

# Package ‘hrdtools’

September 27, 2016

**Title** Homologous Recombination Deficiency Tests with Whole Genome Sequencing

**Version** 0.0.0.9000

**Description** Implementation of various tests of homologous recombination deficiency in a single convenient package.

**Depends** R (>= 3.3.0)

**License**

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 5.0.1

## R topics documented:

count_lst . . . . .	2
crosses_centromere . . . . .	2
filter_short_contigs . . . . .	2
filter_worker . . . . .	3
get_centromere_regions . . . . .	3
get_contigs . . . . .	3
get_reference_signatures . . . . .	4
get_subtelomere_coordinate . . . . .	4
get_subtelomere_regions . . . . .	4
import_ranges . . . . .	5
import_snvs . . . . .	5
is_contiguous . . . . .	5
loh_test . . . . .	6
lst_test . . . . .	6
mutation_catalog . . . . .	6
npls_exposures . . . . .	7
npls_exposures_worker . . . . .	7
npls_montecarlo . . . . .	8
run_snv . . . . .	8
run_test . . . . .	9
stitch . . . . .	9
tai_test . . . . .	9
test_multiple . . . . .	10

<b>Index</b>	<b>11</b>
--------------	-----------

---

count_lst	<i>LST Counter</i>
-----------	--------------------

---

**Description**

This function counts the number of LST sites

**Usage**

```
count_lst(contigs, contigSizeThreshold)
```

**Arguments**

contigs	From get_contigs() function
---------	-----------------------------

---

crosses_centromere	<i>Does it Cross the Centromere?</i>
--------------------	--------------------------------------

---

**Description**

This function checks whether a region crosses the centromere

**Usage**

```
crosses_centromere(chr, start, end, centromeres)
```

**Arguments**

chr	The chromosome that the region belongs to
start	The start position of the region
end	The end position of the region
centromeres	Centromere positions from get_centromere_regions()

---

filter_short_contigs	<i>Filter Short Contigs</i>
----------------------	-----------------------------

---

**Description**

Removes short contigs from the provided contigs using specified filters

**Usage**

```
filter_short_contigs(contigs, filterSizeThreshold)
```

**Arguments**

contigs	From get_contigs() function
filterSizeThreshold	Threshold value, below which segments are filtered out

---

filter_worker	<i>Filter Worker</i>
---------------	----------------------

---

**Description**

Does the heavy lifting of filtering

**Usage**

```
filter_worker(contigs, filterSizeThreshold)
```

**Arguments**

contigs	From get_contigs() function
filterSizeThreshold	Threshold value, below which segments are filtered out

---

get_centromere_regions	<i>Get Centromere regions</i>
------------------------	-------------------------------

---

**Description**

This function returns centromere regions as a table

**Usage**

```
get_centromere_regions()
```

---

get_contigs	<i>Get Contigs from Ranges</i>
-------------	--------------------------------

---

**Description**

This function processes a GRanges object and returns a data frame of regions, all of which are contiguous. It does so by filling in all the regions not included in the ranges, by assuming these unreported regions are all heterozygous.

**Usage**

```
get_contigs(gr)
```

**Arguments**

gr	GRanges object, obtained from import_ranges().
----	--

---

```
get_reference_signatures
```

*Retrieving Reference Mutation Signatures*

---

### Description

This function imports reference mutation signatures to serve as a comparison point when performing non-negative least squares decomposition.

### Usage

```
get_reference_signatures(path = NULL)
```

### Arguments

path	String indicating path to reference signatures table file. Defaults to bundled 30-signatures.
------	---

---

```
get_subtelomere_coordinate
```

*Get Subtelomere Coordinate*

---

### Description

Convenience function that retrieves the start or end coordinate of a specific subtelomere

### Usage

```
get_subtelomere_coordinate(chr, subtelomere, startOrEnd)
```

### Arguments

chr	The chromosome of interest
subtelomere	A subtelomeres object from get_subtelomere_regions()
startOrEnd	Takes on value either "start" or "end" depending on which position to return

---

```
get_subtelomere_regions
```

*Get Subtelomere data*

---

### Description

This function returns subtelomere regions as a table.

### Usage

```
get_subtelomere_regions()
```

---

import_ranges	<i>Imports ranges output from APOLLOH</i>
---------------	---

---

### Description

This function returns a GRanges object based on the data contained in a TSV. This TSV can be output from APOLLOH or similar program.

### Usage

```
import_ranges(path, genomeVersion = "hg19")
```

---

import_snvs	<i>SNV Importer This function imports SNVs from a tab-delimited file. The file must be formatted in at least 4 tab-separated columns. The first four columns of the file must be chromosome number (without 'chr'), position, reference base, mutant base.</i>
-------------	--

---

### Description

SNV Importer This function imports SNVs from a tab-delimited file. The file must be formatted in at least 4 tab-separated columns. The first four columns of the file must be chromosome number (without 'chr'), position, reference base, mutant base.

### Usage

```
import_snvs(path, genomeVersion = "hg19")
```

### Arguments

path	Path to the tab-delimited SNV data file
------	---

---

is_contiguous	<i>Contiguity test</i>
---------------	------------------------

---

### Description

Returns TRUE if set of ranges provided is contiguous (has no gaps)

### Usage

```
is_contiguous(contigs)
```

### Arguments

contigs	From get_contigs() function
---------	-----------------------------

---

loh_test	<i>The HRD-LOH Test</i>
----------	-------------------------

---

**Description**

This function runs the Telomeric Allelic Imbalance test (HRD-LOH).

**Usage**

```
loh_test(gr)
```

**Arguments**

gr	GRanges object obtained from import_ranges()
----	--

---

lst_test	<i>The HRD-LST Test</i>
----------	-------------------------

---

**Description**

This function carries out the large scale transition (HRD-LST) test on a GRanges object.

**Usage**

```
lst_test(gr)
```

**Arguments**

gr	GRanges object, obtained from import_ranges().
----	--

---

mutation_catalog	<i>Obtain the Mutational Catalog / Spectrum</i>
------------------	---

---

**Description**

This function returns the mutational catalogue / mutational spectrum of a cancer genome.

**Usage**

```
mutation_catalog(vr)
```

**Arguments**

vr	VRanges file, obtained from import_snvs()
----	---

---

nnls_exposures	<i>Calculate signature exposures</i>
----------------	--------------------------------------

---

**Usage**

```
nnls_exposures(subjectMotifs, refSignature, fractions = FALSE,
               montecarlo = FALSE, iterations = 2000)
```

**Arguments**

subjectMotifs	The mutation catalog of a subject derived from mutation_catalog()
refSignature	The reference signature table, which can be imported by get_reference_signatures()
fractions	If TRUE, will return results as a fraction of all mutations rather than number of mutations.
montecarlo	If TRUE, will perform Monte Carlo simulation to obtain 95 \itemiterationsThe number of iterations of Monte Carlo to run This function determines the signature exposures using non-negative least squares.

---

nnls_exposures_worker	<i>NNLS Worker Function</i>
-----------------------	-----------------------------

---

**Description**

Performs the NNLS calculation itself.

**Usage**

```
nnls_exposures_worker(subjectMotifs, refSignature, fractions)
```

**Arguments**

subjectMotifs	Mutation Catalog from get_mutation_catalog()
refSignature	Reference signatures from get_reference_signatures()
fractions	If TRUE, will return exposures as a fraction of total mutation burden

---

nnls_montecarlo	<i>Monte Carlo Simulation of NNLS</i>
-----------------	---------------------------------------

---

### Description

Performs Monte Carlo simulation to obtain NNLS exposure confidence intervals

### Usage

```
nnls_montecarlo(subjectMotifs, refSignature, fractions, iterations,
               alpha = 0.05)
```

### Arguments

subjectMotifs	Mutation Catalog from get_mutation_catalog()
refSignature	Reference signatures from get_reference_signatures()
fractions	If TRUE, will return exposures as a fraction of total mutation burden
iterations	Number of iterations to run
alpha	Confidence level to retrieve confidence intervals at (default 0.05)

---

run_snv	<i>Deciphers SNV Signatures</i>
---------	---------------------------------

---

### Description

This function loads an SNV data file and runs the mutation signature deciphering process on it. It returns the output of nnls\_exposures.

### Usage

```
run_snv(snv_file, genome = "hg19", silent = FALSE)
```

### Arguments

snv_file	Path to an SNV file, suitable for import_snvs()
genome	ID of the genome being used (default: 'hg19')
silent	If TRUE, does not print exposures output (default: 'FALSE')



---

run_test	<i>Runs the LOH, TAI, and LST Tests</i>
----------	---

---

**Description**

This function loads an LOH segments file and performs the three HRD tests on it.

**Usage**

```
run_test(loh_file, genome = "hg19", silent = FALSE)
```

**Arguments**

loh_file	Path to an LOH file suitable for import_ranges()
genome	ID of the genome being used (default: 'hg19')
silent	If TRUE, does not print HRD scores, just returns them (default: 'FALSE')

---

stitch	<i>Combine Contigs</i>
--------	------------------------

---

**Description**

This function combines neighbouring contigs based on a few set criteria.

**Usage**

```
stitch(contigs)
```

**Arguments**

contigs	Contigs output from get_contigs().
---------	------------------------------------

---

tai_test	<i>The HRD-TAI Test</i>
----------	-------------------------

---

**Description**

This function runs the Telomeric Allelic Imbalance test (HRD-TAI).

**Usage**

```
tai_test(gr)
```

**Arguments**

gr	GRanges object obtained from import_ranges()
----	--

---

test_multiple	<i>Run the HRD Tests on Multiple Files</i>
---------------	--

---

**Description**

This is a convenience function to run\_test() on multiple files.

**Usage**

```
test_multiple(loh_files, output_file, genome = "hg19", multicore = TRUE)
```

**Arguments**

output_file	Path to an output file where a table of the results will be stored.
genome	ID of the genome being used (default: 'hg19')
snv_file	Character vector of paths to SNV files
silent	If TRUE, does not print exposures output (default: 'FALSE')

# Index

count\_lst, [2](#)  
crosses\_centromere, [2](#)  
  
filter\_short\_contigs, [2](#)  
filter\_worker, [3](#)  
  
get\_centromere\_regions, [3](#)  
get\_contigs, [3](#)  
get\_reference\_signatures, [4](#)  
get\_subtelomere\_coordinate, [4](#)  
get\_subtelomere\_regions, [4](#)  
  
import\_ranges, [5](#)  
import\_snvs, [5](#)  
is\_contiguous, [5](#)  
  
loh\_test, [6](#)  
lst\_test, [6](#)  
  
mutation\_catalog, [6](#)  
  
nnls\_exposures, [7](#)  
nnls\_exposures\_worker, [7](#)  
nnls\_montecarlo, [8](#)  
  
run\_snv, [8](#)  
run\_test, [9](#)  
  
stitch, [9](#)  
  
tai\_test, [9](#)  
test\_multiple, [10](#)