The doBy package – yet another utility package

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Abstract The doBy is one of several general utility packages on CRAN. We illustrate two main features of the package: The ability to making groupwise computations and the ability to compute linear estimates, contrasts and least-squares means.

Introduction

The doBy package (which first appeared on CRAN in 2006) grew out of a need to calculate groupwise summary statistics (much in the spirit of PROC SUMMARY of the SAS system, (SAS Institute Inc., 2020)). The name doBy comes from the need to **do** some computations on data which is stratified **By** the value of some variables. Today the package contains many additional utilities.

In this paper we focus 1) on the "doing by" functions and 2) on functions related to linear estimates and contrasts.

Related functionaly

When it comes to data handling, doBy is nowhere nearly as powerful as more contemporary packages, such as those in the tidyverse eco system, (Wickham et al., 2019). The aggregate function in base R provides functionality similar to doBys summaryBy function. Another package to be mentioned in this connection is data.table, Dowle and Srinivasan (2019). On the other hand, doBy is based on classical data structures that are unlikely to undergo sudden changes. Moreover, it can be hypothesized that the data handling functions in doBy remain appealing to a group of users because of their simplicity.

Functions related to groupwise computations

A working dataset - the CO2 data

The CO2 data frame comes from an experiment on the cold tolerance of the grass species *Echinochloa crus-galli*. To limit the amount of output we modify names and levels of variables as follows

```
data(CO2)
CO2 <- within(CO2, {
   Treat = Treatment; Treatment = NULL
   levels(Treat) = c("nchil", "chil"); levels(Type) = c("Que", "Mis")
CO2 <- subset(CO2, Plant %in% c("Qn1", "Qc1", "Mn1", "Mc1"))
dim(CO2)
#> [1] 28 5
head(CO2, 4)
#>
    Plant Type conc uptake Treat
      Qn1 Que 95 16.0 nchil
      Qn1 Que 175
                     30.4 nchil
                250
                      34.8 nchil
      Qn1
           Que
      Qn1 Que 350
                     37.2 nchil
```

The summaryBy function

The summaryBy function is used for calculating quantities like *the mean and variance of numerical variables* x *and* y *for each combination of two factors* A *and* B\$. Notice: A functionality similar to summaryBy is provided by aggregate from base R, but summaryBy offers additional features.

```
myfun1 \leftarrow function(x)\{c(m=mean(x), s=sd(x))\}
summaryBy(cbind(conc, uptake, lu=log(uptake)) ~ Plant, data=CO2, FUN=myfun1)
```

```
#>
    Plant conc.m conc.s uptake.m uptake.s lu.m
           435 317.7 33.23
#> 1
     Qn1
                                8.215 3.467 0.3189
                      29.97
            435 317.7
                                8.335 3.356 0.3446
#> 2
     Qc1
#> 3
           435 317.7 26.40 8.694 3.209 0.4234
     Mn1
#> 4
     Mc1
           435 317.7 18.00
                                4.119 2.864 0.2622
```

The convention is that variables that do not appear in the dataframe (e.g. log(uptake)) must be named (here as lu). Various shportcuts are available, e.g. the following, where left hand side dot refers to "all numeric variables" while the right hand side dot refers to "all factor variables". Writing 1 on the right hand side leads to computing over the entire dataset:

```
summaryBy(. ~ ., data=CO2, FUN=myfun1)
#>
    Plant Type Treat conc.m conc.s uptake.m uptake.s
#> 1
      Qn1 Que nchil 435 317.7
                                    33.23
#> 2
      Qc1 Que chil
                       435 317.7
                                    29.97
                                             8.335
                       435 317.7
                                    26.40
                                             8.694
      Mn1 Mis nchil
      Mc1 Mis chil
                       435 317.7
                                    18.00
                                             4.119
summaryBy(. ~ 1, data=CO2, FUN=myfun1)
    conc.m conc.s uptake.m uptake.s
      435 299.6
                    26.9
                             9.189
```

Spcifications as formulas and lists

The convention for the "By"-functions is that a two sided formula like can be written in two ways:

```
cbind(x, y) ~ A + B
list(c("x", "y"), c("A", "B"))
```

Some "By"-functions only take a right hand sided formula as input. Such a formula can also be written in two ways:

```
~ A + B
c("A", "B")
```

The list-form / vector-form is especially useful if a function is invoked programatically. Hence the calls to summaryBy above can also be made as

```
summaryBy(list(c("conc", "uptake", "lu=log(uptake)"), "Plant"), \ data=CO2, \ FUN=myfun1) \\ summaryBy(list(c("."), c(".")), \ data=CO2, \ FUN=myfun1) \\ summaryBy(list(c("."), c("1")), \ data=CO2, \ FUN=myfun1) \\
```

The orderBy function

Ordering (or sorting) a data frame is possible with the orderBy function. Suppose we want to order the rows of the the CO2 data by increasing values of conc and decreasing value of uptake (within conc):

```
x1 <- orderBy(~ conc - uptake, data=CO2)
head(x1)
#>
     Plant Type conc uptake Treat
#> 1
       Qn1 Que 95
                     16.0 nchil
                 95
                      14.2 chil
#> 22
       Qc1
            Que
#> 43
           Mis
                 95
                      10.6 nchil
       Mn1
#> 64
       Mc1
            Mis
                 95
                      10.5 chil
#> 2
       Qn1
           Que 175
                      30.4 nchil
#> 23
      Qc1 Que 175
                      24.1 chil
```

Following the remarks about specification in "By"-functions, an equivalent form is:

```
orderBy(c("conc", "-uptake"), data=CO2)
```

The splitBy function

Suppose we want to split CO2 into a list of dataframes:

```
x1 <- splitBy(~ Plant + Type, data=CO2)
x1

#> listentry Plant Type
#> 1  Qn1|Que  Qn1  Que
#> 2  Qc1|Que  Qc1  Que
#> 3  Mn1|Mis  Mn1  Mis
#> 4  Mc1|Mis  Mc1  Mis
```

The result is a list (with a few additional attributes):

```
lapply(x1, head, 2)
#> $'Qn1|Que'
#> Plant Type conc uptake Treat
#> 1 Qn1 Que 95 16.0 nchil
#> 2 Qn1 Que 175
                   30.4 nchil
#>
#> $'Qc1|Que'
#> Plant Type conc uptake Treat
#> 22 Qc1 Que 95 14.2 chil
#> 23 Qc1 Que 175
                    24.1 chil
#>
#> $`Mn1|Mis`
#> Plant Type conc uptake Treat
#> 43 Mn1 Mis 95 10.6 nchil
#> 44 Mn1 Mis 175 19.2 nchil
#>
#> $`Mc1|Mis`
#> Plant Type conc uptake Treat
#> 64 Mc1 Mis 95 10.5 chil
#> 65 Mc1 Mis 175 14.9 chil
```

The subsetBy function

Suppose we want to select those rows within each treatment for which the uptake is larger than 75% quantile of uptake (within the treatment). This is achieved by:

The transformBy function

The transformBy function is analogous to the transform function except that it works within groups. For example:

The 1mBy function

The 1mBy function allows for fitting linear models to different strata of data (the vertical bar is used for defining groupings of data):

The result is a list with a few additional attributes and the list can be processed further as e.g.

Functions related linear estimates and contrasts

A linear function of a p-dimensional parameter vector β has the form

$$C = L\beta$$

where L is a $q \times p$ matrix which we call the Linear Estimate Matrix or simply LE-matrix. The corresponding linear estimate is $\hat{C} = L\hat{\beta}$. A linear hypothesis has the form $H_0: L\beta = m$ for some q dimensional vector m.

A working dataset - the ToothGrowth data

The response is the length of odontoblasts cells (cells responsible for tooth growth) in 60 guinea pigs. Each animal received one of three dose levels of vitamin C (0.5, 1, and 2 mg/day) by one of two delivery methods, (orange juice (coded as OJ) or ascorbic acid (a form of vitamin C and (coded as VC)). The dataset is balanced with 10 measurements for each combination of dose and supp. To illustrated certain points in what follows we make data unbalance by removing the some rows of the dataframe:

```
data("ToothGrowth")
ToothGrowth <- transform(ToothGrowth, dose = factor(dose))
ToothGrowth <- ToothGrowth[-(1:3), ]
head(ToothGrowth, 4)

#> len supp dose
#> 4 5.8 VC 0.5
#> 5 6.4 VC 0.5
#> 6 10.0 VC 0.5
#> 7 11.2 VC 0.5
```

The interaction plot indicates some interaction between dose and supp. This is also supported by a formal test:

```
tooth1 <- lm(len ~ dose + supp, data=ToothGrowth)
tooth2 <- lm(len ~ dose * supp, data=ToothGrowth)
anova(tooth1, tooth2)

#> Analysis of Variance Table
#>
#> Model 1: len ~ dose + supp
#> Model 2: len ~ dose * supp
```

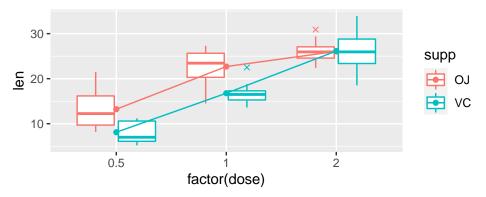


Figure 1: Interaction plot for the ToothGrowth data. The average 'len' for each group is a dot. Boxplot outliers are crosses.

```
#> Res.Df RSS Df Sum of Sq F Pr(>F)
#> 1     53 789
#> 2     51 685 2     104 3.88 0.027
```

Computing linear estimates

For now, we focus on the additive model. Consider computing the estimated length for each dose of orange juice (OJ): One option: Construct the LE–matrix L directly and then invoke linest:

```
L <- matrix(c(1, 0, 0, 0, 0, 1, 1, 0, 0, 1, 0, nrow=3, byrow=T)
```

The matrix *L* can be generated as follows:

```
L <- LE_matrix(tooth1, effect="dose", at=list(supp="0J"))
```

The estimates can be computed directly as

```
L %*% coef(tooth1)
```

but we do not obtain standard errors etc. this way. Instead we can invoke linest

```
c1 <- linest(tooth1, L)</pre>
coef(c1)
#>
     estimate std.error statistic df p.value
       12.59 1.027 12.26 53 4.239e-17
#> 1
#> 2
       21.52
                1.004
                           21.43 53 8.969e-28
#> 3
       27.88
              1.004
                        27.77 53 2.958e-33
confint(c1)
    0.025 0.975
#> 1 10.53 14.65
#> 2 19.50 23.53
#> 3 25.87 29.90
```

The function esticon has been part of doBy for many years while linest is a newer addition. The functionality, however, is similar:

```
c1 <- esticon(tooth1, L)
c1

#> estimate std.error statistic p.value beta0 df
#> [1,] 12.59 1.03 12.26 0.00 0.00 53
#> [2,] 21.52 1.00 21.43 0.00 0.00 53
#> [3,] 27.88 1.00 27.77 0.00 0.00 53
```

Least-squares means (LS-means)

A related question could be: What is the estimated length for each dose if we ignore the source of vitamin C (i.e. whether it is OJ or VC). One approach would be to fit a model in which source does not appear:

```
tooth0 <- update(tooth1, . ~ . - supp)
L0 <- LE_matrix(tooth0, effect="dose")
#>
       (Intercept) dose1 dose2
         1
#> [1,]
                    0
                          0
#> [2,]
               1
                    1
#> [3,]
               1
                     0
                          1
linest(tooth0, L=L0)
#> Coefficients:
#> estimate std.error statistic
                                    df p.value
#> [1,] 11.124 1.027 10.830 54.000
                        20.841 54.000
                  0.947
                                            0
#> [2,] 19.735
               0.947 27.562 54.000
#> [3,] 26.100
                                            0
```

An alternative would be to stick to the original model but compute the estimate for an "average vitamin C source". That would correspond to giving weight 1/2 to each of the two vitamin C source parameters. However, as one of the parameters is already set to zero to obtain identifiability, we obtain the LE-matrix L as

```
L1 \leftarrow matrix(c(1, 0, 0, 0.5,
              1, 1, 0, 0.5,
              1, 0, 1, 0.5), nrow=3, byrow=T)
linest(tooth1, L=L1)
#> Coefficients:
      estimate std.error statistic
                                       df p.value
#> [1,] 10.809 0.940 11.496 53.000
#> [2,] 19.735
                    0.863 22.873 53.000
                                                0
#> [3,] 26.100
                    0.863 30.249 53.000
                                                0
```

Such a particular linear estimate is sometimes called a least-squares mean, an LSmean, a marginal mean or a population mean. Notice: One may generate L automatically with

```
L1 <- LE_matrix(tooth1, effect="dose")
L1
       (Intercept) dose1 dose2 suppVC
#> [1,]
             1 0 0 0.5
#> [2,]
                1
                     1
                           0
                                0.5
#> [3.]
                1
                     0
                          1
                                0.5
```

Notice: One may obtain the LSmean directly as:

```
LSmeans(tooth1, effect="dose")
```

```
#> Coefficients:
#> estimate std.error statistic
                                      df p.value
#> [1,]
      10.809 0.940 11.496 53.000
                                               0
                   0.863 22.873 53.000
#> [2,]
        19.735
                                               0
                   0.863 30.249 53.000
#> [3,]
         26.100
                                               0
  which is the same as
L <- LE_matrix(tooth1, effect="dose")
linest(tooth1, L=L)
```

Interaction model

For a model with interactions, the LSmeans are

```
LSmeans(tooth2, effect="dose")
#> Coefficients:
#> estimate std.error statistic
                                  df p.value
#> [1,] 10.672 0.903 11.819 51.000
                  0.819 24.085 51.000
#> [2,] 19.735
#> [3,] 26.100
                  0.819 31.853 51.000
  In this case, the LE-matrix is
L <- LE_matrix(tooth2, effect="dose")
L
      (Intercept) dose1 dose2 suppVC dose1:suppVC dose2:suppVC
#> [1,]
        1 0 0 0.5
                                   0.0
#> [2,]
               1
                    1
                         0
                              0.5
                                         0.5
#> [3,]
               1
                     0
                         1
                              0.5
                                         0.0
```

Using (transformed) covariates

Covariates are fixed at their average value unles the at=...-argument is used. For example, conc is fixed at the average value:

0.0

0.5

```
co2.lm1 <- lm(uptake ~ conc + Type + Treat, data=CO2)
lsm1 <- LSmeans(co2.lm1, effect="Treat")</pre>
1sm1
#> Coefficients:
#> estimate std.error statistic      df p.value
#> [1,] 29.81 1.35 22.16 24.00
#> [2,] 23.99
                   1.35
                           17.83 24.00
lsm1$L
      (Intercept) conc TypeMis Treatchil
#> [1,]
          1 435 0.5
#> [2,]
               1 435
                          0.5
                                     1
```

A special issue arises in connection with transformed covariates. Consider:

```
co2.lm2 <- lm(uptake ~ conc + I(conc^2) + log(conc) + Type + Treat, data=CO2)
lsm2 <- LSmeans(co2.lm2, effect="Treat")</pre>
1sm2
#> Coefficients:
#> estimate std.error statistic
                                   df p.value
#> [1,] 33.416 0.932 35.858 22.000
#> [2,] 27.587
                  0.932
                          29.603 22.000
1sm2$L
     (Intercept) conc I(conc^2) log(conc) TypeMis Treatchil
#> [1,]
        1 435 189225 6.075 0.5 0
#> [2,]
               1 435
                         189225
                                   6.075
                                            0.5
                                                      1
```

Above I (conc^2) is the the square of the average of conc - not the average of the squared values of conc. Likewise log(conc) is the log of the average of conc - not the average of the log of conc, cfr. this:

```
c(mean(CO2$conc)^2, mean(CO2$conc^2))
#> [1] 189225 275754
c(log(mean(CO2$conc)), mean(log(CO2$conc)))
```

```
#> [1] 6.075 5.819
```

To make computations based on the average value of the square of conc and the average of the log of conc do

```
co2.lm3 <- lm(uptake ~ conc + conc2 + log.conc + Type + Treat,
            data=transform(CO2, conc2=conc^2, log.conc=log(conc)))
lsm3 <- LSmeans(co2.lm3, effect="Treat")</pre>
lsm3
#> Coefficients:
#> estimate std.error statistic
                                      df p.value
#> [1,] 29.814 0.737 40.479 22.000
#> [2,] 23.986
                   0.737
                           32.566 22.000
1sm3$I
#>
       (Intercept) conc conc2 log.conc TypeMis Treatchil
#> [1,]
        1 435 275754 5.819 0.5 0
#> [2,]
                1 435 275754
                                5.819
                                         0.5
                                                    1
   If we want to evaluate the LS-means at conc=700 then we can do:
lsm4 <- LSmeans(co2.lm3, effect="Treat", at=list(conc=700, conc2=700^2, log.conc=log(700)))</pre>
1sm4
#> Coefficients:
#>
     estimate std.error statistic df p.value
#> [1,]
        34.54 1.19 29.02 22.00
                                            0
#> [2,]
                    1.19
                            24.12 22.00
          28.71
1sm4$L
      (Intercept) conc conc2 log.conc TypeMis Treatchil
          1 700 490000 6.551 0.5
#> [1,]
                                                    0
                1 700 490000
                                6.551
#> [2,]
                                         0.5
                                                    1
```

Alternative models

The functions esticon, linest, LSmeans' etc. are available for a range of model classes. We illustrate a few below: We may decide to treat supp as a random effect. This leads to a *linear mixed effects model* as implemented in Bates et al. (2015):

```
library(lme4)
tooth.mix <- lmer(len ~ dose + (1|supp), data=ToothGrowth)</pre>
LSmeans(tooth.mix, effect="dose")
#> Coefficients:
#> [1,] 10.84 1.95 5.56 1.43 0.06
#> [2,]
                1.91
                       10.32 1.33
        19.73
                                    0 03
#> [3,]
        26.10
                1.91
                       13.65 1.33
                                   0.02
```

Notice here that the parameter estimates themselves are similar to those of a linear model (had data been completely balanced, the estimates would have been identical). However, the standard errors of the the estimates are much larger under the mixed model. This is due to supp being treated as a random effect. Notice that the degrees of freedom by default are adjusted using a Kenward–Roger approximation (provided that pbkrtest package (Halekoh and Højsgaard, 2014) is installed). Adustment of degrees of freedom is controlled with the adjust of argument.

We can also calculate LS-means for e.g. a Poisson or a gamma model in a *generalized linear model* setting. Notice that the LS-means are on the scale of the linear predictor - not on the scale of the response.

```
tooth.gam <- glm(len \sim dose + supp, family=Gamma("identity"), data=ToothGrowth) LSmeans(tooth.gam, effect="dose")
```

Likewise, for *generalized estimating equations* as implemented in the geepack package, (Halekoh et al., 2006), we get:

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