# The Escherichia coli Core Model: Modular Energetic Bond Graph Analysis of Pentose Phosphate Pathways

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Note: this is the EcoliPPP.ipynb notebook. The PDF version "The Escherichia coli Core Model: Modular Energetic Bond Graph Analysis of Pentose Phosphate Pathways" is available here.

• This is the version without reaction potentials

#### 1 Introduction

The Network Thermodynamics/Bond Graph approach of Oster et al. (1971, 1973), extended by Gawthrop and Crampin (2016, 2014, 2017), to modelling biomolecular systems developed independently from the stoichiometric approach Palsson (2006, 2011, 2015).

However, the conceptual point of intersection of the two approaches is the fact that the stoichiometric matrix is the modulus of the conceptual multiport transformer linking reactions to species. This was pointed out by Cellier and Greifeneder (2009). This means that the two approaches are complementary and each can build on the strengths of the other.

In particular, as discussed here, the Bond Graph approach adds energy to stoichiometry.

This notebook focuses on building modular models of metabolism and consequent pathway analysis based on the Escherichia coli Core Model Orth et al. (2010); in particular, the Glycolysis and Pentose Phosphate portion is extracted and analysed. Following the discussion in the textbook of Garrett and Grisham (2017), section 22.6d, various possible pathways are examined by choosing appropriate chemostats and flowstats. Gawthrop and Crampin (2018)

Assuming steady-state conditions, the corresponding pathway potentials Gawthrop (2017) are derived.

### 1.1 Import some python code

The bond graph analysis uses a number of Python modules:

```
[1]: ## Paths
NeedPath=True
if NeedPath:
    import sys
    sys.path += ['/usr/lib/python3/dist-packages']
```

```
[2]: ## Maths library
     import numpy as np
     import scipy
     ## BG tools
     import BondGraphTools as bgt
     ## BG stoichiometric utilities
     import stoich as st
     ## Stoichiometric conversion
     import CobraExtract as Extract
     import stoichBondGraph as stbg
     ## Potentials
     import phiData
     ## Faraday constant
     import scipy.constants as con
     F = con.physical_constants['Faraday constant'][0]
     ## Display
     import IPython.display as disp
     import copy
     ## Allow output from within functions
     from IPython.core.interactiveshell import InteractiveShell
     InteractiveShell.ast_node_interactivity = "all"
     import importlib as imp
     ## Units etc
     factor = 1
     units = ['mV', 'kJ']
     ## Control output
     quiet = True
     computePhi = True
```

```
showMu = True
```

#### 2 Extract the model

#### 2.1 Extract full ecoli core model from the CobraPy representation

[3]: sm = Extract.extract(cobraname='textbook', Remove=['\_C','\_\_'],

```
negReaction=['RPI', 'PGK', 'PGM', 'SUCOAS'], quiet=quiet)
print(sm['reaction'])
Extracting stoichiometric matrix from: textbook
Cobra Model name: e_coli_core BondGraphTools name: e_coli_core_abg
Extract. Integer only handles one non-integer per reaction
Multiplying reaction BIOMASS_ECOLIORE ( 12 ) by 0.6684491978609626 to avoid non-
integer species 3PG (2)
Multiplying reaction CYTBD ( 15 ) by 2.0 to avoid non-integer species 02 ( 55 )
Multiplying reaction PGK (54) by -1
Multiplying reaction PGM (56) by -1
Multiplying reaction RPI (65) by -1
Multiplying reaction SUCOAS (69) by -1
['ACALD', 'ACALDT', 'ACKR', 'ACONTA', 'ACONTB', 'ACT2R', 'ADK1', 'AKGDH',
'AKGT2R', 'ALCD2X', 'ATPM', 'ATPS4R', 'BIOMASS_ECOLIORE', 'CO2T', 'CS', 'CYTBD',
'D_LACT2', 'ENO', 'ETOHT2R', 'FBA', 'FBP', 'FORT2', 'FORTI', 'FRD7', 'FRUPTS2',
'FUM', 'FUMT2_2', 'G6PDH2R', 'GAPD', 'GLCPTS', 'GLNS', 'GLNABC', 'GLUDY',
'GLUN', 'GLUSY', 'GLUT2R', 'GND', 'H2OT', 'ICDHYR', 'ICL', 'LDH_D', 'MALS',
'MALT2_2', 'MDH', 'ME1', 'ME2', 'NADH16', 'NADTRHD', 'NH4T', 'O2T', 'PDH',
'PFK', 'PFL', 'PGI', 'PGK', 'PGL', 'PGM', 'PIT2R', 'PPC', 'PPCK', 'PPS', 'PTAR',
'PYK', 'PYRT2', 'RPE', 'RPI', 'SUCCT2_2', 'SUCCT3', 'SUCDI', 'SUCOAS', 'TALA',
'THD2', 'TKT1', 'TKT2', 'TPI']
```

# 2.2 Extract Glycolysis, Pentose Phosphate Pathways and TCA (using PDH and PDH)

```
[4]: name = 'GlyPPP_abg'
  reaction = []

## Glycolysis
  reaction += ['PGI', 'PFK', 'FBA', 'TPI']

## Pentose Phosphate
  reaction += ['G6PDH2R', 'PGL', 'GND', 'RPI', 'TKT2', 'TALA', 'TKT1', 'RPE']

## Create submodel
  sGlyPPP = Extract.choose(sm, reaction=reaction)
```

```
[5]: ## Create stoichiometry
import GlyPPP_abg
S = st.stoich(GlyPPP_abg.model(),quiet=quiet)
```

#### 2.3 Display the extracted reactions

[6]: disp.Latex(st.sprintrl(sGlyPPP,chemformula=True))

[6]:

$$G_6P \stackrel{PGI}{\longleftarrow} F_6P$$
 (1)

$$ATP + F_6P \xrightarrow{PFK} ADP + FDP + H$$
 (2)

$$FDP \stackrel{FBA}{\longleftrightarrow} DHAP + G_3P$$
 (3)

$$DHAP \stackrel{TPI}{\longleftarrow} G_3P \tag{4}$$

$$G_6P + NADP \xrightarrow{G_6PDH_2R} {}_6PGL + H + NADPH$$
 (5)

$$_{6}PGL + H_{2}O \stackrel{PGL}{\longleftrightarrow} _{6}PGC + H$$
 (6)

$$_{6}$$
PGC + NADP  $\stackrel{\text{GND}}{\longleftarrow}$  CO<sub>2</sub> + NADPH + RU<sub>5</sub>PD (7)

$$RU_5PD \stackrel{RPI}{\longleftarrow} R_5P \tag{8}$$

$$E_4P + XU_5PD \xrightarrow{TKT_2} F_6P + G_3P$$
 (9)

$$G_3P + S_7P \xrightarrow{TALA} E_4P + F_6P \tag{10}$$

$$R_5P + XU_5PD \xrightarrow{TKT_1} G_3P + S_7P$$
 (11)

$$RU_5PD \stackrel{RPE}{\longleftarrow} XU_5PD \tag{12}$$

#### 2.4 Code to analyse pathways defined by chemostats and flowstats

```
[7]: ## Analyse pathways defined by chemostats and flowstats
def ch(name):
    return '\\ch{'+name+'}'

def pathway(bg,phi,chemostats,flowstats=[],computePhi=False,verbose=False):
    """ Analyse pathways
    """

print('Chemostats:',sorted(chemostats))
print('Flowstats:', sorted(flowstats))
## Stoichiometry
## Create stoichiometry from bond graph.
s = st.stoich(bg,quiet=True)

## Stoichiometry with chemostats
sc = st.statify(s,chemostats=chemostats,flowstats=flowstats)

## Pathway stoichiometry
sp = st.path(s,sc)

## Print info
```

```
if verbose:
        for stat in sorted(chemostats):
            print(ch(stat)+',')
    ## Energetics
    if computePhi:
        Phi,Phip = energetics(s,sp,phi)
       #print('Phi units: kJ/mol')
          fac = -F/1000
#
#
          units='~\si{\kilo\joule\per\mol}'
        units = '~\si{\volt}'
        print(st.sprintp(sc))
        disp.Latex(st.sprintrl(sp,chemformula=True,Phi=Phip,showMu=showMu))
        #return s,sc,sp,Phi*fac,Phip*fac,units
        return s,sc,sp,Phip
    else:
        print(st.sprintrl(sp,chemformula=True))
        Phip = 0
        return s,sc,sp,Phip
def Pathway(S,chemostats,flowstats=[],verbose=False):
    """ Analyse pathways
    print('Chemostats:',sorted(chemostats))
    print('Flowstats:', sorted(flowstats))
    ## Stoichiometry
    ## Create stoichiometry from bond graph.
    #s = st.stoich(bg, quiet=True)
    s = copy.copy(S)
    ## Stoichiometry with chemostats
    sc = st.statify(s,chemostats=chemostats,flowstats=flowstats)
    ## Pathway stoichiometry
    sp = st.path(s,sc)
    ## Print info
    if verbose:
        for stat in sorted(chemostats):
            print(ch(stat)+',')
    print(st.sprintrl(sp,chemformula=True))
    return s,sc,sp
```

# 3 Analyse Pentose Phosphate Pathway with Glycolysis

The pathways are isolated by using appropriate (zero-flow) flowstats. For compatibility with Garrett and Grisham (2017, § 18.2) the pathways start from G6P (Glucose 6-phosphate).

#### 3.1 Common chemostats

```
[8]: def Chemostats(start='G6P',end=None):
    chemostats = ['ADP','ATP','C02','H','H20','NADP','NADPH']
    chemostats += [start]
    if end is not None:
        chemostats += end
    return chemostats
```

#### 3.2 R<sub>5</sub>P and NADPH generation

- This pathway is isolated by setting PGI and TKT2 as flowstats and the product R<sub>5</sub>P is added to the chemostat list.
- It is isolated from the TCA cycle by replacing the connecting reactions (PDH and PFL) by flowstats.

```
[9]: imp.reload(st)
    print('R5P and NADPH generation')
    chemostats = Chemostats(start='G6P',end=['R5P'])
    flowstats = ['PGI','TKT2']
    s,sc,sp = Pathway(S,chemostats,flowstats=flowstats)
    disp.Latex(st.sprintrl(sp,chemformula=True))
```

[9]: <module 'stoich' from
 '/home/peterg/WORK/Research/SystemsBiology/lib/python/stoich.py'>

```
R5P and NADPH generation
Chemostats: ['ADP', 'ATP', 'CO2', 'G6P', 'H', 'H2O', 'NADP', 'NADPH', 'R5P']
Flowstats: ['PGI', 'TKT2']
\begin{align}
\ch{G6P + H2O + 2 NADP &<>[ pr1 ] CO2 + 2 H + 2 NADPH + R5P }
\end{align}
```

[9]:

$$G_6P + H_2O + 2NADP \stackrel{pr_1}{\iff} CO_2 + 2H + 2NADPH + R_5P$$
 (13)

- The pathway reaction P<sub>1</sub> corresponds to the R<sub>5</sub>P and NADPH synthesis discussed in comment 1 of Garrett and Grisham (2017), p787.
- It is isolated from the TCA cycle by replacing the connecting reactions (PDH and PFL) by flowstats.

#### 3.3 $R_5P$ generation

• This pathway is isolated by setting GAPD and G6PDH2R as flowstats and the product  $R_5P$  is added to the chemostat list.

```
[10]: print('R5P generation')
    chemostats = Chemostats(start='G6P',end=['R5P'])
    flowstats = ['G6PDH2R']
    s,sc,sp = Pathway(S,chemostats,flowstats=flowstats)
    disp.Latex(st.sprintrl(sp,chemformula=True))

R5P generation
    Chemostats: ['ADP', 'ATP', 'C02', 'G6P', 'H', 'H20', 'NADP', 'NADPH', 'R5P']
    Flowstats: ['G6PDH2R']
    \begin{align}
    \ch{ADP + H + 6 R5P &<>[ pr1 ] ATP + 5 G6P }
    \end{align}
```

[10]:

$$ADP + H + 6R_5P \xrightarrow{pr_1} ATP + 5G_6P$$
 (14)

## 3.4 NADPH generation

- This pathway is isolated by setting GAPD as a flowstat.
- It is isolated from the TCA cycle by replacing the connecting reactions (PDH and PFL) by flowstats.

```
[11]: print('NADPH generation')
    chemostats = Chemostats(start='G6P')
    flowstats = []
    s,sc,sp = Pathway(S,chemostats,flowstats=flowstats)
    disp.Latex(st.sprintrl(sp,chemformula=True))

NADPH generation
    Chemostats: ['ADP', 'ATP', 'CO2', 'G6P', 'H', 'H2O', 'NADP', 'NADPH']
    Flowstats: []
    \begin{align}
```

\ch{ADP + G6P + 6 H2O + 12 NADP &<>[ pr1 ] ATP + 6 CO2 + 11 H + 12 NADPH }

[11]:

$$ADP + G_6P + 6H_2O + 12NADP \xrightarrow{pr_1} ATP + 6CO_2 + 11H + 12NADPH$$
 (15)

• The pathway reaction pr<sub>1</sub> corresponds to the NADPH synthesis discussed in comment 3 of Garrett and Grisham (2017), p787.

[]:

#### References

\end{align}

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