

# The amyloid cascade hypothesis: an updated critical review

Kasper P. Kepp,<sup>1</sup> Nikolaos K. Robakis,<sup>2</sup> Poul F. Høilund-Carlson,<sup>3,4</sup> Stefano L. Sensi<sup>5,6</sup> and Bryce Vissel<sup>7,8</sup>

Results from recent clinical trials of antibodies that target amyloid- $\beta$  (A $\beta$ ) for Alzheimer's disease have created excitement and have been heralded as corroboration of the amyloid cascade hypothesis. However, while A $\beta$  may contribute to disease, genetic, clinical, imaging and biochemical data suggest a more complex aetiology.

Here we review the history and weaknesses of the amyloid cascade hypothesis in view of the new evidence obtained from clinical trials of anti-amyloid antibodies. These trials indicate that the treatments have either no or uncertain clinical effect on cognition. Despite the importance of amyloid in the definition of Alzheimer's disease, we argue that the data point to A $\beta$  playing a minor aetiological role.

We also discuss data suggesting that the concerted activity of many pathogenic factors contribute to Alzheimer's disease and propose that evolving multi-factor disease models will better underpin the search for more effective strategies to treat the disease.

- 1 Section of Biophysical and Biomedical chemistry, DTU Chemistry, Technical University of Denmark, 2800 Kongens Lyngby, Denmark
- 2 Icahn School of Medicine at Mount Sinai Medical Center, New York, NY 10029, USA
- 3 Department of Nuclear Medicine, Odense University Hospital, 5000 Odense C, Denmark
- 4 Department of Clinical Research, University of Southern Denmark, 5000 Odense C, Denmark
- 5 Center for Advanced Studies and Technology—CAST, and Institute for Advanced Biotechnology (ITAB), University G. d'Annunzio of Chieti-Pescara, Chieti, 66013, Italy
- 6 Department of Neuroscience, Imaging, and Clinical Sciences, University G. d'Annunzio of Chieti-Pescara, Chieti, 66013, Italy
- 7 St Vincent's Hospital Centre for Applied Medical Research, St Vincent's Hospital, Sydney, 2010, Australia
- 8 School of Clinical Medicine, UNSW Medicine and Health, St Vincent's Healthcare Clinical Campus, Faculty of Medicine and Health, Sydney, NSW 2052, Australia

Correspondence to: Kasper P. Kepp

Section of Biophysical and Biomedical chemistry, DTU Chemistry, Technical University of Denmark, Kemitorvet 206, DK 2800 Kongens Lyngby, Denmark

E-mail: kpj@kemi.dtu.dk

**Keywords:** Alzheimer's disease;  $\beta$ -amyloid; aducanumab; lecanemab; amyloid cascade hypothesis

## Introduction

Fifty-five to sixty million people are estimated to suffer from dementia, and perhaps 30–40 million of them are clinically estimated to have Alzheimer's disease (AD). Dementia kills in the order of a million people every year globally and is one of the most significant tolls on healthcare budgets in the developed world.<sup>1–3</sup> Despite

decades of intense research on AD-type dementia, available treatments have short-lived effects.<sup>1,4–6</sup>

The idea that amyloid- $\beta$  (A $\beta$ ) causes AD emerged in the 1980s<sup>7–9</sup> and has dominated research into treatments since then (the amyloid cascade hypothesis, ACH).<sup>10,11</sup> The controversial approval of Biogen's antibody aducanumab (Aduhelm®, ADU) by the Food and Drug Administration (FDA)<sup>12–15</sup> renewed concerns about the ACH, whereas

on the other hand, a clinical trial using the antibody lecanemab in November 2022 showed possible beneficial effects.<sup>16</sup> For patients and families, this is an important sign of hope, and for some scientists the results are perceived as support for the ACH.<sup>17,18</sup>

AD is a complex disorder with age as the leading risk factor and a broad clinical spectrum of symptoms.<sup>19–22</sup> Many risk modifiers and biochemical pathways contribute to disease.<sup>23–26</sup> AD exhibits diverse histopathological phenotypes that include not just the ‘hallmarks’ brain amyloid consisting of A $\beta$  peptides<sup>27</sup> and tau depositions,<sup>28–30</sup> but also neurodegeneration, the deposition of TAR DNA-binding protein-43 (TDP-43),  $\alpha$ -synuclein and other misfolded proteins.<sup>31–33</sup> In addition, oxidative tissue damage,<sup>34–37</sup> inflammation,<sup>38,39</sup> metabolic abnormalities,<sup>40–45</sup> atherosclerosis,<sup>46–49</sup> cardiovascular effects,<sup>50,51</sup> cerebral hypoperfusion<sup>52,53</sup> and imbalances of intraneuronal calcium<sup>54–56</sup> and other metal ions<sup>25,57–60</sup> participate in the pathology of AD.

AD mainly occurs in the population as sporadic AD (SAD; >95% of cases) with many genetic and lifestyle risk modifiers<sup>61,62</sup> developing slowly first as mild cognitive impairment (MCI),<sup>20,63,64</sup> and subsequently with loss of episodic memory, cognitive skill and identity.<sup>1,65</sup> A few per cent of cases are inherited as autosomal dominant early-onset familial AD (FAD), typically beginning at 30–65 years, whereas SAD usually develops after 65 years.<sup>66,67</sup> The FAD cases are associated with mutations in the genes encoding presenilin-1 (PS1),<sup>68–71</sup> presenilin-2 (PS2),<sup>72</sup> and the amyloid- $\beta$  precursor protein (APP).<sup>73–77</sup> Despite causing a small subset of AD, FAD mutations have profoundly influenced AD research due to their appealing genotype-phenotype relationships.<sup>11,78–81</sup>

In contrast to these complex aetiologies, the original ACH suggested that the disease is the consequence of a ‘machine failure’ in the form of overload of brain A $\beta$  depositions. This view substantially impacted AD research, funding and drug development efforts<sup>82</sup> despite consistent concerns expressed by many in the scientific community,<sup>19,83–88</sup> especially after repeated drug failures from 2014 and forward.<sup>89–95</sup>

With the aim of presenting the history, data and discussions of the ACH in a critical context of the recent antibody trials, we systematically searched the literature for papers on the ACH and its history and criticism. Our review led to a list of 15 data-related distinct discussion points necessary for the debate on the hypothesis and how to move forward from the current state of available data.

## The amyloid cascade hypothesis

### Early foundation and molecular and genetic basis

The ACH can be traced back to the first half of the 1980s. Prusiner<sup>96</sup> and Masters et al.<sup>97</sup> noted analogies between brain amyloid deposition in AD patients and proteinopathies found in Creutzfeldt-Jakob diseases, implying a causal gain-of-toxic function. Glenner and Wong<sup>8</sup> purified and sequenced amyloid peptides from cerebral blood vessels (cerebrovascular amyloid, CVA) of AD and Down syndrome patients. They also found that both amyloid forms contained the same primary sequence of  $\beta$ -protein (now known as A $\beta$  peptides) and predicted a genetic risk locus at chromosome 21.<sup>98</sup> In 1985, Masters and colleagues<sup>9</sup> purified plaque A $\beta_{40}$ , characterized the N-terminal sequence and related it to the plaques in Down syndrome. The first amyloid-based aetiology of AD was proposed by Glenner and associates in the mid-1980s and posited that CVA compromises the blood-brain barrier and harms neurons.<sup>7,99</sup> With reports indicating that A $\beta$  could be cytotoxic,<sup>100,101</sup> Hardy and Higgins<sup>10</sup> formulated the ACH proposing that A $\beta$  plaque deposition

was the main pathogenic factor of AD. A $\beta$  derives from the transmembrane  $\beta$ -APP cloned in 1987 by several groups<sup>102–105</sup> and mutations in the genes encoding APP<sup>106</sup> and PS1<sup>68</sup> may modify the production of A $\beta$ , which is thought to cause early-onset FAD. This was interpreted as supporting the ACH rather than generating interest in the cellular functions of the two proteins and their role in the neurodegeneration of AD.<sup>68,107,108</sup>

In the non-amyloidogenic pathway, APP is cleaved by  $\alpha$ -secretase in the middle of the A $\beta$  sequence within APP, precluding the formation of A $\beta$ .<sup>109</sup> However, APP is also cleaved by  $\beta$ -secretase to yield the 99-residue C-terminal fragment of APP (APP-C99) that includes the A $\beta$  sequence.<sup>110–112</sup> A $\beta$  peptides of variable length are formed during consecutive cleavages of APP-C99 by  $\gamma$ -secretase, the four-subunit intramembrane di-aspartyl protease complex having PS1 or PS2 as its multipass transmembrane catalytic subunit.<sup>10,113–116</sup> The cleavage occurs along two pathways, yielding the peptides A $\beta_{49}$   $\rightarrow$  A $\beta_{46}$   $\rightarrow$  A $\beta_{43}$   $\rightarrow$  A $\beta_{40}$   $\rightarrow$  A $\beta_{37}$  and A $\beta_{48}$   $\rightarrow$  A $\beta_{45}$   $\rightarrow$  A $\beta_{42}$   $\rightarrow$  A $\beta_{38}$ . The physiological significance of these cleavage products remains poorly understood,<sup>116–119</sup> but evidence indicates that APP and its processing may have important physiological functions.<sup>120–124</sup> This view is also supported by recent findings indicating that A $\beta_{42}$  is associated with normal cognition and preservation of hippocampal volume.<sup>125,126</sup> However, longer peptides (e.g. A $\beta_{42}$ ) are more hydrophobic, toxic and aggregate more easily,<sup>127</sup> and a surplus of longer peptides (e.g. the A $\beta_{42}$ /A $\beta_{40}$  ratio) has thus been proposed as a key driver of the pathogenic process.<sup>118,128–131</sup>

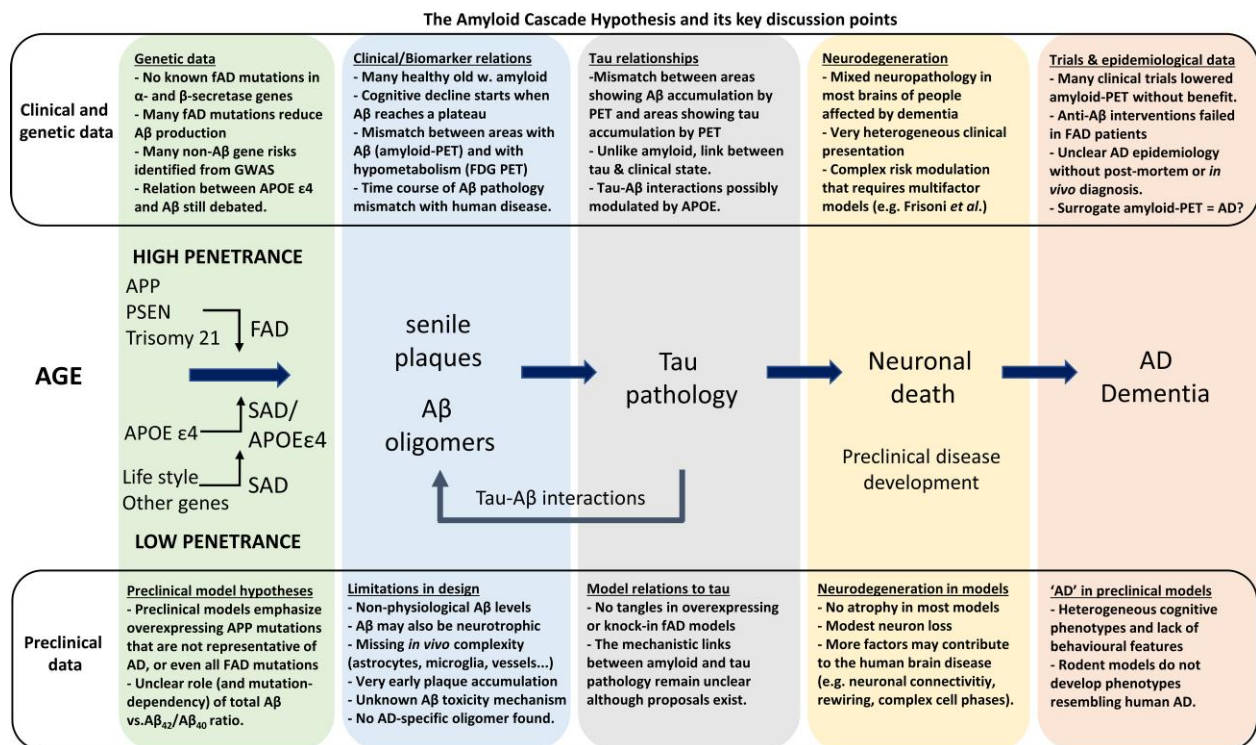
The tendency of many FAD mutations to cause autosomal dominant AD<sup>76,79,132,133</sup> is often interpreted as a gain-of-toxic function.<sup>134</sup> However, loss-of-function cannot be ruled out as a cause of FAD. For example, in protein misfolding diseases, a peptide’s misfolding can cause loss of that protein’s normal function while at the same time, generate a toxic interaction with other parts of the cell.<sup>135</sup> Also, one wild-type allele is not necessarily enough to fully compensate for the loss-of-function of the inactivated allele, thereby causing a loss-of-function disorder. This is especially true over time, so this argument is relatively weak when considering a gradually evolving age-dependent disease such as AD.<sup>136</sup> An important example of loss-of-function leading to dementia is progranulin (PRGN) mutations that cause a 50% reduction of the protein (haploinsufficiency), leading to autosomal dominant neurodegeneration and frontotemporal dementia.<sup>137</sup>

Two main therapeutic avenues have been explored based on the ACH: modifying the production of A $\beta$  peptides or neutralizing them after formation. Many pharmaceutical companies have developed  $\beta$ - and  $\gamma$ -secretase inhibitors or, more recently, due to clinical adverse effects of many of these,<sup>138,139</sup> modulators of  $\gamma$ -secretase, changing the relative production of the various forms of A $\beta$ ,<sup>140–148</sup> or antibodies intended to neutralize the formed toxic peptides.<sup>149–152</sup>

### Current versions of the amyloid cascade hypothesis

The current version of the ACH<sup>11,153,154</sup> differs in several ways from the original one proposed in the 1990s<sup>10</sup> and is summarized in Fig. 1. It describes the oligomers as the most likely pathogenic species, in contrast to plaques, and considers the role of risk factors that modulate penetrance, for example, apolipoprotein E (APOE) and tau-A $\beta$  interactions.<sup>154</sup>

As studies found that senile plaque loads do not correlate with cognitive decline<sup>155–157</sup> and higher A $\beta$  production was not a phenotype of many FAD mutations,<sup>158,159</sup> attention moved to other forms of A $\beta$  as probably more important to disease. Notably, soluble



**Figure 1 Overview of the amyloid cascade hypothesis and its related controversies.** The amyloid cascade hypothesis<sup>11,80,153</sup> has been updated into a model in which APOE  $\epsilon$ 4 and tau pathology affect total penetrance (Frisoni *et al.*<sup>154</sup>). A $\beta$  = amyloid- $\beta$ ; AD = Alzheimer's disease; FAD = familial Alzheimer's disease; GWAS = genome-wide association studies; SAD = sporadic Alzheimer's disease.

oligomers of A $\beta$ <sup>100</sup> are more cytotoxic than fibrillar A $\beta$  plaques.<sup>160–162</sup> The selective toxicity of oligomers in cellular assays was taken as further support for this hypothesis.<sup>163–165</sup> The deposits of senile plaques consist of post-translationally modified, oxidized and truncated forms of A $\beta$  in fibrillar  $\beta$ -sheet structures representing several years of production of A $\beta$ <sup>166</sup> and represent end points of aggregation processes that initiate before clinical onset<sup>167</sup> and are probably less directly involved in pathogenesis compared to their precursor oligomers.<sup>168</sup>

Exposure of the hydrophobic parts of A $\beta$  is likely to cause aggregation and interactions with membranes and other molecules.<sup>169–171</sup> Nevertheless, the molecular mechanisms, location and context of A $\beta$  aggregates causing AD remain elusive despite suggestions<sup>172</sup> that these aggregates cause permeabilization of cell membranes,<sup>169,173–176</sup> dysfunction of neuronal potentiation,<sup>165</sup> oxidative stress,<sup>177</sup> vascular effects,<sup>46,50</sup> disruption of prion-protein interactions and dysfunction of the NMDA receptors,<sup>178</sup> abnormalities of the respiratory chains of mitochondria<sup>179–181</sup> and effects on neuronal calcium and zinc dyshomeostasis.<sup>60,182,183</sup>

An important modification of the ACH is to account for the more complex pathology and many small cumulative genetic, vascular and lifestyle-related factors,<sup>184,185</sup> i.e. abandoning a single-cause determinism of toxic proteinopathy.<sup>44</sup> The 'probabilistic' version of the ACH<sup>154</sup> incorporates additional factors into a model that still retains A $\beta$  as a main causal agent but distinguishes three types of AD (Fig. 1): (i) FAD with very high penetrance due to mutations in PS1, PS2 and APP; (ii) SAD with APOE  $\epsilon$ 4 genotype as an additional risk increasing penetrance; and (iii) SAD without an APOE  $\epsilon$ 4 genotype, and with the latest typical age of onset. The concept of using risk-modifying penetrance resembles other multi-risk models,<sup>44</sup> but the identification of three

distinct types of AD makes it a novel and straightforward extension of the ACH.

Another trend is the appreciation of a need for understanding the normal functions of APP, A $\beta$  and presenilin, and the possibility that some loss-of-function pathways can be viewed in connection with the ACH.<sup>11</sup> In combination, these studies reflect a more recent tendency towards a hypothesis that accommodates multiple modifiable genetic, vascular and lifestyle risk factors<sup>154</sup> and loss of the physiological functions of the proteins involved in the proteinopathy, rather than considering the mutant FAD genes as simply pathogenic.<sup>11</sup>

### Criticism of the amyloid cascade hypothesis

The main criticisms of the ACH have often been stated to be the lack of correlation between cognition and amyloid deposition<sup>19</sup> and the modest clinical effects of anti-amyloid drugs.<sup>90,186</sup> However, as discussed below, there are many other anomalies, for example, in relation to the genetic data and increasing evidence of other disease risk factors that should weigh heavily.<sup>19,86,89,90,92,136,162,187</sup> Criticism of the ACH has been continuous<sup>19,84,86,89,136,188</sup> but markedly intensified in 2014–2017 after the failure of several high-profile anti-amyloid drug candidates.<sup>89,90,92–94,187,189,190</sup> Some of the main discussion points of the ACH that have emerged are summarized in Table 1 and discussed below. A list of associated representative statements regarding the hypothesis is presented in Box 1.

### Evidence of a complex aetiology

In contrast to the complex clinical and neuropathological phenotypes of AD, the original version of the ACH proposed a simple cause-effect relationship between amyloid deposition and

Table 1 Criticism of the amyloid cascade hypothesis, and its counter-arguments

Criticism number	Observation	Context	Notes/statements	Counter-argument
1	Evidence of a complex aetiology <sup>19,191</sup>	Many risk factors, heterogeneous results and clinical spectrum should be addressed <sup>92,192</sup>	Selective interpretation of genetic, biochemical and clinical data	New models begin to account for more risks modulating penetrance <sup>154</sup>
2	The ageing human brain differs from mice	Much evidence based on rodent models <sup>193,194</sup> with distinct neurology and ageing	Antibodies tend to reverse cognitive decline in mice but not in humans. <sup>90</sup> Mouse A $\beta$ confounds results <sup>195</sup>	Primates are expensive and some mechanisms can still be shown in mice
3	Neglect of normal roles of APP, A $\beta$ and PS1 <sup>85,92,123,136,196</sup>	A $\beta$ is neurotrophic at normal concentrations, but toxic at high concentrations <sup>197,198</sup>	Soluble A $\beta_{42}$ correlates with cognitive function <sup>125,126</sup> elaborate splicing of APP is clearly there for a reason <sup>123</sup>	$\gamma$ -Secretase modulators or antibodies may be made selective to harmful forms of A $\beta$
4	No established causal molecular species; the mechanism of A $\beta_{42}$ /A $\beta_{40}$ ratio is obscure <sup>92</sup>	Despite decades of research, the molecular mode of pathogenesis of A $\beta$ is unclear <sup>11,92,199,200</sup>	The structure, mechanism and cellular location of pathogenic species not established; some oligomers may be artefacts <sup>201</sup>	Some oligomers have been proposed but very hard to identify <i>in vivo</i> mechanism <sup>136,162</sup>
5	Poor correlation of A $\beta$ to clinical outcome <sup>157,202</sup>	A $\beta$ in many asymptomatic people <sup>155–157,203</sup> could suggest A $\beta$ is not causal <sup>19,57,90,92,188,204</sup>	At the minimum additional modulators of disease required. <sup>204,205</sup> One proposal is tau-A $\beta$ interactions <sup>154</sup>	Preclinical <sup>1206</sup> or cognitive reserves, <sup>186</sup> oligomers rather than plaques <sup>162,189</sup> or tau-A $\beta$ effects <sup>154</sup>
6	Cytotoxicity is not the same as human pathogenicity <sup>86,187</sup>	Studies often used 1000-fold physiological A $\beta$ concentration <sup>19,86,92,187</sup>	A $\beta$ is sub-nanomolar concentrations, toxicity studies often micromolar <sup>19,92</sup>	Some studies <sup>207</sup> showed toxicity at physiological concentrations
7	Absence of $\alpha/\beta$ secretase FAD mutations <sup>92</sup>	If processing of A $\beta$ was central such mutations are expected	Only seen in APP close to cleavage site, suggesting PS1/PS2 more central	Not considered to the author's knowledge
8	Absence of mutations in key A $\beta$ degrading metalloproteases <sup>92</sup>	Hypomorphic mutations in main A $\beta$ degrading proteases should confer risk	Not seen	Not considered to the author's knowledge
9	Many FAD mutations lead to less A $\beta$ <sup>123,158</sup>	Suggests that 'cascade' of A $\beta$ is not causal <sup>204</sup>	Data e.g. in Sun et al. for PS1 mutations <sup>159</sup>	Both A $\beta$ and A $\beta_{42}$ /A $\beta_{40}$ ratio gives disease (i.e. nearly all mutations)
10	Many more FAD mutations in PS1 than elsewhere	Indicates PS1/ $\gamma$ -secretase is more central to disease <sup>208</sup>	Impaired APP cleavage leading to plaques could be correlation, not cause	Most PS1 mutations affect APP cleavage <sup>159</sup>
11	APP mutation have diverse effects <sup>131</sup>	Some APP FAD mutations lower, others raise A $\beta$	Swedish mutation increases A $\beta$ but is not representative of FAD mutations	Emphasis on longer peptides such as A $\beta_{42}$ and A $\beta_{43}$ <sup>209–212</sup>
12	Accounting for the physiological regulators of A $\beta$ balance	A $\beta$ /APP is bound and modified by metal ions <sup>120,213–218</sup>	Metal ions bind, regulate production, aggregation and clearance of A $\beta$ <sup>57,59,122,219–225</sup>	Even within amyloid hypothesis it seems relevant to account for such regulators <sup>122</sup>
13	Interpretation of Down syndrome dementia as due to APP <sup>226</sup>	APP overexpression associates with dementia in Down syndrome patients.	>200 genes on chromosome 21, including BACE2 and APP but also genes of SOD1, proteostasis, calcium binding and ion channel proteins	Regardless of other genes, the excess APP and plaque pathology is consistent with the hypothesis
14	Apparent A $\beta$ removal has not improved cognition <sup>90,92,227,228</sup>	'Bayesian meta-analysis of these trial data provides strong evidence of absence of a therapeutic effect'. <sup>227</sup>	Autopsy studies of patients who received A $\beta$ immunization indicate no slowed progression <sup>228</sup>	The hypothesis is ok, but trials failed to target the right species at the right time <sup>189</sup>
15	Blocking A $\beta$ producing enzymes impairs cognition <sup>139</sup>	Not unexpected, because of A $\beta$ 's proposed functions <sup>85,123,197,229</sup>	$\beta$ - and $\gamma$ -secretase inhibitors cause adverse effects <sup>89</sup>	Not because of neurotrophic A $\beta$ but other substrates <sup>139</sup>

neurodegeneration. However, AD is a complex disorder as indicated by the involvement of many genetic risk factors.<sup>92,192</sup> In the homozygote state, the ApoE  $\epsilon 4$  allele increases the AD risk by up to 15-fold.<sup>235–237</sup> Many other genetic risk factors have been identified from genome-wide association studies (GWAS),<sup>238,239</sup> including for example, ABCA7, BIN1, CLU, GAB2, GALP, PICALM and TREM2 that seem implicated in broader functions, such as, lipid metabolism

and immune functions.<sup>75,239–241</sup> Lifestyle-related risk factors such as body mass index,<sup>242,243</sup> diabetes,<sup>244–246</sup> hypertension<sup>247</sup> and depression<sup>248,249</sup> also seem to affect and modify the disorder.<sup>250,251</sup> It has been estimated that 40% of dementia cases are caused by 12 modifiable risk factors.<sup>252</sup> These complex pathways then need to be seen in the context of gradual ageing and oxidative stress-induced neuronal insults.<sup>34–37,230</sup> One way to do this is a



**Box 1 Statements about the amyloid cascade hypothesis, in chronological order**

'This is a controversial theory, however, primarily because there is a poor correlation between the concentrations and distribution of amyloid depositions in the brain and several parameters of AD pathology, including degree of dementia, loss of synapses, loss of neurons and abnormalities of the cytoskeleton'.<sup>19</sup>

'Clearly, it is time to rethink this position and to propose instead that future approaches should focus upon altering the age-related sensitivity of the neuronal environment to insults involving such factors as inflammation and oxidative stress'.<sup>230</sup>

'A detailed review of the relevant data led us to conclude that some data, particularly those from transgenic mice, are inconsistent with the predictions of the amyloid hypothesis. Instead, most data are consistent with the notion that amyloid- $\beta$  (A $\beta$ ) peptide is neuroprotective'.<sup>84</sup>

'By analogy, individuals suffering from altitude sickness nearly always have elevated levels of hemoglobin. However, while hemoglobin is toxic to cells in culture and increased erythropoiesis at sea level can be deadly, it is clear that the increases in hemoglobin occurring at altitude are beneficial. Amyloid, like hemoglobin, may also be beneficial, in this case, following neuronal stress or disease'.<sup>86</sup>

'Implicit in the amyloid hypothesis is that the A $\beta$  peptide harbors neurotoxic properties. Yet, the precise mechanism by which A $\beta$  exerts these putative toxic effects on neurons remains unclear'.<sup>27</sup>

'While the debate over the validity of the amyloid cascade hypothesis will no doubt continue, it remains likely that there are other critical factors playing a role in AD pathogenesis. Metal ions are one such possibility'.<sup>231</sup>

'the amyloid cascade hypothesis has been so modified over time that it is now impossible to confirm or deny. The hypothesis now states, in effect, that invisible molecules target invisible structures. Still relevant, however, are multiple factors that surely cast some doubt but have either been rationalized or overlooked'.<sup>87</sup>

'This elusive soluble A $\beta$  species is in danger of becoming a way to explain inconsistencies in existing models without applying the scientific rigor needed to make real progress'.<sup>162</sup>

'The findings that SDS promotes A $\beta$  dimerization have significant implications for the putative role of low-order oligomers in AD pathogenesis and draw into question the utility of oligomeric A $\beta$  as a therapeutic target'.<sup>201</sup>

'We show here however that the central conclusion of the amyloid hypothesis, that A $\beta$  is the cause of AD is, at very least, premature'.<sup>89</sup>

'it is increasingly likely that A $\beta$  is merely a marker, a later consequence of upstream changes that lead to neuronal and synaptic losses'.<sup>204</sup>

'while there is fear in the field over the consequences of rejecting it outright, clinging to an inaccurate disease model is the option we should fear most'.<sup>90</sup>

'the large and solid body of evidence accumulated to date points to the waning of the amyloid cascade hypothesis and indicates that the scientific community should still devote its utmost efforts to identify the real culprit of Alzheimer's disease'.<sup>93</sup>

'We're all disappointed, but there are concrete reasons why each failure occurred'.<sup>232</sup>

'Are we going to tie up the whole field for another 15 to 20 years?'<sup>232</sup>

'The general view is that these are the right drugs, but they're too late'.<sup>232</sup>

'A holistic view of the available data does not support an unequivocal conclusion that A $\beta$  has a central or unique role in AD'.<sup>94</sup>

'Diagnostic and therapeutic approaches rooted in the 'amyloid hypothesis' have permeated every fiber of AD research for decades by creating a domain of deceiving intellectual monopoly. These unjustified efforts have affected the decisions of funding agencies and editorial offices, and as such, have prevented the exploration of new ideas and avenues of research'.<sup>233</sup>

'As a result, dementia research is estimated to be 15–30 years behind where it could be with conflicting data ignored in favor of the amyloid dogma'.<sup>234</sup>

'annual NIH support for studies labeled amyloid, oligomer, and Alzheimer's' has risen from near zero to \$287 million in 2021'.<sup>88</sup>

model that assumes that some of these disease modulators contribute to aggravating A $\beta$  pathology.<sup>184</sup> Recent formulations of the hypothesis have therefore moved towards incorporating these modulators in a penetrance model, with three forms of AD emphasizing the APOE  $\epsilon$ 4 genotype in particular, as summarized in Fig 1.<sup>154</sup>

**Relying on preclinical familial Alzheimer's disease models**

Much support for the ACH came from transgenic rodent models expressing FAD mutants. However, given the marked differences in the ageing processes and neurological development of rodents, results obtained in transgenic mice are easily over-interpreted.<sup>193,194</sup> For example, antibodies tend to reverse cognitive decline in mice but not humans.<sup>90</sup> Even at the level of amyloid aggregation and associated pathology, mouse A $\beta$  itself differs by three substitutions from human A $\beta$ , and endogenous mouse A $\beta$  aggregates to form

amyloid at a much lower rate than human A $\beta$ .<sup>195</sup> To overcome the problem, murine AD models have used over-expressed mutant human APP, but this is a non-physiological setting that mainly explores the human peptide's interaction with the host murine brain, as discussed previously.<sup>89,136,253</sup> One of the issues is that AD does not generally feature APP overexpression, which may cause toxicity by itself,<sup>136,254</sup> rather than by a mechanism that relates to AD. Thus, while most models are wrong and only some useful, the distinct ageing and neurological features of humans limit the value of pathogenic processes observed in mice.

**Physiological roles of APP, A $\beta$  and PS1/ $\gamma$ -secretase have been neglected**

The ACH is not concerned with the normal function of APP, A $\beta$  and PS1 and why humans feature APP and its complex cleavage by

secretases in the first place.<sup>85,123,255–257</sup> APP is expressed as a chondroitin sulphate proteoglycan, which has important brain functions.<sup>258</sup> Plausible functions of APP<sup>124,213</sup> were established at the same time as the ACH, and A $\beta$  concentrations below nanomolar (as encountered within cells) are neurotrophic while higher concentrations (as achieved in many models) are toxic.<sup>197,198,259</sup> Nevertheless, the original ACH focused exclusively on A $\beta$  toxicity. While monomeric A $\beta$  seems important for normal brain function,<sup>85,119,123,196,205,260–262</sup> the exact functions remain elusive but may include effects on neuronal signalling in the synaptic cleft,<sup>229,263</sup> anti-microbial function,<sup>264–266</sup> potassium channel modulation<sup>267</sup> and inhibition of blood–brain barrier leaking.<sup>265</sup> Whereas the N-terminal half of A $\beta$  is hydrophilic, the cleaved C-terminal half is hydrophobic, making A $\beta_{42}$  more hydrophobic than A $\beta_{40}$ .<sup>268</sup> Thus, senile plaques tend to be enriched in longer A $\beta$  peptides.<sup>269</sup> Although the potential beneficial roles of A $\beta$  were ignored, toxicity data,<sup>197,259</sup> plaque deposits and FAD mutations pointed to possible loss-of-function<sup>123</sup> and foresaw the adverse effects of indiscriminate amyloid reduction by antibodies and  $\gamma$ -secretase inhibitors.<sup>256,270</sup> Also, A $\beta$  functions in innate immunity<sup>271,272</sup> may be explored in the context of the role of infections in AD.<sup>273–278</sup> A recent study concluded that soluble A $\beta_{42}$  is associated with normal cognitive function, and low A $\beta_{42}$  with cognitive impairment,<sup>125,126</sup> consistent with a loss-of-function of APP/A $\beta$ .<sup>123</sup> There is also good evidence that many PS1 FAD mutations reduce the  $\gamma$ -secretase-mediated epsilon cleavage of many substrates beyond APP-C99, including NOTCH1 and cadherins,<sup>279</sup> and the resulting cytosolic peptides regulate gene expression necessary for neuron function.<sup>280–285</sup>

### No established causal molecular amyloid- $\beta$ mechanism

The ACH has not established the precise pathogenic molecular form of A $\beta$  (some forms of oligomers<sup>164,286,287</sup>), the detailed location of the process in the brain (arguably synapses<sup>288,289</sup>), and the exact molecular mode of pathogenesis.<sup>11,92,199,200</sup> Current strategies assume that we can identify the specific pathogenic form of A $\beta$  and thereby, rationally develop specific antibodies targeting the relevant structural epitopes. Low molecular mass oligomers, such as dimers, may be artefacts of sodium dodecyl sulphate (SDS)-induced dimerization.<sup>201</sup> At the very least, it is clear that A $\beta$  cannot cause disease by itself but needs a modifier. Even if some forms of oligomers are pathogenic, it has been hard to identify relevant *in vivo* mechanisms,<sup>162</sup> and the specific mechanism that makes the A $\beta_{42}$ /A $\beta_{40}$  ratio, rather than the total amount of A $\beta_{42}$  or A $\beta_{40}$ , biologically significant remains obscure.<sup>92</sup>

### Amyloid biomarkers and their relation to Alzheimer's disease

The ACH proposes that A $\beta$  deposition should be associated with disease,<sup>11,80</sup> possibly with a time delay that is heterogeneous in the type of risk profile of the individual patient.<sup>154</sup> Asymptomatic people could reflect preclinical disease states,<sup>206</sup> but it seems clear that additional modulators are involved.<sup>57,136,186,204,205</sup>

A $\beta$  deposition is measured by PET using tracers such as <sup>11</sup>C-labelled Pittsburgh Compound-B<sup>290</sup> or <sup>18</sup>F-florbetapir, <sup>18</sup>F-florbetaben and <sup>18</sup>F-flutemetamol; with distinct chemistry, affinity and specificity.<sup>233,291–295</sup> Whereas amyloid-PET has high sensitivity for  $\beta$ -sheet structured fibrillar depositions (typically 80–100%),<sup>296,297</sup> complex visual and quantitative assessment confounded by spillover from tracer uptake in white matter and partial volume effects complicate use.<sup>295,298</sup> Thus, very heterogeneous specificities (46–88%) with a median estimate of 58% were found

in the Cochrane review on <sup>11</sup>C-labelled Pittsburgh Compound-B for MCI who eventually convert to AD.<sup>299</sup> Meta-analysis supports a consensus that cross-sectional A $\beta$ -cognition relationships are modest among healthy elderly.<sup>300</sup> Although A $\beta$  is found in many parts of the brain, AD generally begins in the entorhinal/parahippocampal cortices and the hippocampus.<sup>301,302</sup> A $\beta$  deposits correlate only modestly with clinical outcomes.<sup>157,202,303</sup> As regularly mentioned,<sup>57,89,90,204</sup> 30% or more of asymptomatic older people have high loads of A $\beta$  plaques,<sup>155–157,203</sup> and many of these are enough to satisfy AD diagnosis criteria.<sup>304–306</sup> Also, a significant fraction of amyloid-PET positive subjects remain cognitively healthy at 5 years follow-up,<sup>307</sup> and radiotracer uptake is also confounded by covariates such as sleep patterns.<sup>308,309</sup>

The fluid biomarkers offer useful complementary insight, most notably the CSF A $\beta_{42}$ /A $\beta_{40}$  ratio with a clear tendency of decline with age and in AD.<sup>206,304,310</sup> The A $\beta_{42}$ /A $\beta_{40}$  ratio is typically inversely related to the amyloid PET signal, which is interpreted as due to plaques being enriched in the longer more hydrophobic and aggregation-prone A $\beta_{42}$  peptide.<sup>311</sup> However, this relationship has not been robust in antibody trials that offer the highest level of evidence (randomized controlled trials) for these relationships. While most antibodies lower amyloid-PET, bapinezumab and gantenerumab did not clearly affect CSF A $\beta_{42}$ /A $\beta_{40}$ .<sup>312–314</sup> The inverse relationship between the two biomarkers is strong for aducanumab and lecanemab.<sup>16,315</sup>

In recognition that tau, at least, is also involved, the clinical criteria for AD diagnosis by the Consortium to Establish a Registry for Alzheimer's Disease (CERAD)<sup>316</sup> include Braak tau pathology and A $\beta$  pathology not directly captured by standard amyloid PET.<sup>293</sup> The National Institute of Aging-Alzheimer's Association (NIA-AA) diagnostic guidelines for AD criteria included in 2011, amyloid positivity as one element of AD diagnosis, at least in research,<sup>317</sup> and more recently, in practice.<sup>65</sup> The International Working Group (IWG) emphasized clinical markers as well, although maintaining that amyloid (and tau) positivity together with specific AD phenotypes is a precondition for diagnosis.<sup>318</sup>

The correlation between amyloid and tau pathology is complex and not fully understood, with tau pathology typically correlating better with clinical course.<sup>184</sup> A recent paper by Frisoni et al.<sup>154</sup> includes a detailed discussion of this 'spatial paradox' and the tau-amyloid relationship that may be modulated by APOE  $\epsilon$ 4 genotype.<sup>319</sup> It seems likely as more data accrue that the complexity will not be explained by amyloid, tau and APO  $\epsilon$ 4 alone.

### Cell toxicity is not the same as human pathogenicity

Some of the main data supporting the ACH originated from studies evaluating A $\beta$  effects on cell viability or *in vitro* aggregation assays.<sup>86,187</sup> However, many reports of A $\beta$  toxicity<sup>320</sup> and aggregation tendency<sup>321</sup> occurred at micromolar concentrations,<sup>136</sup> although biological concentrations of the peptides are in the (sub) nanomolar range, i.e. the models used amounts of peptide that represent perhaps 1 year of total brain production administered instantaneously at 1000-fold higher than biological concentrations.<sup>19,86,187,322,323</sup> This is unlikely to occur with a slowly developing, age-induced human disease of the brain. Although some toxic modes at physiologically relevant concentrations were reported,<sup>207</sup> the implications of these studies for a complex neurodegenerative disorder remain highly doubtful, and even if the used cells had ageing processes and phenotypes that resemble human neurons,<sup>324,325</sup> the multicellular context and interplay of connected neurons, astrocytes, microglia, etc. is not caught by such models,<sup>38,89,326</sup> including the central role played by synapses in this interplay.<sup>89,289,327</sup>

### Absence of familial Alzheimer's disease mutants of $\alpha$ - and $\beta$ -secretase

One of the most surprising claims is that the FAD genetic evidence for the ACH is almost indisputable.<sup>328</sup> Accordingly, PS1 and APP mutations indicate a role for these genes in FAD, but do not prove that a gain-of-function mechanism or that  $A\beta$  is central to the aetiology. Meanwhile, no known mutations in the  $\alpha$ - or  $\beta$ -secretases appear to associate with FAD,<sup>92</sup> despite the crucial roles these two enzymes play in determining whether  $A\beta$  is formed. Mutations are found only in PS1, the catalytic subunit of  $\gamma$ -secretase making the final cleavage in the production of  $A\beta$  but also cleaving more than a hundred other substrates.<sup>118,329</sup> If  $A\beta$  (or an associated cleavage product of APP) were central to AD pathogenesis, one would expect FAD mutations in  $\alpha$ - and  $\beta$ -secretases that control the non-amyloidogenic versus amyloidogenic pathways to modulate the risk of AD.

### Absence of mutations in key amyloid- $\beta$ degrading metalloproteases

If  $A\beta$  overload and aggregation were critical to AD aetiology, one would expect hypomorphic mutations in the main proteases responsible for degrading  $A\beta$  intracellularly and in the extracellular matrix<sup>330</sup> to confer risks of AD, yet this is not the case to any large extent.<sup>92</sup> These proteases include, for example insulin-degrading enzyme,<sup>331,332</sup> neprilysin<sup>333–335</sup> and other zinc proteases.<sup>57,333</sup> The absence of such mutations suggests that impaired clearance of  $A\beta$  is not a substantial problem, supporting that  $A\beta$  has either a spectator, minor or even a loss-of-function role in AD.<sup>125,136</sup>

### Most familial Alzheimer's disease mutations reduce amyloid- $\beta$ due to loss-of-function

For a long time, a cornerstone of the ACH was that FAD mutations lead to more  $A\beta$  (e.g. the Swedish mutation<sup>336</sup>). However, most FAD-causing PS1 variants are hypomorphic, displaying impaired enzyme activity<sup>123,158,279</sup> (as expected on evolutionary grounds, as the enzyme is already optimized), yet often increase the  $A\beta_{42}/A\beta_{40}$  ratio.<sup>134,159,337–339</sup> These hypomorphic phenotypes of PS1/PS2 FAD mutants<sup>134,159,337–339</sup> substantially challenge a gain-of-function hypothesis.<sup>204</sup> The Fit-Stay-Trim (FIST) model of  $\gamma$ -secretase<sup>338,340–342</sup> explains why the loss-of-function increases the  $A\beta_{42}/A\beta_{40}$  ratio: the PS1 transmembrane helix 'fingers' produce two conformation states, a semi-open conformation with good affinity for APP-C99, maximal staying time and trimming to shorter  $A\beta$  peptides, and an open conformation with looser binding, less staying time, less and inaccurate cleavage and higher  $A\beta_{42}/A\beta_{40}$  ratios.<sup>67,159,338,342,343</sup> Despite high heterogeneity in clinical outcomes, assayed  $A\beta_{42}/A\beta_{40}$  ratios of mutants<sup>159</sup> seem to correlate with the age of symptom onset for carriers of the PS1 mutations.<sup>344</sup> This could imply that impaired presenilin/ $\gamma$ -secretase function may be clinically significant, as suggested by adverse effects of  $\gamma$ -secretase inhibitors,<sup>5,345</sup> and the presenilin hypothesis.<sup>208,346–348</sup> An inhibition of function by FAD mutations could affect both  $\gamma$ -secretase-independent presenilin processes<sup>349–354</sup> and the  $\gamma$ -secretase cleavage of many substrates involved in neuronal homeostasis.<sup>114,284,329,355</sup>

### Familial Alzheimer's disease mutations suggest that presenilin/ $\gamma$ -secretase could be central

FAD mutations are more common in PS1/PS2 than in APP, and PS1 mutations are often more severe.<sup>76</sup> This suggests that PS1/ $\gamma$ -secretase function could be important to AD pathogenesis.<sup>208,348,356</sup> Although the ACH is supported by FAD mutations,<sup>162</sup>

many of them have insignificant or unclear effects on APP-C99 processing.<sup>158,159</sup> If APP-C99 processing were central, one would expect mutations in all secretases and arguably in other parts of APP not directly related to PS1 interaction. In contrast, most FAD mutations scatter across PS1 and tend to impair protein compactness and stability,<sup>338</sup> which explains their effect on APP-C99 cleavage.<sup>342</sup> The (much fewer) APP mutations are all located in areas that involve PS1 engagement. A slow membrane protease, like  $\gamma$ -secretase, occupied with more APP-C99, could spend less time on other substrates, for example, Notch.<sup>357</sup> Thus, we need to explore whether FAD APP mutations work by disrupting PS1 function rather than the other way around. The original paper on the ACH also discussed the possible impact of other APP cleavage products than  $A\beta$ .<sup>10</sup> For example, soluble APP fragment (sAPP) has important functions that could be affected by aberrant APP cleavage.<sup>358</sup>

### The heterogeneous APP mutation effects

A central support for the ACH is the existence of APP mutations that cause FAD, located in the neighbourhood of (e.g. the Swedish and London mutations) or within the  $A\beta$  (e.g. the Dutch and Italian mutation) portion of APP.<sup>131</sup> The Swedish double mutation produces very high  $A\beta$  levels and has been much used in AD mouse models as it fits the 'overload' concept of the early hypothesis well.<sup>193</sup> Correspondingly, the protective A2T variant (using  $A\beta$  sequence numbering)<sup>359</sup> was taken as a strong support of the ACH,<sup>153</sup> as a useful counter-example to the Swedish mutation and the same-site pathogenic substitution A2V.<sup>360</sup> However, these examples are not representative of the FAD mutations in APP, whose effects are extremely heterogeneous.<sup>131</sup> In response to the observation of many FAD phenotypes with lower overall amyloid production, the emphasis turned to the  $A\beta_{42}/A\beta_{40}$  ratio.<sup>209–212</sup> While some APP mutations increase this ratio, others seem to lower it. A2V and D7N lead to  $A\beta_{42}/A\beta_{40}$  ratios similar to wild-type, but E22G, E22K and E22Q lower the  $A\beta_{42}/A\beta_{40}$  ratio.<sup>131</sup> The London mutation causes more severe (earlier onset) disease despite producing much less  $A\beta$  than the Swedish mutation.<sup>188</sup> Some variants have high  $A\beta$  aggregation tendencies, others low. Some mutations are more toxic in cell assays than the wild-type, others not significantly so.<sup>131</sup> Accordingly, the Swedish mutation used in much AD research is not representative of FAD APP mutation phenotypes broadly.<sup>131</sup>

### Accounting for the physiological regulators of amyloid- $\beta$ balance

As the ACH revolves around  $A\beta$  production, degradation, homeostasis and aggregation, it would be reasonable to include biochemical regulators of this balance. As one well-documented group of regulators, metal ions  $Cu^{2+}$  and  $Zn^{2+}$  bind naturally to APP and  $A\beta$  in high-affinity sites and modulate the aggregation and production of  $A\beta$ ,<sup>120,213–218</sup> as documented in many reviews.<sup>57,59,122,219–225</sup> As mentioned above, no FAD mutations are found in the  $A\beta$ -degrading enzymes, but furthermore, these proteases are typically zinc proteases, indicating perhaps a role of zinc in regulating these pathways.<sup>57,361,362</sup> It would seem natural to include these regulators, and possibly others, in the disease models.<sup>57,121,231,363,364</sup>

### Interpretation of Down syndrome dementia as due to amyloid overload

One of the early claims of evidence in favour of the ACH<sup>9</sup> was the development of dementia and plaque burden in patients with Down syndrome, which have three copies of chromosome 21 where



the APP gene is situated.<sup>365</sup> However, there are more than 200 genes on chromosome 21, including BACE2 and APP but also the gene coding for Cu, Zn-superoxide dismutase 1 (SOD1) whose mutations cause familial amyotrophic lateral sclerosis,<sup>366,367</sup> proteostasis genes, calcium-binding proteins and ion channels important to neurological homeostasis; all these genes may be upregulated in a process that could give dementia in trisomy-21 by many pathways. A proposed alternative explanation is that the additional chromosome (or parts of it) exhausts proteostasis in neurons,<sup>44</sup> accelerating disease processes that lead to neurodegeneration by proteostatic dysfunction.<sup>368,369</sup> In this case, aberrant amyloid deposition would be a marker of such dysfunction, not the primary cause of dementia.

### Amyloid- $\beta$ antibodies: small benefits with adverse effects

The ultimate test of any disease mechanism is the development of treatments based on it, but this approach has repeatedly failed.<sup>5,90–92,189,191,227,228,370</sup> High profile antibodies before lecanemab, such as solanezumab<sup>151,371</sup> and bapineuzumab,<sup>313</sup> did not show cognitive benefit<sup>151</sup> and produced adverse effects<sup>150</sup> despite lowering A $\beta$  levels as assessed by changes in CSF.<sup>372,373</sup> As with aducanumab, subsequent *post hoc* analysis of solanezumab data was interpreted as possible clinical benefit,<sup>374,375</sup> which led to an expensive extra trial (EXPEDITION 3) before giving up.<sup>376,377</sup> This has been explained by suggesting that the trials have been incorrectly designed and targeted irrelevant forms of A $\beta$  too late in the disease process.<sup>189,226,232</sup> Yet, many trials targeting MCI also failed,<sup>139,378</sup> and a 2019 report of autopsy follow-up performed 4 months to 14 years after the first active A $\beta$  immunization showed plaque removal but progression to severe dementia in most AD patients.<sup>228</sup> In March 2023, it was announced that solanezumab targeting pre-symptomatic subjects only showing PET-evidence of brain amyloidosis (the A4 study)<sup>379</sup> did not slow cognitive decline. In trials specifically addressing the most direct genotype-phenotype relation of the hypothesis via studying FAD patients, antibodies gantenerumab or solanezumab, despite targeting A $\beta$ , also failed to produce clinical effect versus placebo; solanezumab in some patients generated even worse outcomes than placebo.<sup>380</sup>

Also, amyloid-related imaging abnormalities (ARIA) in monoclonal antibody trials are a major concern.<sup>150</sup> When administering aducanumab, ARIA affected 41% of participants versus 10% in the placebo group, including microhaemorrhages in 19% (placebo group: 7%) and siderosis in 15% of patients (placebo group: 2%).<sup>381</sup> ARIA was consistently more frequent in the high-dose group, indicating a dose-dependent adverse effect of the highly specific A $\beta$  antibody.<sup>381</sup> ApoE  $\epsilon$ 4 carriers had these adverse effects twice as often as non-carriers.<sup>381</sup> Gantenerumab also showed no effect but led to vasogenic oedema or inflammation in some patients,<sup>382</sup> and ARIA risk was higher for APOE  $\epsilon$ 4 genotypes in the gantenerumab<sup>312</sup> and bapineuzumab trials.<sup>313</sup> It also speaks against a strong disease-modifying role that while ARIA follows a clear dose-response relationship with biomarker changes,<sup>151,381</sup> the dose-response correlation for clinical benefits on cognition in the same trials tend to be weaker.

### Blocking amyloid- $\beta$ producing enzymes often impairs cognition

While  $\beta$ - and  $\gamma$ -secretase inhibitors failed to show clinical effects, they often caused adverse effects,<sup>139</sup> which could be due to the inhibition of action towards other substrates or other functional APP cleavage products such as sAPP,<sup>383,384</sup> with A $\beta$  deposition being a correlating observation. Blocking A $\beta$  producing enzymes

commonly impaired cognition,<sup>89,139</sup> raising the possibility that normal protein functions need to be preserved.<sup>85,123,197,229</sup> Recent Bayesian analysis of historic anti-amyloid trials indicates strong evidence for a null hypothesis.<sup>227,385</sup> We note that this analysis included trials without precise target engagement by biomarkers or advanced stage patients for whom treatment may come too late.<sup>386</sup> However, even for the newer monoclonal antibody trials of donanemab, aducanumab and lecanemab with highly selected early-disease cohorts and robust biomarker data, the benefits were below the threshold typically considered clinically meaningful<sup>387,388</sup> and the risk/benefit ratio remained uncertain.<sup>389</sup>

### Amyloid-PET as a diagnostic tool

Concerns have been raised that surrogate-based approvals without clear clinical benefit<sup>13</sup> could negatively impact dementia research<sup>94,234</sup> and efforts to combat other diseases such as cancer.<sup>390</sup> Yet, the field increasingly focuses on amyloid removal as a surrogate of treatment benefit. This is potentially a concern because the ACH has not yet been proven true, and because techniques used to measure amyloid have limitations: the heterogeneous and modest specificity,<sup>299</sup> differences between tracers, and large non-specific PET signals raise questions about the appropriate use and limitations of amyloid PET, not only for diagnostic purposes, but also in therapeutic trials.<sup>153,203,233,292,293</sup> Amyloid-PET currently has limited ability to detect amyloid over time in relation to disease course and post-therapy evaluations.<sup>294,391,392</sup> There are concerns about the spatial resolution being too low to identify low concentration plaque in early disease<sup>203,291,293</sup> and tracer uptake in white matter, spill-over and partial volume effects may complicate analysis.<sup>291–293,393</sup>

PET mainly targets fibrillar  $\beta$ -sheet structures and is unlikely to measure the soluble pathogenic forms of A $\beta$ <sup>168,207,394</sup> that may vary in morphology and contain less  $\beta$ -sheet structure.<sup>292</sup> Reduced uptake of tracers can also be due to many other causes than removal of plaque deposits, including morphological changes or changing the conformation of the plaques.<sup>293,294,395</sup> Combined with the substantial indication of no clinical effect of many other immunizations,<sup>227</sup> FDA approval of a surrogate PET marker with a median specificity of 58% according to the Cochrane topic review<sup>299</sup> seems erroneous due to suboptimal specificity of the surrogate<sup>293,395</sup> and the underlying causal premise that it reflects disease state.<sup>155–157,203,304,305</sup>

### Aducanumab: a case study of unmaterialized hope

The focus on amyloid-PET as a surrogate for determining disease status and assessing the clinical benefit of anti-amyloid therapies has especially raised concern in relation to aducanumab.<sup>13</sup>

Aducanumab is a monoclonal antibody that selectively targets A $\beta$  oligomers over monomers *in vitro*, and it is hoped, also *in vivo*.<sup>396</sup> The rationale was to reduce the pathogenicity of oligomers and the seeding by oligomers of fibrillar plaques.<sup>397</sup> Results of early aducanumab trials were promising as the drug significantly reduced the amount of brain amyloid depositions.<sup>398,399</sup> The following two, nearly identical, phase 3 trials, named EMERGE and ENGAGE, yielded contradictory clinical results (little versus no effect) and were halted based on futility analysis from the first half of the enrolled patients as announced on 21 March 2019.<sup>400,401</sup> However, on 22 October 2019, Biogen announced that *post hoc* analysis of trial data<sup>315</sup> indicated that EMERGE met its primary clinical end point for more prolonged exposures of aducanumab, while ENGAGE did not. Based on this, Biogen sought FDA approval.<sup>402,403</sup> FDA then approved aducanumab as Aduhelm via the accelerated



approval pathway, based on reduction in amyloid plaques as a surrogate of clinical effect despite weak evidence of such,<sup>404</sup> a decision that caused widespread controversy.<sup>13,14,405–408</sup>

The move by FDA executives to use amyloid-PET tracer uptake as a surrogate of clinical efficacy assumes that the ACH has been proven. The surrogate claim has never been tested scientifically as it was not submitted to the advisory panel.<sup>13,409,410</sup> Setting aside the limitations of measuring amyloid and the dispute over the ACH itself, the amyloid-clinical relationship is well accepted as being confounded by additional interactions such as APOE  $\epsilon$ 4 genotype and tau pathology,<sup>154,184</sup> i.e. FDA's claim that reduced amyloid-PET tracer uptake is a surrogate of clinical effect lacks scientific support.

### Lecanemab: promise with limitations

Lecanemab,<sup>411</sup> a humanized form of the mAb158 mouse antibody attacking protofibrils in mouse models,<sup>412</sup> although not meeting its primary end points in phase 2, showed promising clinical results in follow-up analysis.<sup>411</sup> Lecanemab was made specific towards the Arctic APP mutation (E22G) that presumably forms protofibrils very quickly and seems to bind protofibril more strongly than aducanumab.<sup>413</sup> The published 18-month study trial data<sup>16</sup> indicated a statistically significant clinical effect on several end points. A significant outcome was observed for the primary end point [change in the 18-point Clinical Dementia Rating–Sum of Boxes (CDR-SB) score] and for secondary end points, such as the Alzheimer's Disease Assessment Scale (ADAS-cog14).<sup>16</sup>

In terms of CDR-SB scores, patients receiving lecanemab declined by 1.21 points, while patients receiving placebo declined by 1.66 points. This is a difference of 0.45 on an 18-point scale (27% relative effect but only 2–3% absolute effect). For the ADAS-cog14 outcomes, the difference was –1.44.<sup>16</sup> Although statistical significance was maintained or even expanded at late time points, this effect might reverse or reduce after the trial period. Whether the observed 0.45-point improvement compared to controls translates into a clinically meaningful difference is uncertain<sup>414</sup>: establishing clinical meaningful changes can be tricky, but for the primary end point CDR-SB score a cut-off with real-world clinical implications has been estimated to be ~1.<sup>387,388</sup> Moreover, lecanemab is not superior to the acetylcholinesterase inhibitor donepezil, which for example, has shown a 2-point difference versus placebo on the ADAS-cog scale,<sup>415</sup> despite being symptomatic rather than amyloid-modifying.

Lecanemab was also estimated to provide cognitive and functional symptom improvement corresponding to half a year disease progression based on data obtained during the trial.<sup>16</sup> However, this depends on whether the trajectory of any potential benefits continues after 18 months. Similarly, the cumulative adverse effects of ongoing treatment are unknown and could ultimately curtail treatment time.

Also of note are the very heterogeneous subgroup results. For some end points, simultaneous use of symptomatic medicine gave as large an effect as lecanemab. Males had much more benefit across all end points, whereas females exhibited little benefit (12% for the primary end point), a large and unexplained effect. Also, lecanemab showed by far the smallest effect in younger patients (6% for the primary end point in < 65 years), in contrast to the assumption that the drugs must be applied early to work. Data for composite groups (e.g. young females, female APOE  $\epsilon$ 4 carriers) were missing, but the effects separately suggest that efficacy could approach zero in many groups. The lower benefit to APOE  $\epsilon$ 4 carriers is unexplained. This heterogeneity is important for labelling of the drug (as some patients may have no benefit) and for illustrating

that population covariates affect disease development as much as Lecanemab itself.

Lecanemab was originally welcomed as associated with relatively less adverse effects than some other antibodies,<sup>411</sup> but 12.6% of treated patients still displayed ARIA.<sup>16</sup> Also, although analysis was not described in detail in the paper,<sup>16</sup> previous MRI data indicated consistent brain volume changes.<sup>411</sup> Of concern, during the trial period, at least two deaths have been reported.<sup>416,417</sup> The lecanemab study indicated similar death rates in the treatment and placebo group (one more in the latter) and no deaths associated with ARIA,<sup>16</sup> but independent autopsies<sup>417</sup> indicated otherwise, so careful investigation of these red flags is needed. A third death indicates a risk associated with the use of anticoagulants (heparin) simultaneously with lecanemab administration.<sup>418</sup>

Although the authors did perform sensitivity analysis for this possibility,<sup>16</sup> unblinding of patients due to ARIA protocols could potentially cause bias in the cognitive performance scores that would correspond to a weakening of the clinical disease progression delay.<sup>419</sup> Of further interest are the adverse events that caused participants to drop out of treatment. This number was twice as large in the treatment group compared to the placebo group,<sup>16</sup> and the outcomes of these patients need to be analysed in total together with the long-term outcomes of participants, as essential information for the treatment perspective. The efficacy was evaluated against a consistently reducing cohort during the trial period, not the final cohort at 18 months, which may also produce bias.<sup>420</sup>

Finally, it is very important to understand the mechanisms behind both the beneficial and adverse effects (ARIA<sup>381,421</sup>) on the molecular level. It is, for example, possible that the benefits are not due to removal of oligomers or protofibrils *per se*, but for example, reconstitution of functional monomers<sup>123</sup> or A $\beta$ -independent pathways, depending on the molecular affinity and specificity of the antibodies versus other molecular targets. A $\beta$ <sub>42</sub> increased substantially in the CSF due to lecanemab treatment,<sup>16</sup> which for a given level of amyloidosis has recently been associated with a net positive clinical effect by itself,<sup>125,126</sup> consistent with loss-of-function aetiologies of the disease.<sup>84,86,136,188,422</sup> Thus, the contributions of beneficial and harmful forms of A $\beta$  need attention when dissecting the information provided by the data. The clinical effects were not correlated with the change in amyloid load and were much more modest,<sup>16</sup> i.e. only an absolute 2–3% clinical effect as elaborated above.

### Donanemab: debated efficacy

Donanemab is another monoclonal antibody developed by Eli Lilly that also bears some promise in recent trials.<sup>423,424</sup> Like the trials of lecanemab and aducanumab, the donanemab trial used early-disease cohorts and consistent use of biomarker data. Mintun *et al.*<sup>423</sup> indicated a borderline significant outcome for their primary end point (the iADRS scale<sup>425</sup>), which is a composite of two of the other four scores used as secondary end points [CDR-SB, ADAS-Cog13, ADCS-iADL and Mini-Mental State Examination (MMSE)] in none of which the change after 76 weeks was significant. The change in the primary end point iADRS score after 76 weeks was –6.86 versus –10.06 with placebo, i.e. a difference of 3.2 (equal to 3.2/149 or 0.22%). However, clinically meaningful changes in iADRS are estimated to be >5 for MCI and >9 for mild AD dementia.<sup>426</sup>

At the same time, treatment-related ARIA-E and ARIA-H were frequent in treated versus placebo patients: 26.7% versus 0.8% and 8.4% versus 3.2%, respectively. As with lecanemab, unexplained brain volume changes and possible atrophy associated

with treatment are of concern and need to be understood,<sup>423</sup> noting a new meta-analysis that finds these effects to be systematic and significant.<sup>427</sup>

Also, the small (below clinically meaningful) benefits may be affected by potential biases from unblinding due to ARIA protocols<sup>419</sup> and differential dropout biases.<sup>420</sup> However, these effects are all below the threshold typically considered clinically meaningful<sup>387,388</sup> and the risk/benefit ratio remains uncertain.<sup>389</sup> Recently, the FDA declined accelerated approval for donanemab due to too few patient data, but traditional approval will likely be sought by Lilly based on the confirmatory phase 3 TRAILBLAZER-ALZ 2 trial later in 2023.<sup>428</sup> However, as for aducanumab, the cost-effectiveness of donanemab for treating mild AD is uncertain and plausibly unfavourable,<sup>429</sup> and the current debate on the monoclonal antibody trials<sup>389,414,430–432</sup> is likely to continue. The additional TRAILBLAZER-ALZ trial results coming over the next years will add useful insight to the current debate on monoclonal antibody treatments.

## Important considerations regarding anti-amyloid therapies

It is difficult to ascertain the extent to which amyloid removal can be considered beneficial, given the early data from the clinical trials. While currently approved treatments, such as donepezil, show benefits in clinical studies,<sup>415,433</sup> improvements in clinical practice are limited and even then, last for a limited time. The benefit of A $\beta$  antibodies should be measured against the same real-world benchmarks, considering their significant adverse effects in some patients and the fact that they seem to work in distinct subsets of patients only, and the modest effects of these drugs may not be cost-effective for early AD.<sup>429</sup> We need more work to determine the reproducibility of results, alerted by the different aducanumab results in the parallel, similarly designed EMERGE and ENGAGE trials.<sup>401,406</sup> We should also work to understand the mechanisms behind the adverse effects (ARIA) and the paradoxical brain volume changes and concern about atrophy triggered by these compounds,<sup>427</sup> as well as the specific mechanisms behind any potential clinical benefits.

There will be pressure to apply the drugs well outside their tested parameters, and there is concern that the risk-benefit analysis would not favour such unfettered use.<sup>429,434</sup> The patients in the CLARITY trial<sup>16</sup> were a carefully selected and followed cohort, and excluded the dropouts noted above, arguably increasing the tendency towards a positive benefit in the trial. The field will need to carefully follow real-world clinical experience with these drugs, including the long-term risk profile and magnitudes of meaningful, beneficial impact on specific patient groups with diverse real-life experiences of dementia. Caution on the implementation of these drugs into practice is warranted until these analyses have been generated.

In summary, it would be unwarranted at this stage to double down on the ACH based on the current data. We are concerned that enormous pressure is leading to fast-track approval of anti-amyloid therapies, fuelled by a view that A $\beta$  is synonymous with dementia in AD, as implied by the FDA's scientifically untested adoption of the amyloid-PET surrogate for clinical efficacy.<sup>13,295</sup> We do not yet know the long-term effects of lecanemab use (positive or negative) or whether effects persist or revert after the drug is ceased. We recommend follow-up of patients enrolled in these trials over the next decade and careful assessments of real-world use, to address these questions.

## Moving forward: options for integrating theories and therapies

Lack of therapeutic success in AD and other major neurodegenerative disorders such as Parkinson's disease and amyotrophic lateral sclerosis may relate to the absence of convergent molecular models of disease and strict definitions of disease criteria, and the lack of clear distinction between pathology and pathogenicity. No monocausal treatment has ever produced disease reversal, stagnation or even general patient-wide robust clinical effects.<sup>227</sup>

It is crucial to acknowledge the historic importance of the ACH in establishing AD as a single disorder that may be targeted therapeutically.<sup>89,94</sup> The hypothesis followed a series of exceptional observations, discoveries and reasoning. It was supported by the best available evidence at the time.<sup>11</sup> However, the data reviewed above suggest a need to replace the ACH with a more complex hypothesis integrating the many polygenic, epigenetic, environmental, vascular, neuroinflammatory and metabolic factors, ideally in more predictive holistic models.<sup>154,185</sup> The misfolding of proteins may be a sign, not always cause, of molecular abnormalities.<sup>94,136,218</sup> More realistic models that acknowledge the many modulators of the disease<sup>44,154,185</sup> yield room for individual disease trajectories based on the patient's personal aggregate risk.

Understanding the function of APP is essential, as it may increase our understanding of the main pathways and mechanisms involved in FAD. The function of presenilins is important for lysosome proteolysis. Notably, PS1 mutations impair this function<sup>435</sup> and functions of the vascular system.<sup>436</sup> The accumulation of undegraded waste proteins and peptides resembles the pathology of lysosomal storage diseases that would assign senile plaques to a dysfunction of the proteasome and lysosome,<sup>110,437</sup> with a plausible role of presenilins in controlling calcium access to the lysosomes and preserving proteostasis competency.<sup>435,437,438</sup>

We should consider that patients exhibit individual, overlapping but heterogeneous disease trajectories.<sup>49,89,92,94,136</sup> Ignoring these multiple factors and pathways in favour of magic 'silver bullets' is detrimental. We urge great care and caution in considering the whole individual and reiterate the need for developing and applying individualized care and treatment programmes for patients.

The next decade will be an exciting time to be in AD research as we work towards new understandings and effective therapeutics. Efforts are now required to establish the molecular granularity and aetiological heterogeneity but relatedness of condition subtypes as framing the complexity of the disease, including both loss- and gain-of-function pathways,<sup>19,123,422,439</sup> with ageing being a main factor of pathogenicity.<sup>57,89,205</sup>

Hypotheses relating to infections, vascular disease and dysmetabolism must be explored further as possible drivers, among others.<sup>266,275,440</sup> In these scenarios amyloid may, for example, be a marker of increased APP turnover, as a response to insult rather than a disease driver. Importantly, a recent trend in drug pipelines towards more diverse disease pathways<sup>82</sup> lends promise of more complex and novel future applications of treatment regimes.

The ageing human proteome features reduced gene expression related to synaptic function, calcium homeostasis and vesicular transport but increased expression of genes involved in inflammation and stress tolerance,<sup>441</sup> a human ageing phenotype largely absent in research models. Chemical ageing is relatively straightforward to include in preclinical models and is known to modulate A $\beta$  production and clearance and aggregation.<sup>57,219,442</sup>

On the regenerative front, promising areas are emerging. Converging data support a role for insulin signalling in

dementia.<sup>443,444</sup> These pathways offer opportunities to integrate metabolic pathways with other protein-specific pathways. For instance, glucagon-like peptide-1 receptor (GLP-1R) analogues have been tested in preclinical models and clinical trials.<sup>445,446</sup> Additional therapeutic venues, ideally in combination, can be expected from non-neuronal targets like neuroinflammation and the microglia-astrocyte axis.<sup>26,440</sup>

Finally, we envision that data-backed artificial intelligence and systems biology tools can help address the challenge of unravelling personalized disease trajectories. These approaches have helped define degenerative subprocesses inside and outside the CNS. For instance, growing evidence supports a gut-brain connection,<sup>447,448</sup> microbiota's role in neurodegeneration,<sup>266,275</sup> and the role of lipid and energy-related dysmetabolism in AD.<sup>40,44,185,449</sup> Artificial intelligence may combine the various personal-medicine trajectories and account for the biochemical and clinical heterogeneity in perhaps more complete, realistic and predictive AD models.

## Concluding remarks

Motivated by the recent trials of monoclonal antibodies against AD, we have reviewed the history of the ACH and genetic, biochemical, biomarker and clinical data underlying it. We suggest that these data show that A $\beta$ , while important in AD pathology, plays a minor role in the aetiology and thus treatment of the disease. Despite decades of research, the normal functions of APP and A $\beta$  and the precise pathogenic mechanism and structure of A $\beta$  have not been established. Whether clinical trials produce positive or adverse results, we are largely in the dark regarding the molecular underpinnings of these outcomes.

Anti-amyloid therapies may at best support a notion that the amyloid cascade participates in some pathologically relevant processes of AD. However, A $\beta$ -pathways as targeted in the current form by antibodies in no way account for the aetiological complexity and real-world clinical needs of patients, and thus AD is better described within a modern network-based anti-reductionist disease view.<sup>450</sup>

To resolve the current therapeutic impasse, efforts are required to explore the molecular granularity of condition subtypes to capture the full complexity of the disease and the many disease trajectories and converging biochemical pathways and complex cellular interactions in the brain. We envision that artificial intelligence-aided approaches along with systems biology and precision medicine can help to refocus our attention toward generating a significant impact on the lives of the millions of patients affected by this terrible disease.

## Funding

This work did not receive any specific grant from funding agencies in the public, commercial or not-for-profit sectors. B.V. is supported by the Boyarsky, Roth, Perlstein, Battersby and Howland-Rose Families, who pose no conflict of interest. S.L.S. has received grant support from the Alzheimer's Association, The Italian Ministry of Health, and the Italian Ministry of Research.

## Competing interests

The authors report no competing interests.

## References

1. Blennow K, de Leon MJ, Zetterberg H. Alzheimer's disease. *Lancet*. 2015;368:387–403.
2. Masters CL, Bateman R, Blennow K, Rowe CC, Sperling RA, Cummings JL. Alzheimer's disease. *Nat Rev Dis Prim*. 2015;1:15056.
3. Nichols E, Szeke CEI, Vollset SE, et al. Global, regional, and national burden of Alzheimer's disease and other dementias, 1990–2016: A systematic analysis for the Global Burden of Disease Study 2016. *Lancet Neurol*. 2019;18:88–106.
4. Bäckman K, Joas E, Waern M, et al. 37 Years of body mass Index and dementia: Effect modification by the APOE genotype: Observations from the prospective population study of women in Gothenburg, Sweden. *J Alzheimer's Dis*. 2015;48:1119–1127.
5. Karran E, Hardy J. A critique of the drug discovery and phase 3 clinical programs targeting the amyloid hypothesis for Alzheimer disease. *Ann Neurol*. 2014;76:185–205.
6. Querfurth HW, LaFerla FM. Alzheimer's disease. *N Engl J Med*. 2010;362:329–344.
7. Wong CW, Quaranta V, Glenner GG. Neuritic plaques and cerebrovascular amyloid in Alzheimer disease are antigenically related. *Proc Natl Acad Sci U S A*. 1985;82:8729–8732.
8. Glenner GG, Wong CW. Alzheimer's disease: Initial report of the purification and characterization of a novel cerebrovascular amyloid protein. *Biochem Biophys Res Commun*. 1984;120:885–890.
9. Masters CL, Simms G, Weinman NA, Multhaup G, McDonald BL, Beyreuther K. Amyloid plaque core protein in Alzheimer disease and down syndrome. *Proc Natl Acad Sci U S A*. 1985;82:4245–4249.
10. Hardy JA, Higgins GA. Alzheimer's disease: The amyloid cascade hypothesis. *Science*. 1992;256:184–185.
11. Hardy J. Alzheimer's disease: The amyloid cascade hypothesis —An update and reappraisal. *J Alzheimer's Dis*. 2006;9:151–153.
12. Mullard A. FDA Approval for biogen's aducanumab sparks Alzheimer disease firestorm. *Nat Rev Drug Discov*. 2021;20:496.
13. Alexander GC, Knopman DS, Emerson SS, et al. Revisiting FDA approval of aducanumab. *N Engl J Med*. 2021;385:769–771.
14. Perlmutter JS. FDA's green light, science's red light. *Science*. 2021;372:1371.
15. Tagliaianni F, Tiraboschi P, Federico A. Alzheimer's disease: The controversial approval of aducanumab. *Neurol Sci*. 2021;42:3069–3070.
16. van Dyck CH, Swanson CJ, Aisen P, et al. Lecanemab in early Alzheimer's disease. *N Engl J Med*. 2023;388:9–21.
17. Mahase E. Lecanemab trial finds slight slowing of cognitive decline, but clinical benefits are uncertain. *BMJ*. 2022;379:o2912.
18. Collins TR. Neurologists react to lecanemab news with swirl of excitement. *Caution. Neurol Today*. 2022;22:5–6.
19. Neve RL, Robakis NK. Alzheimer's disease: A re-examination of the amyloid hypothesis. *Trends Neurosci*. 1998;21:15–19.
20. Karantzoulis S, Galvin JE. Distinguishing Alzheimer's disease from other major forms of dementia. *Expert Rev Neurother*. 2011;11:1579–1591.
21. Boyle PA, Yu L, Wilson RS, Leurgans SE, Schneider JA, Bennett DA. Person-specific contribution of neuropathologies to cognitive loss in old age. *Ann Neurol*. 2018;83:74–83.
22. Boyle PA, Yu L, Leurgans SE, et al. Attributable risk of Alzheimer's dementia attributed to age-related neuropathologies. *Ann Neurol*. 2019;85:114–124.
23. Holmes C. Genotype and phenotype in Alzheimer's disease. *Br J Psychiatry*. 2002;180:131–134.
24. Tanzi RE. The genetics of Alzheimer disease. *Cold Spring Harb Perspect Med*. 2012;2:a006296.
25. Carreiras M, Mendes E, Perry M, Francisco A, Marco-Contelles J. The multifactorial nature of Alzheimer's disease for



- developing potential therapeutics. *Curr Top Med Chem*. 2013;13:1745–1770.
26. Heneka MT, Carson MJ, El Khoury J, et al. Neuroinflammation in Alzheimer's disease. *Lancet Neurol*. 2015;14:388–405.
  27. Tanzi RE, Bertram L. Twenty years of the Alzheimer's disease amyloid hypothesis: A genetic perspective. *Cell*. 2005;120:545–555.
  28. Bramblett GT, Goedert M, Jakes R, Merrick SE, Trojanowski JQ, Lee VMY. Abnormal tau phosphorylation at Ser396 in Alzheimer's disease recapitulates development and contributes to reduced microtubule binding. *Neuron*. 1993;10:1089–1099.
  29. Augustinack JC, Schneider A, Mandelkow EM, Hyman BT. Specific tau phosphorylation sites correlate with severity of neuronal cytopathology in Alzheimer's disease. *Acta Neuropathol*. 2002;103:26–35.
  30. Avila J. Tau phosphorylation and aggregation in Alzheimer's disease pathology. *FEBS Lett*. 2006;580:2922–2927.
  31. Arai T, Mackenzie IRA, Hasegawa M, et al. Phosphorylated TDP-43 in Alzheimer's disease and dementia with Lewy bodies. *Acta Neuropathol*. 2009;117:125–136.
  32. Higashi S, Iseki E, Yamamoto R, et al. Concurrence of TDP-43, tau and  $\alpha$ -synuclein pathology in brains of Alzheimer's disease and dementia with Lewy bodies. *Brain Res*. 2007;1184:284–294.
  33. Twohig D, Nielsen HM.  $\alpha$ -synuclein in the pathophysiology of Alzheimer's disease. *Mol Neurodegener*. 2019;14:1–19.
  34. Pappolla MA, Omar RA, Kim KS, Robakis NK. Immunohistochemical evidence of oxidative [corrected] stress in Alzheimer's disease. *Am J Pathol*. 1992;140:621.
  35. Hoyer S. Oxidative metabolism deficiencies in brains of patients with Alzheimer's disease. *Acta Neurol Scand*. 1996;94(S165):18–24.
  36. Nunomura A, Perry G, Aliev G, et al. Oxidative damage is the earliest event in Alzheimer disease. *J Neuropathol Exp Neurol*. 2001;60:759–767.
  37. Gibson GE, Huang H-M. Oxidative stress in Alzheimer's disease. *Neurobiol Aging*. 2005;26:575–578.
  38. Zhang F, Jiang L. Neuroinflammation in Alzheimer's disease. *Neuropsychiatr Dis Treat*. 2015;11:243–256.
  39. Vilalta A, Brown GC. Neurophagy, the phagocytosis of live neurons and synapses by glia, contributes to brain development and disease. *FEBS J*. 2018;285:3566–3575.
  40. Hoyer S. Brain glucose and energy metabolism abnormalities in sporadic Alzheimer disease. Causes and consequences: An update. *Exp Gerontol*. 2000;35(9–10):1363–1372.
  41. Razay G, Raza G, Vreugdenhil A. Obesity in middle age and future risk of dementia: Midlife obesity increases risk of future dementia. *Br Med J*. 2005;331:455.
  42. Diaz R. Obesity: Overweight as a risk factor for dementia. *Nat Rev Endocrinol*. 2009;5:587.
  43. Dahl AK, Hassing LB. Obesity and cognitive aging. *Epidemiol Rev*. 2013;35:22–32.
  44. Kepp KP. A quantitative model of human neurodegenerative diseases involving protein aggregation. *Neurobiol Aging*. 2019;80:46–55.
  45. Robbins J, Busquets O, Tong M, de la Monte SM. Dysregulation of insulin-linked metabolic pathways in Alzheimer's disease: Co-factor role of apolipoprotein E  $\epsilon$ 4. *J Alzheimer's Dis reports*. 2020;4:479–493.
  46. Kalback W, Esh C, Castaño EM, et al. Atherosclerosis, vascular amyloidosis and brain hypoperfusion in the pathogenesis of sporadic Alzheimer's disease. *Neurol Res*. 2004;26:525–539.
  47. Yarchoan M, Xie SX, Kling MA, et al. Cerebrovascular atherosclerosis correlates with Alzheimer pathology in neurodegenerative dementias. *Brain*. 2012;135:3749–3756.
  48. Wendell CR, Waldstein SR, Ferrucci L, O'Brien RJ, Strait JB, Zonderman AB. Carotid atherosclerosis and prospective risk of dementia. *Stroke*. 2012;43:3319–3324.
  49. Høiland-Carlson PF, Revheim M-E, Alavi A. Alzheimer's disease at a crossroad: Time to part from amyloid to more promising aspects—Atherosclerosis for a start. *J Alzheimer's Dis*. 2022;88:455–458.
  50. Love S, Miners JS. Cerebrovascular disease in ageing and Alzheimer's disease. *Acta Neuropathol*. 2016;131:645–658.
  51. Ronnema E, Zethelius B, Lannfelt L, Kilander L. Vascular risk factors and dementia: 40-year follow-up of a population-based cohort. *Dement Geriatr Cogn Disord*. 2011;31:460–466.
  52. Thal DR, Griffin WST, de Vos RAI, Ghebremedhin E. Cerebral amyloid angiopathy and its relationship to Alzheimer's disease. *Acta Neuropathol*. 2008;115:599–609.
  53. Toda N, Ayajiki K, Okamura T. Obesity-Induced cerebral hypoperfusion derived from endothelial dysfunction: One of the risk factors for Alzheimer's disease. *Curr Alzheimer Res*. 2014;11:733–744.
  54. Bezprozvanny I, Mattson MP. Neuronal calcium mishandling and the pathogenesis of Alzheimer's disease. *Trends Neurosci*. 2008;31:454–463.
  55. Khachaturian ZS. Hypothesis on the regulation of cytosol calcium concentration and the aging brain. *Neurobiol Aging*. 1987;8:345–346.
  56. Smith IF, Green KN, LaFerla FM. Calcium dysregulation in Alzheimer's disease: Recent advances gained from genetically modified animals. *Cell Calcium*. 2005;38(3–4):427–437.
  57. Kepp KP. Bioinorganic chemistry of Alzheimer's disease. *Chem Rev*. 2012;112:5193–5239.
  58. Morris JK, Honea RA, Vidoni ED, Swerdlow RH, Burns JM. Is Alzheimer's disease a systemic disease? *Biochim Biophys Acta—Mol Basis Dis*. 2014;1842:1340–1349.
  59. Kepp KP, Squitti R. Copper imbalance in Alzheimer's disease: Convergence of the chemistry and the clinic. *Coord Chem Rev*. 2019;397:168–187.
  60. Corona C, Pensalfini A, Frazzini V, Sensi SL. New therapeutic targets in Alzheimer's disease: Brain deregulation of calcium and zinc. *Cell Death Dis*. 2011;2:e176.
  61. Beydoun MA, Beydoun HA, Gamaldo AA, Teel A, Zonderman AB, Wang Y. Epidemiologic studies of modifiable factors associated with cognition and dementia: Systematic review and meta-analysis. *BMC Public Health*. 2014;14:643.
  62. Mayeux R. Epidemiology of neurodegeneration. *Annu Rev Neurosci*. 2003;26:81–104.
  63. Bäckman L, Jones S, Berger AK, Laukka EJ, Small BJ. Multiple cognitive deficits during the transition to Alzheimer's disease. *J Intern Med*. 2004;256:195–204.
  64. Arnáiz E, Almkvist O. Neuropsychological features of mild cognitive impairment and preclinical Alzheimer's disease. *Acta Neurol Scand*. 2003;107:34–41.
  65. McKhann GM, Knopman DS, Chertkow H, et al. The diagnosis of dementia due to Alzheimer's disease: Recommendations from the national institute on aging-Alzheimer's association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's Dement*. 2011;7:263–269.
  66. Van Cauwenbergh C, Van Broeckhoven C, Sleegers K. The genetic landscape of Alzheimer disease: Clinical implications and perspectives. *Genet Med*. 2016;18(5):421–430. doi:10.1038/gim.2015.117
  67. Tang N, Dehury B, Kepp KP. Computing the pathogenicity of Alzheimer's disease presenilin 1 mutations. *J Chem Inf Model*. 2019;59:858–870.

68. Sherrington R, Rogaev EI, Liang Y, et al. Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease. *Nature*. 1995;375:754–760.
69. George-Hyslop PS, Haines J, Rogaev E, et al. Genetic evidence for a novel familial Alzheimer's disease locus on chromosome 14. *Nat Genet*. 1992;2:330–334.
70. Van Broeckhoven C, Backhovens H, Cruts M, et al. Mapping of a gene predisposing to early-onset Alzheimer's disease to chromosome 14q24.3. *Nat Genet*. 1992;2:335–339.
71. Levy-Lahad E, Wasco W, Poorkaj P, et al. Candidate gene for the chromosome 1 familial Alzheimer's disease locus. *Science*. 1995;269:973–977.
72. Sherrington R, Froelich S, Sorbi S, et al. Alzheimer's disease associated with mutations in presenilin 2 is rare and variably penetrant. *Hum Mol Genet*. 1996;5:985–988.
73. Goate A, Chartier-Harlin MC, Mullan M, et al. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nature*. 1991;349:704–706.
74. George-Hyslop PH S, Tanzi RE, Polinsky RJ, et al. The genetic defect causing familial Alzheimer's disease maps on chromosome 21. *Science*. 1987;235:885–890.
75. Hollingworth P, Harold D, Jones L, Owen MJ, Williams J. Alzheimer's disease genetics: Current knowledge and future challenges. *Int J Geriatr Psychiatry*. 2011;26:793–802.
76. Ryman DC, Acosta-Baena N, Aisen PS, et al. Symptom onset in autosomal dominant Alzheimer disease: A systematic review and meta-analysis. *Neurology*. 2014;83:253–260.
77. Campion D, Dumanchin C, Hannequin D, et al. Early-onset autosomal dominant Alzheimer disease: Prevalence, genetic heterogeneity, and mutation spectrum. *Am J Hum Genet*. 1999;65:664–670.
78. Bekris LM, Yu CE, Bird TD, Tsuang DW. Genetics of Alzheimer disease. *J Geriatr Psychiatry Neurol*. 2010;23:213–227.
79. Bateman RJ, Aisen PS, De Strooper B, et al. Autosomal-dominant Alzheimer's disease: A review and proposal for the prevention of Alzheimer's disease. *Alzheimer's Res Ther*. 2011;2:1.
80. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. *Science*. 2002;297:353–356.
81. Karran E, Mercken M, De Strooper B. The amyloid cascade hypothesis for Alzheimer's disease: An appraisal for the development of therapeutics. *Nat Rev Drug Discov*. 2011;10:698–712.
82. Cummings J, Lee G, Zhong K, Fonseca J, Taghva K. Alzheimer's disease drug development pipeline: 2021. *Alzheimer's Dement Transl Res Clin Interv*. 2021;7:e12179.
83. Smith MA, Joseph JA, Arson PG. Tracking the culprit in Alzheimer's disease. *Ann N Y Acad Sci*. 2000;924:35–38.
84. Bishop GM, Robinson SR. The amyloid hypothesis: Let sleeping dogmas lie? *Neurobiol Aging*. 2002;23:1101–1105.
85. Smith MA, Casadesus G, Joseph JA, Perry G. Amyloid- $\beta$  and  $\tau$  serve antioxidant functions in the aging and Alzheimer brain. *Free Radic Biol Med*. 2002;33:1194–1199.
86. Lee H, Casadesus G, Zhu X, Takeda A, Perry G, Smith MA. Challenging the amyloid cascade hypothesis: Senile plaques and amyloid- $\beta$  as protective adaptations to Alzheimer disease. *Ann N Y Acad Sci*. 2004; 1019(1):1–4.
87. Castellani RJ, Smith MA. Compounding artefacts with uncertainty, and an amyloid cascade hypothesis that is 'too big to fail'. *J Pathol*. 2011;224:147–152.
88. Pillar C. Blots on a field? *Science*. 2022;377:358–363.
89. Morris GP, Clark IA, Vissel B. Inconsistencies and controversies surrounding the amyloid hypothesis of Alzheimer's disease. *Acta Neuropathol Commun*. 2014;2:135.
90. Herrup K. The case for rejecting the amyloid cascade hypothesis. *Nat Neurosci*. 2015;18:794–799.
91. Harrison JR, Owen MJ. Alzheimer's disease: The amyloid hypothesis on trial. *Br J Psychiatry*. 2016;208:1–3.
92. Kepp KP. Ten challenges of the amyloid hypothesis of Alzheimer's disease. *J Alzheimer's Dis*. 2017;55:447–457.
93. Ricciarelli R, Fedele E. The amyloid cascade hypothesis in Alzheimer's disease: It's time to change our mind. *Curr Neuroparmacol*. 2017;15:926–935.
94. Morris GP, Clark IA, Vissel B. Questions concerning the role of amyloid- $\beta$  in the definition, aetiology and diagnosis of Alzheimer's disease. *Acta Neuropathol*. 2018;136:663–689.
95. Castello MA, Soriano S. On the origin of Alzheimer's disease. Trials and tribulations of the amyloid hypothesis. *Ageing Res Rev*. 2014;13:10–12.
96. Prusiner SB. Some speculations about prions, amyloid, and Alzheimer's disease. *N Engl J Med*. 1984;310:661–663.
97. Masters CL, Gajdusek DC, Gibbs CJJ. The familial occurrence of creutzfeldt-jakob disease and Alzheimer's disease. *Brain*. 1981; 104:535–558.
98. Glenner GG, Wong CW. Alzheimer's disease and down's syndrome: Sharing of a unique cerebrovascular amyloid fibril protein. *Biochem Biophys Res Commun*. 1984;122:1131–1135.
99. Glenner GG, Wong C. Amyloidosis in Alzheimer's disease and down's syndrome. In: Davies P and Finch CE, editors. *Molecular neuropathology of aging*: Cold Spring Harbor Laboratory Press; 1987. p 253–265.
100. Yankner BA, Dawes LR, Fisher S, Villa-Komaroff L, Oster-Granite ML, Neve RL. Neurotoxicity of a fragment of the amyloid precursor associated with Alzheimer's disease. *Science*. 1989;245:417–420.
101. Lorenzo A, Yankner BA. Beta-amyloid neurotoxicity requires fibril formation and is inhibited by Congo red. *Proc Natl Acad Sci U S A*. 1994;91:12243–12247.
102. Kang J, Lemaire HG, Unterbeck A, et al. The precursor of Alzheimer's disease amyloid A4-protein resembles a cell-surface receptor. *Nature*. 1987;325:733–736.
103. Tanzi RE, Gusella JF, Watkins PC, et al. Amyloid beta protein gene: CDNA, mRNA distribution, and genetic linkage near the Alzheimer locus. *Science*. 1987; 235:880–884.
104. Robakis NK, Ramakrishna N, Wolfe G, Wisniewski HM. Molecular cloning and characterization of a cDNA encoding the cerebrovascular and the neuritic plaque amyloid peptides. *Proc Natl Acad Sci U S A*. 1987;84:4190–4194.
105. Goldgaber D, Lerman MI, McBride OW, Saffiotti U, Gajdusek DC. Characterization and chromosomal localization of a cDNA encoding brain amyloid of Alzheimer's disease. *Science*. 1987;235:877–880.
106. Levy E, Carman MD, Fernandez-Madrid IJ, et al. Mutation of the Alzheimer's disease amyloid gene in hereditary cerebral hemorrhage, Dutch type. *Science*. 1990;248:1124–1126.
107. Citron M, Oltersdorf T, Haass C, et al. Mutation of the  $\beta$ -amyloid precursor protein in familial Alzheimer's disease increases  $\beta$ -protein production. *Nature*. 1992;360:672–674.
108. Scheuner D, Eckman C, Jensen M, et al. Secreted amyloid beta-protein similar to that in the senile plaques of Alzheimer's disease is increased in vivo by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. *Nat Med*. 1996;2: 864–870.
109. Anderson JP, Esch FS, Keim PS, Sambamurti K, Lieberburg I, Robakis NK. Exact cleavage site of Alzheimer amyloid precursor in neuronal PC-12 cells. *Neurosci Lett*. 1991;128:126–128.
110. Haass C, Koo EH, Mellon A, Hung AY, Selkoe DJ. Targeting of cell-surface  $\beta$ -amyloid precursor protein to lysosomes: Alternative processing into amyloid-bearing fragments. *Nature*. 1992;357:500–503.

111. Wolfe MS. Processive proteolysis by  $\gamma$ -secretase and the mechanism of Alzheimer's disease. *Biol Chem.* 2012;393:899–905.
112. Vassar R, Bennett BD, Babu-Khan S, et al. Beta-secretase cleavage of Alzheimer's amyloid precursor protein by the transmembrane aspartic protease BACE. *Science.* 1999;286:735–741.
113. Cai T, Tomita T. Structure-activity relationship of presenilin in  $\gamma$ -secretase-mediated intramembrane cleavage. *Semin Cell Dev Biol.* 2020;105:102–109.
114. De Strooper B, Iwatsubo T, Wolfe MS. Presenilins and  $\gamma$ -secretase: Structure, function, and role in Alzheimer disease. *Cold Spring Harb Perspect Med.* 2012;2:a006304.
115. Zhang Y, Thompson R, Zhang H, Xu H. APP Processing in Alzheimer's disease. *Mol Brain.* 2011;4:3.
116. Takami M, Nagashima Y, Sano Y, et al.  $\gamma$ -Secretase: Successive tripeptide and tetrapeptide release from the transmembrane domain of  $\beta$ -carboxyl terminal fragment. *J Neurosci.* 2009;29:13042–13052.
117. Qi-Takahara Y, Morishima-Kawashima M, Tanimura Y, et al. Longer forms of amyloid beta protein: Implications for the mechanism of intramembrane cleavage by gamma-secretase. *J Neurosci.* 2005;25:436–445.
118. Steiner H, Fukumori A, Tagami S, Okochi M. Making the final cut: Pathogenic amyloid- $\beta$  peptide generation by  $\gamma$ -secretase. *Cell Stress.* 2018;2:292–310.
119. Morley JE, Farr SA, Nguyen AD, Xu F. What is the physiological function of amyloid-Beta protein? *J Nutr Health Aging.* 2019;23:225–226.
120. Multhaup G, Schlicksupp A, Hesse L, et al. The amyloid precursor protein of Alzheimer's disease in the reduction of copper(II) to copper(I). *Science.* 1996;271:1406–1409.
121. Multhaup G. Amyloid precursor protein, copper and Alzheimer's disease. *Biomed Pharmacother.* 1997;51:105–111.
122. Kepp KP. Alzheimer's disease: How metal ions define  $\beta$ -amyloid function. *Coord Chem Rev.* 2017;351:127–159.
123. Kepp KP. Alzheimer's disease due to loss of function: A new synthesis of the available data. *Prog Neurobiol.* 2016;143:36–60.
124. White AR, Multhaup G, Maher F, et al. The Alzheimer's disease amyloid precursor protein modulates copper-induced toxicity and oxidative stress in primary neuronal cultures. *J Neurosci.* 1999;19:9170–9179.
125. Sturchio A, Dwivedi AK, Young CB, et al. High cerebrospinal amyloid- $\beta$  42 is associated with normal cognition in individuals with brain amyloidosis. *eClinicalMedicine.* 2021;38:100988.
126. Sturchio A, Dwivedi AK, Malm T, et al. High soluble amyloid- $\beta$  42 predicts normal cognition in amyloid-positive individuals with Alzheimer's disease-causing mutations. *J Alzheimer's Dis.* 2022;90:333–348.
127. Hilbich C, Kisters-Woike B, Reed J, Masters CL, Beyreuther K. Aggregation and secondary structure of synthetic amyloid  $\beta$ A4 peptides of Alzheimer's disease. *J Mol Biol.* 1991;218:149–163.
128. Fukumori A, Fluhrer R, Steiner H, Haass C. Three-amino acid spacing of presenilin endoproteolysis suggests a general step-wise cleavage of gamma-secretase-mediated intramembrane proteolysis. *J Neurosci.* 2010;30:7853–7862.
129. Bolduc DM, Montagna DR, Seghers MC, Wolfe MS, Selkoe DJ. The amyloid-beta forming tripeptide cleavage mechanism of  $\gamma$ -secretase. *Elife.* 2016;5:e17578.
130. Golde TE, Estus S, Younkin LH, Selkoe DJ, Younkin SG. Processing of the amyloid protein precursor to potentially amyloidogenic derivatives. *Science.* 1992;255:728–730.
131. Tiwari MK, Kepp KP.  $\beta$ -Amyloid pathogenesis: Chemical properties versus cellular levels. *Alzheimer's Dement.* 2016;12:184–194.
132. Suarez-Calvet M, Belbin O, Pera M, et al. Autosomal-dominant Alzheimer's disease mutations at the same codon of amyloid precursor protein differentially alter abeta production. *J Neurochem.* 2014;128:330–339.
133. Shea YF, Chu LW, Chan AOK, Ha J, Li Y, Song YQ. A systematic review of familial Alzheimer's disease: Differences in presentation of clinical features among three mutated genes and potential ethnic differences. *J Formos Med Assoc.* 2016;115:67–75.
134. Chévez-Gutiérrez L, Bammens L, Benilova I, et al. The mechanism of  $\gamma$ -secretase dysfunction in familial Alzheimer disease. *EMBO J.* 2012;31:2261–2274.
135. Winklhofer KF, Tatzelt J, Haass C. The two faces of protein misfolding: Gain-and loss-of-function in neurodegenerative diseases. *EMBO J.* 2008;27:336–349.
136. Robakis NK. Mechanisms of AD neurodegeneration may be independent of abeta and its derivatives. *Neurobiol Aging.* 2011;32:372–379.
137. Mackenzie IRA, Baker M, Pickering-Brown S, et al. The neuropathology of frontotemporal lobar degeneration caused by mutations in the progranulin gene. *Brain.* 2006; 129(Pt):3081–3090.
138. Kumar D, Ganeshpurkar A, Kumar D, Modi G, Gupta SK, Singh SK. Secretase inhibitors for the treatment of Alzheimer's disease: Long road ahead. *Eur J Med Chem.* 2018;148:436–452.
139. Karran E, De Strooper B. The amyloid hypothesis in Alzheimer disease: New insights from new therapeutics. *Nat Rev Drug Discov.* 2022;21:306–318.
140. Tate B, McKee TD, Loureiro RMB, et al. Modulation of gamma-secretase for the treatment of Alzheimer's disease. *Int J Alzheimers Dis.* 2012;2012:210756.
141. Cai T, Yonaga M, Tomita T. Activation of  $\gamma$ -secretase trimming activity by topological changes of transmembrane domain 1 of presenilin 1. *J Neurosci.* 2017;37:12272–12280.
142. Oehlrich D, Berthelot DJ-C, Gijzen HJM.  $\gamma$ -Secretase modulators as potential disease modifying anti-Alzheimer's drugs. *J Med Chem.* 2010;54:669–698.
143. Rynearson KD, Tanzi RE, Wagner SL. Discovery of potent gamma secretase modulators for the treatment of Alzheimer's disease. In: *Translational neuroscience: Fundamental approaches for neurological disorders.* Springer US; 2016:359–368.
144. Johnson DS, Pettersson M.  $\gamma$ -secretase modulators as A $\beta$ 42-lowering pharmacological agents to treat Alzheimer's disease. In: *Alzheimer's disease II. Topics in medicinal chemistry.* Springer; 2017:87–118.
145. Crump CJ, Johnson DS, Li YM. Development and mechanism of  $\gamma$ -secretase modulators for Alzheimer's disease. *Biochemistry.* 2013;52:3197–3216.
146. Oehlrich D, Berthelot DJC, Gijzen HJM.  $\gamma$ -Secretase modulators as potential disease modifying anti-Alzheimer's drugs. *J Med Chem.* 2011;54:669–698.
147. Golde TE, Koo EH, Felsenstein KM, Osborne BA, Miele L.  $\gamma$ -Secretase inhibitors and modulators. *Biochim Biophys Acta—Biomembr.* 2013;1828:2898–2907.
148. Wolfe MS.  $\gamma$ -Secretase inhibitors and modulators for Alzheimer's disease. *J Neurochem.* 2012; 120(SUPPL.):89–98.
149. Wilcock DM, Colton CA. Anti-Amyloid-beta immunotherapy in Alzheimer's disease: Relevance of transgenic mouse studies to clinical trials. *J Alzheimers Dis.* 2008;15:555–569.
150. Castellani RJ, Perry G. Pathogenesis and disease-modifying therapy in Alzheimer's disease: The flat line of progress. *Arch Med Res.* 2012;43:694–698.
151. Doody RS, Thomas RG, Farlow M, et al. Phase 3 trials of solanezumab for mild-to-moderate Alzheimer's disease. *N Engl J Med.* 2014;370:311–321.
152. Lao K, Zhang R, Luan J, Zhang Y, Gou X. Therapeutic strategies targeting amyloid- $\beta$  receptors and transporters in Alzheimer's disease. *J Alzheimers Dis.* 2021;79:1429–1442.



153. Selkoe DJ, Hardy J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol Med*. 2016;8:595–608.
154. Frisoni GB, Altomare D, Thal DR, et al. The probabilistic model of Alzheimer disease: The amyloid hypothesis revised. *Nat Rev Neurosci*. 2022;23:53–66.
155. Aizenstein HJ, Nebes RD, Saxton JA, et al. Frequent amyloid deposition without significant cognitive impairment among the elderly. *Arch Neurol*. 2008;65:1509–1517.
156. Bennett DA, Schneider JA, Arvanitakis Z, et al. Neuropathology of older persons without cognitive impairment from two community-based studies. *Neurology*. 2006;66:1837–1844.
157. Giannakopoulos P, Herrmann FR, Bussière T, et al. Tangle and neuron numbers, but not amyloid load, predict cognitive status in Alzheimer's disease. *Neurology*. 2003;60:1495–1500.
158. Shioi J, Georgakopoulos A, Mehta P, et al. FAD Mutants unable to increase neurotoxic A $\beta$ 42 suggest that mutation effects on neurodegeneration may be independent of effects on abeta. *J Neurochem*. 2007;101:674–681.
159. Sun L, Zhou R, Yang G, Shi Y. Analysis of 138 pathogenic mutations in presenilin-1 on the in vitro production of A $\beta$ 42 and A $\beta$ 40 peptides by  $\gamma$ -secretase. *Proc Natl Acad Sci U S A*. 2016;114:E476–E485.
160. Kaye R, Head E, Thompson JL, et al. Common structure of soluble amyloid oligomers implies common mechanism of pathogenesis. *Science*. 2003;300:486–489.
161. Lesne S, Koh MT, Kotilinek L, et al. A specific amyloid-beta protein assembly in the brain impairs memory. *Nature*. 2006;440:352–357.
162. Benilova I, Karran E, De Strooper B. The toxic A $\beta$  oligomer and Alzheimer's disease: An emperor in need of clothes. *Nat Neurosci*. 2012;15:349–357.
163. Shankar GM, Li S, Mehta TH, et al. Amyloid-beta protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory. *Nat Med*. 2008;14:837–842.
164. Cleary JP, Walsh DM, Hofmeister JJ, et al. Natural oligomers of the amyloid- $\beta$  protein specifically disrupt cognitive function. *Nat Neurosci*. 2005;8:79–84.
165. Walsh DM, Klyubin I, Fadeeva J V, et al. Naturally secreted oligomers of amyloid beta protein potently inhibit hippocampal long-term potentiation in vivo. *Nature*. 2002;416:535–539.
166. Bateman RJ, Munsell LY, Morris JC, Swarm R, Yarasheski KE, Holtzman DM. Human amyloid-beta synthesis and clearance rates as measured in cerebrospinal fluid in vivo. *Nat Med*. 2006;12:856–861.
167. Jack Jr CR, Knopman DS, Jagust WJ, et al. Tracking pathophysiological processes in Alzheimer's disease: An updated hypothetical model of dynamic biomarkers. *Lancet Neurol*. 2013;12:207–216.
168. Walsh DM, Selkoe DJ. A beta oligomers - a decade of discovery. *J Neurochem*. 2007;101(5):1172–1184.
169. Sciacca MFM, Kotler SA, Brender JR, Chen J, Lee DK, Ramamoorthy A. Two-step mechanism of membrane disruption by A $\beta$  through membrane fragmentation and pore formation. *Biophys J*. 2012;103:702–710.
170. Brender JR, Heyl DL, Samiseti S, et al. Membrane disordering is not sufficient for membrane permeabilization by islet amyloid polypeptide: Studies of IAPP(20–29) fragments. *Phys Chem Chem Phys*. 2013;15:8908–8915.
171. Tiwari MK, Kepp KP. Modeling the aggregation propensity and toxicity of amyloid- $\beta$  variants. *J Alzheimer's Dis*. 2015;47:215–229.
172. Götz J, Eckert A, Matamalas M, Ittner LM, Liu X. Modes of A $\beta$  toxicity in Alzheimer's disease. *Cell Mol Life Sci*. 2011;68:3359–3375.
173. Glabe CG. Common mechanisms of amyloid oligomer pathogenesis in degenerative disease. *Neurobiol Aging*. 2006;27:570–575.
174. Kotler SA, Walsh P, Brender JR, Ramamoorthy A. Differences between amyloid- $\beta$  aggregation in solution and on the membrane: Insights into elucidation of the mechanistic details of Alzheimer's disease. *Chem Soc Rev*. 2014;43:8–10.
175. Matsuzaki K. How do membranes initiate Alzheimer's disease? Formation of toxic amyloid fibrils by the amyloid  $\beta$ -protein on ganglioside clusters. *Acc Chem Res*. 2014;47:2397–2404.
176. Korshavn KJ, Satriano C, Lin Y, et al. Reduced lipid bilayer thickness regulates the aggregation and cytotoxicity of amyloid- $\beta$ . *J Biol Chem*. 2017;292:4638–4650.
177. Lecanu L, Greeson J, Papadopoulos V. Beta-amyloid and oxidative stress jointly induce neuronal death, amyloid deposits, gliosis, and memory impairment in the rat brain. *Pharmacology*. 2006;76:19–33.
178. You H, Tsutsui S, Hameed S, et al. A $\beta$  neurotoxicity depends on interactions between copper ions, prion protein, and N-methyl-D-aspartate receptors. *Proc Natl Acad Sci U S A*. 2012;109:1737–1742.
179. Caspersen C, Wang N, Yao J, et al. Mitochondrial A $\beta$ : A potential focal point for neuronal metabolic dysfunction in Alzheimer's disease. *FASEB J*. 2005;19:2040–2041.
180. Lustbader JW, Cirilli M, Lin C, et al. ABAD Directly links abeta to mitochondrial toxicity in Alzheimer's disease. *Science*. 2004;304:448–452.
181. Manczak M, Anekonda TS, Henson E, Park BS, Quinn J, Reddy PH. Mitochondria are a direct site of A beta accumulation in Alzheimer's disease neurons: Implications for free radical generation and oxidative damage in disease progression. *Hum Mol Genet*. 2006;15:1437–1449.
182. Arispe N, Rojas E, Pollard HB. Alzheimer Disease amyloid beta protein forms calcium channels in bilayer membranes: Blockade by tromethamine and aluminum. *Proc Natl Acad Sci U S A*. 1993;90:567–571.
183. Bhatia R, Lin H, Lal R. Fresh and globular amyloid beta protein (1–42) induces rapid cellular degeneration: Evidence for AbetaP channel-mediated cellular toxicity. *FASEB J*. 2000;14:1233–1243.
184. Musiek ES, Holtzman DM. Three dimensions of the amyloid hypothesis: Time, space, and “wingmen.”. *Nat Neurosci*. 2015;18:800–806.
185. Massetti N, Russo M, Franciotti R, et al. A machine learning-based holistic approach to predict the clinical course of patients within the Alzheimer's disease Spectrum. *J Alzheimer's Dis*. 2022;85:1639–1655.
186. Golde TE, Schneider LS, Koo EH. Anti-A $\beta$  therapeutics in Alzheimer's disease: The need for a paradigm shift. *Neuron*. 2011;69:203–213.
187. Sorrentino P, Iuliano A, Polverino A, Jacini F, Sorrentino G. The dark sides of amyloid in Alzheimer's disease pathogenesis. *FEBS Lett*. 2014;588:641–652.
188. Pimplikar SW, Nixon RA, Robakis NK, Shen J, Tsai L-H. Amyloid-independent mechanisms in Alzheimer's disease pathogenesis. *J Neurosci*. 2010;30:14946–14954.
189. Rosenblum WI. Why Alzheimer trials fail: Removing soluble oligomeric beta amyloid is essential, inconsistent, and difficult. *Neurobiol Aging*. 2014;35:969–974.
190. Barnard ND, Bush AI, Ceccarelli A, et al. Dietary and lifestyle guidelines for the prevention of Alzheimer's disease. *Neurobiol Aging*. 2014;35:S74–S78.
191. Teich AF, Arancio O. Is the amyloid hypothesis of Alzheimer's disease therapeutically relevant? *Biochem J*. 2012;446:165–177.
192. Butterfield DA, Halliwell B. Oxidative stress, dysfunctional glucose metabolism and Alzheimer disease. *Nat Rev Neurosci*. 2019;20:148–160.
193. Puzzo D, Gulisano W, Palmeri A, Arancio O. Rodent models for Alzheimer's disease drug discovery. *Expert Opin Drug Discov*. 2015;10:703–711.

194. Schwab C, Hosokawa M, McGeer PL. Transgenic mice overexpressing amyloid beta protein are an incomplete model of Alzheimer disease. *Exp Neurol*. 2004;188:52–64.
195. Steffen J, Krohn M, Schmitlick C, et al. Expression of endogenous mouse APP modulates  $\beta$ -amyloid deposition in hAPP-transgenic mice. *Acta Neuropathol Commun*. 2017;5:49.
196. Morley JE, Farr SA, Banks WA, Johnson SN, Yamada KA, Xu L. A physiological role for amyloid- $\beta$  protein: Enhancement of learning and memory. *J Alzheimer's Dis*. 2010;19:441–449.
197. Yankner BA, Duffy LK, Kirschner DA. Neurotrophic and neurotoxic effects of amyloid beta protein: Reversal by tachykinin neuropeptides. *Science*. 1990;250:279–282.
198. Whitson JS, Selkoe DJ, Cotman CW. Amyloid  $\beta$  protein enhances the survival of hippocampal neurons in vitro. *Science*. 1989;243:1488–1490.
199. He G, Luo W, Li P, et al. Gamma-secretase activating protein is a therapeutic target for Alzheimer's disease. *Nature*. 2010;467:95–98.
200. Wolfe MS.  $\gamma$ -Secretase as a target for Alzheimer's disease. *Curr Top Med Chem*. 2002;2:371–383.
201. Watt AD, Perez KA, Rembach A, et al. Oligomers, fact or artefact? SDS-PAGE induces dimerization of  $\beta$ -amyloid in human brain samples. *Acta Neuropathol*. 2013;125:549–564.
202. Villemagne VL, Pike KE, Ch  telat G, et al. Longitudinal assessment of  $a\beta$  and cognition in aging and Alzheimer disease. *Ann Neurol*. 2011;69:181–192.
203. H  ilund-Carlsen PF, Barrio JR, Gjedde A, Werner TJ, Alavi A. Circular inference in dementia diagnostics. *J Alzheimer's Dis*. 2018;63:69–73.
204. Drachman DA. The amyloid hypothesis, time to move on: Amyloid is the downstream result, not cause, of Alzheimer's disease. *Alzheimer's Dement*. 2014;10:372–380.
205. Zhu X, Raina AK, Perry G, Smith MA. Alzheimer's disease: The two-hit hypothesis. *Lancet Neurol*. 2004;3:219–226.
206. Jack CRJ, Knopman DS, Jagust WJ, et al. Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. *Lancet Neurol*. 2010;9:119–128.
207. Townsend M, Shankar GM, Mehta T, Walsh DM, Selkoe DJ. Effects of secreted oligomers of amyloid beta-protein on hippocampal synaptic plasticity: A potent role for trimers. *J Physiol*. 2006; 572(Pt ):477–492.
208. Shen J, Kelleher RJ. The presenilin hypothesis of Alzheimer's disease: Evidence for a loss-of-function pathogenic mechanism. *Proc Natl Acad Sci U S A*. 2007;104:403–409.
209. Saito T, Suemoto T, Brouwers N, et al. Potent amyloidogenicity and pathogenicity of  $A\beta_{43}$ . *Nat Neurosci*. 2011;14:1023–1032.
210. Kretner B, Trambauer J, Fukumori A, et al. Generation and deposition of  $A\beta_{43}$  by the virtually inactive presenilin-1 L435F mutant contradicts the presenilin loss-of-function hypothesis of Alzheimer's disease. *EMBO Mol Med*. 2016;8:458–465.
211. Veugelen S, Saito T, Saido TC, Ch  vez-Guti  rrez L, De Strooper B. Familial Alzheimer's disease mutations in presenilin generate amyloidogenic  $a\beta$  peptide seeds. *Neuron*. 2016;90:410–416.
212. Trambauer J, Rodr  guez Sarmiento RM, Fukumori A, Feederle R, Baumann K, Steiner H.  $A\beta_{43}$ -producing PS 1 FAD mutants cause altered substrate interactions and respond to  $\gamma$ -secretase modulation. *EMBO Rep*. 2020;21:e47996.
213. Hesse L, Behr D, Masters CL, Multhaup G. The beta A4 amyloid precursor protein binding to copper. *FEBS Lett*. 1994;349:109–116.
214. Dong J, Atwood CS, Anderson VE, et al. Metal binding and oxidation of amyloid- $\beta$  within isolated senile plaque cores: Raman microscopic evidence. *Biochemistry*. 2003;42:2768–2773.
215. Cherny RA, Atwood CS, Xilinas ME, et al. Treatment with a copper-zinc chelator markedly and rapidly inhibits  $\beta$ -amyloid accumulation in Alzheimer's disease transgenic mice. *Neuron*. 2001;30:665–676.
216. Bush AI, Multhaup G, Moir RD, et al. A novel zinc(II) binding site modulates the function of the beta A4 amyloid protein precursor of Alzheimer's disease. *J Biol Chem*. 1993;268:16109–16112.
217. Bush AI, Pettingell WH, Multhaup G, et al. Rapid induction of Alzheimer  $a\beta$  amyloid formation by zinc. *Science*. 1994;265:1464–1467.
218. Cuajungco MP, Goldstein LE, Nunomura A, et al. Evidence that the  $\beta$ -amyloid plaques of Alzheimer's disease represent the redox-silencing and entombment of  $a\beta$  by zinc. *J Biol Chem*. 2000;275:19439–19442.
219. Jomova K, Vondrakova D, Lawson M, Valko M. Metals, oxidative stress and neurodegenerative disorders. *Mol Cell Biochem*. 2010;345(1–2):91–104.
220. Opazo CM, Greenough MA, Bush AI. Copper: From neurotransmission to neuroproteostasis. *Front Aging Neurosci*. 2014;6:143.
221. Bush AI. The metal theory of Alzheimer's disease. *Rev Lit Arts Am*. 2013;33:277–281.
222. Hureau C. Coordination of redox active metal ions to the amyloid precursor protein and to amyloid- $\beta$  peptides involved in Alzheimer disease. Part 1: An overview. *Coord Chem Rev*. 2012; 256:2164–2174.
223. Hureau C, Dorlet P. Coordination of redox active metal ions to the amyloid precursor protein and to amyloid- $\beta$  peptides involved in Alzheimer disease. Part 2: Dependence of Cu (II) binding sites with  $a\beta$  sequences. *Coord Chem Rev*. 2012;256:2175–2187.
224. Fallor P. Copper and zinc binding to amyloid- $\beta$ : Coordination, dynamics, aggregation, reactivity and metal-ion transfer. *ChemBioChem*. 2009;10:2837–2845.
225. Viles JH. Metal ions and amyloid fiber formation in neurodegenerative diseases. Copper, zinc and iron in Alzheimer's, Parkinson's and prion diseases. *Coord Chem Rev*. 2012;256:2271–2284.
226. Haass C, Selkoe D. If amyloid drives Alzheimer disease, why have anti-amyloid therapies not yet slowed cognitive decline? *Plos Biol*. 2022;20:e3001694.
227. Richard E, den Brok MGHE, van Gool WA. Bayes Analysis supports null hypothesis of anti-amyloid beta therapy in Alzheimer's disease. *Alzheimer's Dement*. 2021;17:1051–1055.
228. Nicoll JAR, Buckland GR, Harrison CH, et al. Persistent neuropathological effects 14 years following amyloid- $\beta$  immunization in Alzheimer's disease. *Brain*. 2019;142:2113–2126.
229. Abramov E, Dolev I, Fogel H, Cicciotosto GD, Ruff E, Slutsky I. Amyloid-beta as a positive endogenous regulator of release probability at hippocampal synapses. *Nat Neurosci*. 2009;12: 1567–1576.
230. Josepha J, Shukitt-Hale B, Denisova NA, Martin A, Perry G, Smith MA. Copernicus revisited: Amyloid beta in Alzheimer's disease. *Neurobiol Aging*. 2001;22:131–146.
231. Biran Y, Masters CL, Barnham KJ, Bush AI, Adlard PA. Pharmacotherapeutic targets in Alzheimer's disease. *J Cell Mol Med*. 2009;13:61–86.
232. Makin S. The amyloid hypothesis on trial. *Nature*. 2018;559:S4.
233. Johnson KA, Minoshima S, Bohnen NI, et al. Appropriate use criteria for amyloid PET: A report of the amyloid imaging task force, the society of nuclear medicine and molecular imaging, and the Alzheimer's association. *J Nucl Med*. 2013;54:476–490.
234. Mullane K, Williams M. Alzheimer's disease beyond amyloid: Can the repetitive failures of amyloid-targeted therapeutics inform future approaches to dementia drug discovery? *Biochem Pharmacol*. 2020;177:113945.
235. Corder EH, Saunders AM, Strittmatter WJ, et al. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science*. 1993;261:921–923.

236. Kim KY, Wood BE, Wilson MI. Risk factors for Alzheimer's diseases: An overview for clinical practitioners. *Consult Pharm.* 2005;20:224–230.
237. Bickeböllner H, Campion D, Brice A, et al. Apolipoprotein E and Alzheimer disease: Genotype-specific risks by age and sex. *Am J Hum Genet.* 1997;60:439–446.
238. Medway C, Morgan K. Review: The genetics of Alzheimer's disease; putting flesh on the bones. *Neuropathol Appl Neurobiol.* 2014;40:97–105.
239. Bertram L, Tanzi RE. Genome-wide association studies in Alzheimer's disease. *Hum Mol Genet.* 2009;18(R2):R137–R145.
240. Harold D, Abraham R, Hollingworth P, et al. Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. *Nat Genet.* 2009;41:1088–1093.
241. Karch CM, Goate AM. Alzheimer's disease risk genes and mechanisms of disease pathogenesis. *Biol Psychiatry.* 2015;77:43–51.
242. Whitmer RA, Gunderson EP, Quesenberry CP, Zhou J, Yaffe K. Body mass index in midlife and risk of Alzheimer disease and vascular dementia. *Curr Alzheimer Res.* 2007;4:103–109.
243. Kivipelto M, Ngandu T, Fratiglioni L, et al. Obesity and vascular risk factors at midlife and the risk of dementia and Alzheimer disease. *Arch Neurol.* 2005;62:1556–1560.
244. Burdo JR, Chen Q, Calcutt NA, Schubert D. The pathological interaction between diabetes and presymptomatic Alzheimer's disease. *Neurobiol Aging.* 2009;30:1910–1917.
245. Ott A, Stolk RP, Hofman A, VanHarskamp F, Grobbee DE, Breteler MMB. Association of diabetes mellitus and dementia: The rotterdam study. *Diabetologia.* 1996;39:1392–1397.
246. de la Monte SM. The full Spectrum of Alzheimer's disease is rooted in metabolic derangements that drive type 3 diabetes. *Adv Exp Med Biol.* 2019;1128:45–83.
247. Massaia M, Di Ceva AP, Cappa MBG, et al. Risk factors for dementia of Alzheimer's type: A case-control, retrospective evaluation. *Arch Gerontol Geriatr.* 2001;7:253–259.
248. Sanmugam K. Depression is a risk factor for Alzheimer disease-review. *Res J Pharm Technol.* 2015;8:1056.
249. Dufouila C, Seshadri S, Chene G. Cardiovascular risk profile in women and dementia. *J Alzheimers Dis.* 2014;42(Suppl. 4):S353–63.
250. da Silva S L, Vellas B, Elemans S, et al. Plasma nutrient status of patients with Alzheimer's disease: Systematic review and meta-analysis. *Alzheimers Dement.* 2014;10:485–502.
251. Woodward MC. Prevention of Alzheimer's disease and other dementias. *J Pharmacol.* 2003;33:138–143.
252. Livingston G, Huntley J, Sommerlad A, et al. Dementia prevention, intervention, and care: 2020 report of the lancet commission. *Lancet.* 2020;396:413–446.
253. Mucke L, Masliah E, Yu G-Q, et al. High-level neuronal expression of  $\text{A}\beta$ 1–42 in wild-type human amyloid protein precursor transgenic mice: Synaptotoxicity without plaque formation. *J Neurosci.* 2000;20:4050–4058.
254. Bolognesi B, Lehner B. Protein overexpression: Reaching the limit. *Elife.* 2018;7:e39804.
255. Neve RL, McPhie DL, Chen Y. Alzheimer's disease: A dysfunction of the amyloid precursor protein. Published on the world wide web on 11 September 2000. *Brain Res.* 2000; 886:54–66.
256. Plant LD, Boyle JP, Smith IF, Peers C, Pearson HA. The production of amyloid beta peptide is a critical requirement for the viability of central neurons. *J Neurosci.* 2003;23:5531–5535.
257. Korte M, Herrmann U, Zhang X, Draguhn A. The role of APP and APLP for synaptic transmission, plasticity, and network function: Lessons from genetic mouse models. *Exp brain Res.* 2012; 217(3–4):435–440.
258. Tsuchida K, Shioi J, Yamada S, et al. Appican, the proteoglycan form of the amyloid precursor protein, contains chondroitin sulfate E in the repeating disaccharide region and 4-O-sulfated galactose in the linkage region. *J Biol Chem.* 2001; 276:37155–37160.
259. Pearson HA, Peers C. Physiological roles for amyloid beta peptides. *J Physiol.* 2006; 575(Pt ):5–10.
260. Lee H, Zhu X, Castellani RJ, Nunomura A, Perry G, Smith MA. Amyloid- $\beta$  in Alzheimer disease: The null versus the alternate hypotheses. *J Pharmacol Exp Ther.* 2007;321:823–829.
261. Soucek T, Cumming R, Dargusch R, Maher P, Schubert D. The regulation of glucose metabolism by HIF-1 mediates a neuroprotective response to amyloid beta peptide. *Neuron.* 2003;39:43–56.
262. Zou K, Gong J-S, Yanagisawa K, Michikawa M. A novel function of monomeric amyloid  $\beta$ -protein serving as an antioxidant molecule against metal-induced oxidative damage. *J Neurosci.* 2002;22:4833–4841.
263. Kamenetz F, Tomita T, Hsieh H, et al. APP Processing and synaptic function. *Neuron.* 2003;37:925–937.
264. Kumar DKV, Choi SH, Washicosky KJ, et al. Amyloid- $\beta$  peptide protects against microbial infection in mouse and worm models of Alzheimer's disease. *Sci Transl Med.* 2016;8:340ra72–340ra72.
265. Brothers HM, Gosztyla ML, Robinson SR. The physiological roles of amyloid- $\beta$  peptide hint at new ways to treat Alzheimer's disease. *Front Aging Neurosci.* 2018;10:118.
266. Miklossy J. Emerging roles of pathogens in Alzheimer disease. *Expert Rev Mol Med.* 2011;13:1–34.
267. Yu H, Li Z, Zhang H, Wang X. Role of potassium channels in  $\text{A}\beta$ 1–40-activated apoptotic pathway in cultured cortical neurons. *J Neurosci Res.* 2006;84:1475–1484.
268. Yan Y, Wang C.  $\text{A}\beta$ 42 is more rigid than  $\text{A}\beta$ 40 at the C terminus: Implications for  $\text{A}\beta$  aggregation and toxicity. *J Mol Biol.* 2006; 364:853–862.
269. Roher AE, Lowenson JD, Clarke S, et al. Structural alterations in the peptide backbone of beta-amyloid core protein may account for its deposition and stability in Alzheimer's disease. *J Biol Chem.* 1993;268:3072–3083.
270. Perry G, Cash AD, Smith MA. Alzheimer Disease and oxidative stress. *J Biomed Biotechnol.* 2002;2:120–123.
271. Halle A, Hornung V, Petzold GC, et al. The NALP3 inflammasome is involved in the innate immune response to amyloid- $\beta$ . *Nat Immunol.* 2008;9:857–865.
272. Salminen A, Ojala J, Kauppinen A, Kaarniranta K, Suuronen T. Inflammation in Alzheimer's disease: Amyloid- $\beta$  oligomers trigger innate immunity defence via pattern recognition receptors. *Prog Neurobiol.* 2009;87:181–194.
273. Bu X-L, Yao X-Q, Jiao S-S, et al. A study on the association between infectious burden and Alzheimer's disease. *Eur J Neurol.* 2015;22:1519–1525.
274. Lövhelm H, Gilthorpe J, Adolfsson R, Nilsson L-G, Elgh F. Reactivated herpes simplex infection increases the risk of Alzheimer's disease. *Alzheimer's Dement.* 2015;11:593–599.
275. Maheshwari P, Eslick GD. Bacterial infection and Alzheimer's disease: A meta-analysis. *J Alzheimer's Dis.* 2015;43:957–966.
276. Cestari JAF, Fabri GMC, Kalil J, et al. Oral infections and cytokine levels in patients with Alzheimer's disease and mild cognitive impairment compared with controls. *J Alzheimer's Dis.* 2016;52:1479–1485.
277. Itzhaki RF, Lathe R, Balin BJ, et al. Microbes and Alzheimer's disease. *J Alzheimer's Dis JAD.* 2016;51:979.
278. Itzhaki RF, Lin W-R, Shang D, Wilcock GK, Faragher B, Jamieson GA. Herpes simplex virus type 1 in brain and risk of Alzheimer's disease. *Lancet.* 1997;349:241–244.



279. Marambaud P, Wen PH, Dutt A, et al. A CBP binding transcriptional repressor produced by the PS1/ $\epsilon$ -cleavage of N-cadherin is inhibited by PS1 FAD mutations. *Cell*. 2003;114:635–645.
280. Siebel C, Lendahl U. Notch signaling in development, tissue homeostasis, and disease. *Physiol Rev*. 2017;97:1235–1294.
281. Bentahir M, Nyabi O, Verhamme J, et al. Presenilin clinical mutations can affect gamma-secretase activity by different mechanisms. *J Neurochem*. 2006;96:732–742.
282. Walker ES, Martinez M, Brunkan AL, Goate A. Presenilin 2 familial Alzheimer's disease mutations result in partial loss of function and dramatic changes in  $A\beta$  42/40 ratios. *J Neurochem*. 2005;92:294–301.
283. Watanabe H, Shen J. Dominant negative mechanism of presenilin-1 mutations in FAD. *Proc Natl Acad Sci U S A*. 2017;114:12635–12637.
284. Okochi M, Steiner H, Fukumori A, et al. Presenilins mediate a dual intramembranous  $\gamma$ -secretase cleavage of notch-1. *EMBO J*. 2002;21:5408–5416.
285. Barthet G, Georgakopoulos A, Robakis NK. Cellular mechanisms of  $\gamma$ -secretase substrate selection, processing and toxicity. *Prog Neurobiol*. 2012;98:166–175.
286. Klyubin I, Betts V, Welzel AT, et al. Amyloid protein dimer-containing human CSF disrupts synaptic plasticity: Prevention by systemic passive immunization. *J Neurosci*. 2008;28:4231–4237.
287. Walsh DM, Selkoe DJ. Deciphering the molecular basis of memory failure in Alzheimer's disease. *Neuron*. 2004;44:181–193.
288. Lacor PN, Buniel MC, Chang L, et al. Synaptic targeting by Alzheimer's-related amyloid beta oligomers. *J Neurosci*. 2004;24:10191–10200.
289. Coleman PD, Yao PJ. Synaptic slaughter in Alzheimer's disease. *Neurobiol Aging*. 2003;24:1023–1027.
290. Klunk WE, Engler H, Nordberg A, et al. Imaging brain amyloid in Alzheimer's disease with Pittsburgh compound-B. *Ann Neurol*. 2004;55:306–319.
291. Moghbel MC, Saboury B, Basu S, et al. Amyloid- $\beta$  imaging with PET in Alzheimer's disease: Is it feasible with current radiotracers and technologies? *Eur J Nucl Med Mol Imaging*. 2012;39:202–208.
292. Kepe V, Moghbel MC, Långström B, et al. Amyloid- $\beta$  positron emission tomography imaging probes: A critical review. *J Alzheimer's Dis*. 2013;36:613–631.
293. Alavi A, Barrio JR, Werner TJ, Khosravi M, Newberg A, Høilund-Carlsen PF. Suboptimal validity of amyloid imaging-based diagnosis and management of Alzheimer's disease: Why it is time to abandon the approach. *Eur J Nucl Med Mol Imaging*. 2020;47:225–230.
294. Høilund-Carlsen PF, Werner TJ, Alavi A, Revheim M-E. Aducanumab-related amyloid-related imaging abnormalities: Paeon or lament? *Clin Nucl Med*. 2022;47:625–626.
295. Høilund-Carlsen PF, Revheim M-E, Alavi A, Satyamurthy N, Barrio JR. Amyloid PET: A questionable single primary surrogate efficacy measure on Alzheimer immunotherapy trials. *J Alzheimer's Dis*. 2022;90:1395–1399.
296. Morris E, Chalkidou A, Hammers A, Peacock J, Summers J, Keevil S. Diagnostic accuracy of 18F amyloid PET tracers for the diagnosis of Alzheimer's disease: A systematic review and meta-analysis. *Eur J Nucl Med Mol Imaging*. 2016;43:374–385.
297. Rice I, Bisdas S. The diagnostic value of FDG and amyloid PET in Alzheimer's disease—A systematic review. *Eur J Radiol*. 2017;94:16–24.
298. Alavi A, Werner TJ, Høilund-Carlsen PF, Zaidi H. Correction for partial volume effect is a must, not a luxury, to fully exploit the potential of quantitative PET imaging in clinical oncology. *Mol imaging Biol*. 2018;20:1–3.
299. Zhang S, Smailagic N, Hyde C, et al. 11 C-PIB-PET for the early diagnosis of Alzheimer's disease dementia and other dementias in people with mild cognitive impairment (MCI). *Cochrane Database Syst Rev*. 2014;2014:CD010386.
300. Cohen AD, Landau SM, Snitz BE, Klunk WE, Blennow K, Zetterberg H. Fluid and PET biomarkers for amyloid pathology in Alzheimer's disease. *Mol Cell Neurosci*. 2019;97:3–17.
301. Gomez-Isla T, Hollister R, West H, et al. Neuronal loss correlates with but exceeds neurofibrillary tangles in Alzheimer's disease. *Ann Neurol*. 1997;41:17–24.
302. Schmitz C, Rutten BPF, Pielon A, et al. Hippocampal neuron loss exceeds amyloid plaque load in a transgenic mouse model of Alzheimer's disease. *Am J Pathol*. 2004;164:1495–1502.
303. Bush AI, Tanzi RE. Therapeutics for Alzheimer's disease based on the metal hypothesis. *Neurotherapeutics*. 2008;5:421–432.
304. Bouwman FH, Schoonenboom NSM, Verwey NA, et al. CSF Biomarker levels in early and late onset Alzheimer's disease. *Neurobiol Aging*. 2009;30:1895–1901.
305. Price JL, McKeel DWJ, Buckles VD, et al. Neuropathology of non-demented aging: Presumptive evidence for preclinical Alzheimer disease. *Neurobiol Aging*. 2009;30:1026–1036.
306. Clifford R, JJ, Therneau TM, Weigand SD, et al. Prevalence of biologically vs clinically defined Alzheimer spectrum entities using the national institute on aging–Alzheimer's association research framework. *JAMA Neurol*. 2019;76:1174–1183.
307. Arenaza-Urquijo EM, Przybelski SA, Lesnick TL, et al. The metabolic brain signature of cognitive resilience in the 80+: Beyond Alzheimer pathologies. *Brain*. 2019;142:1134–1147.
308. Shokri-Kojori E, Wang G-J, Wiers CE, et al.  $\beta$ -Amyloid accumulation in the human brain after one night of sleep deprivation. *Proc Natl Acad Sci U S A*. 2018;115:4483–4488.
309. Kang J-E, Lim MM, Bateman RJ, et al. Amyloid- $\beta$  dynamics are regulated by orexin and the sleep-wake cycle. *Science*. 2009;326:1005–1007.
310. Hansson O, Lehmann S, Otto M, Zetterberg H, Lewczuk P. Advantages and disadvantages of the use of the CSF amyloid  $\beta$  ( $A\beta$ ) 42/40 ratio in the diagnosis of Alzheimer's disease. *Alzheimers Res Ther*. 2019;11:34.
311. Kang J-H, Korecka M, Toledo JB, Trojanowski JQ, Shaw LM. Clinical utility and analytical challenges in measurement of cerebrospinal fluid amyloid- $\beta$ 1–42 and  $\tau$  proteins as Alzheimer disease biomarkers. *Clin Chem*. 2013;59:903–916.
312. Ostrowitzki S, Lasser RA, Dorflinger E, et al. A phase III randomized trial of gantenerumab in prodromal Alzheimer's disease. *Alzheimers Res Ther*. 2017;9:1–15.
313. Salloway S, Sperling R, Fox NC, et al. Two phase 3 trials of bapineuzumab in mild-to-moderate Alzheimer's disease. *N Engl J Med*. 2014;370:322–333.
314. Vandenberghe R, Rinne JO, Boada M, et al. Bapineuzumab for mild to moderate Alzheimer's disease in two global, randomized, phase 3 trials. *Alzheimers Res Ther*. 2016;8:1–13.
315. Haeblerlein S B, Aisen PS, Barkhof F, et al. Two randomized phase 3 studies of aducanumab in early Alzheimer's disease. *J Prev Alzheimer's Dis*. 2022;9:197–210.
316. Fillenbaum GG, van Belle G, Morris JC, et al. Consortium to establish a registry for Alzheimer's disease (CERAD): The first twenty years. *Alzheimers Dement*. 2008;4:96–109.
317. McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical diagnosis of Alzheimer's disease: Report

- of the NINCDS-ADRDA work group under the auspices of department of health and human services task force on Alzheimer's disease. *Neurology*. 1984;34:939.
318. Dubois B, Villain N, Frisoni GB, et al. Clinical diagnosis of Alzheimer's disease: Recommendations of the international working group. *Lancet Neurol*. 2021;20:484–496.
  319. Gomes LA, Hipp SA, Rijal Upadhya A, et al. A $\beta$ -induced acceleration of Alzheimer-related  $\tau$ -pathology spreading and its association with prion protein. *Acta Neuropathol*. 2019;138:913–941.
  320. Qiang W, Yau W-M, Luo Y, Mattson MP, Tycko R. Antiparallel beta-sheet architecture in Iowa-mutant beta-amyloid fibrils. *Proc Natl Acad Sci U S A*. 2012;109:4443–4448.
  321. Lomakin A, Teplow DB, Kirschner DA, Benedek GB. Kinetic theory of fibrillogenesis of amyloid beta-protein. *Proc Natl Acad Sci U S A*. 1997;94:7942–7947.
  322. Seubert P, Vigo-Pelfrey C, Esch F, et al. Isolation and quantification of soluble Alzheimer's beta-peptide from biological fluids. *Nature*. 1992;359:325–327.
  323. Galasko D, Chang L, Motter R, et al. High cerebrospinal fluid tau and low amyloid beta42 levels in the clinical diagnosis of Alzheimer disease and relation to apolipoprotein E genotype. *Arch Neurol*. 1998;55:937–945.
  324. Arber C, Lovejoy C, Wray S. Stem cell models of Alzheimer's disease: Progress and challenges. *Alzheimers Res Ther*. 2017;9:1–17.
  325. Duyckaerts C, Potier M-C, Delatour B. Alzheimer Disease models and human neuropathology: Similarities and differences. *Acta Neuropathol*. 2008;115:5–38.
  326. Perez-Nievas BG, Stein TD, Tai HC, et al. Dissecting phenotypic traits linked to human resilience to Alzheimer's pathology. *Brain*. 2013;136:2510–2526.
  327. Reddy PH, Mani G, Park BS, et al. Differential loss of synaptic proteins in Alzheimer's disease: Implications for synaptic dysfunction. *J Alzheimers Dis*. 2005;7:103–180.
  328. Musiek ES, Gomez-Isla T, Holtzman DM. Aducanumab for Alzheimer disease: The amyloid hypothesis moves from bench to bedside. *J Clin Invest*. 2021;131:e154889.
  329. Haapasalo A, Kovacs DM. The many substrates of presenilin/ $\gamma$ -secretase. *J Alzheimer's Dis*. 2011;25:3–28.
  330. Saido T, Leissring MA. Proteolytic degradation of amyloid  $\beta$ -protein. *Cold Spring Harb Perspect Med*. 2012;2:a006379.
  331. Qiu WQ, Walsh DM, Ye Z, et al. Insulin-degrading enzyme regulates extracellular levels of amyloid beta-protein by degradation. *J Biol Chem*. 1998;273:32730–32738.
  332. Bulloj A, Leal MC, Xu H, Castaño EM, Morelli L. Insulin-degrading enzyme sorting in exosomes: A secretory pathway for a key brain amyloid- $\beta$  degrading protease. *J Alzheimer's Dis*. 2010;19:79–95.
  333. Malgieri G, Grasso G. The clearance of misfolded proteins in neurodegenerative diseases by zinc metalloproteases: An inorganic perspective. *Coord Chem Rev*. 2014;260:139–155.
  334. Miners JS, Barua N, Kehoe PG, Gill S, Love S. Abeta-degrading enzymes: Potential for treatment of Alzheimer disease. *J Neuropathol Exp Neurol*. 2011;70:944–959.
  335. Carson JA, Turner AJ. B-amyloid catabolism: Roles for neprilysin (NEP) and other metallopeptidases? *J Neurochem*. 2002;81:1–8.
  336. Haass C, Lemere CA, Capell A, et al. The Swedish mutation causes early-onset Alzheimer's disease by [beta]-secretase cleavage within the secretory pathway. *Nat Med*. 1995;1:1291–1296.
  337. De Strooper B. Lessons from a failed  $\gamma$ -secretase Alzheimer trial. *Cell*. 2014;159:721–726.
  338. Somavarapu AK, Kepp KP. Loss of stability and hydrophobicity of presenilin 1 mutations causing Alzheimer's disease. *J Neurochem*. 2016;137:101–111.
  339. Cacquevel M, Aeschbach L, Houacine J, Fraering PC. Alzheimer's disease-linked mutations in presenilin-1 result in a drastic loss of activity in purified  $\gamma$ -secretase complexes. *PLoS One*. 2012;7:1–13.
  340. Somavarapu AK, Kepp KP. Membrane dynamics of  $\gamma$ -secretase provides a molecular basis for  $\beta$ -amyloid binding and processing. *ACS Chem Neurosci*. 2017;8:2424–2436.
  341. Tang N, Somavarapu AK, Kepp KP. Molecular recipe for  $\gamma$ -secretase modulation from computational analysis of 60 active compounds. *ACS Omega*. 2018;3:18078–18088.
  342. Mehra R, Kepp KP. Understanding familial Alzheimer's disease: The fit-stay-trim mechanism of  $\gamma$ -secretase. *Wiley Interdiscip Rev Comput Mol Sci*. 2022;12:e1556.
  343. Mehra R, Kepp KP. Computational analysis of Alzheimer-causing mutations in amyloid precursor protein and presenilin 1. *Arch Biochem Biophys*. 2019;678:108168.
  344. Tang N, Kepp KP. A $\beta$ 42/A $\beta$ 40 ratios of presenilin 1 mutations correlate with clinical onset of Alzheimer's disease. *J Alzheimer's Dis*. 2018;66:939–945.
  345. De Strooper B, Chávez Gutiérrez L. Learning by failing: Ideas and concepts to tackle  $\gamma$ -secretases in Alzheimer's disease and beyond. *Annu Rev Pharmacol Toxicol*. 2015;55:419–437.
  346. Xia D, Watanabe H, Wu B, et al. Presenilin-1 knockin mice reveal loss-of-function mechanism for familial Alzheimer's disease. *Neuron*. 2015;85:967–981.
  347. Kelleher RJ, Shen J. Presenilin-1 mutations and Alzheimer's disease. *Proc Natl Acad Sci U S A*. 2017;114:629–631.
  348. Saura CA, Choi S-Y, Beglopoulos V, et al. Loss of presenilin function causes impairments of memory and synaptic plasticity followed by age-dependent neurodegeneration. *Neuron*. 2004;42:23–36.
  349. Baki L, Shioi J, Wen P, et al. PS1 Activates PI3K thus inhibiting GSK-3 activity and tau overphosphorylation: Effects of FAD mutations. *EMBO J*. 2004;23:2586–2596.
  350. Wu B, Yamaguchi H, Lai FA, Shen J. Presenilins regulate calcium homeostasis and presynaptic function via ryanodine receptors in hippocampal neurons. *Proc Natl Acad Sci U S A*. 2013;110:15091–15096.
  351. Tu H, Nelson O, Bezprozvanny A, et al. Presenilins form ER Ca<sup>2+</sup> leak channels, a function disrupted by familial Alzheimer's disease-linked mutations. *Cell*. 2006;126:981–993.
  352. Supnet C, Bezprozvanny I. Presenilins function in ER calcium leak and Alzheimer's disease pathogenesis. *Cell Calcium*. 2011;50:303–309.
  353. Greenough MA. The role of presenilin in protein trafficking and degradation—Implications for metal homeostasis. *J Mol Neurosci*. 2016;60:289–297.
  354. Das HK, Tchédre K, Mueller B. Repression of transcription of presenilin-1 inhibits  $\gamma$ -secretase independent ER Ca<sup>2+</sup> leak that is impaired by FAD mutations. *J Neurochem*. 2012;122:487–500.
  355. Georgakopoulos A, Litterst C, Ghersi E, et al. Metalloproteinase/presenilin1 processing of ephrinB regulates EphB-induced src phosphorylation and signaling. *EMBO J*. 2006;25:1242–1252.
  356. Xia D, Kelleher RJ, Shen J. Loss of A $\beta$ 43 production caused by presenilin-1 mutations in the knockin mouse brain. *Neuron*. 2016;90:417–422.
  357. Dehury B, Tang N, Mehra R, Blundell TL, Kepp KP. Side-by-side comparison of notch- and C83 binding to  $\gamma$ -secretase in a complete membrane model at physiological temperature. *RSC Adv*. 2020;10:31215–31232.
  358. Bour A, Little S, Dodart J-C, Kelche C, Mathis C. A secreted form of the  $\beta$ -amyloid precursor protein (sAPP695) improves spatial recognition memory in OF1 mice. *Neurobiol Learn Mem*. 2004;81:27–38.

359. Jonsson T, Atwal JK, Steinberg S, et al. A mutation in APP protects against Alzheimer's disease and age-related cognitive decline. *Nature*. 2012;488:96–99.
360. Maloney JA, Bainbridge T, Gustafson A, et al. Molecular mechanisms of Alzheimer disease protection by the A673T allele of amyloid precursor protein. *J Biol Chem*. 2014;289:30990–31000.
361. Fahrenholz F, Gilbert S, Kojro E, Lammich S, Postina R. Alpha-secretase activity of the disintegrin metalloprotease ADAM 10. Influences of domain structure. *Ann N Y Acad Sci*. 2000;920:215–222.
362. Asai M, Hattori C, Szabo B, et al. Putative function of ADAM9, ADAM10, and ADAM17 as APP alpha-secretase. *Biochem Biophys Res Commun*. 2003;301:231–235.
363. Sensi SL, Granzotto A, Siotto M, Squitti R. Copper and zinc dysregulation in Alzheimer's disease. *Trends Pharmacol Sci*. 2018;39:1049–1063.
364. Hung YH, Bush AI, Cherny RA. Copper in the brain and Alzheimer's disease. *J Biol Inorg Chem*. 2010;15:61–76.
365. Robakis NK, Wisniewski HM, Jenkins EC, et al. Chromosome 21q21 sublocalisation of gene encoding beta-amyloid peptide in cerebral vessels and neuritic (senile) plaques of people with Alzheimer disease and down syndrome. *Lancet*. 1987;1:384–385.
366. Carri M, Cozzolino M. SOD1 And mitochondria in ALS: A dangerous liaison. *J Bioenerg Biomembr*. 2011;43:593–599.
367. Renton AE, Chiò A, Traynor BJ. State of play in amyotrophic lateral sclerosis genetics. *Nat Neurosci*. 2014;17:17–23.
368. Kitamura A, Inada N, Kubota H, et al. Dysregulation of the proteasome increases the toxicity of ALS-linked mutant SOD1. *Genes Cells*. 2014;19:209–224.
369. Balch WE, Morimoto RI, Dillin A, Kelly JW. Adapting proteostasis for disease intervention. *Science*. 2008;319:916–919.
370. Kim CK, Lee YR, Ong L, Gold M, Kalali A, Sarkar J. Alzheimer's disease: Key insights from two decades of clinical trial failures. *J Alzheimer's Dis*. 2022;87:83–100.
371. Farlow M, Arnold SE, Van Dyck CH, et al. Safety and biomarker effects of solanezumab in patients with Alzheimer's disease. *Alzheimer's Dement*. 2012;8:261–271.
372. Blennow K, Zetterberg H, Rinne JO, et al. Effect of immunotherapy with bapineuzumab on cerebrospinal fluid biomarker levels in patients with mild to moderate Alzheimer disease. *Arch Neurol*. 2012;69:1002–1010.
373. Imbimbo BP, Lucca U, Watling M. Can anti- $\beta$ -amyloid monoclonal antibodies work in autosomal dominant Alzheimer disease? *Neurol Genet*. 2021;7:e535.
374. Tayeb HO, Murray ED, Price BH, Tarazi FI. Bapineuzumab and solanezumab for Alzheimer's disease: Is the 'amyloid cascade hypothesis' still alive? *Expert Opin Biol Ther*. 2013;13:1075–1084.
375. Siemers ER, Sundell KL, Carlson C, et al. Phase 3 solanezumab trials: Secondary outcomes in mild Alzheimer's disease patients. *Alzheimer's Dement*. 2016;12:110–120.
376. Honig LS, Vellas B, Woodward M, et al. Trial of solanezumab for mild dementia due to Alzheimer's disease. *N Engl J Med*. 2018;378:321–330.
377. Le Couteur DG, Hunter S, Brayne C. Solanezumab and the amyloid hypothesis for Alzheimer's disease. *Bmj*. 2016;355:i6771.
378. Ayton S, Bush AI.  $\beta$ -amyloid: The known unknowns. *Ageing Res Rev*. 2021;65:101212.
379. Sperling RA, Rentz DM, Johnson KA, et al. The A4 study: Stopping AD before symptoms begin? *Sci Transl Med*. 2014;6:228fs13.
380. Salloway S, Farlow M, McDade E, et al. A trial of gantenerumab or solanezumab in dominantly inherited Alzheimer's disease. *Nat Med*. 2021;27:1187–1196.
381. Salloway S, Chalkias S, Barkhof F, et al. Amyloid-Related imaging abnormalities in 2 phase 3 studies evaluating aducanumab in patients with early Alzheimer disease. *JAMA Neurol*. 2022;79:13–21.
382. Ostrowitzki S, Deptula D, Thurfjell L, et al. Mechanism of amyloid removal in patients with Alzheimer disease treated with gantenerumab. *Arch Neurol*. 2012;69:198–207.
383. Lannfelt L, Basun H, Wahlund L-O, Rowe BA, Wagner SL. Decreased  $\alpha$ -secretase-cleaved amyloid precursor protein as a diagnostic marker for Alzheimer's disease. *Nat Med*. 1995;1:829–832.
384. Barger SW, Harmon AD. Microglial activation by Alzheimer amyloid precursor protein and modulation by apolipoprotein E. *Nature*. 1997;388:878–881.
385. Costa T, Cauda F. A Bayesian reanalysis of the phase III aducanumab (ADU) trial. *J Alzheimer's Dis*. 2021;87:1009–1012.
386. Levin J, Vögler J, Quiroz YT, et al. Testing the amyloid cascade hypothesis: Prevention trials in autosomal dominant Alzheimer disease. *Alzheimer's Dement*. 2022;18:2687–2698.
387. Andrews JS, Desai U, Kirson NY, Zichlin ML, Ball DE, Matthews BR. Disease severity and minimal clinically important differences in clinical outcome assessments for Alzheimer's disease clinical trials. *Alzheimer's Dement Transl Res Clin Interv*. 2019;5:354–363.
388. Lansdall CJ, McDougall F, Butler LM, et al. Establishing clinically meaningful change on outcome assessments frequently used in trials of mild cognitive impairment due to Alzheimer's disease. *J Prev Alzheimer's Dis*. 2023;10:9–18.
389. Villain N, Planche V, Levy R. High-clearance anti-amyloid immunotherapies in Alzheimer's disease. Part 1: Meta-analysis and review of efficacy and safety data, and medico-economical aspects. *Rev Neurol (Paris)*. 2022;178:1011–1030.
390. Lythgoe MP, Prasad V. How the US food and drug administration's approval of aducanumab for Alzheimer's disease has implication for oncology and beyond. *Eur J Cancer*. 2021;157:68–70.
391. Chételat G, Arbizu J, Barthel H, et al. Amyloid-PET and 18F-FDG-PET in the diagnostic investigation of Alzheimer's disease and other dementias. *Lancet Neurol*. 2020;19:951–962.
392. Wassef HR, Colletti PM. Re: Aducanumab-related ARIA: Paeon or lament? *Clin Nucl Med*. 2023;48:168–169.
393. Jack CR, Barrio JR, Kepe V. Cerebral amyloid PET imaging in Alzheimer's disease. *Acta Neuropathol*. 2013;126:643–657.
394. Ladiwala ARA, Litt J, Kane RS, et al. Conformational differences between two amyloid beta oligomers of similar size and dissimilar toxicity. *J Biol Chem*. 2012;287:24765–24773.
395. Høiland-Carlsen PF, Alavi A. Aducanumab (marketed as aduhelm) approval is likely based on misinterpretation of PET imaging data. *J Alzheimer's Dis*. 2021;84:1457–1460.
396. Arndt JW, Qian F, Smith BA, et al. Structural and kinetic basis for the selectivity of aducanumab for aggregated forms of amyloid- $\beta$ . *Sci Rep*. 2018;8:1–16.
397. Sevigny J, Chiao P, Bussièrè T, et al. The antibody aducanumab reduces  $\text{A}\beta$  plaques in Alzheimer's disease. *Nature*. 2016;537:50–56.
398. Sevigny J, Chiao P, Williams L, et al. Aducanumab (BIIB037), an anti-amyloid beta monoclonal antibody, in patients with prodromal or mild Alzheimer's disease: Interim results of a randomized, double-blind, placebo-controlled, phase 1b study. *Alzheimer's Dement*. 2015;11:P277.
399. Viglietta V, O'Gorman J, Williams L, et al. Randomized, double-blind, placebo-controlled studies to evaluate treatment with aducanumab (BIIB037) in patients with early Alzheimer's disease: Phase 3 study design (S1. 003). *Neurology*. 2016;86(16 Supplement):S1–003.



400. Selkoe DJ. Alzheimer Disease and aducanumab: Adjusting our approach. *Nat Rev Neurol*. 2019;15:365–366.
401. Knopman DS, Jones DT, Greicius MD. Failure to demonstrate efficacy of aducanumab: An analysis of the EMERGE and ENGAGE trials as reported by biogen, December 2019. *Alzheimer's Dement*. 2021;17:696–701.
402. Howard R, Liu KY. Questions EMERGE as biogen claims aducanumab turnaround. *Nat Rev Neurol*. 2020;16:63–64.
403. Schneider L. A resurrection of aducanumab for Alzheimer's disease. *Lancet Neurol*. 2020;19:111–112.
404. Dunn B, Stein P, Temple R, Cavazzoni P. An appropriate use of accelerated approval—Aducanumab for Alzheimer's disease. *N Engl J Med*. 2021;385:856–857.
405. Hollmann P. Update: FDA approval of biogen's aducanumab. *Geriatr Nurs (Minneap)*. 2022;43:318–319.
406. Schneider LS. Editorial: Aducanumab trials EMERGE but don't ENGAGE. *J Prev Alzheimer's Dis*. 2022;9:193–196.
407. McCleery J, Quinn TJ. Aducanumab and the certainty of evidence. *Age Ageing*. 2021;50:1899–1900.
408. Mahase E. Three FDA advisory panel members resign over approval of Alzheimer's drug. *BMJ*. 2021;373:n1503.
409. Alexander GC, Emerson S, Kesselheim AS. Evaluation of aducanumab for Alzheimer disease: Scientific evidence and regulatory review involving efficacy, safety, and futility. *JAMA*. 2021;325:1717–1718.
410. Lundebjerg NE, Hollmann PA, Supiano MA. Of education and public policy: Aducanumab. *J Am Geriatr Soc*. 2022;70:81–84.
411. Swanson CJ, Zhang Y, Dhadda S, et al. A randomized, double-blind, phase 2b proof-of-concept clinical trial in early Alzheimer's disease with lecanemab, an anti- $\alpha\beta$  protofibril antibody. *Alzheimers Res Ther*. 2021;13:1–14.
412. Tucker S, Möller C, Tegerstedt K, et al. The murine version of BAN2401 (mAb158) selectively reduces amyloid- $\beta$  protofibrils in brain and cerebrospinal fluid of tg-ArcSwe mice. *J Alzheimers Dis*. 2015;43:575–588.
413. Söderberg L, Johannesson M, Nygren P, et al. Lecanemab, aducanumab, and gantenerumab—Binding profiles to different forms of amyloid-Beta might explain efficacy and Side effects in clinical trials for Alzheimer's disease. *Neurotherapeutics*. 2022;20:195–206.
414. Thambisetty M, Howard R. Lecanemab trial in AD brings hope but requires greater clarity. *Nat Rev Neurol*. 2023;19:132–133.
415. Rogers SL, Doody RS, Mohs RC, Friedhoff LT, Group DS. Donepezil improves cognition and global function in Alzheimer disease: a 15-week, double-blind, placebo-controlled study. *Arch Intern Med*. 1998;158:1021–1031.
416. Christensen J. Experimental Alzheimer's drug may have contributed to death of study participant, according to reports. CNN. Published 28 October 2022. <https://edition.cnn.com/2022/10/28/health/alzheimers-drug-lecanemab-trial/index.html>
417. Piller C. Second death linked to potential antibody treatment for Alzheimer's disease. *Science*. Published 27 November 2022. Accessed 20 January 2023. <https://www.science.org/content/article/second-death-linked-potential-antibodytreatment-alzheimer-s-disease>
418. Piller C. Scientists tie third clinical trial death to experimental Alzheimer's drug. *Science*. Published 21 December 2022. Accessed 20 January 2023. <https://www.science.org/content/article/scientists-tiethird-clinical-trial-death-experimental-alzheimer-s-drug>
419. Gleason A, Ayton S, Bush AI. Unblinded by the light: ARIA in Alzheimer's clinical trials. *Eur J Neurol*. 2021;28:e1.
420. Bell ML, Kenward MG, Fairclough DL, Horton NJ. Differential dropout and bias in randomised controlled trials: When it matters and when it may not. *BMJ*. 2013;346:e8668.
421. Sperling RA, Jack Jr CR, Black SE, et al. Amyloid-related imaging abnormalities in amyloid-modifying therapeutic trials: Recommendations from the Alzheimer's association research roundtable workgroup. *Alzheimer's Dement*. 2011;7:367–385.
422. Atwood CS, Bishop GM, Perry G, Smith MA. Amyloid- $\beta$ : A vascular sealant that protects against hemorrhage? *J Neurosci Res*. 2002;70:356.
423. Mintun MA, Lo AC, Duggan Evans C, et al. Donanemab in early Alzheimer's disease. *N Engl J Med*. 2021;384:1691–1704.
424. Alawode DOT, Heslegrave AJ, Fox NC, Zetterberg H. Donanemab removes Alzheimer's plaques: What is special about its target? *Lancet Heal Longev*. 2021;2:e395–e396.
425. Wessels AM, Siemers ER, Yu P, et al. A combined measure of cognition and function for clinical trials: The integrated Alzheimer's disease rating scale (iADRS). *J Prev Alzheimer's Dis*. 2015;2:227.
426. Wessels AM, Rentz DM, Case M, Lauzon S, Sims JR. Integrated Alzheimer's disease rating scale: Clinically meaningful change estimates. *Alzheimer's & Dement Transl Res & Clin Interv*. 2022;8:e12312.
427. Alves F, Kallinowski P, Ayton S. Accelerated brain volume loss caused by anti- $\beta$ -amyloid drugs: A systematic review and meta-analysis. *Neurology*. 2023;100:e2114–e2124.
428. Zimmer J, Solomon P, Evans CD, et al. TRAILBLAZER-ALZ 2: A phase 3 study to assess safety and efficacy of donanemab in early symptomatic Alzheimer's disease (P18-3.005). *Neurology*. 2022;98(18 suppl):1688.
429. Ross EL, Weinberg MS, Arnold SE. Cost-effectiveness of aducanumab and donanemab for early Alzheimer disease in the US. *JAMA Neurol*. 2022;79:478–487.
430. Temp AGM, Ly A, van Doorn J, et al. A Bayesian perspective on biogen's aducanumab trial. *Alzheimer's Dement*. 2022;18:2341–2351.
431. The Lancet. Lecanemab for Alzheimer's disease: Tempering hype and hope. *Lancet*. 2022;400:1899.
432. Walsh S, Merrick R, Richard E, Nurock S, Brayne C. Lecanemab for Alzheimer's disease. *bmj*. 2022;379:o3010.
433. Seltzer B, Zolnouni P, Nunez M, et al. Efficacy of donepezil in early-stage Alzheimer disease: A randomized placebo-controlled trial. *Arch Neurol*. 2004;61:1852–1856.
434. Daly T, Herrup K, Espay AJ. An ethical argument for ending human trials of amyloid-lowering therapies in Alzheimer's disease. *AJOB Neurosci*. Published online 5 October 2022. <https://pubmed.ncbi.nlm.nih.gov/36197130/>
435. Lee J-H, Yu WH, Kumar A, et al. Lysosomal proteolysis and autophagy require presenilin 1 and are disrupted by Alzheimer-related PS1 mutations. *Cell*. 2010;141:1146–1158.
436. Yoon Y, Voloudakis G, Doran N, et al. PS1 FAD mutants decrease ephrinB2-regulated angiogenic functions, ischemia-induced brain neovascularization and neuronal survival. *Mol Psychiatry*. 2021;26:1996–2012.
437. McBrayer M, Nixon RA. Lysosome and calcium dysregulation in Alzheimer's disease: Partners in crime. *Biochem Soc Trans*. 2013;41:1495–1502.
438. Area-Gomez E, De Groof AJC, Boldogh I, et al. Presenilins are enriched in endoplasmic reticulum membranes associated with mitochondria. *Am J Pathol*. 2009;175:1810–1816.
439. Espay AJ, Sturchio A, Schneider LS, Ezzat K. Soluble amyloid- $\beta$  consumption in Alzheimer's disease. *J Alzheimer's Dis*. 2021;82:1403–1415.
440. Brown GC. The endotoxin hypothesis of neurodegeneration. *J Neuroinflammation*. 2019;16:180.
441. Lu T, Pan Y, Kao S-Y, et al. Gene regulation and DNA damage in the ageing human brain. *Nature*. 2004;429:883–891.

442. Greenough MA, Camakaris J, Bush AI. Metal dyshomeostasis and oxidative stress in Alzheimer's disease. *Neurochem Int.* 2013;62:540–555.
443. De la Monte SM. Type 3 diabetes is sporadic Alzheimer's disease: Mini-review. *Eur Neuropsychopharmacol.* 2014;24:1–7.
444. Willette AA, Bendlin BB, Starks EJ, et al. Association of insulin resistance with cerebral glucose uptake in late middle-aged adults at risk for Alzheimer disease. *JAMA Neurol.* 2015;72:1013–1020.
445. Talbot K, Wang H-Y. The nature, significance, and glucagon-like peptide-1 analog treatment of brain insulin resistance in Alzheimer's disease. *Alzheimer's Dement.* 2014;10:S12–S25.
446. Reich N, Hölscher C. The neuroprotective effects of glucagon-like peptide 1 in Alzheimer's and Parkinson's disease: An in-depth review. *Front Neurosci.* 2022;16:970925.
447. Kesika P, Suganthy N, Sivamaruthi BS, Chaiyasut C. Role of gut-brain axis, gut microbial composition, and probiotic intervention in Alzheimer's disease. *Life Sci.* 2021;264:118627.
448. Kohler C A, Maes M, Slyepchenko A, et al. The gut-brain axis, including the microbiome, leaky gut and bacterial translocation: Mechanisms and pathophysiological role in Alzheimer's disease. *Curr Pharm Des.* 2016;22:6152–6166.
449. McAuley MT, Kenny RA, Kirkwood TBL, Wilkinson DJ, Jones JLL, Miller VM. A mathematical model of aging-related and cortisol induced hippocampal dysfunction. *BMC Neurosci.* 2009;10:26.
450. Greene JA, Loscalzo J. Putting the patient back together-social medicine, network medicine, and the limits of reductionism. *N Engl J Med.* 2017;377:2493–2499.