

# Human Intracranial Brain Observations Player

## User Manual

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### Abstract

This document is the user manual of the Human Intracranial Brain Observations Player (HiBoP). HiBoP is being developed by the Centre de Recherche en Neurosciences de Lyon (CRNL), with funds from the Human Brain Project (HBP) to visualize large datasets of intracranial electroencephalography (iEEG) data. This document was written for the 2.1.0 version of HiBoP.

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## 1 HiBoP project

A **Project** links together anatomical and functional data from a set of patients. If you have an anatomical description of the patients (3D mesh of the brain, MRI and coordinates of the sites of the electrodes) and the respective iEEG data, you can visualize them in 4D (3D space + time). The first thing to do with HiBoP is to create or open a Project. This can be done within HiBoP (section 2), but also using a third-party software which automatically generates the files and the hierarchy, such as the HiBoP Database Manager (appendix A).

On your Hard Drive, the Project consists of a folder with several subfolders (Figure 1). Each file in this subfolder is encoded with the json format, and thus can be edited (at your own risks) using any text editor. However this is not recommended, as any coma missing may cause the file to be incorrectly interpreted.

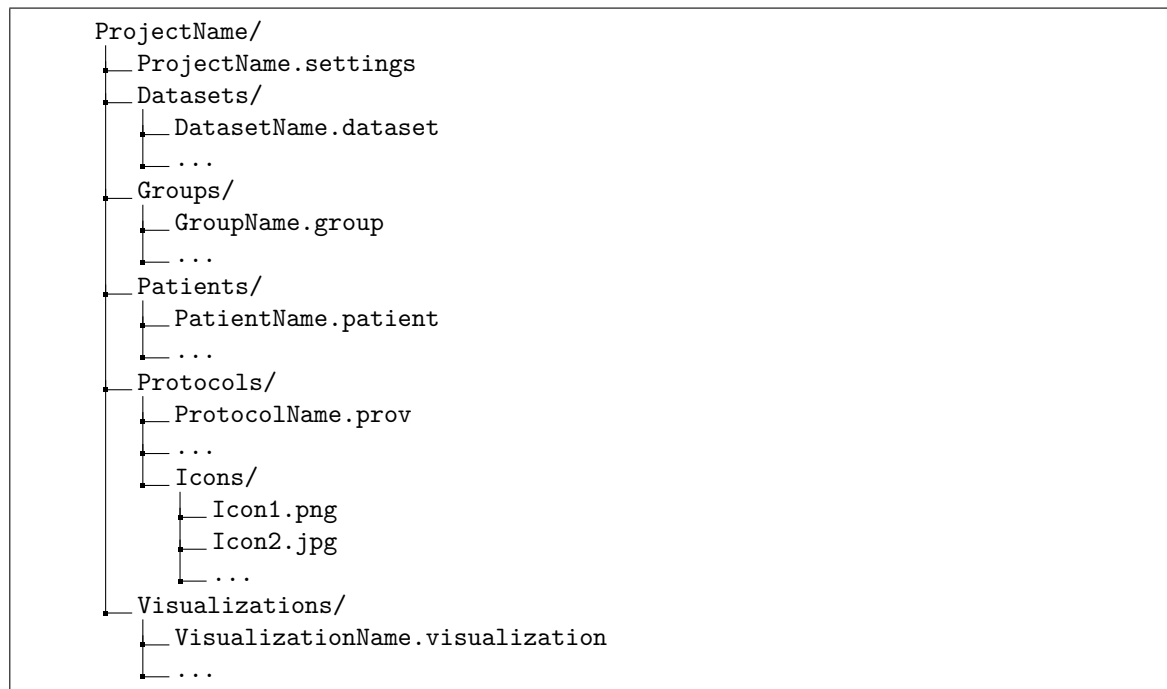


Figure 1: Project directory hierarchy

### 1.1 Settings

The .settings file at the root of the projet folder describes some project settings (Figure 2). This file contains a default path for the anatomical data of the patients and a default path for the functional data (iEEG).

```
{
  "ID": "41001a8a-23a5-409b-8b87-099dbb727a18",
  "Name": "ProjectName",
  "PatientDatabase": "Y:/BrainVisaDB/Epilepsy",
  "LocalizerDatabase": "Y:/LOCA_DATABASE"
}
```

Figure 2: Example project settings file

## 1.2 Patients

This subfolder contains .patient files, which contains metadata of the patients and full paths towards anatomical files for each patient (Figure 3). A 3D mesh can be a single mesh (only one file) or a left-right mesh (one file for the left part and one for the right part of the brain). These files are in GiftI format (.gii). You can also provide additional files to show MarsAtlas parcels (.gii), or a transformation file (to align the mesh and its respective MRI, .trm). The MRIs must be in the NIfTI-1 format (.nii). The implantations must be in the PTS format. The connectivities files are CCEP (amplitude / latencies) text files.

```
{
  "ID": "LYONNEURO_2013_CHAj",
  "Name": "CHAj",
  "Date": 2013,
  "Place": "LYONNEURO",
  "Brain": {
    "Meshes": [
      {
        "$type": "HBP.Data.Anatomy.LeftRightMesh, Assembly-CSharp",
        "ID": "bb1ee4e1-b96b-4973-814d-fb30c4d0d512",
        "Name": "Grey matter",
        "LeftHemisphere": "[...]/LYONNEURO_2013_CHAj_Left.gii",
        "RightHemisphere": "[...]/LYONNEURO_2013_CHAj_Right.gii",
        "LeftMarsAtlasHemisphere": "",
        "RightMarsAtlasHemisphere": "",
        "Transformation": "[...]/LYONNEURO_2013_CHAj.trm"
      }
    ],
    "MRIs": [
      {
        "Name": "Preimplantation",
        "File": "[...]/LYONNEURO_2013_CHAj.nii"
      }
    ],
    "Connectivities": [],
    "Implantations": [
      {
        "Name": "Patient",
        "File": "[...]/LYONNEURO_2013_CHAj.pts",
        "MarsAtlas": "[...]/LYONNEURO_2013_CHAj.csv"
      },
      {
        "Name": "MNI",
        "File": "[...]/LYONNEURO_2013_CHAj_MNI.pts",
        "MarsAtlas": "[...]/LYONNEURO_2013_CHAj.csv"
      }
    ],
    "Epilepsy": {
      "Type": 5
    }
  }
}
```

Figure 3: Example patient file

### 1.3 Groups

This subfolder contains .group files, which are simple lists of patients (Figure 4).

```
{
  "ID": "{813cbd0a-550c-40de-95c7-b387626f9a96}",
  "Name": "VISU",
  "Patients": [
    "Gre_2013_BONn",
    "Gre_2015_BARr",
    "LYONNEURO_2013_CARj",
    "LYONNEURO_2017_PECv",
    "Gre_2015_PATa",
    "LYONNEURO_2014_REId"
  ]
}
```

Figure 4: Example group file

### 1.4 Protocols

This subfolder contains .prov files (Figure 5). The .prov extension stands for **protocol visualization**. Typically, a functional data file in HiBoP is a set of time series (one for each channel) with data for each sample from the beginning to the end of the recording session. Such data are stored in the .eeg/.eeg.ent<sup>1</sup> files. In addition, information about the events of the experimental paradigm is stored in .pos files : event codes occurred at specific samples (e.g. an event of type « 10 » occurred at sample 134567). The .prov file instructs HiBoP on how to epoch the data based on the .pos file. The idea is to create blocs of epochs, each of them centered around a specific type of events, called **main events** (i.e. consider all events with code « 10 » and extract for each of them a window ranging from 200 ms before to 700 ms after that event). Prov files define the blocs you want to visualize (code of the main event), how you want to call them, how you want to sort epochs within a bloc, etc. When you define a new bloc, you must specify the main event (its code) and you can specify secondary events. Secondary events are useful when you want to sort trials: say that the task shows a picture (of type 10 = faces, 20 = landscapes, or 30 = fruits) and that participants have to press one or two buttons to specify whether the picture shows a fruit (1 = « yes », 2 = « no »). You would create for instance a bloc with all epochs around an event of type « 10 » (faces) and sort those epochs according to reaction time (the latency of event 1 or 2 relative to event 10 within each epoch). To do that in HiBoP, you would specify a secondary event in the « face » bloc with type 1 and 2 (i.e. all events of type 1 or 2), then write the « sorting rule » C0L1: which means that you want to sort trials according to the code of the main event (here 10, so no sorting necessary), then for all ex-aequo epochs, use the relative latency of the secondary event (1 or 2, relative to the main event 10). You could set the main event to 10 and 20 and 30 to include in your bloc « faces », « landscapes » and « fruits », then the « sorting rule » C0L1 would sort trials according to the main event code first (all faces, then all landscapes, then all fruits) and then, within a given category, sort according to reaction time (secondary event 1 and 2) The baseline is used for the trial matrices, which can include a baseline correction. This baseline correction is described in preferences (section 2.1). The rest of the .prov file is straightforward : you can ask HiBoP to show .png or .jpg files at specific latencies relative to the main event.

---

<sup>1</sup>ELAN format – ELAN is freely available at <http://elan.lyon.inserm.fr>

```

{
  "ID": "{9fd88c3e-ce7f-4c1c-83b1-f4d1edcbfe5a}",
  "Name": "LEC1",
  "Blocs": [
    {
      "IllustrationPath": "./Protocols/Icons/LEC1_SEM.jpg",
      "ID": "d3ce596a-8918-4ba1-829d-5aa73efe060e",
      "Name": "SEMAN",
      "Position": {
        "Row": 1,
        "Column": 1
      },
      "Sort": "COL1",
      "Window": {
        "Start": -500.0,
        "End": 3500.0
      },
      "Baseline": {
        "Start": -200.0,
        "End": 0.0
      },
      "Events": [
        {
          "Name": "SEM",
          "Codes": [
            10
          ],
          "Type": 0
        },
        {
          "Name": "RESPONSE",
          "Codes": [
            1,
            2
          ],
          "Type": 1
        }
      ],
      "Scenario": {
        "Icons": [
          {
            "IllustrationPath": "./Protocols/Icons/LEC1_SEM.jpg",
            "Name": "PIC",
            "Window": {
              "Start": 0.0,
              "End": 2000.0
            }
          }
        ]
      }
    }
  ]
}

```

Figure 5: Example prov file (one bloc of the LEC1 LOCA)

## 1.5 Datasets

This subfolder contains .dataset files, which summarize and link together anatomical and functional data for a given cognitive task (Figure 6). A dataset is a list of data information, each of them connecting one patient (and its anatomical data) with functional data (.eeg and .pos files) for a single experiment. Each data information can be normalized indepently from what is set in the preferences (see Section 2.1 for detailed information about normalization).

```

{
  "ID": "{dfad2720-a0ec-49d8-b552-787f28c78c3b}",
  "Name": "VISU_dataset",
  "Protocol": "{9d12c695-7ce6-49d4-82d0-a9da12ba776a}",
  "Data": [
    {
      "Name": "alpha_beta",
      "Patient": "Gre_2013_BONn",
      "Measure": "EEG data",
      "EEG": "[...]/Gre_2013_BONn_VISU_f8f24_ds8_sm0.eeg",
      "POS": "[...]/Gre_2013_BONn_VISU_ds8.pos",
      "Normalization": 4
    },
    {
      "Name": "alpha_beta",
      "Patient": "Gre_2015_BARr",
      "Measure": "EEG data",
      "EEG": "[...]/GRE_2015_BARr_VISU_f8f24_ds16_sm0.eeg",
      "POS": "[...]/GRE_2015_BARr_VISU_ds16.pos",
      "Normalization": 4
    },
    {
      "Name": "alpha_beta",
      "Patient": "Gre_2014_MADl",
      "Measure": "EEG data",
      "EEG": "[...]/GRE_2014_MADl_VISU_f8f24_ds8_sm0.eeg",
      "POS": "[...]/GRE_2014_MADl_VISU_ds8.pos",
      "Normalization": 4
    }
  ]
}

```

Figure 6: Example dataset file

## 1.6 Visualizations

This subfolder contains .visualization files (Figure 7), which describe the contents of a visualization: it is composed of patients and columns. Each column allow one to visualize either the anatomy of the chosen patients or iEEG data for a specific protocol/bloc/dataset/data. The also stores the configuration of the visualization or of a particular column or site.

```

{
  "ID": "25c11585-fb23-4c70-b4d3-504f9851da11",
  "Name": "Visualization",
  "Patients": [
    "Gre_2009_CHIm",
    "Gre_2009_MWAm",
    "LYONNEURO_2013_CARj",
    "LYONNEURO_2013_DUMp"
  ],
  "Configuration": {
    "Brain Color": 15,
    "Brain Cut Color": 14,
    "Colormap": 13,
    "Mesh Part": 2,
    "Mesh": "MNI",
    "MRI": "MNI",
    "Implantation": "MNI",
    "Edges": false,
    "Sites": false,
    "MRI Min": 0.0,
    "MRI Max": 1.0,
    "Camera Type": 0,
    "Cuts": [],
    "Views": []
  },
  "Columns": [
    {
      "Dataset": "{dfad2720-a0ec-49d8-b552-787f28c78c3b}",
      "Protocol": "{9d12c695-7ce6-49d4-82d0-a9da12ba776a}",
      "Bloc": "3916b0a0-7ebc-4934-94e5-2308bf511f7c",
      "Label": "alpha_beta",
      "Configuration": {
        "ConfigurationBySite": {},
        "Site Gain": 1.0,
        "Site Maximum Influence": 15.0,
        "Transparency": 0.8,
        "Span Min": 0.0,
        "Middle": 0.0,
        "Span Max": 0.0,
        "Regions Of Interest": []
      }
    }
  ]
}

```

Figure 7: Example visualization file

## 2 HiBoP description

This section will describe in details how to use HiBoP to create a HiBoP project and how to visualize the iEEG data. To open HiBoP, double click on HiBoP.exe (for Windows; for Linux and MacOS, read the README.md file located next to the executable). If you have any doubt about what an interface element does, you can hover it for a second and a tooltip will appear telling you what it does.

### 2.1 Preferences

A lot of global parameters shared through all projects can be set using the preference menu (Figure 8). These parameters are stored in a file located next to the HiBoP executable (GeneralSettings.txt).

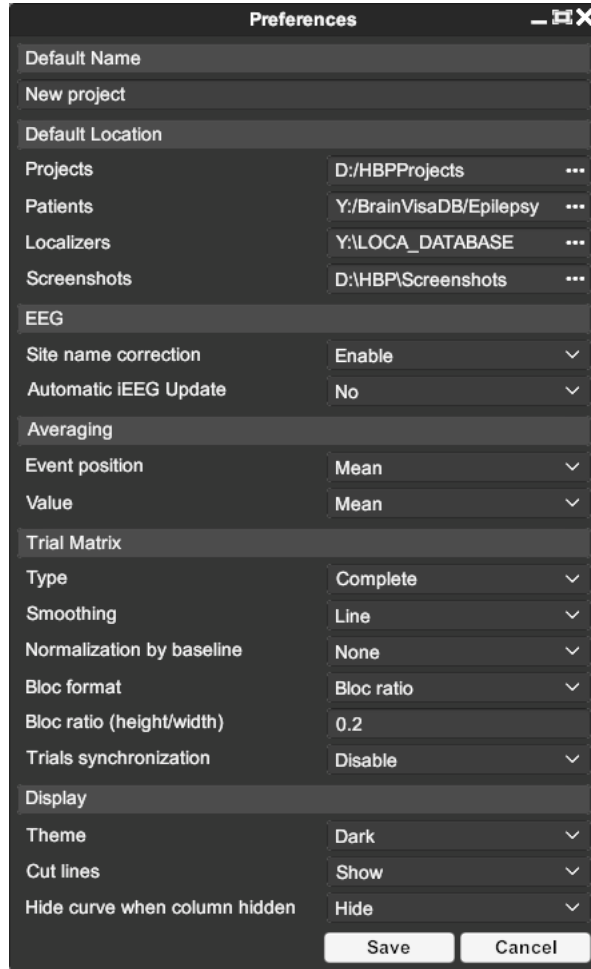


Figure 8: Preferences

- **Default Name** : Default name for a new project
- **Projects** : Default path to search for projects
- **Patients** : Default path to patients database
- **Localizers** : Default path to functional data
- **Screenshots** : Default path to save screenshots
- **Site name correction** : Change the name of a certain pattern of site names (for instance "xp4" becomes "X'4")
- **Automatic iEEG Update** : Update the iEEG values on the brain automatically if they are outdated
- **Event position** : How to average the secondary events position
- **Value** : How to average the values
- **Type** : Show every matrices of a selected protocol or only blocs chosen in columns
- **Smoothing** : Should the matrices be smoothed horizontally (to have a better resolution)
- **Normalization by baseline** : The following formula is applied to each value  $v$

$$v = \frac{v - \mu_b}{\sigma_b}$$



where  $b$  is an array of the baseline values depending on this parameter,  $\mu_b$  is their mean and  $\sigma_b$  is their standard deviation

- **Bloc format** : How the trial matrices should be displayed (parameters depend on this parameter, try yourself to see which one is better for you)
- **Trial synchronization** : Should the matrices be synchronized when selecting specific trials
- **Theme** : Theme of the software (light theme is still under development)
- **Cut lines** : Should the cut lines appear on the cuts ?
- **Hide curve when column hidden** : Should the curves of the hidden column appear ?

## 2.2 Project management

### 2.2.1 Creating, opening and saving a project

When creating a new project, you have to specify the name of your new project, the location where it will be saved, the location of the anatomical database and the location of the functional database (Figure 9). The two last fields are optional.

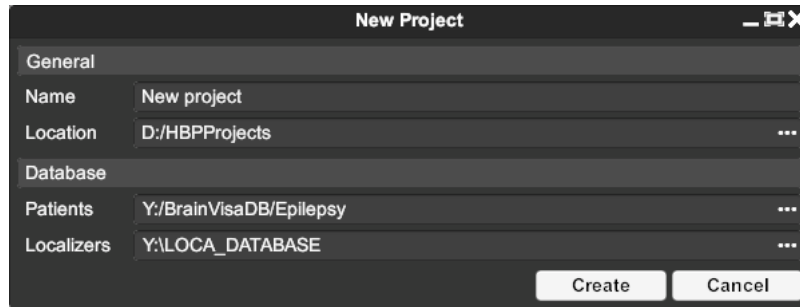


Figure 9: Create a new project

In order to open a previously created project (Figure 10), you have to tell where your projects are located. Once these are loaded, you can either double click on the project you want to open or select the project and then click on Open. You can also sort projects using specific parameters.

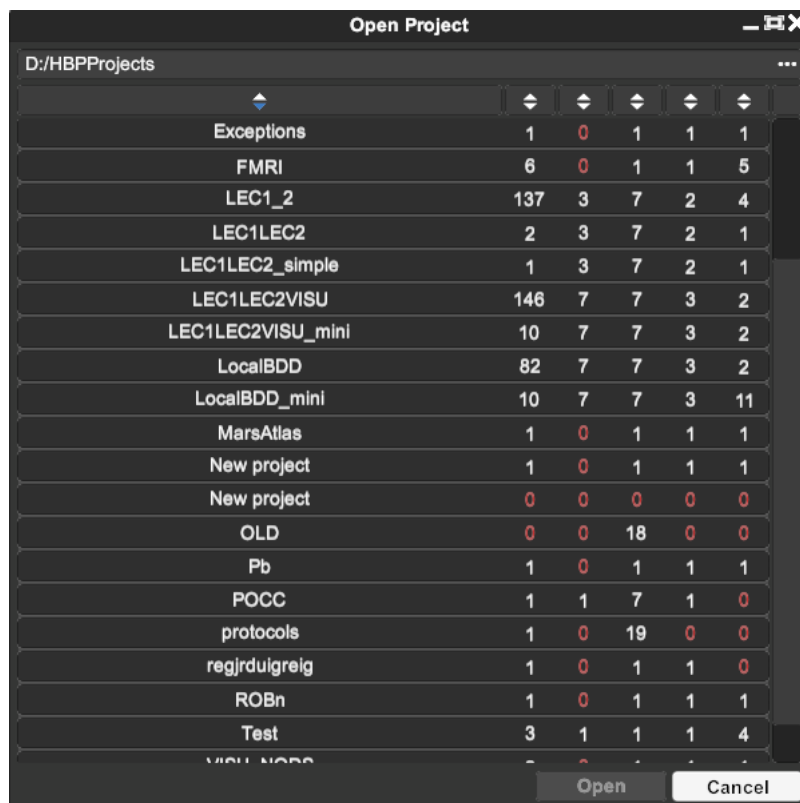


Figure 10: Open a project

If you want to save a project to a different location or with a different name than when it was open, you can use the "Save as" menu (Figure 11).

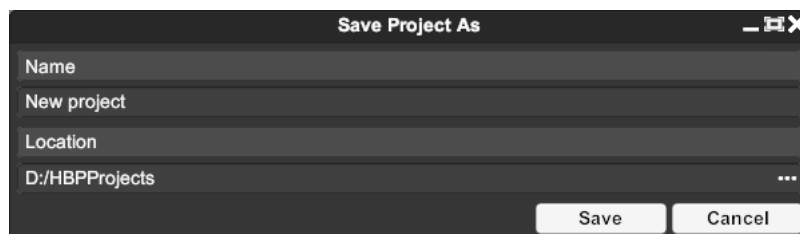


Figure 11: Save a project as

All of these menus are located in the File menu at the top of the screen.

### 2.2.2 Patients management

To add patients to the opened project, you have to use the patients manager (Figure 12). The window has two panels: the left panel displays all available patients in the database (found in the folder you specify at the top of the window) whereas the right panel displays patients added to this project. For each patient, numbers indicate the status of their respective anatomical data (Meshes, MRIs, Implantations and Connectivities). To add or remove patients, you can either select them in any list and use the arrows to add or remove them, or you can create new patients from scratch using the "+" button in the center. In order for the anatomical database to be correctly loaded, you have to use a specific hierarchy described in appendix B.

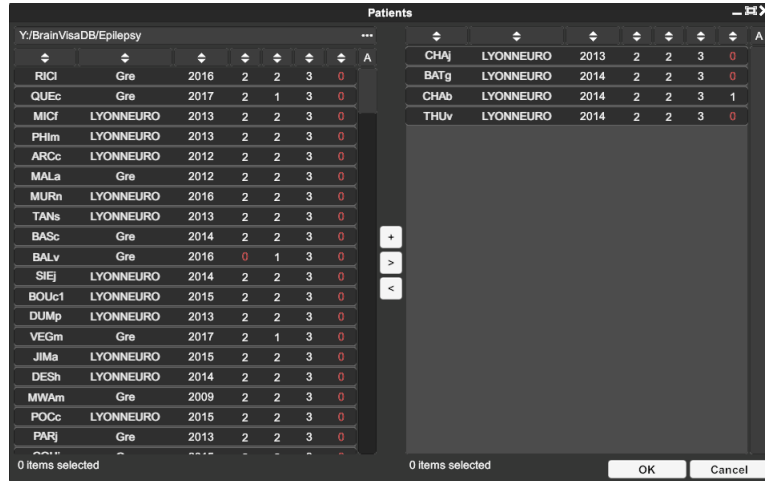


Figure 12: Patients manager

To edit any loaded patient, you can double click<sup>2</sup> on it. This will open the patient modifier window (Figure 13) which will allow you to edit anything concerning the metadata or the anatomical data of this patient. You can navigate between anatomical data types using the tabs in the middle of the window.

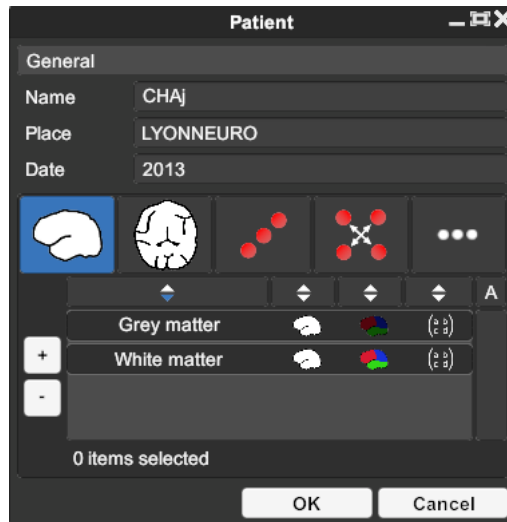


Figure 13: Patient modifier

If you want to add a mesh, you can use the "+" button located to the left; if you want to edit a mesh, you can double click the mesh item, and this will open a mesh modifier window (Figure 14) which will allow you to edit the path to the GIFTI files or to the transformation file.

<sup>2</sup>The process of double clicking an item to edit it is common to most windows of HiBoP

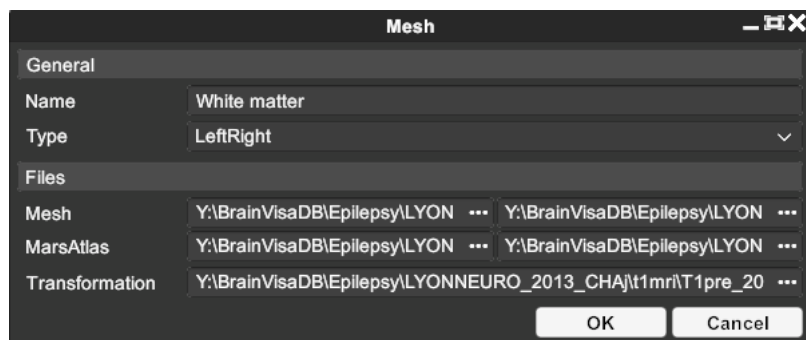


Figure 14: Mesh modifier

As stated in section 1, you can also create groups of patients to add them more easily to the visualizations. This is done through the groups manager window (Figure 15). To create a new group, use the "+" button on the left. To remove existing groups, select the group(s) you want to remove, and use the "-" button on the left<sup>3</sup>. Adding and removing patients from a group is very similar to the patients manager window (Figure 16).

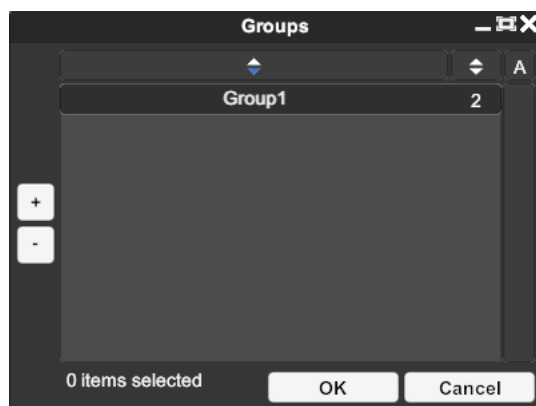


Figure 15: Groups manager

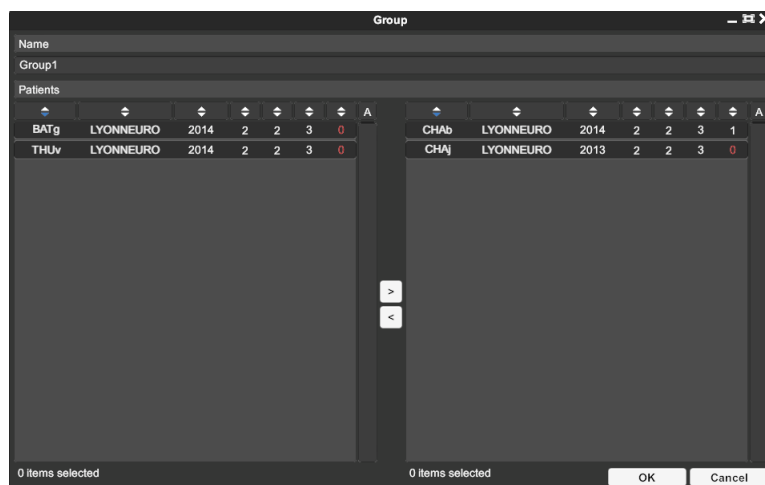


Figure 16: Group modifier

<sup>3</sup>The process of using "+" button to add and "-" button to remove is common to most windows of HiBoP

### 2.2.3 Data management

First thing to do concerning data management is to create a protocol that will epoch the iEEG data. You can do that using the protocol manager window (Figure 17). You can also import an existing .prov file to the project by using the "Import..." button on the bottom.

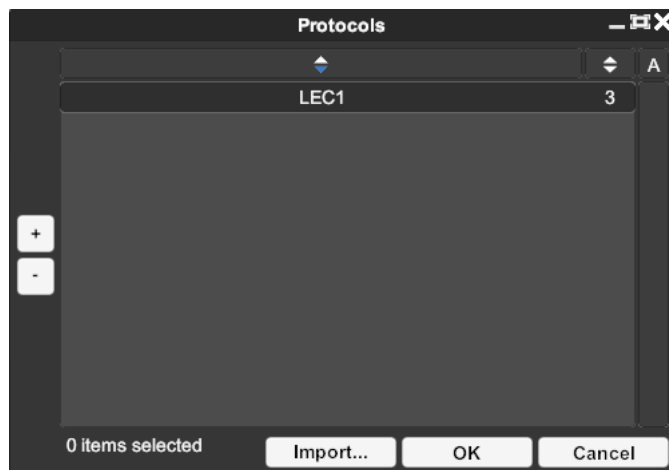


Figure 17: Protocols manager

Editing a protocol consists of defining the blocs for this protocol. This is done with the protocol modifier window (Figure 18). There are two types of blocs in a protocol: a main bloc (first column) will be used for metadata and main epoching, whereas a secondary bloc (second and third columns) will be used to epoch around a second or a third main event (this is used if you want to hide a pause between two important data). To create a new bloc, click on an empty box. To edit a bloc, double click on an existing bloc.

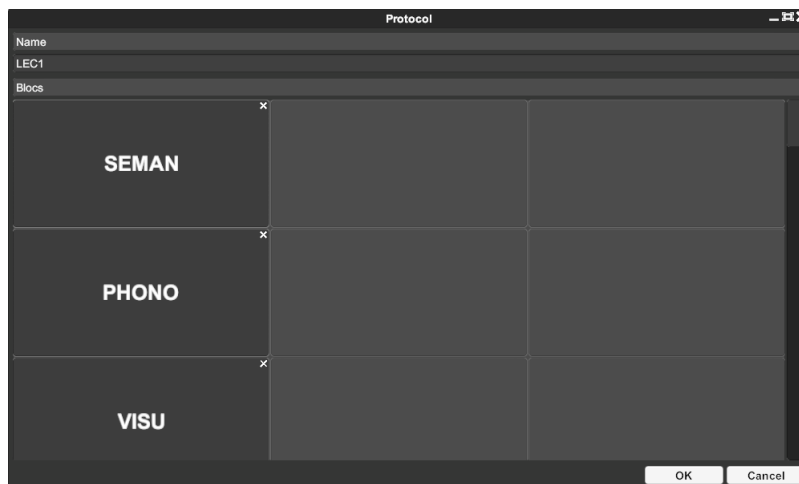


Figure 18: Protocol modifier

A bloc can be edited with the bloc modifier window (Figure 19). There you can specify the name, the sorting method, the window and the baseline window, as well as events and icons for the iconic scenario, which can be edited by double clicking on them. To add an illustration to the bloc, you can click on the icon placeholder in the general category.

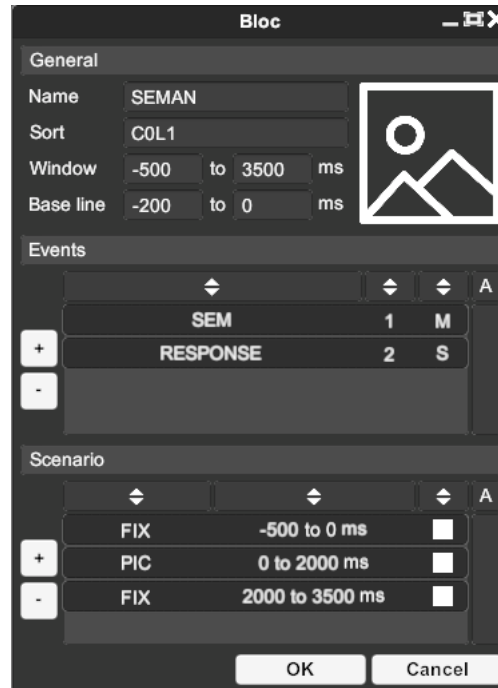


Figure 19: Bloc modifier

When your protocols are correctly defined, you are ready to create your datasets. This will be done using the datasets manager window (Figure 20).

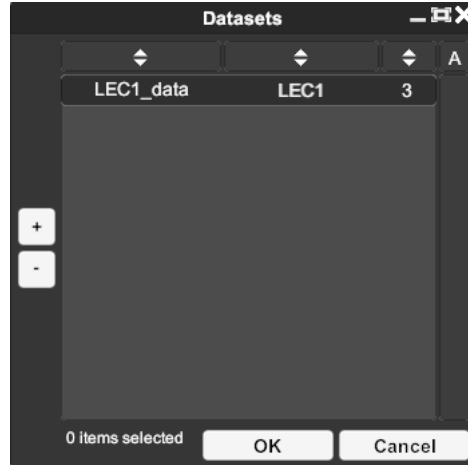


Figure 20: Datasets manager

For a given dataset, you need to give it a name and assign a protocol. All the data included in this dataset will be epoched regarding the chosen protocol. With the dataset modifier window (Figure 21), you can add and edit data information. Each data information has a little colored box which indicates the status of the data information : red means it is unusable, whereas green means you can visualize it. You can click on this box to have more information about the data information.

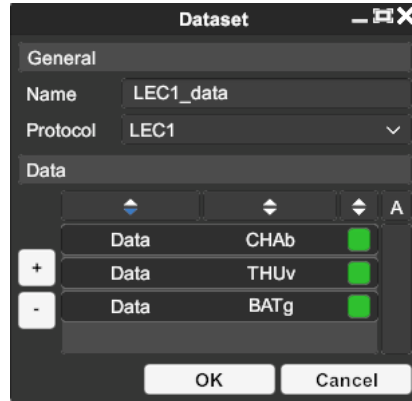


Figure 21: Dataset modifier

A data information has a name and is linked to a patient. A single dataset can not have two data information with the same pair [name, patient]. In order to visualize different data from the same dataset in the same column of a visualization, you have to give them the same name (thus the two patients concerned by these data will be different). Then, you have to fill in the paths to the functional data files (.eeg and .pos). The label in front of the EEG file path corresponds to the label of the data you want to extract from the EEG file (usually, it is "EEG data"). You can also change how this particular data will be normalized (this overwrites the global parameter for normalization).

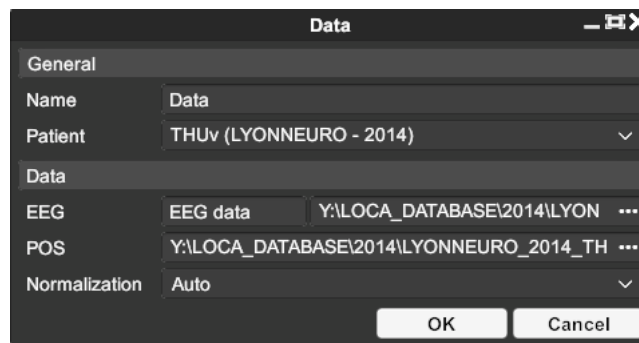


Figure 22: Data information modifier

#### 2.2.4 Visualizations management

When everything is set up, you can create visualizations. This is done using the visualization manager window (Figure 23).

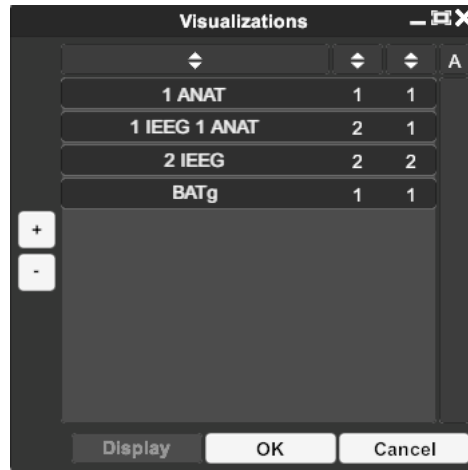


Figure 23: Visualizations manager

A visualization has a name, patients and columns. To edit all of this information, you can use the visualization modifier window (Figure 24). To add patients to the visualization, use the arrows (same process than adding patients to the project). To add a column, click on the "+" button under the "Columns" panel. You can then edit the column by setting the name, the type (anatomy only or iEEG), the protocol, the bloc, the dataset and the data name to be used in this column. Do not forget that if multiple patients are added to the visualization, they must have at least one data information with the same name.

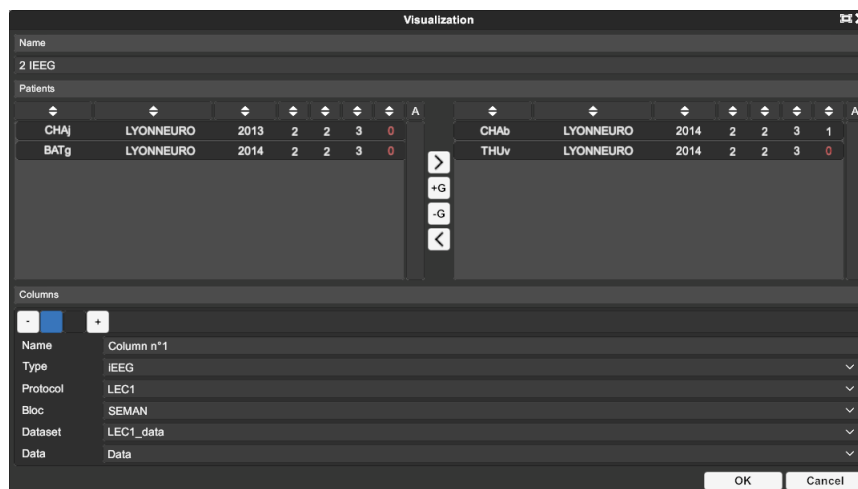


Figure 24: Visualization modifier

Once your visualizations are ready to be displayed, you can select them and click the "Display" button.



## 2.3 3D Visualization

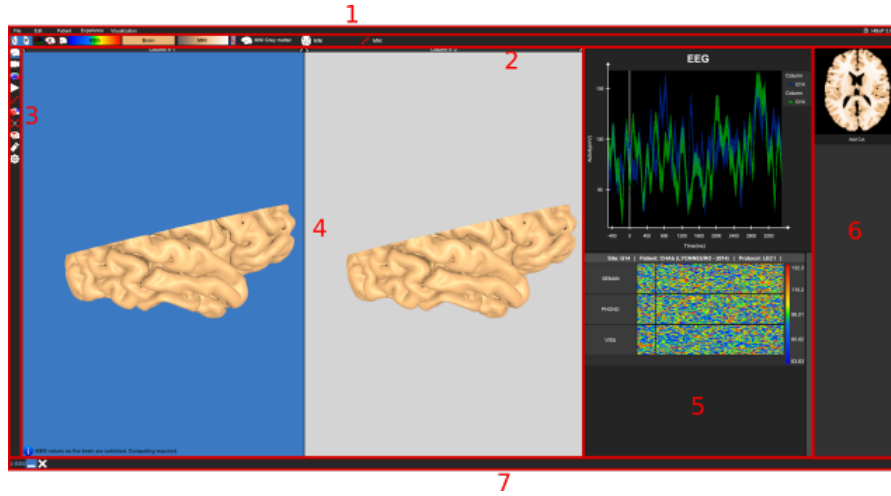


Figure 25: HiBoP user interface

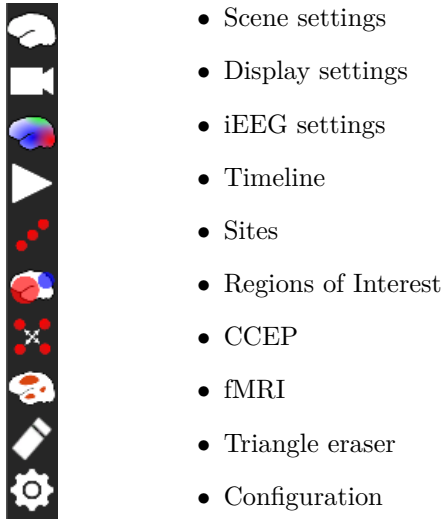
The main user interface of HiBoP can be decomposed as follow (Figure 25) :

- 1 - Main menu (data management, see section 2 for more information)
- 2 - Toolbar (used to do actions on the 3D scene, more information in later sections)
- 3 - Toolbar selector (to select which toolbar you want to use)
- 4 - 3D visualization (main visualization window)
- 5 - Graphs and trial matrices (precise information of iEEG data, see section 2.4)
- 6 - Cut panel (manage cuts, see section 2.3.1)
- 7 - Opened visualizations (used to minimize or close opened visualizations)

Elements in parts 4, 5 and 6 can be resized using the black handlers between them. Columns of the 3D visualisations can be moved by drag and dropping their labels at the top of the column. You can interact with the scene in many ways :

- Left click on a view to select the corresponding view / column / scene or left click on a site / on a ROI to select it.
- Right click and drag to rotate the brain surface
- Middle click and drag to translate the brain surface
- Use the scroll wheel to control the zoom
- Drag and drop the name of a column to swap it with another column or to insert it between two columns

The left panel allows you to change between toolbars. Each toolbar has its own functions, which will be explained in more details in later sections.



### 2.3.1 Cuts

Before getting into the toolbars, let's talk about cuts. Indeed, HiBoP allows you to cut the 3D mesh (using the MRI to fill the cut plane) with the right panel. To add a cut, click on the "Add cut" button. You can change parameters on the cut after clicking on it to open the parameters panel (Figure 26). You can change its orientation (Axial, Coronal, Sagittal or Custom), you can flip it (Axial, Coronal and Sagittal) or set up a custom normal (Custom), you can change the position of the cut using the slider or the "+" and "-" buttons for more precision. To delete a cut, open the parameters panel and click on the little cross on the top-right of the cut.



Figure 26: Cut parameters panel

The following sections will explain the different functionalities of each toolbar.

### 2.3.2 Scene settings



- Display both parts, only the left part or only the right part of the selected mesh (only works for left-right meshes).
- Display mars atlas on the mesh (only works if the mars atlas field of the mesh is filled).

- Display the edges of the mesh.
- Switch between the soft cut mode (keep everything behind a cut) and the hard cut mode (cut everything in front of a cut).
- Change the colormap of the iEEG values.
- Change the color of the brain.
- Change the color of the cuts.
- Change the threshold of the MRI values (contrast). Use the sliders to change these values.
- Change the mesh (MNI meshes are available no matter the number of patients, the patient meshes are only available if the patient is alone in the visualization).
- Change the MRI (patient MRIs are only available if the patient is alone in the visualization).
- Change the implantation (an implantation is available only if each patient of the visualization has an implantation with the same name)

### 2.3.3 Display settings



- Add or remove a view (maximum 5 views).
- Set up standard views (3 views with 3 different points of view).
- Reset the camera position of the selected view.
- Reset the size of the views (if resized using the handlers).
- Automatic rotation of the brain (you can set the rotation speed with the slider).
- Change the camera behaviour (T = trackball camera, O = orbital camera).
- Take a raw screenshot of the scene which will be saved in the default screenshot directory (set in preferences).
- Save the scene as files in a directory named after the name of the visualization in the default screenshot directory (one image per opened view, one image per cut, one image and one svg for the graph, one image for the trial matrices, one csv for each curve containing the values).

### 2.3.4 iEEG settings



- Trigger global mode (any changes will be applied to all columns)
- Change the size of the sites
- Change the size of the influence sphere of the sites (to compute the iEEG on the brain surface)
- Change the transparency of the iEEG value on the brain surface
- Change the threshold values for the iEEG (anything less than the minimum value will be of the same color as the minimum value, the same apply for the maximum value; the rest of the values will increase linearly from minimum to middle, then decrease linearly from middle to maximum). You can change these values by either using the sliders or editing the numbers in the boxes.
- Compute the iEEG values on the brain surface or clean the brain.

### 2.3.5 Timeline



- Trigger global mode (any changes will be applied to all columns).
- Make the timeline play automatically.
- Increase the current timeline ID by a specific amount of samples (this also controls the speed of the play, X samples will be played each second).
- Loop the timeline (when playing automatically).
- Slider to control the timeline precisely; the red mark indicates the main event, the blue marks indicates the secondary events (as defined in the .prov file).

### 2.3.6 Sites



- Selected site name.
- Display all sites in the scene.
- Hide blacklisted sites.
- Change the state of some sites (you can select which sites using filters). Available states are :
  - Excluded : an excluded site is not in the iEEG computations, but it remains visible in the scene
  - Highlighted : a highlighted site appears without transparency in the scene
  - Marked : a marked site appears in another color when the iEEG are not computed (the behaviour is the same as an included site)
  - Blacklisted : a blacklisted site will be barely visible and not included in any calculation
- Display a list of all sites with their respective state. Click on the name of a site to select it.
- Compare two sites : select a site, click on this button, then select another site to compare them.
- Load the single patient scene corresponding to the patient whose site is selected.
- Cut approximately around the select site.
- Copy the states of the sites of the selected column to every other columns.
- Save the states of the sites in a .csv file.
- Load a .csv file with the states of the sites to the selected column.

### 2.3.7 Regions of Interest

Regions of Interest (ROI) are useful if you only want to study a small group of sites. They are composed of multiple spheres with different positions and radiuses. If the option "Show All Sites" of the sites toolbar is not enabled, only the sites of the selected ROI will be included in the iEEG computations on the brain. A curve will also be generated for the selected ROI of each column (section 2.4).



- Add a new ROI in the selected column.
- Select a ROI for the selected column.
- Change the name of the selected ROI.
- Remove the selected ROI.
- Select a sphere of the ROI.
- Delete the selected ROI.
- Copy the selected ROI to other columns.
- Save the selected ROI to a .roi file.
- Load a .roi file to the selected column.

After adding a ROI to the selected column, you can left click on the mesh to add spheres to this ROI. You can select a sphere using either the toolbar or by left clicking on it. You can change the size of the selected sphere with the scroll wheel.

### 2.3.8 CCEP

This toolbar will allow you to enable CCEP mode (which will disable iEEG mode) and choose from the Connectivity files which one you want to display. To select a source, simply select the site you want as source.

### 2.3.9 fMRI



- Add or remove a fMRI to the anatomic MRI. If iEEG values are computed, the fMRI will not be displayed.
- Change the threshold values of the fMRI.
- Change the transparency of the fMRI.

### 2.3.10 Triangle eraser



- Reset the triangle eraser.
- Choose how you want to erase triangles (one triangle, cylinder, zone). To erase triangles, left click on the mesh where you want to erase them.
- Expand the erased zone.
- Invert the erased zone.
- Revert last action.

### 2.3.11 Configuration

This toolbar allow you to save, load or reset the configuration of the selected scene. Beware, saving the configuration will just save it to the loaded visualization: to write the changes in the corresponding file, you still have to save the whole project.

## 2.4 Graphs and Trial Matrices

The middle panel allows you to examine more precise information about the iEEG data of specific sites. You can open it using the handlers or if it is closed, it will automatically open once you select a site.

### 2.4.1 Graphs

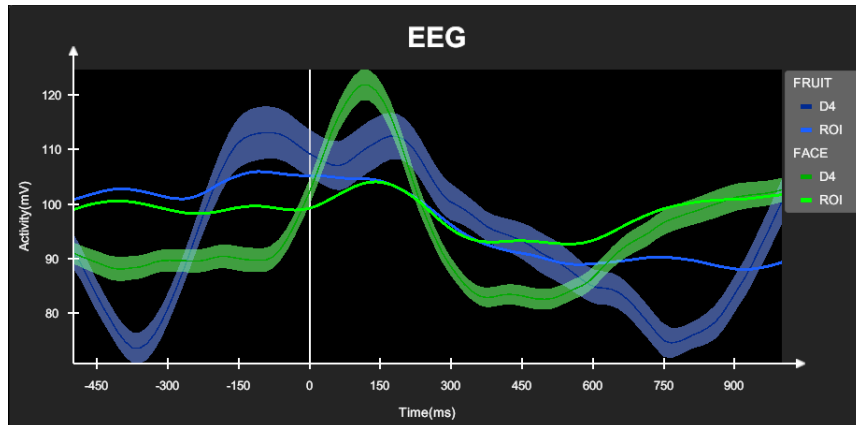


Figure 27: Graph for a protocol and two blocs (site D4 and ROI)

Curves are generated for each column (or not minimized columns, depending on the chosen preferences) :

- One curve for the last selected site (the column in which the site has been selected does not matter)
- One curve for the compared site if you are comparing two sites using the "Compare Site" tool in the site toolbar
- One curve for the ROI if it exists. The ROI is considered as a single site which has the combined activity of all sites (excluding excluded or blacklisted sites) within it.

An example of graph can be found at Figure 27. A curve is the mean or median (depending on global preferences) of the iEEG activity of a site. The shapes around the single site curves correspond to the standard error of the mean (SEM). You can interact with the graph in order to get the most of it :

- You can hover a point with your mouse to see precise coordinates of this point.
- You can zoom in and out using the mouse wheel and left click and drag to move the graph.
- You can left click on an axis to precisely edit the viewport of the curve. This small window also has an "Auto" button; this functionality adapts the viewport so you can see every points of each curve.
- You can hide a curve by left clicking on the corresponding item in the legend.

### 2.4.2 Trial matrices

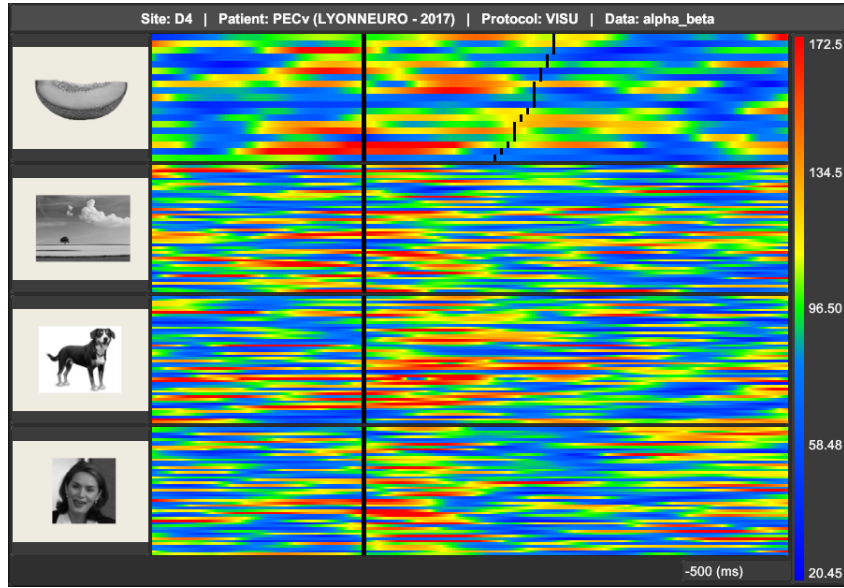


Figure 28: Trial matrix of a site

Trial matrices are useful to inspect iEEG activity of a site trial by trial. Trials are sorted according to the sorting method of a protocol bloc. You can change the color scale by clicking on it (same process as changing the viewport of the graph). Again, you can hover the matrix to get precise information about the hovered section (like in graphs). You can also select specific trials; only the selected trials will be taken into account when computing the values of the corresponding curve. To select trials, you can :

- Select a single trial by left clicking on it
- Select multiple trials by either shift left clicking on the trials you want to select, or by pressing the left click button dragging the mouse and releasing the button.
- Move the selection with the scroll wheel

## A HiBoP Database Manager

The HiBoP Database Manager (DB Manager) can be used to manage the anatomical and functional databases (as long as they respect the hierarchy described in appendix B), create HiBoP projects using huge amounts of data in no time or convert the databases to the Brain Imaging Data Structure (BIDS) format<sup>4</sup>. The main window (Figure 29) of the DB Manager has two main columns. The left column allows you to select patients to visualize their anatomical data, the right column is for functional data.

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<sup>4</sup>More information about BIDS can be found at <http://bids.neuroimaging.io/>

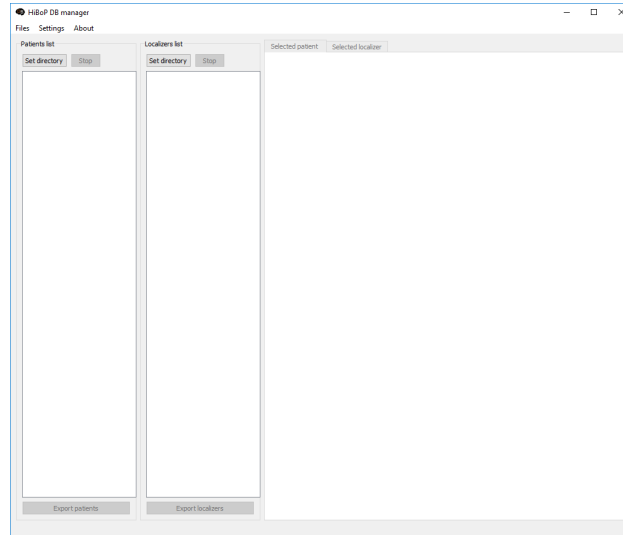


Figure 29: Main window of the DB Manager

## A.1 Anatomical data

Click « Set Directory » in the first column to select the folder containing the anatomical database. After choosing a folder, patients found within that database will appear in the first column. Selecting one patient will display in the middle panel (Figure 30) a list of files types and their status (present in green or absent in red). Some of the files can be visualized or opened directly from that interface. The display allows the user to quickly browse through patients and identify missing files.



Selected patient

Selected localizer

Infos

Name: BATg

Date: 2014

Place: LYONNEURO

Find associated localizer

	Ctrl/c	Exists	Open
T1mri directory	Path		Dir
Implantation directory	Path		Dir
Report	Path		PDF
Implantation schema	Path		PDF

Pre 2014-6-2

Information

	Ctrl/c	Exist	Open
Pre directory	Path		Dir
MRI (*.nii)	Path		File
APC (*.APC)	Path		File
Align fMRI			
Transformations (*.trn)	Path		Dir
To scanner based	Path		File
Implantations (*.pts)	Path		Dir
Pre scanner-based	Path		File
ACPC	Path		File
MNI	Path		File
Mars Atlas	Path		File
Mesher (*.gii)	Path		Dir
Head	Path		File
Lhemi	Path		File
Rhemi	Path		File
Lwhite	Path		File
Rwhite	Path		File
Lwhite mars parcel	Path		File
Rwhite mars parcel	Path		File

Figure 30: Patient panel

## A.2 Functional data

Click « Set Directory » in the second column to select the folder containing the functional database. Once loaded, the database can be inspected patient by patient in a way similar to anatomical data (Figure 31). Missing files can be quickly identified this way.

Selected patient

Selected localizer

Infos

Place : Gre

Date : 2009

Name : Gre\_2009\_Chim

Find associated patient

Experiences

LEC1

LEC2

VISU

Ctrl/c

Exists

Open

Exp dir

Path

Dir

Focus experience

TRC

Path

File

EEG

Path

File

ENT

Path

File

POS

Path

File

POS DS

Path

File

f8f24

sm list

Ctrl/c

Exists

Open

sm0

Path

File

sm250

Path

File

sm500

Path

File

sm1000

Path

File

sm2500

Path

File

sm5000

Path

File

f50f150

sm list

Ctrl/c

Exists

Open

sm0

Path

File

sm250

Path

File

sm500

Path

File

sm1000

Path

File

sm2500

Path

File

sm5000

Path

File

Figure 31: Localizer (iEEG data) panel

### A.3 Exporting data

A panel in the right part (Figure 32) of the window allows the user to filter patients. By checking boxes, you can quickly identify patients matching certain criteria (e.g. all patients with task LEC2, with gamma envelopes and anatomical MRIs).

**Filters**

**I. LOCALIZER**

1. Year

☒ 2009 ☒ 2010 ☒ 2011 ☒ 2012  
☒ 2013 ☒ 2014 ☒ 2015 ☒ 2016  
☒ 2017

2. Experience

☒ All ☐ Any

☒ VISU ☐ MVIS ☐ AUDI ☒ LEC1  
☒ LEC2 ☐ MOTO ☐ ARFA ☐ REST  
☐ MCSE ☐ MASS ☐ NORM ☐ STAB  
☐ MARA ☐ MLAH ☐ MVEB

3. File

☒ EEG computed  
☐ TRC present  
☐ POS present

☒ enable frequency filter ☒ enable temporal smoothing filter

4. Frequency

☐ f8f24 ☒ f50f150

5. Temporal smoothing

☐ has trials  
☒ sm0 ☐ sm250 ☐ sm500  
☐ sm1000 ☐ sm2500 ☐ sm5000

**II. PATIENT**

☒ pre-implantation exists ☐ post-implantation exists

1. Pre-implantation

☒ IRM ☒ Sites MNI ☒ Sites SB  
☒ Transfo pre\_to\_SB ☒ Meshes  
☐ APC

2. Post-implantation

☒ IRM ☒ Sites SB  
☒ Transfo post\_to\_pre

3. Mars-atlas

☐ enable atlas filter Edit filter

**RESULTS (patients associated with localizers filtered)**

☒ LYONNEURO\_2013\_DUMP  
☒ LYONNEURO\_2013\_PECp  
☒ Gre\_2014\_MARj  
☒ Gre\_2013\_CHAp  
☒ Gre\_2015\_BARr  
☒ Gre\_2015\_BESd  
☒ Gre\_2014\_MADl  
☒ Gre\_2014\_BASc  
☒ LYONNEURO\_2013\_CHAf  
☒ LYONNEURO\_2013\_CHAj  
☒ Gre\_2014\_BARf  
☒ Gre\_2013\_MANa  
☒ Gre\_2015\_COJj  
☒ Gre\_2014\_WOJp  
☒ LYONNEURO\_2013\_LARa  
☒ LYONNEURO\_2013\_LARj

Initial: 82  
After filter: 82  
Checked: 82

Check all  
Uncheck all  
Export filtered patients  
Export filtered localizers  
Export to BIDS  
Create HiBoP project

Figure 32: Export panel

You can filter using the following parameters:

- **Year:** Choose patients of specific years.
- **Experience:** Choose patients based on the tasks they performed.
- **Files:** Choose patients with specific functional files (.TRC, .eeg, .pos)
- **Frequency:** Choose patients with processed data using specific frequency range<sup>5</sup>.
- **Smoothing:** Choose patients with data with a specific temporal smoothing.
- **Patient:** Choose patients with specific anatomical criteria: pre implantation files, post implantation files, presence of a site in a specific Mars Atlas area etc.

Once you have selected the patients you want to study (using the boxes next to their names; a red name means this patient does not match one of the parameters you chose), you can create a HiBoP project using these patients, export these patients to BIDS format, export the anatomical files (section A.3.1) or the functional files (section A.3.2) to another folder.

### A.3.1 Anatomical data

If you click on the "Export patients" button, you will see the window presented in figure 33. There you can filter the patients even further and choose which files you want to export (MRI, Implantations, Meshes etc.). When you are ready to export, click on the "Start export" button and select the folder that will be containing your exported anatomical database. You can cancel the export at any time.

<sup>5</sup>Hilbert transform: f8f24 combines the alpha and beta frequency bands, f50f150 corresponds to the gamma band.

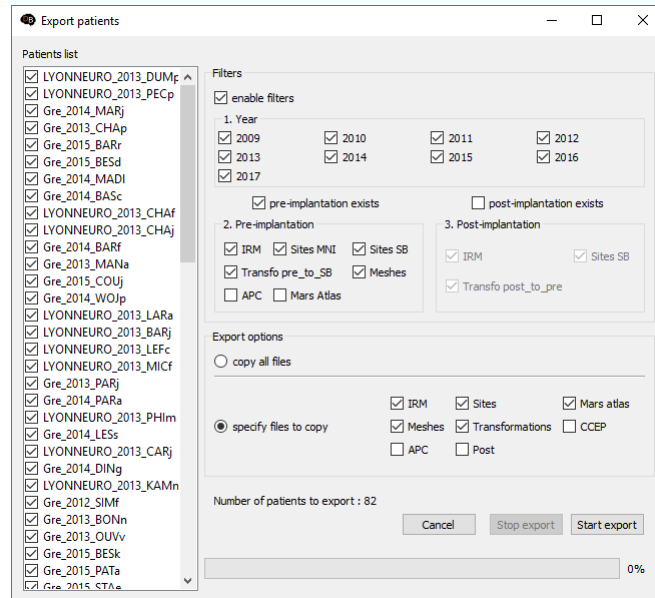


Figure 33: Export patients window

### A.3.2 Functional data

Exporting functional data is very similar to exporting anatomical data (Figure 34). The main difference concerns which file you want to export:

- **copy all files:** this option will export anything found in the functional database no matter what you selected in the filters (for the patients you checked).
- **minimal copy:** this option will only export what has been checked (e.g. if you checked VISU, f50f150, sm0 it will only export the files corresponding to these parameters).

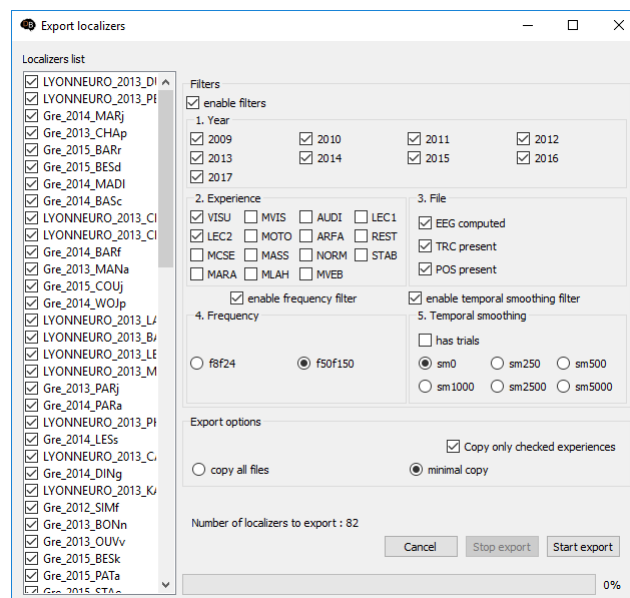


Figure 34: Export localizers window

## A.4 Resources files

A folder called "resources" is present next to the executable of the DB Manager. It contains useful files for the DB Manager to work. These files can either be edited using a text editor or, for some files only, you can edit them safely using Settings/Edit experiences sub-menu. They contain:

- **broadman\_areas.txt**: links between the name of a broadman area and its respective index
- **corr\_nomenclature.txt**: links between the names of the same patient in the anatomical database (right) and in the functional database (left)
- **Downsamplings.txt**: list of possible values for the downsamplings
- **Experiences.txt**: list of experiences to find in the functional database
- **Frequencies.txt**: list of possible values for frequency ranges
- **localizers.tsv**: metadata for the localizers used to convert the database to BIDS
- **mars\_atlas\_index.csv**: links between the name of a mars atlas area and its respective index
- **TemporalSmoothings.txt**: list of possible values for the temporal smoothings
- **Years.txt**: list of possible values for the years
- **Images localizers**: this folder contains illustrations for the localizers (these images are exported into the HiBoP project created with the DB Manager)
- **Protocols**: this folder contains .prov files readable by HiBoP to describe the localizers (these protocol files are exported into the HiBoP project created with the DB Manager)

## B HiBoP Database Hierarchy

In order to use HiBoP and the DB Manager with automatic loading of patients, a specific hierarchy of the database must be respected. There is a specific hierarchy for the anatomical database (usable in both HiBoP and the Database Manager) and for the functional database (usable only in the Database Manager). These are presented in the following sections. Note that BIDS format will be supported in a near future.

## B.1 Anatomical database



Figure 35: Anatomical database hierarchy

On the figure 35:

- **<PATIENT>** corresponds to the patient ID, which is of the form **<PLACE>\_<DATE>\_<NAME>** (e.g. LYONNEURO\_2014\_THUv)
- **<PLACE>** is where the patient data comes from (e.g. LYONNEURO)
- **<DATE>** is the year the data has been acquired (e.g. 2014)
- **<NAME>** is the anonymized name of the patient (e.g. THUv)
- **<DATEFULL>** is the complete date (e.g. 2015-5-20)

## B.2 Functional database

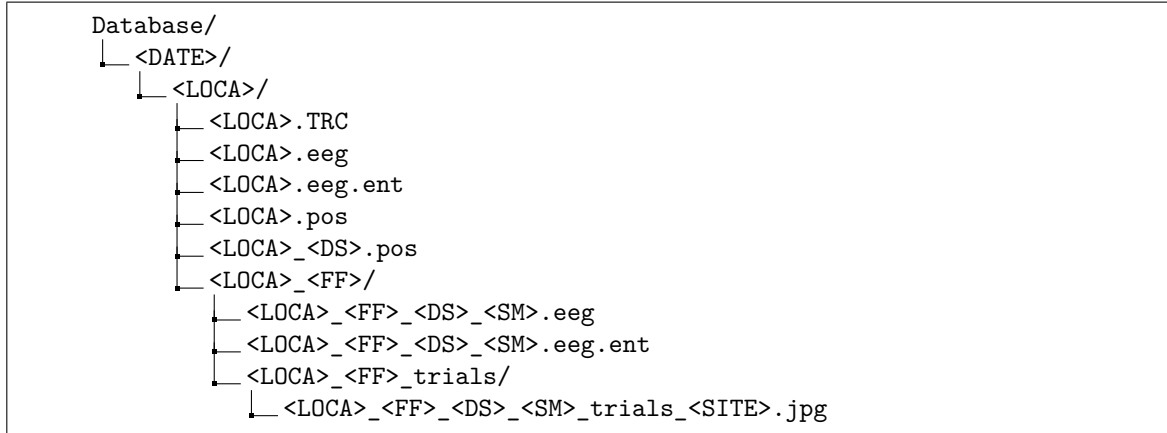


Figure 36: Functional database hierarchy

The figure 36 explains the hierarchy for the data we commonly use. It may not be adapted to every kinds of iEEG data. On this figure:

- **<LOCA>** is the name of the localizer which is usually of the form **<PATIENT>\_<EXP>** (e.g. LYONNEURO\_2014\_THUV\_LEC1). **<PATIENT>** is usually the same as in the anatomical database; if their names are different (case sensitive), you need to create a link in the corr\_nomenclature.txt file of the DB Manager if you wish to use this database with the DB Manager.
- **<PATIENT>** corresponds to the patient ID, which is of the form **<PLACE>\_<DATE>\_<NAME>** (e.g. LYONNEURO\_2014\_THUV)
- **<PLACE>** is where the patient data comes from (e.g. LYONNEURO)
- **<DATE>** is the year the data has been acquired (e.g. 2014)
- **<NAME>** is the anonymized name of the patient (e.g. THUV)
- **<FF>** is the frequency range of the processed data (e.g. f8f24 for alpha-beta or f50f150 for gamma)
- **<DS>** is the downsampling frequency of the processed data (e.g. ds8, ds16 or ds32)
- **<SM>** is the temporal smoothing of the processed data (e.g. sm0, sm250, sm500, sm1000, sm2500 or sm5000)
- **<SITE>** is the name of the channel (e.g. E5)