

The doBy package – yet another utility package

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Abstract The doBy is one of several general utility packages on CRAN. We illustrate two main features of the package: The ability to making groupwise computations and the ability to compute linear estimates, contrasts and least-squares means.

Introduction

The package originally grew out of a need to calculate groupwise summary statistics (much in the spirit of PROC SUMMARY of the SAS system, (SAS Institute Inc., 2020)). The name doBy comes from the need to **do** some computations on data which is stratified **By** the value of some variables. The doBy package (Højsgaard and Halekoh, 2020) appeared on CRAN in 2006. Today the package contains many additional utilities.

When it comes to data handling, doBy is nowhere nearly as powerful as more contemporary packages, such as those in the tidyverse eco system, (Wickham et al., 2019). On the other hand, doBy is based on classical data structures that are unlikely to undergo sudden changes. Moreover, it can be hypothesized that the data handling functions in doBy remain appealing to a group of users because of their simplicity in use.

In this paper we focus 1) on the “doing by” functions and 2) on functions related to linear estimates and contrasts.

Functions related to groupwise computations

A working dataset - the C02 data

The C02 data frame comes from an experiment on the cold tolerance of the grass species *Echinochloa crus-galli*. To limit the amount of output we modify names and levels of variables as follows

```
data(C02)
C02 <- within(C02, {
  Treat <- Treatment
  Treatment <- NULL
  levels(Treat) <- c("nchil", "chil")
  levels(Type) <- c("Que", "Mis")
})
C02 <- subset(C02, Plant %in% c("Qn1", "Qc1", "Mn1", "Mc1"))
dim(C02)

#> [1] 28 5

head(C02, 4)

#>   Plant Type conc uptake Treat
#> 1  Qn1  Que   95   16.0 nchil
#> 2  Qn1  Que  175   30.4 nchil
#> 3  Qn1  Que  250   34.8 nchil
#> 4  Qn1  Que  350   37.2 nchil
```

The summaryBy function

The summaryBy function is used for calculating quantities like *the mean and variance of numerical variables x and y for each combination of two factors A and B*\$. Notice: A functionality similar to summaryBy is provided by aggregate from base R, but summaryBy offers additional features.

```
myfun1 <- function(x){c(m=mean(x), s=sd(x))}
summaryBy(cbind(conc, uptake, lu=log(uptake)) ~ Plant, data=C02, FUN=myfun1)

#>   Plant conc.m conc.s uptake.m uptake.s lu.m lu.s
#> 1  Qn1    435  317.7    33.23    8.215 3.467 0.3189
```

```
#> 2   Qc1   435  317.7   29.97   8.335 3.356 0.3446
#> 3   Mn1   435  317.7   26.40   8.694 3.209 0.4234
#> 4   Mc1   435  317.7   18.00   4.119 2.864 0.2622

## same as
## aggregate(cbind(conc, uptake, log(uptake)) ~ Plant, data=C02, FUN=myfun1)
```

The convention is that variables that do not appear in the dataframe (e.g. `log(uptake)`) must be named (here as `lu`). Various shortcuts are available, e.g. the following, where left hand side dot refers to “all numeric variables” while the right hand side dot refers to “all factor variables”. Writing 1 on the right hand side leads to computing over the entire dataset:

```
summaryBy(. ~ ., data=C02, FUN=myfun1)

#>   Plant Type Treat conc.m conc.s uptake.m uptake.s
#> 1   Qn1  Que nchil   435  317.7   33.23   8.215
#> 2   Qc1  Que  chil   435  317.7   29.97   8.335
#> 3   Mn1 Mis nchil   435  317.7   26.40   8.694
#> 4   Mc1 Mis  chil   435  317.7   18.00   4.119

summaryBy(. ~ 1, data=C02, FUN=myfun1)

#>   conc.m conc.s uptake.m uptake.s
#> 1   435  299.6   26.9   9.189
```

Formulas and lists

The convention for the “By”-functions is that a two sided formula like can be written in two ways:

```
cbind(x, y) ~ A + B
list(c("x", "y"), c("A", "B"))
```

Some “By”-functions only take a right hand sided formula as input. Such a formula can also be written in two ways:

```
~ A + B
c("A", "B")
```

The list-form / vector-form is especially useful if a function is invoked programmatically. Hence the calls to `summaryBy` above can also be made as

```
summaryBy(list(c("conc", "uptake", "lu=log(uptake)"), "Plant"), data=C02, FUN=myfun1)
summaryBy(list(c("."), c(".")), data=C02, FUN=myfun1)
summaryBy(list(c("."), c("1")), data=C02, FUN=myfun1)
```

The orderBy function

Ordering (or sorting) a data frame is possible with the `orderBy` function. Suppose we want to order the rows of the C02 data by increasing values of `conc` and decreasing value of `uptake` (within `conc`):

```
x1 <- orderBy(~ conc - uptake, data=C02)
head(x1)

#>   Plant Type conc uptake Treat
#> 1   Qn1  Que  95  16.0 nchil
#> 22  Qc1  Que  95  14.2  chil
#> 43  Mn1 Mis  95  10.6 nchil
#> 64  Mc1 Mis  95  10.5  chil
#> 2   Qn1  Que 175  30.4 nchil
#> 23  Qc1  Que 175  24.1  chil
```

Following the remarks about specification in “By”-functions, an equivalent form is:

```
orderBy(c("conc", "-uptake"), data=C02)
```

The splitBy function

Suppose we want to split C02 into a list of dataframes:

```
x1 <- splitBy(~ Plant + Type, data=C02)
x1

#>   listentry Plant Type
#> 1   Qn1|Que   Qn1  Que
#> 2   Qc1|Que   Qc1  Que
#> 3   Mn1|Mis   Mn1  Mis
#> 4   Mc1|Mis   Mc1  Mis
```

The result is a list (with a few additional attributes):

```
lapply(x1, head, 2)

#> $`Qn1|Que`
#>   Plant Type conc uptake Treat
#> 1   Qn1  Que   95   16.0 nchil
#> 2   Qn1  Que  175   30.4 nchil
#>
#> $`Qc1|Que`
#>   Plant Type conc uptake Treat
#> 22  Qc1  Que   95   14.2  chil
#> 23  Qc1  Que  175   24.1  chil
#>
#> $`Mn1|Mis`
#>   Plant Type conc uptake Treat
#> 43  Mn1  Mis   95   10.6 nchil
#> 44  Mn1  Mis  175   19.2 nchil
#>
#> $`Mc1|Mis`
#>   Plant Type conc uptake Treat
#> 64  Mc1  Mis   95   10.5  chil
#> 65  Mc1  Mis  175   14.9  chil
```

The subsetBy function

Suppose we want to select those rows within each treatment for which the uptake is larger than 75% quantile of uptake (within the treatment). This is achieved by:

```
x2 <- subsetBy(~ Treat, subset=uptake > quantile(uptake, prob=0.75), data=C02)
head(x2, 4)

#>           Plant Type conc uptake Treat
#> nchil.4   Qn1  Que  350   37.2 nchil
#> nchil.6   Qn1  Que  675   39.2 nchil
#> nchil.7   Qn1  Que 1000   39.7 nchil
#> nchil.49  Mn1  Mis 1000   35.5 nchil
```

The transformBy function

The transformBy function is analogous to the transform function except that it works within groups. For example:

```
x3 <- transformBy(~ Treat, data=C02,
                  minU=min(uptake), maxU=max(uptake),
                  range=diff(range(uptake)))
head(x3, 4)

#>           Plant Type conc uptake Treat minU maxU range
#> nchil.1   Qn1  Que   95   16.0 nchil 10.6 39.7  29.1
#> nchil.2   Qn1  Que  175   30.4 nchil 10.6 39.7  29.1
#> nchil.3   Qn1  Que  250   34.8 nchil 10.6 39.7  29.1
#> nchil.4   Qn1  Que  350   37.2 nchil 10.6 39.7  29.1
```

The lmBy function

The `lmBy` function allows for fitting linear models to different strata of data (the vertical bar is used for defining groupings of data):

```
m <- lmBy(uptake ~ conc | Treat, data=C02)
coef(m)

#>      (Intercept)      conc
#> nchil      20.82 0.02067
#> chil       17.02 0.01602
```

The result is a list with a few additional attributes and the list can be processed further as e.g.

```
lapply(m, function(z) coef(summary(z)))

#> $nchil
#>      Estimate Std. Error t value Pr(>|t|)
#> (Intercept) 20.82342    3.092430   6.734 2.092e-05
#> conc        0.02067    0.005889   3.510 4.304e-03
#>
#> $chil
#>      Estimate Std. Error t value Pr(>|t|)
#> (Intercept) 17.01814    3.668315   4.639 0.0005709
#> conc        0.01602    0.006986   2.293 0.0407168
```

Functions related linear estimates and contrasts

A linear function of a p -dimensional parameter vector β has the form

$$C = L\beta$$

where L is a $q \times p$ matrix which we call the Linear Estimate Matrix or simply LE-matrix. The corresponding linear estimate is $\hat{C} = L\hat{\beta}$. A linear hypothesis has the form $H_0 : L\beta = m$ for some q dimensional vector m .

A working dataset - the ToothGrowth data

The response is the length of odontoblasts cells (cells responsible for tooth growth) in 60 guinea pigs. Each animal received one of three dose levels of vitamin C (0.5, 1, and 2 mg/day) by one of two delivery methods, (orange juice (coded as OJ) or ascorbic acid (a form of vitamin C and (coded as VC)). The dataset is balanced with 10 measurements for each combination of dose and supp. To illustrate certain points in what follows we make data unbalanced by removing some rows of the dataframe:

```
data("ToothGrowth")
ToothGrowth <- transform(ToothGrowth, dose = factor(dose))
ToothGrowth <- ToothGrowth[-(1:3), ]
head(ToothGrowth, 4)

#>   len supp dose
#> 4  5.8   VC  0.5
#> 5  6.4   VC  0.5
#> 6 10.0   VC  0.5
#> 7 11.2   VC  0.5
```

The interaction plot indicates some interaction between dose and supp. This is also supported by a formal test:

```
tooth1 <- lm(len ~ dose + supp, data=ToothGrowth)
tooth2 <- lm(len ~ dose * supp, data=ToothGrowth)
anova(tooth1, tooth2)

#> Analysis of Variance Table
#>
#> Model 1: len ~ dose + supp
#> Model 2: len ~ dose * supp
```

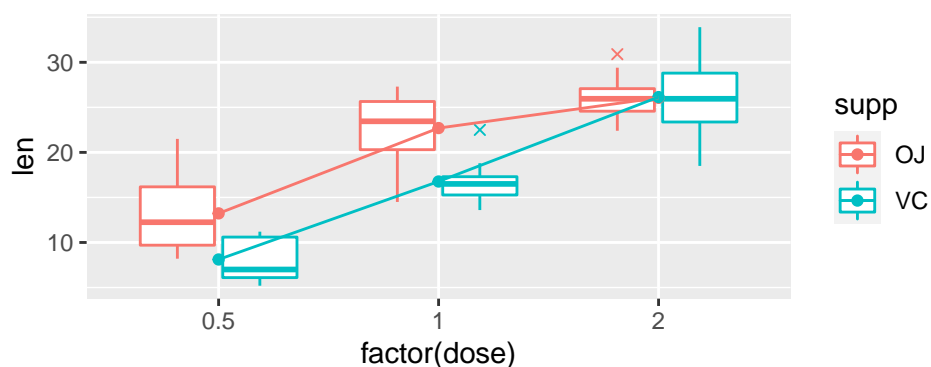


Figure 1: Interaction plot for the ToothGrowth data. The average ‘len’ for each group is a dot. Boxplot outliers are crosses.

```
#>   Res.Df RSS Df Sum of Sq    F Pr(>F)
#> 1     53 789
#> 2     51 685   2      104 3.88 0.027
```

Computing linear estimates

For now, we focus on the additive model. Consider computing the estimated length for each dose of orange juice (OJ): One option: Construct the LE-matrix L directly and then invoke `linest`:

```
L <- matrix(c(1, 0, 0, 0,
              1, 1, 0, 0,
              1, 0, 1, 0), nrow=3, byrow=T)
```

The matrix L can be generated as follows:

```
L <- LE_matrix(tooth1, effect="dose", at=list(supp="OJ"))
```

The estimates can be computed directly as

```
L %*% coef(tooth1)
```

but we do not obtain standard errors etc. this way. Instead we can invoke `linest`

```
c1 <- linest(tooth1, L)
coef(c1)
```

```
#>   estimate std.error statistic df    p.value
#> 1    12.59    1.027    12.26 53 4.239e-17
#> 2    21.52    1.004    21.43 53 8.969e-28
#> 3    27.88    1.004    27.77 53 2.958e-33
```

```
confint(c1)
```

```
#>   0.025 0.975
#> 1 10.53 14.65
#> 2 19.50 23.53
#> 3 25.87 29.90
```

The function `esticon` has been part of `doBy` for many years while `linest` is a newer addition. The functionality, however, is similar:

```
c1 <- esticon(tooth1, L)
c1
```

```
#>      estimate std.error statistic p.value beta0 df
#> [1,]    12.59    1.03    12.26  0.00  0.00 53
#> [2,]    21.52    1.00    21.43  0.00  0.00 53
#> [3,]    27.88    1.00    27.77  0.00  0.00 53
```

Least-squares means (LS-means)

A related question could be: What is the estimated length for each dose if we ignore the source of vitamin C (i.e. whether it is OJ or VC). One approach would be to fit a model in which source does not appear:

```
tooth0 <- update(tooth1, . ~ . - supp)
L0 <- LE_matrix(tooth0, effect="dose")
L0

#>      (Intercept) dose1 dose2
#> [1,]           1     0     0
#> [2,]           1     1     0
#> [3,]           1     0     1

linest(tooth0, L=L0)

#> Coefficients:
#>      estimate std.error statistic    df p.value
#> [1,]   11.124    1.027   10.830 54.000      0
#> [2,]   19.735    0.947   20.841 54.000      0
#> [3,]   26.100    0.947   27.562 54.000      0
```

An alternative would be to stick to the original model but compute the estimate for an “average vitamin C source”. That would correspond to giving weight 1/2 to each of the two vitamin C source parameters. However, as one of the parameters is already set to zero to obtain identifiability, we obtain the LE-matrix L as

```
L1 <- matrix(c(1, 0, 0, 0.5,
               1, 1, 0, 0.5,
               1, 0, 1, 0.5), nrow=3, byrow=T)
linest(tooth1, L=L1)

#> Coefficients:
#>      estimate std.error statistic    df p.value
#> [1,]   10.809    0.940   11.496 53.000      0
#> [2,]   19.735    0.863   22.873 53.000      0
#> [3,]   26.100    0.863   30.249 53.000      0
```

Such a particular linear estimate is sometimes called a *least-squares mean*, an *LSmean*, a *marginal mean* or a *population mean*. Notice: One may generate L automatically with

```
L1 <- LE_matrix(tooth1, effect="dose")
L1
```

```
#>      (Intercept) dose1 dose2 suppVC
#> [1,]           1     0     0     0.5
#> [2,]           1     1     0     0.5
#> [3,]           1     0     1     0.5
```

Notice: One may obtain the LSmean directly as:

```
LSmeans(tooth1, effect="dose")

#> Coefficients:
#>      estimate std.error statistic    df p.value
#> [1,]   10.809    0.940   11.496 53.000      0
#> [2,]   19.735    0.863   22.873 53.000      0
#> [3,]   26.100    0.863   30.249 53.000      0
```

which is the same as

```
L <- LE_matrix(tooth1, effect="dose")
linest(tooth1, L=L)
```

Interaction model

For a model with interactions, the LSmeans are

```
LSmeans(tooth2, effect="dose")
```

```
#> Coefficients:
#>      estimate std.error statistic    df p.value
#> [1,]   10.672    0.903   11.819 51.000      0
#> [2,]   19.735    0.819   24.085 51.000      0
#> [3,]   26.100    0.819   31.853 51.000      0
```

In this case, the LE-matrix is

```
L <- LE_matrix(tooth2, effect="dose")
L
#>      (Intercept) dose1 dose2 suppVC dose1:suppVC dose2:suppVC
#> [1,]           1     0     0     0.5           0.0           0.0
#> [2,]           1     1     0     0.5           0.5           0.0
#> [3,]           1     0     1     0.5           0.0           0.5
```

Using (transformed) covariates

Below, the covariate conc is fixed at the average value:

```
co2.lm1 <- lm(uptake ~ conc + Type + Treat, data=C02)
LSmeans(co2.lm1, effect="Treat")
```

```
#> Coefficients:
#>      estimate std.error statistic    df p.value
#> [1,]    29.81     1.35    22.16 24.00      0
#> [2,]    23.99     1.35    17.83 24.00      0
```

If we use $\log(\text{conc})$ instead we will get an error when calculating LS-means because $\log(\text{conc})$ is not a variable in the dataframe. Instead one can do:

```
co2.lm2 <- lm(uptake ~ log.conc + Type + Treat,
              data=transform(C02, log.conc=log(conc)))
LSmeans(co2.lm2, effect="Treat")
```

```
#> Coefficients:
#>      estimate std.error statistic    df p.value
#> [1,]    29.814     0.934    31.938 24.000      0
#> [2,]    23.986     0.934    25.694 24.000      0
```

This also highlights what is computed: The average of the log of conc; not the log of the average of conc. In a similar spirit consider

```
co2.lm3 <- lm(uptake ~ conc + I(conc^2) + Type + Treat, data=C02)
LSmeans(co2.lm3, effect="Treat")
```

```
#> Coefficients:
#>      estimate std.error statistic    df p.value
#> [1,]    33.24     1.29    25.81 23.00      0
#> [2,]    27.41     1.29    21.29 23.00      0
```

Above $I(\text{conc}^2)$ is the average of the squared values of conc; not the square of the average of conc, cfr. the following.

```
co2.lm4 <- lm(uptake ~ conc + conc2 + Type + Treat, data=
              transform(C02, conc2=conc^2))
LSmeans(co2.lm4, effect="Treat")
```

```
#> Coefficients:
#>      estimate std.error statistic    df p.value
#> [1,]    29.81     1.02    29.27 23.00      0
#> [2,]    23.99     1.02    23.55 23.00      0
```

If we want to evaluate the LS-means at `conc=10` then we can do:

```
LSmeans(co2.lm4, effect="Treat", at=list(conc=10, conc2=100))
```

```
#> Coefficients:
#>      estimate std.error statistic    df p.value
#> [1,]    14.67     2.23      6.58 23.00      0
#> [2,]     8.84     2.23      3.96 23.00      0
```

Alternative models

Linear mixed effects model

For the sake of illustration we treat `supp` as a random effect:

```
library(lme4)
tooth.mix <- lmer(len ~ dose + (1|supp), data=ToothGrowth)
LSmeans(tooth.mix, effect="dose")

#> Coefficients:
#>      estimate std.error statistic    df p.value
#> [1,]    10.84     1.95      5.56  1.43  0.06
#> [2,]    19.73     1.91     10.32  1.33  0.03
#> [3,]    26.10     1.91     13.65  1.33  0.02
```

Notice here that the parameter estimates themselves are similar to those of a linear model (had data been completely balanced, the estimates would have been identical). However, the standard errors of the the estimates are much larger under the mixed model. This comes from that `supp` is treated as a random effect. Notice that the degrees of freedom by default are adjusted using a Kenward–Roger approximation (provided that `pbkrtest` package ([Halekoh and Højsgaard, 2014](#)) is installed). Adjustment of degrees of freedom is controlled with the `adjust.df` argument.

Generalized linear models and generalized estimating equations

We can calculate LS-means for e.g. a Poisson or a gamma model. Notice that the LS-means are on the scale of the linear predictor - not on the scale of the response.

```
tooth.gam <- glm(len ~ dose + supp, family=Gamma, data=ToothGrowth)
LSmeans(tooth.gam, effect="dose")

#> Coefficients:
#>      estimate std.error statistic p.value
#> [1,]  0.09086   0.00567   16.02215      0
#> [2,]  0.05104   0.00294   17.33506      0
#> [3,]  0.03880   0.00224   17.30448      0
```

Similarly, for gee-type models we get:

```
library(geepack)
tooth.gee <- geeglm(len ~ dose, id=supp, family=Gamma, data=ToothGrowth)
LSmeans(tooth.gee, effect="dose")

#> Coefficients:
#>      estimate std.error statistic p.value
#> [1,] 8.99e-02  1.42e-02  6.35e+00      0
#> [2,] 5.07e-02  5.38e-03  9.41e+00      0
#> [3,] 3.83e-02  4.15e-05  9.23e+02      0
```

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Bibliography

- U. Halekoh and S. Højsgaard. A kenward-roger approximation and parametric bootstrap methods for tests in linear mixed models – the R package pbkrtest. *Journal of Statistical Software*, 59(9):1–30, 2014. URL <http://www.jstatsoft.org/v59/i09/>. [p8]
- S. Højsgaard and U. Halekoh. *doBy: Groupwise Statistics, LSmeans, Linear Contrasts, Utilities*, 2020. URL <http://people.math.aau.dk/~sorenh/software/doBy/>. R package version 4.6.6. [p1]
- SAS Institute Inc. *Base SAS 9.4 Procedures Guide, Seventh Edition*, April 2020. [p1]
- H. Wickham, M. Averick, J. Bryan, W. Chang, L. D. McGowan, R. François, G. Golemund, A. Hayes, L. Henry, J. Hester, M. Kuhn, T. L. Pedersen, E. Miller, S. M. Bache, K. Müller, J. Ooms, D. Robinson, D. P. Seidel, V. Spinu, K. Takahashi, D. Vaughan, C. Wilke, K. Woo, and H. Yutani. Welcome to the tidyverse. *Journal of Open Source Software*, 4(43):1686, 2019. doi: 10.21105/joss.01686. [p1]

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