Analysis of three nuclear families with a single pediatric-cancer case using linked-read whole genome sequencing

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## **VCF** Processing

For each step of VCF filtering, we used a Docker image that encapsulated the software tools as well as Python scripts (available on github) to process the data. These Python scripts either used custom code or a combination of custom code and existing software. Some of the scripts used during the VCF filtering process were adapted from CompoundHetVIP.

The files used as part of our analyses are available at (to be updated).

A more detailed description of each step is available in the manuscript.

## Filter out poor quality variants for each sample and each file type

Initial files being filtered are those that were output by *Long Ranger*. *Long Ranger* output a phased SNP VCF, a phased mid-scale deletion VCF, and a phased large-scale SV VCF for each sample.

```
# Phased SNP files
docker run -d -v /Data:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/keep_passed_variants.py \
  PedFam/*/outs/phased variants.vcf.gz \
  PedFam_analysis/phased_files/ \
  PedFam analysis/passed variants summary.tsv \
  > PedFam_analysis/keep_passed_variants.out
# Phased dels files
docker run -d -v /Data:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/keep_passed_variants.py \
  PedFam/*/outs/dels.vcf.gz \
  PedFam_analysis/dels_files/ \
  PedFam_analysis/passed_dels_summary.tsv \
  > PedFam_analysis/keep_passed_dels.out
# Phased sus files
docker run -d -v /Data:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/keep_passed_variants.py \
  PedFam/*/outs/large_svs.vcf.gz \
  PedFam analysis/svs files/ \
  PedFam_analysis/passed_svs_summary.tsv \
  > PedFam_analysis/keep_passed_svs.out
```

#### Combine sample files

```
# Filtered SNP files
docker run -d -v /Data:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/concat_merge_phased_vcf.py \
  PedFam_analysis/phased_files/ \
  PedFam_analysis/combined.vcf.gz \
  --output_fam_file PedFam_analysis/combined.fam \
  --concat files n \
  --merge_files all \
  > PedFam_analysis/concat_merge_phased_vcf.out
# Filtered dels files
docker run -d -v /Data:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/concat_merge_phased_vcf.py \
  PedFam_analysis/dels_files/ \
  PedFam_analysis/dels_combined.vcf.gz \
  --output_fam_file PedFam_analysis/dels_combined.fam \
  --concat_files n \
  --merge_files none \
  > PedFam_analysis/concat_merge_dels.out
# Filtered SVs files
docker run -d -v /Data:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/concat_merge_phased_vcf.py \
  PedFam analysis/svs files/ \
  PedFam_analysis/svs_combined.vcf.gz \
  --output_fam_file PedFam_analysis/svs_combined.fam \
  --concat_files n \
  --merge files none \
  > PedFam_analysis/concat_merge_svs.out
```

#### Normalized and left aligned with vt tools

\*Only the combined SNP file was left aligned and normalized. The SV and del combined files were not able to be left aligned and normalized due to limitations in the reference genomes used.

```
# Combined SNP file
docker run -d -v /Data:/proj -v /Data/PedFam_analysis/references:/references -w /proj \
   -t dmill903/compound-het-vip:1.0 \
   python3 PedFam_analysis/PedFam/scripts/vt_split_trim_left_align.py \
   PedFam_analysis/combined.vcf.gz \
   PedFam_analysis/combined_vt.vcf.gz \
   PedFam_analysis/vt_split_trim_left_align.out
```

#### Annotate

```
# Combined, normalized SNP file
docker run -d -v /Data:/proj \
  -v /Data/PedFam_analysis/references/snpEff_data:/snpEff/./data/GRCh38.86 -w /proj \
```

```
-t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/annotate.py \
  PedFam_analysis/combined_vt.vcf.gz \
  PedFam_analysis/combined_annotated.vcf \
  > PedFam_analysis/annotate.out
# Combined del file
docker run -d -v /Data:/proj \
  -v /Data/PedFam_analysis/references/snpEff_data:/snpEff/./data/GRCh38.86 -w /proj \
  -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/annotate.py \
  PedFam_analysis/dels_combined.vcf.gz \
  PedFam_analysis/dels_annotated.vcf \
  > PedFam_analysis/annotate_dels.out
# Combined SV file
docker run -d -v /Data:/proj \
  -v /Data/PedFam_analysis/references/snpEff_data:/snpEff/./data/GRCh38.86 -w /proj \
  -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/annotate.py \
  PedFam_analysis/svs_combined.vcf.gz \
  PedFam_analysis/svs_annotated.vcf \
  > PedFam_analysis/annotate_svs.out
```

#### Load files into a GEMINI Database

```
# Annotated SNP file
docker run -d -v /Data:/proj \
  -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/gemini_load.py \
  PedFam_analysis/combined_annotated.vcf \
  PedFam_analysis/combined.db \
  --fam_file PedFam_analysis/combined.fam \
  > PedFam_analysis/gemini_load.out
# Annotated del file
docker run -d -v /Data:/proj \
  -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/gemini_load.py \
  PedFam analysis/dels annotated.vcf \
  PedFam_analysis/dels_combined.db \
  --fam_file PedFam_analysis/dels_combined.fam \
  > PedFam_analysis/gemini_load_dels.out
# Annotated SV file
docker run -d -v /Data:/proj \
  -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/gemini_load.py \
  PedFam_analysis/svs_annotated.vcf \
  PedFam_analysis/svs_combined.db \
  --fam_file PedFam_analysis/svs_combined.fam \
 > PedFam_analysis/gemini_load_svs.out
```

# Create a file that contains global gnomAD minor allele frequencies, global 1000 Genomes Project minor allele frequences, and CADD scores

This file is used to provide each variant (when possible) with minor allele frequency, and a CADD score. File is output as gnomAD\_1K\_cadd\_GRCh38.tsv.gz.

```
docker run -d -v /Data/PedFam_analysis:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
   python3 PedFam/scripts/create_gnomAD_1K_cadd_file.py \
        create_gnomAD_1K_cadd_file.out
```

#### Identify compound heterozygous variants

```
# SNP GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/identify_CH_variants.py \
  PedFam analysis/combined.db \
  PedFam analysis/PedFam CH cadd20 maf01.tsv \
  PedFam_analysis/gnomAD_1K_cadd_GRCh38.tsv.gz \
  --fam_file PedFam_analysis/combined.fam \
  --cadd 20 \
  --af 0.01 \
  > PedFam analysis/identify CH variants.out
# del GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/identify_CH_variants.py \
  PedFam_analysis/dels_combined.db \
  PedFam_analysis/PedFam_CH_cadd20_maf01_dels.tsv \
  PedFam_analysis/gnomAD_1K_cadd_GRCh38.tsv.gz \
  --fam_file PedFam_analysis/dels_combined.fam \
  --cadd 20 \
  --af 0.01 \
  --impact filter only y \
  > PedFam_analysis/identify_CH_variants_dels.out
# SV GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam analysis/PedFam/scripts/identify CH variants.py \
  PedFam analysis/svs combined.db \
  PedFam_analysis/PedFam_CH_cadd20_maf01_svs.tsv \
  PedFam_analysis/gnomAD_1K_cadd_GRCh38.tsv.gz \
  --fam_file PedFam_analysis/svs_combined.fam \
  --cadd 20 \
  --af 0.01 \
  --impact_filter_only y \
  > PedFam_analysis/identify_CH_variants_svs.out
```

## Identify de novo variants

```
# SNP GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/identify_deNovo_variants.py \
  PedFam analysis/combined.db \
  PedFam analysis/PedFam deNovo cadd20 maf01.tsv \
  PedFam_analysis/gnomAD_1K_cadd_GRCh38.tsv.gz \
  PedFam_analysis/combined.fam \
  --cadd 20 \
  --af 0.01 \
  > PedFam_analysis/identify_deNovo_variants.out
# del GEMINI databaase
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/identify_deNovo_variants.py \
  PedFam_analysis/dels_combined.db \
 PedFam analysis/PedFam deNovo cadd20 maf01 dels.tsv \
  PedFam_analysis/gnomAD_1K_cadd_GRCh38.tsv.gz \
  PedFam analysis/dels combined.fam \
  --cadd 20 \
  --af 0.01 \
  --impact filter only y \
  > PedFam_analysis/identify_deNovo_variants_dels.out
# SV GEMINI databaase
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/identify_deNovo_variants.py \
  PedFam_analysis/svs_combined.db \
  PedFam_analysis/PedFam_deNovo_cadd20_maf01_svs.tsv \
  PedFam_analysis/gnomAD_1K_cadd_GRCh38.tsv.gz \
  PedFam_analysis/svs_combined.fam \
  --cadd 20 \
  --af 0.01 \
  --impact filter only y \
  > PedFam_analysis/identify_deNovo_variants_svs.out
```

#### Identify homozygous alternate variants

```
# SNP GEMINI databaase
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
    python3 PedFam_analysis/PedFam/scripts/identify_homAlt_variants.py \
    PedFam_analysis/combined.db \
    PedFam_analysis/PedFam_homAlt_cadd20_maf01.tsv \
    PedFam_analysis/gnomAD_1K_cadd_GRCh38.tsv.gz \
    --fam_file PedFam_analysis/combined.fam \
    --cadd 20 \
    --af 0.01 \
    PedFam_analysis/identify_homAlt_variants.out

# del GEMINI databaase
```

```
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/identify_homAlt_variants.py \
  PedFam_analysis/dels_combined.db \
  PedFam_analysis/PedFam_homAlt_cadd20_maf01_dels.tsv \
  PedFam_analysis/gnomAD_1K_cadd_GRCh38.tsv.gz \
  --fam_file PedFam_analysis/dels_combined.fam \
  --cadd 20 \
  --af 0.01 \
  --impact filter only y \
  > PedFam_analysis/identify_homAlt_variants_dels.out
# SV GEMINI databaase
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/identify_homAlt_variants.py \
  PedFam_analysis/svs_combined.db \
  PedFam_analysis/PedFam_homAlt_cadd20_maf01_svs.tsv \
  PedFam_analysis/gnomAD_1K_cadd_GRCh38.tsv.gz \
  --fam_file PedFam_analysis/svs_combined.fam \
  --cadd 20 \
  --af 0.01 \
  --impact_filter_only y \
  > PedFam_analysis/identify_homAlt_variants_svs.out
```

#### Identify heterozygous variants

```
# SNP GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/identify_het_variants.py \
  PedFam_analysis/combined.db \
  PedFam_analysis/PedFam_het_cadd20_maf01.tsv \
  PedFam_analysis/gnomAD_1K_cadd_GRCh38.tsv.gz \
  --fam file PedFam analysis/combined.fam \
  --cadd 20 \
  --af 0.01 \
  > PedFam_analysis/identify_het_variants.out
# del GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/identify_het_variants.py \
  PedFam_analysis/dels_combined.db \
  PedFam_analysis/PedFam_het_cadd20_maf01_dels.tsv \
  PedFam_analysis/gnomAD_1K_cadd_GRCh38.tsv.gz \
  --fam_file PedFam_analysis/dels_combined.fam \
  --cadd 20 \
  --af 0.01 \
  --impact_filter_only y \
  > PedFam_analysis/identify_het_variants_dels.out
# SV GEMINI databaase
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam_analysis/PedFam/scripts/identify_het_variants.py \
```

```
PedFam_analysis/svs_combined.db \
PedFam_analysis/PedFam_het_cadd20_maf01_svs.tsv \
PedFam_analysis/gnomAD_1K_cadd_GRCh38.tsv.gz \
--fam_file PedFam_analysis/svs_combined.fam \
--cadd 20 \
--af 0.01 \
--impact_filter_only y \
> PedFam_analysis/identify_het_variants_svs.out
```

## Pediatric Cancer Prediction Analysis

#### **Functions**

```
# Combines dataframes, as long as there is information in the dataframes
combine df <- function(aList) {</pre>
  newList <- list()</pre>
  for (df in aList) {
    if (nrow(df) > 0) {
      newList <- c(newList, list(df))</pre>
    }
  newDf <- bind_rows(newList)</pre>
  return(newDf)
}
# Reads in a csv and removes parental samples
read_tsv_custom <- function(inputFile) {</pre>
  return(read_tsv(inputFile, col_types = cols(alt = col_character(),
                                                ref = col_character(),
                                                af gnomAD = col character(),
                                                cadd = col_character())) %>%
           dplyr::filter(!sample %in% c("4478_SP_0023", "4478_SP_0024",
                                            "4478_SP_0030", "4478_SP_0031",
                                            "4478_SP_0036", "4478_SP_0037")))
}
# Variation of the other custom csv script. For csv's that don't have all the
# colums that are seen in the other csv script.
read_tsv_custom_2 <- function(inputFile) {</pre>
  return(read_tsv(inputFile) %% dplyr::filter(!sample %in% c("4478-SP-0023",
                                                                 "4478-SP-0024",
                                                                 "4478-SP-0030",
                                                                 "4478-SP-0031",
                                                                 "4478-SP-0036".
                                                                 "4478-SP-0037")))
}
# Returns only the astrocytoma samples
astrocytoma_sample_filter <- function(inputDf) {</pre>
  return(dplyr::filter(inputDf, sample %in% c("4478_SP_0025", "4478_SP_0026",
```

```
"4478_SP_0027", "4478_SP_0028",
                                                "4478_SP_0029")))
}
# Returns only the wilms tumor samples
wilms_sample_filter <- function(inputDf) {</pre>
  return(dplyr::filter(inputDf, sample %in% c("4478_SP_0032", "4478_SP_0033",
                                                "4478_SP_0034", "4478_SP_0035")))
}
# Returns only the lymphoma samples
lymphoma_sample_filter <- function(inputDf) {</pre>
  return(dplyr::filter(inputDf, sample %in% c("4478_SP_0038", "4478_SP_0039",
                                                "4478_SP_0040", "4478_SP_0041",
                                                "4478_SP_0042", "4478_SP_0043",
                                                "4478 SP 0044")))
}
# Adds a column to the input dataframe that indicates the disease type
add_disease_column <- function(inputDf) {</pre>
  astrocytoma temp <- dplyr::filter(inputDf, sample %in% c("25", "26", "27",
                                                              "28", "29")) %>%
    mutate(disease = "low-grade astrocytoma")
  lymphoma_temp <- dplyr::filter(inputDf, sample %in% c("38", "39", "40", "41",
                                                          "42", "43", "44")) %>%
    mutate(disease = "Burkitt's lymphoma")
  wilms_temp <- dplyr::filter(inputDf, sample %in% c("32", "33", "34", "35")) %>%
    mutate(disease = "Wilms tumor")
  return(bind_rows(astrocytoma_temp, lymphoma_temp, wilms_temp))
}
# Summarize the variants to the gene level
gene_level_filter <- function(df, disease) {</pre>
  tempDf <- select(df, sample, gene)</pre>
  finalDf <- add column(tempDf, 'disease' = disease)</pre>
 return(finalDf)
}
# Summarize how many variants per sample
variant_level_filter <- function(df) {</pre>
  tempDf <- filter(df, variant_type != "Compound Heterozygous") %>%
    group_by(sample, variant_type, class) %>% summarise(variants = n())
  tempDf2 <- filter(df, variant_type == "Compound Heterozygous") %>%
    group_by(sample, variant_type, class, gene) %>%
    summarise(variants = length(unique(gene))) %>% select(!gene)
  finalDf <- bind_rows(tempDf, tempDf2)</pre>
  return(finalDf)
}
```

```
# Summarize shared variants
shared_variant_filter <- function(df) {</pre>
  tempDf <- group_by(df, chrom, start, ref, alt, family, sample) %>%
    summarise(shared variants = n())
  tempDf <- group_by(tempDf, chrom, start, ref, alt, family) %>%
    summarise(shared_variants = n())
 return(tempDf)
}
# add family ID to df
add_family_ID <- function(df){</pre>
  fam1 <- filter(df, sample %in% c("4478_SP_0025", "4478_SP_0026",</pre>
                                    "4478_SP_0027", "4478_SP_0028",
                                    "4478_SP_0029")) %>%
    mutate(family = "Family 1")
  fam2 <- filter(df, sample %in% c("4478_SP_0032", "4478_SP_0033",</pre>
                                    "4478_SP_0034", "4478_SP_0035")) %>%
    mutate(family = "Family 2")
  fam3 <- filter(df, sample %in% c("4478_SP_0038", "4478_SP_0039",
                                    "4478_SP_0040", "4478_SP_0041",
                                    "4478_SP_0042", "4478_SP_0043",
                                    "4478 SP 0044")) %>%
    mutate(family = "Family 3")
  return(bind_rows(fam1, fam2, fam3))
}
```

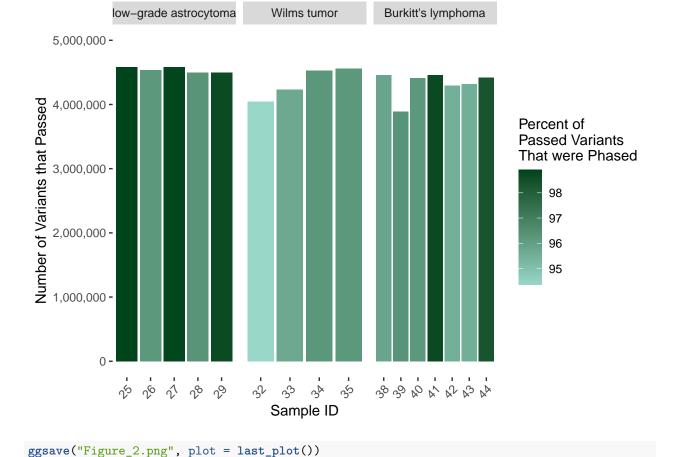
#### Variant Stats of Passed and Phased variants

The passed\_variants\_summary.tsv, passed\_dels\_summary.tsv, and passed\_svs\_summary.tsv were generated with the keep\_passed\_variants.py script.

## [1] "Average number of variants that passed quality filters: 4395811"

## Create figure of passed/phased numbers

This code chunk was used to create Figure 2 in the manuscript.



## Read in CH data and separate by disease

```
wilms_CH_gene_level <- gene_level_filter(wilms_CH_data, "wilms")

# Filter down to lymphoma data and filter to gene level
lymphoma_CH_data <- lymphoma_sample_filter(combined_CH_all)
lymphoma_CH_gene_level <- gene_level_filter(lymphoma_CH_data, "lymphoma")</pre>
```

## Read in de Novo data and separate by disease

```
# Read in de novo data and generate new columns with variant type and class
combined_deNovo_data_vars <- read_tsv_custom("PedFam_deNovo_cadd20_maf01.tsv") %>%
  add_column(variant_type = "De Novo", class = "SNP")
combined_deNovo_data_svs <- read_tsv_custom("PedFam_deNovo_cadd20_maf01_svs.tsv") %>%
  add column(variant type = "De Novo", class = "SV")
combined_deNovo_data_dels <- read_tsv_custom("PedFam_deNovo_cadd20_maf01_dels.tsv") %>%
  add_column(variant_type = "De Novo", class = "indel")
# Combine all imported data into a single dataframe
combined deNovo all <- combine df(list(combined deNovo data vars,
                                       combined deNovo data svs,
                                       combined_deNovo_data_dels))
# Filter down to astrocytoma data and filter to gene level
astrocytoma deNovo data <- astrocytoma sample filter(combined deNovo all)
astrocytoma_deNovo_gene_level <- gene_level_filter(astrocytoma_deNovo_data,
                                                    "astrocytoma")
# Filter down to wilms data and filter to gene level
wilms_deNovo_data <- wilms_sample_filter(combined_deNovo_all)</pre>
wilms_deNovo_gene_level <- gene_level_filter(wilms_deNovo_data, "wilms")</pre>
# Filter down to lymphoma data and filter to gene level
lymphoma_deNovo_data <- lymphoma_sample_filter(combined_deNovo_all)</pre>
lymphoma_deNovo_gene_level <- gene_level_filter(lymphoma_deNovo_data, "lymphoma")
```

## Read in homozygous alternate data and separate by disease

```
# Read in homozygous alternate data and generate new columns with variant type
# and class
combined_homAlt_data_vars <- read_tsv_custom("PedFam_homAlt_cadd20_maf01.tsv") %>%
   add_column(variant_type = "Homozygous Alternate", class = "SNP")

combined_homAlt_data_svs <- read_tsv_custom("PedFam_homAlt_cadd20_maf01_svs.tsv") %>%
   add_column(variant_type = "Homozygous Alternate", class = "SV")

combined_homAlt_data_dels <- read_tsv_custom("PedFam_homAlt_cadd20_maf01_dels.tsv") %>%
   add_column(variant_type = "Homozygous Alternate", class = "indel")
```

## Read in heterozygous data and separate by disease

```
# Read in heterozygous data and generate new columns with variant type and class
combined_het_data_vars <- read_tsv_custom("PedFam_het_cadd20_maf01.tsv") %>%
  add_column(variant_type = "Heterozygous", class = "SNP")
combined_het_data_svs <- read_tsv_custom("PedFam_het_cadd20_maf01_svs.tsv") %>%
  add_column(variant_type = "Heterozygous", class = "SV")
combined_het_data_dels <- read_tsv_custom("PedFam_het_cadd20_maf01_dels.tsv") %>%
  add_column(variant_type = "Heterozygous", class = "indel")
# Combine all imported data into a single dataframe
combined_het_all <- combine_df(list(combined_het_data_vars,</pre>
                                     combined_het_data_svs, combined_het_data_dels))
# Filter down to astrocytoma data and filter to gene level
astrocytoma het data <- astrocytoma sample filter(combined het all)
astrocytoma_het_gene_level <- gene_level_filter(astrocytoma_het_data, "astrocytoma")
# Filter down to wilms data and filter to gene level
wilms_het_data <- wilms_sample_filter(combined_het_all)</pre>
wilms_het_gene_level <- gene_level_filter(wilms_het_data, "wilms")</pre>
# Filter down to lymphoma data and filter to gene level
lymphoma_het_data <- lymphoma_sample_filter(combined_het_all)</pre>
lymphoma_het_gene_level <- gene_level_filter(lymphoma_het_data, "lymphoma")</pre>
```

## Combine all data regardless of data type, and summarise at gene and variant levels

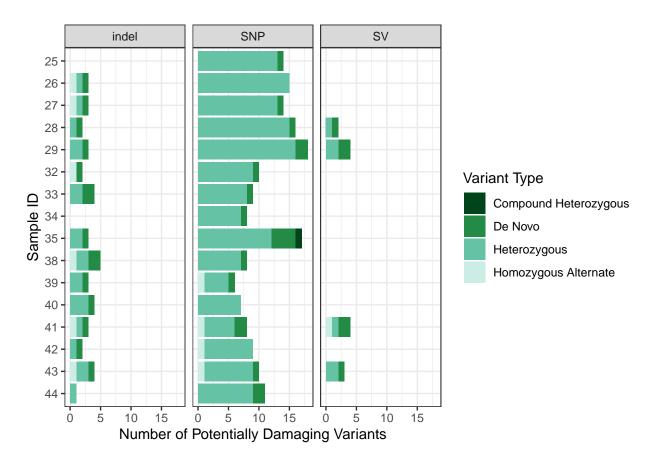
This code chunk was used to generate data for Table 1 in the manuscript and Supplementary Table 1.

```
#Combine all raw data
combined_data_all <- combine_df(list(combined_homAlt_all, combined_deNovo_all,</pre>
                                     combined het all, combined CH all))
# Combine all data at gene level
gene_level_all <- combine_df(list(astrocytoma_CH_gene_level,</pre>
                                  astrocytoma_deNovo_gene_level,
                                  astrocytoma_homAlt_gene_level,
                                  astrocytoma_het_gene_level,
                                  lymphoma_CH_gene_level,
                                  lymphoma_deNovo_gene_level,
                                  lymphoma het gene level,
                                  lymphoma_homAlt_gene_level,
                                  wilms_CH_gene_level, wilms_deNovo_gene_level,
                                  wilms_het_gene_level, wilms_homAlt_gene_level))
# Gene level summary (how many samples each potentially damaged gene has)
gene_level_summary_all <- group_by(gene_level_all, disease, gene) %>%
  summarise(num_samples_with_damage = length(unique(sample)))
write_tsv(gene_level_summary_all, "gene_level_summary.tsv")
gene_level_summary_all
## # A tibble: 84 x 3
## # Groups: disease [3]
##
      disease
                  gene
                           num_samples_with_damage
##
      <chr>
                  <chr>
                                              <int>
## 1 astrocytoma ACADS
                                                  1
## 2 astrocytoma ACOXL
                                                  4
## 3 astrocytoma ALG1L2
                                                  1
## 4 astrocytoma BST1
                                                  5
## 5 astrocytoma CBWD5
                                                  3
## 6 astrocytoma CRNKL1
                                                  3
## 7 astrocytoma DAPL1
                                                  3
## 8 astrocytoma DUOXA1
                                                  3
## 9 astrocytoma FAM8A1
                                                  2
## 10 astrocytoma GOLGA6L1
## # ... with 74 more rows
# Gene and sample level summary (how many genes have x number of samples with a
# variant in that gene)
shared_genes_summary <- group_by(gene_level_summary_all, disease,</pre>
                                 num_samples_with_damage) %>%
  summarise(genes_with_num_damaged_genes = length(unique(gene)))
shared_genes_summary
```

```
## # A tibble: 14 x 3
## # Groups: disease [3]
     disease num_samples_with_damage genes_with_num_damaged_genes
##
     <chr>
                                   <int>
## 1 astrocytoma
## 2 astrocytoma
                                       2
                                                                   5
                                       3
## 3 astrocytoma
## 4 astrocytoma
                                       4
                                                                   6
## 5 astrocytoma
                                       5
                                                                   3
## 6 lymphoma
                                       1
                                                                  14
## 7 lymphoma
                                                                   8
                                       3
## 8 lymphoma
                                                                   4
                                       4
## 9 lymphoma
                                                                   4
## 10 lymphoma
                                       5
                                                                   2
## 11 wilms
                                       1
                                                                  11
                                       2
## 12 wilms
                                                                   5
## 13 wilms
                                       3
                                                                   4
                                       4
                                                                   2
## 14 wilms
# Find any shared variants
shared_variants <- shared_variant_filter(add_family_ID(combined_data_all))</pre>
# Summarise shared variants
shared_variants_summary <- group_by(shared_variants, family, shared_variants) %>%
 summarise(total = n())
shared_variants_summary
## # A tibble: 14 x 3
## # Groups: family [3]
##
     family shared_variants total
##
     <chr>
                       <int> <int>
## 1 Family 1
                          1
                                 12
## 2 Family 1
                                  7
## 3 Family 1
                            3
## 4 Family 1
                            4
                                  6
                            5
## 5 Family 1
                                3
## 6 Family 2
                           1 12
## 7 Family 2
                            2
                                5
## 8 Family 2
                            3
                                 4
## 9 Family 2
## 10 Family 3
                            1 15
## 11 Family 3
                                 8
## 12 Family 3
                            3
                                  5
## 13 Family 3
                                  5
## 14 Family 3
                                  1
```

# Generate a Figure showing a variant level summary that shows variant type and class

This code chunk was used to generate Figure 3 for the manuscript.



```
# Save figure
ggsave("Figure_3.png", plot = last_plot())
```

#### Find genes that have 2 or less siblings with a variant

This code chunk provided a list of genes for each family that was used as input into VarElect.

```
# Astrocytoma data
paste("astrocytoma: ", str_c(filter(gene_level_summary_all,
                                     disease == "astrocytoma" &
                                       num_samples_with_damage <= 2)$gene,</pre>
                              collapse = ","))
## [1] "astrocytoma: ACADS, ALG1L2, FAM8A1, GOLGA6L1, GOLGA6L2, GUCA1C, HLA-DRB1, KRT76, RP11-294C11.1, SIRPB1,
paste("Number of genes to input to VarEelct for astrocytoma: ",
      length(filter(gene_level_summary_all, disease == "astrocytoma" &
                      num_samples_with_damage <= 2)$gene))</pre>
## [1] "Number of genes to input to VarEelct for astrocytoma: 15"
# Lymphoma data
paste("lymphoma: ", str_c(filter(gene_level_summary_all, disease == "lymphoma" &
                                    num_samples_with_damage <= 2)$gene,</pre>
                           collapse = ","))
## [1] "lymphoma: ADGRA2,Clorf50,C2orf16,CCDC179,CSNK1A1,DCHS1,DNTTIP2,ENPEP,GLYATL1,ITGB4,KRTAP10-12,
paste("Number of genes to input to VarEelct for lymphoma: ",
      length(filter(gene_level_summary_all, disease == "lymphoma" &
                      num_samples_with_damage <= 2)$gene))</pre>
## [1] "Number of genes to input to VarEelct for lymphoma: 22"
# Wilms data
paste("wilms: ", str_c(filter(gene_level_summary_all, disease == "wilms" &
                                 num_samples_with_damage <= 2)$gene,</pre>
                       collapse = ","))
## [1] "wilms: ANKRD36C,CYP2A13,DPY19L3,FAM8A1,GOSR2,HLA-DRB1,HLA-DRB5,PAH,PCK1,PCK2,PNPLA7,RP1L1,SIPA
paste("Number of genes to input to VarEelct for wilms: ",
      length(filter(gene_level_summary_all, disease == "wilms" &
                      num_samples_with_damage <= 2)$gene))</pre>
## [1] "Number of genes to input to VarEelct for wilms:
```

Figure how how many unique genes there were before and after sibling filtering

## [1] "Across all families, there were 47 unique genes that had a potentially damaing variant in up

```
paste("Across all famlies, there were ",
    length(unique(gene_level_summary_all$gene)),
    " unique genes with potentially damaging variants.")
```

## [1] "Across all famlies, there were 72 unique genes with potentially damaging variants."

Create a vector of known cancer genes using COSMIC

```
cosmic <- read_tsv("known_genes/Census_allThu Apr 16 21_44_49 2020.tsv")
cosmic_all <- cosmic$'Gene Symbol'</pre>
```

Check to see if any of the children had a potentially damaing variant in a known cancer gene

```
cancer_genes <- c()
for(gene in combined_data_all$gene){
  if (gene %in% cosmic_all & !gene %in% cancer_genes) {
    cancer_genes <- c(cancer_genes, gene)
  }
}
paste("The following genes are known cancer genes and were identified in at least on child across all f
    str_c(cancer_genes, collapse = ", "))</pre>
```

## [1] "The following genes are known cancer genes and were identified in at least on child across all :