

# mTADA

- I. Introduction
- II. Requirements
- III. An example: joint analysis of DD and EE DNMs.
  - Load the source codes
  - Read the data and single-trait parameters
  - Set parameters for two traits.
  - Run mTADA
  - Get results
- Citation

This notebook describes steps used to jointly analyze two traits by mTADA.

## I. Introduction

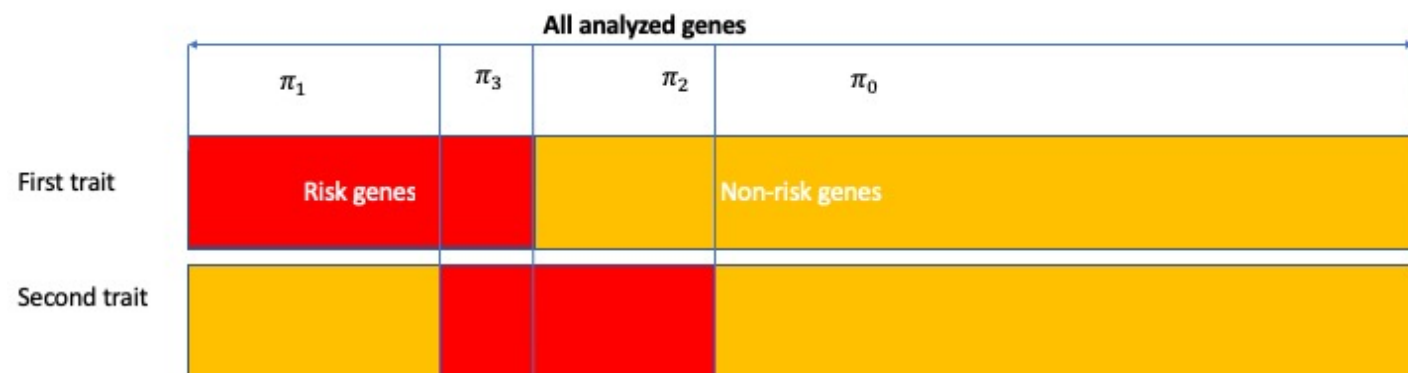
mTADA jointly analyze de novo mutations (DNMs) of two traits to 1) estimate the gene-level genetic overlap of the two traits; 2) report shared and specific risk genes; and 3) identify additional risk genes for each analyzed trait.

The method requires genetic parameters from single-trait analyses (the third and fourth columns in Table 1 below). Users can obtain single-trait parameters from extTADA/TADA methods.

**Table 1.** mTADA model for one variant category at the  $i^{th}$  gene.

Hypothesis	Proportion	First trait	Second trait
$H_0$	$\pi_0$	$x_{i1} \sim \text{Poisson}(2N_1\mu_i)$	$x_{i2} \sim \text{Poisson}(2N_2\mu_i)$
$H_1$	$\pi_1$	$x_{i1} \sim \text{Poisson}(2N_1\gamma_1\mu_i);$ $\gamma_1 \sim \text{Gamma}(\bar{\gamma}_1\beta_1, \beta_1)$	$x_{i2} \sim \text{Poisson}(2N_2\mu_i)$
$H_2$	$\pi_2$	$x_{i1} \sim \text{Poisson}(2N_1\mu_i)$	$x_{i2} \sim \text{Poisson}(2N_2\gamma_2\mu_i); \gamma_2 \sim$ $\text{Gamma}(\bar{\gamma}_2\beta_2, \beta_2)$
$H_3$	$\pi_3$	$x_{i1} \sim \text{Poisson}(2N_1\gamma_1\mu_i);$ $\gamma_1 \sim \text{Gamma}(\bar{\gamma}_1\beta_1, \beta_1)$	$x_{i2} \sim \text{Poisson}(2N_2\gamma_2\mu_i); \gamma_2 \sim$ $\text{Gamma}(\bar{\gamma}_2\beta_2, \beta_2)$

**Figure 1.** mTADA framework.



## Data for reproducible analyses

Data used in the main manuscript are inside the folder data (data):

1. FullDataSet\_DenovoMutations\_for\_mTADA.txt (data/FullDataSet\_DenovoMutations\_for\_mTADA.txt): all gene-level de novo mutations.
2. SingleTrait\_Parameters.txt (data/SingleTrait\_Parameters.txt): all single-trait parameters. We used extTADA to estimate these parameters.

*Note:* Users can re-run all these single-trait analyses by following an example here: <https://github.com/hoangtn/extTADA> (<https://github.com/hoangtn/extTADA>).

## II. Requirements

mTADA is written in R. Other R packages are required to run mTADA:

- rstan: <https://mc-stan.org/rstan/> (<https://mc-stan.org/rstan/>).
- locfit: <https://cran.r-project.org/web/packages/locfit/index.html> (<https://cran.r-project.org/web/packages/locfit/index.html>).

## III. An example: joint analysis of DD and EE DNMs.

Only one function mTADA (in the **Run mTADA** section) is used to obtain results. However, some additional steps are described here.

Software versions were used in the example below: R version 3.5.2, locfit version 1.5-9.1, and rstan version 2.18.2.

### Load the source codes

```
dataDir <- "../data/"
source("script/mTADA.R")
```

```
## locfit 1.5-9.1    2013-03-22
```

### Read the data and single-trait parameters

```
## De novo data
data <- read.table(paste0(dataDir, "FullDataSet_DenovoMutations_for_mTADA.txt"), header =
  = TRUE, as.is = TRUE)
## Single-trait parameters
sPar <- read.table(paste0(dataDir, "SingleTrait_Parameters.txt"), as.is = TRUE, header =
  TRUE)

trait1 = "DD"
trait2 = "EE"
##Take a quick look at the single-trait parameters of DD and EE
sPar[grep(trait1, sPar[, 1]), ] ##Trait 1
```

	Parameter <chr>	EstimatedValue <dbl>
8	DD_pi[1]	0.02936283
9	DD_hyperGammaMeanDN[1]	22.31762802
10	DD_hyperGammaMeanDN[2]	86.03966530
11	DD_hyperBetaDN[1]	0.82594514
12	DD_hyperBetaDN[2]	0.80689775
5 rows		

```
sPar[grep(trait2, sPar[, 1]), ] ##Trait 2
```

	Parameter <chr>	EstimatedValue <dbl>
18	EE_pi[1]	0.01548789
19	EE_hyperGammaMeanDN[1]	51.08181282
20	EE_hyperGammaMeanDN[2]	65.15189031
21	EE_hyperBetaDN[1]	0.80906448
22	EE_hyperBetaDN[2]	0.80774192
5 rows		

## Set parameters for two traits.

As described above, mTADA needs single-trait parameters:

- the number of trios:  $n_{trio}$ ;
- the mean and dispersion parameters of relative risks:  $\bar{\gamma}_j$  and  $\beta_j$  ( $j=1, 2$ );
- the proportion of risk genes:  $\pi_1^S$  and  $\pi_2^S$ .

**All these parameters are shown above.**

```

### Trait-1 INFORMATION
ntrio1 = 4293 #family numbers
p1 = 0.02936283 #The proportion of risk genes, this is p1S
meanGamma1 = c(22.31762802, 86.03966530) #Mean Gamma of two categories
beta1 = c(0.82594514, 0.80689775) #Beta values inside the distribution  $RR \sim \text{Gamma}(\text{meanRR} * \text{beta}, \text{beta})$ 
dataT1 <- data[, paste0(c("dn_damaging_", "dn_lof_"), trait1)] #De novo data
muDataT1 <- data[, c("mut_damaging", "mut_lof")] #Mutation data of the first trait
#####
### Trait-2 INFORMATION
ntrio2 = 356
p2 = 0.01548789 #This is p2S
meanGamma2 = c(51.08181282, 65.15189031)
beta2 = c(0.80906448, 0.80774192)
dataT2 <- data[, paste0(c("dn_damaging_", "dn_lof_"), trait2)]
muDataT2 <- muDataT1

```

## Run mTADA

In this example, we only use a small number of iterations and two MCMC chains. However, users can change these parameters to obtain more reliable results.

```

nIteration = 2000 #This should be higher to obtain better results.
nChain = 2 #The number of MCMC chains

#####MAIN ANALYSIS
mTADAresults <- mTADA(geneName = data[, 1],
  #####Trait-1 information
  ntrio1 = ntrio1, # Trio number of Trait 1
  p1 = p1, #Risk-gene proportion of Trait 1
  dataDN1 = data.frame(dataT1), #De novo data of Trait 1
  mutRate1 = data.frame(muDataT1), # Mutation rates of Trait 1
  hyperGammaMeanDN1 = c(meanGamma1), # Mean relative risks of Trait 1
  hyperBetaDN01 = beta1, #NULL, #array(c(1, 1)),
  #####Trait-2 information
  ntrio2 = ntrio2, # Trio number of Trait 2
  p2 = p2, #Risk-gene proportion of Trait 2
  dataDN2 = data.frame(dataT2), # De novo data of Trait 2
  mutRate2 = data.frame(muDataT2), # Mutation rates of Trait 2
  hyperGammaMeanDN2 = c(meanGamma2), # Mean relative risks of Trait 2
  hyperBetaDN02 = beta2, #NULL, #array(c(1, 1)),
  #####Other parameters
  nIteration = nIteration,
  useMCMC = TRUE, #If FALSE, it will use the 'Variational Bayes' approach.
  nChain = nChain
)

```

```

## No information for core numbers (nCore); therefore, nCore = nChain: 2 core(s) is/are used

```

```
## Loading required package: ggplot2
```

```
## Loading required package: StanHeaders
```

```
## rstan (Version 2.18.2, GitRev: 2e1f913d3ca3)
```

```
## For execution on a local, multicore CPU with excess RAM we recommend calling  
## options(mc.cores = parallel::detectCores()).  
## To avoid recompilation of unchanged Stan programs, we recommend calling  
## rstan_options(auto_write = TRUE)
```

```
## =====  
## Building the model  
## =====
```

```
##  
## =====Use MCMC=====
```

```
## recompiling to avoid crashing R session
```

```
## ====  
## Only pi, alpha and hyper parameters are estimated in this step  
## The method does not calculate HPDs for hyper betas, just their medians  
## ==
```

## Get results

mTADA's output includes:

1. data: main gene-level results (posterior probabilities for the four models as described in the main manuscript: PP0, PP1, PP2 and PP3).
2. probModel: a vector of  $\pi_j$ , ( $j = 0..3$ ) in Table 1.
3. pars: the estimated value and credible interval of  $\pi_3$  (described as p12 in the our code).
4. mcmcData: MCMC sampling results for  $\pi_3$ .

The most important information is from data. **Users can use this information to obtain top prioritized genes for downstream analyses (e.g., top shared/specific genes, top genes for each trait).** However, we will also take a quick look at all these information.

### Results for downstream analyses (gene-level posteior probabilities of four models)

```
fData <- mTADAResults$data ## Full analysis results of the two-trait analysis.  
head(fData)
```

geneN...	dn_damaging_...	dn_lof_DD	dn_damaging_EE	dn_lof_EE	NO	BOTH
<fctr>	<int>	<int>	<int>	<int>	<dbl>	<dbl>
1 A1BG	0	0	0	0	0.9782988	0.0027703687
2 A1BG-AS1	0	0	0	0	0.9645411	0.0058484751
3 A1CF	0	0	0	0	0.9892214	0.0006241505
4 A2M	0	0	1	0	0.7675001	0.0023974666
5 A2M-AS1	0	0	0	0	0.9635476	0.0060760422
6 A2ML1	0	0	0	0	0.9918274	0.0002016731

6 rows | 1-9 of 10 columns

### Genes with PP3 > 0.8 (Posterior probabilities of Model 3)

```
fData[fData$BOTH > 0.8, ]
```

geneN...	dn_damaging_...	dn_lof_DD	dn_damaging_EE	dn_lof_EE	NO	BOT
<fctr>	<int>	<int>	<int>	<int>	<dbl>	<db
2348 CACNA1A	5	0	2	0	3.186042e-04	0.993066
3201 CHD2	0	6	0	1	4.709061e-10	0.934235
6254 GABBR2	2	0	2	0	2.568316e-03	0.949980
6265 GABRB3	2	0	2	0	9.424303e-04	0.978920
6610 GNAO1	4	1	2	0	1.637585e-08	0.998330
7165 HECW2	5	1	1	0	1.970360e-06	0.887266
7426 HNRNPU	0	7	0	1	8.952701e-13	0.933556
8283 KCNQ2	9	0	2	0	3.319964e-13	0.998132
8284 KCNQ3	3	0	1	0	4.396331e-03	0.909428
10146 MLL	1	26	1	0	1.507451e-48	0.862002

1-10 of 14 rows | 1-8 of 10 columns

Previous **1** 2 Next

### Genes with PP1 > 0.8 (Posterior probabilities of Model 1)

```
fData[fData$FIRST > 0.8, ]
```

geneN...	dn_damaging_...	dn_lof_DD	dn_damaging_EE	dn_lof_EE	NO	BC
<fctr>	<int>	<int>	<int>	<int>	<dbl>	<c
347 ADNP	1	19	0	0	4.074985e-37	0.192735
681 ANKRD11	0	32	0	0	2.286217e-60	0.132585

	geneN...	dn_damaging_...	dn_lof_DD	dn_damaging_EE	dn_lof_EE	NO	BO
	<fctr>	<int>	<int>	<int>	<int>	<dbl>	<c
1000	ARID1A	1	2	0	0	8.434492e-02	0.111468
1001	ARID1B	0	30	0	0	1.696572e-56	0.139999
1002	ARID2	0	3	0	0	2.186807e-03	0.167429
1153	ASXL1	0	4	0	0	1.839357e-05	0.165438
1155	ASXL3	0	14	0	0	2.517375e-25	0.192215
1317	AUTS2	0	4	0	0	1.130829e-05	0.177120
1450	BCL11A	2	3	0	0	7.439678e-07	0.183144
1630	BRPF1	0	4	0	0	7.080246e-05	0.126899

1-10 of 78 rows | 1-8 of 10 columns

Previous 1 2 3 4 5 6 ... 8 Next

## Genes with PP2 > 0.8 (Posterior probabilities of Model 2)

```
fData[fData$SECOND > 0.8, ]
```

	geneN...	dn_damaging_...	dn_lof_DD	dn_damaging_EE	dn_lof_EE	NO	BOT
	<fctr>	<int>	<int>	<int>	<int>	<dbl>	<dbl>
14671	SCN1A	2	0	4	4	1.998619e-12	0.114279

1 row | 1-8 of 10 columns

## Use mTADA's results for single-trait analyses.

We can obtain single-trait results by summing PP1 and PP3 (Trait 1) or PP2 and PP3 (Trait 2).

### Trait 1

```
fData[, 'pTrait1'] <- fData[, 'BOTH'] + fData[, 'FIRST']
fData1 <- fData[fData$pTrait1 > 0.8, ]
head(fData1[, c(1:5, 10)])
```

	geneNa...	dn_damaging_DD	dn_lof_DD	dn_damaging_EE	dn_lof_EE	pTrait1
	<fctr>	<int>	<int>	<int>	<int>	<dbl>
347	ADNP	1	19	0	0	1.0000000
447	AHDC1	0	8	0	0	1.0000000
681	ANKRD11	0	32	0	0	1.0000000
1000	ARID1A	1	2	0	0	0.9152718
1001	ARID1B	0	30	0	0	1.0000000
1002	ARID2	0	3	0	0	0.9977987

6 rows

## Trait 2

```
fData[, 'pTrait2'] <- fData[, 'BOTH'] + fData[, 'SECOND']
fData2 <- fData[fData$pTrait2 > 0.8, ]
head(fData2[, c(1:5, 11)])
```

	geneNa... <fctr>	dn_damaging_DD <int>	dn_lof_DD <int>	dn_damaging_EE <int>	dn_lof_EE <int>	pTrait2 <dbl>
2348	CACNA1A	5	0	2	0	0.9956184
3201	CHD2	0	6	0	1	0.9342339
6254	GABBR2	2	0	2	0	0.9956821
6265	GABRB3	2	0	2	0	0.9974236
6610	GNAO1	4	1	2	0	0.9983307
7165	HECW2	5	1	1	0	0.8872665

6 rows

## Other information

Some additional information can be obtained from mTADA's results.

```
pCI <- mTADAResults$pars ## Genetic parameters
piValue <- mTADAResults$probModel ## Posterior probabilities of genes for four models
mcmcResult <- mTADAResults$mcmcData ##MCMC results
```

## The proportions of risk genes

*piValue* is a vector of  $\pi$  values. In the result below, pNO, pFIRST, pSECOND, and pBOTH are  $\pi_0$ ,  $\pi_1$ ,  $\pi_2$  and  $\pi_3$  respectively in **Table 1**.

piValue

```
##           pNO           pFIRST           pSECOND           pBOTH
## 0.961659367 0.022852743 0.008977803 0.006510087
```

## Estimated information of $\pi_3$ .

Credible-interval information is from *pCI*.

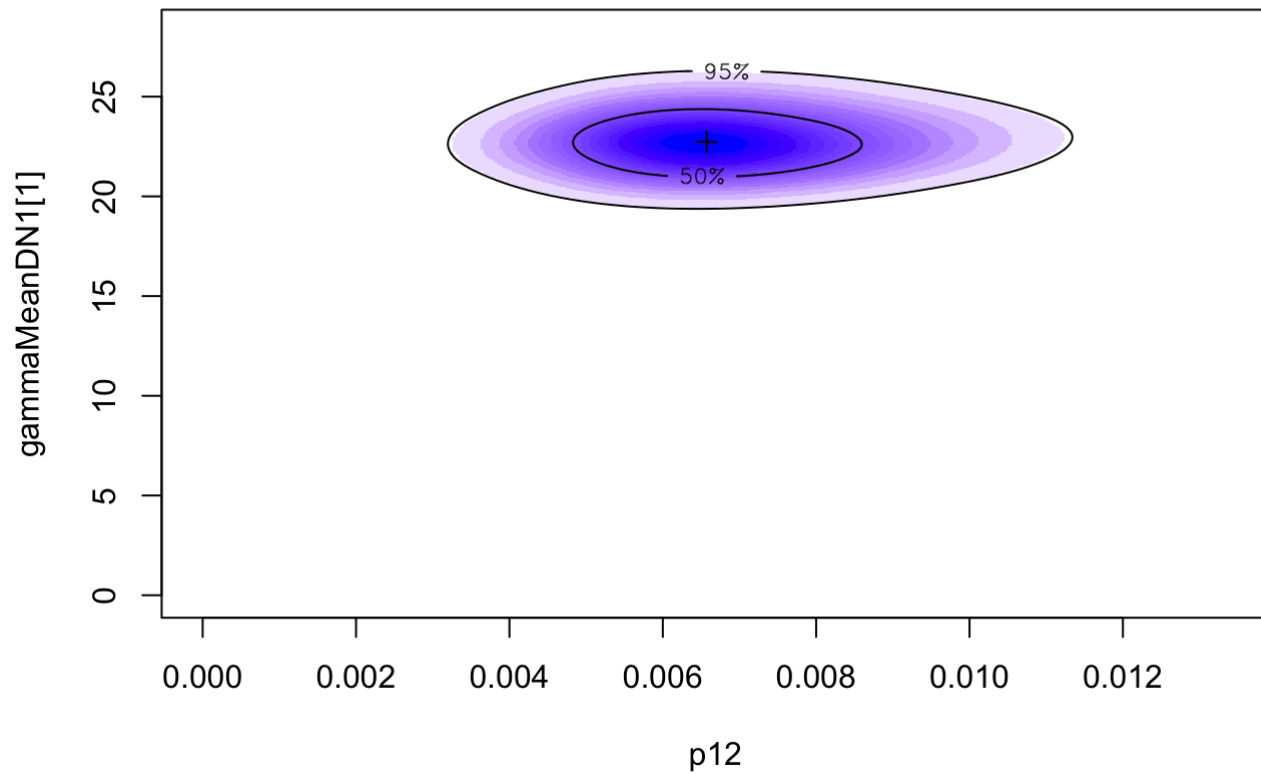
```
pCI ## Mode: estimated values; CI: credible interval with low (l) and upper (u) values
```

```
##           Mode           lCI           uCI
## p12           0.006510087 0.003824866 0.01023632
## gammaMeanDN1[1] 22.750038264 20.051680175 25.50273351
```

To check the convergent information of  $\pi_3$ , we can visualize MCMC results.



```
## p12 is pi3 in the model
plotParHeatmap1(mcmcResult = mcmcResult, pars = c('p12', 'gammaMeanDN1[1]'))
```



## Citation

**mTADA: a framework for identifying risk genes from de novo mutations in multiple traits.** Hoang T. Nguyen, Amanda Dobbyn, Ruth C. Brown, Brien P. Riley, Joseph Buxbaum, Dalila Pinto, Shaun M Purcell, Patrick F Sullivan<sup>8</sup>, Xin He, Eli A. Stahl.