CSS Seed Predator

Jenna Loesberg

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Data analysis for CSS paper:

Which sites to drop:

- sites where more than 5 seeds were missing from Exclusion dish
- \bullet sites where less than 10 or more than 70 seeds taken
- sites where seeds taken are less than 10 different from Exclusion

```
# Selecting exclusion dishes with 5 or less seeds missing, there are 94
exc <- seed %>%
   filter(Treatment == "C") #101 observations

exc <- exc %>%
   filter(Total_eaten <=5) #94 observations

# get unique ID for those sites:
exc_codes <- exc %>%
   select(GPS_code)
# add back the rest of the data
seed <- semi_join(seed, exc_codes) #seed 3 = without those Exc sites</pre>
```

Joining, by = "GPS_code"

```
# BR and A only dishes with seed eaten no. between 10 and 70

BR <- seed %>%
filter(Treatment == "BR+") %>% #88 dishes
filter(Total_eaten %in% (10:70)) #52 dishes

A <- seed %>%
filter(Treatment == "A+") %>% #90 dishes
filter(Total_eaten %in% (10:70)) #5 dishes

ABR <- rbind(BR, A)
# ABR: These are the sites we're keeping.

# do any of these sites have <10 difference with control?

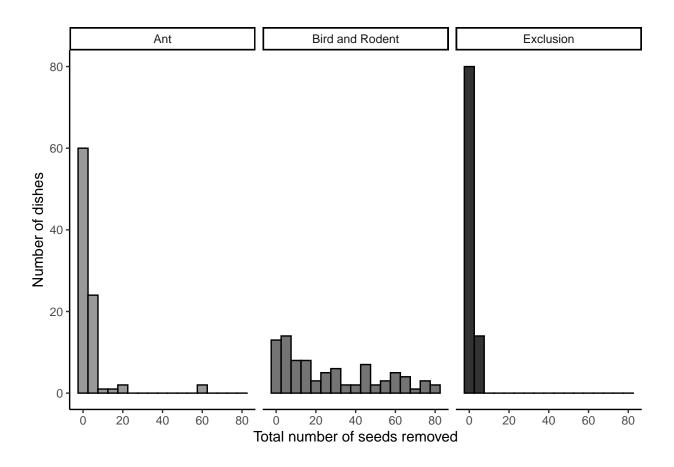
GPS <- ABR %>%
select(GPS_code) #these are the sites that we need control numbers for

GPS <- semi_join(exc, GPS) #control sites for each site that meet criteria
```

```
## Joining, by = "GPS_code"
```

```
ABR <- rbind(GPS, ABR)
ABR2 <- ABR
#one site has a too small seed difference between C and trt:
ABR <- ABR %>%
 filter(GPS_code != "JL_Feb15_S2") %>%
  arrange(GPS_code, Treatment)
# These are the sites that do not have camera data: let's exclude them from the analyses
ABR <- ABR %>%
 filter(GPS_code != "May22_S2") %>%
 filter(GPS_code != "Nov5_S4") %>%
 filter(GPS_code != "Oct10_S1") %>%
 filter(GPS_code != "Sep17_S4") %>%
 filter(GPS_code != "Ap23_S2") %>%
 filter(GPS_code != "Ap23_S4")
#ABR has all of the sites that we're including! Now getting just A and BR
Global_final <- ABR %>%
filter(Treatment != "C")
```

Creating histograms of seeds missing for appendix (before sites are cleaned for preference analysis):



#ggsave("figures/histogram.png", width = 8, height = 4)

creating Manly alpha indices:

```
Global_final["prey"] <- 20 #starting amount = 20</pre>
Global_final <- Global_final %>%
  mutate(salv.pro = salvia.left/prey, #proportion of starting amount
         enc.pro = encelia_left/prey,
         brom.pro = bromus left/prey,
         bras.pro = brassica_left/prey)
#add small vlaue to zeros to take log:
Global_final$salv.pro[Global_final$salv.pro == 0] <- 0.001</pre>
Global_final$enc.pro[Global_final$enc.pro == 0] <- 0.001</pre>
Global_final$brom.pro[Global_final$brom.pro == 0] <- 0.001</pre>
Global_final$bras.pro[Global_final$bras.pro == 0] <- 0.001</pre>
Global_final <- Global_final %>%
  mutate(salvia = log(salv.pro)/(log(salv.pro)+log(enc.pro)+log(brom.pro)+log(bras.pro)), #manly calc
         encelia = log(enc.pro)/(log(salv.pro)+log(enc.pro)+log(brom.pro)+log(bras.pro)),
         bromus = log(brom.pro)/(log(salv.pro)+log(enc.pro)+log(brom.pro)+log(bras.pro)),
         brassica = log(bras.pro)/(log(salv.pro)+log(enc.pro)+log(brom.pro)+log(bras.pro)))
#here I will split it up into A, BR and global
A_final <- Global_final %>%
```

```
filter(Treatment == "A+")
BR_final <- Global_final %>%
filter(Treatment == "BR+")
```

This is in wide format, need to switch to long

```
BR_final <- BR_final %>%
    select(GPS_code, salvia, encelia, bromus, brassica) %>%
    pivot_longer(cols = c(salvia:brassica), names_to = "species", values_to = "index")
A_final <- A_final %>%
    select(GPS_code, salvia, encelia, bromus, brassica) %>%
    pivot_longer(cols = c(salvia:brassica), names_to = "species", values_to = "index")
Global_final <- Global_final %>%
    select(Treatment, GPS_code, salvia, encelia, bromus, brassica) %>%
    pivot_longer(cols = c(salvia:brassica), names_to = "species", values_to = "index")
```

T-tests for each seed species. Are the true means different from 0.25?

```
bromus <- Global_final %>%
 filter(species == "bromus") %>%
  select(index)
t.test(bromus, mu = 0.25)
##
##
  One Sample t-test
## data: bromus
## t = -2.124, df = 49, p-value = 0.03874
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.1214085 0.2464405
## sample estimates:
## mean of x
## 0.1839245
brassica <- Global_final %>%
 filter(species == "brassica") %>%
  select(index)
t.test(brassica, mu = 0.25)
##
## One Sample t-test
##
## data: brassica
## t = 5.0737, df = 49, p-value = 6.005e-06
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.3560355 0.4951191
## sample estimates:
## mean of x
## 0.4255773
```

```
salvia <- Global_final %>%
  filter(species == "salvia") %>%
  select(index)
t.test(salvia, mu = 0.25)
## One Sample t-test
##
## data: salvia
## t = -0.14028, df = 49, p-value = 0.889
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.1943824 0.2983593
## sample estimates:
## mean of x
## 0.2463709
encelia <- Global_final %>%
 filter(species == "encelia") %>%
  select(index)
t.test(encelia, mu = 0.25)
##
## One Sample t-test
## data: encelia
## t = -5.7254, df = 49, p-value = 6.182e-07
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.1069666 0.1812880
## sample estimates:
## mean of x
## 0.1441273
Confidence intervals for graph:
BR_sum <- BR_final %>%
  group_by(species) %>%
  summarize(mean = mean(index), sd = sd(index), n = n(), se = sd/sqrt(n))
## `summarise()` ungrouping output (override with `.groups` argument)
Global_sum <- Global_final %>%
  group_by(species) %>%
  summarize(mean = mean(index),
            sd = sd(index),
            n = n(),
            se = sd/sqrt(n),
            lower.ci = mean-1.96*sd/sqrt(n),
            upper.ci = mean+1.96*sd/sqrt(n))
```

`summarise()` ungrouping output (override with `.groups` argument)

```
A_sum <- A_final %>%
group_by(species) %>%
summarize(mean = mean(index), sd = sd(index), n = n(), se = sd/sqrt(n))
```

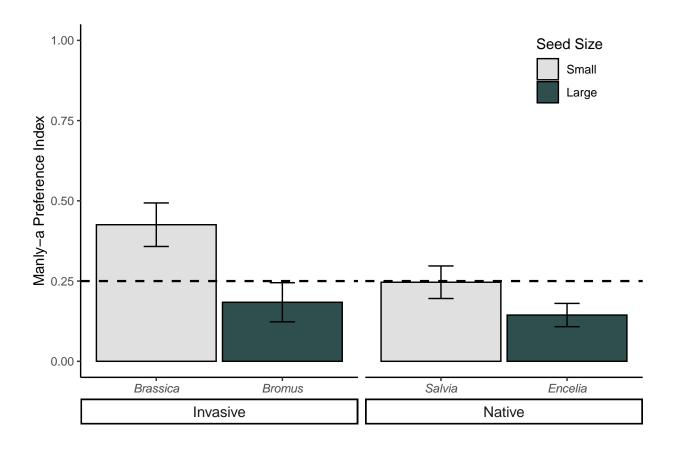
`summarise()` ungrouping output (override with `.groups` argument)

```
Global_sum <- left_join(Global_sum, info, by = "species")
Global_sum$species[Global_sum$species == "brassica"] <- "Brassica"
Global_sum$species[Global_sum$species == "bromus"] <- "Bromus"
Global_sum$species[Global_sum$species == "encelia"] <- "Encelia"
Global_sum$species[Global_sum$species == "salvia"] <- "Salvia"
```

Global plot:

```
Global sum %>%
  mutate(species = as.factor(species),
         size = as.factor(size)) %>%
  ggplot(aes(x = factor(species, levels = c("Brassica", "Bromus", "Salvia", "Encelia")), y = mean))+
  geom_col(aes(fill = size), width = .95, color = "black")+
  #geom_point(data = Global_final, aes(species, index))+
  scale_fill_manual(values=c("darkslategrey", "gray88"), guide = guide_legend(reverse=TRUE))+
  geom_hline(aes(linetype = "Preference Threshold"), yintercept=0.25,linetype="dashed", color = "black"
  \#scale\_linetype\_manual(name = "", values = c(1, 1), guide = guide\_legend(override.aes = "black")) +
  geom_errorbar(aes(ymin = lower.ci, ymax = upper.ci), width = 0.2)+
  expand_limits(y = c(0, 1))+
  labs(y= "Manly-\u03B1 Preference Index", x = "", fill = "Seed Size")+
  facet_grid(.~origin, scales = "free", switch = "x", space = "free")+
  theme(strip.placement = "outside",
         legend.position = c(0.87, 0.87),
         axis.text.x = element text(face = "italic"),
         strip.text.x = element_text(size = 11))
```

Warning: geom hline(): Ignoring `mapping` because `yintercept` was provided.



```
#ggsave("figures/GlobalManly.png", width = 7, height = 5)
# n = 50
```

Add in the granivore species information from the camera:

site_species <- site_species %>%
 filter(Sp_ate != "none") %>%
 mutate(Sp_ate = as.factor(Sp_ate),

```
site_species <- species %>%
    select(c(Run, GPS_code, Site, Sp_ate))

site_species <- left_join(Global_final, site_species, by = "GPS_code")
site_species$Sp_ate <- ifelse(site_species$Treatment == "A+", "ant", site_species$Sp_ate)
site_species <- left_join(site_species, info)

## Joining, by = "species"

species specific plot:

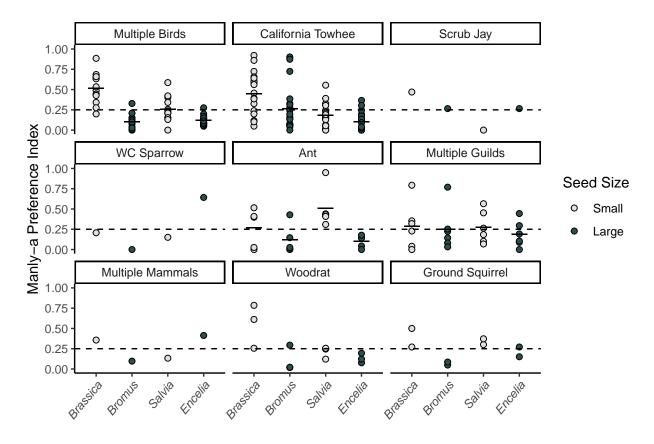
labels <- c(ant = "Ant", CA_Towee = "California Towhee", GS = "Ground Squirrel", mixed_bird = "Multiple")</pre>
```

```
species = as.factor(species),
    species = factor(species, levels = c("brassica", "bromus", "salvia", "encelia")),
    Sp_ate = factor(Sp_ate, levels = c("mixed_bird", "CA_Towee", "scrub_jay", "WC_sparrow", "ant",

species_stat <- site_species %>%
    group_by(species, Sp_ate) %>%
    summarize(mean = mean(index)) %>%
    summarize(mean = mean(index)) %>%
    filter(Sp_ate == "mixed_bird"| Sp_ate == "CA_Towee"|Sp_ate == "ant"| Sp_ate == "multipred" )
```

`summarise()` regrouping output by 'species' (override with `.groups` argument)

```
site_species %>%
  group_by(species) %>%
  ggplot(aes(species, index))+
  geom_point(aes(fill = size), shape = 21, size = 1.8)+
  geom_hline(yintercept=0.25,linetype="dashed", color = "black", size = .5)+
  expand_limits(y = c(0, 1))+
  facet_wrap(~Sp_ate, labeller = labeller(Sp_ate = labels), ncol = 3)+
  labs(y= "Manly-\u03B1 Preference Index", x = "", fill = "Seed Size")+
  theme(axis.text.x=element_text(angle=50, hjust=1)) +
  geom_point(data = species_stat, aes(y = mean, x = species), shape = 95, size = 7, color = "#0000000")+
  scale_x_discrete(labels = c("Brassica", "Bromus", "Salvia", "Encelia"))+ #just changes names
  theme(axis.text.x = element_text(face = "italic"))+
  scale_fill_manual(values=c("darkslategrey", "gray88"), guide = guide_legend(reverse=TRUE))
```



```
#ggsave("figures/specices_preference_1.png", width = 7, height = 6 )
```

One sample t-tests for all of those means:

• Multiple birds:

```
mult_birds_seeds <- site_species %>%
  filter(Sp_ate == "mixed_bird")
MBbird_bromus <- mult_birds_seeds %>%
  filter(species == "bromus") %>%
  select(index)
t.test(MBbird_bromus, mu = 0.25)
##
## One Sample t-test
##
## data: MBbird_bromus
## t = -5.2284, df = 11, p-value = 0.0002818
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.03816417 0.16367926
## sample estimates:
## mean of x
## 0.1009217
MBbird_brassica <- mult_birds_seeds %>%
  filter(species == "brassica") %>%
  select(index)
t.test(MBbird_brassica, mu = 0.25)
##
## One Sample t-test
## data: MBbird_brassica
## t = 4.7584, df = 11, p-value = 0.0005916
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.3941274 0.6422041
## sample estimates:
## mean of x
## 0.5181657
MBbird_salvia <- mult_birds_seeds %>%
  filter(species == "salvia") %>%
  select(index)
t.test(MBbird_salvia, mu = 0.25)
##
   One Sample t-test
##
```

```
## data: MBbird_salvia
## t = 0.18705, df = 11, p-value = 0.855
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.1584539 0.3585517
## sample estimates:
## mean of x
## 0.2585028
MBbird_encelia <- mult_birds_seeds %>%
  filter(species == "encelia") %>%
  select(index)
t.test(MBbird_encelia, mu = 0.25)
##
##
   One Sample t-test
##
## data: MBbird_encelia
## t = -6.5473, df = 11, p-value = 4.151e-05
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.07951814 0.16530134
## sample estimates:
## mean of x
## 0.1224097
  • towhee:
towhee_seeds <- site_species %>%
  filter(Sp_ate == "CA_Towee")
towhee_bromus <- towhee_seeds %>%
  filter(species == "bromus") %>%
  select(index)
t.test(towhee_bromus, mu = 0.25)
##
##
  One Sample t-test
##
## data: towhee_bromus
## t = 0.24616, df = 18, p-value = 0.8083
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.1323390 0.3988928
## sample estimates:
## mean of x
## 0.2656159
towhee_brassica <- towhee_seeds %>%
  filter(species == "brassica") %>%
  select(index)
t.test(towhee_brassica, mu = 0.25)
```

```
##
## One Sample t-test
##
## data: towhee_brassica
## t = 3.2774, df = 18, p-value = 0.004185
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.3212697 0.5758160
## sample estimates:
## mean of x
## 0.4485428
towhee_salvia <- towhee_seeds %>%
  filter(species == "salvia") %>%
  select(index)
t.test(towhee_salvia, mu = 0.25)
##
## One Sample t-test
##
## data: towhee_salvia
## t = -2.0414, df = 18, p-value = 0.05615
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.1127865 0.2519708
## sample estimates:
## mean of x
## 0.1823786
towhee_encelia <- towhee_seeds %>%
  filter(species == "encelia") %>%
  select(index)
t.test(towhee_encelia, mu = 0.25)
## One Sample t-test
## data: towhee_encelia
## t = -5.7936, df = 18, p-value = 1.725e-05
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.0503241 0.1566012
## sample estimates:
## mean of x
## 0.1034626
  • ant:
ant_seeds <- site_species %>%
  filter(Sp_ate == "ant")
ant_bromus <- ant_seeds %>%
```

```
filter(species == "bromus") %>%
  select(index)
t.test(ant_bromus, mu = 0.25)
##
##
   One Sample t-test
##
## data: ant_bromus
## t = -1.5835, df = 4, p-value = 0.1885
\#\# alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## -0.1043772 0.3469653
## sample estimates:
## mean of x
## 0.1212941
ant_brassica <- ant_seeds %>%
  filter(species == "brassica") %>%
  select(index)
t.test(ant_brassica, mu = 0.25)
##
##
  One Sample t-test
##
## data: ant_brassica
## t = 0.18353, df = 4, p-value = 0.8633
\mbox{\tt \#\#} alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## -0.02716817 0.56640434
## sample estimates:
## mean of x
## 0.2696181
ant_salvia <- ant_seeds %>%
  filter(species == "salvia") %>%
  select(index)
t.test(ant_salvia, mu = 0.25)
##
##
   One Sample t-test
##
## data: ant_salvia
## t = 2.2786, df = 4, p-value = 0.08492
\mbox{\tt \#\#} alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.1938395 0.8202618
## sample estimates:
## mean of x
## 0.5070507
```

```
ant_encelia <- ant_seeds %>%
  filter(species == "encelia") %>%
  select(index)
t.test(ant_encelia, mu = 0.25)
##
##
   One Sample t-test
##
## data: ant_encelia
## t = -4.3643, df = 4, p-value = 0.01202
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.007908171 0.196166167
## sample estimates:
## mean of x
## 0.1020372
  • Multiple taxa:
mult_taxa_seeds <- site_species %>%
  filter(Sp_ate == "multipred")
mult_taxa_bromus <- mult_taxa_seeds %>%
  filter(species == "bromus") %>%
  select(index)
t.test(mult_taxa_bromus, mu = 0.25)
##
##
   One Sample t-test
##
## data: mult_taxa_bromus
## t = 0.0038641, df = 5, p-value = 0.9971
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## -0.03014113 0.53098463
## sample estimates:
## mean of x
## 0.2504217
mult_taxa_brassica <- mult_taxa_seeds %>%
  filter(species == "brassica") %>%
  select(index)
t.test(mult_taxa_brassica, mu = 0.25)
##
##
    One Sample t-test
##
## data: mult_taxa_brassica
## t = 0.33556, df = 5, p-value = 0.7508
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
```

```
## -0.01083297 0.58915510
## sample estimates:
## mean of x
## 0.2891611
mult_taxa_salvia <- mult_taxa_seeds %>%
  filter(species == "salvia") %>%
  select(index)
t.test(mult_taxa_salvia, mu = 0.25)
##
##
   One Sample t-test
##
## data: mult_taxa_salvia
## t = 0.28074, df = 5, p-value = 0.7902
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.06467411 0.48076958
## sample estimates:
## mean of x
## 0.2727218
mult_taxa_encelia <- mult_taxa_seeds %>%
  filter(species == "encelia") %>%
  select(index)
t.test(mult_taxa_encelia, mu = 0.25)
##
## One Sample t-test
##
## data: mult_taxa_encelia
## t = -0.94961, df = 5, p-value = 0.3859
## alternative hypothesis: true mean is not equal to 0.25
## 95 percent confidence interval:
## 0.01903776 0.35635292
## sample estimates:
## mean of x
## 0.1876953
Now onto the community graph...
Do non-granivore species need to be taken out? Yep
# take out non-granivore species
species2 \leftarrow species[-c(5, 18, 21, 26, 34, 35, 36)]
Changing the dataframe into a useable format:
species2 <- species</pre>
species2[species2 == 0] <- NA</pre>
#species_n <- species2 %>%
```

select(-c(Site, Days_Out)) %>%

```
# summarize_all(n, na.rm = TRUE)
species3 <- species2 %>%
    summarize_if(is.numeric, sum, na.rm=TRUE) # add up all of the observations

species3 <- species3 %>%
    pivot_longer(cols = -c(Site, Days_Out), values_to = "number", names_to = c("species", "type"), names
species3[is.na(species3)] <- 0

species4 <- species3 %>%
    pivot_wider(names_from = "type", values_from = "number") %>%
    select(-c(Site, Days_Out))
species4[is.na(species4)] <- 0
species4 <- species4%>%
    group_by(species) %>%
    mutate(ratio = Eat/(Visit+Eat))
```

Need number of sites each each was observed at:

```
table <- read.csv("C:/Users/Jenna/Dropbox/Jenna/seed predator/CSS_SeedPredator/species_table.csv")
```

Formatted table:

```
table <- table %>% arrange(taxa, SciName) %>% select(-taxa)
# make all column integers so they are centered in table:
#table$Eat <- as.numeric(table$Eat)
#table$Visit <- as.numeric(table$Visit)
#table$SitesPresent <- as.numeric(table$SitesPresent)

tbl <- gt(data = table)
tbl</pre>
```

SciName	CommonName	Eat	Visit	ratio	SitesPresent
Aphelocoma californica	Scrub Jay	11	16	0.40740741	15
Callipepla californica	Quail	0	21	0.00000000	9
Catharus ustulatus	Swansons Thrush	0	29	0.00000000	3
Haemorhous mexicanus	House Finch		4	0.00000000	2
Melozone crissalis	California Towhee	173	204	0.45888594	68
Mimus polyglottos	Mocking Bird	0	1	0.00000000	1
Pipilo maculatus	Spotted Towhee	1	7	0.12500000	7
Setophaga coronata	Yellow Rumped Warbler	1	0	1.00000000	1
Toxostoma redivivum	California Thrasher	17	58	0.22666667	34
Zenaida macroura	Morning Dove	0	2	0.00000000	2
Zonotrichia leucophrys	White Crowned Sparrow	34	120	0.22077922	22
NA	Unidentified Bird	0	3	0.00000000	3
Sylivagus audubonii	Desert Cottontail	7	373	0.01842105	66
Dipodomys californicus	Kangaroo Rat	6	62	0.08823529	17
Neotoma spp.	Wood Rat	27	183	0.12857143	29
Otospermophilus beecheyi	Ground Squirrel	9	246	0.03529412	9
Peromyscus spp.	Deer Mouse	3	31	0.08823529	14
NA	Unidentified Rat	0	3	0.00000000	2

```
tbl1 <- tbl %>%
  tab_row_group(group = "Bird", rows = 1:12) %>%
  tab_row_group(group = "Rodent", rows = 14:18) %>%
  tab_row_group(group = "Lagomorph", rows = 13) %>%
  row_group_order(groups = c("Bird", "Rodent", "Lagomorph")) %>%
  tab_stubhead(label = "Group") %>%
  tab_spanner(label = "Type of Observtion", columns = vars(Eat, Visit)) %>%
  fmt_number(column = vars(ratio), decimals = 2) %>%
  cols_label(SciName = "Scientific Name", CommonName = "Common Name", Eat = "Seed Predation", ratio = ":
  tab_style(style = cell_text(style = "italic"), locations = cells_body(columns = vars(SciName))) %>%
  fmt_missing(columns = 1:2, missing_text = "---") %>%
  cols_align(align = "center") %>%
  cols_width(everything() ~ px(90)) %>%
  cols_width(ends_with("Name") ~ px(150))
```

Warning in min(rows_matched): no non-missing arguments to min; returning Inf

Warning in max(rows_matched): no non-missing arguments to max; returning -Inf

		Type of Observtion		
Scientific Name	Common Name	Seed Predation	Visit	Ratio of predation events to total events
Bird				
Aphelocoma californica	Scrub Jay	11	16	0.41
Callipepla californica	Quail	0	21	0.00
Catharus ustulatus	Swansons Thrush	0	29	0.00
Haemorhous mexicanus	House Finch	0	4	0.00
Melozone crissalis	California Towhee	173	204	0.46
Mimus polyglottos	Mocking Bird	0	1	0.00
Pipilo maculatus	Spotted Towhee	1	7	0.12
Setophaga coronata	Yellow Rumped Warbler	1	0	1.00
Toxostoma redivivum	California Thrasher	17	58	0.23
Zenaida macroura	Morning Dove	0	2	0.00
Zonotrichia leucophrys	White Crowned Sparrow	34	120	0.22
_	Unidentified Bird	0	3	0.00
Rodent				
Dipodomys californicus	Kangaroo Rat	6	62	0.09
Neotoma spp.	Wood Rat	27	183	0.13
Otospermophilus beecheyi	Ground Squirrel	9	246	0.04
Peromyscus spp.	Deer Mouse	3	31	0.09
_	Unidentified Rat	0	3	0.00
Lagomorph			-	
Sylivagus audubonii	Desert Cottontail	7	373	0.02

#tbl1 %>% gtsave("figures/fig3_Feb18.png")