# geNet package

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check\_input\_geNet

checking input df Function

# **Description**

function to check the format of the input binary df. It is automatically called by the geNet() function.

# Usage

```
check_input_geNet(binary_df)
```

# **Arguments**

binary\_df

a binary dataframe to be checked. Mandatory argument.

#### Value

None

#### **Examples**

```
## Not run: check_input_df(binary_df)
```

```
check_input_visualization
```

check\_input\_visualization

# Description

function to check the format of the input binary df. It is automatically called by the geNet() function.

# Usage

```
check_input_visualization(data)
```

# Arguments

data

output of the geNet() function (list of two ffdf objects)

#### Value

error message if checking fails

```
## Not run: check_input_visualization(data)
```

contract\_network 3

# **Description**

function to contract the visnetwork dataframe. It is automatically called by the plot\_visnetwork() function if contract\_net=T.

# Usage

```
contract_network(data, seed = 123, show_label = F, show_legend = T)
```

#### **Arguments**

data visnetwork dataframe generated by the geNet algorithm. Mandatory argument.

seed set seed of the nodes

show\_label should the labels of the nodes be visible? default to False

show\_legend should the legend be visible? default to True

#### Value

list of two objects:

- visnet: object of class "visNetwork" generated from the contracted data.
- data\_contracted: a compressed version of the input data object. It is generated by contracting the nodes of each cluster in one vertex.

This dataset reports only the nodes contained in each cluster and the connections between the clusters

#### **Description**

function to export the geNet output. It is automatically called by the geNet() function if export\_results=T

```
export_geNet_result(geNet_output, out_directory, format = "csv")
```

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# **Arguments**

geNet\_output the object generated by the geNet function. It is a list of two ffdf objects.

• nodes: ffdf object containing the information about the nodes

• edges: ffdf object containing the information about the edges

out\_directory

the directory to write the output

format

the user can export the geNet output in .csv format or in .ffdata format. In both cases, two files are generated. One containing the nodes information (e.g., clustering association), and the other one containing edges information (e.g., connections weight). Default to "csv".

#### Value

In case of the csv format, two csv files are generated in the output directory

- nodes\_geNet\_output.csv: it contains the information about the nodes
- edges\_geNet\_output.csv: it contains the information about the edges

#### **Examples**

```
## Not run: export_geNet_result(geNet_output,out_directory="./",format="csv")
# export output in current working directory "./"
## End(Not run)
```

filter\_id

function to filter the genes ID

#### **Description**

function to filter the genes ID from data. It is called automatically by the visualize\_network() function

#### Usage

```
filter_id(data, selected_id)
```

#### **Arguments**

data List of two ffdf objects (format generated by the geNet() function ). Mandatory

argument

selected\_id select the ids to filter data. Optional argument.

#### Value

data: data object filtered

final\_score 5

final_score	function to format the final df of scores	

# Description

function to format the final df of scores. The adjustment and filtering steps are executed. It is automatically called by the geNet() function

# Usage

```
final_score(
  final_df_phi_score,
  sel_weight = "coeff",
  pval_thr_pos = 0.01,
  pval_thr_neg = 0.1
)
```

# **Arguments**

```
final_df_phi_score

the dataframe of scores generated by the generate_final_df() function. Mandatory argument

sel_weight select the type of weights of the edges. Possible values: logpvalue: choose the negative log-pvalues as weights of the edges coeff: choose the correlation coefficients as weights of the edges Default to "coeff"

pval_thr_pos threhold p-value positive edges. Default to 0.01.

pval_thr_neg threshold p-value negative edges. Default to 0.1.
```

#### Value

Object of class "ffdf", containing the connections between the nodes. The connections have been filtered and correctly formatted to be converted in an igraph object

```
## Not run: final_score(final_df_phi_score,sel_weight="coeff")
```

6 generate\_final\_df

 ${\tt generate\_df\_scores} \qquad {\tt generate\_df\_scores}$ 

#### **Description**

function to generate the weights of the edges given the combinations. It is automatically called by the geNet() function. the correlation coefficient and its pvalue is calculated using the cor.test function from the stat package. See the relative documentation for additional details.

#### Usage

```
generate_df_scores(all_combs, binary_matrix, signif_test = "cor_test")
```

# **Arguments**

all\_combs matrix of combinations

binary\_matrix binary matrix of presence/absences genes

signif\_test significance test to use.

- cor\_test: use Pearson correlation test approach (faster)
- chisquare: use chi-square test approach (slower) Read vignettes for additional details about the significance tests used.

#### Value

Object of class "dataframe" containing the connections between the nodes and relative weights

generate\_final\_df generate\_final\_df function to generate the final df of scores based on the weights of edges generated by the generate\_df\_scores\_ff() function. It is called automatically by the geNet() function

#### **Description**

```
generate_final_df
```

function to generate the final df of scores based on the weights of edges generated by the generate\_df\_scores\_ff() function. It is called automatically by the geNet() function

```
generate_final_df(binary_matrix, n_cores = 1, test = "cor_test")
```

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# **Arguments**

binary\_matrix binary matrix of presence/absences genes n\_cores number of cores to use. Default to 1.

test significance test to use.

cor\_test: use Pearson correlation test approach (faster)
chisquare: use chi-square test approach (slower)

#### Value

Object of class "ffdf" containing the connections between the nodes and relative weights

# **Examples**

```
## Not run: generate_final_df(binary_matrix,progress=F,n_cores=1,test="cor_test")
```

generate\_freqs\_genes Calculate genes frequency

**Description** 

function to calculate the percentage of occurrence of every gene across all strains.

#### Usage

```
generate_freqs_genes(input_binary_df)
```

# **Arguments**

```
input_binary_df
```

Object of class "dataframe", containing the absence/presence of genes

#### Value

Object of class "dataframe". The first column reports the ID of the nodes/genes, the second column reports the frequency across all strains

```
## Not run: generate_freqs_genes(input_binary_df)
```

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geNet

Execution geNet algorithm

# **Description**

function to execute the geNet algorithm on a binary dataframe of genes occurrences (presence/absence data)

### Usage

```
geNet(
  input_binary_df,
  clust_method = "infomap",
  type_weight = "coeff",
  cores = 1,
  out_dir,
  export_results = F,
  test_pvalue = "cor_test",
  pval_thr_pos = 0.01,
  pval_thr_neg = 0.1
)
```

#### **Arguments**

input\_binary\_df

input binary dataframe containing genes occurrences. Mandatory argument. input\_binary\_df is an object of class "dataframe". The rownames are the strains names, the columns names are unique genes/nodes IDs.

clust\_method

clustering method. Default to "infomap".

type\_weight

possible values:

- logpvalue: the weights of the edges will be the negative log adjusted p-values
- coeff: the weights of the edges will be the correlation coefficients values Note: This is true only for the positive edges. The negative edges will have 0 weights in both cases. The negative edges doesn't influence the topology of the network or the clustering.

cores number of cores to use for parallel processing
out\_dir if export\_results=T, select output directory
export\_results exporting geNet output in csv files?
test\_pvalue significance test to use.

• cor\_test: use Pearson correlation test approach (faster)
• chisquare: use chi-square test approach (slower)

pval\_thr\_pos threhold p-value positive edges. Default to 0.01.

pval\_thr\_pos threhold p-value positive edges. Default to 0.01.

pval\_thr\_neg threshold p-value negative edges. Default to 0.1.

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#### Value

list of two objects of class "ffdf".

- nodes: reports info about the nodes and clustering
- edges: reports info about the connections between the nodes This format is designed to be used with the visNetwork package.

# **Examples**

```
## Not run: geNet(input_binary_df,clust_method="infomap",type_weight="coeff",cores=4)
```

gen\_network\_obj

generate igraph network

# **Description**

function to generate the igraph network object based on the final scores. It is called automatically by the geNet() function.

#### Usage

```
gen_network_obj(final_df_score)
```

# **Arguments**

final\_df\_score the dataframe of scores generated by the final\_score() function. Mandatory argument.

#### Value

Object of class "igraph"

gen\_visnetwork\_data

gen\_visnetwork\_data

# Description

function to get the object to generate the visnetwork. It is automatically called by the geNet() function.

```
gen_visnetwork_data(igraph_network)
```

#### **Arguments**

igraph\_network igraph object. Mandatory argument.

# Value

list of two objects of class "ffdf".

- nodes: reports info about the nodes and clustering
- edges: reports info about the connections between the nodes This format is designed to be used with the visNetwork package.

# **Description**

function to get the colors based on the ID of the nodes

#### **Usage**

```
get_colors_based_on_gene_ID(data, vec_gene_ids = "None", col_nodes = "black")
```

# **Arguments**

data List of two ffdf objects (format generated by the geNet() function )

vec\_gene\_ids vector of genes IDs to color. Mandatory argument.

col\_nodes color of the nodes that match the specified IDs. Default to "black".

#### Value

Object of class "dataframe", which reports the new colors for each node of the network

```
## Not run: get_colors_based_on_gene_ID(data,vec_gene_ids="None",col_nodes="black")
```

# **Description**

function to get the colors based on the size of the nodes

#### Usage

```
get_colors_based_on_size(
  data,
  size_thr,
  direction = "less",
  col_nodes = "black"
)
```

# **Arguments**

data List of two ffdf objects (format generated by the geNet() function )
size\_thr size threshold. Mandatory argument.

direction values greater/less than (or equal) the selected threshold

col\_nodes color of the nodes that match the specified condition. Default to "black".

#### Value

data\_new\_colors: object of class "dataframe", which reports the new colors for each node of the network

# Examples

```
## Not run: get_colors_based_on_size(data,size_thr=10,direction="less",col_nodes="black")
```

# Description

function to predict the colors based on the strains occurrence of genes i.e., show nodes/genes associated with the selected strains (i.e., that gene is present in that strains)

#### **Usage**

```
get_colors_based_on_strains(
  input_binary_df,
  data,
  strains_names,
  col_nodes = "black"
)
```

#### **Arguments**

input\_binary\_df

Object of class "dataframe", containing presence/absence of genes. Mandatory

argument.

data visnetwork dataframe generated by the geNet algorithm. Mandatory argument. strains\_names the vector containing the names of the strains to visualize. Mandatory argument.

col\_nodes Color of the nodes matched with the strains. Default to "black".

#### Value

Object of class "dataframe", which reports the new colors for each node of the network

# **Examples**

```
## Not run: get_colors_based_on_strains(input_binary_df,data,strains_names=c("strain_x"),col_nodes="black")
```

#### **Description**

function to get the gene IDs based on the annotation (tooltip column)

#### Usage

```
get_gene_ID_based_on_tooltip(data, vec_tooltip)
```

#### **Arguments**

data List of two ffdf objects (format generated by the geNet() function )

vec\_tooltip vector of annotation to match with the gene IDs. Mandatory argument.

#### Value

Object of class "dataframe", which reports the genes ID for the specified annotation keywords.

#### **Examples**

```
## Not run: get_gene_ID_based_on_tooltip(data,vec_tooltip=c("protein","secretion"))
# Get the nodes IDs whose annotation reports the word "protein" and "secretion"

## End(Not run)

get_groups_based_on_annotation

get groups based on the annotation
```

#### **Description**

function to get the groups based on the annotation (tooltip column)

#### Usage

```
get_groups_based_on_annotation(data, select_annotation)
```

#### **Arguments**

```
data List of two ffdf objects (format generated by the geNet() function ) select_annotation vector of annotation words to match with the groups
```

#### Value

Object of class "vector", which reports the groups selected

# **Examples**

# **Description**

function to get the groups based on a clustering algorithm Note: the color of the nodes is based on the group (using the function get\_new\_colors\_based\_on\_groups()) to evaluate the clustering method we use the modularity measure. The modularity of a graph with respect to some division (or vertex types) measures how good the division is, or how separated are the different vertex types from each other.

#### Usage

```
get_groups_based_on_clustering(igraph_network, method = "infomap")
```

#### **Arguments**

igraph\_network igraph network generated by the gen\_igraph\_network() function. Mandatory argument.

method clustering method:

- louvain: predict clusters using the Louvain algorithm. The "modularity" is the objective function to maximize. Small clusters may be hidden. It detects only the bigger clusters.
- infomap: predict clusters using the Infomap algorithm. The "map equation" is the objective function to minimize. Compared to the Louvain algorithm, it is able to identify smaller clusters (subclusters)
- · fastgreedy: predicts clusters using the Fastgreedy algorithm
- walktrap: predicts clusters using the Walktrap algorithm

Default to "infomap" to detect smaller clusters. The Louvain algorithm is faster than the Infomap algorithm in large networks.

#### Value

Object of class "dataframe", which reports the genes ID associated with the new groups

#### **Examples**

```
## Not run: get_groups_based_on_clustering(igraph_network,method="infomap")
```

# Description

function to get the groups based on the ID of the nodes

#### Usage

```
get_groups_based_on_gene_ID(data, vec_gene_ids)
```

#### **Arguments**

data List of two ffdf objects (format generated by the geNet() function )

vec\_gene\_ids vector of genes IDs to which the groups must be matched. Mandatory argument.

# Value

Object of class "dataframe", which reports the groups for the specified genes ID.

# **Examples**

```
## Not run: get_groups_based_on_gene_ID(data,vec_gene_ids=c("group_200","group_253"))
# Get the groups for the nodes IDs "group_200" and "group_253"
## End(Not run)
```

# **Description**

function to get the group names based on the number of nodes in each group.

# Usage

```
get_group_names_based_on_n_nodes(data)
```

#### **Arguments**

data

List of two ffdf objects (format generated by the geNet() function )

#### Value

Object of class "dataframe", which reports the old names associated with the new names

# **Examples**

```
## Not run: get_group_names_based_on_n_nodes(data)
```

#### **Description**

function to get the group names based on the annotation of the nodes (tooltip column)

```
get_group_names_based_on_tooltip(data)
```

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#### **Arguments**

data

List of two ffdf objects (format generated by the geNet() function )

#### Value

Object of class "dataframe", which reports the old names associated with the new names

# **Examples**

```
## Not run: get_group_names_based_on_tooltip(data)
```

get\_legend\_df

get\_legend\_df

# **Description**

function to get the legend of the groups. It is automatically called by the plot\_visnetwork() function.

#### Usage

```
get_legend_df(data)
```

#### **Arguments**

data

list of two ffdf objects generated by the geNet algorithm. Mandatory argument.

# Value

Object of class "dataframe", which reports the legend information of the nodes

get\_mappings

get\_mappings

# **Description**

function to get the correct mapping to contract the network. It is automatically called by the contract\_network() function.

#### Usage

```
get_mappings(igraph_net, data)
```

# Arguments

igraph\_net

igraph object. Mandatory argument.

data

list of two ffdf objects generated by the geNet algorithm. Mandatory argument.

# Value

Object of "vector" containing the mapping codes.

# Description

function to get the colors based on the groups

#### Usage

```
get_new_colors_based_on_groups(df_groups)
```

#### **Arguments**

df\_groups

object of class "dataframe", which reports the genes ID associated with the groups. Mandatory argument.

#### Value

Object of class "dataframe", which reports the gene IDs and the groups associated with the colors

# **Examples**

```
## Not run: get_new_colors_based_on_groups(df_groups)
```

mod\_color\_layer

 $mod\_color\_layer$ 

# **Description**

function to modify the color layer (colors of the node)

```
mod_color_layer(data, data_new_colors, no_matching_col = "old")
```

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#### **Arguments**

```
data_new_colors

dataframe that reports the new colors for each node

no_matching_col

default color in case of no matching (i.e. gene IDs of the network not reported in data_new_colors)

• old: the color of no matching nodes is the old color.

• white: the color of no matching nodes is white ("white" is a reserved color)
```

#### Value

data: data object modified with the new color layer

Default to "white".

# **Examples**

```
## Not run: mod_color_layer(data,data_new_colors,no_matching_col="white")
```

mod\_group\_layer

mod\_group\_layer

# **Description**

function to modify the group layer (influence also the structure of the contracted network)

#### Usage

```
mod_group_layer(data, data_new_groups = NULL)
```

# Arguments

```
data List of two ffdf objects (format generated by the geNet() function )
data_new_groups
dataframe that reports the new group for each node
```

#### Value

data: data object modified with the new group layer

```
## Not run: mod_group_layer(data,data_new_groups)
```

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mod\_group\_names

mod\_group\_names

# **Description**

function to modify the names of the group

# Usage

```
mod_group_names(data, df_new_groups_names = NULL)
```

#### **Arguments**

```
data  List \ of \ two \ ffdf \ objects \ (format \ generated \ by \ the \ geNet() \ function \ )   df\_new\_groups\_names
```

dataframe that reports the new names for each old group name

# Value

data: data object modified with the new group names for the current group layer

# **Examples**

```
## Not run: mod_group_names(data,df_new_groups_names)
```

mod\_size\_layer

mod\_size\_layer

# **Description**

function to modify the size layer (size of each node)

#### Usage

```
mod_size_layer(data, data_new_size = NULL)
```

# **Arguments**

data List of two ffdf objects (format generated by the geNet() function )
data\_new\_size dataframe that reports the size for each node

#### Value

data: data object modified with the new size layer

```
## Not run: mod_size_layer(data,data_new_size)
```

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```
mod\_tooltip\_visnetwork \\ mod\_tooltip\_visnetwork
```

# **Description**

function to modify the tooltip layer of the network

# Usage

```
mod_tooltip_visnetwork(data, data_new_tooltip = NULL)
```

# **Arguments**

```
data List of two ffdf objects (format generated by the geNet() function )

data_new_tooltip

dataframe that reports the new tooltip string for each node
```

#### Value

data: data object modified with the new tooltip layer

# **Examples**

```
## Not run: mod_tooltip_visnetwork(data,data_new_tooltip)
```

plot\_visnetwork

plot\_visnetwork

# **Description**

function to plot the visnetwork object based on the data input

```
plot_visnetwork(
  data,
  show_label = F,
  show_legend = T,
  seed = 123,
  name_net = "None",
  show_negative = F,
  contract_net = T,
  size_opt = "fixed"
)
```

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# **Arguments**

data list of two ffdf objects generated by the geNet algorithm. Mandatory argument.

show\_label show the labels of the nodes? show\_legend show the legend of groups?

seed set the seed of the current instance

name\_net name of the network show\_negative show negative edges?

contract\_net Should the network be contracted? Default to True

• fixed: the size of the node is equal to 8 (size layer values ignored)

• size\_opt: the size of the node is proportional to the size layer values

Default to "fixed". Note: if the there are too many nodes the size of the node is

reduced regardless the option specified by the user.

#### Value

data: data object modified with the new size layer

# **Examples**

```
## Not run:
plot_visnetwork(data,show_label=F,
show_legend=T,seed=123,name_net="None",
show_negative=F,contract_net=T,
size_opt="fixed")
## End(Not run)
```

visualize\_network

visualize network

# **Description**

function to visualize the network based on the connections and different conditions

```
visualize_network(
  data,
  select_group = NULL,
  select_tooltip = "None",
  select_size = "None",
  clust_method = "None",
  select_ID = NULL
)
```

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# Arguments

data List of two ffdf objects (format generated by the geNet() function ). Mandatory

argument

select\_group select the groups to filter data. Optional argument.

select\_tooltip select the annotation keywords to filter data. Optional argument. Note: the

tooltip values are the values that appear when hovering the nodes

select\_size select the size of the nodes to filter data. Optional argument.

clust\_method select the clustering method. Default to Infomap. select\_ID select the IDs to filter data. Optional argument.

#### Value

data: data object filtered

```
## Not run:
data_subsetted<-visualize_network(data,select_group=c("group_1","group_5"))
data_subsetted<-visualize_network(data,select_tooltip=c("protein","secretion"))
data_subsetted<-visualize_network(data,clust_method="louvain")
## End(Not run)</pre>
```

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