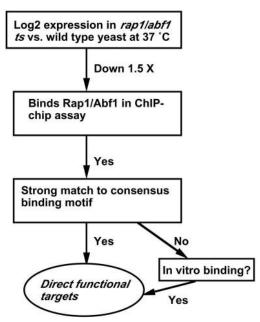
Prac9: Background

Abf1 is general regulatory factor (RFs) that contribute to transcriptional activation of a large number of genes, as well as to replication, silencing and telomere structure in yeast In spite of their widespread roles in transcription, the scope of their functional targets genome-wide has not been previously determined

Yarragudi et al use microarrays to examine the contribution of these essential RFs to transcription genome-wide, by using mutants that dissociate from their binding sites at 37C

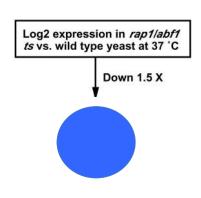
Yarragudi et al. Genome-wide analysis of transcriptional dependence and probable target sites for Abf1 and Rap1 in Saccharomyces cerevisiae. Nucleic Acids Res. 35(1) 2007.

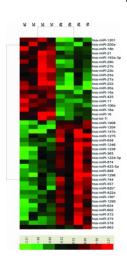
Bioinformatics: Identify targets



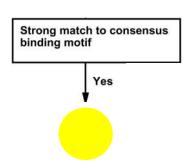
From: Yarragudi et al. Genome-wide analysis of transcriptional dependence and probable target sites for Abf1 in Saccharomyces cerevisiae. *Nucleic Acids Res.* 35(1) 2007.

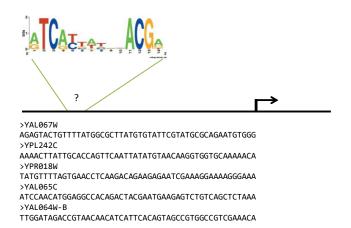
Differential expression: Identify targets





DNA binding: Identify targets





Fishers Exact Test

- Quantify statistical significance of an association between two properties
- Used gene set enrichment

	Has Property	Does not have property	Row total
In gene set of interest	а	b	a+b
Not in gene set	С	d	c+d
Column total	a+c	b+d	a+b+c+d

- α or significance level a probability which is fixed in advance of making the hypothesis test.
- If the observed p-value is smaller than the significance level then the null hypothesis is rejected.
- Null hypothesis
- "Drug x is not indicative of chaperone regulator activity"

Exercise 1 Code

import stats

2 genes are annotated as negatives

c=2

#14 genes in our negative set

d = 14 - 2

Positive set of genes

Positives = set({"YPL106C", "YOL081W", "YOR027W", "YOR299W", "YNL006W", "YNL007C", "YLL039C", "YLR216C"})

Genes annotated with GO Term

has_property=set({"YER048C", "YIL016W", "YLR090W", "YOR027W", "YMR161W", "YNL064C", "YNL281W", "YDR214W", "YPL106C", "YNL007C", "YNL227C"})

We need to overlap Positives and has_property

a= # Fill me in here

#number of positives-a

b= Fill me in here

Print b

pval = stats.getFETpval(a, b, c, d,
left=False)

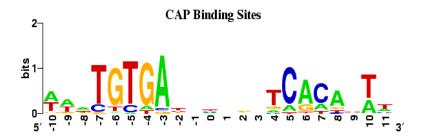
print pval

- Provide the p-value and the significance level you are using.
- And a statement (reject or not reject null hypothesis)

- seqs=readFastaFile("yeast_promoters.fa")
- print len(seqs)
- Hint: look at SCPD as a source
- 1-2 lines (How they are biologically sensible)

- Visualizing motifs using "logo"
- Shows sequence conservation
- Frequency of residue

Example

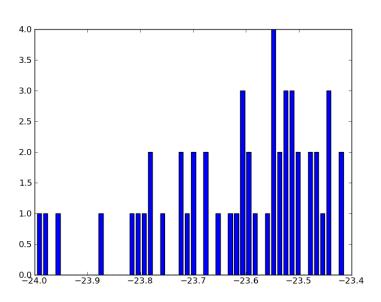


- Column 10
- Why do you only see G and A (few lines)
- Think about how PWMs are constructed
- Why total height is around 1bit (1bit is half of 2 bits)
- Think about what the height indicates.

Exercise 5 code

```
>>> bind_map = {}
>>> for s in yeast_prom: # yeast_prom is an array of sequences
#Insert condition here !!!
bind_map[s.name] = abf1_pwm.maxscore(s)[0] # save score
```

Try to aim for this graph



- histogram
- Provide the list of 50 target genes
- Few lines (explaining your reason for the threshold)

Abf1 SOFT file

```
:dataset_update_date = mar בי צטעט
SUBSET = GDS3198_1
!subset_dataset_id = GDS3198
!subset_description = wild type
!subset_sample_id = GSM140786,GSM140800,GSM140801
!subset_type = genotype/variation
^SUBSET = GDS3198 2
!subset_dataset_id = GDS3198
!subset description = Abf1 mutant
!subset_sample_id = GSM140802,GSM140803,GSM140804
!subset_type = genotype/variation
^DATASET = GDS3198
#ID_REF = Platform reference identifier
#IDENTIFIER = identifier
#GSM140786 = Value for GSM140786: Abf1 wt 37 C rep1; src: Abf1 wild type control
#GSM140800 = Value for GSM140800: Abf1 wt 37 C rep2; src: Abf1 wild type control
#GSM140801 = Value for GSM140801: Abf1 wt 37C rep3; src: Abf1 wild type control
#GSM140802 = Value for GSM140802: Abf1 ts 37 C rep1; src: Abf1 ts mutant
#GSM140803 = Value for GSM140803: Abf1 ts 37 C rep2; src: Abf1 ts mutant
#GSM140804 = Value for GSM140804: Abf1 ts 37 C rep3; src: Abf1 ts mutant
!dataset_table_begin
ID REF IDENTIFIER GSM140786
                                GSM140800
                                            GSM140801
                                                        GSM140802
                                                                    GSM140803
                                                                                GSM140804
10000_at
           YLR331C 24.600 24.800 2.800
                                            28.500 31.900 23.900
10001_at
           MID2
                    1725.400
                                1485.400
                                            1723.000
                                                        1891.900
                                                                    1236.700
                                                                                1572.500
10002 i at RPS25B 3201.000
                                3320.100
                                            3851.900
                                                        4330.000
                                                                    4849.700
                                                                                4194.800
```

Provide probe and gene numbers...

g1 = ge.readGEOFile('GDS3198.soft', id_column = 0)

g2 = ge.readGEOFile('GDS3198.soft', id_column = 1) Hint: getGenes() and len() may be useful.

- Code
- Pairing (WT/mutants)
- Mention the transformations (ie. Log)
- How you filtered the top 100 and lowest 100
- Hint: indexing was useful.

```
Code
result = sorted(meanfold.items(), key=lambda v: v[1])
print '======== Wildtype may down-regulate ========'
for r in result[0:100]:
#Fill me in I am only one condition:
    print r[0]
print '======== Wildtype may up-regulate ======='
for r in result[-1:-100:-1]:
    # fill me in I am only 1 condition
    print r[0]
```

Exercise 8 cont'd

Provide the gene list of 50 genes like so.

====== Wildtype may down-regulate ======

ATG29

YCLWOMEGA2

YLL067C

CDA1

YAL064W-B

YHR145C

YPR078C

RTG1

YOLCDELTA2

SPR3

YLR279W

...

- Submit: A simple explanation (1-2 lines) why it is useful
- Hint: Consider multiple hypothesis testing
- (i.e. testing n terms)

Exercise 10 +11

- For Q10 Bind_map may be useful.
- For Q11 Store the gene symbols
- Provide
- Significant GO Terms

- Helpful link:
- http://www.yeastgenome.org/cgibin/locus.fpl?locus=abf1
- Use get_GO_description