Context-dependent host-microbe interactions in stochastic environments

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This manuscript was compiled on June 24, 2022

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Along with increases in average temperatures, global climate change is driving increases in the variability of precipitation events, temperature extremes, and droughts (1–3). Thus discerning the effects of variability on population dynamics and interactions between species is pivotal to forecasting the future of ecological systems. Classic ecological theory predicts that long-term population growth rates will be reduced by environmental variability (4, 5). This stochastic variability means that populations can expect to experience both good and bad years. The long-term stochastic growth rate (λ_s), which is the long-run geometric mean of annual growth rates, captures this varability; the geometric mean will always be less than expected from the mean growth rate alone.

Following Lewontin and Cohen (4), λ_s can be approximated as:

$$log(\lambda_s) \approx log(\overline{\lambda}) - \frac{\sigma^2}{2\overline{\lambda}^2} \qquad [1]$$

Where $\overline{\lambda}$ is the mean of annual population growth rates (λ_t) and σ^2 is the variance (4). Populations will increase over time if λ_s is greater than 1, and can be expected to decrease if λ_s is less than 1. Here, there are two pathways to increase λ_s: (1) increasing the mean growth rate, and/or (2) reducing the variance in growth rates. That both mean and variance can determine fitness underlies understanding of which aspects of a species' life history influence its success (6) and has important implications for population viability analysis (7).

Buffering population growth from variance is

Reducing variance and buffering populations from variability is a potentially important mechanisms determining fitness. Anything that limits the negative effects of bad years, while being neutral or costly in good years has the potential to decrease the impact of interannual environmental variability on population dynamics because it would limit variance. * Whether species interactions contribute to variance buffering† is an underexplored question (8).

Host-associated microbes are ubiquitous in nature. Across a broad range of taxa‡, microbial symbionts provide their hosts with protection from environmental stresses including drought, extreme temperatures, and enemies (9). Commonly, the benefits from these symbioses are context-dependent where the magnitude of interaction benefit depends on environmental conditions (10). This can make it difficult to quantify the net effect of a given interaction, but it also allows for the possibility that interaction strength can vary through time (cite). Symbionts may provide benefits under harsh conditions when they are needed by their hosts, but be neutral or even costly under benign conditions (cite). Over time, this may lead symbiont-associated organisms to experience a reduction in variation in vital rates by reducing the frequency of extreme years (conceptual figure). Embracing context-dependence in this way, we reveal a novel mechanism by which symbionts can act as mutualists that may come to be of increasing importance in a more variable future.

Using long-term data from experimental grass-fungal endophyte plots§, we test the hypothesis that context-dependent benefits of microbial symbionts buffer hosts from the fitness consequences of environmental variability. Specifically, we ask first how fungal endophytes influence the mean and interannual variance of their hosts' vital rates; next, we ask if these vital rate effects buffer variance in fitness and, if so, what is the relative importance of variance buffering vs. mean effects in the overall fitness impact of the symbiosis. With 14 years of demographic data, we employ structured, stochastic population models for seven species of cool-season grass hosts that are commonly infected with fungal endophytes¶ (*Lolium arundinaceum*, *Festuca subverticillata*, *Elymus virginicus*, and *Elymus villosus*, *Poa alsodes* and *Poa sylvestris*). These long-term data, in which each annual census is a sample of weather variation, allow us to construct a climate-explicit population models, which we use to evaluate the importance of buffering under forecasted changes in the mean and variance of climate drivers.

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§ *This needs context. No one will know what this means.*

¶ *I think we need a more thorough description of the experiment here - it's a novel experiment, at least in temporal scale, so we will want to sell it.*

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* *Not sure this is helpful. Obviously, "Anything" is mutualism, but if you don't know that yet then "Anything" will be confusing. But I do think you need some bridge to symbiosis.*

† *This should be defined.*

‡ *You only cite one paper on plant-fungal interactions – so not very convincing as a broad range of taxa*

Please provide details of author contributions here.

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Across species, we find that variance buffering by endophytes contributes (percentage) to population growth rates. While the effect is generally weaker than effects on the mean, we found that buffering was common in the most sensitive vital rates, and was most important for xxx species with xxx life history.

Results

Discussion

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Table 1. Comparison of the fitted potential energy surfaces and ab initio benchmark electronic energy calculations

Species	CBS	CV	G3
1. Acetaldehyde	0.0	0.0	0.0
2. Vinyl alcohol	9.1	9.6	13.5
3. Hydroxyethylidene	50.8	51.2	54.0

nomenclature for the TSs refers to the numbered species in the table.

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$$(x+y)^3 = (x+y)(x+y)^2$$
$$= (x+y)(x^2 + 2xy + y^2) \qquad [2]$$
$$= x^3 + 3x^2y + 3xy^3 + x^3.$$

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Materials and Methods

Natural history of grass-endophyte symbiosis. To study the effects of context-dependent microbial symbiosis, we focused on *Epichloë* fungal endophytes, which live in the aboveground tissue of many species of cool-season grasses and grow into their hosts' seeds where they can be transmitted to the next generation. This vertical transmission couples host and symbiont fitness and leads to the expectation that the interaction be mutualistic, else the fungi cause their host to be selected out of the population (11–13). While there are demonstrated benefits against herbivory(14) and under drought stress (15) in some species, these interactions are commonly context-dependent (16, 17).

Plant propagation and endophyte removal. Seeds from naturally infected populations of seven species of cool-season grasses (Agrostis perennans, Elymus villosus, Elymus virginicus, Festuca subverticillata, Lolium arundinaceum, Poa alsodes, and Poa sylvestris[**]) were collected in the Spring of 2006??????[††] from Lilly Dickie Woods (Lat,Lon) and Bayles Road (Lat,Lon) in Brown Co. IN. Seeds with shared maternal ancestry were either experimentally disinfected by heat treatments[‡‡] or left naturally infected to reduce confounding genotype effects. Seeds were surface sterilized with XXXX and cold stratified for XXXX weeks, then germinated in a growth chamber for XXXX weeks. Seedlings were then transferred to the greenhouse at Indiana University and allowed to grow for XXXX weeks. We confirmed the endophyte status of these plants using leaf peels, where tissue from the leaf sheet is stained with aniline blue lactic acid and examined for the presence of fungal hyphae (18), and

vegetatively propogated clones of similar sizes from the plants[§§]. These clones were used to establish the experimental plots, and cloning reduces the potential for negative effects of heat treatments (cite).

Experimental design and data collection. In 2007[¶¶], we established 10 3x3 plots for *Lolium arundinaceum, Festuca subverticillata, Elymus virginicus*, and *Elymus villosus* and 18 plots for *Poa alsodes* and *Poa sylvestris*. For each species, an equal number of plots were randomly assigned to each endophyte status, E+ or E-, and was planted with only symbiotic or symbiont-free plants respectively. Each plot was planted with 25?[***] evenly spaced individuals and each plant marked with aluminum tags.

In each Summer starting in 2007, we censused all original transplants and any recruits for survival, growth and reproduction. After clearing out leaf litter, for each plant alive in the previous year, we marked its survival. We measured the size of each plant as a count of the number of tillers. Further, we collected reproductive data by counting the number of reproductive tillers, and then counting the number of seed-bearing spikelets on up to three of those reproductive tillers. In XXXX year, we took additional counts of seeds per inflorescence (list of species) or seeds per spikelet (list of species). Together, we use these measurements to estimate seed production. In each year, we also survey for and mark any unmarked individuals. New recruits are typically a size of one tiller and non-reproductive, but we also find and mark any individuals who may have been missed in previous censuses.

We typically expect plots of each endophyte status to maintain their status as the fungus is almost entirely vertically transmitted, and plots are spaced at least XX m apart, limiting the possibility for unwanted dispersal between plots or horizontal transmission of the fungus. Seeds from reproductive individuals are opportunistically taken and scored for their endophyte status. These scores reflect a XXXX% faithfullness of recruits to their expected endophyte status (Supplement data)[†††]

In sum, this individual-level demographic dataset covers 14 years and contains 30,XXXX individual transition-years.[‡‡‡]

Demographic modeling. Armed with this demographic data, we next constructed size-structured, stochastic population models. This demographic model describes transitions between sizes (measured as a count of tillers) from one year to the next. For all species, we include a 1 year reproductive delay in the population model following the observation that these newly recruited plants are rarely observed flowering in their first year.

$$\mathbf{n}_{t+1} = \mathbf{A}\mathbf{n}_t \qquad [3]$$

where $\mathbf{n}_{t+1}$ is a vector of abundances across sizes in year t+1 for each species and endophyte status.

$$\mathbf{n}_{t+1} = \begin{bmatrix} size_i \\ . \\ . \\ . \\ size_N \end{bmatrix} \qquad [4]$$

[**] *Italicize species names.*

[††] *check with Jenn*

[‡‡] *need methods for temp, duration, etc.*

[§§] *not sure this happened*

[¶¶] *dates?*

[***] *I think 20, check data.*

[†††] *We had these data in the original LTREB proposal.*

[‡‡‡] *Move this earlier and also mention at the end of intro. The total number of plant-years is impressive.*

and **A** is expressed as a N+1 x N+1 matrix:

$$\mathbf{A} = \begin{bmatrix} size_{sldg} & \cdot & \cdot & \cdot & size_{sldg_N} \\ \cdot & \cdot & \cdot & \cdot & \cdot \\ \cdot & \cdot & \cdot & \cdot & \cdot \\ \cdot & \cdot & \cdot & \cdot & \cdot \\ \cdot & \cdot & \cdot & \cdot & size_N \end{bmatrix} \quad [5]$$

Model description and estimation. We modeled the effect of endophyte symbiosis on mean and variance of vital rates by fitting generalized linear mixed models (GLMM) to the long-term data with year and plot random effects. We fit all vital rate models in a hierarchical Bayesian framework using Rstan (**?**), allowing us to propagate uncertainty from the vital rate estimates to our population model [19].

The probability of survival and flowering are recorded as successes or failures and consequently are modeled as Bernoulli processes. We modeled growth (measured as the number of tillers in year t+1), the number of flowering tillers, and the number of spikelets per inflorescence with the zero-truncated Poisson-Inverse Gaussian distribution. Where possible, each of these size dependent vital rates are modeled with the same structure of linear predictor (μ)

For example, growth ($G_{i,t1}$) of a given individual (i) in year t+1 is modeled as:

$$G_{i,t1} \sim P(IG(\mu_{s,e}, \lambda_{s,e})) \quad [6]$$

Similarly, survival $S_{i,t1}$ in year t+1 is modeled as:

$$S_{i,t1} \sim Bernoulli(\mu_{s,e}) \quad [7]$$

Where μ, for each species (s), is a linear function of the logarithm of plant size in year t(t), the plot level endophyte status (e), the status as an initial experimental transplant plant(r), along with random effects to account for plot(p), and year random effects specific to each species and endophyte status. Thus μ can be written:

$$\mu_{s,e} = \beta_s^1 + \beta_s^2 log(size_t) + \beta_{s,e}^3 + \beta_r^4$$
$$+\tau + \rho$$
$$\tau \sim N(0, \sigma_{s,e}) \quad [8]$$
$$\rho \sim N(0, \sigma_p)$$

For all species, we account for a reproductive delay by modeling seedling growth and survival separately from adult growth and survival. Seedlings are those plants that are recruited into the plot in a given year, and typically have only one tiller. This means that they do not have size dependence. So, for seedlings, growth ($G_{1,t1}^{sdlg}$) is modelled as:

$$G_{1,t1}^{sdlg} \sim P(IG(\mu_{s,e}^{sdlg}, \lambda_{s,e})) \quad [9]$$

Similarly, survival ($S_{i,t1}^{sdlg}$) in year t+1 is modeled as:

$$S_{i,t1}^{sdlg} \sim Bernoulli(\mu_{s,e}^{sdlg}) \quad [10]$$

Here, $\mu_{s,e}^{sdlg}$ is the linear function for these seedling specific vital rates. It does not include size-dependence or an effect to account for the initial benefits of greenhouse rearing.

$$\mu_{s,e}^{sdlg} = \beta_s^1 + \beta_{s,e}^3$$
$$+\tau + \rho$$
$$\tau \sim N(0, \sigma_{s,e}) \quad [11]$$
$$\rho \sim N(0, \sigma_p)$$

As a default, we ran each vital rate model for 2500 warm-up and 2500 MCMC sampling iterations with three chains using rStan. We assessed model convergence with trace plots of posterior chains and and other model diagnostic statistics (). For those models that show poor convergence, we extend the MCMC sampling to include 5000 warm-up and 5000 sampliing iterations; this was only necessary for seedling growth in our climate-implicit analysis. For each of these vital rate models, we graphically check model fit with posterior predictive checks using simulated data from 500 posterior draws and comparing with the observed data (See supplement?). These checks provide evidence that our models are accurately recreating size-specific growth, survival, and reproduction patterns in our data across endophyte treatments.

We calculate the effect of endophytes on mean and variance in growth rates by assembliing matrix models following equation [3] with and without endophyte effects for each year based on 500 posterior draws from our vital rate models.

Stochastic growth rate simulation experiment. We decompose the contribution of endophyte to long-termThe stochastic growth rate is approximated as the long-run expectation of annual growth rates [20].

$$log(\lambda_s) \ E[log(\Sigma(\mathbf{n}_{t+1})/\Sigma(\mathbf{n}_t))] \quad [12]$$

Estimating climate drivers of environmental context-dependence.

Climate data.

Climate-explicit Model description and estimation.

Climate-explicit Model assessment.

Forecasting under alternative climate forcings. We used statistics

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