# Toward a methodology for evaluating DNA variants in nuclear families: data processing and analysis

**Dustin Miller** 

06/10/2021

#### **VCF Processing**

For each step of VCF filtering, we used a Docker image that encapsulated the software tools as well as Python scripts (available on github) to process the data. These Python scripts either used custom code or a combination of custom code and existing software. Some of the scripts used during the VCF filtering process were adapted from CompoundHetVIP.

The BAM files used as part of our analyses are available from the European Genome-Phenome Archive (accession: EGAS00001005321).

A more detailed description of each processing step is available in the manuscript.

#### Filter out poor quality variants for each sample and each file type

Initial files being filtered are those that were output by *Long Ranger*. *Long Ranger* output a phased SNP VCF, a phased mid-scale deletion VCF, and a phased large-scale SV VCF for each sample.

```
# Phased SNP files
docker run -d -v /Data:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/keep passed variants.py \
  PedFam/*/outs/phased_variants.vcf.gz \
  phased files/ \
  passed variants summary.tsv \
  > keep_passed_variants.out
# Phased dels files
docker run -d -v /Data:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/keep passed variants.py \
  PedFam/*/outs/dels.vcf.gz \
  dels files/ \
  passed_dels_summary.tsv \
  > keep_passed_dels.out
# Phased svs files
docker run -d -v /Data:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/keep passed variants.py \
  PedFam/*/outs/large_svs.vcf.gz \
```

```
svs_files/ \
passed_svs_summary.tsv \
> keep_passed_svs.out
```

#### **Combine sample files**

```
# Filtered SNP files
docker run -d -v /Data:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/concat merge phased vcf.py \
  phased_files/ \
  combined.vcf.gz \
  --output fam file combined.fam \
  --concat files n \
  --merge files all \
  > concat_merge_phased_vcf.out
# Filtered dels files
docker run -d -v /Data:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/concat_merge_phased_vcf.py \
  dels files/ \
  dels combined.vcf.gz \
  --output_fam_file dels_combined.fam \
  --concat files n \
  --merge_files none \
  > concat_merge_dels.out
# Filtered SVs files
docker run -d -v /Data:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/concat merge phased vcf.py \
  svs files/ \
  svs combined.vcf.gz \
  --output_fam_file svs_combined.fam \
  --concat_files n \
  --merge files none \
  > concat_merge_svs.out
```

#### Normalized and left aligned with vt tools

\*Only the combined SNP file was left aligned and normalized. The SV and del combined files were not able to be left aligned and normalized due to limitations in the reference genomes used.

```
# Combined SNP file
docker run -d -v /Data:/proj -v /Data/references:/references -w /proj \
   -t dmill903/compound-het-vip:1.0 \
   python3 PedFam/scripts/vt_split_trim_left_align.py \
   combined.vcf.gz \
   combined_vt.vcf.gz \
   > vt_split_trim_left_align.out
```

#### **Annotaate**

```
# Combined, normalized SNP file
docker run -d -v /Data:/proj \
  -v /Data/references/snpEff_data:/snpEff/./data/GRCh38.86 -w /proj \
  -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/annotate.py \
  combined vt.vcf.gz \
  combined annotated.vcf \
  > annotate.out
# Combined del file
docker run -d -v /Data:/proj \
  -v /Data/references/snpEff_data:/snpEff/./data/GRCh38.86 -w /proj \
  -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/annotate.py \
  dels combined.vcf.gz \
  dels annotated.vcf \
  > annotate_dels.out
# Combined SV file
docker run -d -v /Data:/proj \
  -v /Data/references/snpEff data:/snpEff/./data/GRCh38.86 -w /proj \
  -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/annotate.py \
  svs combined.vcf.gz \
  svs_annotated.vcf \
 > annotate svs.out
```

#### Load files into a GEMINI Database

```
# Annotated SNP file
docker run -d -v /Data:/proj \
  -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/gemini_load.py \
  combined annotated.vcf \
  combined.db \
  --fam file combined.fam \
  > gemini_load.out
# Annotated del file
docker run -d -v /Data:/proj \
  -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/gemini_load.py \
  dels annotated.vcf \
  dels combined.db \
  --fam_file dels_combined.fam \
  > gemini load dels.out
# Annotated SV file
docker run -d -v /Data:/proj \
```

```
-w /proj -t dmill903/compound-het-vip:1.0 \
python3 PedFam/scripts/gemini_load.py \
svs_annotated.vcf \
svs_combined.db \
--fam_file svs_combined.fam \
> gemini_load_svs.out
```

# Create a file that contains global gnomAD minor allele frequencies, global 1000 Genomes Project minor allele frequences, and CADD scores

This file was used to provide each variant (when possible) with minor allele frequency, and a CADD score. File was output as gnomAD\_1K\_cadd\_GRCh38.tsv.gz.

```
docker run -d -v /Data:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
   python3 PedFam/scripts/create_gnomAD_1K_cadd_file.py \
        create_gnomAD_1K_cadd_file.out
```

#### **Identify compound heterozygous variants**

```
# SNP GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/identify_CH_variants.py \
  combined.db \
  PedFam CH cadd20 maf01.tsv \
  gnomAD 1K cadd GRCh38.tsv.gz \
  --fam file combined.fam \
  --cadd 20 \
  --af 0.01 \
  > identify CH variants.out
# del GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/identify CH variants.py \
  dels combined.db \
  PedFam_CH_cadd20_maf01_dels.tsv \
  gnomAD 1K cadd GRCh38.tsv.gz \
  --fam_file dels_combined.fam \
  --cadd 20 \
  --af 0.01 \
  --impact_filter_only y \
  > identify CH variants dels.out
# SV GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/identify CH variants.py \
  svs combined.db \
  PedFam CH cadd20 maf01 svs.tsv \
  gnomAD_1K_cadd_GRCh38.tsv.gz \
```

```
--fam_file svs_combined.fam \
--cadd 20 \
--af 0.01 \
--impact_filter_only y \
> identify_CH_variants_svs.out
```

#### Identify de novo variants

```
# SNP GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/identify_deNovo_variants.py \
  combined.db \
  PedFam deNovo cadd20 maf01.tsv \
  gnomAD_1K_cadd_GRCh38.tsv.gz \
  combined.fam \
  --cadd 20 \
  --af 0.01 \
  > identify_deNovo_variants.out
# del GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/identify_deNovo_variants.py \
  dels combined.db \
  PedFam_deNovo_cadd20_maf01_dels.tsv \
  gnomAD_1K_cadd_GRCh38.tsv.gz \
  dels combined.fam \
  --cadd 20 \
  --af 0.01 \
  --impact_filter_only y \
  > identify_deNovo_variants_dels.out
# SV GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/identify deNovo variants.py \
  svs combined.db \
  PedFam_deNovo_cadd20_maf01_svs.tsv \
  gnomAD_1K_cadd_GRCh38.tsv.gz \
  svs combined.fam \
  --cadd 20 \
  --af 0.01 \
  --impact filter only y \
  > identify_deNovo_variants_svs.out
```

#### **Identify homozygous alternate variants**

```
# SNP GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
    python3 PedFam/scripts/identify_homAlt_variants.py \
    combined.db \
    PedFam_homAlt_cadd20_maf01.tsv \
    gnomAD_1K_cadd_GRCh38.tsv.gz \
```

```
--fam file combined.fam \
  --cadd 20 \
  --af 0.01 \
  > identify_homAlt_variants.out
# del GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/identify_homAlt_variants.py \
  dels combined.db \
  PedFam_homAlt_cadd20_maf01_dels.tsv \
  gnomAD_1K_cadd_GRCh38.tsv.gz \
  --fam_file dels_combined.fam \
  --cadd 20 \
  --af 0.01 \
  --impact filter only y \
  > identify_homAlt_variants_dels.out
# SV GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/identify_homAlt_variants.py \
  svs combined.db \
  PedFam_homAlt_cadd20_maf01_svs.tsv \
  gnomAD 1K cadd GRCh38.tsv.gz \
  --fam file svs combined.fam \
  --cadd 20 \
  --af 0.01 \
  --impact filter only y \
 > identify_homAlt_variants_svs.out
```

#### **Identify heterozygous variants**

```
# SNP GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/identify het variants.py \
  combined.db \
  PedFam_het_cadd20_maf01.tsv \
  gnomAD 1K cadd GRCh38.tsv.gz \
  --fam_file combined.fam \
  --cadd 20 \
  --af 0.01 \
  > identify_het_variants.out
# del GEMINI database
docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
  python3 PedFam/scripts/identify_het_variants.py \
  dels combined.db \
  PedFam het cadd20 maf01 dels.tsv \
  gnomAD 1K cadd GRCh38.tsv.gz \
  --fam file dels combined.fam \
  --cadd 20 \
```

```
--af 0.01 \
--impact_filter_only y \
> identify_het_variants_dels.out

# SV GEMINI database

docker run -d -v /Data/:/proj -w /proj -t dmill903/compound-het-vip:1.0 \
    python3 PedFam/scripts/identify_het_variants.py \
    svs_combined.db \
    PedFam_het_cadd20_maf01_svs.tsv \
    gnomAD_1K_cadd_GRCh38.tsv.gz \
    --fam_file svs_combined.fam \
    --cadd 20 \
    --af 0.01 \
    --impact_filter_only y \
    identify_het_variants_svs.out
```

#### **Pediatric Cancer Prediction Analysis with R (and a little Python)**

#### **Functions**

```
# Combines dataframes, as long as there is information in the dataframes
combine df <- function(aList) {</pre>
 newList <- list()</pre>
 for (df in aList) {
    if (nrow(df) > 0) {
     newList <- c(newList, list(df))</pre>
   }
 }
 newDf <- bind rows(newList)</pre>
 return(newDf)
# Reads in a csv and removes parental samples
read tsv custom <- function(inputFile) {</pre>
 return(read_tsv(inputFile, col_types = cols(alt = col_character(),
                                             ref = col character(),
                                             af gnomAD = col character(),
                                             cadd = col_character())) %>%
          }
# Variation of the other custom csv script. For csv's that don't have all the
# columns that are seen in the other csv script.
read_tsv_custom_2 <- function(inputFile) {</pre>
  return(read tsv(inputFile) %>% dplyr::filter(!sample %in% c("4478-SP-0023",
                                                             "4478-SP-0024"
                                                            "4478-SP-0030",
```

```
"4478-SP-0031",
                                                             "4478-SP-0036",
                                                             "4478-SP-
0037")))
}
# Returns only the astrocytoma samples
astrocytoma sample filter <- function(inputDf) {</pre>
 return(dplyr::filter(inputDf, sample %in% c("4478 SP 0025", "4478 SP 0026",
                                             "4478_SP_0027", "4478_SP_0028",
                                             "4478 SP 0029")))
}
# Returns only the wilms tumor samples
wilms sample filter <- function(inputDf) {</pre>
 return(dplyr::filter(inputDf, sample %in% c("4478_SP_0032", "4478_SP_0033",
                                             "4478 SP 0034".
"4478 SP 0035")))
}
# Returns only the lymphoma samples
lymphoma sample_filter <- function(inputDf) {</pre>
 "4478 SP 0044")))
}
# Adds a column to the input dataframe that indicates the disease type
add disease column <- function(inputDf) {</pre>
 astrocytoma temp <- dplyr::filter(inputDf, sample %in% c("25", "26", "27",
                                                          "28", "29")) %>%
   mutate(disease = "low-grade astrocytoma")
 lymphoma_temp <- dplyr::filter(inputDf, sample %in% c("38", "39", "40",
"41",
                                                       "42", "43", "44"))
%>%
   mutate(disease = "Burkitt's lymphoma")
 wilms temp <- dplyr::filter(inputDf, sample %in% c("32", "33", "34", "35"))</pre>
%>%
   mutate(disease = "Wilms tumor")
 return(bind rows(astrocytoma temp, lymphoma temp, wilms temp))
}
# Summarize the variants to the gene level
gene_level_filter <- function(df, disease) {</pre>
```

```
tempDf <- select(df, sample, gene)</pre>
  finalDf <- add column(tempDf, `disease` = disease)</pre>
  return(finalDf)
}
# Summarize how many variants per sample
variant_level_filter <- function(df) {</pre>
  tempDf <- filter(df, variant_type != "Compound Heterozygous") %>%
    group by(sample, variant type, class) %>% summarise(variants = n())
  tempDf2 <- filter(df, variant type == "Compound Heterozygous") %>%
    group_by(sample, variant_type, class, gene) %>%
    summarise(variants = length(unique(gene))) %>% select(!gene)
  finalDf <- bind_rows(tempDf, tempDf2)</pre>
  return(finalDf)
}
variant level filter 2 <- function(df) {</pre>
  finalDf <- group_by(df, sample, variant_type, class) %>% summarise(variants
= n())
  return(finalDf)
}
# Summarize shared variants
shared_variant_filter <- function(df) {</pre>
  tempDf <- group_by(df, chrom, start, ref, alt, family, sample) %>%
    summarise(shared_variants = n())
  tempDf <- group_by(tempDf, chrom, start, ref, alt, family) %>%
    summarise(shared variants = n())
  return(tempDf)
}
# add family ID to df
add_family_ID <- function(df){</pre>
 fam1 <- filter(df, sample %in% c("4478_SP_0025", "4478_SP_0026",</pre>
                                   "4478_SP_0027", "4478_SP_0028", "4478_SP_0029", "25", "26", "27",
                                    "28", "29")) %>%
    mutate(family = "Family 1")
  "32", "33", "34", "35")) %>%
    mutate(family = "Family 2")
  fam3 <- filter(df, sample %in% c("4478_SP_0038", "4478_SP_0039",
                                    "4478_SP_0040", "4478_SP_0041",
```

```
"4478_SP_0042", "4478_SP_0043",

"4478_SP_0044", "38", "39", "40",

"41", "42", "43", "44")) %>%

mutate(family = "Family 3")

return(bind_rows(fam1, fam2, fam3))
}
```

#### **Variant Stats of Passing and Phased variants**

The passed\_variants\_summary.tsv, passed\_dels\_summary.tsv, and passed\_svs\_summary.tsv were generated with the keep\_passed\_variants.py script.

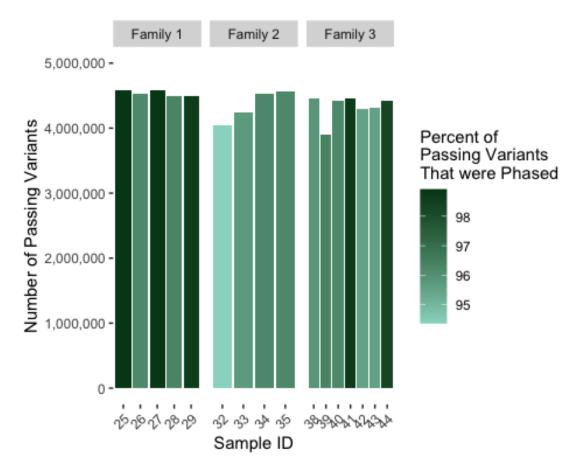
```
passed_variants <- read_tsv_custom_2("passed_variants_summary.tsv") %>%
  arrange(sample)
passed_dels <- read_tsv_custom_2("passed_dels_summary.tsv") %>%
arrange(sample)
passed svs <- read tsv custom 2("passed svs summary.tsv") %>% arrange(sample)
passed_combined <- bind_rows(passed_variants, passed_dels, passed_svs) %>%
  group by(sample) %>% summarise all(sum) %>%
  mutate(passed_percentage = (num_passed/total_variants) * 100,
           passed phased percentage = (num passed phased/num passed) * 100)
%>%
  add_disease_column()
write_tsv(passed_combined, "passed_variant_summary")
passed_tidy <- gather(passed_combined, category, "value", 2:6)</pre>
paste("Average number of total variants sequenced: ",
      mean(passed_combined$total_variants))
## [1] "Average number of total variants sequenced: 6058006.5"
paste("Average number of variants that passed quality filters: ",
      mean(passed combined$num passed))
## [1] "Average number of variants that passed quality filters: 4395811"
```

#### **Create figure of passing/phased numbers**

This code chunk was used to create Figure 2 in the manuscript.

```
passed_combined <- add_family_ID(passed_combined)

ggplot(passed_combined, aes(as.character(sample), num_passed)) +
    geom_col(aes(fill = passed_phased_percentage)) +</pre>
```



ggsave("Figure\_2.png", plot = last\_plot())

#### Read in CH data and separate by disease

```
# Read in CH data and generate new columns with variant type and class
combined_CH_data_vars <- read_tsv_custom("PedFam_CH_cadd20_maf01.tsv") %>%
   add_column(variant_type = "Compound Heterozygous", class = "SNP")

combined_CH_data_svs <- read_tsv_custom("PedFam_CH_cadd20_maf01_svs.tsv") %>%
   add_column(variant_type = "Compound Heterozygous", class = "SV")

combined_CH_data_dels <- read_tsv_custom("PedFam_CH_cadd20_maf01_dels.tsv")</pre>
```

```
%>%
  add column(variant type = "Compound Heterozygous", class = "indel")
# Combine all imported data into a single dataframe
combined_CH_all <- combine_df(list(combined_CH_data_vars,</pre>
                                    combined CH data svs,
combined_CH_data_dels))
# Filter down to astrocytoma data and filter to gene level
astrocytoma CH data <- astrocytoma sample filter(combined CH all)</pre>
astrocytoma CH gene level <- gene level filter(astrocytoma CH data,
"astrocytoma")
# Filter down to wilms data and filter to gene level
wilms_CH_data <- wilms_sample_filter(combined_CH_all)</pre>
wilms_CH_gene_level <- gene_level_filter(wilms_CH_data, "wilms")</pre>
# Filter down to Lymphoma data and filter to gene level
lymphoma_CH_data <- lymphoma_sample_filter(combined_CH_all)</pre>
lymphoma CH gene level <- gene level filter(lymphoma CH data, "lymphoma")</pre>
Read in de Novo data and separate by disease
# Read in de novo data and generate new columns with variant type and class
combined deNovo data vars <-
read_tsv_custom("PedFam_deNovo_cadd20_maf01.tsv") %>%
  add column(variant type = "De Novo", class = "SNP")
combined deNovo data svs <-
read tsv custom("PedFam deNovo cadd20 maf01 svs.tsv") %>%
  add column(variant type = "De Novo", class = "SV")
combined deNovo data dels <-
read tsv custom("PedFam deNovo cadd20 maf01 dels.tsv") %>%
  add column(variant type = "De Novo", class = "indel")
# Combine all imported data into a single dataframe
combined_deNovo_all <- combine_df(list(combined_deNovo_data_vars,</pre>
                                        combined deNovo data svs,
                                        combined deNovo data dels))
# Filter down to astrocytoma data and filter to gene level
astrocytoma deNovo data <- astrocytoma sample filter(combined deNovo all)</pre>
astrocytoma deNovo gene level <- gene level filter(astrocytoma deNovo data,
                                                     "astrocytoma")
# Filter down to wilms data and filter to gene level
wilms_deNovo_data <- wilms_sample_filter(combined_deNovo_all)</pre>
wilms deNovo gene level <- gene level filter(wilms deNovo data, "wilms")</pre>
```

```
# Filter down to Lymphoma data and filter to gene Level
lymphoma_deNovo_data <- lymphoma_sample_filter(combined_deNovo_all)
lymphoma_deNovo_gene_level <- gene_level_filter(lymphoma_deNovo_data,
"lymphoma")</pre>
```

#### Read in homozygous alternate data and separate by disease

```
# Read in homozygous alternate data and generate new columns with variant
type
# and class
combined_homAlt_data_vars <-</pre>
read tsv custom("PedFam homAlt cadd20 maf01.tsv") %>%
  add column(variant type = "Homozygous Alternate", class = "SNP")
combined homAlt data svs <-
read_tsv_custom("PedFam_homAlt_cadd20_maf01_svs.tsv") %>%
  add column(variant type = "Homozygous Alternate", class = "SV")
combined homAlt data dels <-
read_tsv_custom("PedFam homAlt cadd20 maf01 dels.tsv") %>%
  add_column(variant_type = "Homozygous Alternate", class = "indel")
# Combine all imported data into a single dataframe
combined homAlt all <- combine df(list(combined homAlt data vars,
                                        combined homAlt data svs.
                                        combined homAlt data dels))
# Filter down to astrocytoma data and filter to gene level
astrocytoma homAlt data <- astrocytoma sample filter(combined homAlt all)</pre>
astrocytoma_homAlt_gene_level <- gene_level_filter(astrocytoma_homAlt_data,</pre>
                                                     "astrocytoma")
# Filter down to wilms data and filter to gene level
wilms_homAlt_data <- wilms_sample_filter(combined_homAlt_all)</pre>
wilms homAlt gene level <- gene level filter(wilms homAlt data, "wilms")</pre>
# Filter down to Lymphoma data and filter to gene level
lymphoma homAlt data <- lymphoma sample filter(combined homAlt all)</pre>
lymphoma_homAlt_gene_level <- gene_level_filter(lymphoma_homAlt_data,</pre>
"lymphoma")
```

#### Read in heterozygous data and separate by disease

```
# Read in heterozygous data and generate new columns with variant type and
class
combined_het_data_vars <- read_tsv_custom("PedFam_het_cadd20_maf01.tsv") %>%
    add_column(variant_type = "Heterozygous", class = "SNP")

combined_het_data_svs <- read_tsv_custom("PedFam_het_cadd20_maf01_svs.tsv")
%>%
    add_column(variant_type = "Heterozygous", class = "SV")
```

```
combined_het_data_dels <- read_tsv_custom("PedFam_het_cadd20_maf01_dels.tsv")</pre>
  add column(variant type = "Heterozygous", class = "indel")
# Combine all imported data into a single dataframe
combined het all <- combine df(list(combined het data vars,
                                     combined_het_data_svs,
combined het data dels))
# Filter down to astrocytoma data and filter to gene level
astrocytoma het data <- astrocytoma sample filter(combined het all)</pre>
astrocytoma het gene level <- gene level filter(astrocytoma het data,
"astrocytoma")
# Filter down to wilms data and filter to gene level
wilms het data <- wilms sample filter(combined het all)</pre>
wilms_het_gene_level <- gene_level_filter(wilms_het_data, "wilms")</pre>
# Filter down to Lymphoma data and filter to gene Level
lymphoma het data <- lymphoma sample filter(combined het all)</pre>
lymphoma het gene level <- gene level filter(lymphoma het data, "lymphoma")</pre>
```

### Combine all data regardless of data type, and summarise at gene and variant levels

This code chunk was used to generate data for Table 1 in the manuscript and Supplementary Table 1.

```
#Combine all raw data
combined data all <- combine df(list(combined homAlt all,</pre>
combined_deNovo_all,
                                      combined_het_all, combined_CH_all))
write tsv(combined data all, "combined data all.tsv")
# Combine all data at gene level
gene level all <- combine df(list(astrocytoma CH gene level,
                                   astrocytoma deNovo gene level,
                                   astrocytoma_homAlt_gene_level,
                                   astrocytoma het gene level,
                                   lymphoma_CH_gene_level,
                                   lymphoma_deNovo_gene_level,
                                   lymphoma het_gene_level,
                                   lymphoma_homAlt_gene_level,
                                   wilms_CH_gene_level,
wilms deNovo gene level,
                                  wilms het gene level,
wilms homAlt gene level))
# Gene level summary (how many samples each potentially damaged gene has)
```

```
gene level summary all <- group by(gene level all, disease, gene) %>%
  summarise(num samples with damage = length(unique(sample)))
write tsv(gene level summary all, "gene level summary.tsv")
gene level summary all
## # A tibble: 84 x 3
## # Groups:
               disease [3]
##
      disease
                  gene
                           num samples with damage
##
      <chr>>
                                              <int>
                  <chr>>
## 1 astrocytoma ACADS
                                                  1
                                                  4
## 2 astrocytoma ACOXL
## 3 astrocytoma ALG1L2
                                                  1
## 4 astrocytoma BST1
                                                  5
                                                  3
## 5 astrocytoma CBWD5
## 6 astrocytoma CRNKL1
                                                  3
                                                  3
## 7 astrocytoma DAPL1
## 8 astrocytoma DUOXA1
                                                  3
                                                  2
## 9 astrocytoma FAM8A1
                                                  1
## 10 astrocytoma GOLGA6L1
## # ... with 74 more rows
# Gene and sample level summary (how many genes have x number of samples with
а
# variant in that gene)
shared_genes_summary <- group_by(gene_level_summary_all, disease,</pre>
                                  num_samples_with_damage) %>%
  summarise(genes_with_num_damaged_genes = length(unique(gene)))
shared_genes_summary
## # A tibble: 14 x 3
## # Groups: disease [3]
##
                  num_samples_with_damage genes_with_num_damaged_genes
      disease
##
      <chr>>
                                     <int>
                                                                   <int>
## 1 astrocytoma
                                                                      10
                                         1
                                         2
                                                                       5
## 2 astrocytoma
## 3 astrocytoma
                                         3
                                                                       6
## 4 astrocytoma
                                         4
                                                                       6
## 5 astrocytoma
                                         5
                                                                       3
                                         1
## 6 lymphoma
                                                                      14
## 7 lymphoma
                                         2
                                                                       8
## 8 lymphoma
                                         3
                                                                       4
                                         4
                                                                       4
## 9 lymphoma
## 10 lymphoma
                                         5
                                                                       2
                                         1
## 11 wilms
                                                                      11
## 12 wilms
                                         2
                                                                       5
## 13 wilms
                                         3
                                                                       4
## 14 wilms
                                         4
                                                                       2
```

```
# Find any shared variants
shared variants <- shared variant filter(add family ID(combined data all))
# Summarize shared variants
shared variants summary <- group by(shared variants, family, shared variants)
  summarise(total = n())
shared variants summary
## # A tibble: 14 x 3
## # Groups: family [3]
     family shared_variants total
##
     <chr>
                       <int> <int>
## 1 Family 1
                            1
                                 12
## 2 Family 1
                                  5
                            2
## 3 Family 1
                            3
                                  7
## 4 Family 1
                            4
                                  6
                            5
## 5 Family 1
                                 3
                            1
                                 12
## 6 Family 2
## 7 Family 2
                           2
                                 5
## 8 Family 2
                            3
                                  4
## 9 Family 2
                            4
                                  2
## 10 Family 3
                            1
                                 15
                            2
                                  8
## 11 Family 3
## 12 Family 3
                            3
                                  5
                            4
                                  5
## 13 Family 3
## 14 Family 3
                                  1
```

### Use python to retain pertinent information from the 'combined\_data\_all.tsv' file and combine the chromosome and position columns

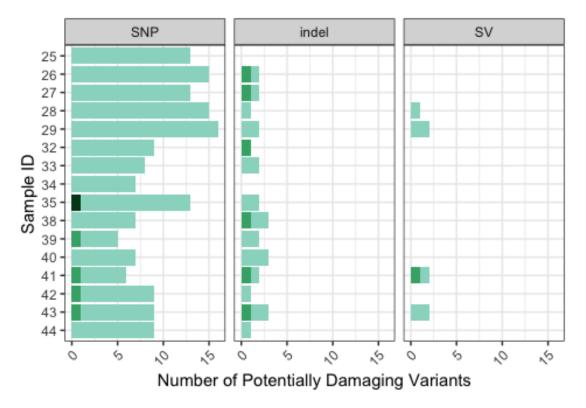
```
sample dict = {}
with open('combined_data_all.tsv') as input_file:
  header = input_file.readline()
  header_list = header.rstrip("\n").split("\t")
  variant_type_index = header_list.index("variant_type")
  sample_index = header_list.index("sample")
  start index = header list.index("start")
  class_index = header_list.index("class")
  chrom_index = header_list.index("chrom")
  for line in input_file:
    line_list = line.rstrip("\n").split("\t")
    variant_type = line_list[variant_type_index]
    if variant type == "Heterozygous":
      variant_type = "Simple Heterozygous"
    sample = line list[sample index]
    start = line_list[start_index]
    chrom = line_list[chrom_index]
```

```
variant class = line list[class index]
    pos = chrom + ": " + start
    # Summarize variant class and type for each variant for each sample
    if sample not in sample dict:
      sample_dict[sample] = {pos: {variant_class: [variant_type]}}
    elif sample in sample_dict and pos not in sample_dict[sample]:
      sample dict[sample][pos] = {variant class :[variant type]}
    elif sample in sample_dict and pos in sample_dict[sample] \
      and variant_class not in sample_dict[sample][pos]:
      sample dict[sample][pos][variant class] = [variant type]
    elif sample in sample dict and pos in sample dict[sample] \
      and variant class in sample dict[sample][pos] and variant type \
      not in sample_dict[sample][pos][variant_class]:
      sample_dict[sample][pos][variant_class].append(variant_type)
with open('combined_data_updated.tsv', 'wt') as output_file:
  output_file.write("sample\tpos\tclass\tvariant_type\n")
  for sample, pos_dict in sample_dict.items():
    for pos, variant class dict in pos dict.items():
      for variant class, variant list in variant class dict.items():
        variant list.sort()
        # Retains the variant type, removes "de novo" from the type if found
        variant_string = variant_list[-1]
        # Retains "compound heterozygous" only, does not include more
specific
        # variant types contributing to the compound heterozygous variant
        if "Compound Heterozygous" in variant_string:
          variant_string = variant_list[0]
        output_file.write("{}\t{}\t{}\n".format(sample, pos,
variant class, variant string))
```

### Generate a Figure showing a variant level summary that shows variant type and class

This code chunk was used to generate Figure 3 for the manuscript.

```
levels = c("SNP", "indel", "SV"))
variant level summary all$variant type <-
factor(variant_level_summary_all$variant_type, levels = c("Simple")
Heterozygous", "Homozygous Alternate", "Compound Heterozygous"))
# Generate figure
ggplot(variant_level_summary_all, aes(variants, sample, variant_type, class))
  geom col(aes(variants, sample, fill = variant type)) +
  facet_wrap(~class, scales = "fixed") +
  theme_bw() +
  theme(panel.background = element_rect(fill= "white"),
        axis.text.y = element_text(hjust = 1),
        axis.text.x = element_text(angle = 45, hjust = 1), legend.position =
"bottom") +
  labs(y="Sample ID", x="Number of Potentially Damaging Variants",
       fill="Variant Type") +
  scale_fill_manual(values = rev(c("#00441b", "#41ae76", "#99d8c9"))) +
  scale_y_discrete(limits = rev)
```



riant Type Simple Heterozygous Homozygous Alternate Compound He

```
# Save figure
ggsave("Figure_3.png", plot = last_plot())
```

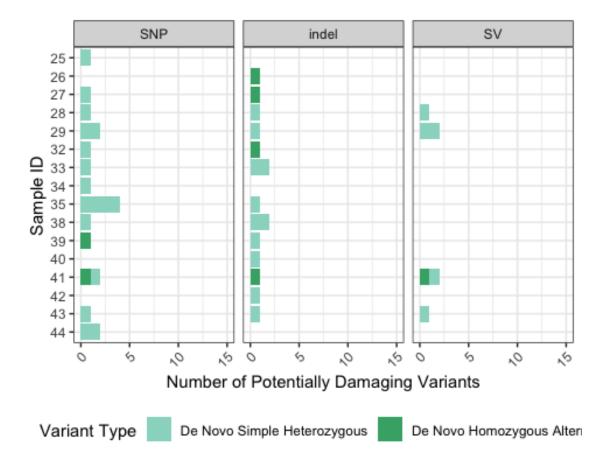
### Use python to retain de novo variants from the 'combined\_data\_all.tsv' file and combine the chromosome and position columns

```
sample dict = {}
with open('combined data all.tsv') as input file:
  header = input file.readline()
  header_list = header.rstrip("\n").split("\t")
  variant type index = header list.index("variant type")
  sample index = header list.index("sample")
  start_index = header_list.index("start")
  class_index = header_list.index("class")
  chrom_index = header_list.index("chrom")
  for line in input file:
    line_list = line.rstrip("\n").split("\t")
    variant_type = line_list[variant_type_index]
    if variant_type == "Heterozygous":
      variant_type = "Simple Heterozygous"
    sample = line_list[sample_index]
    start = line list[start index]
    chrom = line list[chrom index]
    variant class = line list[class index]
    pos = chrom + ": " + start
    # Summarize variant class and type for each variant for each sample
    if sample not in sample_dict:
      sample dict[sample] = {pos: {variant class: [variant type]}}
    elif sample in sample dict and pos not in sample dict[sample]:
      sample_dict[sample][pos] = {variant_class :[variant_type]}
    elif sample in sample dict and pos in sample dict[sample] \
      and variant class not in sample dict[sample][pos]:
      sample_dict[sample][pos][variant_class] = [variant_type]
    elif sample in sample dict and pos in sample dict[sample] \
      and variant class in sample dict[sample][pos] and variant type \
      not in sample_dict[sample][pos][variant_class]:
      sample dict[sample][pos][variant class].append(variant type)
with open('combined data de novo updated.tsv', 'wt') as output file:
  output file.write("sample\tpos\tclass\tvariant type\n")
  for sample, pos_dict in sample_dict.items():
    for pos, variant class dict in pos dict.items():
      for variant class, variant list in variant class dict.items():
        variant_list.sort()
        variant_string = " ".join(variant_list)
        if "Compound Heterozygous" in variant string:
          variant_string = variant_string.replace("Compound Heterozygous",
        # outputs de novo variants. For compound heterozygous variants, only
the
        # variant types that contribute to the compound heterozygous variant
```

## Generate a Figure showing a variant level summary that shows de novo variant type and class

This code chunk was used to generate Figure 4 for the manuscript.

```
# Update sample names and add disease column
combined data de novo updated <-
read tsv("combined data de novo updated.tsv")
combined_data_de_novo_updated <- mutate(combined_data_de_novo_updated,
                                sample = str replace(sample, "4478 SP 00",
""))
combined_data_de_novo_updated <-</pre>
add disease column(combined data de novo updated)
# de novo variant level summary (how many of each type of variant each sample
variant_level_summary_de_novo <-</pre>
variant level filter 2(combined data de novo updated)
variant level summary de novo <-
add_disease_column(variant_level_summary_de_novo)
# Change order that classes appear in graph
variant level summary de novo$class <-
factor(variant level summary de novo$class, levels = c("SNP", "indel", "SV"))
variant_level_summary_de_novo$variant_type <-</pre>
factor(variant level summary de novo$variant type, levels = c("De Novo Simple
Heterozygous", "De Novo Homozygous Alternate"))
# Generate figure
ggplot(variant level summary de novo, aes(variants, sample, variant type,
class)) +
  geom col(aes(variants, sample, fill = variant type)) +
  facet_wrap(~class, scales = "fixed") +
  theme bw() +
  theme(panel.background = element rect(fill= "white"),
        axis.text.y = element text(hjust = 1),
        axis.text.x = element_text(angle = 45, hjust = 1), legend.position =
"bottom") +
  labs(y="Sample ID", x="Number of Potentially Damaging Variants",
       fill="Variant Type") +
  scale_fill_manual(values = rev(c("#41ae76", "#99d8c9"))) +
  scale y discrete(limits = rev) +
  scale_x_continuous(limits = c(0, 15))
```



```
# Save figure
ggsave("Figure_4.png", plot = last_plot())
```

### Find genes that have 2 or less siblings with a variant

This code chunk provided a list of genes for each family that was used as input into *VarElect*.

```
# Lymphoma data
paste("lymphoma: ", str_c(filter(gene_level_summary_all, disease ==
"lymphoma" &
                                    num samples with damage <= 2)$gene,
                           collapse = ","))
## [1] "lymphoma:
ADGRA2, Clorf50, C2orf16, CCDC179, CSNK1A1, DCHS1, DNTTIP2, ENPEP, GLYATL1, ITGB4, KRTA
P10-12, LILRA2, RGPD3, RNU7-
167P, SIRPB1, SPAG11B, TNNT3, TRMT1, USP17L15, Y RNA, ZNF826P, ZNF99"
paste("Number of genes to input to VarEelct for lymphoma: ",
      length(filter(gene_level_summary_all, disease == "lymphoma" &
                       num_samples_with_damage <= 2)$gene))</pre>
## [1] "Number of genes to input to VarEelct for lymphoma: 22"
# Wilms data
paste("wilms: ", str c(filter(gene level summary all, disease == "wilms" &
                                 num_samples_with_damage <= 2)$gene,</pre>
                        collapse = ","))
## [1] "wilms: ANKRD36C,CYP2A13,DPY19L3,FAM8A1,GOSR2,HLA-DRB1,HLA-
DRB5, PAH, PCK1, PCK2, PNPLA7, RP1L1, SIPA1L3, TRPM3, Y RNA, ZAN"
paste("Number of genes to input to VarEelct for wilms: ",
      length(filter(gene_level_summary_all, disease == "wilms" &
                       num samples with damage <= 2)$gene))</pre>
## [1] "Number of genes to input to VarEelct for wilms: 16"
```

### Figure how how many unique genes there were before and after sibling filtering

#### Create a vector of known cancer genes using COSMIC

```
cosmic <- read_tsv("known_genes/Census_allThu Apr 16 21_44_49 2020.tsv")
cosmic_all <- cosmic$`Gene Symbol`</pre>
```

# Check to see if any of the children had a potentially damaing variant in a known cancer gene

```
cancer_genes <- c()
for(gene in combined_data_all$gene){
  if (gene %in% cosmic_all & !gene %in% cancer_genes) {
    cancer_genes <- c(cancer_genes, gene)
  }
}
paste("The following genes are known cancer genes and were identified in at least on child across all families: ",
    str_c(cancer_genes, collapse = ", "))
### [1] "The following genes are known cancer genes and were identified in at least on child across all families: RGPD3, CRNKL1"</pre>
```