

# Prac9: Background

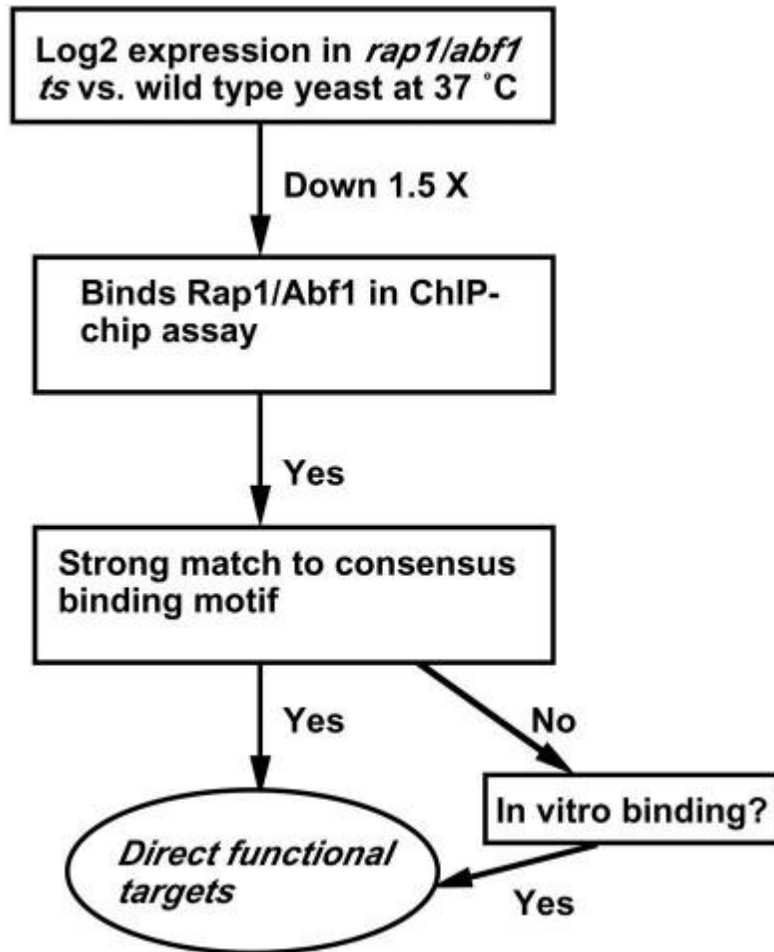
Abf1 is general regulatory factor (RFs) that contribute to transcriptional activation of a large number of genes, as well as to replication, silencing and telomere structure in yeast

In spite of their widespread roles in transcription, the scope of their functional targets genome-wide has not been previously determined

Yarragudi et al use microarrays to examine the contribution of these essential RFs to transcription genome-wide, by using mutants that dissociate from their binding sites at 37C

Yarragudi et al. Genome-wide analysis of transcriptional dependence and probable target sites for Abf1 and Rap1 in *Saccharomyces cerevisiae*. *Nucleic Acids Res.* 35(1) 2007.

# Bioinformatics: Identify targets

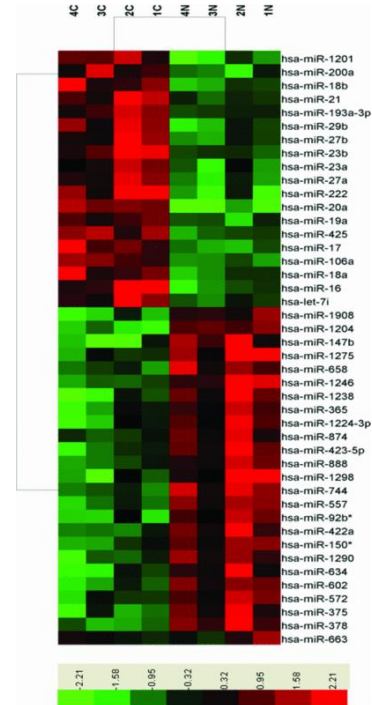
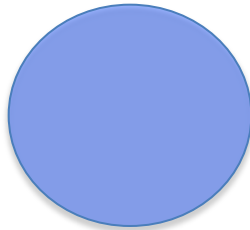


From: Yarragudi et al. Genome-wide analysis of transcriptional dependence and probable target sites for Abf1 in *Saccharomyces cerevisiae*. *Nucleic Acids Res.* 35(1) 2007.

# Differential expression: Identify targets

Log2 expression in *rap1/abf1*  
*ts* vs. wild type yeast at 37 °C

Down 1.5 X



# DNA binding: Identify targets



```
>YAL067W
AGAGTACTGTTTTATGGCGCTTATGTGTATTCGTATGCGCAGAATGTGGG
>YPL242C
AAAAC TTATTGCACCAGTTCAATTATATGTAACAAGGTGGTGCAAAAACA
>YPR018W
TATGTTTTAGTGAACCTCAAGACAGAAGAGAATCGAAAGGAAAAGGGAAA
>YAL065C
ATCCAACATGGAGGCCACAGACTACGAATGAAGAGTCTGTCAGCTCTAAA
>YAL064W-B
TTGGATAGACCGTAACAACATCATTACAGTAGCCGTGGCCGTCGAAACA
```

# Fishers Exact Test

- Quantify statistical significance of an association between two properties
- Used gene set enrichment

	Has Property	Does not have property	Row total
In gene set of interest	a	b	a+b
Not in gene set	c	d	c+d
Column total	a+c	b+d	a+b+c+d

# Exercise 1

- $\alpha$  or significance level – a probability which is fixed in advance of making the hypothesis test.
- If the observed p-value is smaller than the significance level then the null hypothesis is rejected.
- Null hypothesis
- "Drug x is not indicative of chaperone regulator activity"

# Exercise 1 Code

```
import stats
```

```
# 2 genes are annotated as negatives
```

```
c=2
```

```
#14 genes in our negative set
```

```
d= 14 -2
```

```
# Positive set of genes
```

```
Positives = set({"YPL106C", "YOL081W",  
"YOR027W", "YOR299W", "YNL006W", "YNL007C",  
"YLL039C", "YLR216C"})
```

```
# Genes annotated with GO Term
```

```
has_property=set({"YER048C", "YIL016W",  
"YLR090W", "YOR027W", "YMR161W", "YNL064C",  
"YNL281W", "YDR214W", "YPL106C", "YNL007C",  
"YNL227C"})
```

```
# We need to overlap Positives and has_property
```

```
a= # Fill me in here
```

```
#number of positives-a
```

```
b= Fill me in here
```

```
Print b
```

```
pval = stats.getFETpval(a, b, c, d,  
left=False)
```

```
print pval
```

# Exercise 1

- Provide the p-value and the significance level you are using.
- And a statement (reject or not reject null hypothesis)



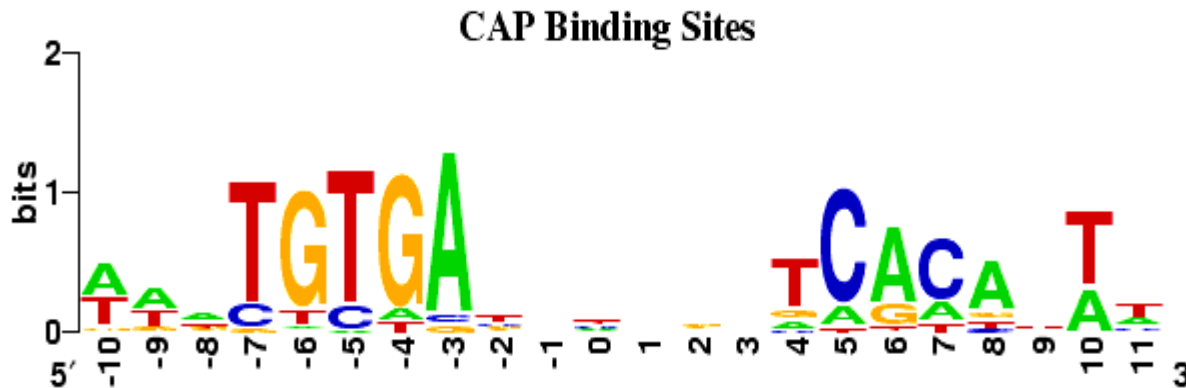
## Exercise 2

- `seqs=readFastaFile("yeast_promoters.fa")`
- `print len(seqs)`
- Hint: look at SCPD as a source
- 1-2 lines (How they are biologically sensible)

# Exercise 3

- Visualizing motifs using “logo”
- Shows sequence conservation
- Frequency of residue

Example



# Exercise 3

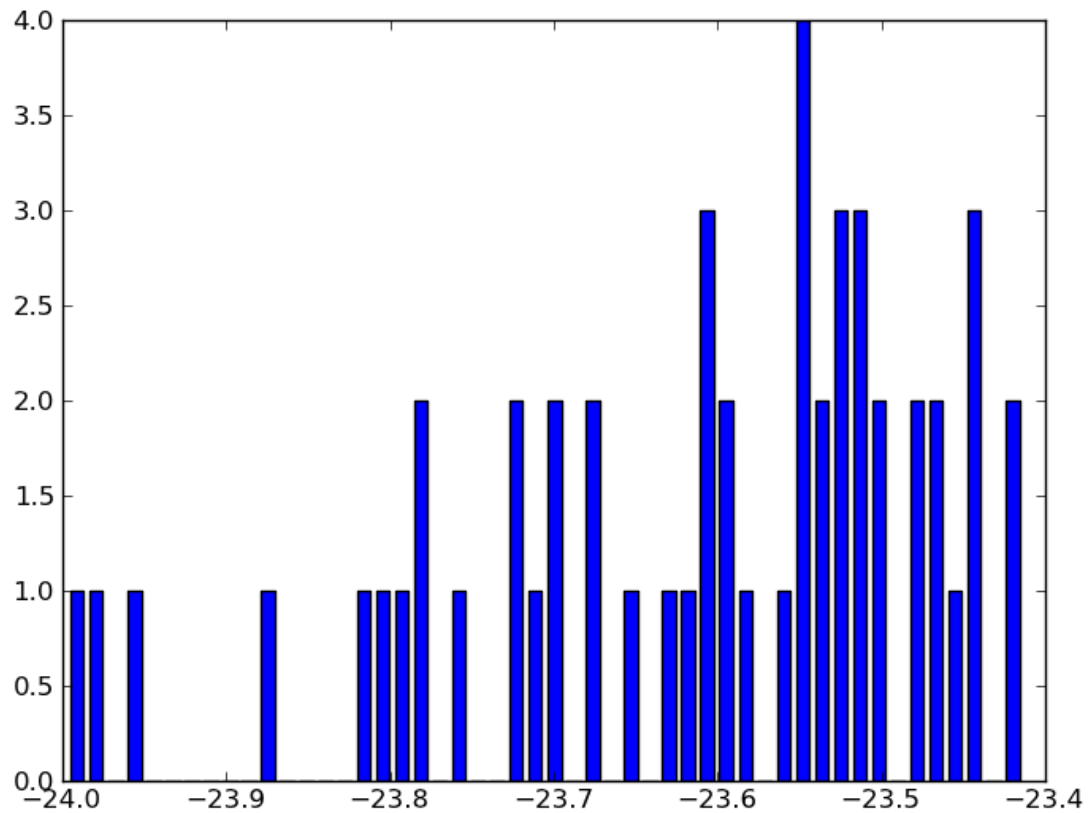
- Column 10
- Why do you only see G and A (few lines)
- Think about how PWMs are constructed
- Why total height is around 1bit (1bit is half of 2 bits)
- Think about what the height indicates.

# Exercise 5 code

```
>>> bind_map = {}  
>>> for s in yeast_prom: # yeast_prom is an array of sequences  
    #Insert condition here !!!  
    bind_map[s.name] = abf1_pwm.maxscore(s)[0] # save score
```

# Exercise 5

Try to aim for this graph



# Exercise 5

- histogram
- Provide the list of 50 target genes
- Few lines (explaining your reason for the threshold)

# Abf1 SOFT file

```
!dataset_update_date = mar 19 2008
^SUBSET = GDS3198_1
!subset_dataset_id = GDS3198
!subset_description = wild type
!subset_sample_id = GSM140786,GSM140800,GSM140801
!subset_type = genotype/variation
^SUBSET = GDS3198_2
!subset_dataset_id = GDS3198
!subset_description = Abf1 mutant
!subset_sample_id = GSM140802,GSM140803,GSM140804
!subset_type = genotype/variation
^DATASET = GDS3198
#ID_REF = Platform reference identifier
#IDENTIFIER = identifier
#GSM140786 = Value for GSM140786: Abf1 wt 37 C rep1; src: Abf1 wild type control
#GSM140800 = Value for GSM140800: Abf1 wt 37 C rep2; src: Abf1 wild type control
#GSM140801 = Value for GSM140801: Abf1 wt 37C rep3; src: Abf1 wild type control
#GSM140802 = Value for GSM140802: Abf1 ts 37 C rep1; src: Abf1 ts mutant
#GSM140803 = Value for GSM140803: Abf1 ts 37 C rep2; src: Abf1 ts mutant
#GSM140804 = Value for GSM140804: Abf1 ts 37 C rep3; src: Abf1 ts mutant
!dataset_table_begin
ID_REF  IDENTIFIER  GSM140786  GSM140800  GSM140801  GSM140802  GSM140803  GSM140804
10000_at  YLR331C  24.600  24.800  2.800  28.500  31.900  23.900
10001_at  MID2  1725.400  1485.400  1723.000  1891.900  1236.700  1572.500
10002_i_at  RPS25B  3201.000  3320.100  3851.900  4330.000  4849.700  4194.800
```

# Exercise 6

Provide probe and gene numbers...

```
g1 = ge.readGEOFile('GDS3198.soft', id_column = 0)
```

```
g2 = ge.readGEOFile('GDS3198.soft', id_column = 1)
```

Hint: `getGenes()` and `len()` may be useful.



# Exercise 7

- Code
- Pairing (WT/mutants)
- Mention the transformations (ie. Log)
- How you filtered the top 100 and lowest 100
- Hint: indexing was useful.

# Exercise 8

Code

```
result = sorted(meanfold.items(), key=lambda v: v[1])
print '===== Wildtype may down-regulate ====='
for r in result[0:100]:
    #Fill me in I am only one condition:
        print r[0]
print '===== Wildtype may up-regulate ====='
for r in result[-1:-100:-1]:
    # fill me in I am only 1 condition
        print r[0]
```

# Exercise 8 cont'd

Provide the gene list of 50 genes like so.

===== Wildtype may down-regulate =====

ATG29

YCLWOMEGA2

YLL067C

CDA1

YAL064W-B

YHR145C

YPR078C

RTG1

YOLCDELTA2

SPR3

YLR279W

...

# Exercise 9

- Submit: A simple explanation (1-2 lines) why it is useful
- Hint: Consider multiple hypothesis testing
- (i.e. testing  $n$  terms)

# Exercise 10 +11

- For Q10 Bind\_map may be useful.
- For Q11 Store the gene symbols
- Provide
- Significant GO Terms

# Exercise 12

- Helpful link:
- <http://www.yeastgenome.org/cgi-bin/locus.fpl?locus=abf1>
- Use get\_GO\_description