

# frodyma\_2020

March 31, 2020

## 1 RNA-seq analysis – Frodyma, et. al.; 2020

The following notebook details the analyses performed to analyze the RNA-seq data of the PGC1 or ERR knockdown in HCT116 cells.

Raw data was processed and differential expression analysis using [XPRESSyourself](#).

### 1.0.1 Sequencing Methods

cDNA libraries of RNA libraries were prepared using \_\_\_\_\_. cDNA libraries were sequenced at \_\_\_\_\_ using \_\_\_\_\_. Raw sequence data has been deposited as GSE#####.

### 1.0.2 Processing and Analysis Methods

Raw FASTQ files were processed using XPRESSpipe v 0.4.1.

Script executed as `sbatch process.sh`

```
#!/bin/bash
#SBATCH --time=72:00:00
#SBATCH --nodes=1
#SBATCH -o /scratch/general/lustre/$USER/slurmjob-%j
#SBATCH --partition=kingspeak

#set up the temporary directory
SCRDIR=/scratch/general/lustre/$USER/$SLURM_JOBID
mkdir -p $SCRDIR

FILES=/scratch/general/lustre/$USER/danielle_files/input/files

# Source FASTA files obtained as:
# for X in 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 X Y; do wget ftp://ftp.en.
# Source GTF file obtained as:
# wget ftp://ftp.ensembl.org/pub/release-99/gtf/homo_sapiens/Homo_sapiens.GRCh38.99.gtf.gz
REF=/scratch/general/lustre/$USER/references/human_reference_pe_v99

mkdir $SCRDIR/input
```

```

mkdir $SCRDIR/output

cp $FILES/*.fastq $SCRDIR/input/

cd $SCRDIR/.

xpresspipe makeReference -o $REF -f $REF/genome_fastas -g $REF/transcripts.gtf --sjdbOverhang 100

xpresspipe peRNAseq -i $SCRDIR/input -o $SCRDIR/output -r $REF --gtf $REF/transcripts.gtf -e f

mkdir $SCRDIR/cufflinks
xpresspipe count -i $SCRDIR/output/alignments_genome -o $SCRDIR/cufflinks -g $REF/transcripts.gtf

```

### 1.0.3 Import dependencies

```

[1]: import pandas as pd
import numpy as np
import xpressplot as xp
import matplotlib
import matplotlib.pyplot as plt
%matplotlib inline

```

## 1.1 Differential expression analysis

### 1.1.1 Compile counts

1. Samples were run on 4 lanes. Processed read data are compiled for analysis at biological replicate level.
2. Export as two separate tables for the different experimental groups and the control for differential expression analysis

```

[2]: counts = pd.read_csv(
    "./data/frodyama_count_table.tsv",
    sep='\t',
    index_col=0)
counts.columns = [x.split('_')[0] for x in counts.columns.tolist()]
counts = counts.groupby(counts.columns, axis=1).sum()

```

```

[3]: # Export ERRA counts
ERRa_counts = counts[[
    'HCT116siCONT1',
    'HCT116siCONT2',
    'HCT116siERRa7',
    'HCT116siERRa8']]

```

```

ERRa_counts.to_csv(
    './data/counts_erra.txt',
    sep='\t')

# Export metadata
ERRa_meta = pd.DataFrame()
ERRa_meta['samples'] = [
    'HCT116siCONT1',
    'HCT116siCONT2',
    'HCT116siERRa7',
    'HCT116siERRa8']
ERRa_meta['conditions'] = [
    'control',
    'control',
    'experiment',
    'experiment']
ERRa_meta = ERRa_meta.set_index('samples')
del ERRa_meta.index.name

ERRa_meta.to_csv(
    './data/metadata_erra.txt',
    sep='\t')

```

```

[4]: # Export PGC1b counts
PGC1B_counts = counts[[
    'HCT116siCONT1',
    'HCT116siCONT2',
    'HCT116sipgc1b2',
    'HCT116sipgc1b6']]

PGC1B_counts.to_csv(
    './data/counts_pgc1b.txt',
    sep='\t')

# Export metadata
PGC1B_meta = pd.DataFrame()
PGC1B_meta['samples'] = [
    'HCT116siCONT1',
    'HCT116siCONT2',
    'HCT116sipgc1b2',
    'HCT116sipgc1b6']
PGC1B_meta['conditions'] = [
    'control',
    'control',
    'experiment',
    'experiment']
PGC1B_meta = PGC1B_meta.set_index('samples')

```

```
del PGC1B_meta.index.name

PGC1B_meta.to_csv(
    './data/metadata_pgc1b.txt',
    sep='\t')
```

### 1.1.2 Perform differential expression analysis

Uses DESeq2 v1.22.1 via the XPRESSpipe wrapper

*# ERRa*

```
$ xpresspipe diffxpress -i ./data/counts_erra.txt -s ./data/metadata_erra.txt --design condition
```

*# PGC1b*

```
$ xpresspipe diffxpress -i ./data/counts_pgc1b.txt -s ./data/metadata_pgc1b.txt --design condition
```

*# Rename gene IDs as gene names*

```
$ xpresspipe convertNames -i ./data/counts_erra_diffx.tsv -g ~/Desktop/Homo_sapiens.GRCh38.99.genes
```

```
$ xpresspipe convertNames -i ./data/counts_pgc1b_diffx.tsv -g ~/Desktop/Homo_sapiens.GRCh38.99.genes
```

### 1.1.3 Import differential expression data

```
[5]: err_file = "./data/counts_erra_diffx_renamed.tsv"
err_data = pd.read_csv(
    err_file,
    sep='\t',
    index_col=0)
del err_data.index.name
```

```
[6]: pgc1_file = "./data/counts_pgc1b_diffx_renamed.tsv"
pgc1_data = pd.read_csv(
    pgc1_file,
    sep='\t',
    index_col=0)
del pgc1_data.index.name
```

### 1.1.4 RNA-seq Count Analysis

Genes of interest: - PCK2 (ENSG00000100889) - SLC1A4 (ENSG00000115902) - ESRRA (ENSG00000173153) - PPARGC1B (ENSG00000155846)

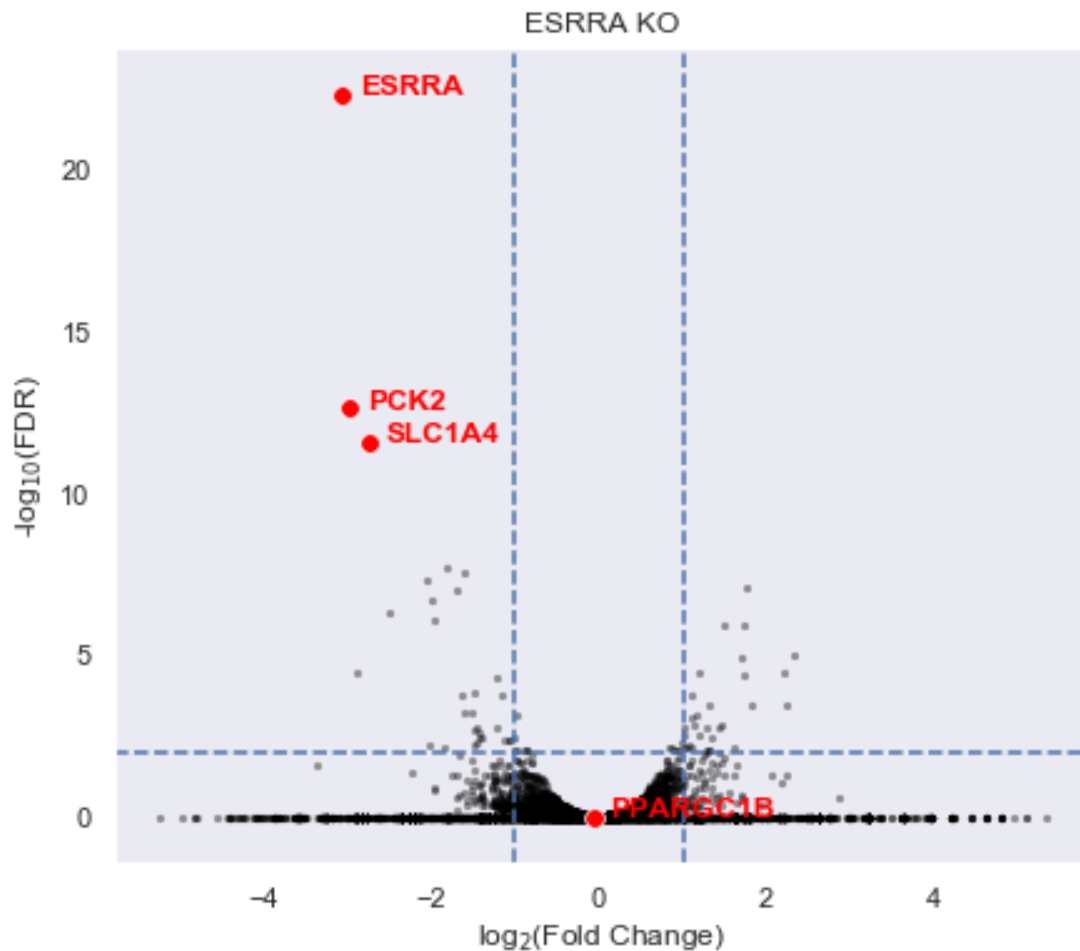
```
[7]: highlight_points = [
    'PCK2',
    'SLC1A4',
    'ESRRA',
```

```
'PPARGC1B']
```

## ERRA (ESRRA)

```
[8]: xp.rna_volcano(  
    err_file,  
    highlight_points=highlight_points,  
    highlight_color='red',  
    label_points=highlight_points,  
    y_threshold=2,  
    x_threshold=[-1,1],  
    size=10,  
    highlight_size=60,  
    alpha=0.4,  
    figsize=(7,6),  
    title="ESRRA KO")
```

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### PGC1B (PPARGC1B)

```
[9]: xp.rna_volcano(  
    pgc1_file,  
    highlight_points=highlight_points,  
    highlight_color='red',  
    label_points=highlight_points,  
    y_threshold=2,  
    x_threshold=[-1,1],  
    size=10,  
    highlight_size=60,  
    alpha=0.4,  
    figsize=(7,6),  
    title="PPARGC1B KO")
```

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