BPL_TEST2_Batch_calibration script with PyFMI

The key library PyFMI is installed.

After the installation a small application BPL_TEST2_Batch_calibration is loaded and run. You can continue with this example if you like.

```
!lsb_release -a # Actual VM Ubuntu version used by Google

→ No LSB modules are available.

    Distributor ID: Ubuntu
    Description:
                    Ubuntu 22.04.4 LTS
                    22.04
    Release:
    Codename:
                    jammy
%env PYTHONPATH=
→ env: PYTHONPATH=
!python --version
→ Python 3.11.11
!wget https://repo.anaconda.com/miniconda/Miniconda3-py311_24.11.1-0-Linux-x86_64.sh
!chmod +x Miniconda3-py311_24.11.1-0-Linux-x86_64.sh
!bash ./Miniconda3-py311_24.11.1-0-Linux-x86_64.sh -b -f -p /usr/local
import sys
sys.path.append('/usr/local/lib/python3.11/site-packages/')
   --2025-03-25 16:46:25-- https://repo.anaconda.com/miniconda/Miniconda3-py311 24.11.1-0-Linux-x86 64.sh
    Resolving repo.anaconda.com (repo.anaconda.com)... 104.16.32.241, 104.16.191.158, 2606:4700::6810:20f1, ...
    Connecting to repo.anaconda.com (repo.anaconda.com)|104.16.32.241|:443... connected.
    HTTP request sent, awaiting response... 200 OK
    Length: 145900576 (139M) [application/octet-stream]
    Saving to: 'Miniconda3-py311_24.11.1-0-Linux-x86_64.sh'
    Miniconda3-py311_24 100%[===========] 139.14M
                                                                         in 1.2s
    2025-03-25 16:46:26 (116 MB/s) - 'Miniconda3-py311_24.11.1-0-Linux-x86_64.sh' saved [145900576/145900576]
    PRFFTX=/usr/local
    Unpacking payload ...
    Installing base environment...
    Preparing transaction: ...working... done
    Executing transaction: ...working... done
    installation finished.
!conda update -n base -c defaults conda --yes
   Channels:
     defaults
    Platform: linux-64
    Collecting package metadata (repodata.json): done
```

Package Plan

environment location: /usr/local

added / updated specs:

Solving environment: done

conda

The following packages will be downloaded:

package	build	
ca-certificates-2025.2.25 certifi-2025.1.31 openssl-3.0.16	h06a4308_0 py311h06a4308_0 h5eee18b_0	129 KB 163 KB 5.2 MB
	Total:	5.5 MB

The following packages will be UPDATED:

Downloading and Extracting Packages: | 5.2 MB | 163 KB 0% 0/1 [00:00<?, ?it/s] 0% 0/1 [00:00<?, ?it/s] | : | : openssl-3.0.16 certifi-2025.1.31 ca-certificates-2025 | 129 KB 1: 0% 0/1 [00:00<?, ?it/s] | 5.2 MB | 163 KB 0% 0.002982593950162064/1 [00:00<00:40, 41.09s/it] openssl-3.0.16 | : 10% 0.0984012204057609/1 [00:00<00:01, 1.26s/it] certifi-2025.1.31 ca-certificates-2025 | 129 KB | : 100% 1.0/1 [00:00<00:00, 1.17it/s] | : 100% 1.0/1 [00:00<00:00, 1.17it/s] ca-certificates-2025 | 129 KB certifi-2025.1.31 | 163 KB | : 100% 1.0/1 [00:00<00:00, 1.26s/it]

Preparing transaction: done Verifying transaction: done Executing transaction: done

!conda --version
!python --version

conda 24.11.1 Python 3.11.11

!conda config --set channel_priority strict

!conda install -c conda-forge pyfmi --yes # Install the key package



Preparing transaction: done Verifying transaction: done Executing transaction: done

Now specific installation and the run simulations. Start with connecting to Github. Then upload the four files:

- FMU BPL_TEST2_Batch_linux_om_me.fmu
- Setup-file BPL_TEST2_Batch_explore.py

%%bash

git clone https://github.com/janpeter19/BPL_TEST2_Batch_calibration

Cloning into 'BPL_TEST2_Batch_calibration'...

%cd BPL_TEST2_Batch_calibration

/content/BPL_TEST2_Batch_calibration

BPL_TEST2_Batch_calibration - demo

Author: Jan Peter Axelsson

This notebook shows the possibilities for calibration of the model BPL_TEST2_Batch using scipy.optimize.minimize() routine. There are several different methods to choose between. In this notebook we work with simulated data.

The text-book model of batch cultivation we simulate is the following where S is substrate, X is cell concentration, and V is volume of the broth

$$\frac{d(VS)}{dt} = -q_S(S) \cdot VX$$

$$\frac{d(VX)}{dt} = \mu(S) \cdot VX$$

and where specific cell growth rate μ and substrate uptake rate q_S are

$$\mu(S) = Y \cdot q_S(S)$$

$$q_S(S) = q_S^{max} \frac{S}{K_s + S}$$

where Y is the yield, $q_S^{\it max}$ is the maximal specific substrate uptake rate and K_s is the corresponding saturation constant.

The parameter estimation is done with optimization methods that only require evaluation of the missmatch between simulation with given parameters and data. At start the allowed range for each parameter is given. The method used for optimization is Nelder-Mead but can easily be changed [1].

In the near future the FMU may provide first derivative gradient informaion, that will make it possible to choose corresponding method of minimize() for improved performance. This possibility is related to the upgrade to the FMI-standard ver 3.0 for the Modelica compiler.

The Python package PyFMI [2] that is the base for FMU-explore has a simplified built-in functionality for parameter estimation that also use scipy.optimize.minimize(). However, there is estimatation functionally but the purpose seems to only address smaller examples. There is for instance no support to handle models that takes sub-models from libraries and necessary changes of default parameters not to be estimated. Therefore we here define a Python function evaluate() that facilitate the formulation of the parameter estimation and also bring flexibility to choice of optimization method, default Nelder-Mead.

run -i BPL_TEST2_Batch_explore.py

→ Linux - run FMU pre-comiled OpenModelica

Model for the process has been setup. Key commands:

```
- change of parameters and initial values
     - par()
     - init()
                    - change initial values only

    simulate and plot

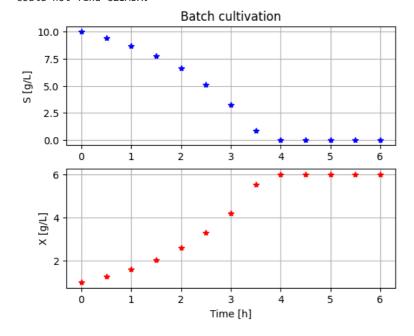
     - simu()
                   - make a new plot
     - newplot()
     - show()
                    - show plot from previous simulation
                    - display parameters and initial values from the last simulation
     - disp()
                   - describe culture, broth, parameters, variables with values/units
     - describe()
    Note that both disp() and describe() takes values from the last simulation
    and the command process_diagram() brings up the main configuration
    Brief information about a command by help(), eg help(simu)
    Key system information is listed with the command system_info()
# Adjust the size of diagrams
plt.rcParams['figure.figsize'] = [15/2.54, 12/2.54]
```

1 Generate data later used for parameter estimation

```
# Data generated
simulationTime = 6.0
par(Y=0.50, qSmax=1.00, Ks=0.1)
init(V_start=1.0, VS_start=10, VX_start=1.0)
newplot(plotType='Demo_2')
simu(simulationTime, options=opts_data)
```

import pandas as pd

Could not find cannot import name 'dopri5' from 'assimulo.lib' (/usr/local/lib/python3.11/site-packages/assimulo.lib' (/u



```
# Store data in a DataFrame for later use
data = pd.DataFrame(data={'time':sim_res['time'], 'X':sim_res['bioreactor.c[1]'], 'S':sim_res['bioreactor.c[2]']})
data
```

_		time	Х	S
	0	0.0	1.000000	1.000000e+01
	1	0.5	1.269848	9.438455e+00
	2	1.0	1.615795	8.719839e+00
	3	1.5	2.050445	7.800734e+00
	4	2.0	2.601038	6.626389e+00
	5	2.5	3.297304	5.128962e+00
	6	3.0	4.195962	3.229259e+00
	7	3.5	5.524388	8.813998e-01
	8	4.0	6.000000	-2.037810e-08
	9	4.5	6.000000	2.960320e-10
	10	5.0	6.000000	1.200938e-10
	11	5.5	6.000000	2.363337e-10
	12	6.0	6.000000	-1.553435e-10

2 Simulation with initial guess of parameters compared with data

Here we define the parameters that should be estimated and specify allowed ranges. Nominal parameters are chosen as the mid-point of the allowed parameter range.

Simulation with these nominal parameter set and compare with data give an idea of who well the model fit data.

```
# Parameters to be estimated using parDict names and their bounds
parEstim = ['Y', 'qSmax', 'Ks']
parBounds = [(0.4, 0.8), (0.7, 1.3), (0.05, 0.20)]
parEstim_0 = [np.mean(parBounds[k]) for k in range(len(parBounds))]
# Simulation with nominal parameters
newplot(plotType='Demo_1')
par(Y=parEstim_0[0], qSmax=parEstim_0[1], Ks=parEstim_0[2])
simu(simulationTime)
# Show data
ax1.plot(data['time'], data['S'],'b*')
ax2.plot(data['time'], data['X'],'r*')
plt.show()
\overline{2}
                                  Batch cultivation
        10.0
         7.5
     S [g/L]
         5.0
         2.5
         0.0
                        i
               0
                                                                    6
           6
        [√6] ×
           2
```

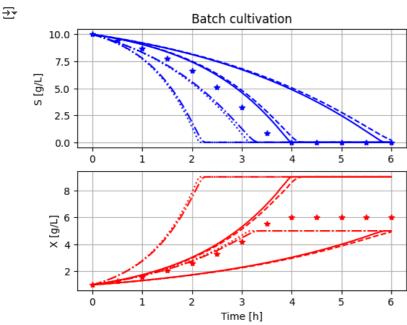
4

3

Time [h]

```
# Simulation over the parameter ranges given
newplot(plotType='Demo_1')
for Y_value in parBounds [0]:
    for qSmax_value in parBounds[1]:
        for Ks_value in parBounds[2]:
            par(Y=Y_value, qSmax=qSmax_value, Ks=Ks_value)
            simu(simulationTime)

# Show data
ax1.plot(data['time'], data['S'],'b*')
ax2.plot(data['time'], data['X'],'r*')
plt.show()
```



Simulation over the different parameter combinations of the parameter bounds shows that data is "covered" and we have good hope to find a parameter combination that fits data well.

3 Parameter estimation

Here we use the scipy.optimize.minimize() procedure which contain a family of different methods [1]. The default method is Nelder-Mead and is robust for fitting a model to data. Further we have chosen to work with bounds for the parameters to be estimated and the initial guess is chosen as the middle point in parameter space.

```
# Optimization routine import
import scipy.optimize
# Parameters to be estimated using parDict names and their bounds
extra_args = (parEstim, data, fmu_model, simulationTime)
# Modified evaluation function tailored for Python optimization algorithms
def objective(x, parEstim, data=data, fmu_model=fmu_model, simulationTime=simulationTime):
    """The parameter list is tailored for scipy optimization algorithms interface,
       where the first parameter x is an array with parameters that are tuned
       and evalauted and parEstim is a list of the names of these parameters.
       The code can be made 20-30% faster, but loner, using pyfmi-commands directly."""
    # Update parameters and simulate
    for i, p in enumerate(parEstim): par(**{p:x[i]})
    simu(simulationTime, options=opts_fast)
    # Calculate loss function V
    V['X'] = np.linalg.norm(data['X'] - np.interp(data['time'], sim_res['time'], sim_res['bioreactor.c[1]'])) \\
    V['S'] = np.linalg.norm(data['S'] - np.interp(data['time'], sim_res['time'], sim_res['bioreactor.c[2]']))
    return V['X'] + V['S']
```

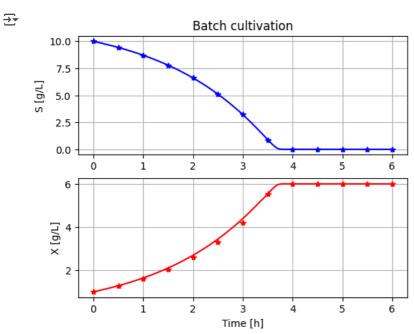
```
import time
```

The estimated parameters result.x are very close to the original values and no surprise.

4 Simulation with estimated parameters compared with data

```
newplot(plotType='Demo_1')
par(Y=result.x[0], qSmax=result.x[1], Ks=result.x[2])
simu(simulationTime)

# Show data
ax1.plot(data['time'], data['S'],'b*')
ax2.plot(data['time'], data['X'],'r*')
plt.show()
```



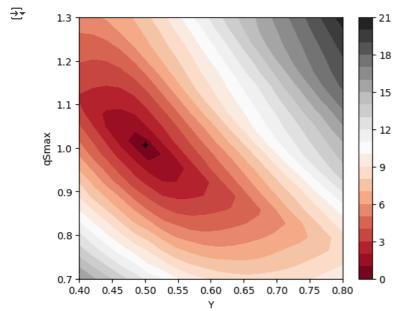
```
# The estimated parameters are for i in range(len(parEstim)): print(parEstim[i],':', result.x[i])

→ Y: 0.4999727558733863
qSmax: 1.0073152667279195
Ks: 0.14380564144282715
```

5 Analysis of the loss function

The problem is small and analysis of the loss function brings some insight. From the diagram above showing parameter sweep over combinations min- and max-parameters we see that the parameter K_s has little influence. Let use set that a fixed value and then plot the loss function in the parameters Y and qSmax. We do this by go through all the parametera combinations and evaluate each of them.

```
# Sweep through Y and qSmax variation and store the value of the loss-function for each
nY = 20
nqSmax = 20
V = np.zeros((nY, nqSmax))
Y = np.linspace(parBounds[0][0],parBounds[0][1],nY)
qSmax = np.linspace(parBounds[1][0],parBounds[1][1],nqSmax)
for j in range(nY):
    for k in range(nqSmax):
        V[k,j] = objective([Y[j], qSmax[k], 0.1], parEstim)
# Contour plot
plt.figure()
plt.clf
plt.subplot(1,1,1)
plt.contourf(Y, qSmax, V, 20, cmap='RdGy')
plt.plot(result.x[0], result.x[1],'k+')
plt.colorbar()
plt.ylabel('qSmax')
plt.xlabel('Y')
plt.show()
```



We see the following in the contour diagram of the loss function simplified:

- The minima is unique in the range of parmaters we study. This is good news.
- The contour plot is ellipsoid and rather narrow. The more narrow the ellipsoid the more difficult and more time it takes to converge to the minima.
- The direction of the ellipsoid axis indicate the correlation you may get between the two parameters during the minimization process.

Note that the form of the contour plot change with the parameters (and initial values) of the actual proces. You can see the impact by changing the parameters in "cell # 4" where data is generated and then just choose to run that cell and the cells below. No need to restart the notebook.

6 Summary

A choice was made to work with allowed ranges of parameters to be estimated and a start value was defined as the center point in this parameter space. There are only three methods available in optimize.minimize() that can handle bounds on parameters.

An evaluate() function was created that define how the difference beween simulation and data is measured. The function is rather transparent and easy to modify and you may want to change weight on the loss in S and X, for instance. Here they have so far equal weight.

The FMU-explore workspace dictionaries partDict[] and parLocation[] are useful also here and simplify the code for the evaluation() function. But we also use the detailed PyFMI-functions to administrate and set parameters of the actual simulation.

The call optimize.minimize() has several parameters and can easily be modified, for instance change of method. For fitting a model to data Nelder-Mead is ao a robust and good choice, but can be somewhat slow.

The estimated parameters were close to perfect!

The contour plot of the simplified loss function shows that the minima is unique and should not be difficult too difficult to obtain. More narrow elliptical contour plots would indicate difficulties. Multiple local minima would also be a problem.

7 References

[1] Scipy Reference guide on optimize.minimize() here

Start coding or generate with AI.

[2] Andersson, C., Åkesson, J., Fuhrer C.: "PyFMI: A Python package for simulation of coupled dynamic models with the functional mock-up interface", Centre for Mathematical Sciences, Lund University, Report LUTFNA-5008-2016, 2016.

Appendix

```
describe('parts')
['bioreactor', 'bioreactor.culture']
describe('MSL')
→ MSL: 3.2.3 – used components: none
system_info()
\overline{z}
    System information
     -OS: Linux
     -Python: 3.11.11
     -Scipy: 1.14.1
     -PyFMI: 2.16.3
     -FMU by: OpenModelica Compiler OpenModelica 1.25.0~dev-133-ga5470be
     -FMI: 2.0
     -Type: FMUModelME2
     -Name: BPL.Examples_TEST2.Batch
     -Generated: 2024-11-06T21:35:34Z
     -MSL: 3.2.3
     -Description: Bioprocess Library version 2.3.0
     -Interaction: FMU-explore version 1.0.0
```