (of the overall experimental system or a relevant subsystem), then you need to be able to normalize your results with respect to the difference in sizes.

4.1.5 Model geometry

In many cases, experimental vesicles are modeled with planar bilayer simulations. This may raise questions about the

meaning of the results, as vesicular membranes are the result of a balance of positive strain on the outer leaflet and negative strain on the inner leaflet, while planar membranes have on average zero strain on each leaflet. Furthermore, vesicles often have a different number of outer and inner leaflet lipids. Rigorously speaking, there are mathematical transformations to convert data between vesicles and the corresponding planar bilayer. For more information on how to do this, see Abel Transforms. However, large enough vesicles are also locally flat, so a planar membrane can be a good approximant. The extensivity of the experimental vesicles can be used as a guide for the simulation size (or the size to which you normalize your results).

4.2 Pre-simulation considerations (selection of MD settings)

Technically, *NVE* (pure MD) is the correct ensemble for all MD simulations, and is strictly the only ensemble in which pure dynamics are observed (cf. https://github.com/MobleyLab/basic_simulation_training). To roughly conserve energy and prevent drift, the integration settings (time step, etc.) matter. Since periodic center of mass removal can hide integrator artifacts, removal should ideally only occur at the start of the simulation [93]. However, as mentioned earlier, proper control over membrane phase and tension often necessitates the use of thermostats and barostats. In some cases, this (as well as the method for calculation of long-range electrostatics) can lead to significant drift when the center of mass motion is not removed [93]. It has been shown, however, that weak-coupling thermostats and barostats and periodic center of mass motion removal have a negligible role on lipid membrane dynamics, and that these thermodynamic controls rigorously correspond to the associated statistical ensemble [79].

Additionally, thermostats can potentially affect hydrodynamic interactions. In general, MD thermostats that periodically randomize velocities disrupt velocity correlations, and therefore hydrodynamic flows [93]. For membranes, this can significantly affect in-plane lipid correlations and perturb lipid lateral diffusivities. In particular, the Langevin thermostat (sometimes recommended for implicit solvent coarse-grained (IS CG) models to nonspecifically account for otherwise absent solvent collisions) does not technically conserve momentum [26]. Other stochastic dynamics thermostats do conserve momentum, and have been built to accurately capture long-range hydrodynamics for coarse-grained and implicit solvent models [82, 93].

There are some additional subtleties to barostat compressibilities for lipid membrane simulations. Inverse to some other interfacial simulations (e.g. SAMs on an inorganic sur-

face in water), where the in-plane compressibilities are set to zero to preserve hydrocarbon area per molecule, tilt, and density, membrane simulations are usually set to be compressible in the *xy* plane, and sometimes even incompressible in *z* (especially for IS CG models).

4.3 Preparation of initial configurations

In terms of templating methods, CHARMM-GUI is perhaps the most commonly used package. CHARMM-GUI packs lipids from a library, then relaxes atom clashes on its own. During the building and equilibration schemes, CHARMM-GUI performs internal checks for "disaster structures" that can range from ring penetration (molecular chains going through rings) to flipped chiralities of lipid backbones, and additionally includes built-in restraints to maintain chirality. That said, the user should still perform spot checks and, if necessary, conduct more extensive structural analysis. An alternative is the OptimalMembraneGenerator from LOOS, which is newer and less commonly used. OptimalMembraneGenerator operates in the NAMD package only, and packs lipids from a library, but uses a scaling procedure such that no disaster structures result. For multicomponent membranes, there are some tools to help build membranes given the desired lipid types and proportions. One example is insane.py [83] (<http://www.cgmartini.nl/index.php/downloads/tools/239-insane>) for the Martini CG force field, which like OptimalMembraneGenerator also uses a scaling procedure to avoid bad structures. Insane is well established in user controls, and largely prevents user errors in setup; the downside, therefore, is that it sacrifices user creativity and customizability.

Self-assembly, or even combining pre-existing bilayers to make a larger one, can result in lipid number asymmetric membranes. While there is an overall propensity for the lipid number to equalize for a planar, tensionless membrane with a single component of lipids, the process can be extremely slow. In fact, lipid leaflets are symmetrized typically through collective motions to form a pore, which then facilitates flip flop. Single lipid leaflet crossing is a highly activated event. Membranes are generally fairly stable to minor lipid number asymmetries (area per lipid mismatches of up to 5%) [60]. Still, in general, unless there is an explicit reason for studying lipid number asymmetry, one should ensure a leaflet symmetric membrane.

Depending on the application, the two build methods can vary significantly in their efficiency and final outcomes. In Table 1, we outline some major advantages and disadvantages of both.

Table 1. Membrane building methods: a cross-comparison

Method	Advantages	Disadvantages
Templating	<ul style="list-style-type: none"> • Directly construct a sane-looking bilayer • Efficient 	<ul style="list-style-type: none"> • Doesn't capture preferential segregation of multicomponent bilayers, which can be slow to emerge • You need to know what's in what leaflet, etc.; can lead to user bias if arrangement is not known
Self-Assembly	<ul style="list-style-type: none"> • "Natural": lets things assemble the way they want • Great if you know the overall system composition but not distribution (across leaflets, within leaflets, etc.) • Easy (at least with CG models) – scatter molecules and run simulation 	<ul style="list-style-type: none"> • Less reproducible – bilayers won't necessarily be symmetric, and leaflets won't necessarily have same composition • Can be problematic with small systems • Relatively expensive

4.4 Post-simulation considerations (property validation)

4.4.1 Nuances to dynamical property calculations

Subdiffusion, a distinct form of anomalous diffusive motion characterized by long-range correlations in time or space, has been recognized in many biological systems, including diffusion in the crowded cytoplasm, the internal dynamics of proteins, and the gating of ion channels [27, 44, 56, 88]. The physical origin of subdiffusion, and whether it is truly present, remains controversial for some systems [73]. However, there is growing evidence for transient subdiffusion in the lateral dynamics of lipids in phospholipid bilayers [55]. The subdiffusive regime has been shown to exist between the ballistic and random (linear MSD) walk regimes, spanning as many as five orders of magnitude in time and timescales reaching many seconds in multicomponent membranes. While this potentially has crucial consequences for dynamical validation of membrane models, its application to a robust protocol for multiscale lipid models is at this point unestablished.

There is a major box size dependence for dynamic properties in MD simulation. In general, diffusive dynamics in confined, periodic simulation systems are perturbed relative to the macroscopic limit, and can be corrected through the application of hydrodynamic theories. Lateral diffusive dynamics in lipid membranes suffer from significant finite size effects (factor of 3 to 4 for AA MD) [40, 79] relative to bulk dynamics in a homogeneous fluid (~10-20%) [92]. Because of longer-ranged hydrodynamic correlations for membranes, convergence is expected to be even slower than an inverse box length convergence in 3D [14]. Camley et al. has adapted the Periodic Saffman-Dellbrück theory describing hydrodynamics of a periodically-replicated membrane suspended in an infinite bulk fluid for cylinders spanning a single leaflet

(i.e. lipids). The model additionally accounts for the influence of interleaflet friction. Crucially, it has been shown that not only the lateral dimension, but also the transverse dimension (solvent thickness) plays a large role in the convergence to macroscopic diffusive dynamics [79]. The theory shows that typical simulation dimensions are much too small for macroscopic dynamical estimates, but allows for extrapolation and comparison with experiment. The theory also effectively implies a variational principle for capturing diffusivities with AA simulations of reasonably-sized systems: if the force field and other settings (integration, ensemble, long-range electrostatics, etc.) are correct, then the simulated diffusivities are expected to underestimate experiment [79]. However, if the desire is to compare to some biological system, it may be useful to normalize to a different finite system size. Experiments have shown that, through an anchored protein picket model, proteins anchored by the membrane cytoskeleton can slow effective lipid diffusion due to both steric hindrance and circumferential slowing, a hydrodynamic friction-like effect [53]. This has been found to be consistent with a characteristic domain size of 150-500 nm [53, 68] (cf. Section 4.1.4).

Experimental diffusivity estimates can vary for a number of reasons. First, diffusivities can be determined from a variety of methods, including fluorescence techniques (e.g. fluorescence recovery after photobleaching, FRAP, and fluorescence correlation spectroscopy, FCS), quasi-elastic neutron scattering, EPR, and NMR, amongst others. The time scale of study can vary significantly with different methods; for example, EPR and NMR measure motions over distances and time scales around > 0.1 nm and ns-ms, much longer neutron scattering at 0.1-10 nm and < 1 ns. While different regimes of diffusion may exist across different lag time scales (most notably, non-classical subdiffusion; see above), fits and com-

parisons from very different time scales can also be prone to significant statistical error. In some of these techniques (e.g. FCS, FRAP), the use of labeled lipids instead of the normal lipids biases the calculations for lipid diffusion. In addition to the increased drag that the label introduces, dynamics can also be impacted by label concentration. Dynamics can be particularly slow for supported lipid bilayers, the physics of which can also be very different from those of the simulation due to enhanced interleaflet friction. Otherwise, results are highly dependent on temperature, hydration content, pH, ionic strength, and experimental setup [63].

4.4.2 Evaluation of major techniques for mechanical property calculation

In general terms, we outline the merits of each umbrella of approaches for calculating mechanical properties (Table 2).

For most of our mechanical property method recommendations here, we focus on the "equilibrium fluctuations" and "stress profile" classes of techniques. However, there are several alternative methods based on "biased/active deformations" techniques. For the area compressibility modulus, the bilayer can be actively stretched via different simulations in the NP_zAT ensemble; the surface tension can be evaluated at each area, and K_A is calculated from the derivative $K_A^{app} = A_p(\partial\gamma/\partial A_p)_T$ [24, 94]. For the bending modulus, there are several techniques [23, 29, 59, 74]. Umbrella sampling has been used to enforce large undulation modes [59], but this study experienced difficulty separating bending contributions from those of stretching. The membrane tether stretching approach in general applies to larger curvature deformations, with radii of curvature down to the membrane thickness, and converges to undulation spectrum results in the small deformation limit. Most recently, the buckling technique was introduced to overcome limitations of membrane tether stretching techniques (including problems with handling explicit solvent), and provides insight into the enthalpy-entropy breakdown of bending contributions and therefore the local temperature dependence on κ [23, 74].

The bending modulus can be experimentally determined from a variety of techniques. This includes fluctuation analysis (e.g. flicker spectroscopy), micropipette aspiration and the low-tension stress-strain relationship, tether stretching (with magnetic fields or optical tweezers [65, 91], x-ray scattering, and neutron spin echo measurements. The bending modulus provides one example where parallel experimental and simulation calculation methods is useful. Simulation results from the undulation and lipid director spectra have been found to agree well with flicker experiments, as both are based on equilibrium fluctuations. These deviate significantly from micropipette aspiration, which is based on "biasing/active deformations" [78].

In the simulation community, as in most of the experimental community, bending is treated as an elastic deformation, so the stress-strain relationship is independent of the rate at which the bending strain is applied. However, there have been various suggestions, with experiment evidence, that membranes may exhibit viscoelasticity with short-time transient responses [11, 91]. Due to the complete lack of simulation infrastructure for assessing membrane viscoelasticity at this time, we defer any recommendations on studying it.

4.4.3 Techniques for leaflet identification & leaflet-dependent properties

Lipid bilayers are made up of two molecularly-thick leaflets in a fluctuating membrane embedded in three dimensions. Identifying which lipid is in which leaflet at any given time is useful for identifying the local bilayer midplane, and therefore a host of bilayer and individual monolayer properties. One method to identify leaflets involves first determining the height function of the entire bilayer in high-resolution gridspace (cf. recommended procedure in Watson et al. J. Chem. Phys., 2011, Appendix C), then going back and sorting into leaflets. However, this can be difficult, especially for membranes with large deformations and complicated morphologies. One analysis package that overcomes these complications is the Fast Analysis Toolbox for Simulations of Lipid Membranes (FATSLiM). FATSLiM is a Python-based package designed to work with GROMACS that, for every simulation frame, can estimate the normal for every lipid via principal component analysis of each lipid and its neighbors. Therefore, it can approximate the membrane surface in a morphology-independent manner (e.g. can be applied to planar and vesicular membranes alike, unlike the packages APL@Voro, MEM-BPLUGIN, and GridMATMD), and can determine membrane leaflets, thickness, and area per lipid. FATSLiM is both efficient and low-memory-consuming (relative to APL@Voro), and documentation is available at: <http://pythonhosted.org/fatslim> [13].

One class of metrics that require leaflet identification are those associated with lipid coordination. The lipid coordination number can be used in phase transitions and coexistence to distinguish between fluid and gel phases, which have markedly different values, and to characterize the segregation of multicomponent bilayers (e.g. does cholesterol segregate with lipid X vs Y). The coordination number is rigorously determined from cumulative integration of the in-plane (2D) radial distribution function to some coordination cutoff distance. Unless interested otherwise, histograms should be binned separately for each leaflet (otherwise, liquid and solid structural signatures are convoluted and lack meaning) by the in-plane 2D radial distance (as opposed to standard RDFs in 3D). (There are several other structural, thermodynamic, and

Table 2. Methods for calculating mechanical properties: a broad cross-comparison

Class of Methods	Advantages	Disadvantages
Equilibrium Fluctuations	<ul style="list-style-type: none"> • Theoretically consistent/rigorous and elegant (admitted directly from Landau-Ginzburg and related approaches, e.g. Canham-Helfrich) • No additional user input required (just run the simulation!) • Well documented and generally the preferred choice in the membrane theory and simulation community 	<ul style="list-style-type: none"> • Can only study and apply results to small deformation limit; not necessarily relevant to strong deformations [23] • Can take a long time for statistics to converge / fluctuations to develop [29] • Low signal-to-noise ratio • Grid analysis in post-processing can be expensive
Local Thermodynamics / Stress Profile	<ul style="list-style-type: none"> • Theoretically motivated • No additional user input required (just run the simulation!) • Consistent with rationale for simulation pressure coupling scheme and Laplace tension (and therefore frame tension), i.e. equation for justifying tension settings in simulation is a specific case (Equation 1) • The only route to calculating certain properties (tension, monolayer spontaneous curvature) • Profile also be used to understand local stresses and molecular driving forces 	<ul style="list-style-type: none"> • Rigorous implementation (although codes exist); expensive voxel analysis in post-processing • Certain moments give you combinations of properties rather than individual ones, and are therefore dependent on other techniques • Slow convergence • Low signal-to-noise ratio (seeking small numbers often from largely-fluctuating ones)
Biased/Active Deformations	<ul style="list-style-type: none"> • Theoretically motivated • Applicable to large-scale deformations (potentially more physically relevant), with a broader range of permissible deformations overall; capable of addressing nonlinear and higher-order effects on mechanical properties at higher deformations [29] • Often more efficient, due to lesser importance of fluctuations and therefore also lesser importance of finite size effects [29] • High signal-to-noise ratio 	<ul style="list-style-type: none"> • Requires additional user input: some biasing scheme and/or nontrivial simulation setup (e.g. tether) for the calculation

dynamic techniques for detecting and characterizing phase transitions and coexistence outlined above, including thickness, area per lipid, the deuterium NMR order parameter, and lipid lateral diffusivity, amongst others.)

4.4.4 Other properties & relevant resources

In order to simulate biologically-relevant transmembrane voltage gradients, there are several possible simulation "tricks." The major problem is the need to use PBCs in simulations, which prevent the setup of a charge gradient. A well-accepted solution is to simulate two bilayers in a single simulation cell, separating the salt baths for the charge gradient. For more details, see the work of Sachs, Crozier, and Woolf [70].

At present, free energy calculations and rare events & importance sampling methods are beyond the current scope

of this document. We recognize that free energy calculations and advanced sampling strategies are extremely promising for studies of pore formation, membrane fusion and other collective phenomena of lipid membranes. For more information, see Smirnova et al. [75] and its references[50, 78].

5 Author Contributions

DJS initiated this document, and set up the complete layout of Version 0.1, ensuring comprehensive and consistent content (including consistency in terminology and nomenclature). DJS compiled and curated the major content of Sections 1, 2, 3, and 4, including the guidance on Model Selection, Pre-Simulation Considerations, Preparation of Initial Configurations, and Property Validation, and also the simulation

graphics, equations, checklist itself, and Tables 1 and 2. JBK and AJS added to the initial document by DJS and expanded based on their areas of expertise.

For a more detailed description of author contributions, see the GitHub issue tracking and changelog at https://github.com/davesmith4398/best_practice_membranes.

6 Other Contributions

The authors thank Alan Grossfield at the University of Rochester Medical Center for general guidance and specific resources incorporated into this document.

For a more detailed description of contributions from the community and others, see the GitHub issue tracking and changelog at https://github.com/davesmith4398/best_practice_membranes.

7 Potentially Conflicting Interests

The authors declare no competing financial interest.

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References

- [1] **Allen WJ**, Lemkul JA, Bevan DR. GridMAT-MD: A grid-based membrane analysis tool for use with molecular dynamics. *Journal of Computational Chemistry*. 2009 sep; 30(12):1952–1958. <http://doi.wiley.com/10.1002/jcc.21172>, doi: 10.1002/jcc.21172.
- [2] **Arnarez C**, Uusitalo JJ, Masman MF, Ingólfsson HI, de Jong DH, Melo MN, Periole X, de Vries AH, Marrink SJ. Dry Martini, a Coarse-Grained Force Field for Lipid Membrane Simulations with Implicit Solvent. *Journal of Chemical Theory and Computation*. 2015 jan; 11(1):260–275. <http://dx.doi.org/10.1021/ct500477k>, doi: 10.1021/ct500477k. <http://pubs.acs.org/doi/full/10.1021/ct500477k>, doi: 10.1021/ct500477k. <http://pubs.acs.org/doi/pdfplus/10.1021/ct500477k>, doi: 10.1021/ct500477k.
- [3] **Ben-Shaul A**. Molecular Theory of Chain Packing, Elasticity and Lipid-Protein Interaction in Lipid Bilayers. In: *Handbook of Biological Physics*, vol. 1; 1995.p. 359–401. <http://linkinghub.elsevier.com/retrieve/pii/S1383812106800242>, doi: 10.1016/S1383-8121(06)80024-2.
- [4] **Berkowitz ML**. Detailed molecular dynamics simulations of model biological membranes containing cholesterol. *Biochimica et Biophysica Acta (BBA) - Biomembranes*. 2009 jan; 1788(1):86–96. <http://dx.doi.org/10.1016/j.bbamem.2008.09.009><http://linkinghub.elsevier.com/retrieve/pii/S0005273608002927>, doi: 10.1016/j.bbamem.2008.09.009.
- [5] **Brannigan G**, Brown FLH. A Consistent Model for Thermal Fluctuations and Protein-Induced Deformations in Lipid Bilayers. *Biophysical Journal*. 2006 mar; 90(5):1501–1520. <http://dx.doi.org/10.1529/biophysj.105.075838><http://linkinghub.elsevier.com/retrieve/pii/S0006349506723414>, doi: 10.1529/biophysj.105.075838.
- [6] **Brannigan G**, Lin LCL, Brown FLH. Implicit solvent simulation models for biomembranes. *European Biophysics Journal*. 2006 jan; 35(2):104–124. <http://link.springer.com/10.1007/s00249-005-0013-y>, doi: 10.1007/s00249-005-0013-y.
- [7] **Brannigan G**, Phillips PF, Brown FLH. Flexible lipid bilayers in implicit solvent. *Physical Review E*. 2005 jul; 72(1):011915. <http://link.aps.org/doi/10.1103/PhysRevE.72.011915><http://link.aps.org/doi/10.1103/PhysRevE.72.011915>, doi: 10.1103/PhysRevE.72.011915.
- [8] **Braun AR**, Brandt EG, Edholm O, Nagle JF, Sachs JN. Determination of Electron Density Profiles and Area from Simulations of Undulating Membranes. *Biophysical Journal*. 2011 may; 100(9):2112–2120. <http://dx.doi.org/10.1016/j.bpj.2011.03.009><http://linkinghub.elsevier.com/retrieve/pii/S0006349511003195>, doi: 10.1016/j.bpj.2011.03.009.
- [9] **Brochard-Wyart F**, de Gennes PG, Sandre O. Transient pores in stretched vesicles: role of leak-out. *Physica A: Statistical Mechanics and its Applications*. 2000 apr; 278(1-2):32–51. <http://linkinghub.elsevier.com/retrieve/pii/S0378437199005592>, doi: 10.1016/S0378-4371(99)00559-2.
- [10] **Brooks BR**, Brooks CL, Mackerell AD, Nilsson L, Petrella RJ, Roux B, Won Y, Archontis G, Bartels C, Boresch S, Caflisch A, Caves L, Cui Q, Dinner AR, Feig M, Fischer S, Gao J, Hodoscek M, Im W, Kuczera K, et al. CHARMM: the biomolecular simulation program. *Journal of computational chemistry*. 2009 jul; 30(10):1545–614. <http://www.ncbi.nlm.nih.gov/pubmed/19444816><http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC2810661>, doi: 10.1002/jcc.21287.
- [11] **Brown FLH**. Continuum simulations of biomembrane dynamics and the importance of hydrodynamic effects. *Quarterly Reviews of Biophysics*. 2011 nov; 44(04):391–432. <http://www.ncbi.nlm.nih.gov/pubmed/21729348>http://www.journals.cambridge.org/abstract/_S0033583511000047, doi: 10.1017/S0033583511000047.
- [12] **Brown FLH**. Elastic Modeling of Biomembranes and Lipid Bilayers. *Annual Review of Physical Chemistry*. 2008 may; 59(1):685–712. <http://www.ncbi.nlm.nih.gov/pubmed/18173377><http://www.annualreviews.org/doi/10.1146/annurev.physchem.59.032607.093550>, doi: 10.1146/annurev.physchem.59.032607.093550.
- [13] **Buchoux S**. FATSliM: a fast and robust software to analyze MD simulations of membranes. *Bioinformatics*. 2017 jan; 33(1):133–134. <https://academic.oup.com/bioinformatics/article-lookup/>

- [doi/10.1093/bioinformatics/btw563](https://doi.org/10.1093/bioinformatics/btw563), doi: 10.1093/bioinformatics/btw563.
- [14] **Camley Ba**, Brown FLH. Diffusion of complex objects embedded in free and supported lipid bilayer membranes: role of shape anisotropy and leaflet structure. *Soft Matter*. 2013; 9(19):4767. <http://xlink.rsc.org/?DOI=c3sm00073g>, doi: 10.1039/c3sm00073g.
 - [15] **Canham PB**. The minimum energy of bending as a possible explanation of the biconcave shape of the human red blood cell. *Journal of Theoretical Biology*. 1970 jan; 26(1):61–81. <http://linkinghub.elsevier.com/retrieve/pii/S0022519370800327>, doi: 10.1016/S0022-5193(70)80032-7.
 - [16] **Case DA**, Brozell SR, Cerutti DS, Cheatham, III TE, Cruzeiro VWD, Darden TA, Duke RE, Ghoreishi D, Gohlke H, Goetz AW, Greene D, Harris R, Homeyer N, Izadi S, Kovalenko A, Lee TS, LeGrand S, Li P, Lin C, Liu J, et al., AMBER 2018. University of California, San Francisco; 2018.
 - [17] **Castro-Román F**, Benz RW, White SH, Tobias DJ. Investigation of Finite System-Size Effects in Molecular Dynamics Simulations of Lipid Bilayers. *The Journal of Physical Chemistry B*. 2006 nov; 110(47):24157–24164. <http://pubs.acs.org/doi/abs/10.1021/jp064746g>, doi: 10.1021/jp064746g.
 - [18] **Cheng X**, Jo S, Lee HS, Klauda JB, Im W. CHARMM-GUI Micelle Builder for Pure/Mixed Micelle and Protein/Micelle Complex Systems. *Journal of Chemical Information and Modeling*. 2013 aug; 53(8):2171–2180. <http://www.ncbi.nlm.nih.gov/pubmed/23865552><http://pubs.acs.org/doi/10.1021/ci4002684>, doi: 10.1021/ci4002684.
 - [19] **Cooke IR**, Deserno M. Solvent-free model for self-assembling fluid bilayer membranes: Stabilization of the fluid phase based on broad attractive tail potentials. *The Journal of Chemical Physics*. 2005 dec; 123(22):224710. <http://aip.scitation.org/doi/10.1063/1.2135785>, doi: 10.1063/1.2135785.
 - [20] **Deserno M**. Mesoscopic Membrane Physics: Concepts, Simulations, and Selected Applications. *Macromolecular Rapid Communications*. 2009 may; 30(9-10):752–771. <http://doi.wiley.com/10.1002/marc.200900090>, doi: 10.1002/marc.200900090.
 - [21] **Diamant H**. Model-free thermodynamics of fluid vesicles. *Physical Review E*. 2011 dec; 84(6):061123. <http://link.aps.org/doi/10.1103/PhysRevE.84.061123><https://link.aps.org/doi/10.1103/PhysRevE.84.061123>, doi: 10.1103/PhysRevE.84.061123.
 - [22] **Dickson CJ**, Madej BD, Skjevik ÅA, Betz RM, Teigen K, Gould IR, Walker RC. Lipid14: The Amber Lipid Force Field. *Journal of Chemical Theory and Computation*. 2014 feb; 10(2):865–879. <http://pubs.acs.org/doi/10.1021/ct4010307>, doi: 10.1021/ct4010307.
 - [23] **Diggins P**, Deserno M. Determining the bending modulus of a lipid membrane by simulating buckling. *The Journal of Chemical Physics*. 2013 jun; 138(21):214110. <http://aip.scitation.org/doi/10.1063/1.4808077>, doi: 10.1063/1.4808077.
 - [24] **Feller SE**, Zhang Y, Pastor RW. Computer simulation of liquid/liquid interfaces. II. Surface tension-area dependence of a bilayer and monolayer. *The Journal of Chemical Physics*. 1995 dec; 103(23):10267–10276. <http://aip.scitation.org/doi/10.1063/1.469928>, doi: 10.1063/1.469928.
 - [25] **Goetz R**, Gompper G, Lipowsky R. Mobility and Elasticity of Self-Assembled Membranes. *Physical Review Letters*. 1999 jan; 82(1):221–224. <https://link.aps.org/doi/10.1103/PhysRevLett.82.221>, doi: 10.1103/PhysRevLett.82.221.
 - [26] **Goga N**, Rzepiela AJ, de Vries AH, Marrink SJ, Berendsen HJC. Efficient Algorithms for Langevin and DPD Dynamics. *Journal of Chemical Theory and Computation*. 2012 oct; 8(10):3637–3649. <http://pubs.acs.org/doi/10.1021/ct3000876>, doi: 10.1021/ct3000876.
 - [27] **Goychuk I**, Hänggi P. Fractional diffusion modeling of ion channel gating. *Physical Review E*. 2004 nov; 70(5):051915. <https://link.aps.org/doi/10.1103/PhysRevE.70.051915>, doi: 10.1103/PhysRevE.70.051915.
 - [28] **Hamm M**, Kozlov M. Elastic energy of tilt and bending of fluid membranes. *The European Physical Journal E*. 2000 dec; 3(4):323–335. <http://link.springer.com/article/10.1007/s101890070003><http://link.springer.com/10.1007/s101890070003><http://link.springer.com/10.1007/s101890070003>, doi: 10.1007/s101890070003.
 - [29] **Harmandaris VA**, Deserno M. A novel method for measuring the bending rigidity of model lipid membranes by simulating tethers. *The Journal of Chemical Physics*. 2006 nov; 125(20):204905. <http://aip.scitation.org/doi/10.1063/1.2372761>, doi: 10.1063/1.2372761.
 - [30] **Helfrich W**. Elastic properties of lipid bilayers: theory and possible experiments. *Zeitschrift für Naturforschung Teil C: Biochemie, Biophysik, Biologie, Virologie*. 1981 mar; 28(11):693–703. <http://doi.wiley.com/10.1002/mus.880040211><http://www.ncbi.nlm.nih.gov/pubmed/4273690>, doi: 10.1002/mus.880040211.
 - [31] **Hu M**, Briguglio JJ, Deserno M. Determining the Gaussian Curvature Modulus of Lipid Membranes in Simulations. *Biophysical Journal*. 2012 mar; 102(6):1403–1410. <http://dx.doi.org/10.1016/j.bpj.2012.02.013><http://linkinghub.elsevier.com/retrieve/pii/S0006349512002147>, doi: 10.1016/j.bpj.2012.02.013.
 - [32] **Hu M**, de Jong DH, Marrink SJ, Deserno M. Gaussian curvature elasticity determined from global shape transformations and local stress distributions: a comparative study using the MARTINI model. *Faraday Discuss*. 2013; 161:365–382. <http://xlink.rsc.org/?DOI=C2FD20087B>, doi: 10.1039/C2FD20087B.
 - [33] **Ingólfsson HI**, Melo MN, van Eerden FJ, Arnarez C, Lopez CA, Wassenaar TA, Periole X, de Vries AH, Tieleman DP, Marrink SJ. Lipid Organization of the Plasma Membrane. *Journal of the American Chemical Society*. 2014 oct; 136(41):14554–14559. <http://pubs.acs.org/doi/10.1021/ja507832e>, doi: 10.1021/ja507832e.
 - [34] **Israelachvili JN**. *Intermolecular and Surface Forces*. Waltham, MA: Elsevier; 2011.
 - [35] **Jämbeck JPM**, Lyubartsev AP. Derivation and Systematic Validation of a Refined All-Atom Force Field for Phosphatidylcholine Lipids. *The Journal of Physical Chemistry B*. 2012 mar;

- 116(10):3164–3179. <http://pubs.acs.org/doi/10.1021/jp212503e>, doi: 10.1021/jp212503e.
- [36] **Jo S**, Kim T, Iyer VG, Im W. CHARMM-GUI: A web-based graphical user interface for CHARMM. *Journal of Computational Chemistry*. 2008 aug; 29(11):1859–1865. <http://www.ncbi.nlm.nih.gov/pubmed/20928852><http://doi.wiley.com/10.1002/jcc.21576><http://doi.wiley.com/10.1002/jcc.20945>, doi: 10.1002/jcc.20945.
- [37] **Jo S**, Lim JB, Klauda JB, Im W. CHARMM-GUI Membrane Builder for Mixed Bilayers and Its Application to Yeast Membranes. *Biophysical Journal*. 2009 jul; 97(1):50–58. <http://dx.doi.org/10.1016/j.bpj.2009.04.013><http://www.ncbi.nlm.nih.gov/pubmed/19580743><http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC2711372><http://linkinghub.elsevier.com/retrieve/pii/S0006349509007917>, doi: 10.1016/j.bpj.2009.04.013.
- [38] **Khakbazi P**, Klauda JB. Probing the importance of lipid diversity in cell membranes via molecular simulation. *Chemistry and Physics of Lipids*. 2015 nov; 192:12–22. <http://dx.doi.org/10.1016/j.chemphyslip.2015.08.003><http://linkinghub.elsevier.com/retrieve/pii/S0009308415300347>, doi: 10.1016/j.chemphyslip.2015.08.003.
- [39] **Kirkwood JG**, Buff FP. The Statistical Mechanical Theory of Surface Tension. *The Journal of Chemical Physics*. 1949 mar; 17(3):338–343. <http://aip.scitation.org/doi/10.1063/1.1747248>, doi: 10.1063/1.1747248.
- [40] **Klauda JB**, Brooks BR, Pastor RW. Dynamical motions of lipids and a finite size effect in simulations of bilayers. *The Journal of Chemical Physics*. 2006 oct; 125(14):144710. <http://aip.scitation.org/doi/10.1063/1.2354486>, doi: 10.1063/1.2354486.
- [41] **Klauda JB**, Kučerka N, Brooks BR, Pastor RW, Nagle JF. Simulation-Based Methods for Interpreting X-Ray Data from Lipid Bilayers. *Biophysical Journal*. 2006 apr; 90(8):2796–2807. <http://linkinghub.elsevier.com/retrieve/pii/S0006349506724614>, doi: 10.1529/biophysj.105.075697.
- [42] **Klauda JB**, Venable RM, Freites JA, O'Connor JW, Tobias DJ, Mondragon-Ramirez C, Vorobyov I, MacKerell AD, Pastor RW. Update of the CHARMM All-Atom Additive Force Field for Lipids: Validation on Six Lipid Types. *The Journal of Physical Chemistry B*. 2010 jun; 114(23):7830–7843. <http://pubs.acs.org/doi/abs/10.1021/jp101759q>, doi: 10.1021/jp101759q.
- [43] **König S**, Sackmann E. Molecular and collective dynamics of lipid bilayers. *Current Opinion in Colloid & Interface Science*. 1996 feb; 1(1):78–82. <http://linkinghub.elsevier.com/retrieve/pii/S1359029496800471>, doi: 10.1016/S1359-0294(96)80047-1.
- [44] **Kou SC**, Xie XS. Generalized Langevin Equation with Fractional Gaussian Noise: Subdiffusion within a Single Protein Molecule. *Physical Review Letters*. 2004 oct; 93(18):180603. <https://link.aps.org/doi/10.1103/PhysRevLett.93.180603>, doi: 10.1103/PhysRevLett.93.180603.
- [45] **Leftin A**, Brown MF. An NMR database for simulations of membrane dynamics. *Biochimica et Biophysica Acta (BBA) - Biomembranes*. 2011 mar; 1808(3):818–839. <http://dx.doi.org/10.1016/j.bbamem.2010.11.027><http://linkinghub.elsevier.com/retrieve/pii/S0005273610004128>, doi: 10.1016/j.bbamem.2010.11.027.
- [46] **Lindahl E**, Edholm O. Mesoscopic Undulations and Thickness Fluctuations in Lipid Bilayers from Molecular Dynamics Simulations. *Biophysical Journal*. 2000 jul; 79(1):426–433. [http://dx.doi.org/10.1016/S0006-3495\(00\)76304-1](http://dx.doi.org/10.1016/S0006-3495(00)76304-1)<http://linkinghub.elsevier.com/retrieve/pii/S0006349500763041>, doi: 10.1016/S0006-3495(00)76304-1.
- [47] **Lukat G**, Krüger J, Sommer B. APL@Voro: A Voronoi-Based Membrane Analysis Tool for GROMACS Trajectories. *Journal of Chemical Information and Modeling*. 2013 nov; 53(11):2908–2925. <http://pubs.acs.org/doi/10.1021/ci400172g>, doi: 10.1021/ci400172g.
- [48] **Marrink SJ**, Risselada HJ, Yefimov S, Tieleman DP, de Vries AH. The MARTINI Force Field: Coarse Grained Model for Biomolecular Simulations. *The Journal of Physical Chemistry B*. 2007 jul; 111(27):7812–7824. <http://www.ncbi.nlm.nih.gov/pubmed/17569554><http://pubs.acs.org/doi/abs/10.1021/jp071097f>, doi: 10.1021/jp071097f.
- [49] **Marrink SJ**, Tieleman DP. Perspective on the Martini model. *Chemical Society Reviews*. 2013 aug; 42(16):6801. <http://www.ncbi.nlm.nih.gov/pubmed/23708257><http://xlink.rsc.org/?DOI=c3cs60093a>, doi: 10.1039/c3cs60093a.
- [50] **Marrink SJ**, de Vries AH, Mark AE. Coarse Grained Model for Semiquantitative Lipid Simulations. *The Journal of Physical Chemistry B*. 2004 jan; 108(2):750–760. <http://pubs.acs.org/doi/abs/10.1021/jp036508g>, doi: 10.1021/jp036508g.
- [51] **May ER**, Narang A, Kopelevich DI. Role of molecular tilt in thermal fluctuations of lipid membranes. *Physical Review E*. 2007 aug; 76(2):021913. <http://link.aps.org/doi/10.1103/PhysRevE.76.021913><https://link.aps.org/doi/10.1103/PhysRevE.76.021913>, doi: 10.1103/PhysRevE.76.021913.
- [52] **Monje-Galvan V**, Klauda JB. Modeling Yeast Organelle Membranes and How Lipid Diversity Influences Bilayer Properties. *Biochemistry*. 2015 nov; 54(45):6852–6861. <http://pubs.acs.org/doi/10.1021/acs.biochem.5b00718>, doi: 10.1021/acs.biochem.5b00718.
- [53] **Morone N**, Fujiwara T, Murase K, Kasai RS, Ike H, Yuasa S, Usukura J, Kusumi A. Three-dimensional reconstruction of the membrane skeleton at the plasma membrane interface by electron tomography. *The Journal of Cell Biology*. 2006 sep; 174(6):851–862. <http://www.jcb.org/lookup/doi/10.1083/jcb.200606007>, doi: 10.1083/jcb.200606007.
- [54] **Moroz JD**, Nelson P. Dynamically stabilized pores in bilayer membranes. *Biophysical Journal*. 1997 may; 72(5):2211–2216. <http://arxiv.org/abs/cond-mat/9610072>[http://dx.doi.org/10.1016/S0006-3495\(97\)78864-7](http://dx.doi.org/10.1016/S0006-3495(97)78864-7)<http://linkinghub.elsevier.com/retrieve/pii/S0006349597788647>, doi: 10.1016/S0006-3495(97)78864-7.
- [55] **Munguira I**, Casuso I, Takahashi H, Rico F, Miyagi A, Chami M, Scheuring S. Glasslike Membrane Protein Diffusion in a Crowded Membrane. *ACS Nano*. 2016 feb; 10(2):2584–2590. <http://pubs.acs.org/doi/10.1021/acsnano.5b07595>, doi: 10.1021/acsnano.5b07595.
- [56] **Nagle JF**. Long tail kinetics in biophysics? *Biophysical Journal*. 1992 aug; 63(2):366–370. <http://linkinghub.elsevier.com/retrieve/pii/S0006349592816028>, doi: 10.1016/S0006-3495(92)81602-8.

- [57] **Nagle JF**. Area/lipid of bilayers from NMR. *Biophysical Journal*. 1993 may; 64(5):1476–1481. [http://dx.doi.org/10.1016/S0006-3495\(93\)81514-5](http://dx.doi.org/10.1016/S0006-3495(93)81514-5)<http://linkinghub.elsevier.com/retrieve/pii/S0006349593815145>, doi: 10.1016/S0006-3495(93)81514-5.
- [58] **Olila OHS**, Risselada HJ, Louhivuori M, Lindahl E, Vattulainen I, Marrink SJ. 3D Pressure Field in Lipid Membranes and Membrane-Protein Complexes. *Physical Review Letters*. 2009 feb; 102(7):078101. <https://link.aps.org/doi/10.1103/PhysRevLett.102.078101>, doi: 10.1103/PhysRevLett.102.078101.
- [59] **den Otter WK**, Briels WJ. The bending rigidity of an amphiphilic bilayer from equilibrium and nonequilibrium molecular dynamics. *The Journal of Chemical Physics*. 2003 mar; 118(10):4712–4720. <http://aip.scitation.org/doi/10.1063/1.1543941>, doi: 10.1063/1.1543941.
- [60] **Park S**, Beaven AH, Klauda JB, Im W. How Tolerant are Membrane Simulations with Mismatch in Area per Lipid between Leaflets? *Journal of Chemical Theory and Computation*. 2015 jul; 11(7):3466–3477. <http://pubs.acs.org/doi/10.1021/acs.jctc.5b00232>, doi: 10.1021/acs.jctc.5b00232.
- [61] **Peliti L**, Leibler S. Effects of Thermal Fluctuations on Systems with Small Surface Tension. *Physical Review Letters*. 1985 apr; 54(15):1690–1693. <https://link.aps.org/doi/10.1103/PhysRevLett.54.1690>, doi: 10.1103/PhysRevLett.54.1690.
- [62] **Phillips JC**, Braun R, Wang W, Gumbart J, Tajkhorshid E, Villa E, Chipot C, Skeel RD, Kalé L, Schulten K. Scalable molecular dynamics with NAMD. *Journal of Computational Chemistry*. 2005 dec; 26(16):1781–1802. <http://doi.wiley.com/10.1002/jcc.20289>, doi: 10.1002/jcc.20289.
- [63] **Poger D**, Caron B, Mark AE. Validating lipid force fields against experimental data: Progress, challenges and perspectives. *Biochimica et Biophysica Acta (BBA) - Biomembranes*. 2016 jul; 1858(7):1556–1565. <http://dx.doi.org/10.1016/j.bbamem.2016.01.029><http://linkinghub.elsevier.com/retrieve/pii/S0005273616300281>, doi: 10.1016/j.bbamem.2016.01.029.
- [64] **Poger D**, Van Gunsteren WF, Mark AE. A new force field for simulating phosphatidylcholine bilayers. *Journal of Computational Chemistry*. 2010 apr; 31(6):1117–1125. <http://doi.wiley.com/10.1002/jcc.21396>, doi: 10.1002/jcc.21396.
- [65] **Pontes B**, Monzo P, Gauthier NC. Membrane tension: A challenging but universal physical parameter in cell biology. *Seminars in Cell & Developmental Biology*. 2017 nov; 71:30–41. <https://doi.org/10.1016/j.semcdb.2017.08.030><http://linkinghub.elsevier.com/retrieve/pii/S1084952116302932>, doi: 10.1016/j.semcdb.2017.08.030.
- [66] **Ramakrishnan N**, Sunil Kumar PB, Radhakrishnan R. Mesoscale computational studies of membrane bilayer remodeling by curvature-inducing proteins. *Physics Reports*. 2014 oct; 543(1):1–60. <http://dx.doi.org/10.1016/j.physrep.2014.05.001><http://linkinghub.elsevier.com/retrieve/pii/S0370157314002038>, doi: 10.1016/j.physrep.2014.05.001.
- [67] **Rawicz W**, Olbrich KC, McIntosh T, Needham D, Evans E. Effect of Chain Length and Unsaturation on Elasticity of Lipid Bilayers. *Biophysical Journal*. 2000 jul; 79(1):328–339. <http://linkinghub.elsevier.com/retrieve/pii/S0006349500762953>, doi: 10.1016/S0006-3495(00)76295-3.
- [68] **Ritchie K**, Iino R, Fujiwara T, Murase K, Kusumi A. The fence and picket structure of the plasma membrane of live cells as revealed by single molecule techniques (Review). *Molecular Membrane Biology*. 2003 jan; 20(1):13–18. <http://taylorandfrancis.metapress.com/openurl.asp?genre=article&id=doi:10.1080/09687680307076>, doi: 10.1080/09687680307076.
- [69] **Rodgers JM**, Sørensen J, de Meyer FJM, Schiøtt B, Smit B. Understanding the Phase Behavior of Coarse-Grained Model Lipid Bilayers through Computational Calorimetry. *The Journal of Physical Chemistry B*. 2012 feb; 116(5):1551–1569. <http://pubs.acs.org/doi/10.1021/jp207837v>, doi: 10.1021/jp207837v.
- [70] **Sachs JN**, Crozier PS, Woolf TB. Atomistic simulations of biologically realistic transmembrane potential gradients. *The Journal of Chemical Physics*. 2004; 121(22):10847. <http://scitation.aip.org/content/aip/journal/jcp/121/22/10.1063/1.1826056>, doi: 10.1063/1.1826056.
- [71] **Safran SA**. *Statistical Thermodynamics of Surfaces, Interfaces, and Membranes*. Boca Raton, FL: Westview Press; 2018.
- [72] **Sapp K**, Shlomovitz R, Maibaum L. Seeing the Forest in Lieu of the Trees. In: *Annual Reports in Computational Chemistry*, vol. 10; 2014.p. 47–76. <http://linkinghub.elsevier.com/retrieve/pii/B9780444633781000033>, doi: 10.1016/B978-0-444-63378-1.00003-3.
- [73] **Saxton MJ**. Wanted: A Positive Control for Anomalous Subdiffusion. *Biophysical Journal*. 2012 dec; 103(12):2411–2422. <http://dx.doi.org/10.1016/j.bpj.2012.10.038><http://linkinghub.elsevier.com/retrieve/pii/S0006349512011927>, doi: 10.1016/j.bpj.2012.10.038.
- [74] **Shiba H**, Noguchi H. Estimation of the bending rigidity and spontaneous curvature of fluid membranes in simulations. *Physical Review E*. 2011 sep; 84(3):031926. <https://link.aps.org/doi/10.1103/PhysRevE.84.031926>, doi: 10.1103/PhysRevE.84.031926.
- [75] **Smirnova YG**, Fuhrmans M, Barragan Vidal IA, Müller M. Free-energy calculation methods for collective phenomena in membranes. *Journal of Physics D: Applied Physics*. 2015 sep; 48(34):343001. <http://stacks.iop.org/0022-3727/48/i=34/a=343001?key=crossref.0e98c1485ea1fe32cef3171b29196341>, doi: 10.1088/0022-3727/48/34/343001.
- [76] **Tieleman DP**, Marrink SJ, Berendsen HJC. A computer perspective of membranes: molecular dynamics studies of lipid bilayer systems. *Biochimica et Biophysica Acta (BBA) - Reviews on Biomembranes*. 1997 nov; 1331(3):235–270. <http://linkinghub.elsevier.com/retrieve/pii/S0304415797000087>, doi: 10.1016/S0304-4157(97)00008-7.
- [77] **Tolpekina TV**, den Otter WK, Briels WJ. Simulations of stable pores in membranes: System size dependence and line tension. *The Journal of Chemical Physics*. 2004; 121(16):8014. <http://scitation.aip.org/content/aip/journal/jcp/121/16/10.1063/1.1796254>, doi: 10.1063/1.1796254.

- [78] **Venable RM**, Brown FLH, Pastor RW. Mechanical properties of lipid bilayers from molecular dynamics simulation. *Chemistry and Physics of Lipids*. 2015 nov; 192:60–74. <http://dx.doi.org/10.1016/j.chemphyslip.2015.07.014><http://linkinghub.elsevier.com/retrieve/pii/S0009308415300190>, doi: 10.1016/j.chemphyslip.2015.07.014.
- [79] **Venable RM**, Ingólfsson HI, Lerner MG, Perrin BS, Camley BA, Marrink SJ, Brown FLH, Pastor RW. Lipid and Peptide Diffusion in Bilayers: The Saffman–Delbrück Model and Periodic Boundary Conditions. *The Journal of Physical Chemistry B*. 2017 apr; 121(15):3443–3457. <http://pubs.acs.org/doi/10.1021/acs.jpcb.6b09111>, doi: 10.1021/acs.jpcb.6b09111.
- [80] **Vermeer LS**, de Groot BL, Réat V, Milon A, Czaplicki J. Acyl chain order parameter profiles in phospholipid bilayers: computation from molecular dynamics simulations and comparison with 2H NMR experiments. *European Biophysics Journal*. 2007 nov; 36(8):919–931. <http://link.springer.com/10.1007/s00249-007-0192-9>, doi: 10.1007/s00249-007-0192-9.
- [81] **Waheed Q**, Edholm O. Undulation Contributions to the Area Compressibility in Lipid Bilayer Simulations. *Biophysical Journal*. 2009 nov; 97(10):2754–2760. <http://dx.doi.org/10.1016/j.bpj.2009.08.048><http://linkinghub.elsevier.com/retrieve/pii/S0006349509014386>, doi: 10.1016/j.bpj.2009.08.048.
- [82] **Wang Y**, Sigurdsson JK, Brandt E, Atzberger PJ. Dynamic implicit-solvent coarse-grained models of lipid bilayer membranes: Fluctuating hydrodynamics thermostat. *Physical Review E*. 2013 aug; 88(2):023301. <https://link.aps.org/doi/10.1103/PhysRevE.88.023301>, doi: 10.1103/PhysRevE.88.023301.
- [83] **Wassenaar TA**, Ingólfsson HI, Böckmann RA, Tieleman DP, Marrink SJ. Computational Lipidomics with insane : A Versatile Tool for Generating Custom Membranes for Molecular Simulations. *Journal of Chemical Theory and Computation*. 2015 may; 11(5):2144–2155. <http://pubs.acs.org/doi/10.1021/acs.jctc.5b00209>, doi: 10.1021/acs.jctc.5b00209.
- [84] **Watson MC**, Brandt EG, Welch PM, Brown FLH. Determining Biomembrane Bending Rigidities from Simulations of Modest Size. *Physical Review Letters*. 2012 jul; 109(2):028102. <http://link.aps.org/doi/10.1103/PhysRevLett.109.028102><https://link.aps.org/doi/10.1103/PhysRevLett.109.028102>, doi: 10.1103/PhysRevLett.109.028102.
- [85] **Watson MC**, Brown FLH. Interpreting Membrane Scattering Experiments at the Mesoscale: The Contribution of Dissipation within the Bilayer. *Biophysical Journal*. 2010 mar; 98(6):L9–L11. <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=2849052&tool=pmcentrez&rendertype=abstract><http://linkinghub.elsevier.com/retrieve/pii/S000634950901755X>, doi: 10.1016/j.bpj.2009.11.026.
- [86] **Watson MC**, Morriss-Andrews A, Welch PM, Brown FLH. Thermal fluctuations in shape, thickness, and molecular orientation in lipid bilayers. II. Finite surface tensions. *The Journal of Chemical Physics*. 2013 aug; 139(8):084706. <http://www.ncbi.nlm.nih.gov/pubmed/24007028><http://aip.scitation.org/doi/10.1063/1.4818530>, doi: 10.1063/1.4818530.
- [87] **Watson MC**, Penev ES, Welch PM, Brown FLH. Thermal fluctuations in shape, thickness, and molecular orientation in lipid bilayers. *The Journal of Chemical Physics*. 2011 dec; 135(24):244701. <http://scitation.aip.org/content/aip/journal/jcp/135/24/10.1063/1.3660673><http://aip.scitation.org/doi/10.1063/1.3660673>, doi: 10.1063/1.3660673.
- [88] **Weiss M**, Elsner M, Kartberg F, Nilsson T. Anomalous Subdiffusion Is a Measure for Cytoplasmic Crowding in Living Cells. *Biophysical Journal*. 2004 nov; 87(5):3518–3524. <http://linkinghub.elsevier.com/retrieve/pii/S0006349504738163>, doi: 10.1529/biophysj.104.044263.
- [89] **Wohlert J**, den Otter WK, Edholm O, Briels WJ. Free energy of a trans-membrane pore calculated from atomistic molecular dynamics simulations. *The Journal of Chemical Physics*. 2006 apr; 124(15):154905. <http://aip.scitation.org/doi/10.1063/1.2171965>, doi: 10.1063/1.2171965.
- [90] **Wu EL**, Cheng X, Jo S, Rui H, Song KC, Dávila-Contreras EM, Qi Y, Lee J, Monje-Galvan V, Venable RM, Klauda JB, Im W. CHARMM-GUI Membrane Builder toward realistic biological membrane simulations. *Journal of computational chemistry*. 2014 oct; 35(27):1997–2004. <http://www.ncbi.nlm.nih.gov/pubmed/25130509><http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC4165794>, doi: 10.1002/jcc.23702.
- [91] **Wu SH**, Sankhagowit S, Biswas R, Wu S, Povinelli ML, Malmstadt N. Viscoelastic deformation of lipid bilayer vesicles. *Soft Matter*. 2015; 11(37):7385–7391. <http://xlink.rsc.org/?DOI=C5SM01565K>, doi: 10.1039/C5SM01565K.
- [92] **Yeh IC**, Hummer G. System-Size Dependence of Diffusion Coefficients and Viscosities from Molecular Dynamics Simulations with Periodic Boundary Conditions. *The Journal of Physical Chemistry B*. 2004 oct; 108(40):15873–15879. <http://pubs.acs.org/doi/abs/10.1021/jp0477147>, doi: 10.1021/jp0477147.
- [93] **Zgorski A**, Lyman E. Toward Hydrodynamics with Solvent Free Lipid Models: STRD Martini. *Biophysical Journal*. 2016 dec; 111(12):2689–2697. <http://linkinghub.elsevier.com/retrieve/pii/S0006349516310384>, doi: 10.1016/j.bpj.2016.11.010.
- [94] **Zhang Y**, Feller SE, Brooks BR, Pastor RW. Computer simulation of liquid/liquid interfaces. I. Theory and application to octane/water. *The Journal of Chemical Physics*. 1995 dec; 103(23):10252–10266. <http://aip.scitation.org/doi/10.1063/1.469927>, doi: 10.1063/1.469927.
- [95] **Zhelev DV**, Needham D. Tension-stabilized pores in giant vesicles: determination of pore size and pore line tension. *Biochimica et Biophysica Acta (BBA) - Biomembranes*. 1993 apr; 1147(1):89–104. <http://linkinghub.elsevier.com/retrieve/pii/000527369390319U>, doi: 10.1016/0005-2736(93)90319-U.