# trackViewer

LOLLIPOP DANDELION PLOTS FOR METHYLATION STATUS AND MUTATION DATA

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#### **INSTALL THE WORKSHOP PKG**

https://github.com/jianhong/trackViewerBiocAsia2020Workshop https://jianhong.github.io/trackViewerBiocAsia2020Workshop/articles/ trackViewer.html

https://bioconductor.org/packages/trackViewer

https://www.nature.com/articles/s41592-019-0430-y

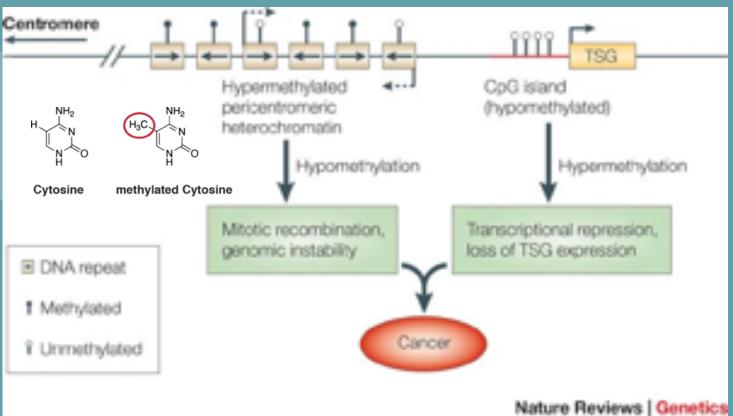
Slides:

https://github.com/jianhong/trackViewerBiocAsia2020Workshop/blob/master/inst/extdata/slides.pdf

verBiocAsia2020Workshop/articles/
trackViewer
-1592-019-0430-y
iewerBiocAsia2020Workshop/blob/

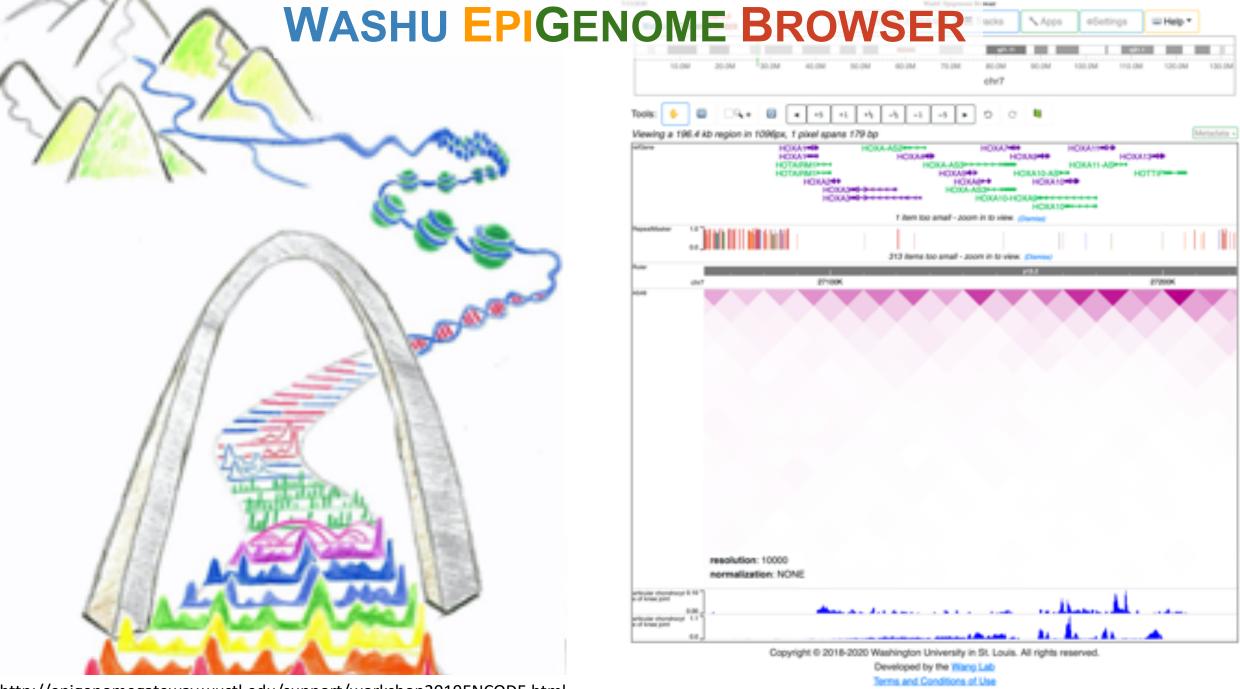


## DNA METHYLATION

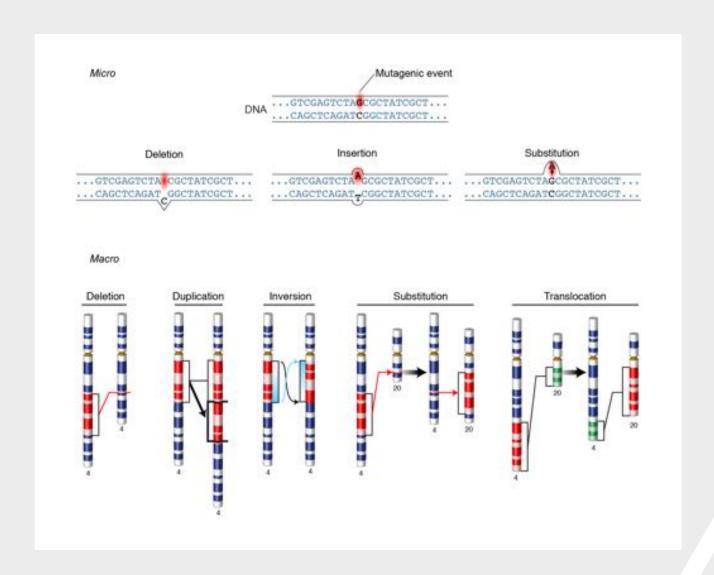


Robertson, K. 2005. doi: 10.1038/nrg1655 http://www.roadmapepigenomics.org/

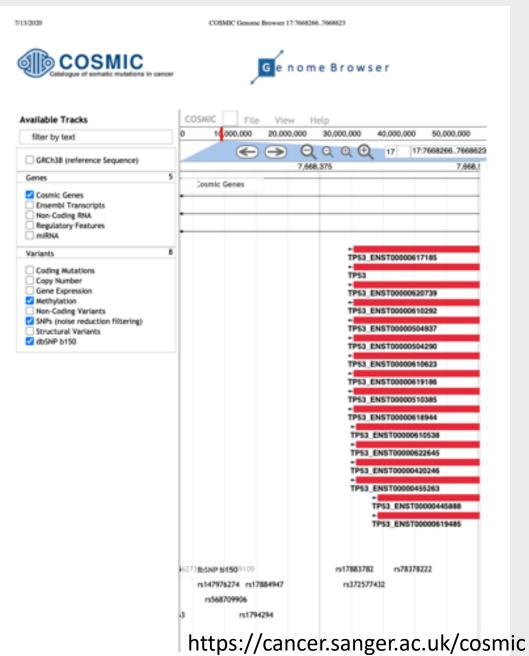




http://epigenomegateway.wustl.edu/support/workshop2019ENCODE.html



## MUTATION

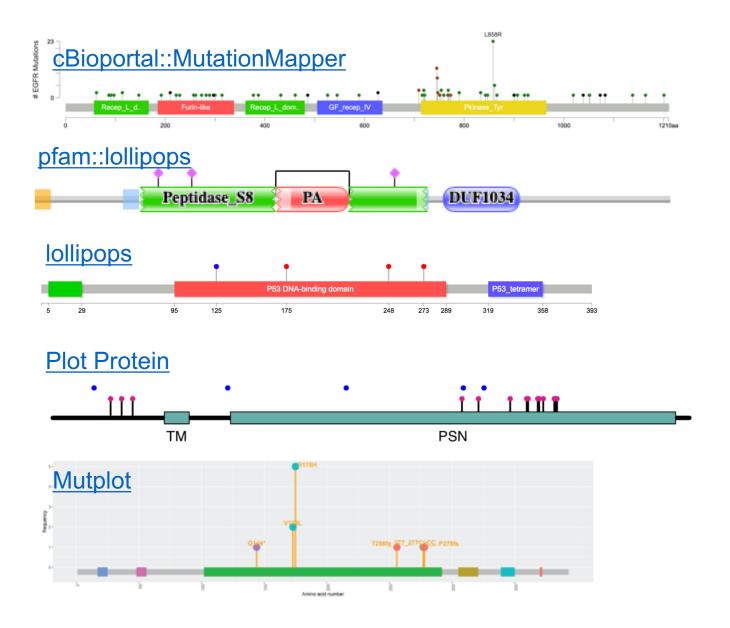




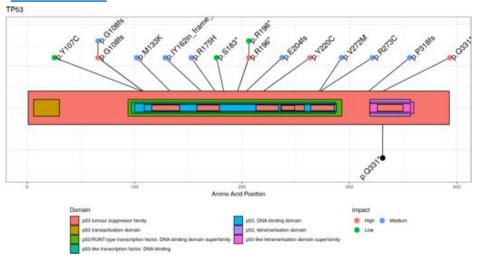
dbSNP contains human single nucleotide variations, microsatellites, and small-scale insertions and deletions along with publication, population frequency, molecular consequence, and genomic and RefSeq mapping information for both common variations and clinical mutations.

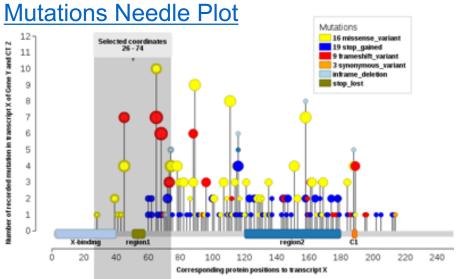
# GENOME BROWSERS AND VIEWERS

#### LOLLIPOP/NEEDLE PLOTS



#### •GenVisR





Jay et.al., 2016. doi: 10.1371/journal.pone.0160519

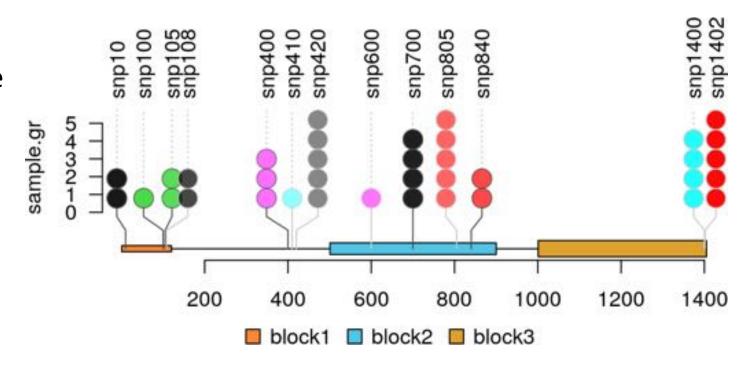
#### WHY trackViewer

#### pros:

- minimized: simple, clean design for the publication
- highly customizable: labels, symbols, colors, opacity and size
- powerful: handle high dense data

#### cons:

- learning curve
- no embedded database



## SNP: Single Nucleotide Poly INDEL: INsertion/DELetion

SNP: Single Nucleotide Polymorphism

Name	Input formats	Output formats	Called variants
Atlas-SNP2	BAM	VCF	SNPs
GATK (UnifiedGenotyper)	BAM	VCF	SNPs, InDels
SAMtools (samtools mpileup)	BAM	VCF	SNPs, InDels
SNVer	BAM	VCF	SNPs, InDels
SOAPsnp	SOAP	Text format, GLFv2	SNPs
Varscan2	Pileup/mpileup	Text format, VCF	SNPs, InDels, CNA

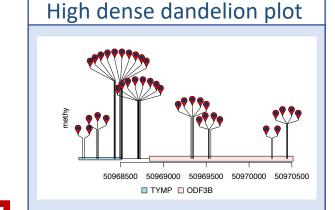
## VCF Variant Call Format

```
##fileformat=VCFv4.3
##fileDate=20090805
##source=myImputationProgramV3.1
##reference=file:///seq/references/1000GenomesPilot-NCBI36.fasta
##contig=<ID=20.length=62435964.assembly=B36.md5=f126cdf8a6e0c7f379d618ff66beb2da.species="Homo sapiens".taxonomy=x>
##phasing=partial
##INFO=<ID=NS, Number=1, Type=Integer, Description="Number of Samples With Data">
##INFO=<ID=DP, Number=1, Type=Integer, Description="Total Depth">
##INFO=<ID=AF, Number=A, Type=Float, Description="Allele Frequency">
##INFO=<ID=AA, Number=1, Type=String, Description="Ancestral Allele">
##INFO=<ID=DB, Number=0, Type=Flag, Description="dbSNP membership, build 129">
##INFO=<ID=H2, Number=0, Type=Flag, Description="HapMap2 membership">
##FILTER=<ID=q10, Description="Quality below 10">
##FILTER=<ID=s50, Description="Less than 50% of samples have data">
##FORMAT=<ID=GT, Number=1, Type=String, Description="Genotype">
##FORMAT=<ID=GO.Number=1.Type=Integer.Description="Genotype Quality">
##FORMAT=<ID=DP, Number=1, Type=Integer, Description="Read Depth">
##FORMAT=<ID=HQ, Number=2, Type=Integer, Description="Haplotype Quality">
#CHROM POS
                         REF ALT
                                    OUAL FILTER INFO
                                                                         FORMAT
                                                                                      NA00001
                                                                                                     NA00002
                                                                                                                    NA00003
20
       14370
              rs6054257 G
                           Α
                                    29 PASS
                                                NS=3;DP=14;AF=0.5;DB;H2 GT:GO:DP:H0 0|0:48:1:51.51 1|0:48:8:51.51 1/1:43:5:...
20
      17330
                                         q10
                                                 NS=3:DP=11:AF=0.017
                                                                         GT:GQ:DP:HQ 0|0:49:3:58.50 0|1:3:5:65.3
                                                                                                                    0/0:41:3
20
      1230237 .
                                    47
                                         PASS
                                                NS=3:DP=13:AA=T
                                                                         GT:GQ:DP:HQ 0|0:54:7:56,60 0|0:48:4:51,51 0/0:61:2
20
       1234567 microsat1 GTC G,GTCT 50
                                         PASS
                                                 NS=3:DP=9:AA=G
                                                                         GT:GQ:DP
                                                                                     0/1:35:4
                                                                                                     0/2:17:2
                                                                                                                    1/1:40:3
```

VCF/BED/BedGraph

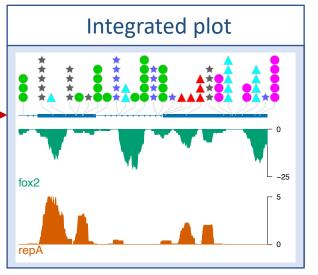
TxDb annotation package

# Low dense lollipop plot ■ TYMP ■ ODF3B

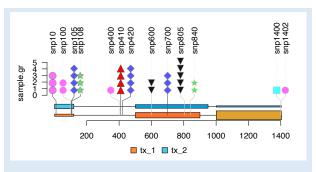


# Coverage plot

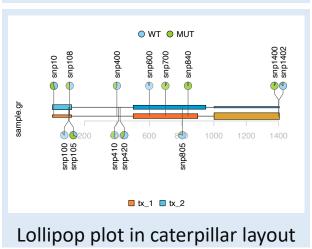


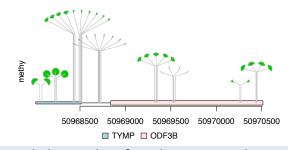


### **VISUALIZATION OF METHYLATION AND MUTATION** DATA IN DIFFERENT STYLES **USING** trackViewer

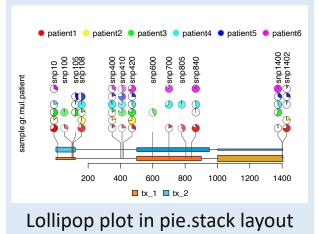


Lollipop plot with different symbols

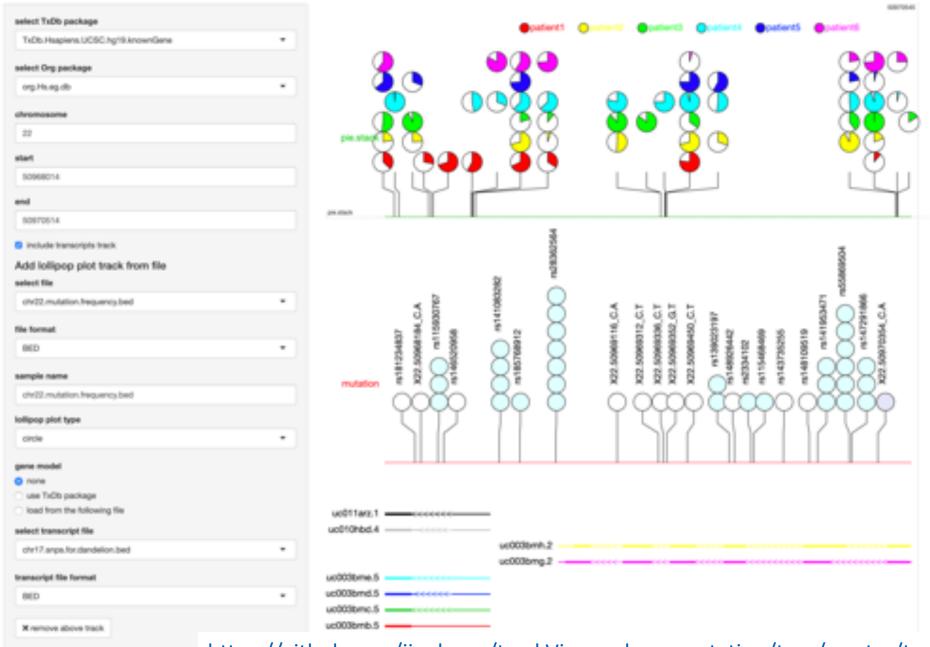




Dandelion plot for depicting dense methylation data



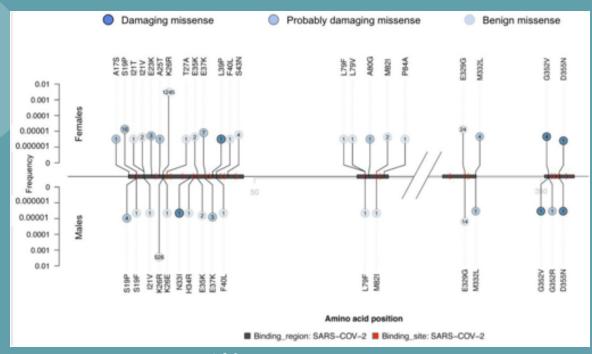
#### trackViewer



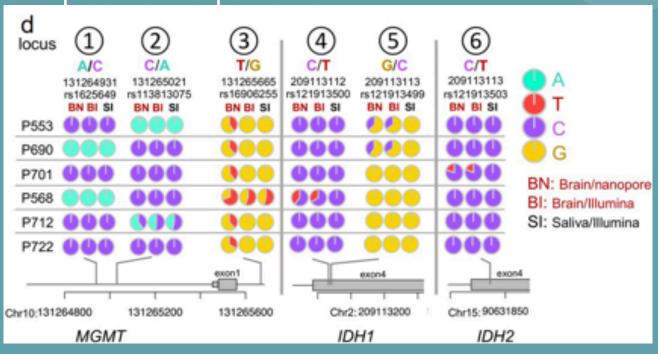
https://github.com/jianhong/trackViewer.documentation/tree/master/trackViewerShinyApp

#### EXAMPLES USING THE trackViewer PACKAGE

Lolliplot of missense variants in the regions of ACE2 that bind to SARS-CoV-2.



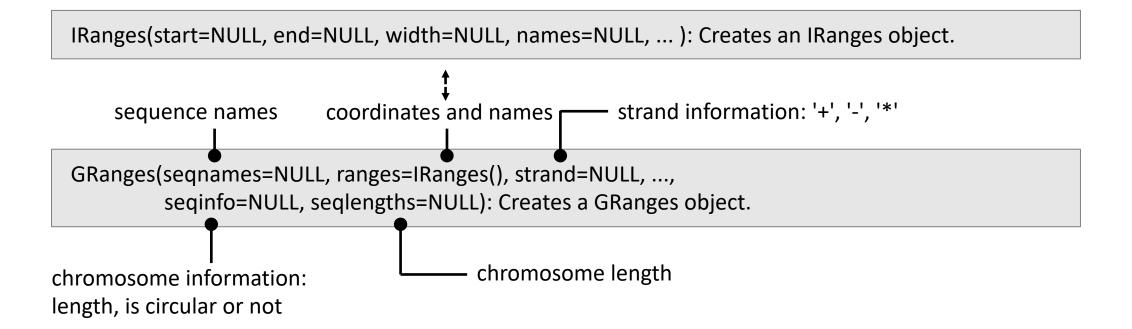
SNVs in MGMT and IDH1/2 in tumor and saliva samples from 6 patients



Cirulli et.al., 2020. doi: 10.1101/2020.04.07.030544

Wongsurawat et.al., 2020. doi: 10.1186/s40478-020-00963-0

## **GRanges Class**



## GOTO VIGNETTE

https://jianhong.github.io/trackViewerBiocAsia2020Workshop/

#### trackViewer CAN ...

generate lollipop plot to depict the methylation and SNP/mutation status

visualize regular read coverage tracks

be easily integrated into standard analysis pipeline for various high-throughput sequencing dataset such as ChIP-seq, RNA-seq, methylation-seq or DNA-seq

produce highly customized figure for publication



