GTO 2: The genomics-proteomics toolkit

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Preface

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¹https://rmarkdown.rstudio.com

²https://bookdown.org

Introduction

Recent advances in DNA sequencing, specifically in next-generation sequencing (NGS), revolutionised the field of genomics, making possible the generation of large amounts of sequencing data very rapidly and at substantially low cost(Mardis, 2017). This new technology also brought with it several challenges, namely in what concerns the analysis, storage, and transmission of the generated sequences(Brouwer et al., 2016, Liu et al. (2012)). As a consequence, several specialised tools were developed throughout the years in order to deal with these challenges.

Firstly, the storage of the raw data generated by NGS experiments is possible by using several file formats, the FASTQ and FASTA are the most commonly used(Zhang, 2016). FASTQ is an extension of the FASTA format, that besides the nucleotide sequence, also stores associated per base quality score and it is considered the standard format for sequencing data storage and exchange(Cock et al., 2009).

Regarding the analysis and manipulation of these sequencing data files many software applications emerged, including fqtools(Droop, 2016), FASTX-Toolkit(Gordon et al., 2010), GALAXY(Afgan et al., 2018), GATK(DePristo et al., 2011), MEGA(Kumar et al., 2016), SeqKit(Shen et al., 2016), among others. Fqtools is a suite of tools to view, manipulate and summarise FASTQ data. This software also identifies invalid FASTQ files(Droop, 2016). GALAXY, in its turn, is an open, web-based scientific platform for analysing genomic data(Goecks et al., 2010). This platform integrates several specialised sets of tools, e.g. for manipulating FASTQ files(Blankenberg et al., 2010). FASTX-Toolkit is a collection of command-line tools to process FASTA and FASTQ files. This toolkit is available in two forms: as a command-line, or integrated into the web-based platform GALAXY (Gordon et al., 2010). SeqKit is another toolkit used to process FASTA and FASTQ files and is available for all major operating systems (Shen et al., 2016). The Genome Analysis Toolkit (GATK) was designed as a structured programming framework

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to simplify the development of analysis tools. However, nowadays, it is a suite of tools focused on variant discovering and genotyping (Van der Auwera et al., 2013). More towards the evolutionary perspectives, Molecular Evolutionary Genetics Analysis (MEGA) software provides tools to analyse DNA and protein sequences statistically (Tamura et al., 2011). Several of these frameworks lack on variety, namely the ability to perform multiple tasks using only one toolkit.

Compression is another important aspect when dealing with high-throughput sequencing data, as it reduces storage space and accelerates data transmission. A survey on DNA compressors and amino acid sequence compression can be found in (Hosseini et al., 2016). Currently, the DNA sequence compressors HiRGC(Liu et al., 2017), iDoComp(Ochoa et al., 2014), GeCo(Pratas et al., 2016), and GDC(Deorowicz et al., 2015) are considered to have the best performance (Hernaez et al., 2019). Of these four approaches, GeCo is the only one that can be used for reference-free and reference-based compression. Furthermore, GeCo can be used as an analysis tool to determine absolute measures for many distance computations and local measures (Pratas et al., 2016).

Amino acid sequences are known to be very hard to compress(Nalbantoglu et al., 2010), however, Hosseini et al.(Hosseini et al., 2019) recently developed AC, a state-of-the-art for lossless amino acid sequence compression. In(Pratas et al., 2018) the authors compared the performance of AC, in terms of bit-rate, to several general-purpose lossless compressors and several protein compressors, using different proteomes. They concluded that in average AC provides the best bit-rates.

Another relevant subject is genomic data simulation. Read simulations tools are fundamental for the development, testing and evaluation of methods and computational tools(Huang et al., 2011, price2017simulome). Despite the availability of a large number of real sequence reads, read simulation data is necessary due to the inability to know the ground truth of real data(Baruzzo et al., 2017). Escalona et al. (Escalona et al., 2016), recently, reviewed 23 NGS simulation tools. XS(Pratas et al., 2014), a FASTQ read simulation tool, stands out in relation to the other 22 simulation tools because it is the only one that does not need a reference sequence. Furthermore, XS is the only opensource tool for simulation of FASTQ reads produced by the four most

1.1 Installation 3

used sequencing machines, Roche-454, Illumina, ABI SOLiD and Ion Torrent.

Although a large number of tools are available for analysing, compressing, and simulation, these tools are specialised in only a specific task. Besides, in many cases the output of one tool cannot be used directly as input for another tool, e.g. the output of a simulation tool cannot always be used directly as input for an analysis tool. Thus, unique software that includes several specialised tools is necessary.

In this document, we describe **GTO2**, a complete toolkit for genomics and proteomics, namely for FASTQ, FASTA and SEQ formats, with many complementary tools. The toolkit is for Unix-based systems, built for ultra-fast computations. **GTO2** supports pipes for easy integration with the sub-programs belonging to **GTO2** as well as external tools. **GTO2** works as **LEGOs**, since it allows the construction of multiple pipelines with many combinations.

GTO2 includes tools for information display, randomisation, edition, conversion, extraction, search, calculation, compression, simulation and visualisation. GTO2 is prepared to deal with very large datasets, typically in the scale of Gigabytes or Terabytes (but not limited). The complete toolkit is an optimised command-line version, using the prefix gto2_ followed by the suffix with the respective name of the program. GTO2 is implemented in C language and it is available, under the MIT license, at https://github.com/cobilab/gto2

1.1 Installation

To install **GTO2** through the GitHub repository:

```
git clone https://github.com/cobilab/gto2.git
cd gto2/src/
make
```

Or by installing them directly using the Cobilab channel from Conda:

```
conda install -c cobilab gto2 -y
```

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1.2 Testing

The examples provided in this document are available in the repository. Therefore, each example can be easily reproduced, which it will also test and validate each tool. To replicate those tests, it can be done in two different ways:

- Running one test for a specific tool:
 - cd gto2/tester/gto2_{tool}
 - sh runExample.sh
- Running the batch of tests for all the tools:
 - cd gto2/tester/
 - sh runAllTests.sh

Some of these tests require internet connection to download external files and it will create new files.

1.3 Execution control

The quality control in Unix/Linux pipelines using GTO's tools is made in three ways:

- Input verification: where the tools verify the format of the input file;
- Stderr logs: Some execution errors are directly sent for the stderr channel.
- Scripting validation: In complex pipelines, the verification of all the tools in the pipeline were executed properly, it is used the PIPESTA-TUS variable, e.g.:

```
gto2_fa_rand_extra_chars < input.fa | \
gto2_fa_to_seq > output.seq
echo "${PIPESTATUS[0]} ${PIPESTATUS[1]}"
0 0
```

2

FASTA Tools

 $2.1 \quad gto2_fa_to_fq$

to do

FASTQ Tools

The toolkit has a set of tools dedicated to manipulating FASTQ files. Some of these tools allow the data conversion to/from different formats, i. e., there are tools designed to convert a FASTQ file into a sequence or a FASTA/Multi-FASTA format, or converting DNA in some of those formats to FASTQ.

There are also tools for data manipulation in this format, which are designed to exclude 'N', remove low quality scored reads, following different metrics and randomize DNA sequences. Succeeding the manipulation, it is also possible to perform analyses over these files, simulations and mutations. The current available tools for FASTQ format analysis and manipulation include:

- gto2_fq_to_fa: to convert a FASTQ file format to a pseudo FASTA file.
- gto2_fq_to_mfa: to convert a FASTQ file format to a pseudo Multi-FASTA file.
- **gto2_fq_exclude_n**: to discard the FASTQ reads with the minimum number of "N" symbols.
- gto2_fq_extract_quality_scores: to extract all the quality-scores from FASTQ reads.
- gto2_fq_info: to analyse the basic information of FASTQ file format.
- gto2_fq_maximum_read_size: to filter the FASTQ reads with the length higher than the value defined.
- gto2_fq_minimum_quality_score: to discard reads with average quality-score below of the defined.
- gto2_fq_minimum_read_size: to filter the FASTQ reads with the length smaller than the value defined.
- gto2_fq_rand_extra_chars: to substitue in the FASTQ files, the DNA sequence the outside ACGT chars by random ACGT symbols.

- gto2_fq_from_seq: to convert a genomic sequence to pseudo FASTQ file format.
- **gto2_fq_mutate**: to create a synthetic mutation of a FASTQ file given specific rates of mutations, deletions and additions.
- **gto2_fq_split**: to split Paired End files according to the direction of the strand ('/1' or '/2').
- gto2 fq pack: to package each FASTQ read in a single line.
- gto2_fq_unpack: to unpack the FASTQ reads packaged using the gto2_fq_pack tool.
- gto2_fq_quality_score_info: to analyse the quality-scores of a FASTQ file.
- gto2_fq_quality_score_min: to analyse the minimal quality-scores of a FASTQ file.
- gto2_fq_quality_score_max: to analyse the maximal quality-scores of a FASTQ file.
- gto2 fq cut: to cut read sequences in a FASTQ file.
- gto2_fq_minimum_local_quality_score_forward: to filter the reads considering the quality score average of a defined window size of bases.
- gto2_fq_minimum_local_quality_score_reverse: to filter the reverse reads, considering the average window size score defined by the bases.
- gto2_fq_xs: a skilled FASTQ read simulation tool, flexible, portable and tunable in terms of sequence complexity.
- gto2_fq_complement: to replace the ACGT bases with their complements in a FASTQ file format.
- **gto2_fq_reverse**: to reverse the ACGT bases order for each read in a FASTQ file format.
- gto2_fq_variation_map: to identify the variation that occours in the sequences relative to the reads or a set of reads.
- gto2_fq_variation_filter: to filter and segments the regions of singularity from the output of gto2_fq_variation_map.
- **gto2**_**fq**_**variation**_**visual**: to depict the regions of singularity using the output from **gto2 fq variation filter** into an SVG image.
- gto2_fq_metagenomics: to measure the similarity between any FASTQ file, independently from the size, against any multi-FASTA database.

3.1 Program gto2_fq_to_fa

The **gto2_fq_to_fa** converts a FASTQ file format to a pseudo FASTA file. However, this tool does not align the sequence, instead, it extracts the sequence and adds a pseudo-header.

For help type:

```
./gto2_fq_to_fa -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The **gto2_fq_to_fa** program needs two streams for the computation, namely the input and output standard. The input stream is a FASTQ file.

The attribution is given according to:

An example of such an input file is:

@SRR001666.1 071112_SLXA-EAS1_s_7:5:1:817:345 length=60
GGGTGATGGCCGCTGCCGATGGCGTCAAATCCCACCAAGTTACCCTTAACAACTTAAGGG

Output

The output of the **gto2_fq_to_fa** program is a FASTA file. Using the input above, an output example of this is the following:

> Computed with Fastq2Fasta
GGGTGATGGCCGCTGCCGATGGCGTCAAATCCCACCAAGTTACCCTTAACAACTTAAGGG
GTTCAGGGATACGACGTTTGTATTTTAAGAATCTGAAGCAGAAGTCGATGATAATACGCG

3.2 Program gto2_fq_to_mfa

The **gto2_fq_to_mfa** converts a FASTQ file format to a pseudo Multi-FASTA file. However, this tool does not align the sequence, instead, it extracts the sequence and adds a pseudo header.

For help type:

```
./gto2_fq_to_mfa -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The gto2_fq_to_mfa program needs two streams for the computation, namely the input and output standard. The input stream is a FASTQ file.

An example of such an input file is:

Output

The output of the **gto2_fq_to_mfa** program is a Multi-FASTA file. Using the input above, an output example of this is the following:

>SRR001666.1 071112_SLXA-EAS1_s_7:5:1:817:345 length=60 GGGTGATGGCCGCTGCCGATGGCGTCAAATCCCACCAAGTTACCCTTAACAACTTAAGGG >SRR001666.2 071112_SLXA-EAS1_s_7:5:1:801:338 length=60 GTTCAGGGATACGACGTTTGTATTTTAAGAATCTGAAGCAGAAGTCGATGATAATACGCG

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3.3 Program gto2_fq_exclude_n

The **gto2_fq_exclude_n** discards the FASTQ reads with the minimum number of "N" symbols, and it will erase the second header (after +), if presented.

For help type:

```
./gto2_fq_exclude_n -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The gto2_fq_exclude_n program needs two streams for the computation, namely the input and output standard. The input stream is a FASTQ file.

```
Usage: ./gto2_fq_exclude_n [options] [[--] args]
   or: ./gto2_fq_exclude_n [options]
It discards the FASTQ reads with the minimum number of "\mathbb{N}"
symbols.
If present, it will erase the second header (after +).
    -h, --help
                            show this help message and exit
Basic options
    -m, --max = < int >
                            The maximum of of "\mathbb{N}" symbols in
                            the read
    < input.fastq
                            Input FASTQ file format (stdin)
    > output.fastq
                           Output FASTQ file format (stdout)
Example: ./gto2_fq_exclude_n -m <max> < input.fastq >
output.fastq
```

```
Console output example :
<FASTQ non-filtered reads>
Total reads : value
Filtered reads : value
```

An example of such an input file is:

Output

The output of the **gto2_fq_exclude_n** program is a set of all the filtered FASTQ reads, followed by the execution report. The execution report only appears in the console.

Using the input above with the max value as 5, an output example for this is the following:

3.4 Program gto2_fq_extract_quality_scores

The gto2_fq_extract_quality_scores extracts all the quality-scores from FASTQ reads.

For help type:

```
./gto2_fq_extract_quality_scores -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The gto2_fq_extract_quality_scores program needs two streams for the computation, namely the input and output standard. The input stream is a FASTQ file.

The attribution is given according to:

```
Usage: ./gto2_fq_extract_quality_scores [options] [[--]args]
    or: ./gto2_fq_extract_quality_scores [options]
```

It extracts all the quality-scores from FASTQ reads.

```
-h, --help show this help message and exit
```

Basic options

```
Example: ./gto2_fq_extract_quality_scores < input.fastq >
output.fastq
```

Console output example:
<FASTQ quality scores>

Total reads : value Total Quality-Scores : value

An example of such an input file is:

Output

The output of the **gto2_fq_extract_quality_scores** program is a set of all the quality scores from the FASTQ reads, followed by the execution report. The execution report only appears in the console. Using the input above, an output example of this is the following:

3.5 Program gto2_fq_info

The **gto2_fq_info** analyses the basic information of FASTQ file format.

For help type:

```
./gto2_fq_info -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The **gto2_fq_info** program needs two streams for the computation, namely the input and output standard. The input stream is a FASTQ file.

```
Usage: ./gto2_fq_info [options] [[--] args]
   or: ./gto2_fq_info [options]
It analyses the basic information of FASTQ file format.
    -h, --help
                          show this help message and exit
Basic options
    < input.fastq
                          Input FASTQ file format (stdin)
    > output
                          Output read information (stdout)
Example: ./gto2_fq_info < input.fastq > output
Output example:
Total reads
                : value
Max read length: value
Min read length: value
Min QS value
                : value
Max QS value
                : value
```

An example of such an input file is:

: value

Output

QS range

The output of the **gto2_fq_info** program is a set of information related to the file read. Using the input above, an output example of this is the following:

```
Total reads : 2
Max read length : 72
```

```
Min read length: 72
Min QS value: 41
Max QS value: 73
QS range: 33
```

3.6 Program gto2_fq_maximum_read_size

The gto2_fq_maximum_read_size filters the FASTQ reads with the length higher than the value defined.

For help type:

```
./gto2_fq_maximum_read_size -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The gto2_fq_maximum_read_size program needs two streams for the computation, namely the input and output standard. The input stream is a FASTQ file.

```
> output.fastq Output FASTQ file format (stdout)
Example: ./gto2_fq_maximum_read_size -s <size> < input.fastq
> output.fastq
Console output example :
<FASTQ non-filtered reads>
Total reads : value
Filtered reads : value
```

An example of such an input file is:

Output

The output of the **gto2_fq_maximum_read_size** program is a set of all the filtered FASTQ reads, followed by the execution report. The execution report only appears in the console.

Using the input above with the size values as 59, an output example for this is the following:

3.7 Program gto2_fq_minimum_quality_score

The **gto2_fq_minimum_quality_score** discards reads with average quality-score below of the defined.

For help type:

```
./gto2_fq_minimum_quality_score -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The gto2_fq_minimum_quality_score program needs two streams for the computation, namely the input and output standard. The input stream is a FASTQ file.

```
Usage: ./gto2 fq minimum quality score [options] [[--] args]
   or: ./gto2_fq_minimum_quality_score [options]
It discards reads with average quality-score below value.
    -h, --help
                          show this help message and exit
Basic options
    -m, --min=<int>
                          The minimum average quality-score
                           (Value 25 or 30 is commonly used)
    < input.fastq</pre>
                           Input FASTQ file format (stdin)
    > output.fastq
                          Output FASTQ file format (stdout)
Example: ./gto2 fq minimum quality score -m <min> <
input.fastq > output.fastq
Console output example:
<FASTQ non-filtered reads>
```

Total reads : value Filtered reads : value

An example of such an input file is:

Output

The output of the **gto2_fq_minimum_quality_score** program is a set of all the filtered FASTQ reads, followed by the execution report. Using the input above with the minimum averge value as 30, an output example of this is the following:

@SRR001666.1 071112_SLXA-EAS1_s_7:5:1:817:345 length=60 GGGTGATGGCCGCTGCCGATGGCGTCAAATCCCACCAAGTTACCCTTAACAACTTAAGGG +

Total reads : 2 Filtered reads : 1

3.8 Program gto2_fq_minimum_read_size

The gto2_fq_minimum_read_size filters the FASTQ reads with the length smaller than the value defined.

For help type:

```
./gto2_fq_minimum_read_size -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The gto2_fq_minimum_read_size program needs two streams for the computation, namely the input and output standard. The input stream is a FASTQ file.

The attribution is given according to:

```
Usage: ./gto2_fq_minimum_read_size [options] [[--] args]
   or: ./gto2 fq minimum read size [options]
It filters the FASTQ reads with the length smaller than the
value defined.
If present, it will erase the second header (after +).
    -h, --help
                          show this help message and exit
Basic options
    -s, --size=<int>
                          The minimum read length
                          Input FASTQ file format (stdin)
    < input.fastq</pre>
                          Output FASTQ file format (stdout)
    > output.fastq
Example: ./gto2 fq minimum read size -s <size> < input.fastq
> output.fastq
Console output example:
<FASTQ non-filtered reads>
Total reads
               : value
Filtered reads : value
```

An example of such an input file is:

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Output

The output of the **gto2_fq_minimum_read_size** program is a set of all the filtered FASTQ reads, followed by the execution report. The execution report only appears in the console. Using the input above with the size values as 55, an output example of this is the following:

Total reads : 2 Filtered reads : 1

3.9 Program gto2_fq_rand_extra_chars

The gto2_fq_rand_extra_chars substitutes the outside ACGT chars by random ACGT symbols in the DNA sequence of FASTQ files.

For help type:

```
./gto2_fq_rand_extra_chars -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The **gto2_fq_rand_extra_chars** program needs two streams for the computation, namely the input and output standard. The input stream is a FASTQ file.

An example of such an input file is:

Output

The output of the **gto2_fq_rand_extra_chars** program is a FASTQ file. Using the input above, an output example of this is the following: @SRR001666.1 071112_SLXA-EAS1_s_7:5:1:817:345 length=60 GTGTGATGGCCGCTGCCGATGGCGCATAATCCCACCAACATACCCTTAACAACTTAAGGG

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3.10 Program gto2_fq_from_seq

The ${\tt gto2_fq_from_seq}$ converts a genomic sequence to pseudo FASTQ file format.

For help type:

```
./gto2_fq_from_seq -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The gto2_fq_from_seq program needs two streams for the computation, namely the input and output standard. The input stream is a sequence group file.

The attribution is given according to:

```
Usage: ./gto2_fq_from_seq [options] [[--] args]
   or: ./gto2_fq_from_seq [options]
It converts a genomic sequence to pseudo FASTQ file format.
    -h, --help
                          show this help message and exit
Basic options
    < input.seq
                          Input sequence file (stdin)
                          Output FASTQ file format (stdout)
    > output.fastq
Optional options
    -n, --name=<str>
                          The read's header
    -1, --lineSize=<int> The maximum of chars for line
Example: ./gto2 fq from seq -l <lineSize> -n <name> <
input.seq > output.fastq
```

An example of such an input file is:

ACAAGACGGCCTCCTGCTGCTGCTCCTCCGGGGCCACGGCCCTGGAGGGTCCACCGCT GCCCTGCTGCCATTGTCCCCGGCCCCACCTAAGGAAAAGCAGCCTCCTGACTTTCCTCGC TTGGGCCGAGACAGCGAGCATATGCAGGAAGCGGCAGGAAGTGGTTTGAGTGGACCTCCG GGCCCCTCATAGGAGAGGAAGCTCGGGAGGTGGCCAGGCGGCAGGAAGCAGGCCAGTGCC GCGAATCCGCGCGCCGGGACAGAATCTCCTGCAAAGCCCTGCAGGAACTTCTTCTGGAAG

Output

The output of the **gto2_fq_from_seq** program is a pseudo FASTQ file. An example, using the size line as 60 and the read's header as "SeqToFastq", for the input, is:

@SeqToFastq2

GCCCTGCTGCCATTGTCCCCGGCCCCACCTAAGGAAAAGCAGCCTCCTGACTTTCCTCGC

 ${\tt TTGGGCCGAGACAGCGAGCATATGCAGGAAGCGGCAGGAAGTGGTTTGAGTGGACCTCCG} + \\$

 $\tt GGCCCTCATAGGAGAGGAAGCTCGGGAGGTGGCCAGGCGGCAGGAAGCAGGCCAGTGCC$

 $\tt GCGAATCCGCGCGCGGGACAGAATCTCCTGCAAAGCCCTGCAGGAACTTCTTCTGGAAG$

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3.11 Program gto2_fq_mutate

The gto2_fq_mutate creates a synthetic mutation of a FASTQ file given specific rates of mutations, deletions and additions. All these parameters are defined by the user, and their are optional.

For help type:

```
./gto2_fq_mutate -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The gto2_fq_mutate program needs two streams for the computation, namely the input and output standard. However, optional settings can be supplied too, such as the starting point to the random generator, and the edition, deletion and insertion rates. Also, the user can choose to use the ACGTN alphabet in the synthetic mutation. The input stream is a FASTQ File.

```
Usage: ./gto2_fq_mutate [options]
                                  [[--] args]
   or: ./gto2_fq_mutate [options]
Creates a synthetic mutation of a FASTQ file given specific
rates of mutations, deletions and additions.
    -h, --help
                    show this help message and exit
Basic options
                    Input FASTQ file format (stdin)
    < input.fasta
    > output.fasta
                    Output FASTQ file format (stdout)
Optional
    -s
                    Starting point to the random generator
                    Defines the mutation rate (default 0.0)
    -m
```

```
-d Defines the deletion rate (default 0.0)
-i Defines the insertion rate (default 0.0)
-a When active, the application uses the ACGTN alphabet

Example: ./gto2_fq_mutate -s <seed> -m <mutation rate>
-d <deletion rate> -i <insertion rate> -a
< input.fastq > output.fastq
```

Output

The output of the **gto2_fq_mutate** program is a FASTQ file whith the synthetic mutation of input file. Using the input above with the seed value as 1 and the mutation rate as 0.5, an output example of this is the following:

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3.12 Program gto2_fq_split

The $\mathbf{gto2}_{\mathbf{fq}}$ split splits Paired End files according to the direction of the strand ('/1' or '/2'). It writes by default singleton reads as forward stands.

For help type:

```
./gto2_fq_split -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The gto2_fq_split program needs a stream for the computation, namely the input standard. The input stream is a FASTQ file.

The attribution is given according to:

```
Usage: ./gto2_fq_split [options] [[--] args]
   or: ./gto2_fq_split [options]
It writes by default singleton reads as forward stands.
    -h, --help
                          show this help message and exit
Basic options
    -f, --forward=<str>
                          Output forward file
    -r, --reverse=<str>
                          Output reverse file
    < input.fastq
                          Input FASTQ file format (stdin)
    > output
                          Output read information (stdout)
Example: ./gto2_fq_split -f <output_forward.fastq>
-r <output_reverse.fastq> < input.fastq > output
Output example :
Total reads
                 : value
Singleton reads
                : value
```

```
Forward reads : value
Reverse reads : value
```

Output

The output of the **gto2_fq_split** program is a set of information related to the file read. Using the input above, an output example of this is the following:

```
Total reads : 2
Singleton reads : 0
Forward reads : 65536
Reverse reads : 1
```

Also, this program generates two FASTQ files, with the reverse and forward reads.

An example of the forward reads, for the input, is:

30 3 FASTQ Tools

3.13 Program gto2_fq_pack

The **gto2_fq_pack** packages each FASTQ read in a single line. It can show the read score first or the dna sequence, depending on the execution mode.

For help type:

```
./gto2_fq_pack -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The gto2_fq_pack program needs two streams for the computation, namely the input and output standard. The input stream is a FASTQ file.

The attribution is given according to:

```
Usage: ./gto2_fq_pack [options] [[--] args]
   or: ./gto2_fq_pack [options]
It packages each FASTQ read in a single line.
    -h, --help
                          show this help message and exit
Basic options
    < input.fastq
                          Input FASTQ file format (stdin)
    > output.fastqpack
                          Output packaged FASTQ file format
                          (stdout)
Optional
    -s, --scores
                          When active, the application show
                          the scores first
Example: ./gto2_fq_pack -s < input.fastq > output.fastqpack
```

Output

The output of the **gto2_fq_pack** program is a packaged FASTQ file. Using the input above, an output example of this is the following:

Another example for the same input, but using the scores first (flag ''s"), is:

3.14 Program gto2_fq_unpack

The gto2_fq_unpack unpacks the FASTQ reads packaged using the gto2_fq_pack tool.

For help type:

```
./gto2_fq_unpack -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The **gto2_fq_unpack** program needs two streams for the computation, namely the input and output standard. The input stream is a packaged FASTQ file.

The attribution is given according to:

```
Usage: ./gto2_fq_unpack [options] [[--] args]
   or: ./gto2 fq unpack [options]
It unpacks the FASTQ reads packaged using the gto2 fq pack
tool.
    -h, --help
                          show this help message and exit
Basic options
    < input.fastq
                          Input FASTQ file format (stdin)
    > output.fastq
                          Output FASTQ file format (stdout)
Optional
    -s, --scores
                          When active, the application show
                          the scores first
Example: ./gto2_fq_unpack -s < input.fastqpack > out.fastq
```

An example of such an input file is:

Output

The output of the **gto2_fq_unpack** program is a FASTQ file. Using the input above, an output example of this is the following:

3.15 Program gto2_fq_quality_score_info

The ${\tt gto2_fq_quality_score_info}$ analyses the quality-scores of a FASTQ file.

For help type:

```
./gto2_fq_quality_score_info -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The gto2_fq_quality_score_info program needs two streams for the computation, namely the input and output standard. The input stream is a FASTQ file.

The attribution is given according to:

```
Usage: ./gto2_fq_quality_score_info [options] [[--] args]
  or: ./gto2_fq_quality_score_info [options]
```

It analyses the quality-scores of a FASTQ file.

```
-h, --help
                           show this help message and exit
Basic options
    < input.fastq
                           Input FASTQ file format (stdin)
    > output
                           Output read information (stdout)
Optional
    -m, --max=<int>
                           The lenght of the maximum window
Example: ./gto2_fq_quality_score_info -m <max> < input.fastq</pre>
> output
Output example :
Total reads
                 : value
Max read length : value
Min read length: value
Min QS value
                 : value
Max QS value
                 : value
QS range
                 : value
```

Output

The output of the **gto2_fq_quality_score_info** program is a set of information related to the file read. Using the input above with the max window value as 30, an output example of this is the following:

```
Total reads : 2
Max read length : 60
```

3.16 Program gto2_fq_quality_score_min

The gto2_fq_quality_score_min analyses the minimal quality-scores of a FASTQ file.

For help type:

```
./gto2_fq_quality_score_min -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The gto2_fq_quality_score_min program needs two streams for the computation, namely the input and output standard. The input stream is a FASTQ file.

The attribution is given according to:

An example of such an input file is:

Output

The output of the **gto2_fq_quality_score_min** program is a set of information related to the file read, considering the minimum quality scores. Using the input above with the max window value as 20, an output example of this is the following:

3.17 Program gto2_fq_quality_score_max

The **gto2_fq_quality_score_max** analyses the maximal quality-scores of a FASTQ file.

For help type:

```
./gto2_fq_quality_score_max -h
```

In the following subsections, we explain the input and output parameters.

Input parameters

The gto2_fq_quality_score_max program needs two streams for the computation, namely the input and output standard. The input stream is a FASTQ file.

The attribution is given according to:

An example of such an input file is:

```
@111 071112_SLXA-EAS1_s_7:5:1:817:345 length=60 1 GNNTGATGGCCGCTGCCGATGGCGNANAATCCCACCAANATACCCTTAACAACTTAAGGG
```

Output

The output of the **gto2_fq_quality_score_max** program is a set of information related to the file read, considering the maximal quality scores. Using the input above with the max window value as 20, an output example of this is the following:

3.18 Program gto2_fq_cut

to do

3.19 Program gto2_fq_minimum_local_quality_score_forward to do

3.20 Program gto2_fq_minimum_local_quality_score_reverse to do

3.21 Program $gto2_fq_xs$

to do

 $3.22 \quad Program \ gto2_fq_complement$

to do

 $3.23 \quad Program~gto2_fq_reverse$

to do

3.24 Program gto2_fq_variation_map

to do

 $3.25 \quad Program\ gto2_fq_variation_filter$

 $3.26 \quad Program~gto2_fq_variation_visual$

to do

 $3.27 \quad Program~gto2_fq_metagenomics$

Amino Acid Tools

4.1 gto2_aa

Genomic Tools

 $5.1 \quad gto2_dna$

General Purpose Tools

6.1 gto2_

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