

ReCentering Psych Stats: Analysis of Variance

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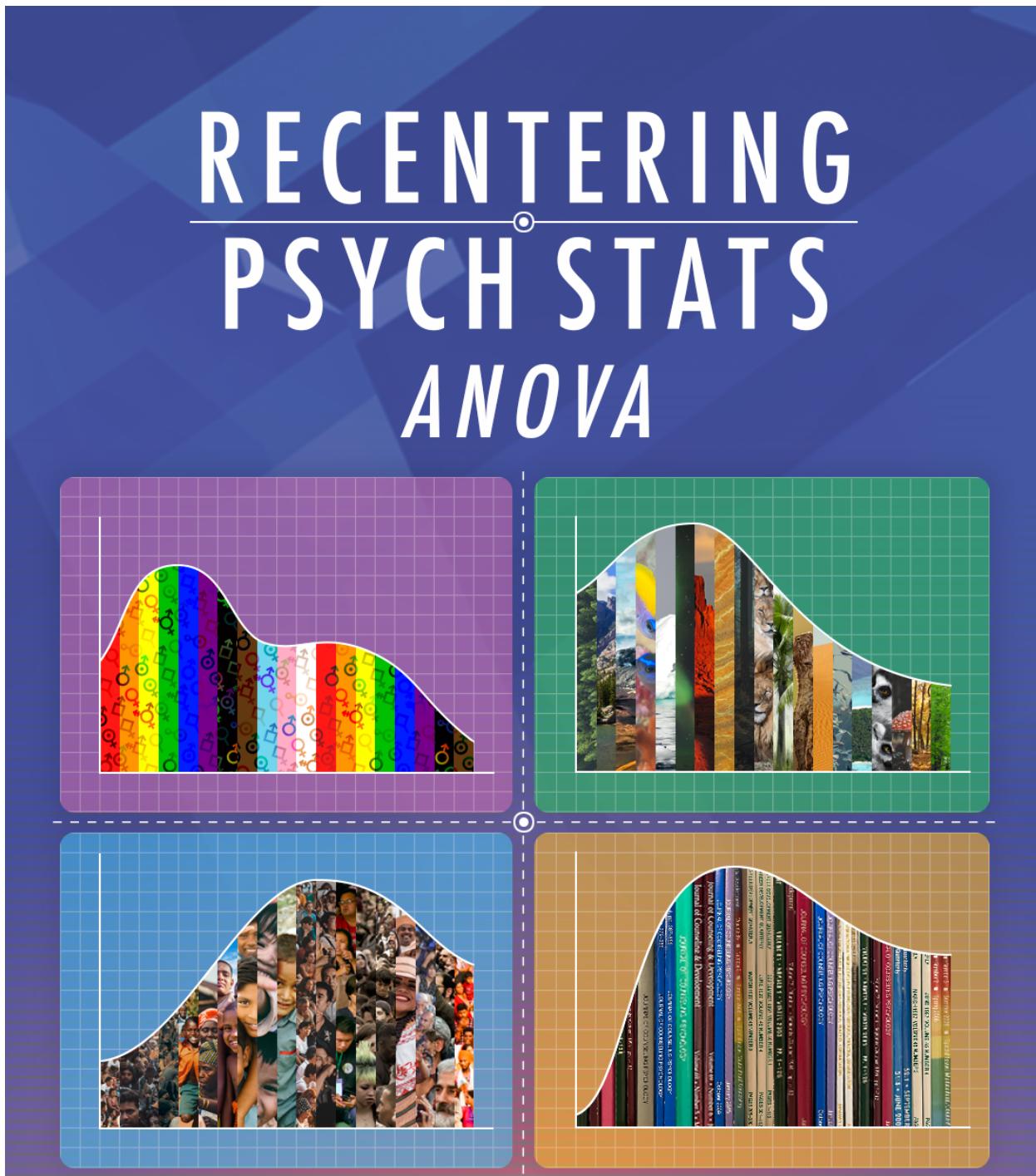
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BOOK COVER



- Formatted as an [html book](#) via GitHub Pages
- As a [PDF](#) available in the [docs](#) folder at the GitHub repository
- As an [ebook](#) available in the [docs](#) folder at the GitHub repository

All materials used in creating this OER are available at its [GitHub repo](#).

As a perpetually-in-progress, open education resource, feedback is always welcome. This IRB-approved (SPU IRB #202102010R, no expiration) [Qualtrics-hosted survey](#) includes formal rating scales, open-ended text boxes, and a portal for uploading attachments (e.g., marked up PDFs). You are welcome to complete only the portions that are relevant to you.

PREFACE

If you are viewing this document, you should know that this is a book-in-progress. Early drafts are released for the purpose teaching my classes and gaining formative feedback from a host of stakeholders. The document was last updated on 14 Jul 2022. Emerging volumes on other statistics are posted on the [ReCentering Psych Stats](#) page at my research team's website.

[Screencasted Lecture Link](#)

To *center* a variable in regression means to set its value at zero and interpret all other values in relation to this reference point. Regarding race and gender, researchers often center male and White at zero. Further, it is typical that research vignettes in statistics textbooks are similarly seated in a White, Western (frequently U.S.), heteronormative, framework. The purpose of this project is to create a set of open educational resources (OER) appropriate for doctoral and post-doctoral training that contribute to a socially responsive pedagogy – that is, it contributes to justice, equity, diversity, and inclusion.

Statistics training in doctoral programs are frequently taught with fee-for-use programs (e.g., SPSS/AMOS, SAS, MPlus) that may not be readily available to the post-doctoral professional. In recent years, there has been an increase and improvement in R packages (e.g., *psych*, *lavaan*) used for analyses common to psychological research. Correspondingly, many graduate programs are transitioning to statistics training in R (free and open source). This is a challenge for post-doctoral psychologists who were trained with other software. This OER will offer statistics training with R and be freely available (specifically in a GitHub repository and posted through GitHub Pages) under a Creative Commons Attribution - Non Commercial - Share Alike license [CC BY-NC-SA 4.0].

Training models for doctoral programs in health service psychology are commonly scholar-practitioner, scientist-practitioner, or clinical-scientist. An emerging model, the *scientist-practitioner-advocacy* training model, incorporates social justice advocacy so that graduates are equipped to recognize and address the sociocultural context of oppression and unjust distribution of resources and opportunities [[Mallinckrodt et al., 2014](#)]. In statistics textbooks, the use of research vignettes engages the learner around a tangible scenario for identifying independent variables, dependent variables, covariates, and potential mechanisms of change. Many students recall examples in Field's [[2012](#)] popular statistics text: Viagra to teach one-way ANOVA, beer goggles for two-way ANOVA, and bushtucker for repeated measures. What if the research vignettes were more socially responsive?

In this OER, research vignettes will be from recently published articles where:

- the author's identity is from a group where scholarship is historically marginalized (e.g.,

- BIPOC, LGBTQ+, LMIC[low-middle income countries]),
- the research is responsive to issues of justice, equity, inclusion, diversity,
 - the lesson's statistic is used in the article, and
 - there is sufficient information in the article to simulate the data for the chapter example(s) and practice problem(s); or it is publicly available.

In training for multicultural competence, the saying, “A fish doesn’t know that it’s wet” is often used to convey the notion that we are often unaware of our own cultural characteristics. In recent months and years, there has been an increased awakening to institutional and systemic factors that contribute to discrimination as a function of race, gender, nationality, class, and so forth. Queuing from the water metaphor, I am hopeful that a text that is recentered in the ways I have described can contribute to *changing the water* in higher education and in the profession of psychology.

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A [GitHub open-source repository](#) contains all of the text and source code for the book, including data and images.

ACKNOWLEDGEMENTS

As a doctoral student at the University of Kansas (1992-1996), I learned that “a foreign language” was a graduation requirement. *Please note that as one who studies the intersections of global, vocational, and sustainable psychology, I regret that I do not have language skills beyond English.* This could have been met with credit from high school, but my rural, mid-Missouri high school did not offer such classes. This requirement would have typically been met with courses taken during an undergraduate program – but my non-teaching degree in the University of Missouri’s School of Education was exempt from this. The requirement could have also been met with a computer language (FORTRAN, C++) – but I did not have any of those either. There was a tiny footnote on my doctoral degree plan that indicated that a 2-credit course, “SPSS for Windows” would substitute for the language requirement. Given that it was taught by my one of my favorite professors, I readily signed up. As it turns out, Samuel B. Green, PhD, was using the course to draft chapters in the textbook [[Green and Salkind, 2014b](#)] that has been so helpful for so many. Unfortunately, Drs. Green (1947 - 2018) and Salkind (1947 - 2017) are no longer with us. I have worn out numerous versions of their text. Another favorite text of mine has been Dr. Barbara Byrne’s [[2016](#)], “Structural Equation Modeling with AMOS.” I loved the way she worked through each problem and paired it with a published journal article, so that the user could see how the statistical evaluation fit within the larger project/article. I took my tea-stained text with me to a workshop she taught at APA and was proud of the signature she added to it. Dr. Byrne created SEM texts for a number of statistical programs (e.g., LISREL, EQS, MPlus). As I was learning R, I wrote Dr. Byrne, asking if she had an edition teaching SEM/CFA with R. She promptly wrote back, saying that she did not have the bandwidth to learn a new statistics package. We lost Dr. Byrne in December 2020. I am so grateful to these role models for their contributions to my statistical training. I am also grateful for the doctoral students who have taken my courses and are continuing to provide input for how to improve the materials.

The inspiration for training materials that re*center statistics and research methods came from the [Academics for Black Survival and Wellness Initiative](#). This project, co-founded by Della V. Mosley, Ph.D., and Pearis L. Bellamy, M.S., made clear the necessity and urgency for change in higher education and the profession of psychology.

At very practical levels, I am indebted to SPU’s Library, and more specifically, SPU’s Education, Technology, and Media Department. Assistant Dean for Instructional Design and Emerging Technologies, R. John Robertson, MSc, MCS, has offered unlimited consultation, support, and connection. Senior Instructional Designer in Graphics & Illustrations, Dominic Wilkinson, designed the logo and bookcover. Psychology and Scholarly Communications Librarian, Kristin Hoffman, MLIS, has provided consultation on topics ranging from OERS to citations. I am also indebted to Associate Vice President, Teaching and Learning at Kwantlen Polytechnic University, Rajiv Jhangiani, PhD. Dr. Jhangiani’s text [[2019](#)] was the first OER I ever used and I was grateful for

his encouraging conversation.

Financial support for this project has been provided the following:

- *Call to Action on Equity, Inclusion, Diversity, Justice, and Social Responsivity Request for Proposals* grant from the Association of Psychology Postdoctoral and Internship Centers (2021-2022).
- *Diversity Seed Grant*, Office of Inclusive Excellence and Advisory Council for Diversity and Reconciliation (ACDR), Seattle Pacific University.
- *ETM Open Textbook & OER Development Funding*, Office of Education, Technology, & Media, Seattle Pacific University.

Chapter 1

Introduction

[Screencasted Lecture Link](#)

1.1 What to expect in each chapter

This textbook is intended as *applied*, in that a primary goal is to help the scientist-practitioner-advocate use a variety of statistics in research problems and *writing them up* for a program evaluation, dissertation, or journal article. In support of that goal, I try to provide just enough conceptual information so that the researcher can select the appropriate statistic (i.e., distinguishing between when ANOVA is appropriate and when regression is appropriate) and assign variables to their proper role (e.g., covariate, moderator, mediator).

This conceptual approach does include occasional, step-by-step, *hand-calculations* (using R to do the math for us) to provide a *visceral feeling* of what is happening within the statistical algorithm that may be invisible to the researcher. Additionally, the conceptual review includes a review of the assumptions about the characteristics of the data and research design that are required for the statistic.

Statistics can be daunting, so I have worked hard to establish a *workflow* through each analysis. When possible, I include a flowchart that is referenced frequently in each chapter and assists the researcher keep track of their place in the many steps and choices that accompany even the simplest of analyses.

As with many statistics texts, each chapter includes a *research vignette*. Somewhat unique to this resource is that the vignettes are selected from recently published articles. Each vignette is chosen with the intent to meet as many of the following criteria as possible:

- the statistic that is the focus of the chapter was properly used in the article,
- the author's identity is from a group where scholarship is historically marginalized (e.g., BIPOC, LGBTQ+, LMIC [low middle income countries]),
- the research has a justice, equity, inclusion, diversity, and social responsibility focus and will contribute positively to a social justice pedagogy, and
- there is sufficient information in the article to simulate the data for the chapter example(s) and practice problem(s); or the data is available in a repository.

In each chapter we employ *R* packages that will efficiently calculate the statistic and the dashboard of metrics (e.g., effect sizes, confidence intervals) that are typically reported in psychological science.

1.2 Strategies for Accessing and Using this OER

There are a number of ways you can access this resource. You may wish to try several strategies and then select which works best for you. I demonstrate these in the screencast that accompanies this chapter.

1. Simply follow along in your preferred format of the book (html, PDF, or ebook) and then
 - open a fresh .rmd file of your own, copying (or retyping) the script and running it
2. Locate the original documents at the [GitHub repository](#). You can
 - open them to simply take note of the “behind the scenes” script
 - copy/download individual documents that are of interest to you
 - clone a copy of the entire project to your own GitHub site and further download it (in its entirety) to your personal workspace. The [GitHub Desktop app](#) makes this easy!
3. Listen to the accompanying lectures (I think sound best when the speed is 1.75). The lectures are being recorded in Panopto and should include the closed captioning.
4. Each time the book is updated, new ebook and PDF versions are also pushed out. You can access these in the “docs” folder at the [GitHub repository](#).
5. Provide feedback to me! If you fork a copy to your own GitHub repository, you can
 - open up an editing tool and mark up the document with your edits,
 - start a discussion by leaving comments/questions, and then
 - sending them back to me by committing and saving. I get an e-mail notifying me of this action. I can then review (accepting or rejecting) them and, if a discussion is appropriate, reply back to you.
 - I am also seeking peer-review feedback at this [Qualtrics-hosted survey](#). You are welcome to complete only the portions that are relevant to you.

1.3 If You are New to R

R can be overwhelming. Jumping right into advanced statistics might not be the easiest way to start. The [Ready_Set_R](#)lesson of this volume provides an introduction and the [waRming up](#)lesson walks through simple data preparation and descriptive statistics.

In the remaining lessons, I have attempted to provide complete code for every step of the process, starting with uploading the data. To help explain what *R* script is doing, I sometimes write it in the chapter text; sometimes leave hashtags-commments in the chunks; and, particularly in the accompanying screencasted lectures, try to take time to narrate what the *R* script is doing.

I've found that, somewhere on the internet, there's almost always a solution to what I'm trying to do. I am frequently stuck and stumped and have spent hours searching the internet for even the tiniest of tasks. When you watch my videos, you may notice that in my R studio, there is a "scRiptuRe" file. I take notes on the solutions and scripts here – using keywords that are meaningful to me so that when I need to repeat the task, I can hopefully search my own prior solutions and find a fix or a hint. You may also find it useful to create a working document of your own tips and tricks.

Chapter 2

Ready_Set_R

[Screencasted Lecture Link](#)

With the goal of creating a common, system-wide approach to using the platform, this lesson was originally created for Clinical and Industrial-Organizational doctoral students who are entering the “stats sequence.” I hope it will be useful for others (e.g., faculty, post-doctoral researchers, and practitioners) who are also making the transition to R.

2.1 Navigating this Lesson

There is about 45 minutes of lecture.

While the majority of R objects and data you will need are created within the R script that sources the chapter, occasionally there are some that cannot be created from within the R framework. Additionally, sometimes links fail. All original materials are provided at the [Github site](#) that hosts the book. More detailed guidelines for ways to access all these materials are provided in the OER’s [introduction](#)

2.1.1 Learning Objectives

Learning objectives from this lecture include the following:

- Downloading/installing R’s parts and pieces.
- Using R-Markdown as the interface for running R analyses and saving the script.
- Recognizing and adopting best practices for “R hygiene.”
- Identifying effective strategies for troubleshooting R hiccups.

2.2 downloading and installing R

2.2.1 So many paRts and pieces

Before we download R, it may be helpful to review some of R’s many parts and pieces.

The base software is free and is available [here](#)

Because R is already on my machine (and because the instructions are sufficient), I will not walk through the demo, but I will point out a few things.

- The “cran” (I think “cranium”) is the *Comprehensive R Archive Network*. In order for R to run on your computer, you have to choose a location – and it should be geographically “close to you.”
 - Follow the instructions for your operating system (Mac, Windows, Linux)
 - You will see the results of this download on your desktop (or elsewhere if you chose to not have it appear there) but you won’t ever use R through this platform.
- **R Studio** is the way in which we operate R. It’s a separate download. Choose the free, desktop, option that is appropriate for your operating system:
- *R Markdown* is the way that many analysts write *script*, conduct analyses, and even write up results. These are saved as .rmd files.
 - In R Studio, open an R Markdown document through File/New File/R Markdown
 - Specify the details of your document (title, author, desired output)
 - In a separate step, SAVE this document [File/Save] into a NEW FILE FOLDER that will contain anything else you need for your project (e.g., the data).
 - *Packages* are at the heart of working in R. Installing and activating packages require writing script.

Note If you have an enterprise-owned machine (e.g., in my specific context, if you are a faculty/staff or have a lab with institution-issued laptops) there can be complications caused by how documents are stored. In recent years we have found that letting the computer choose where to load base R, R Studio, and the packages generally works. The trick, though, is to save R projects (i.e., folder with .rmd files and data) into the OneDrive folder that syncs to your computer. If you have difficulty knitting that is unrelated to code (verified by getting a classmate or colleague to successfully knit it), it is likely because you have saved the files to the local hard drive and not OneDrive. If you continue to have problems I recommend consulting with your computer and technology support office.

2.2.2 oRienting to R Studio (focusing only on the things we will be using first and most often)

R Studio is organized around four panes. These can be sized and rearranged to suit your personal preferences.

- Upper right window
 - Environment: lists the *objects* that are available to you (e.g., dataframes)
- Lower right window
 - *Files*: Displays the file structure in your computer’s environment. Make it a practice to (a) organize your work in small folders and (b) navigating to that small folder that is holding your project when you are working on it.

- *Packages*: Lists the packages that have been installed. If you navigate to it, you can see if it is “on.” You can also access information about the package (e.g., available functions, examples of script used with the package) in this menu. This information opens in the Help window.
- The *Viewer* and *Plots* tabs will be useful, later, in some advanced statistics when we can simultaneously examine output and script in windows that are side-by-side.
- Upper left window
 - If you are using R Markdown, that file lives here and is composed of open space and chunks.
- Lower left window
 - R Studio runs in the Console (the background). Very occasionally, I can find useful troubleshooting information here.
 - More commonly, I open my R Markdown document so that it takes the whole screen.

2.3 best pRactices

Many initial problems in R can be solved with good R hygiene. Here are some suggestions for basic practices. It can be tempting to “skip this.” However, in the first few weeks of class, these are the solutions I am presenting (and repeating, ad nauseum) to my students.

2.3.1 Everything is documented in the .rmd file

Although others do it differently, I put everything in my .rmd file. That is, for uploading data I write the code in my .rmd file. For opening packages, I include the package in my script. I also use the .rmd file to make notes about what I was thinking and why I made the choices I did. I also keep a “bug log” – noting what worked and what did not work. I will also begin my APA style results section directly in the .rmd file.

Why do I do all this? Because when I return to my project hours or years later, I have a permanent record of very critical things like (a) where my data is located, (b) what version I was using, and (c) what package was associated with the functions.

2.3.2 Setting up the file

File organization is a critical key to success:

- Create a project file folder.
- Put the data file in it.
- Open an R Markdown file.
- Save it in the same file folder.
- When your data and .rmd files are in the same folder (not your desktop, but a shared folder) the data can be pulled into the .rmd file without creating a working directory.

2.3.3 Script in chunks and everything else in the “inline text” sections

The R Markdown document is an incredible tool for integrating text, tables, and analyses. This entire OER is written in R Markdown. A central feature of this is “chunks.”

The only thing in the chunks should be script for running R. You can also hashtag-out comments so they won’t run.

You can put almost anything you want in the “inline text with simple formatting.” Syntax for simple formatting in the text areas (e.g., using italics, making headings, bold) is found here: https://rmarkdown.rstudio.com/authoring_basics.html

“Chunks” start and end with three tic marks and will show up in a shaded box. Chunks have three symbols in their upper right. Those controls will disappear and your script will not run if you have replaced them with double or single quotation marks or one or more of the tics are missing.

The easiest way to insert a chunk is to use the INSERT/R command at the top of this editor box. You can also insert a chunk with the keyboard shortcut: CTRL/ALT/i

```
#hashtags let me write comments to remind myself what I did
#here I am simply demonstrating arithmetic (but I would normally be running code)
2021 - 1966
```

[1] 55

2.3.4 Managing packages

As scientist-practitioners (and not coders), we will rely on *packages* to do much of the work. At first you may feel overwhelmed about the large number of packages that are available. Soon, though, you will become accustomed to the ones most applicable to our work (e.g., psych, tidyverse, lavaan, apaTables).

Researchers treat packages differently. In these lectures, I list all the packages we will use in an opening chunk at the beginning of the lecture. When the hashtags are removed, the script will ask R to check to see if the package is installed. If it is, installation is skipped. If it is not, R installs it. Simply remove the hashtag to run the code the first time, then hashtag them back out so R is not always re-checking.

```
# will install the package if not already installed
# if(!require(psych)){install.packages('psych')}
```

To make a package operable, you need to open it. There are two primary ways to do this. The first is to use the library function.

```
#install.packages ("psych")
library (psych)
```

The second way is to place a double colon between the package and function. This second method has become my preferred practice because it helps me remember what package goes with each function. It can also prevent R hiccups when there are identical function names and R does not know which package to use. Below is an example where I might ask for descriptives from the psych package. Because I have not yet uploaded data, I have hashtags it out, making the command inoperable.

```
#psych::describe(mydata)
```

There are exceptions. One is the *tidyverse* package. Some of my script uses pipes (%>%) and they require tidyverse to be activated this is why you will often see me call the tidyverse package with the *library()* function (as demonstrated above.)

2.3.5 Upload the data

When imported (or simulated) properly, data will appear as an object in the global environment.

In the context of this OER, I will be simulating data right in each lesson for use in the lesson. This makes the web-based platform more *portable*. This means that when working the problems in the chapter we do not (a) write the data to a file and (b) import data from files. Because these are essential skills, I will demonstrate this process here – starting with simulating data.

At this point, simulating data is beyond the learning goals I have established for the chapter. I do need to include the code so that we get some data. The data I am simulating is used in the [one-way ANOVA lesson](#). The data is from the Tran and Lee [2014] random clinical trial.

In this simulation, I am simply creating an ID number, a condition (High, Low, Control), and a score on the dependent variable, “Accurate.” More information about this study is included in the [one-way ANOVA chapter](#).

```
# Note, this simulation results in a different dataset than is in the
# OnewayANOVA lesson sets a random seed so that we get the same
# results each time
set.seed(2021)
# sample size, M and SD for each group
Accurate <- c(rnorm(30, mean = 1.18, sd = 0.8), rnorm(30, mean = 1.83,
               sd = 0.58), rnorm(30, mean = 1.76, sd = 0.56))
# set upper bound for DV
Accurate[Accurate > 3] <- 3
# set lower bound for DV
Accurate[Accurate < 0] <- 0
# IDs for participants
ID <- factor(seq(1, 90))
# name factors and identify how many in each group; should be in same
# order as first row of script
COND <- c(rep("High", 30), rep("Low", 30), rep("Control", 30))
# groups the 3 variables into a single df: ID, DV, condition
Acc_sim30 <- data.frame(ID, COND, Accurate)
```

At this point, this data lives only in this .rmd file after the above code is run. Although there are numerous ways to export and import data, I have a preference for two.

2.3.5.1 To and from .csv files

The first is to write the data to a .csv file. In your computer's environment (outside of R), these files are easily manipulated in Excel. I think of them as being "Excel lite" because although Excel can operate them, they lack some of the more advanced features of an Excel spreadsheet.

In the code below, I identify the R object "Acc_sim30" and give it a file name, "to_CSV.csv". This file name must have the .csv extension. I also indicate that it should preserve the column names (but ignore row names; since we don't have row names).

This file will save in the same folder as wherever you are using this .rmd file.

```
# to write it to an outfile as a .csv
write.table(Acc_sim30, file = "to_CSV.csv", sep = ",", col.names = TRUE,
            row.names = FALSE)
```

Importing this object back into the R environment can be accomplished with some simple code. For the sake of demonstration,

```
# to save the df as an .csv (think 'Excel lite') file on your
# computer; it should save in the same file as the .rmd file you are
# working with
from_CSV <- read.csv("to_CSV.csv", header = TRUE)
```

The advantage of working with .csv files is that it is then easy to inspect and manipulate them outside of the R environment. The disadvantage of .csv files is that each time they are imported they lose any formatting you may have meticulously assigned to them.

2.3.5.2 To and from .rds files

While it is easy enough to rerun the code (or copy it from data prep .rmd and paste it into an .rmd you are using for advanced analysis), there is a better way! Saving the data as an R object preserves all of its characteristics.

```
# to save the df as an .rds file on your computer; it should save in
# the same file as the .rmd file you are working with
saveRDS(Acc_sim30, "to_Robject.rds")
```

This file will save to your computer (and you can send it to colleagues). However, it is not easy to "just open it" in Excel. To open an .rds file and use it (whether you created it or it is sent to you by a colleague), use the following code:

```
from_rds <- readRDS("to_Robject.rds")
```

If you are the recipient of an R object, but want to view it as a .csv, simply import the .rds then use the above code to export it as a .csv.

2.3.5.3 From SPSS files

Your data may come to you in a variety of ways. One of the most common is SPSS. The *foreign* package is popular for importing SPSS data. Below is code which would import an SPSS file *if I had created one*. You'll see that this script is hashtags out because I rarely use SPSS and do not have a handy file to demo.

```
# opening an SPSS file requires the foreign package which I opened
# earlier from_SPSS <- foreign::read.spss ('SPSSdata.sav',
# use.value.labels = TRUE, to.data.frame = TRUE)
```

2.4 quick demonstRation

Let's run some simple descriptives. In the script below, I am using the *psych* package. Descriptive statistics will appear for all the data in the dataframe and the output will be rounded to three spaces. Note that rather than opening the psych package with the library function, I have used the double colon convention.

```
round(psych::describe(Acc_sim30), 3)
```

	vars	n	mean	sd	median	trimmed	mad	min	max	range	skew	kurtosis
ID*	1	90	45.50	26.12	45.50	45.50	33.36	1	90	89	0.00	-1.24
COND*	2	90	2.00	0.82	2.00	2.00	1.48	1	3	2	0.00	-1.53
Accurate	3	90	1.52	0.68	1.55	1.54	0.70	0	3	3	-0.19	-0.34
			se									
ID*			2.75									
COND*			0.09									
Accurate			0.07									

Because "ID" is the case ID and COND is the factor (high, low, control), the only variable for which this data is sensible is "Accurate." Nonetheless, this provides an example of how to apply a package's function to a dataset. As we progress through the text we will learn how to manage the data so that we get the specific output we are seeking.

2.5 the knitted file

One of the coolest things about R Markdown is its capacity to *knit* to HTML, PPT, or WORD.

- In this OER, I am writing the lessons in R markdown (.rmd files), with the package *bookdown* as a helper, and knitting the files to the .html format. In prior years, I knitted these documents to .doc formats. There are numerous possibilities!
- The package *papaja* is designed to prepare APA manuscripts where the writing, statistics, and references are all accomplished in a single file. This process contributes to replicability and reproducibility.
- If you are using the PDF or ebook version of this OER you will realize that it is also possible to render to these formats. Albeit slightly more complicated, this is possible, too! More detailed instructions for this are provided in the [extRas](#) mini-volume of [ReCentering Psych Stats](#).

2.6 tRoubleshooting in R maRkdown

Hiccups are normal. Here are some ideas that I have found useful in getting unstuck.

- In a given set of operations, you must run/execute each piece of code in order: every, single, time. That is, all the packages have to be in your library and activated.
 - If you open an .rmd file, you cannot just scroll down to make a boxplot. You need to run any *prerequisite* script (like loading files, putting the data in the global environment, etc.)
 - Lost? Clear your global environment (broom icon in the upper right) and start over. Fresh starts are good.
- Your .rmd file and your data need to be stored in the same file folder. Make unique folders for each project (even if each contains only a few files).
- If you have tried what seems apparent to you and cannot solve your challenge, do not wait long before typing warnings into a search engine. Odds are, you'll get some useful hints in a manner of seconds. Especially at first, these are common errors:
 - The package isn't loaded.
 - The .rmd file hasn't been saved yet, or isn't saved in the same folder as the data.
 - There are errors in punctuation or spelling.
- Restart R (it's quick – not like restarting your computer). I like to restart and clear my output and environment so that I can better track my order of operations.
- If you receive an error indicating that a function isn't working or recognized, and you have loaded the package, type the name of the package in front of the function with two colons (e.g., psych::describe(df)). If multiple packages are loaded with functions that have the same name, R can get confused.

2.7 just *why* have we tRansitioned to R?

- It (or at least it appears to be) is the futuRe.
- SPSS site (and individual) licenses are increasingly expensive and limited (e.g., we need Mplus, AMOS, HLM, or R). As package development for R is exploding, we have tools to “do just about anything.”

- Most graduate psychology programs are scientist/practitioner in nature and include training in “high end” statistics. Yet, many of your employing organizations will not have SPSS. R is a free, universally accessible program, that our graduates can use anywhere.

2.8 stRategies for success

- Engage with R, but don’t let it overwhelm you.
 - The *mechanical is also the conceptual*. Especially while it’s *simpler*, do try to retype the script into your own .rmd file and run it. Track down the errors you are making and fix them.
 - If this stresses you out, move to simply copying the code into the .rmd file and running it. If you continue to have errors, you may have violated one of the best practices above (ask, “Is the package activated?” “Are the data and .rmd files in the same place?” “Is all the prerequisite script run?”).
 - Still overwhelmed? Keep moving forward by (retrieving the original.rmd file from the GitHub repository) opening a copy of the .rmd file and just “run it along” with the lecture. Spend your mental power trying to understand what each piece does so you can translate it for any homework assignments. My suggestions for practice aspire to be parallel to the lecture with no sneaky trix.
- Copy script that works elsewhere and replace it with your datafile, variables, and so forth.
- The learning curve is steep, but not impossible. Gladwell [2008] taught us that it takes about 10,000 hours to get great at something (2,000 to get reasonably competent). Practice. Practice. Practice.
- Updates to R, R Studio, and the packages are necessary, but can also be problematic. Sometimes updates cause programs/script to fail (e.g., “X has been deprecated for version X.XX”). My personal practice is to update R, R Studio, and the packages a week or two before each academic term. I expect that prior scripts may need to be updated or revised with package updates and incongruencies between base R, R Studio, and the packages.
- Embrace your downward dog. And square breathing. Also, walk away, then come back.

2.9 Resources for getting staRted

R for Data Science: <https://r4ds.had.co.nz/>

R Cookbook: <http://shop.oreilly.com/product/9780596809164.do>

R Markdown homepage with tutorials: <https://rmarkdown.rstudio.com/index.html>

R has cheatsheets for everything, here’s the one for R Markdown: <https://www.rstudio.com/wp-content/uploads/2015/02/rmarkdown-cheatsheet.pdf>

R Markdown Reference guide: <https://www.rstudio.com/wp-content/uploads/2015/03/rmarkdown-reference.pdf>

Using R Markdown for writing reproducible scientific papers: <https://libscie.github.io/rmarkdown-workshop/handout.html>

Script for all of Field’s text: <https://studysites.uk.sagepub.com/dsur/study/scriptfi.htm>

LaTeX equation editor: <https://www.codecogs.com/latex/eqneditor.php>

Chapter 3

waRming up

[Screencasted Lecture Link](#)

The beginning of any data analysis means familiarizing yourself with the data, that is, its distributional characteristics as well as its relations.

3.1 Navigating this Lesson

There is about 25 minutes of lecture.

While the majority of R objects and data you will need are created within the R script that sources the chapter, occasionally there are some that cannot be created from within the R framework. Additionally, sometimes links fail. All original materials are provided at the [Github site](#) that hosts the book. More detailed guidelines for ways to access all these materials are provided in the OER's [introduction](#)

3.1.1 Learning Objectives

Learning objectives from this lecture include the following:

- Produce descriptive statistics (means, standard deviations, skew, kurtosis), and correlation matrix, with the *psych* package
- Produce plots of data
- Interpret skew, kurtosis, correlations
- Create an APA Style table and results section that includes means, standard deviations, and correlations

3.1.2 Planning for Practice

This is designed as a “get (or ‘get back’) into it” assignment. You will essentially work through this very same lecture, using the same dataframe – just a different set of continuous variables.

3.1.3 Readings & Resources

In preparing this chapter, I drew heavily from the following resource(s). Other resources are cited (when possible, linked) in the text with complete citations in the reference list.

- Revelle, W. (2021). An introduction to the psych package: Part I: data entry and data description. 60.
 - Revelle is the author/creator of the *psych* package. His tutorial provides both technical and interpretive information. Read pages 1-17.
- Lui, P. P. (2020). Racial microaggression, overt discrimination, and distress: (In)Direct associations with psychological adjustment. *The Counseling Psychologist*, 32.
 - This is the research vignette from which I simulate data that we can use in the lesson and practice problem.

3.2 Research Vignette

We will use simulated data from Lui [2020] for simple data screening. Controlling for overt discrimination, and neuroticism, Lui examined the degree to which racial microaggressions contributed to negative affect, alcohol consumption, and drinking problems in African American, Asian American, and Latinx American college students ($N = 713$).

Using the means, standard deviations, correlation matrix, and group sizes (n) I simulated the data. While the process of simulation is beyond the learning goals of this lesson (you can skip that part), I include it here so that it is easy to work the rest of the script.

```
set.seed(210807) #sets the random seed so that we consistently get the same results
# for practice, you could change (or remove) the random seed and try
# to interpret the results (they should be quite similar) There are
# probably more efficient ways to simulate data. Given the
# information available in the manuscript, my approach was to first
# create the datasets for each of the racial ethnic groups that were
# provided and then binding them together.

# First, the data for the students who identified as Asian American
Asian_mu <- c(1.52, 1.72, 2.69, 1.71, 2.14, 2.35, 2.42)
Asian_stddev <- c(2.52, 2.04, 0.47, 0.7, 0.8, 2.41, 3.36)
Asian_corMat <- matrix(c(1, 0.69, 0.19, 0.28, 0.32, 0.08, 0.23, 0.69, 1,
  0.2, 0.29, 0.33, 0.13, 0.25, 0.19, 0.2, 1, 0.5, 0.5, -0.04, 0.09, 0.28,
  0.29, 0.5, 1, 0.76, 0.04, 0.18, 0.32, 0.33, 0.5, 0.76, 1, 0.1, 0.21,
  0.08, 0.13, -0.04, 0.04, 0.1, 1, 0.42, 0.23, 0.25, 0.09, 0.18, 0.21,
  0.42, 1), ncol = 7)
Asian_covMat <- Asian_stddev %*% t(Asian_stddev) * Asian_corMat

Asian_dat <- MASS::mvrnorm(n = 398, mu = Asian_mu, Sigma = Asian_covMat,
  empirical = TRUE)
```

```

Asian_df <- as.data.frame(Asian_dat)

library(tidyverse)
Asian_df <- rename(Asian_df, OvDisc = V1, mAggr = V2, Neuro = V3, nAff = V4,
  psyDist = V5, Alcohol = V6, drProb = V7)
Asian_df$RacEth <- "Asian"

# Second, the data for the students who identified as Black/African
# American
Black_mu <- c(4.45, 3.84, 2.6, 1.84, 2.1, 2.81, 2.14)
Black_stddev <- c(4.22, 3.08, 0.89, 0.8, 0.81, 2.49, 3.24)
Black_corMat <- matrix(c(1, 0.81, 0.17, 0.15, 0.09, 0.05, -0.16, 0.81,
  1, 0.17, 0.21, 0.11, 0.09, -0.01, 0.17, 0.17, 1, 0.59, 0.54, 0.05,
  0.24, 0.15, 0.21, 0.59, 1, 0.72, 0.12, 0.22, 0.09, 0.11, 0.54, 0.72,
  1, 0.21, 0.4, 0.05, 0.09, 0.05, 0.12, 0.21, 1, 0.65, -0.16, -0.01,
  0.24, 0.22, 0.4, 0.65, 1), ncol = 7)
Black_covMat <- Black_stddev %*% t(Black_stddev) * Black_corMat
Black_dat <- MASS::mvrnorm(n = 133, mu = Black_mu, Sigma = Black_covMat,
  empirical = TRUE)
Black_df <- as.data.frame(Black_dat)
Black_df <- rename(Black_df, OvDisc = V1, mAggr = V2, Neuro = V3, nAff = V4,
  psyDist = V5, Alcohol = V6, drProb = V7)
Black_df$RacEth <- "Black"

# Third, the data for the students who identified as Latinx American
Latinx_mu <- c(1.56, 2.34, 2.69, 1.81, 2.17, 3.47, 2.69)
Latinx_stddev <- c(2.46, 2.49, 0.86, 0.71, 0.78, 2.59, 3.76)
Latinx_corMat <- matrix(c(1, 0.78, 0.27, 0.36, 0.42, -0.06, 0.08, 0.78,
  1, 0.33, 0.26, 0.35, -0.11, -0.02, 0.27, 0.33, 1, 0.62, 0.64, -0.04,
  0.15, 0.36, 0.26, 0.62, 1, 0.81, -0.08, 0.17, 0.42, 0.35, 0.64, 0.81,
  1, -0.06, 0.15, -0.06, -0.11, -0.04, -0.08, -0.06, 1, 0.6, 0.08, -0.02,
  0.15, 0.17, 0.15, 0.6, 1), ncol = 7)
Latinx_covMat <- Latinx_stddev %*% t(Latinx_stddev) * Latinx_corMat
Latinx_dat <- MASS::mvrnorm(n = 182, mu = Latinx_mu, Sigma = Latinx_covMat,
  empirical = TRUE)
Latinx_df <- as.data.frame(Latinx_dat)
Latinx_df <- rename(Latinx_df, OvDisc = V1, mAggr = V2, Neuro = V3, nAff = V4,
  psyDist = V5, Alcohol = V6, drProb = V7)
Latinx_df$RacEth <- "Latinx"

Lui_sim_df <- bind_rows(Asian_df, Black_df, Latinx_df)

```

If you have simulated the data, you can continue using the the “Lui_sim_df” object that we created. In your own research you may wish to save data as a file. Although I will hashtag it out (making it inoperable until the hashtags are removed), here is code to save the simulated data both .csv (think “Excel lite”) and .rds (it retains all the properties we assigned to it in R) files and then bring/import them back into R. For more complete instructions see the [Ready_Set_R](#) lesson.

```
# write the simulated data as a .csv write.table(Lui_sim_df,
# file='Lui_CSV.csv', sep=',', col.names=TRUE, row.names=FALSE) bring
# back the simulated dat from a .csv file df <- read.csv
# ('Lui_CSV.csv', header = TRUE)

# to save the df as an .rds (think 'R object') file on your computer;
# it should save in the same file as the .rmd file you are working
# with saveRDS(Lui_sim_df, 'Lui_RDS.rds') bring back the simulated
# dat from an .rds file df <- readRDS('Lui_RDS.rds')
```

You may have noticed a couple of things in each of these operations

- First, I named the data object to include a “df” (i.e., dataframe).
 - It is a common (but not required) practice for researchers to simply use “df” or “dat” for their R script, no matter the type or source of the data. This practice has advantages (such as making the reuse of code quite easy) and disadvantages (it is easy to get confused about what data is what).
- Second, if you run the code, the updating *replaces* the prior code.
 - While this is irrelevant today (we are saving the same data in different ways), it points out the importance of creating a sensible and systematic *order of operations* in your .rmd files and then knowing where you are in the process.

Because the data is simulated, I can simply use the data I created in the simulation, however, I will go ahead and use the convention of renaming it, “df.”

```
df <- Lui_sim_df
```

Is this R session reading our variables correctly? Check it with the “structure” command.

```
str(df)
```

```
'data.frame': 713 obs. of 8 variables:
 $ OvDisc : num 1.45 4.9 0.45 2.85 2.09 ...
 $ mAggr : num 0.682 4.383 0.225 2.235 1.977 ...
 $ Neuro : num 3.11 3.69 3.5 2.68 2.08 ...
 $ nAff : num 2.32 2.59 2.27 2.28 2.01 ...
 $ psyDist: num 1.83 3.41 2.75 2.11 3.12 ...
 $ Alcohol: num 3.125 4.388 0.999 0.137 -0.575 ...
 $ drProb : num 2.499 -3.23 3.035 -0.858 -1.467 ...
 $ RacEth : chr "Asian" "Asian" "Asian" "Asian" ...
```

Looking back at the Lui [2020] article we can presume the following variables (variable/variable name) to have the following structure (scaling/R scaling):

- Discrimination/OvDis: interval/num
 - theoretically someone could experience no discrimination, but it would still score as 1.0 if the scale is averaged and the respondent marked 1 (*never* or *not at all*) for all the options.
- Microaggressions/mAggr: interval/num
 - theoretically someone could experience no microaggressions, but it would still score as 1.0 if the scale is averaged and the respondent marked 1 (*never*; and then *not at all* [stressful]) for all the options.
- Neuroticism/Neuro: interval/num
- Negative affect/nAff: interval/num
- Psychological distress/psyDist :interval/num
- Hazardous alcohol use/Alcohol: interval/num
- Drinking problems/drProb: interval/num
- Race Ethnicity/RacEth: nominal/chr

Only the Race Ethnicity needs to be transformed from a character (“chr”) variable to a factor. I will use the *mutate()* function in the *dplyr* package to convert the RacEth variable to be a factor with three levels.

```
# A .csv file is uninformed -- it just holds data (and R guesses what
# it is); respecifying the type of variable will likely need to be
# completed each time the file is used.

df <- df %>%
  dplyr::mutate(RacEth = as.factor(RacEth))
```

```
# checking the structure of the data
str(df)
```

```
'data.frame': 713 obs. of 8 variables:
 $ OvDisc : num 1.45 4.9 0.45 2.85 2.09 ...
 $ mAggr : num 0.682 4.383 0.225 2.235 1.977 ...
 $ Neuro : num 3.11 3.69 3.5 2.68 2.08 ...
 $ nAff : num 2.32 2.59 2.27 2.28 2.01 ...
 $ psyDist: num 1.83 3.41 2.75 2.11 3.12 ...
 $ Alcohol: num 3.125 4.388 0.999 0.137 -0.575 ...
 $ drProb : num 2.499 -3.23 3.035 -0.858 -1.467 ...
 $ RacEth : Factor w/ 3 levels "Asian","Black",...: 1 1 1 1 1 1 1 1 1 ...
```

It is possible to work with this data without restructuring them into a “tiny df.” However, this function is often one of the first skills we want to use so I will demonstrate it here.

Manipulating the data is an important skill. I can use the *select()* function to retain three variables that are continuous and then obtain descriptive statistics about them.

```
# I often 'open' the package by placing it (and two colons) in front
# of its function. However, opening tidyverse via the library command
# allows me to use the pipe: %>%
library(tidyverse)
tiny_df <- df %>%
  dplyr::select(mAggr, nAff, psyDist)
```

We can start with simple descriptives. Revelle's [Revelle, 2021] *psych* package was made specifically for much of the type of work we do.

```
psych::describe(tiny_df)
```

	vars	n	mean	sd	median	trimmed	mad	min	max	range	skew	kurtosis
mAggr	1	713	2.27	2.51	2.17	2.19	2.42	-4.42	10.42	14.85	0.35	0.32
nAff	2	713	1.76	0.72	1.75	1.76	0.71	-0.56	4.19	4.75	0.01	0.15
psyDist	3	713	2.14	0.80	2.11	2.13	0.79	-0.10	4.73	4.84	0.12	0.08
			se									
mAggr			0.09									
nAff			0.03									
psyDist			0.03									

One of the easiest way to get "counts" of categorical variables is to use them in the *psych* package's *describeBy()* function. In the script below, I am asking for descriptives of the entire df, but disaggregated by RacEth. I have added the "mat=TRUE" command so that the results are presented in one complete table. This is optional and so omitting the statement or writing "mat=FALSE" can be something you might like to try.

```
psych::describeBy(df, df$RacEth, mat = TRUE)
```

	item	group1	vars	n	mean	sd	median	trimmed	mad
OvDisc1	1	Asian	1	398	1.52	2.52	1.671877	1.536984	2.5083543
OvDisc2	2	Black	1	133	4.45	4.22	4.101902	4.459685	3.7911955
OvDisc3	3	Latinx	1	182	1.56	2.46	1.622515	1.592317	2.1839022
mAggr1	4	Asian	2	398	1.72	2.04	1.629094	1.712366	2.0479218
mAggr2	5	Black	2	133	3.84	3.08	3.560519	3.844107	2.8031478
mAggr3	6	Latinx	2	182	2.34	2.49	2.366613	2.343589	2.5832009
Neuro1	7	Asian	3	398	2.69	0.47	2.664432	2.688114	0.4913231
Neuro2	8	Black	3	133	2.60	0.89	2.691292	2.616584	0.8381441
Neuro3	9	Latinx	3	182	2.69	0.86	2.719094	2.697182	0.8817373
nAff1	10	Asian	4	398	1.71	0.70	1.724998	1.709749	0.6808739
nAff2	11	Black	4	133	1.84	0.80	1.779947	1.837073	0.8226934
nAff3	12	Latinx	4	182	1.81	0.71	1.799092	1.811002	0.6725811
psyDist1	13	Asian	5	398	2.14	0.80	2.117524	2.136579	0.8011511
psyDist2	14	Black	5	133	2.10	0.81	2.001099	2.084985	0.8371713
psyDist3	15	Latinx	5	182	2.17	0.78	2.142896	2.159840	0.7579508
Alcohol1	16	Asian	6	398	2.35	2.41	2.299663	2.324671	2.4620921

Alcohol2	17	Black	6	133	2.81	2.49	2.760749	2.822602	2.1592551	
Alcohol3	18	Latinx	6	182	3.47	2.59	3.353811	3.479536	2.7406457	
drProb1	19	Asian	7	398	2.42	3.36	2.409912	2.418404	3.5040063	
drProb2	20	Black	7	133	2.14	3.24	2.254354	2.158361	3.5583579	
drProb3	21	Latinx	7	182	2.69	3.76	2.528477	2.658522	3.5799107	
RacEth*1	22	Asian	8	398	1.00	0.00	1.000000	1.000000	0.0000000	
RacEth*2	23	Black	8	133	2.00	0.00	2.000000	2.000000	0.0000000	
RacEth*3	24	Latinx	8	182	3.00	0.00	3.000000	3.000000	0.0000000	
			min		max	range		skew	kurtosis	se
OvDisc1			-5.83902804		8.421599	14.260627	-0.078189651	-0.13080404	0.12631619	
OvDisc2			-10.31638613		16.388836	26.705222	-0.125879064	0.57600168	0.36592041	
OvDisc3			-6.57126805		7.625248	14.196516	-0.142542665	0.35807531	0.18234733	
mAggr1			-3.63155800		8.682565	12.314123	0.091336696	-0.18297749	0.10225596	
mAggr2			-4.09305401		10.424468	14.517522	-0.017884129	-0.31873937	0.26706987	
mAggr3			-4.42196168		9.237781	13.659743	-0.025230120	-0.03996127	0.18457108	
Neuro1			1.45287258		3.988193	2.535321	0.070330835	-0.35836361	0.02355897	
Neuro2			-0.28319065		4.738701	5.021892	-0.259085030	0.08300312	0.07717279	
Neuro3			0.01432133		5.263965	5.249644	-0.102274817	0.54931296	0.06374744	
nAff1			-0.29889116		3.613959	3.912850	-0.039998756	-0.12116003	0.03508783	
nAff2			-0.29056580		4.042763	4.333328	0.034482279	-0.25309219	0.06936880	
nAff3			-0.56044730		4.186554	4.747001	0.002283094	0.92439559	0.05262870	
psyDist1			-0.08596817		4.514520	4.600488	0.071251904	-0.12020053	0.04010038	
psyDist2			-0.10126503		4.402106	4.503371	0.151367949	-0.01156587	0.07023591	
psyDist3			-0.07657607		4.734104	4.810680	0.205329737	0.54580657	0.05781745	
Alcohol1			-3.07666900		9.204501	12.281170	0.143207169	-0.37141988	0.12080238	
Alcohol2			-5.55081496		9.730453	15.281268	-0.169805727	0.69454596	0.21591038	
Alcohol3			-3.64260294		10.765453	14.408056	-0.014397205	-0.23205639	0.19198357	
drProb1			-7.97853725		11.688962	19.667499	-0.006618251	-0.07367148	0.16842158	
drProb2			-5.64470469		9.939996	15.584701	-0.063644276	-0.38485480	0.28094363	
drProb3			-7.28910040		10.657865	17.946965	0.028417092	-0.36766984	0.27870974	
RacEth*1			1.00000000		1.000000	0.000000		NaN	NaN	0.00000000
RacEth*2			2.00000000		2.000000	0.000000		NaN	NaN	0.00000000
RacEth*3			3.00000000		3.000000	0.000000		NaN	NaN	0.00000000

3.2.1 Determining Skew and Kurtosis

To understand whether our data is normally distributed, we can look at skew and kurtosis. The skew and kurtosis indices in the *psych* package are reported as *z* scores. Regarding skew, values > 3.0 are generally considered “severely skewed.” Regarding kurtosis, “severely kurtotic” is argued anywhere > 8 to 20 [Kline, 2016].

3.2.2 SPLOM

SPLOM (scatterplot matrix) is a graphical tool that uses multiple scatterplots to determine the correlation (if any) between a set of variables. These plots are organized into a matrix, making it easy to see them all at once.

In the *psych* package, the *pairs.panels()* produces these.

On the diagonal we see:

- histogram of each variable
- superimposed with a normal curve

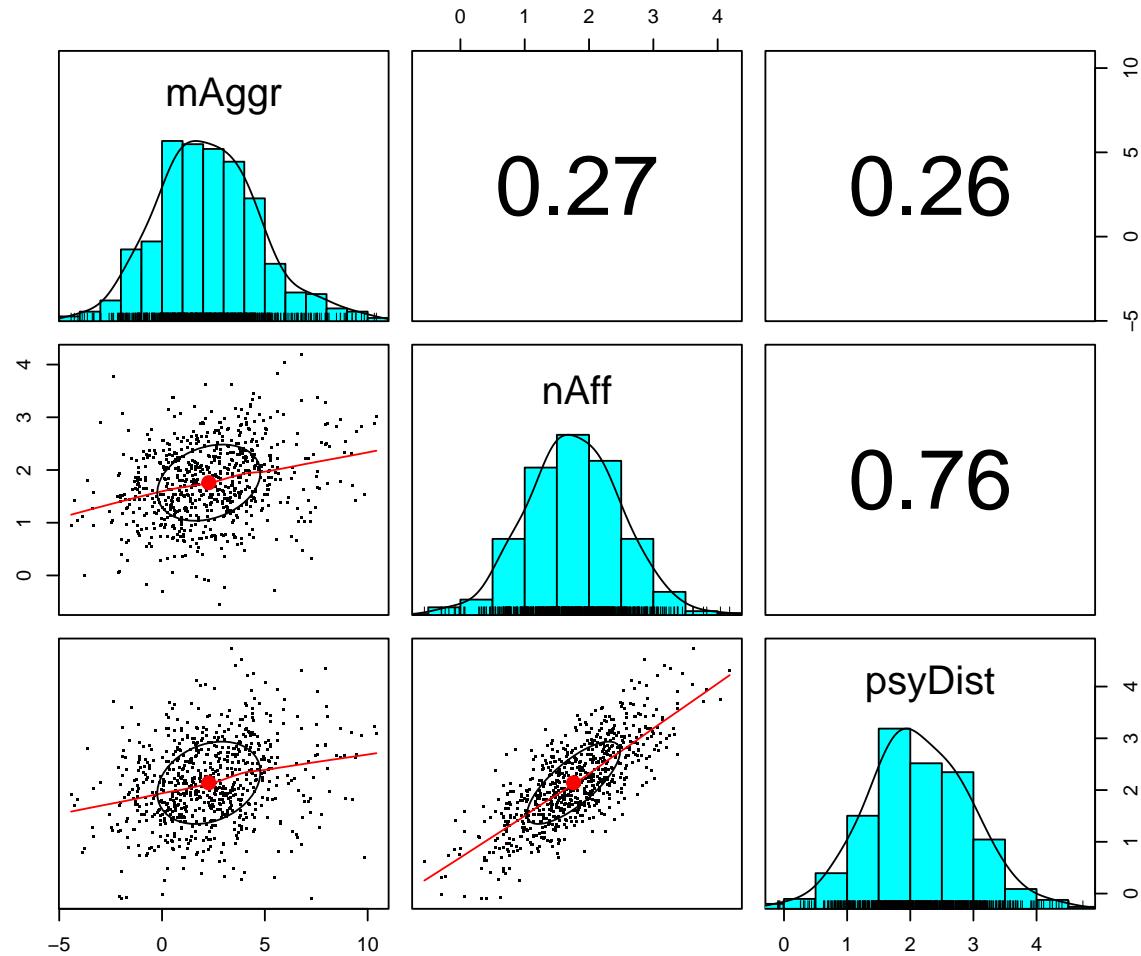
Below the diagonal we see:

- xy scatter plot (not useful for our categorical RacEth row)
- the *lowess* locally fit regression line
- the X axis represents the column variable; the Y axis, the row variable

Above the diagonal we see:

- Pearson correlation

```
# the pch = '.' command produces a cleaner graphic it is especially
# useful when there are lots of variables
psych::pairs.panels(tiny_df, pch = ".")
```



What do we observe?

- All 3 variables look normally distributed
- The relationship between mAggr with nAff and psyDist is quite comparable
- The relationship between nAff and psyDist is very strong
- Are these correlations significant? Revelle [2021] is not a fan of the associated p value, but we can get it several ways (hang tight).

Writing up an APA style results section frequently involves tables. A helpful package for doing this is *apaTables*. An instructional article notes the contributions of tools like this contributing to the *reproducibility* of science by reducing errors made when the author or analyst retypes or copies text from output to the manuscript. When the R script is shared through an open science framework, reproducibility is further enhanced [Stanley and Spence, 2018].

We pass our tiny df to the *apa.cor.table()* function of the *apaTables* package. Commands allow us to specify what is included in the table and whether it should be displayed in the console or saved as a document to the project's folder.

```
# unlike the psych package, this function removes any categorical
# variables
Table1_Cor <- apaTables::apa.cor.table(tiny_df, filename = "Table1_Cor.doc",
  table.number = 1, show.conf.interval = FALSE, landscape = TRUE)
```

The ability to suppress reporting of reporting confidence intervals has been deprecated in this. The function argument `show.conf.interval` will be removed in a later version.

```
# swap in this command to see it in the console
print(Table1_Cor)
```

Table 1

Means, standard deviations, and correlations with confidence intervals

Variable	M	SD	1	2
1. mAggr	2.27	2.51		
2. nAff	1.76	0.72	.27** [.20, .33]	
3. psyDist	2.14	0.80	.26** [.19, .32]	.76** [.73, .79]

Note. M and SD are used to represent mean and standard deviation, respectively.

Values in square brackets indicate the 95% confidence interval.

The confidence interval is a plausible range of population correlations that could have caused the sample correlation (Cumming, 2014).

* indicates $p < .05$. ** indicates $p < .01$.

These types of statistics are often presented in the preliminary results portion of an empirical manuscript. Here is how I might write them up.

3.3 Results

Our sample included Asian American ($n = 398$), Latinx ($n = 182$), and Black ($n = 133$) participants. Visual inspection of the three variables of interest (negative affect, psychological distress, microaggressions) combined with formal evaluation of skewness and kurtosis suggested univariate normality. Correlations between negative affect and psychological distress were quite strong; correlations with each of these variables and microaggressions were moderate. Results are presented in Table 1.

3.4 Practice Problems

The three exercises described below are designed to “meet you where you are” and allow you to challenge your skills depending on your comfort with statistics and R.

Regardless which you choose, you should:

- Create a tiny_df from a larger df
- Calculate and interpret descriptives for the continuously scaled variables
- Create the SPLOM (pairs.panels) of the continuously scaled variables
- Use the *apaTables* package to make an APA style table with means, standard deviations, and correlations
- Write up a mini-results section (APA style)

3.4.1 Problem #1: Change the Random Seed

If this topic feels a bit overwhelming, simply change the random seed in the data simulation (at the very top), then rework the lesson exactly as written. This should provide minor changes to the data (maybe in the second or third decimal point), but the results will likely be very similar.

3.4.2 Problem #2: Swap Variables in the Simulation

Use the simulated data from the Lui [2020] study. However, select three continuous variables (2 must be different from mine) and then conduct the analyses.

3.4.3 Problem #3: Use (or Simulate) Your Own Data

Use data for which you have permission and access. This could be IRB approved data you have collected or from your lab; data you simulate from a published article; data from an open science repository; or data from other chapters in this OER.

3.4.4 Grading Rubric

Regardless which option(s) you chose, use the elements in the grading rubric to guide you through the practice.

Element	Points Poss	Points Earned
1. Create the tiny_df	3	
2. Descriptives of tiny_df	3	
3. SPLOM/pairs.panels	3	
4. apaTables matrix	3	
5. Mini-results	5	
6. Explanation/discussion with a grader	5	
**Totals	22	

Chapter 4

One-way ANOVA

[Screencasted Lecture Link](#)

One-way ANOVA allows the researcher to analyze mean differences between two or more groups on a between-subjects factor. For the one-way ANOVA, each case (i.e., individual, participant) must have scores on two variables: a factor and a dependent variable.

The factor must be categorical in nature, dividing the cases into two or more groups or levels. These levels could be ordered (e.g., placebo, low dose, high dose) or unordered (e.g., cognitive-behavioral, existential, psychodynamic). The dependent variable must be assessed on a quantitative, continuous dimension. The ANOVA F test evaluates whether population means on the dependent variable differ across the levels of the factor.

One-way ANOVA can be used in experimental, quasi-experimental, and field studies. As we work through the chapter, we will examine some of the requirements (assumptions) of the statistic in greater detail.

4.1 Navigating this Lesson

There is about 2 hours of lecture. If you work through the materials with me, plan for another two hours of study.

4.1.1 Learning Objectives

Learning objectives from this lecture include the following:

- Evaluate the statistical assumptions associated with one-way analysis of variance (ANOVA).
- Describe the relationship between model/between-subjects and residual/within-subjects variance.
- Narrate the steps in conducting a formal one-way ANOVA beginning with testing the statistical assumptions through writing up an APA style results section.
- Conduct a one-way ANOVA in R (including calculation of effect sizes and follow-up to the omnibus).
- Conduct a power analysis for a one-way ANOVA.
- Produce an APA style results section.

4.1.2 Planning for Practice

In each of these lessons I provide suggestions for practice that allow you to select one or more problems that are graded in difficulty. The least complex is to change the random seed and rework the problem demonstrated in the lesson. The results *should* map onto the ones obtained in the lecture.

The second option comes from the research vignette. The Tran et al. [2014] vignette has two variables where the authors have conducted one-way ANOVAs. I will demonstrate one (*Accurate*) in this lecture; the second is available as one of the homework options.

As a third option, you are welcome to use data to which you have access and is suitable for two-way ANOVA. In either case the practice options suggest that you:

- test the statistical assumptions
- conduct a one-way ANOVA, including
 - omnibus test and effect size
 - follow-up (pairwise, planned comparisons, polynomial trends)
- write a results section to include a figure and tables

4.1.3 Readings & Resources

In preparing this chapter, I drew heavily from the following resource(s) that are freely available on the internet. Other resources are cited (when possible, linked) in the text with complete citations in the reference list.

- Navarro, D. (2020). Chapter 14: Comparing Several Means (one-Way ANOVA). In [Learning Statistics with R - A tutorial for Psychology Students and other Beginners](#). Retrieved from [https://stats.libretexts.org/Bookshelves/Applied_Statistics/Book%3A_Learning_Statistics_with_R_-_A_tutorial_for_Psychology_Students_and_other_Beginners_\(Navarro\)](https://stats.libretexts.org/Bookshelves/Applied_Statistics/Book%3A_Learning_Statistics_with_R_-_A_tutorial_for_Psychology_Students_and_other_Beginners_(Navarro))
 - Navarro's OER includes a good mix of conceptual information about one-way ANOVA as well as R code. My code/approach is a mix of Green and Salkind's [2014b], Field's [2012], Navarro's [2020b], and other techniques I have found on the internet and learned from my students.
- Crump, M. J. C. (2018). Chapter 5.5.2, Simulating data for one-way between subjects design with 3 levels. In [Programming for Psychologists: Data Creation and Analysis](#). Retrieved from <https://crumplab.github.io/programmingforpsych/simulating-and-analyzing-data-in-r.html#single-factor-anovas-data-simulation-and-analysis>
 - Although this reference is on simulating data, the process of simulation can provide another perspective on one-way ANOVA.
- Tran, A. G. T. T., & Lee, R. M. (2014). You speak English well! Asian Americans' reactions to an exceptionalizing stereotype. *Journal of Counseling Psychology*, 61(3), 484–490. <https://doi.org/10.1037/cou0000034>
 - The source of our research vignette.

4.1.4 Packages

The packages used in this lesson are embedded in this code. When the hashtags are removed, the script below will (a) check to see if the following packages are installed on your computer and, if not (b) install them.

```
# will install the package if not already installed easy plotting for
# simple ANOVA if(!require(gplots)){install.packages('gplots')}
# creating new variables and other handy functions
# if(!require(tidyverse)){install.packages('tidyverse')} a specific
# part of the tidyverse with useful tools for manipulating data
# if(!require(dplyr)){install.packages('dplyr')} for descriptive
# statistics and writing them as csv files
# if(!require(psych)){install.packages('psych')} a number of wrappers
# for ANOVA models; today for evaluating the Shapiro
# if(!require(rstatix)){install.packages('rstatix')} produces effect
# sizes if(!require(lsrr)){install.packages('lsrr')} estimating sample
# sizes and power analysis if(!require(pwr)){install.packages('pwr')}
# produces an APA style table for ANOVAs and other models
# if(!require(apaTables)){install.packages('apaTables')} helps with
# formats like decimals and percentages for inline code
# if(!require(formattable)){install.packages('formattable')} more
# effect size options
# if(!require(effectsize)){install.packages('effectsize')}
```

4.2 Workflow for One-Way ANOVA

The following is a proposed workflow for conducting a one-way ANOVA.

1. Prepare (upload) data.
2. Explore data
 - graphs
 - descriptive statistics
3. Checking distributional assumptions
 - assessing normality via skew, kurtosis, Shapiro Wilks
 - checking for violation of homogeneity of variance assumption with Levene's test; if we violate this we can use Welch's omnibus ANOVA
4. Compute the omnibus ANOVA (remember to use Welch's if Levene's $p < .05$)
5. Compute post-hoc comparisons, planned contrasts, or polynomial trends
6. Managing Type I error
7. Sample size/power analysis (which you should think about first – but in the context of teaching ANOVA, it's more pedagogically sensible, here)

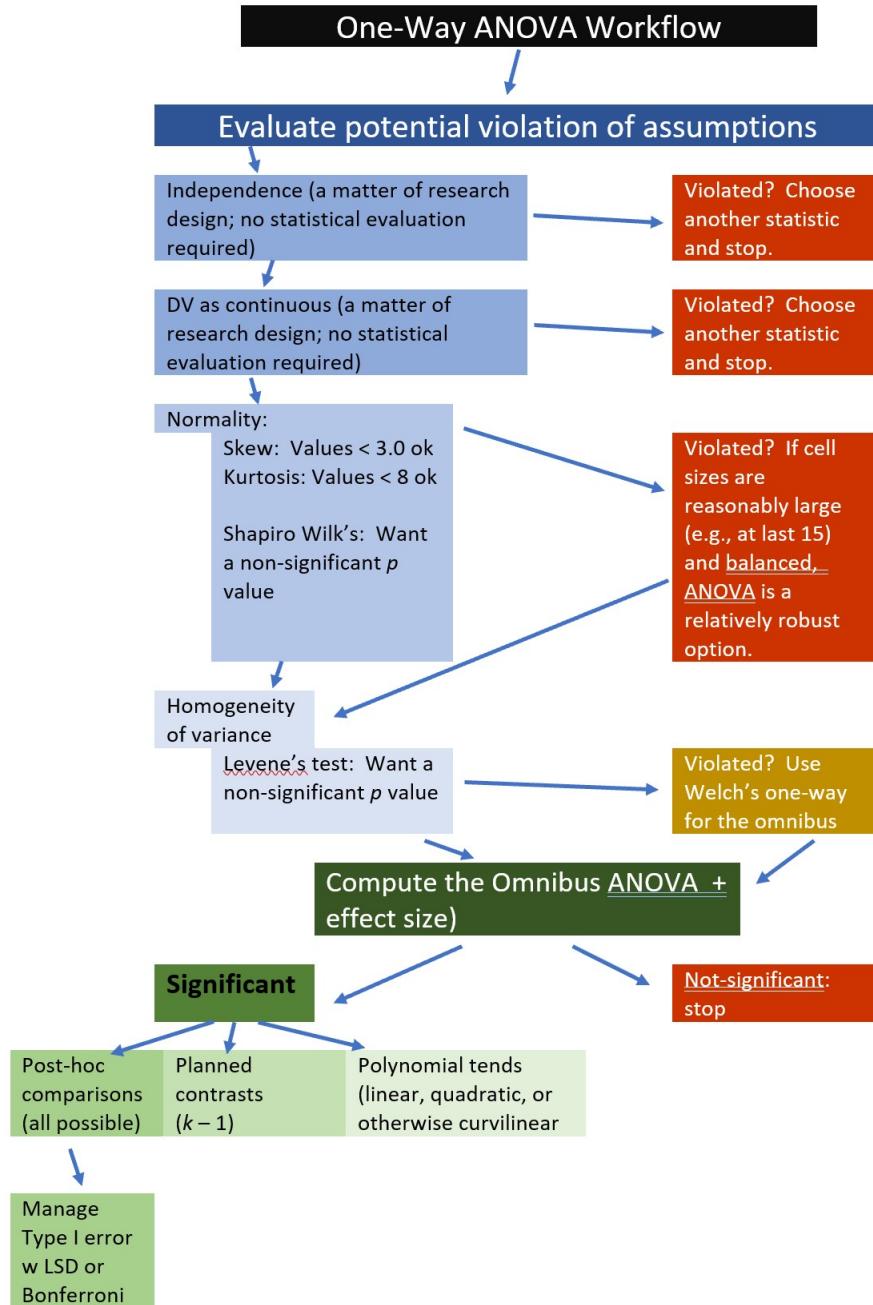


Figure 4.1: A colorful image of a workflow for the one-way ANOVA

4.3 Research Vignette

The *exceptionalizing racial stereotype* is microaggression framed as interpersonally complimentary, but perpetuates negative stereotypical views of a racial/ethnic group. We are using data that is *simulated* from a random clinical trial (RCT) conducted by Tran and Lee [2014].

The one-way ANOVA examples we are simulating represent the post-only design which investigated three levels of the exceptionalizing stereotype in a sample of Asian American participants. This experimental design involved a confederate (posing as a peer) whose parting comment fell into the low racial loading, high racial loading, or control conditions.

COND	Assignment	Manipulation	Post-test Observation
Low racial loading condition ($n = 22$)	Random	Yes: “Nice talking to you. You speak English well.”	Accurate
High racial loading ($n = 23$)	Random	Yes: “Nice talking to you. You speak English well for an Asian.”	Accurate
Control ($n = 23$)	Random	No: “Nice talking to you.”	Accurate

Tran and Lee [2014] reported results from two ANOVAs and 4 ANCOVAs, using a pre-test as a covariate. A preprint of their article is available [here](#).

- **Accurate** is the DV we will be exploring in this lesson. Participants rated how *accurate* they believed their partner’s impression of them was ($0 = \text{very inaccurate}$, $3 = \text{very accurate}$).
- **moreTalk** is the DV suggested as a practice problem. Participants rated how much longer they would continue the interaction with their partner compared to their interactions in general ($-2 = \text{much less than average}$, $0 = \text{average}$, $2 = \text{much more than average}$).

4.3.1 Data Simulation

Simulating data for a one-way ANOVA requires the sample size (rnorm), mean (mean), and standard deviation (sd) for each of the groups [Crump, 2018]. In creating this simulation, I used the data from Table 1 in the Tran and Lee [2014] article. Having worked the problem several times, I made one change. The group sizes in the original study were 23, 22, and 23. To increase the probability that we would have statistically significant results in our worked example, I increased the sample sizes to 30 for each group. In this way we have a perfectly *balanced* (equal cell sizes) design.

```
# Note, this script results in a different simulation than is in the
# ReadySetR lesson sets a random seed so that we get the same results
# each time
set.seed(210820)
# sample size, M and SD for each group
Accurate <- c(rnorm(30, mean = 1.18, sd = 0.8), rnorm(30, mean = 1.83,
```

```

sd = 0.58), rnorm(30, mean = 1.76, sd = 0.56))
# set upper bound for DV
Accurate[Accurate > 3] <- 3
# set lower bound for DV
Accurate[Accurate < 0] <- 0
# sample size, M and SD for each group
moreTalk <- c(rnorm(30, mean = -0.82, sd = 0.91), rnorm(30, mean = -0.39,
               sd = 0.66), rnorm(30, mean = -0.04, sd = 0.71))
# set upper bound for DV
moreTalk[moreTalk > 2] <- 2
# set lower bound for DV
moreTalk[moreTalk < -2] <- -2
# IDs for participants
ID <- factor(seq(1, 90))
# name factors and identify how many in each group; should be in same
# order as first row of script
COND <- c(rep("High", 30), rep("Low", 30), rep("Control", 30))
# groups the 3 variables into a single df: ID#, DV, condition
accSIM30 <- data.frame(ID, COND, Accurate, moreTalk)

```

4.4 Working the Problem

4.4.1 Preparing the Data

Examining the data is important for several reasons. First, we can begin to inspect for any anomalies. Second, if we are confused about what statistic we wish to apply, understanding the characteristics of the data can provide clues.

In R markdown we can

- look at the data by clicking on it, and
- examine its structure with the `str()` function.

Let's do both.

```
str(accSIM30)
```

```
'data.frame': 90 obs. of 4 variables:
 $ ID      : Factor w/ 90 levels "1","2","3","4",...: 1 2 3 4 5 6 7 8 9 10 ...
 $ COND    : chr "High" "High" "High" "High" ...
 $ Accurate: num 0.42 1.123 0.885 1.569 1.831 ...
 $ moreTalk: num -0.64 -2 -0.25 0.146 -0.996 ...
```

If we look at this simple dataset, we see that we see that

- `COND` is a grouping variable) with 3 levels (high, low, control)

- it is presently in “chr” (character) format, it needs to be changed to be a factor.
- **Accurate** is a continuous variable
 - it is presently in “num” (numerical) format, this is an appropriate format.
- **moreTalk** is a continuous variable
 - it is presently in “num” (numerical) format, this is an appropriate format

There are many ways to convert variables to factors; here is one of the simplest.

```
#convert variable to factor
accSIM30$COND <- factor(accSIM30$COND)
```

Let's recheck the structure

```
str(accSIM30)
```

```
'data.frame': 90 obs. of 4 variables:
 $ ID      : Factor w/ 90 levels "1","2","3","4",...: 1 2 3 4 5 6 7 8 9 10 ...
 $ COND    : Factor w/ 3 levels "Control","High",...: 2 2 2 2 2 2 2 2 2 2 ...
 $ Accurate: num  0.42 1.123 0.885 1.569 1.831 ...
 $ moreTalk: num  -0.64 -2 -0.25 0.146 -0.996 ...
```

By default, R orders factors alphabetically. This means, analyses will assume that “Control” (C) is the lowest condition, then “High,” then “Low.” Since these have theoretically ordered values, we want them in the order of “Control,” “Low,” “High.”

Here is the script to create an ordered factor. The order in which the variables are entered in the concatenated list (“c”) establishes the order (e.g., levels).

```
# ordering the factor
accSIM30$COND <- factor(accSIM30$COND, levels = c("Control", "Low", "High"))
```

Again, we can check our work.

```
#another structure check
str(accSIM30)
```

```
'data.frame': 90 obs. of 4 variables:
 $ ID      : Factor w/ 90 levels "1","2","3","4",...: 1 2 3 4 5 6 7 8 9 10 ...
 $ COND    : Factor w/ 3 levels "Control","Low",...: 3 3 3 3 3 3 3 3 3 3 ...
 $ Accurate: num  0.42 1.123 0.885 1.569 1.831 ...
 $ moreTalk: num  -0.64 -2 -0.25 0.146 -0.996 ...
```

Now our variables are suitable for analysis.

At this point, you may wish to export and/or import the data as a .csv (think “Excel lite”) or .rds (R object that preserves the information about the variables – such changing COND to an ordered factor), here is the code to do so. The data should save in the same folder as the .rmd file. Therefore, it is really important (think, “good R hygiene”) to have organized your folders so that your .rmd and data files are co-located.

I have hashtags out the code. If you wish to use it, delete the hashtags. Although I show the .csv code first, my personal preference is to save R data as .rds files. While they aren’t easy to “see” as an independent file, they retain the formatting of the variables. For a demonstration, refer back to the [Ready_Set_R](#) lesson.

```
# write the simulated data as a .csv write.table(accSIM30,
# file='accSIM.csv', sep=',', col.names=TRUE, row.names=FALSE) bring
# back the simulated dat from a .csv file acc_csv <- read.csv
# ('accSIM.csv', header = TRUE)
```

```
# a quick demo to show that the .csv format loses the variable
# formatting str(acc_csv)
```

```
# to save the df as an .rds (think 'R object') file on your computer;
# it should save in the same file as the .rmd file you are working
# with saveRDS(accSIM30, 'accSIM.rds') bring back the simulated dat
# from an .rds file acc_RDS <- readRDS('accSIM.rds')
```

```
# a quick demo to show that the .rds format preserves the variable
# formatting str(acc_RDS)
```

Note that I renamed each of these data objects to reflect the form in which I saved them (i.e., “acc_csv”, “acc_RDS”). If you have followed this step, you will want to rename the file before continuing with the rest of the chapter. Alternatively, you can start from scratch, re-run the code to simulate the data, and skip this portion on importing/exporting data.

```
#accSIM30 <- acc_RDS
#or
#accSIM30 <- acc_csv
```

4.4.2 Exploring the Distributional Characteristics Numerically

We will explore the data such that you will have several tools for future exploration. In this first demonstration I will quickly produce a mean and standard deviation.

These functions are in base R. The *aggregate()* function lets R know we want output by a grouping variable. We then list the variable of interest, a tilda (I think of the word “by”), and then the grouping variable (I think “Accurate by COND”). Finally we list the dataframe and the statistic (e.g., mean or standard deviation). R is case sensitive to check your capitalization if your code fails to execute.

```
aggregate(Accurate ~ COND, accSIM30, mean)
```

```
COND Accurate
1 Control 1.756195
2 Low 1.900116
3 High 1.152815
```

```
aggregate(Accurate ~ COND, accSIM30, sd)
```

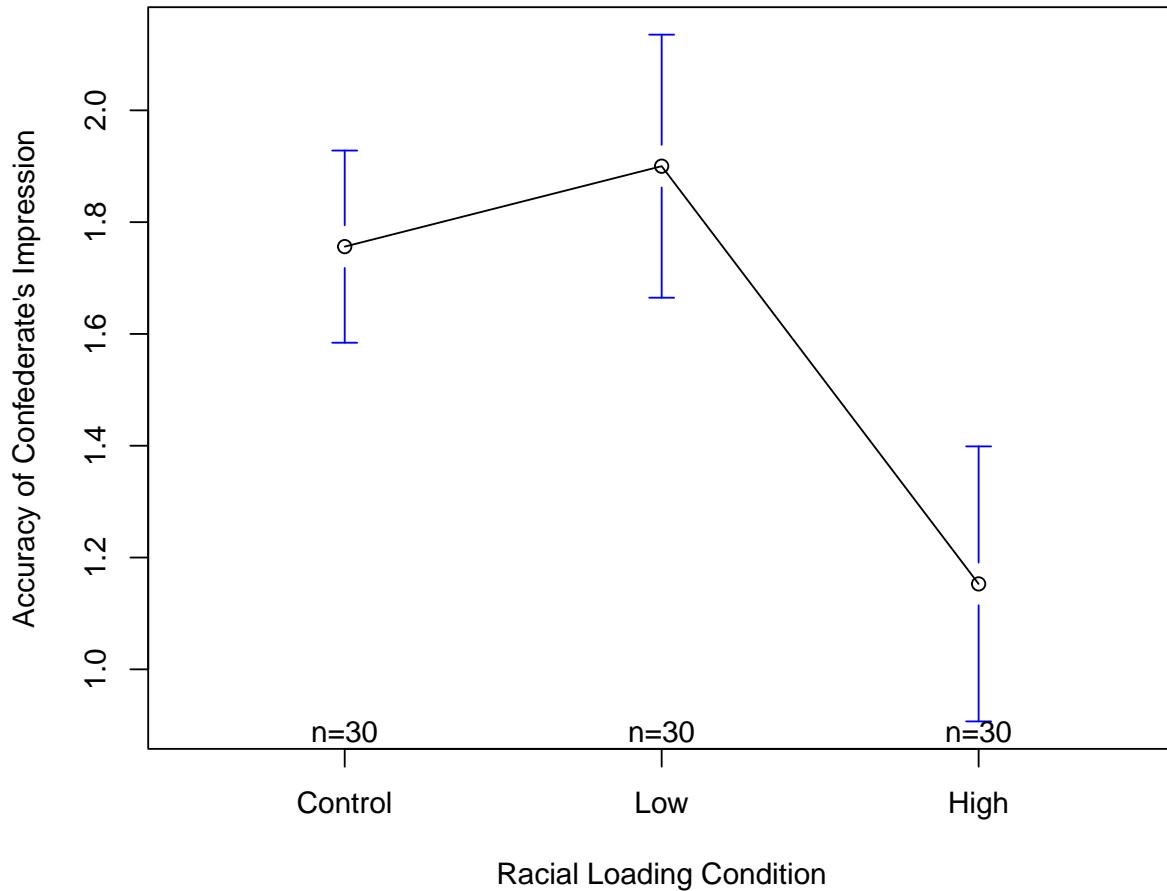
```
COND Accurate
1 Control 0.4603964
2 Low 0.6301138
3 High 0.6587486
```

Before looking at graphs, we can see that racially loaded *high* condition has the lowest accuracy score and the largest variability. Let's produce some helpful figures so that we can visualize this.

4.4.3 Exploring the Distributional Characteristics Graphically

The package *gplots* produces a simple line graph and the script is fairly intuitive. The *plotmeans()* function plots the means with error bars (95% confidence intervals) around the mean. Regarding the confidence intervals, we can think, “How confident are we that the mean is this particular value?” Earlier we noted that the “high racial loading condition” had the lowest mean and the widest variability. Is this apparent from the graph?

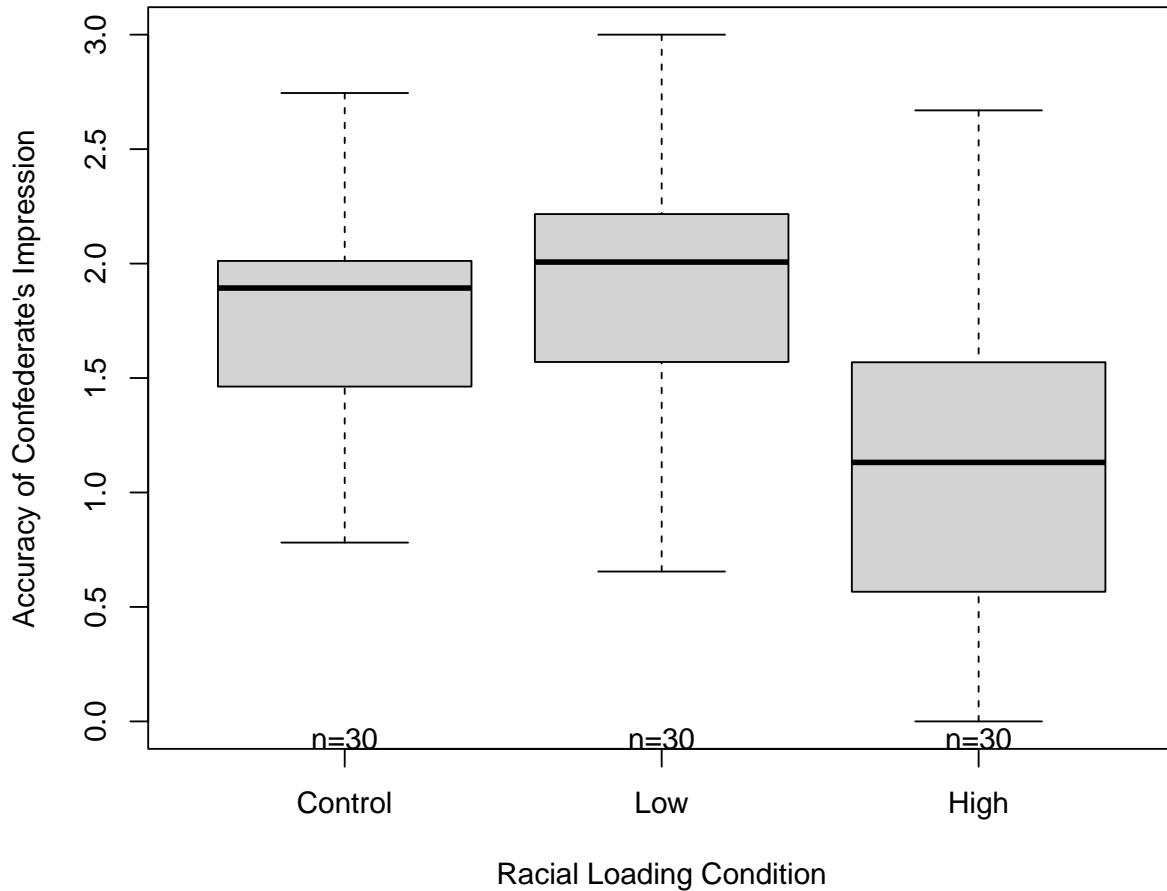
```
# plots DV by IV
gplots::plotmeans(formula = Accurate ~ COND, data = accSIM30, xlab = "Racial Loading Condition",
                  ylab = "Accuracy of Confederate's Impression", n.label = TRUE)
```



```
# this code could be more elegantly written in one row
plotmeans
# (formula = Accurate ~ COND, data = accSIM30, xlab = 'Racial Loading
# Condition', ylab = 'Accuracy of Confederate's Impression', n.label
# = TRUE)
```

Boxplots, with the *boxplot2()* function provide another view of our data. In boxplots the center value is the median. The box spans the *interquartile range* and ranges from the 25th to the 75th percentile. The whiskers cover 1.5 times the interquartile range. When this does not capture the entire range, outliers are represented with dots.

```
gplots::boxplot2(Accurate ~ COND, data = accSIM30, xlab = "Racial Loading Condition",
                 ylab = "Accuracy of Confederate's Impression", n.label = TRUE)
```



From both the boxplot and the linegraph with error bars, we can see that participants in the low racial loading condition have the highest accuracy ratings. This is followed by the control and then high racial loading conditions. Are these differences statistically significant? This is why we need the one-way ANOVA.

4.5 Understanding ANOVA with *Hand Calculations*

ANOVA was developed by Sir Ronald Fisher in the early 20th century. The name is a bit of a misnomer – rather than analyzing *variances*, we are investigating differences in *means* (but the formula does take variances into consideration...stay tuned).

ANOVA falls squarely within the tradition of **null hypothesis significance testing** (NHST). As such, a formal, traditional, ANOVA begins with statements of the null and alternate hypotheses. *Note. In their article, Tran and Lee [2014] do not list such.*

In our example, we would hypothesize that the population means (i.e., Asian or Asian American individuals in the U.S.) are equal:

$$H_O : \mu_1 = \mu_2 = \mu_3$$

There are an number of ways that the H_O could be false. Here are a few:

$$H_{a1} : \mu_1 \neq \mu_2 \neq \mu_3$$

$$H_{a2} : \mu_1 = \mu_2 > \mu_3$$

$$H_{a3} : \mu_1 > \mu_2 > \mu_3$$

The bottom line is that if we have a statistically significant omnibus ANOVA (i.e., the test of the overall significance of the model) and the H_O is false, somewhere between the three levels of the grouping factor, the means are statistically significantly different from each other.

In evaluating the differences between means, one-way ANOVA compares:

- systematic variance to unsystematic variance
- explained to unexplained variation
- experimental effect to the individual differences
- model variance to residual variance
- between group variance to within group variance

The ratio of these variances is the F -ratio.

Navarro [2020a] offers a set of useful figures to compare between- and within-group variation.

When between-group variance (i.e., model variance) is greater than within-group variance (i.e., residual variance) there may be support to suggest that there are statistically significant differences between groups.

Let's examine how variance is partitioned by hand-calculating sums of squares total, model, and residual. Along the way we will use some basic R skills to manipulate the data.

4.5.1 Sums of Squares Total

Sums of squares total represents the total amount of variance within our data. Examining the formula(s; there are variants of each) can help us gain a conceptual understanding of this.

In this first version of the formula we can see that the grand (or overall) mean is subtracted from each individual score, squared, and then summed. This makes sense: *sums of squares, total*.

$$SS_T = \sum (x_i - \bar{x}_{grand})^2$$

In the next version of the formula we see that the sums of square total is the addition of the sums of squares model and residual.

$$SS_T = SS_M + SS_R$$

“Between” and “within” are another way to understand “model” and “residual.” This is reflected in the next formula.

$$SS_T = SS_B + SS_W$$

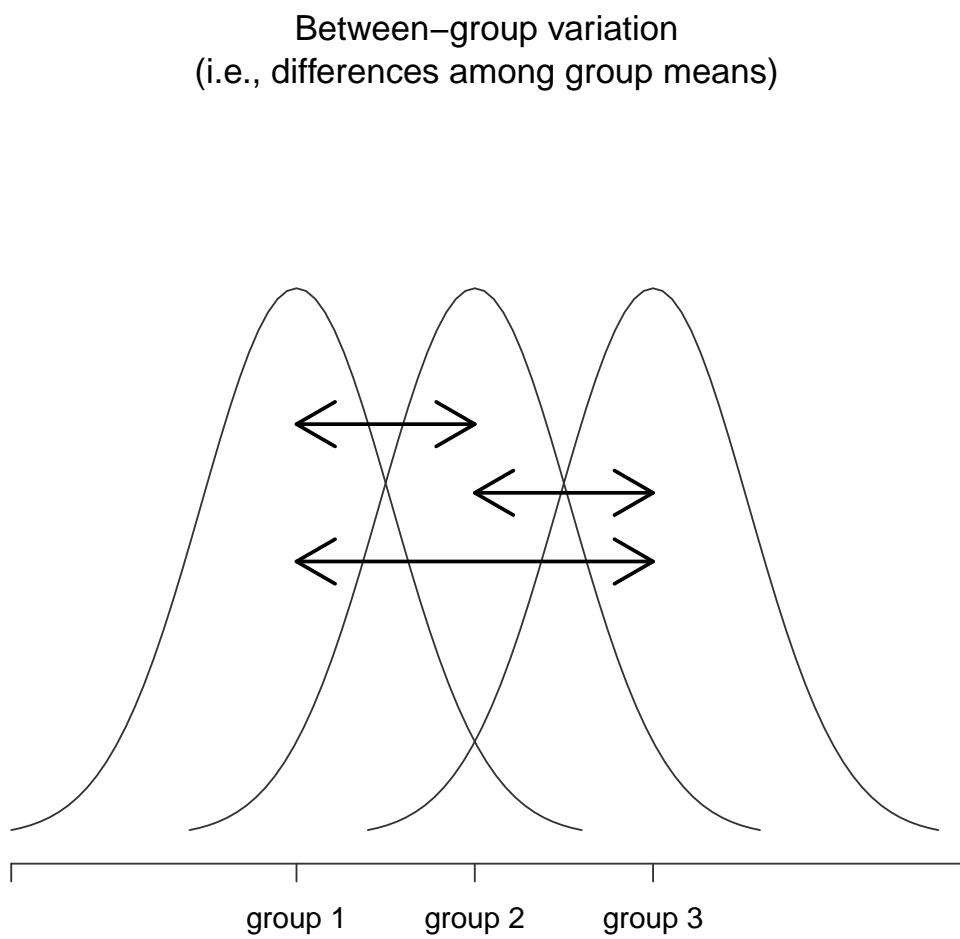


Figure 4.2: Graphical illustration of “between groups” variation

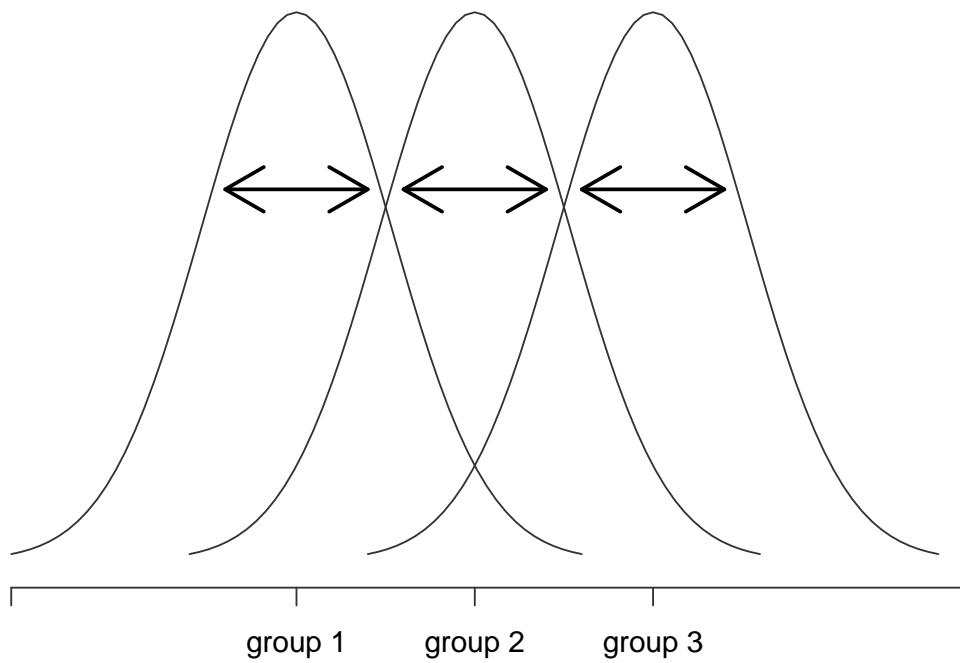


Figure 4.3: Graphical illustration of “within groups” variation

Finally, think of the sums of squares total as the grand variance multiplied by the overall degrees of freedom ($N - 1$).

$$SS_T = s_{grand}^2(n - 1)$$

Let's take a moment to *hand-calculate* SS_T . Not to worry – we'll get R to do the math for us!

Our grand (i.e., overall) mean is

```
GrandMean <- mean(accSIM30$Accurate)
GrandMean
```

```
[1] 1.603042
```

Subtracting the grand mean from each Accurate rating yields a mean difference. In the script below I have used the *mutate()* function from the *dplyr* package (a part of the *tidyverse*) to created a new variable ("m_dev") in the dataframe. The *tidyverse* package is one of the few exceptions that I will open via the library. This is because we need it if we are going to use the pipe (%>%) to string parts of our script together.

```
library(tidyverse)

accSIM30 <- accSIM30 %>%
  dplyr::mutate(m_dev = Accurate-mean(Accurate))

head(accSIM30)
```

	ID	COND	Accurate	moreTalk	m_dev
1	1	High	0.4203896	-0.6398265	-1.18265259
2	2	High	1.1226505	-2.0000000	-0.48039170
3	3	High	0.8852238	-0.2497750	-0.71781837
4	4	High	1.5689439	0.1455637	-0.03409829
5	5	High	1.8307196	-0.9960413	0.22767748
6	6	High	1.8874431	-1.0692978	0.28440098

Pop quiz: What's the sum of our new *m_dev* variable? Let's check.

```
mean(accSIM30$m_dev)
```

```
[1] 0.0000000000000003830065
```

Unless you run the script at the top of this document ("options(scipen=999")"), R will (seemingly selectively) use **scientific e notation** to report your results. The proper value is one where the base number (before the "e") is multiplied by 10, raised to the power shown: 3.830065×10^{17} Another way to think of it is to move the decimal 17 places to the left. In any case, this number is essentially zero.

Back to the point of sums of squares total, the sum of deviations around the grand mean will always be zero. To make them useful, we must square them:

```
accSIM30 <- accSIM30 %>%
  dplyr::mutate(m_devSQ = m_dev^2)

head(accSIM30)

  ID COND Accurate moreTalk      m_dev      m_devSQ
1  1 High 0.4203896 -0.6398265 -1.18265259 1.398667144
2  2 High 1.1226505 -2.0000000 -0.48039170 0.230776185
3  3 High 0.8852238 -0.2497750 -0.71781837 0.515263216
4  4 High 1.5689439  0.1455637 -0.03409829 0.001162694
5  5 High 1.8307196 -0.9960413  0.22767748 0.051837034
6  6 High 1.8874431 -1.0692978  0.28440098 0.080883915
```

If we sum the squared mean deviations we will obtain the total variance (sums of squares total):

```
SST <- sum(accSIM30$m_devSQ)
SST
```

```
[1] 39.67818
```

This value, the sum of squared deviations around the grand mean, is our SS_T . The associated *degrees of freedom* is $N - 1$; in our case this is $90-1 = 89$.

In one-way ANOVA, we divide SS_T into **model/between sums of squares** and **residual/within sums of squares**.

The *model* generally represents the notion that the means are different than each other. We want the variation between our means to be greater than the variation within each of the groups from which our means are calculated.

4.5.2 Sums of Squares for the Model (or Between)

We just determined that the total amount of variation within the data is 39.678 units. From this we can estimate how much of this variation our model can explain. SS_M tells us how much of the total variation can be explained by the fact that different data points come from different groups.

We see this reflected in the formula below, where

- the grand mean is subtracted from each group mean
- this value is squared and multiplied by the number of cases in each group
- these values are summed

$$SS_M = \sum n_k (\bar{x}_k - \bar{x}_{grand})^2$$

To calculate this, we start with the grand mean (previously calculated): 1.603.

We also estimate the group means.

```
GroupMeans <- aggregate(Accurate ~ COND, accSIM30, mean)
GroupMeans
```

COND	Accurate
1 Control	1.756195
2 Low	1.900116
3 High	1.152815

This formula occurs in three chunks, representing the control, low, and high racial loading conditions. In each of the chunks we have the n , group mean, and grand mean.

```
# Calculated by using object names from our calculations
SSM <- nControl * (ControlMean - GrandMean)^2 + nLow * (LowMean - GrandMean)^2 +
      nHigh * (HighMean - GrandMean)^2
SSM
```

```
[1] 9.432
```

```
# calculated by specifying the actual values from our calculations
30 * (1.756 - 1.603)^2 + 30 * (1.9 - 1.603)^2 + 30 * (1.153 - 1.603)^2
```

```
[1] 9.42354
```

```
# Both result in the same
```

This value, SS_M is the amount of variance accounted for by the model; that is, the the amount of variance accounted for by the grouping variable/factor, COND. Degrees of freedom for SS_M is always one less than the number of elements (e.g., groups) used in its calculation ($k - 1$). Because we have three groups, our degrees of freedom for the model is two.

4.5.3 Sums of Squares Residual (or within)

To recap, we know there are 39.678 units of variation to be explained in our data. Our model explains 9.432 of these units. Sums of squares residual tells us how much of the variation cannot be explained by the model. This value is influenced by extraneous factors; some will refer to it as “noise.”

Looking at the formula can assist us in with a conceptual formula. In SS_R we subtract the group mean from each individual member of the group and then square it.

$$SS_R = \sum (x_{ik} - \bar{x}_k)^2$$

Below is another approach to calculating SS_R . In this one the variance for each group is multiplied by its respective degrees of freedom, then summed.

$$SS_R = s_{group1}^2(n - 1) + s_{group2}^2(n - 1) + s_{group3}^2(n - 1)$$

Again, the formula is in three chunks – but this time the calculations are *within-group*. We need the variance (the standard deviation squared) for the calculation.

```
SDs <- aggregate (Accurate ~ COND, accSIM30, sd)
SDs
```

COND	Accurate
1 Control	0.4603964
2 Low	0.6301138
3 High	0.6587486

4.5.3.1 On the relationship between standard deviation and variance

Early in statistics training the difference between standard deviation (s or σ_{n-1}) and variance(s^2 or σ^2) can be confusing. This calculation demonstrates the relationship between standard deviation and variance. Variance is the standard deviation, squared.

```
#when squared, the standard deviation of the control group,
#should equal the variance reported in the next chunk
sdControl^2
```

```
[1] 0.212
```

```
VARs <- aggregate (Accurate ~ COND, accSIM30, var)
VARs
```

COND	Accurate
1 Control	0.2119648
2 Low	0.3970434
3 High	0.4339497

We will use the second formula to calculate SS_R . For each of the groups, we multiply the variance by the respective degrees of freedom for the group ($n - 1$).

```
# Calculated by using object names from our calculations
SSR <- varControl * (nControl - 1) + varLow * (nLow - 1) + varHigh * (nHigh -
1)
```

```
# Re-calculated by specifying the actual values from our calculations
SSR
```

```
[1] 30.246
```

```
0.212 * (30 - 1) + 0.397 * (30 - 1) + 0.434 * (30 - 1)
```

[1] 30.247

Both result in the same

The value for our SS_R is 30.246. Degrees of freedom for the residual is $df_T - df_M$.

- df_T was $N - 1: 90 - 1 = 89$
- df_M was $k - 1: 3 - 1 = 2$
- Therefore, df_R : is $89 - 2 = 87$

4.5.4 Relationship between SS_T , SS_M , and SS_R .

In case it is not clear:

$SS_T = 9.432 + 30.246$

#calculated with object names
 $SSM + SSR$

[1] 39.678

#Re-calculated with the actual values
 $9.432 + 30.247$

[1] 39.679

#Both result in the same

Our SST, calculated from above was 39.678.

4.5.5 Mean Squares Model & Residual

Our estimates of variation were *sums of squares* and are influenced by the number of scores that were summed. We can correct this bias by calculating their average – the *mean squares* or MS . We will use these in the calculation of the F ratio – the statistic that tests if there are significant differences between groups.

Like the constellation of sums of squares, we calculate mean squares for the model (MS_M) and residual(MS_R). Each formula simply divides the corresponding sums of squares by their respective degrees of freedom.

$$MS_M = \frac{SS_M}{df_M}$$

Regarding the calculation of our model mean squares:

- SS_M was 9.432
- df_M was 2
- Therefore, MS_M is:

```
#mean squares for the model
#calculated with object names
MSM <- SSM/dfM
MSM
```

[1] 4.716

```
#Re-calculated with actual values
9.432/2
```

[1] 4.716

```
#Both result in the same
```

$$MS_R = \frac{SS_R}{df_R}$$

Regarding the calculation of our model residual squares:

- SS_R was 30.246
- df_R was 87
- Therefore, MS_R is:

```
#mean squares for the residual
#calculated with object names
MSR <- SSR/ dfR
MSR
```

[1] 0.348

```
#calculated with actual values
30.247/87
```

[1] 0.3476667

```
#Both result in the same
```

4.5.6 Calculating the F Statistic

The F statistic (or F ratio) assesses the ratio (as its name implies) of variation explained by the model to unsystematic factors (i.e., the residual). Earlier we used “between” and “within” language. Especially when we think of our example – where the model is composed of three groups, we can think of the F statistic as assessing the ratio of variation explained by between-subjects differences to within-subjects differences. Navarro’s [2020b] figures (earlier in the chapter) illustrate this well.

$$F = \frac{MS_M}{MS_R}$$

Regarding the calculation of our F -ratio:

- MS_M was 4.716
- MS_R was 0.348
- Therefore, F is:

```
#calculated with object names
Fratio <- MSM / MSR
Fratio
```

[1] 13.566

```
#calculated with actual values
#Both result in the same
4.716/0.348
```

[1] 13.55172

4.5.7 Source Table Games

These last few calculations are actually less complicated than this presentation makes them seem. To better understand the relation between sums of squares, degrees of freedom, and mean squares, let’s play a couple of rounds of *Source Table Games*!

Rules of the game:

- In each case, mean squares are determined by dividing the sums of squares by its respective degrees of freedom.
- The F statistic is determined by dividing MS_M by MS_R

Knowing only two of the values, challenge yourself to complete the rest of the table. Before looking at the answers (below), try to fill in the blanks based on what we have learned so far.

Game	Total (df, $N - 1$)	Model (df, $k - 1$)	Residual (df, $df_T - df_M$)
SS	39.678(89)	9.432(2)	_____
MS	NA	_____	_____

$$F = MS_M/MS_R = \underline{\hspace{2cm}}$$

DON'T PEEK! TRY TO DO THE CALCULATIONS IN THE “SOURCE TABLE GAMES” EXERCISE BEFORE LOOKING AT THESE ANSWERS

Answers	Total (df, $N - 1$)	Model (df, $k - 1$)	Residual (df, $df_T - df_M$)
SS	39.678(89)	9.432(2)	30.246(87)
MS	NA	4.716	0.348

$$F = MS_M/MS_R = 13.566$$

To determine whether or not it is statistically significant, we can check a [table of critical values \[Zach, 2019\]](#) for the F test.

Our example has 2 (numerator) and 87 (denominator) degrees of freedom. Rolling down to the table where $\alpha = .05$, we can see that any F value > 3.11 (a value somewhere between 3.07 and 3.15) will be statistically significant. Our $F = 13.566$, so we have clearly exceeded the threshold. This is our *omnibus F test*.

We can also use a look-up function, which follows this general form: `qf(p, df1, df2, lower.tail=FALSE)`

```
qf(.05, 2, 87, lower.tail=FALSE)
```

```
[1] 3.101296
```

Significance at this level lets us know that there is at least 1 statistically significant difference between our control, low, and high racially loaded conditions. While it is important to follow-up to see where these significant differences lie, we will not do these by hand. Rather, let's rework the problem in R.

4.6 Working the One-Way ANOVA in R

Let's rework the problem in R. We start at the top of the flowchart, evaluating the statistical assumptions.

4.6.1 Evaluating the Statistical Assumptions

All statistical tests have some assumptions about the data. The one-way ANOVA has four assumptions:

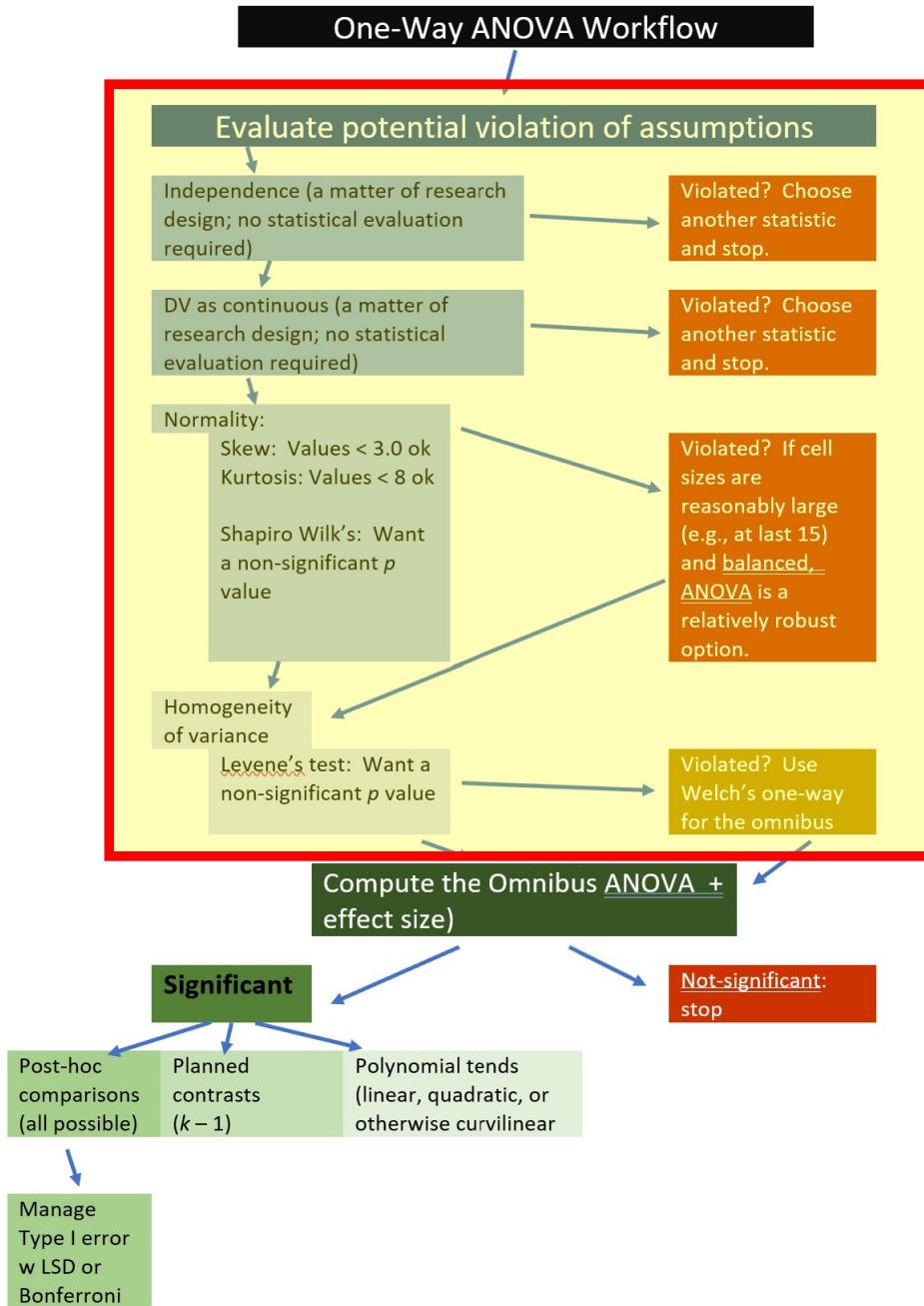


Figure 4.4: An image of the workflow for one-way ANOVA, showing that we are at the beginning: evaluating the potential violation of the assumptions.

- The dependent variable is normally distributed for each of the populations as defined by the different levels of the factor. We will examine this by
 - evaluating skew and kurtosis
 - visually inspecting the distribution
 - conduct a Shapiro Wilks test
 - examine a QQ plot
- The variances of the dependent variable are the same for all populations. This is often termed the *homogeneity of variance* assumption. We will examine this with
 - Levene's Test
- The cases represent *random* samples from the populations and scores on the test variable are *independent* of each other. That is, comparing related cases (e.g., parent/child, manager/employee, time1/time2) violates this assumption and this question would need to be evaluated by a different statistic such as repeated measures ANOVA or dyadic data analysis.
 - *Independence* in observations is a research design issue. ANOVA is not robust to violating this assumption. When observations are correlated/dependent there is a dramatic increase in Type I error.
- The dependent variable is measured on an interval scale.
 - If the dependent variable is categorical, another statistic (such as logistic regression) should be chosen.

4.6.1.1 Is the dependent variable normally distributed across levels of the factor?

From the *psych* package, the *describe()* function can be used to provide descriptive statistics (or, “descriptives”) of continuously scaled variables (i.e., variables measured on the interval or ratio scale). In this simple example, we can specify the specific continuous, DV.

```
#we name the function
#in parentheses we list data source
psych::describe(accSIM30$Accurate)
```

	vars	n	mean	sd	median	trimmed	mad	min	max	range	skew	kurtosis	se
X1	1	90	1.6	0.67	1.73	1.62	0.68	0	3	3	-0.28	-0.48	0.07

If we want descriptives for each level of the grouping variable (factor), we can use the *describeBy()* function of the *psych* package. The order of entry within the script is the DV followed by the grouping variable (IV).

```
# It is unnecessary to create an object, but an object allows you to
# do cool stuff, like write it to a .csv file and use that as a basis
# for APA style tables In this script we can think 'Accurate by COND'
# meaning that the descriptives for accuracy will be grouped by COND
# which is a categorical variable mat = TRUE presents the output in
```

```

# matrix (table) form digits = 3 rounds the output to 3 decimal
# places data = accSIM30 is a different (I think easier) way to
# identify the object that holds the dataframe
des.mat <- psych::describeBy(Accurate ~ COND, mat = TRUE, digits = 3, data = accSIM30)
# Note. Recently my students and I have been having intermittent
# struggles with the describeBy function in the psych package. We
# have noticed that it is problematic when using .rds files and when
# using data directly imported from Qualtrics. If you are having
# similar difficulties, try uploading the .csv file and making the
# appropriate formatting changes. displays the matrix object that we
# just created
des.mat

    item group1 vars   n  mean     sd median trimmed   mad   min   max range
X11     1 Control    1 30 1.756 0.460  1.893   1.767 0.392 0.781 2.745 1.964
X12     2      Low    1 30 1.900 0.630  2.007   1.918 0.458 0.655 3.000 2.345
X13     3     High    1 30 1.153 0.659  1.131   1.128 0.743 0.000 2.669 2.669
          skew kurtosis     se
X11 -0.275   -0.537 0.084
X12 -0.378   -0.465 0.115
X13  0.208   -0.690 0.120

# optional to write it to a .csv file
write.csv(des.mat, file = "Table1.csv")

```

Skew and kurtosis speaks to normal distributions. The skew and kurtosis indices in the *psych* package are reported as *z* scores. Regarding skew, values greater than 3.0 are generally considered “severely skewed.” Regarding kurtosis, “severely kurtotic” is argued to be anywhere greater 8 to 20 [Kline, 2016].

The *Shapiro-Wilks* test evaluates the hypothesis that the distribution of the data as a whole deviates from a comparable normal distribution. If the test is non-significant ($p > .05$) the distribution of the sample is not significantly different from a normal distribution. If, however, the test is significant ($p < .05$), then the sample distribution is significantly different from a normal distribution. The *rstatix* package has a wrapper that can conduct the Shapiro-Wilks test for us.

```

shapiro <- accSIM30 %>%
  group_by(COND) %>%
  rstatix::shapiro_test(Accurate)
shapiro

```

```

# A tibble: 3 x 4
  COND    variable statistic     p
  <fct>  <chr>        <dbl> <dbl>
1 Control Accurate     0.954 0.215
2 Low      Accurate     0.944 0.115
3 High     Accurate     0.980 0.831

```

The p values for the distributions of the dependent variable (accurate) in each of the three conditions are all well above .05. This tells us that the Accurate variable does not deviate from a statistically significant distribution (Control, $W = 0.954$, $p = 0.215$; Low, $W = 0.944$, $p = 0.115$; High, $W = 0.980$, $p = 0.831$).

There are limitations to the Shapiro-Wilks test. As the dataset being evaluated gets larger, the Shapiro-Wilks test becomes more sensitive to small deviations; this leads to a greater probability of rejecting the null hypothesis (null hypothesis being the values come from a normal distribution). Green and Salkind [2014b] advised that ANOVA is relatively robust to violations of normality if there are at least 15 cases per cell and the design is reasonably balanced (i.e., equal cell sizes).

4.6.1.2 Are the variances of the dependent variable similar across the levels of the grouping factor?

The Levene's test evaluates the ANOVA assumption that variances of the dependent variable for each level of the independent variable are similarly distributed. We want this to be non-significant ($p > .05$). If violated, we need to use an ANOVA test that is “robust to the violation of the homogeneity of variance” (e.g., Welch's oneway).

In R, Levene's test is found in the *car* package.

```
# Our set up is similar: Accurate by condition, followed by the
# object that holds the data frame, followed by the instruction to
# center the analysis around the mean
levene <- car::leveneTest(Accurate ~ COND, accSIM30, center = mean)
levene
```

```
Levene's Test for Homogeneity of Variance (center = mean)
  Df F value Pr(>F)
group  2  1.5315  0.222
      87
```

We write the result of the Levene's as $F(2,87) = 1.532$, $p = 0.222$. Because $p > .05$, we know that the result is nonsignificant – that the variances of the three groups are not statistically significantly different than each other.

If the results had been statistically significantly different, we would have needed to use a Welch's F or robust version of ANOVA.

4.6.1.3 Summarizing results from the analysis of assumptions

It is common for an APA style results section to begin with a review of the evaluation of the statistical assumptions. As we have just finished these analyses, I will document what we have learned so far:

A one-way analysis of variance was conducted to evaluate the relationship between degree of racial loading of an exceptionalizing microaggression and the perceived accuracy of a research confederate's impression of the Asian or Asian American participant. The independent variable, condition,

included three levels: control/none, low, and high levels of racial loading. Results of Levene's homogeneity of variance test indicated no violation of the homogeneity of variance assumption ($F[2,87] = 1.532, p = 0.222$). Similarly, results of the Shapiro Wilk's test indicated no violation of the normality assumption in each of the cells (control, $W = 0.954, p = 0.215$; low, $W = 0.944, p = 0.115$; high, $W = 0.980, p = 0.831$).

Now we can move onto computing the omnibus ANOVA. *Omnibus* is the term applied to the first F test that evaluates if all groups have the same mean [Chen et al., 2018]. If this test is not significant there is no evidence in the data to reject the null; that is, there is no evidence to suggest that group means are different. If it is significant – and there are three or more groups – follow-up testing will be needed to determine where the differences lie.

4.6.2 Computing the Omnibus ANOVA

Having met all the assumptions, we are now ready to calculate the omnibus F test.

ANOVA is a special case of the general linear model (regression is a “not so special case” of the general linear model), therefore we use the linear model function, `aov()` to run the analysis.

In the code below, we predict Accuracy from COND (3 levels: control, low, high).

By assigning the results of the `aov()` function to an object (omnibus) we can then use that object (think *model*) in other functions to get details about our analysis.

```
# the script looks familiar, 'Accurate by Condition' DV ~ IV I say,
# 'DV by IV'
omnibus <- aov(Accurate ~ COND, data = accSIM30)
# prints the ANOVA output
summary(omnibus)
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)						
COND	2	9.432	4.716	13.57	0.00000745 ***						
Residuals	87	30.246	0.348								

Signif. codes:	0	'***'	0.001	'**'	0.01	'*'	0.05	'. '	0.1	' '	1

Inserting the `aov()` object (omnibus) into the summary command produces the ANOVA Source Table that we manually created above.

The values we see map onto those we calculated by hand. Our SS_M (9.432) plus SS_R (30.246) sum to equal the SS_T (39.678).

Dividing the two sums of squares by their respective degrees of freedom produces the means squared.

Then, dividing the MS_M (COND) by MS_R (4.716/0.348) provides the F ratio. By using a table of F critical values, we already knew that our F value exceeded the value in the table of critical values. Here we see that $p = 0.000$.

The “ F string” for an APA style results section should be written like this: $F (2, 87) = 13.566, p < .001$.

The object we created with the `aov()` function is capable of producing much information. Applying the `names()` function to the object can give us a list of values within it.

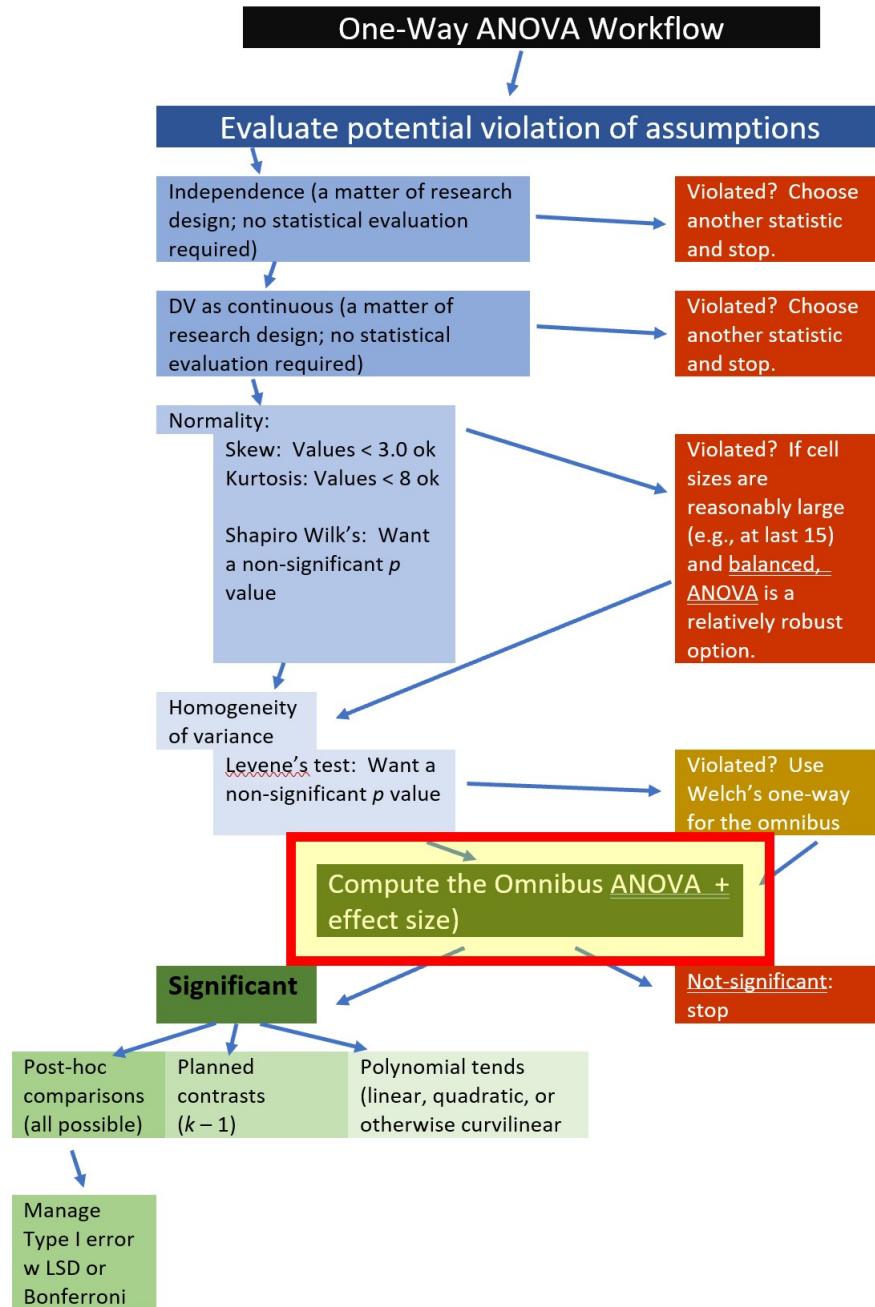


Figure 4.5: An image of the workflow for one-way ANOVA, showing that we are at the stage of computing the omnibus ANOVA.

```
names(omnibus)
```

```
[1] "coefficients"   "residuals"       "effects"        "rank"  
[5] "fitted.values" "assign"          "qr"             "df.residual"  
[9] "contrasts"     "xlevels"         "call"           "terms"  
[13] "model"
```

One of the most commonly used functions to be applied to the *aov()* objects is *summary()*; but it's not the only one. Let's try some other options.

```
model.tables (omnibus, "means")
```

Tables of means

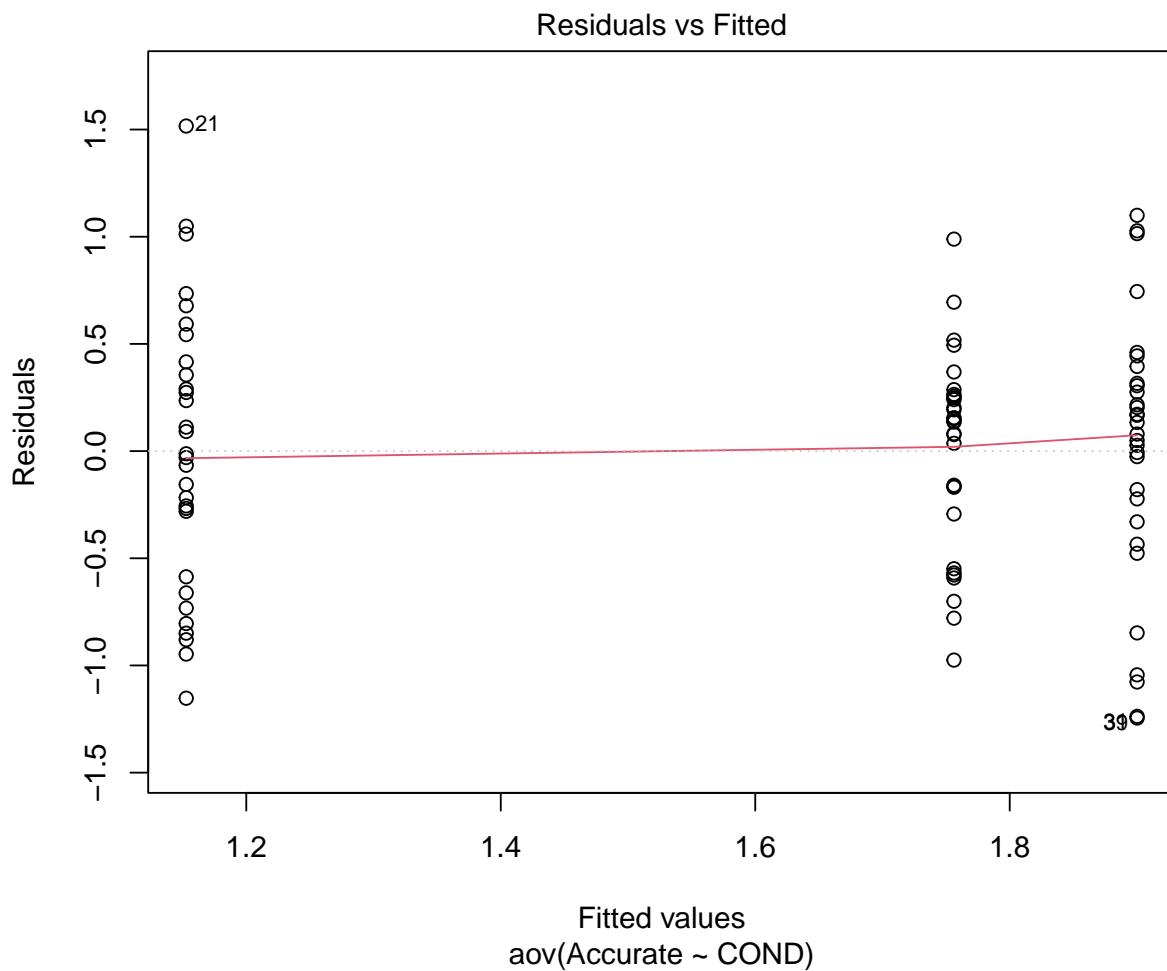
Grand mean

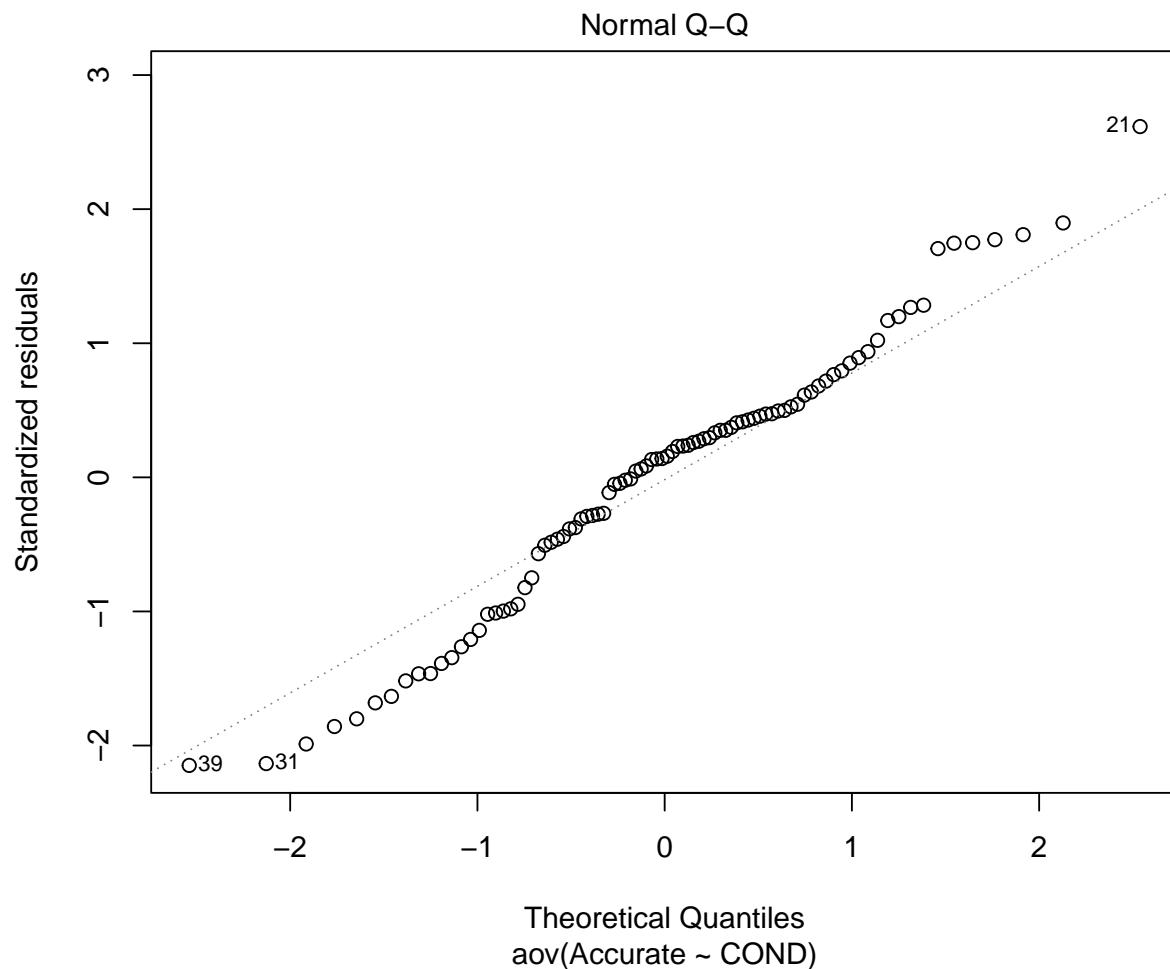
1.603042

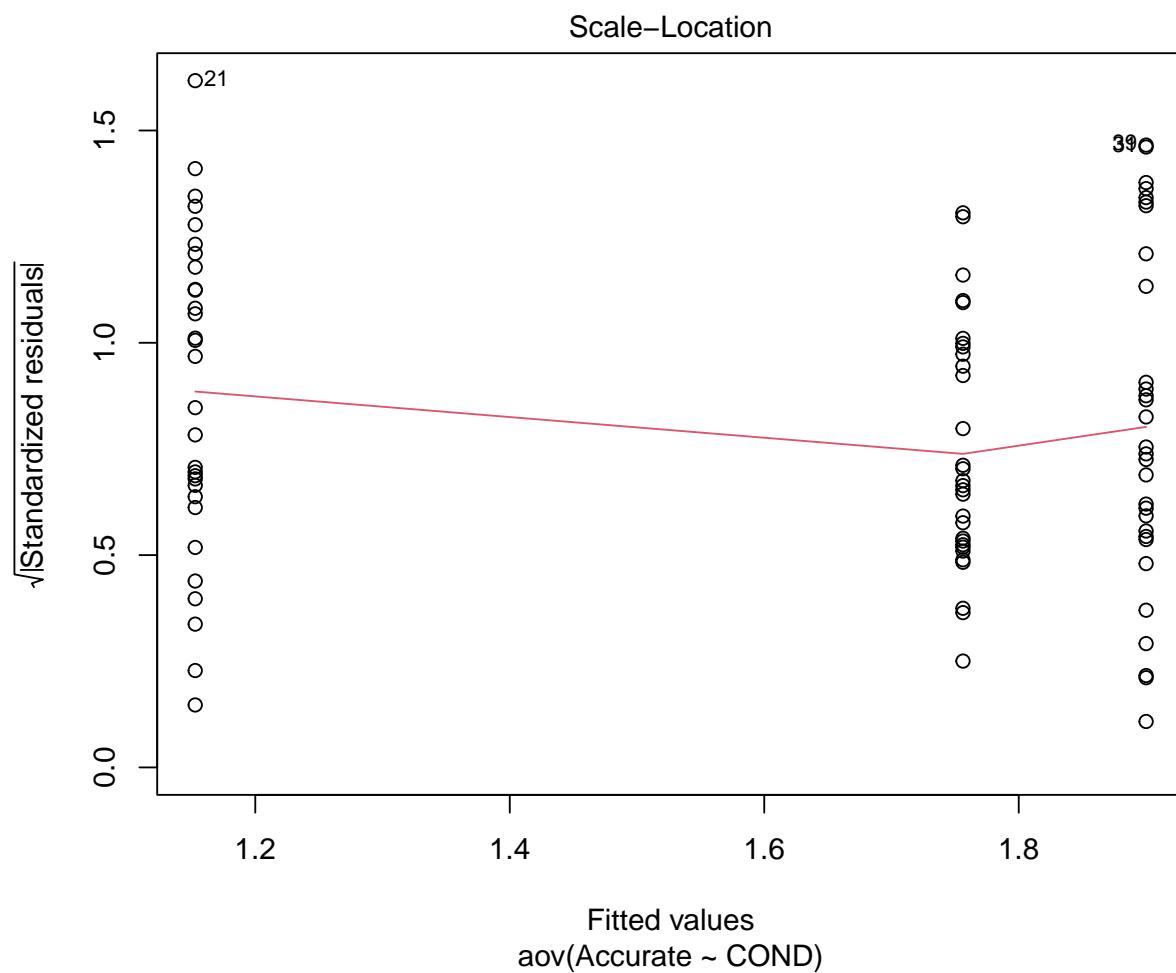
```
COND  
COND  
Control      Low      High  
1.7562  1.9001  1.1528
```

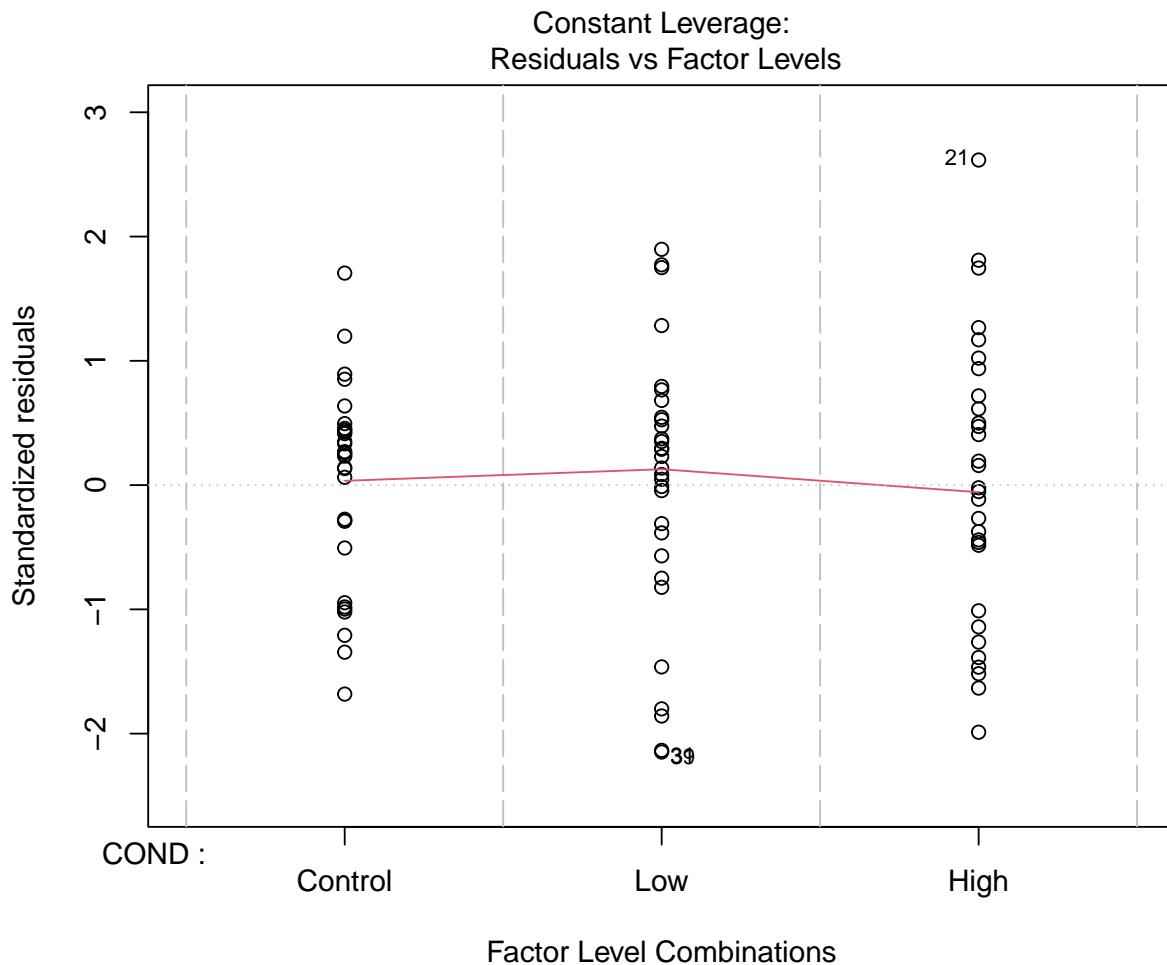
The *aov()* command has a quickplot feature.

```
plot(omnibus)
```









The first of the four plots fits the residuals. We already know from Levene's that we did not violate the homogeneity of variance test. With its straight line, this plot shows an equal spread across the three groups.

When the dots of the Q-Q plot map onto the diagonal, we have some indication of normality of the residuals (we want residuals to be normally distributed).

4.6.2.1 Effect size for the one-way ANOVA

Eta squared is one of the most commonly used measures of effect. It refers to the proportion of variability in the dependent variable/outcome that can be explained in terms of the independent variable/predictor. Traditional interpretive values are similar to the Pearson's r :

- 0.0 = no relationship
- .02 = small
- .13 = medium
- .26 = large

- $1.0 =$ a perfect (one-to-one) correspondence

A useful summary of effect sizes, guide to interpreting their magnitudes, and common usage can be found [here \[Watson, 2020\]](#).

The formula for η^2 is straightforward. If we think back to our hand-calculations of all the sums of squares, we can see that this is the proportion of variance that is accounted for by the model.

$$\eta^2 = \frac{SS_M}{SS_T}$$

Hand calculation, then, is straightforward.:

```
#Calculated using the object names
SSM omnibus / (SSM omnibus + SSR omnibus)
```

```
[1] 0.238
```

```
#Re-calculated by using the numeric values
9.432/(9.432 + 30.246)
```

```
[1] 0.2377136
```

```
#Both options should produce the same result
```

The *lsr* package includes an eta-squared calculator. To use it, we simply insert the model/object we created with the *aov()* function to *lsr*'s *etaSquared()* function.

```
lsr::etaSquared(omnibus)
```

```
eta.sq eta.sq.part
COND 0.2377226 0.2377226
```

Notice that there are two effect sizes. We described eta-squared. Partial eta-squared is the default effect size reported in SPSS. There is a long history of debate about which to use. In certain circumstances (especially in more complicated analyses), partial-eta squared can be a bit more generous (i.e., larger than η^2). Thus, many prefer the reporting of the more cautious η^2 .

In our case, we see no difference between the two values. Differences begin to appear in datasets that are more complicated, such as when sample sizes across the levels of a factor differ.

4.6.2.2 Summarizing results from the omnibus ANOVA

Presenting the APA style results of the omnibus test is very straightforward:

Results indicated a significant effect of COND on accuracy perception $F(2, 87) = 13.566, p < .001$, $\eta^2 = 0.238$.

4.6.3 Follow-up to the Omnibus F

The F -test associated with the one-way ANOVA is the *omnibus* – giving the result for the overall test. Looking at the workflow for the one-way ANOVA we see that if we had had we had a non-significant F , we would have stopped our analysis.

However, if the omnibus F is significant, we know that there is at least one pair of cells where there is a statistically significant difference. We have several ways (each with its own strengths/limitations) to figure out where these differences lie.

A very common option is to conduct post-hoc, pairwise comparisons.

4.6.3.1 OPTION 1: Post-hoc, pairwise, comparisons

Post-hoc, pairwise comparisons are:

- used for exploratory work when no firm hypotheses were articulated a priori,
- used to compare the means of all combinations of pairs of an experimental condition,
- less powerful than planned comparisons b/c strict criterion for significance must be used.

Helpful information about how to conduct post-hoc pairwise comparisons in R can be found at the [UCLA Institute for Digital Research and Education site \[noa\]](#).

```
pairwise.t.test(accSIM30$Accurate, accSIM30$COND, p.adj = "none")
```

```
Pairwise comparisons using t tests with pooled SD

data: accSIM30$Accurate and accSIM30$COND

Control Low
Low 0.34709 -
High 0.00015 0.0000042

P value adjustment method: none
```

```
#can swap "bonf" or "holm" for p.adj
```

The output only provides the p values associated with the mean differences in each of the conditions. We see that $p < .05$ when high is compared to control (0.00015) and high is compared to low (0.0000042). An APA style reporting results of these typically involves referencing the means (often reported in a table of means and standard deviations) or mean differences (hand calculated) with their p values.

Should we be concerned about Type I error?

Recall that *Type I error* is the concern about false positives – that we would incorrectly reject a true null hypothesis (that we would say that there is a statistically significant difference when there is not one). This concern increases when we have a large number of pairwise comparisons.

Green and Salkind [2014b] reviewed three options for managing Type I error.

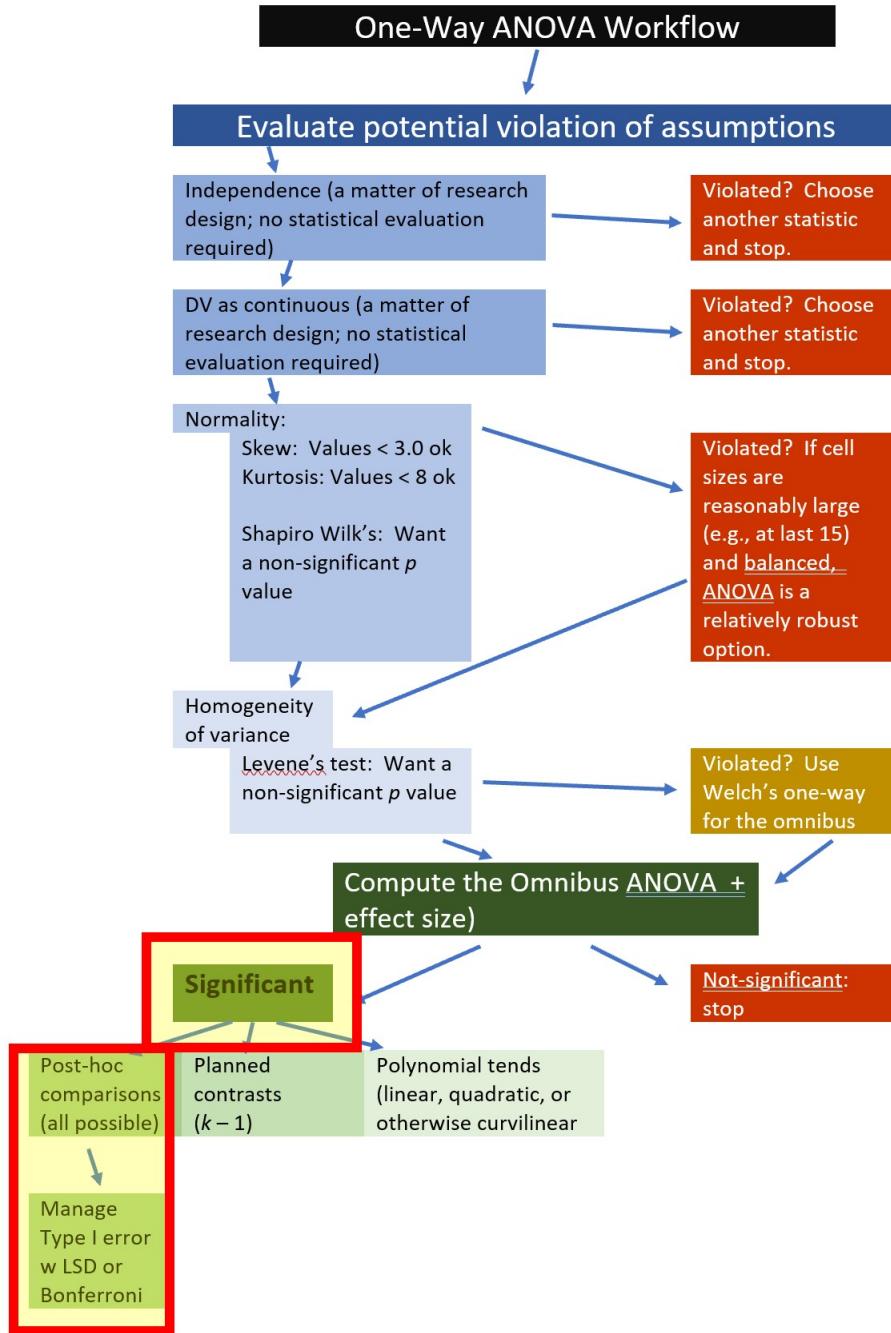


Figure 4.6: An image of the workflow for one-way ANOVA, showing that we are at the stage of following a statistically significant omnibus F test and are now conducting posthoc comparisons.

- Traditional Bonferroni:
 - Adjusts the p value upward by multiplying it (the raw p values) by the number of comparisons being completed. This holds the *total* Type I error rate across these tests to α (usually .05). The traditional Bonferroni is simple and therefore attractive, but when p values hover around .05, it can be too restrictive.
- Holms Sequential Bonferroni:
 - We'll describe this in more detail later. Briefly, it allows us to rank order the comparisons by their p value (smallest to largest). We determine the significance of each p value *sequentially* where the criteria for significance is incrementally relaxed.
- LSD method:
 - Permitted when there are only three pairwise comparisons among three groups, researchers can leave the p values as they are. Since the Tran and Lee [2014] research vignette is one of those circumstances, I will not make adjustments for Type I error. That is, I would claim the LSD and cite Green and Salkind [2014b] as justification for that decision.

There is, though, an even more powerful approach...

4.6.3.2 OPTION 2: Planned contrasts (non-orthogonal)

Another option is to evaluate planned comparisons.

Planned comparisons are

- theory-driven comparisons constructed prior to data collection,
- based on the idea of partitioning the variance created by the overall effect of group differences into gradually smaller portions of variance, and
- more powerful than post-hoc tests.

Planned contrasts involve further considerations regarding the *partitioning of variance*.

- There will always be $k-1$ contrasts.
- Each contrast must involve only two chunks of variance.

Orthogonal contrasts are even more sophisticated. Essential to conducting an orthogonal contrast is the requirement that if a group is singled out in one comparison it should be excluded from subsequent contrasts. The typical, orthogonal scenario with three ordered groups has only two contrasts:

1. Control versus low and high (because control was excluded, it should not reappear in the next contrast)
2. Low versus high

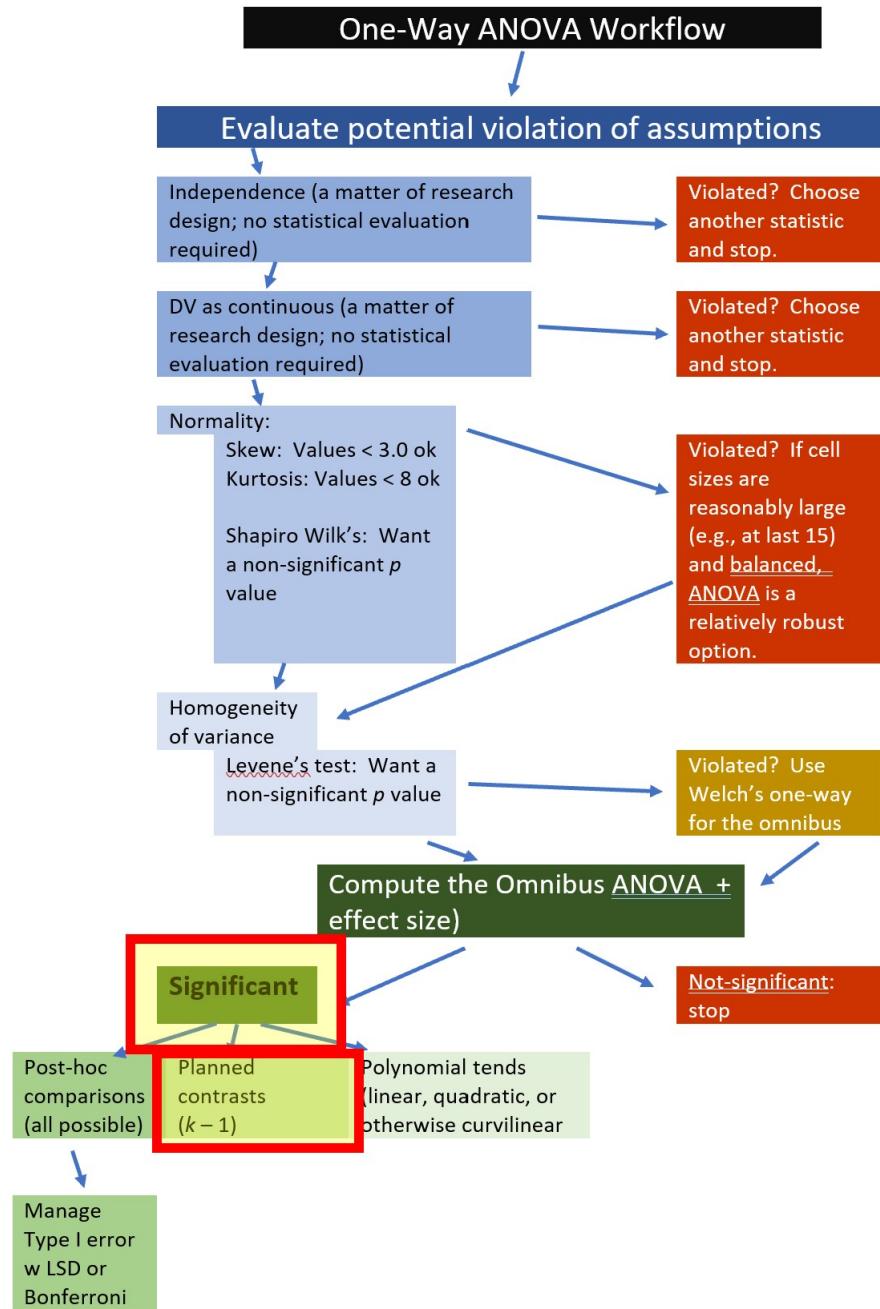


Figure 4.7: An image of the workflow for one-way ANOVA, showing that we are up to a significant omnibus F by conducting planned comparisons

Underlying the `aov()` program is the linear model (“lm”). We could have used it to calculate the omnibus ANOVA, but it has clunky output.

We use it now to retrieve some contrast information. The code below is a planned comparison that uses the coding in the database (created when we formatted COND as an ordered factor) to compare the lowest coded group (control was 1, low was 2, high was 3) to the other two groups.

```
summary.lm(omnibus)
```

```
Call:
aov(formula = Accurate ~ COND, data = accSIM30)

Residuals:
    Min      1Q  Median      3Q     Max 
-1.24533 -0.32092  0.08642  0.30101  1.51646 

Coefficients:
            Estimate Std. Error t value     Pr(>|t|)    
(Intercept)  1.7562     0.1076 16.314 < 0.0000000000000002 *** 
CONDLow       0.1439     0.1522   0.945      0.347095    
CONDHigh     -0.6034     0.1522  -3.963      0.000151 ***  
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1 

Residual standard error: 0.5896 on 87 degrees of freedom
Multiple R-squared:  0.2377,    Adjusted R-squared:  0.2202 
F-statistic: 13.57 on 2 and 87 DF,  p-value: 0.000007446
```

From the above, regression output, we see that there was not a statistically significant difference between CONDControl and CONDLow ($p = 0.347$). There were statistically significant difference when CONDControl (the intercept) was compared to CONDHIGH ($p < .001$).

Note that these tests are conducted as t tests. Why? We are comparing only two groups and can use the t distribution.

Also note that these involved the comparison of the control group to low; then the control to high. This is consistent with an orthogonal contrast in that there are $k - 1$ contrasts (two contrasts with three original groups). However it is inconsistent with the requirement that once a group is singled out, it should not be used in a subsequent contrast. Therefore, it is quite possible we want something different.

4.6.3.3 OPTION 3: Planned contrasts (orthogonal)

Step 1: Specify our contrasts

- Specifying the contrasts means you know their order within the factor
- Early in the data preparation, we created an ordered factor with Control, Low, High as the order.

- We want orthogonal contrasts, this means there will be
 - $k - 1$ contrasts; with three groups we will have two contrasts
 - once we single out a condition for comparison, we cannot use it again.

In *contrast1* we compare the control condition to the combined low and high conditions. In *contrast2* we discard the control condition (it was already singled out) and we compare the low and high conditions.

This is sensible because we likely hypothesize that any degree of racially loaded stereotypes may have a deleterious outcome, so we first compare control to the two conditions with any degree of racial loading. Subsequently, we compare the low and high levels of the factor.

Step 2: Bind them together and check the output to ensure that we've mapped them correctly.

```
# Contrast1 compares Control against the combined effects of Low and
# High.
contrast1 <- c(-2, 1, 1)

# Contrast2 excludes Control; compares Low to High.
contrast2 <- c(0, -1, 1)

# binding the contrasts together
contrasts(accSIM30$COND) <- cbind(contrast1, contrast2)
accSIM30$COND
```

```
[1] High   High   High   High   High   High   High   High   High
[10] High   High   High   High   High   High   High   High   High
[19] High   High   High   High   High   High   High   High   High
[28] High   High   High   Low    Low    Low    Low    Low    Low
[37] Low    Low    Low    Low    Low    Low    Low    Low    Low
[46] Low    Low    Low    Low    Low    Low    Low    Low    Low
[55] Low    Low    Low    Low    Low    Low    Control Control Control
[64] Control Control Control Control Control Control Control Control
[73] Control Control Control Control Control Control Control Control
[82] Control Control Control Control Control Control Control Control
attr("contrasts")
  contrast1 contrast2
Control      -2      0
Low         1     -1
High         1      1
Levels: Control Low High
```

Thinking back to the hand-calculations and contrast mapping, the table of weights that R just produced confirms that

- Contrast 1 compares the control condition against the levels with any racial loading.
- Contrast 2 compares the low and high loadings.

Step 3: Create a new `aov()` model

```
# create a new object, the ANOVA looks the same, but it will now
# consider the contrasts (this is where order-of-operations matters)
accPlanned <- aov(Accurate ~ COND, data = accSIM30)
summary.lm(accPlanned)
```

Call:
`aov(formula = Accurate ~ COND, data = accSIM30)`

Residuals:

Min	1Q	Median	3Q	Max
-1.24533	-0.32092	0.08642	0.30101	1.51646

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	1.60304	0.06215	25.793 < 0.0000000000000002	***
CONDcontrast1	-0.07658	0.04395	-1.742	0.085 .
CONDcontrast2	-0.37365	0.07612	-4.909	0.00000425 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.5896 on 87 degrees of freedom
Multiple R-squared: 0.2377, Adjusted R-squared: 0.2202
F-statistic: 13.57 on 2 and 87 DF, p-value: 0.000007446

```
contrasts(accSIM30$COND) <- cbind(c(-2, 1, 1), c(0, -1, 1))
```

These planned contrasts show that when the control condition is compared to the combined low and high racial loading conditions, there is not a statistically significant difference, $t(87) = -1.742$, $p = 0.085$. However, when the low and high racial loading conditions are compared, there is a statistically significant difference, $t(87) = -4.909$, $p < 0.001$.

4.6.3.4 OPTION 4: Trend (polynomial) analysis

Polynomial contrasts evaluate for the presence of a linear (or curvilinear) pattern in the data. To detect a trend, the data must be coded in an ascending order and it needs to be a sensible, theoretically defensible, comparison. Our data has a theoretically ordered effect (control, low racially loaded condition, high racially loaded condition). Recall that we created an ordered factor when we imported the data.

In a polynomial analysis, the statistical analysis looks across the ordered means to see if they fit a linear or curvilinear shape that is one less than the number of levels. Our factor has three levels, therefore the polynomial contrast can check for a linear shape (.L) or a quadratic (one change in direction) shape (.Q). If we had four levels, the `contr.poly` could check for cubic change (two changes in direction). Conventionally, when more than one trend is significant, we interpret the most complex one (i.e., quadratic over linear).

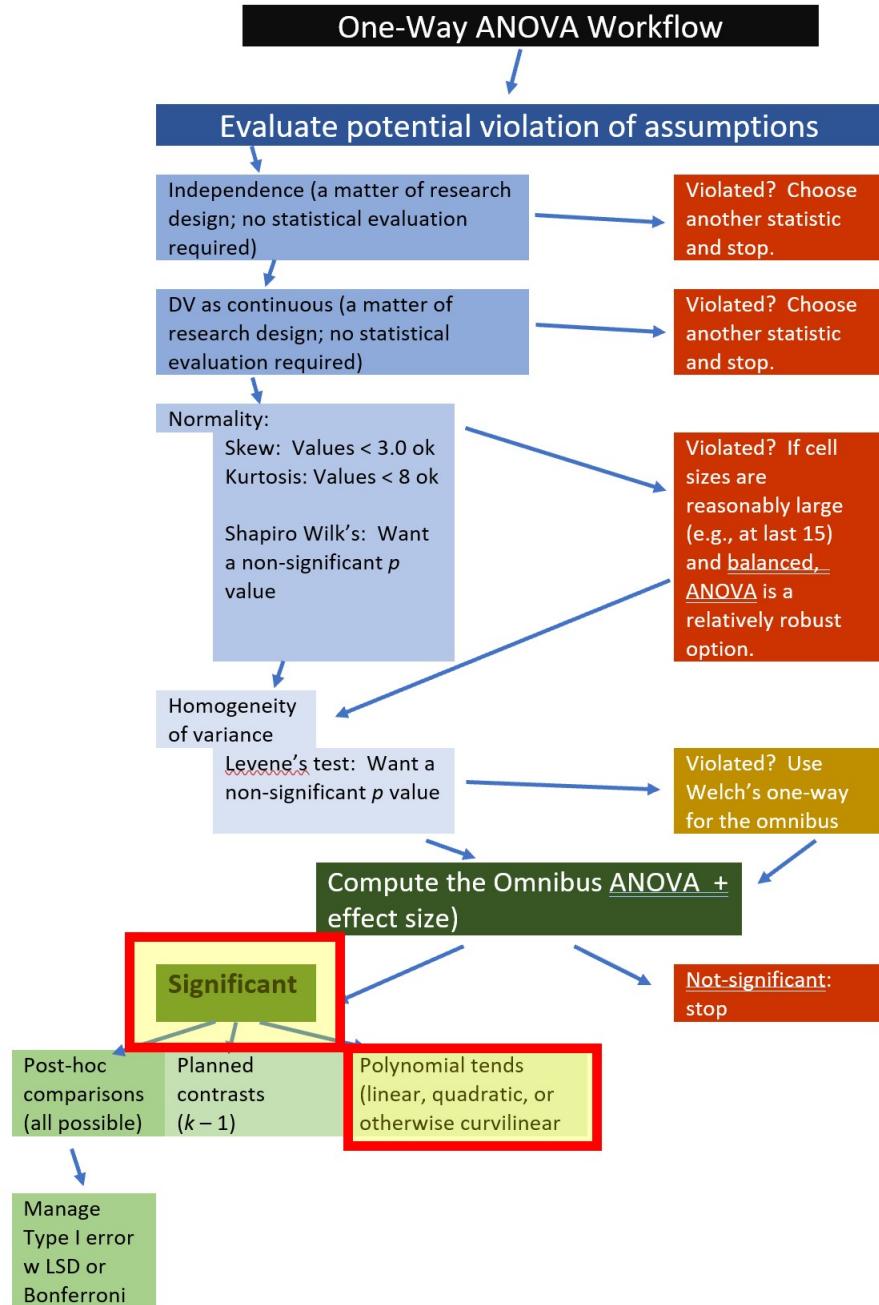


Figure 4.8: An image of the workflow for one-way ANOVA, showing that we are up to a significant omnibus F by assessing for a polynomial trend

```
contrasts(accSIM30$COND) <- contr.poly(3)
accTrend <- aov(Accurate ~ COND, data = accSIM30)
summary.lm(accTrend)
```

Call:
`aov(formula = Accurate ~ COND, data = accSIM30)`

Residuals:

Min	1Q	Median	3Q	Max
-1.24533	-0.32092	0.08642	0.30101	1.51646

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	1.60304	0.06215	25.793	< 0.0000000000000002 ***
COND.L	-0.42665	0.10765	-3.963	0.000151 ***
COND.Q	-0.36384	0.10765	-3.380	0.001087 **

Signif. codes:	0 ***	0.001 **	0.01 *	0.05 .
	''	''	''	''

Residual standard error: 0.5896 on 87 degrees of freedom
Multiple R-squared: 0.2377, Adjusted R-squared: 0.2202
F-statistic: 13.57 on 2 and 87 DF, p-value: 0.000007446

A quick peek back at an early plot shows illustrates the quadratic trend that was supported by the analysis.

Results of our polynomial contrast suggested statistically significant results for both linear $t(87) = -3.963, p < .001$ and quadratic $t(87) = -3.380, p = .001$ trends.

4.6.3.5 Which set of follow-up tests do we report?

It depends! What best tells the story of your data? Here are some things to consider.

- If the post-hoc comparisons are robustly statistically significant (and controlling Type I error is not going to change that significance), I think this provides good information and I would lean toward reporting those.
- If p values are hovering around 0.05, an orthogonal contrast will offer more power because
 - a $k - 1$ comparison will be more powerful
 - *a priori* theory is compelling
- The polynomial can be a useful descriptive addition if there is a linear or quadratic relationship that is sensible or interesting.

Although I would report either the post-hoc or planned contrasts, I will sometimes add a polynomial if it clarifies the result (i.e., there is a meaningful linear or curvilinear pattern essential to understanding the data).

4.6.4 What if we Violated the Homogeneity of Variance test?

The *oneway.test* produces Welch's *F* – a test more robust to violation of the homogeneity of variance assumption. The Welch's approach to attenuating error or erroneous conclusions caused by violations of the homogeneity of variance assumption is to adjust the residual degrees of freedom used to produce the Welch's *F*-ratio.

Another common correction is the Brown and Forsythe *F*-ratio. At this time I have not located and tried an R package that produces this.

```
oneway.test(Accurate ~ COND, data = accSIM30)
```

```
One-way analysis of means (not assuming equal variances)
```

```
data: Accurate and COND
F = 11.569, num df = 2.000, denom df = 56.343, p-value = 0.00006174
```

Note that the denominator *df* is now 56.34 (not 87) and *p* value is a little larger (it was 0.00000745). With its design intended to avoid making a Type I error, the Welch's *F* is more "conservative." While it doesn't matter in this case, if it were a study where the *p* value was closer to .05, it could make a difference. These are some of the tradeoffs made in order to have confidence in the results.

4.7 Power Analysis

Power analysis allows us to determine the sample size required to detect an effect of a given size with a given degree of confidence. Utilized another way, it allows us to determine the probability of detecting an effect of a given size with a given level of confidence. If the probability is unacceptably low, we may want to revise or stop. A helpful overview of power as well as guidelines for how to use the *pwr* package can be found at a [Quick-R website \[Kabacoff, 2017\]](#).

There are four interrelating elements of power:

1. Sample size, *N*
2. Effect size,
 - For one-way ANOVAs, Cohen suggests that *f* values of 0.1, 0.25, and 0.4 represent small, medium, and large effect sizes, respectively.
3. Significance level = $P(\text{Type I error})$,
 - Recall that Type I error is the rejection of a true null hypothesis (a false positive).
 - Stated another way, Type I error is the probability of finding an effect that is not there.
4. Power = $1 - P(\text{Type II error})$,
 - Recall that Type II error is the non-rejection of a false null hypothesis (a false negative).
 - Power is the probability of finding an effect that is there.

If we have any three of these values, we can calculate the fourth.

In Champely's *pwr* package, we can conduct a power analysis for a variety of designs, including the balanced one-way ANOVA (i.e., roughly equal cell sizes) design that we worked in this chapter.

The *pwr.anova.test()* has five parameters:

- $k = \#$ groups
- $n = \text{sample size}$
- $f = \text{effect sizes, where } 0.1/\text{small}, 0.25/\text{medium}, \text{ and } 0.4/\text{large}$
 - In the absence from an estimate from our own data, we make a guess about the expected effect size value based on our knowledge of the literature
- $\text{sig.level} = p$ value that you will use
- $\text{power} = .80$ is the standard value

In the script below, we simply add our values. So long as we have four values, the fifth will be calculated for us.

Because this calculator requires the effect size in the metric of Cohen's f (this is not the same as the F ratio), we need to convert it. The *effectsize* package has a series of converters. We can use the *eta2_to_f()* function.

```
effectsize::eta2_to_f(0.238)
```

```
Registered S3 method overwritten by 'parameters':
  method                  from
  format.parameters_distribution datawizard
```

```
[1] 0.5588703
```

We simply plug this value into the "f =".

```
pwr::pwr.anova.test(k = 3, f = 0.5589, sig.level = 0.05, power = 0.8)
```

```
Balanced one-way analysis of variance power calculation
```

```

k = 3
n = 11.3421
f = 0.5589
sig.level = 0.05
power = 0.8
```

NOTE: n is number in each group

This result suggested that we would need 11 people per group.

If we were unsure about what to expect in terms of our results, we could take a guess. I like to be on the safe(r) side and go with a smaller effect. What would happen if we had a Cohen's f that represented a small effect?

```
pwr::pwr.anova.test(k = 3, f = 0.1, sig.level = 0.05, power = 0.8)
```

```
Balanced one-way analysis of variance power calculation
```

```
k = 3
n = 322.157
f = 0.1
sig.level = 0.05
power = 0.8
```

NOTE: `n` is number in each group

Yikes! We would need over 300 per group!

If effect sizes are new to you, play around with this effect size converter hosted at Psychometrica.de. For examples like this one, use the option labeled, “Transformation of the effect sizes d , r , f , *Odds Ratio*, η^2 , and *Common Language Effect Size (CLES)*.”

4.8 APA Style Results

All that's left to do is *write it up!* APA style results sections in empirical manuscripts are typically accompanied by tables and figures. APA style discourages repeated material and encourages reducing the cognitive load of the reader. For this example, I suggest two tables – one with means and standard deviations of the groups and a second that reports the output from the one-way ANOVA. In an article with multiple statistics, the authors might wish to combine or delete these.

The package `apaTables` can produce journal-ready tables by using the object produced by the `aov()` function. Deciding what to report in text and table is important.

Here I create Table 1 with means and standard deviations (plus a 95% confidence interval around each mean).

```
# table.number = 1 assigns a table number to the top of the table
# filename = 'Table1.doc' writes the table to Microsoft Word and puts
# it in your project folder
apaTables::apa.1way.table(iv = COND, dv = Accurate, show.conf.interval = TRUE,
  data = accSIM30, table.number = 1, filename = "Table1.doc")
```

Table 1

Descriptive statistics for Accurate as a function of COND.

COND	M	M_95%_CI	SD
Control	1.76	[1.58, 1.93]	0.46
Low	1.90	[1.66, 2.14]	0.63
High	1.15	[0.91, 1.40]	0.66

Note. M and SD represent mean and standard deviation, respectively.

LL and UL indicate the lower and upper limits of the 95% confidence interval for the mean, respectively.

The confidence interval is a plausible range of population means that could have caused a sample mean (Cumming, 2014).

I will want to give the values of mean differences (M_{diff}) in the results. I can quickly use R to calculate them here.

```
# calculating mean difference between control and high
1.76 - 1.15
```

[1] 0.61

```
# calculating mean difference between low and high
1.9 - 1.15
```

[1] 0.75

```
# calculating mean difference between control and low
1.76 - 1.9
```

[1] -0.14

Here I create Table 2 with results of the one-way ANOVA. The result in Microsoft Word can be edited (e.g., I would replace the partial-eta squared with η^2) for the journal article.

```
apaTables::apa.aov.table(omnibus, table.number = 2, filename = "Table2.doc")
```

Table 2

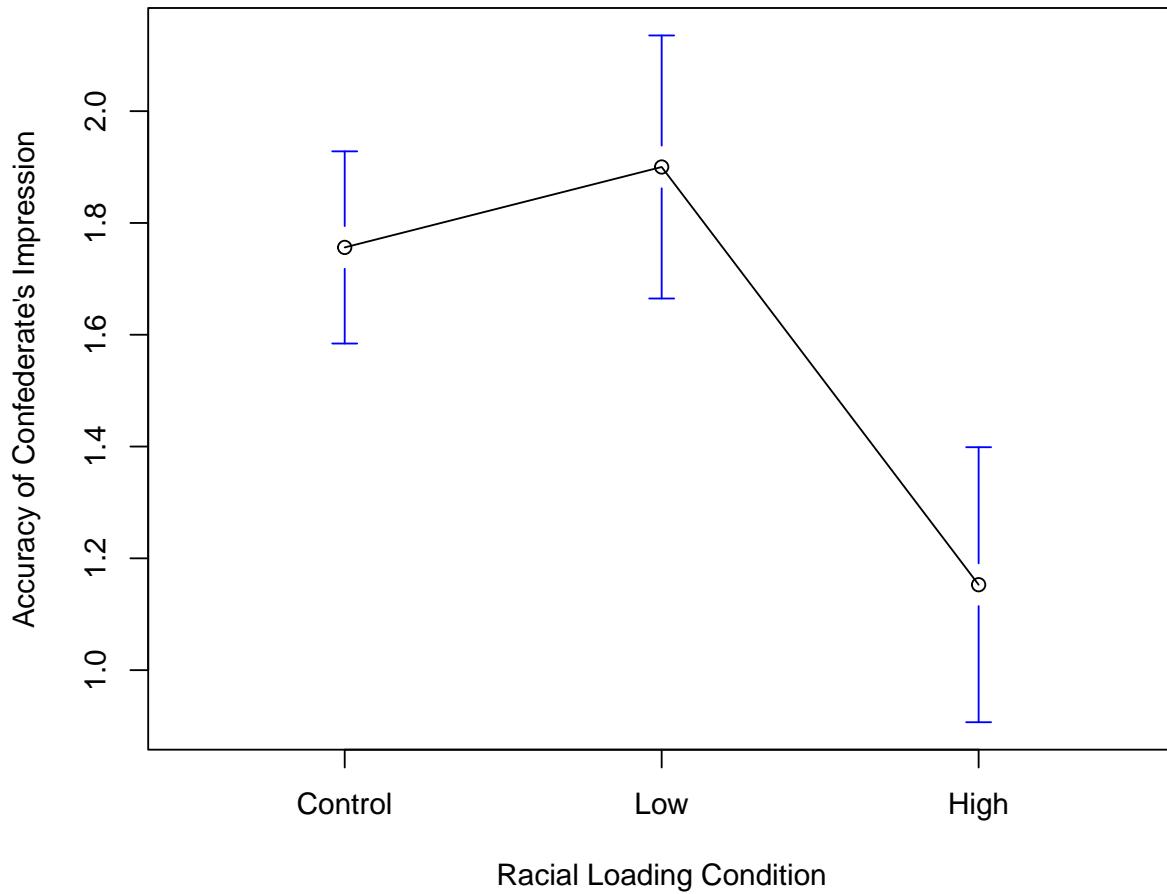
ANOVA results using Accurate as the dependent variable

Predictor	SS	df	MS	F	p	partial_eta2	CI_90_partial_eta2
(Intercept)	92.53	1	92.53	266.15	.000		
COND	9.43	2	4.71	13.57	.000	.24	[.11, .34]
Error	30.25	87	0.35				

Note: Values in square brackets indicate the bounds of the 90% confidence interval for partial

Regarding figures, there are a number of options. I find the `plotmeans()` function (used earlier) to be adequate for the purpose of one-way ANOVA. As we progress through this textbook, I will demonstrate options that offer more and less complexity.

```
gplots:::plotmeans(formula = Accurate ~ COND, data = accSIM30, xlab = "Racial Loading Condition",
                   ylab = "Accuracy of Confederate's Impression", n.label = FALSE)
```



With table and figure at hand, here is how I would write up these results:

A one-way analysis of variance was conducted to evaluate the relationship between degree of racial loading of an exceptionalizing microaggression and the perceived accuracy of a research confederate's impression of the Asian or Asian American participant. The independent variable, COND, included three levels: control/none, low, and high levels of racial loading. Results of Levene's homogeneity of variance test indicated no violation of the homogeneity of variance assumption ($F[2,87] = 1.532, p = 0.222$). Similarly, results of the Shapiro Wilk's test indicated no violation of the normality assumption in each of the cells (Control, $W = 0.954, p = 0.215$; Low, $W = 0.944, p = 0.115$; High, $W = 0.980, p = 0.831$).

Results indicated a significant effect of COND on accuracy perception $F(2, 87) = 13.566, p < .001, \eta^2 = 0.238$. In a series of post-hoc comparisons, there were statistically significant differences between the control and high ($M_{diff} = 0.61, p < .001$) and low and high ($M_{diff} = 0.75, p < 0.001$) conditions, but not the control and low conditions ($M_{diff} = -.14, p = 0.347$). A statistically significant polynomial contrast suggested a quadratic trend across the three, ordered, levels of the conditions ($t[87] = -0.364, p = .001$) such that perception of accuracy increased slightly from the control to low conditions, but decreased more dramatically from low to high. Consequently, it appeared that only the highest degree of racial loading (e.g., “You speak English well for an Asian”) resulted in the decreased perceptions of accuracy of impressions from the confederate. Means and standard deviations are presented in Table 1 and complete ANOVA results are presented in Table 2. Figure 1 provides an illustration of the results.

4.9 A Conversation with Dr. Tran

Doctoral student (and student in one of my classes) Emi Ichimura and I were able to interview the first author (Alisia Tran, PhD) about the article and what it means. Here’s a direct [link](#) to that interview.

Among others, we asked:

- What were unexpected challenges to the research method or statistical analysis?
- What were the experiences of the confederates as they offered the statements in teh racial loading conditions? And in the debriefings, did the research participants share anything more anecdotally in their experiences as research participants?
- What are your current ideas about interventions or methods for mitigating the harm caused by racial microaggressions?
- How do you expect the article to change science, practice, and/or advocacy?

4.10 Practice Problems

In each of these lessons I provide suggestions for practice that allow you to select one or more problems that offer differing levels of difficulty. Whichever you choose, you will focus on these larger steps in one-way ANOVA, including:

- testing the statistical assumptions
- conducting a one-way ANOVA, including
 - omnibus test and effect size
 - follow-up (pairwise, planned comparisons, polynomial trends)
- writing a results section to include a figure and tables

4.10.1 Problem #1: Play around with this simulation.

If one-way ANOVA is new to you, perhaps you just change the number in “set.seed(2021)” from 2021 to something else. Your results should parallel those obtained in the lecture, making it easier for you to check your work as you go.

There are other ways to change the dataset. For example, if you are interested in power, change the sample size to something larger or smaller. If you are interested in variability (i.e., the homogeneity of variance assumption), perhaps you change the standard deviations in a way that violates the assumption.

4.10.2 Problem #2: Conduct a one-way ANOVA with the *moreTalk* dependent variable.

In their study, Tran and Lee [2014] included an outcome variable where participants rated how much longer they would continue the interaction with their partner compared to their interactions in general. The scale ranged from -2 (*much less than average*) through 0 (*average*) to 2 (*much more than average*). This variable is available in the original simulation and is an option for a slightly more challenging analysis.

4.10.3 Problem #3: Try something entirely new.

Using data for which you have permission and access (e.g., IRB approved data you have collected or from your lab; data you simulate from a published article; data from an open science repository; data from other chapters in this OER), complete a one-way ANOVA. Please have at least 3 levels for the predictor variable.

4.10.4 Grading Rubric

Regardless which option(s) you chose, use the elements in the grading rubric to guide you through the practice.

Assignment Component	Points Possible	Points Earned
1. Narrate the research vignette, describing the IV and DV	5	_____
2. Simulate (or import) and format data	5	_____
3. Evaluate statistical assumptions	5	_____
4. Conduct omnibus ANOVA (w effect size)	5	_____
5. Conduct one set of follow-up tests; narrate your choice	5	_____
6. Describe approach for managing Type I error	5	_____
7. APA style results with table(s) and figure	5	_____
8 Explanation to grader	5	_____
Totals	40	_____

4.11 Bonus Reel:

4.11.1 What's with the inline code?

If you are working in the .rmd file you may notice sections that look like this:

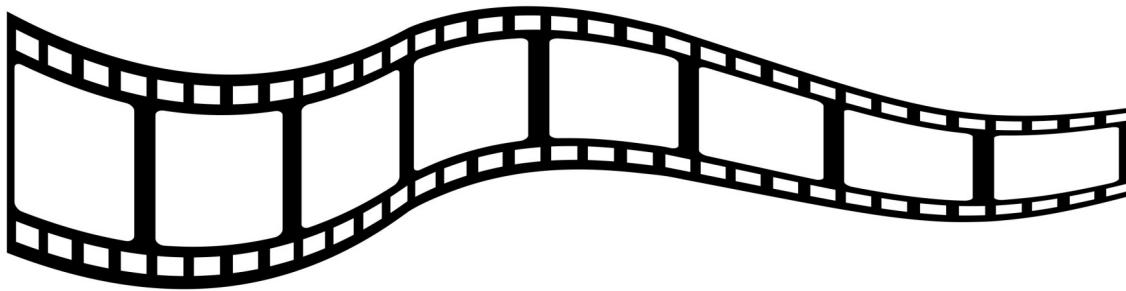


Figure 4.9: Image of a filmstrip signifying that what follows is considered to be supplemental

```
# this script is used for the inline coding in the lesson and,
# although you will want to run it so you can 'feed' the objects into
# later script, there is no 'lesson' relative to this lecture
df1omnibus <- formattable::digits(summary(omnibus)[[1]][1, "Df"], 0)
df1omnibus
df2omnibus <- formattable::digits(summary(omnibus)[[1]][2, "Df"], 0)
df2omnibus
SSMognibus <- formattable::digits(summary(omnibus)[[1]][1, "Sum Sq"], 3)
SSMognibus
SSRomnibus <- formattable::digits(summary(omnibus)[[1]][2, "Sum Sq"], 3)
SSRomnibus
SSTomnibus <- formattable::digits((SSMognibus + SSRomnibus), 3)
SSTomnibus
MSMognibus <- formattable::digits(summary(omnibus)[[1]][1, "Mean Sq"],
  3)
MSMognibus
MSRomnibus <- formattable::digits(summary(omnibus)[[1]][2, "Mean Sq"],
  3)
MSRomnibus
Fomnibus <- formattable::digits(summary(omnibus)[[1]][1, "F value"], 3)
Fomnibus
pomnibus <- formattable::digits(summary(omnibus)[[1]][1, "Pr(>F)"], 3)
pomnibus
```

I'm still learning about this and am using the lessons to practice. In these hidden (from the rendered view) boxes of script, I am capturing the output values as R objects. Later I can use inline code (the object's name, preceded with an "r", surrounded by tics [unfortunately, I cannot demo it in the .rmd file without getting an error]) to insert the value into the lecture notes and results. I'm working up to writing an entire journal article in R. This way, if something changes (e.g., more participants in the Qualtrics survey, a different approach to cleaning data) when I re-run the script everything auto-updates and I'm at less of a risk (or maybe a different kind of risk) for making typographical errors.

Chapter 5

Factorial (Between-Subjects) ANOVA

[Screencasted Lecture Link](#)

In this (somewhat long and complex) lesson we conduct a 3X2 ANOVA. We will

- Work an actual example from the literature.
 - “by hand”, and
 - with R packages
- I will also demonstrate
 - several options for exploring interaction effects, and
 - several options for exploring main effects.
- Exploring these options will allow us to:
 - Gain familiarity with the concepts central to multi-factor ANOVAs.
 - Explore tools for analyzing the complexity in designs.

The complexity is that not all of these things need to be conducted for every analysis. The Two-Way ANOVA Workflow is provided to help you map a way through your own analyses. I will periodically refer to this map so that we can more easily keep track of where we are in the process.

5.1 Navigating this Lesson

There is about 1 hour and 30 minutes hours of lecture. If you work through the materials with me plan for another two hours of study.

While the majority of R objects and data you will need are created within the R script that sources the chapter, occasionally there are some that cannot be created from within the R framework. Additionally, sometimes links fail. All original materials are provided at the [Github site](#) that hosts the book. More detailed guidelines for ways to access all these materials are provided in the OER’s [introduction](#)

5.1.1 Learning Objectives

Learning objectives from this lecture include the following:

- Define, locate, and interpret all the effects associated with two-way ANOVA:
 - main
 - interaction (introducing the concept, *moderator*)
 - simple main effects
- Identify which means belong with which effects. Then compare and interpret them.
 - marginal means
 - individual cell means
 - comparing them
- Map a process/workflow for investigating a factorial ANOVA
- Manage Type I error
- Conduct a power analysis to determine sample size

5.1.2 Planning for Practice

In each of these lessons I provide suggestions for practice that allow you to select one or more problems that are graded in difficulty. The least complex is to change the random seed and rework the problem demonstrated in the lesson. The results *should* map onto the ones obtained in the lecture.

The second option comes from the research vignette. The Ramdhani et al. [2018] article has two dependent variables (DVs; negative and positive evaluation) which are suitable for two-way ANOVAs. I will demonstrate a simulation of one of their 3X2 ANOVAs (negative) in this lecturette. The second dependent variable (positive) is suggested for the moderate level of difficulty.

As a third option, you are welcome to use data to which you have access and is suitable for two-way ANOVA. In either case the practice options suggest that you:

- test the statistical assumptions
- conduct a two-way ANOVA, including
 - omnibus test and effect size
 - report main and interaction effects
 - conduct follow-up testing of simple main effects
- write a results section to include a figure and tables

5.1.3 Readings & Resources

In preparing this chapter, I drew heavily from the following resource(s) that are freely available on the internet. Other resources are cited (when possible, linked) in the text with complete citations in the reference list.

- Navarro, D. (2020). Chapter 16: Factorial ANOVA. In Learning Statistics with R - A tutorial for Psychology Students and other Beginners. Retrieved from [https://stats.libretexts.org/Bookshelves/Applied_Statistics/Book%3A_Learning_Statistics_with_R_-_A_tutorial_for_Psychology_Students_and_other_Beginners_\(Navarro\)](https://stats.libretexts.org/Bookshelves/Applied_Statistics/Book%3A_Learning_Statistics_with_R_-_A_tutorial_for_Psychology_Students_and_other_Beginners_(Navarro))
 - Navarro's OER includes a good mix of conceptual information about one-way ANOVA as well as R code. My code/approach is a mix of Green and Salkind's [2014b], Field's [2012], Navarro's [2020b], and other techniques I have found on the internet and learned from my students.
- Ramdhani, N., Thontowi, H. B., & Ancok, D. (2018). Affective Reactions Among Students Belonging to Ethnic Groups Engaged in Prior Conflict. *Journal of Pacific Rim Psychology*, 12, e2. <https://doi.org/10.1017/prp.2017.22>
 - The source of our research vignette.

5.1.4 Packages

The packages used in this lesson are embedded in this code. When the hashtags are removed, the script below will (a) check to see if the following packages are installed on your computer and, if not (b) install them.

```
# will install the package if not already installed
# if(!require(knitr)){install.packages('knitr')}
# if(!require(psych)){install.packages('psych')}
# if(!require(tidyverse)){install.packages('tidyverse')}
# if(!require(lsr)){install.packages('lsr')}
# if(!require(car)){install.packages('car')}
# if(!require(ggpubr)){install.packages('ggpubr')}
# if(!require(effectsize)){install.packages('effectsize')}
# if(!require(pwr2)){install.packages('pwr2')}
# if(!require(apaTables)){install.packages('apaTables')}
```

5.2 Introducing Factorial ANOVA

My approach to teaching is to address the conceptual as we work problems. That said, there are some critical ideas we should address first.

ANOVA is for experiments (or arguably closely related designs). As we get into the assumptions you'll see that it has some rather restrictive ones (e.g., there should be an equal/equivalent number of cases per cell). To the degree that we violate these assumptions, we should locate alternative statistical approaches where these assumptions are relaxed.

Factorial: a term used when there are two or more independent variables (IVs; the factors). The factors could be between-groups, within-groups, repeated measures, or a combination of between and within.

- **Independent factorial design:** several IVs (predictors/factors) and each has been measured using different participants (between groups).

- **Related factorial design:** several IVs (factors/predictors) have been measured, but the same participants have been used in all conditions (repeated measures or within-subjects).
- **Mixed design:** several IVs (factors/predictors) have been measured. One or more factors uses different participants (between-subjects) and one or more factors uses the same participants (within-subjects). Thus, there is a combination of independent (between) and related (within or repeated) designs.
- Factor naming follows a number/levels convention.
- Today's example is a 3X2 ANOVA. We know there are two factors that have three and two levels, respectively: *ethnicity* has three levels representing the two ethnic groups that were in prior conflict (Marudese, Dayaknese) and a third group who was uninvolved in the conflict (Javanese); *photo stimulus* has two levels representing members of the two ethnic groups that were in prior conflict (Madurese, Dayaknese);

Moderator is what creates an interaction. Below are traditional representations of the *statistical* and *conceptual* figures of interaction effects. We will say that Factor B, *moderates* the relationship between Factor A/IV and the DV.

In a later lesson we work an ANCOVA – where we will distinguish between a *moderator* and a *covariate*. In regression models, you will likely be introduced to the *mediator*.

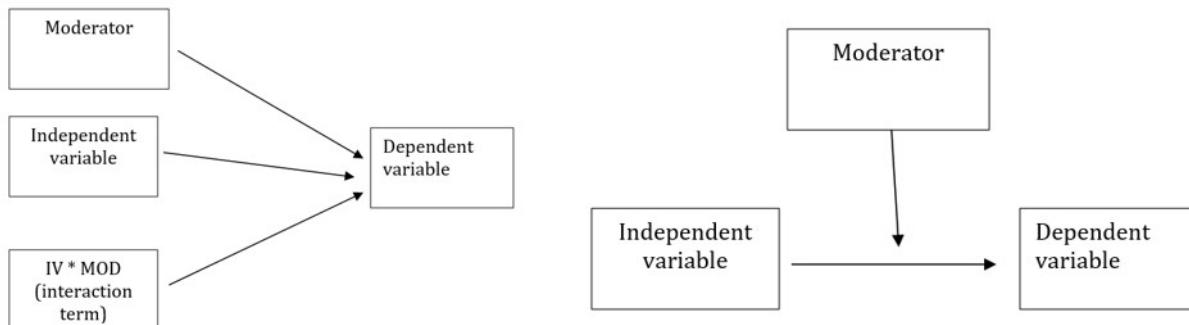


Figure 5.1: Graphic representations of a moderated relationship?

5.2.1 Workflow for Two-Way ANOVA

The following is a proposed workflow for conducting a two-way ANOVA.

Steps of the workflow include:

1. Enter data
 - predictors should be formatted as factors (ordered or unordered); the dependent variable should be continuously scaled
 - understanding the format of data can often provide clues as to which ANOVA/statistic to use
2. Explore data
 - graphing the data

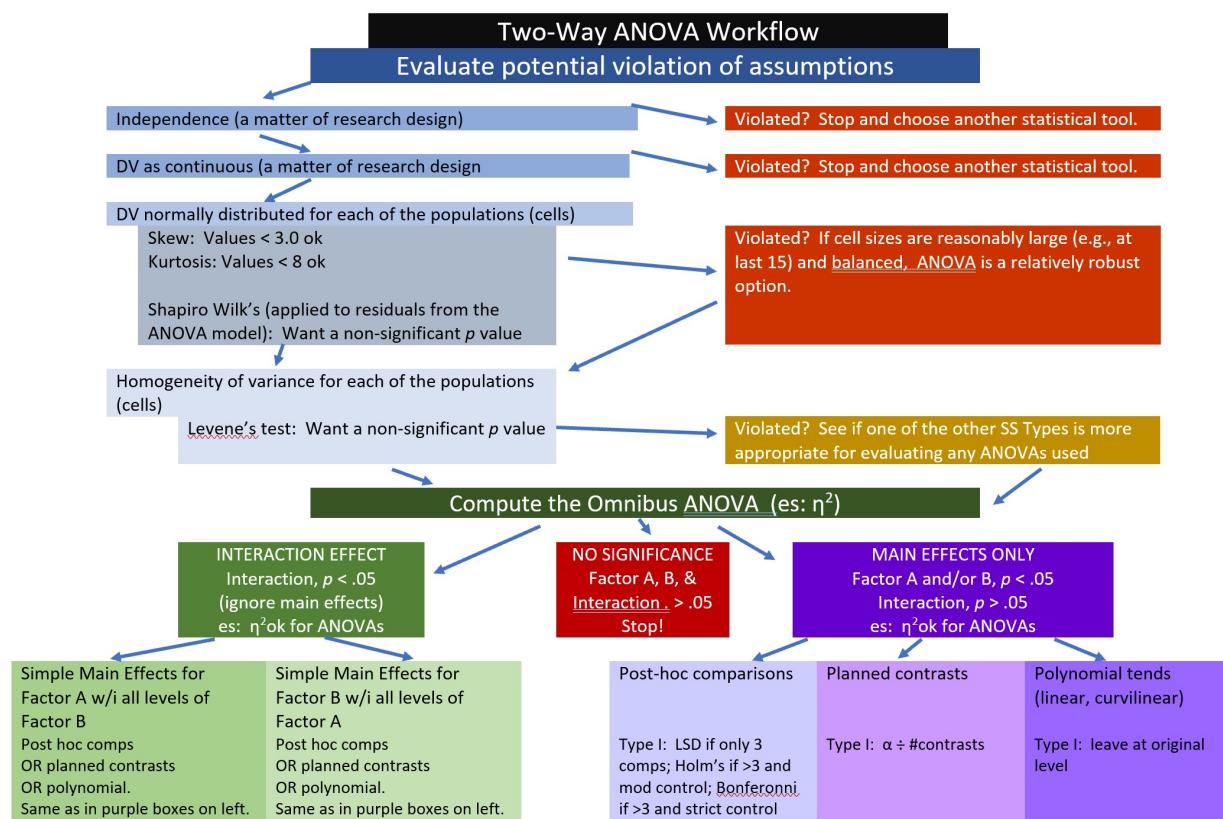


Figure 5.2: An image of a workflow for the two-way ANOVA

- computing descriptive statistics
 - check distributional assumptions
 - use Levene's test to check for homogeneity of variance
 - use Shapiro Wilks to check for normality
3. Construct or choose contrasts
 - select contrasts and specify for all of the IVs in the analysis
 - if you want to use Type III sums of squares, contrasts must be orthogonal
 4. Compute the omnibus ANOVA
 - *depending on what you found in the data exploration phase, you may need to run a robust version of the test*
 5. Follow-up testing based on significant main or interaction effects
 - significant interactions require test of simple main effects which could be further explored with contrasts, posthoc comparisons, and/or polynomials
 - *the exact methods you choose will depend upon the tests of assumptions during data exploration*
 6. Managing Type I error

5.3 Research Vignette

The research vignette for this example was located in Kalimantan, Indonesia and focused on bias in young people from three ethnic groups. The Madurese and Dayaknese groups were engaged in ethnic conflict that spanned 1996 to 2001. The last incidence of mass violence was in 2001 where approximately 500 people (mostly from the Madurese ethnic group) were expelled from the province. Ramdhani et al.'s [2018] research hypotheses were based on the roles of the three ethnic groups in the study. The Madurese appear to be viewed as the transgressors when they occupied lands and took employment and business opportunities from the Dayaknese. Ramdhani et al. also included a third group who were not involved in the conflict (Javanese). The research participants were students studying in Yogyakarta who were not involved in the conflict. They included 39 Madurese, 35 Dyaknese, and 37 Javanese; 83 were male and 28 were female.

In the study [Ramdhani et al., 2018], participants viewed facial pictures of three men and three women (in traditional dress) from each ethnic group (6 photos per ethnic group). Participant were asked, "How do you feel when you see this photo? Please indicate your answers based on your actual feelings." Participants responded on a 7-point Likert scale ranging from 1 (*strongly disagree*) to 7 (*strongly agree*). Higher scores indicated ratings of higher intensity on that scale. The two scales included the following words:

- Positive: friendly, kind, helpful, happy
- Negative: disgusting, suspicious, hateful, angry

Below is script to simulate data for the negative reactions variable from the information available from the manuscript [Ramdhani et al., 2018].

```

library(tidyverse)
set.seed(210731)
# sample size, M and SD for each cell; this will put it in a long
# file
Negative <- round(c(rnorm(17, mean = 1.91, sd = 0.73), rnorm(18, mean = 3.16,
  sd = 0.19), rnorm(19, mean = 3.3, sd = 1.05), rnorm(20, mean = 3, sd = 1.07),
  rnorm(18, mean = 2.64, sd = 0.95), rnorm(19, mean = 2.99, sd = 0.8)), 3)
# sample size, M and SD for each cell; this will put it in a long
# file
Positive <- round(c(rnorm(17, mean = 4.99, sd = 1.38), rnorm(18, mean = 3.83,
  sd = 1.13), rnorm(19, mean = 4.2, sd = 0.82), rnorm(20, mean = 4.19,
  sd = 0.91), rnorm(18, mean = 4.17, sd = 0.6), rnorm(19, mean = 3.26,
  sd = 0.94)), 3)
ID <- factor(seq(1, 111))
Rater <- c(rep("Dayaknese", 35), rep("Madurese", 39), rep("Javanese", 37))
Photo <- c(rep("Dayaknese", 17), rep("Madurese", 18), rep("Dayaknese",
  19), rep("Madurese", 20), rep("Dayaknese", 18), rep("Madurese", 19))
# groups the 3 variables into a single df: ID#, DV, condition
Ramdhani_df <- data.frame(ID, Negative, Positive, Rater, Photo)

```

For two-way ANOVA our variables need to be properly formatted. In our case:

- Negative is a continuously scaled DV and should be *num*
- Positive is a continuously scaled DV and should be *num*
- Rater should be an unordered factor
- Photo should be an unordered factor

```
str(Ramdhani_df)
```

```
'data.frame': 111 obs. of 5 variables:
 $ ID      : Factor w/ 111 levels "1","2","3","4",...: 1 2 3 4 5 6 7 8 9 10 ...
 $ Negative: num  2.768 1.811 0.869 1.857 2.087 ...
 $ Positive: num  5.91 5.23 3.54 5.63 5.44 ...
 $ Rater   : chr  "Dayaknese" "Dayaknese" "Dayaknese" "Dayaknese" ...
 $ Photo    : chr  "Dayaknese" "Dayaknese" "Dayaknese" "Dayaknese" ...
```

Our Negative variable is correctly formatted. Let's reformat Rater and Photo to be factors and ask for the structure again. R's default is to order the factors alphabetically. In this case this is fine. If we had ordered factors such as dosage (placebo, lo, hi) we would want to respecify the order.

```

Ramdhani_df[, "Rater"] <- as.factor(Ramdhani_df[, "Rater"])
Ramdhani_df[, "Photo"] <- as.factor(Ramdhani_df[, "Photo"])
str(Ramdhani_df)

```

```
'data.frame': 111 obs. of 5 variables:
 $ ID      : Factor w/ 111 levels "1","2","3","4",...: 1 2 3 4 5 6 7 8 9 10 ...
 $ Negative: num  2.768 1.811 0.869 1.857 2.087 ...
 $ Positive: num  5.91 5.23 3.54 5.63 5.44 ...
 $ Rater   : Factor w/ 3 levels "Dayaknese","Javanese",...: 1 1 1 1 1 1 1 1 1 ...
 $ Photo   : Factor w/ 2 levels "Dayaknese","Madurese": 1 1 1 1 1 1 1 1 1 ...
```

If you want to export this data as a file to your computer, remove the hashtags to save it (and re-import it) as a .csv (“Excel lite”) or .rds (R object) file. This is not a necessary step.

The code for .csv will likely lose the formatting (i.e., making the Rater and Photo variables factors), but it is easy to view in Excel.

```
# write the simulated data as a .csv write.table(Ramdhani_df,
# file='RamdhaniCSV.csv', sep=',', col.names=TRUE, row.names=FALSE)
# bring back the simulated dat from a .csv file Ramdhani_df <-
# read.csv ('RamdhaniCSV.csv', header = TRUE) str(Ramdhani_df)
```

The code for the .rds file will retain the formatting of the variables, but is not easy to view outside of R.

```
# to save the df as an .rds (think 'R object') file on your computer;
# it should save in the same file as the .rmd file you are working
# with saveRDS(Ramdhani_df, 'Ramdhani_RDS.rds') bring back the
# simulated dat from an .rds file Ramdhani_RDS <-
# readRDS('Ramdhani_RDS.rds') str(Ramdhani_RDS)
```

5.3.1 Preliminary exploration of our research vignette

Let’s first examine the descriptive statistics (e.g., means of the variable, Negative) by group. We can use the *describeBy()* function from the *psych* package.

```
negative.descripts <- psych::describeBy(Negative ~ Rater + Photo, mat = TRUE,
                                         data = Ramdhani_df, digits = 3) #digits allows us to round the output
negative.descripts
```

item	group1	group2	vars	n	mean	sd	median	trimmed	mad	min	
X11	1	Dayaknese	Dayaknese	1	17	1.818	0.768	1.692	1.783	0.694	0.706
X12	2	Javanese	Dayaknese	1	18	2.524	0.742	2.391	2.460	0.569	1.406
X13	3	Madurese	Dayaknese	1	19	3.301	1.030	3.314	3.321	1.294	1.406
X14	4	Dayaknese	Madurese	1	18	3.129	0.156	3.160	3.136	0.104	2.732
X15	5	Javanese	Madurese	1	19	3.465	0.637	3.430	3.456	0.767	2.456
X16	6	Madurese	Madurese	1	20	3.297	1.332	2.958	3.254	1.615	1.211
					max	range	skew	kurtosis	se		
X11					3.453	2.747	0.513	-0.881	0.186		
X12					4.664	3.258	1.205	1.475	0.175		

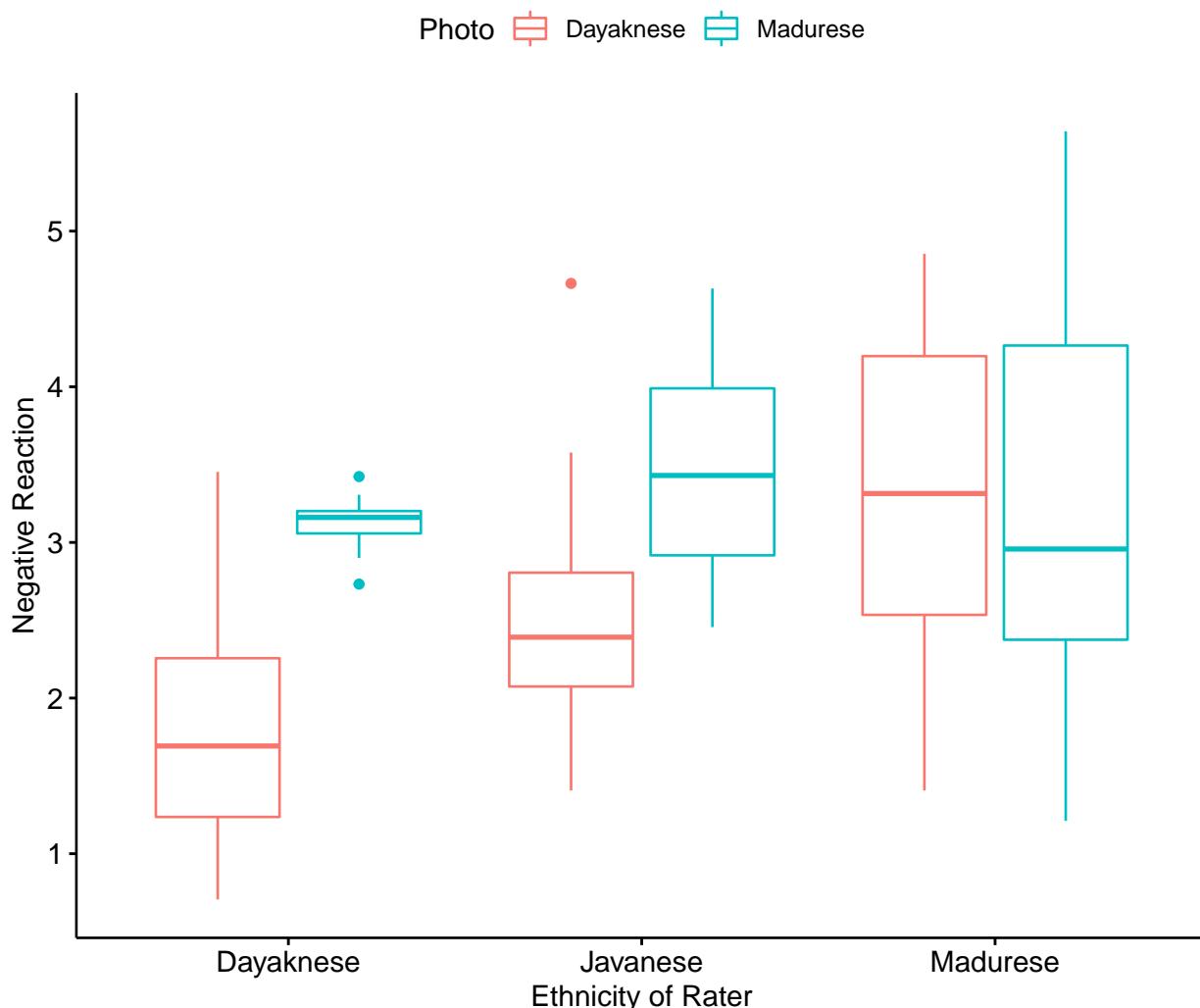
```
X13 4.854 3.448 -0.126   -1.267 0.236
X14 3.423 0.691 -0.623    0.481 0.037
X15 4.631 2.175 -0.010   -1.307 0.146
X16 5.641 4.430  0.215   -1.238 0.298
```

The `write.table()` function can be a helpful way to export output to .csv files so that you can manipulate it into tables.

```
write.table(negative.descripts, file = "NegativeDescripts.csv", sep = ",",
            col.names = TRUE, row.names = FALSE)
```

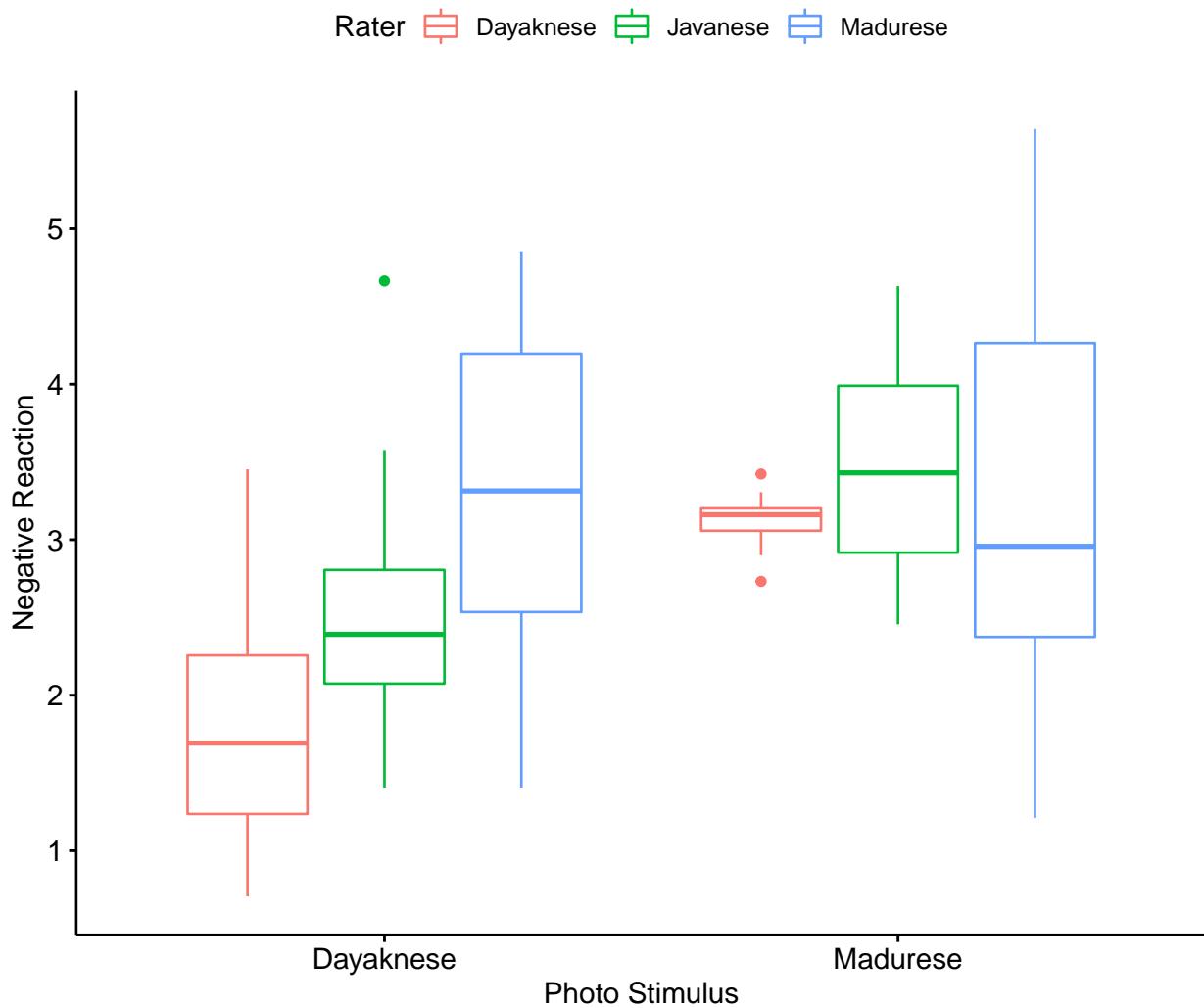
At this stage, it would be useful to plot our data. Figures can assist in the conceptualization of the analysis.

```
ggpubr::ggboxplot(Ramdhani_df, x = "Rater", y = "Negative", color = "Photo",
                   xlab = "Ethnicity of Rater", ylab = "Negative Reaction")
```



Narrating results is sometimes made easier if variables are switched. There is usually not a right or wrong answer. Here is another view, switching the Rater and Photo predictors.

```
ggpubr::ggboxplot(Ramdhani_df, x = "Photo", y = "Negative", color = "Rater",
  xlab = "Photo Stimulus", ylab = "Negative Reaction")
```

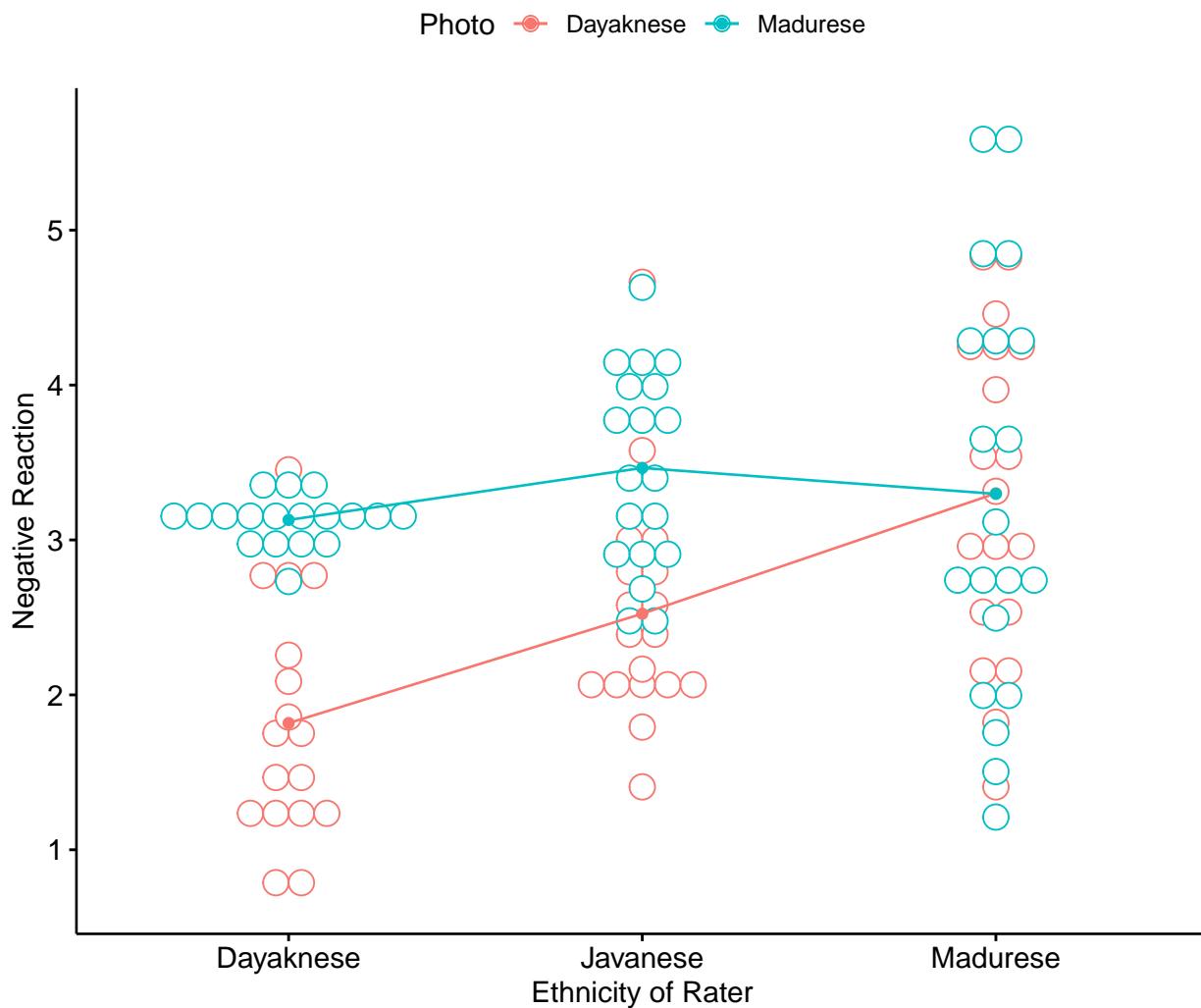


Yet another option plots the raw data as bubbles, the means as lines, and denotes differences in the moderator with color.

```
ggpubr::ggline(Ramdhani_df, x = "Rater", y = "Negative", color = "Photo",
  xlab = "Ethnicity of Rater", ylab = "Negative Reaction", add = c("mean_se",
  "dotplot"))
```

Bin width defaults to 1/30 of the range of the data. Pick better value with `binwidth`.

Warning: Computation failed in `stat_summary()`:
object 'mean_se_' of mode 'function' was not found



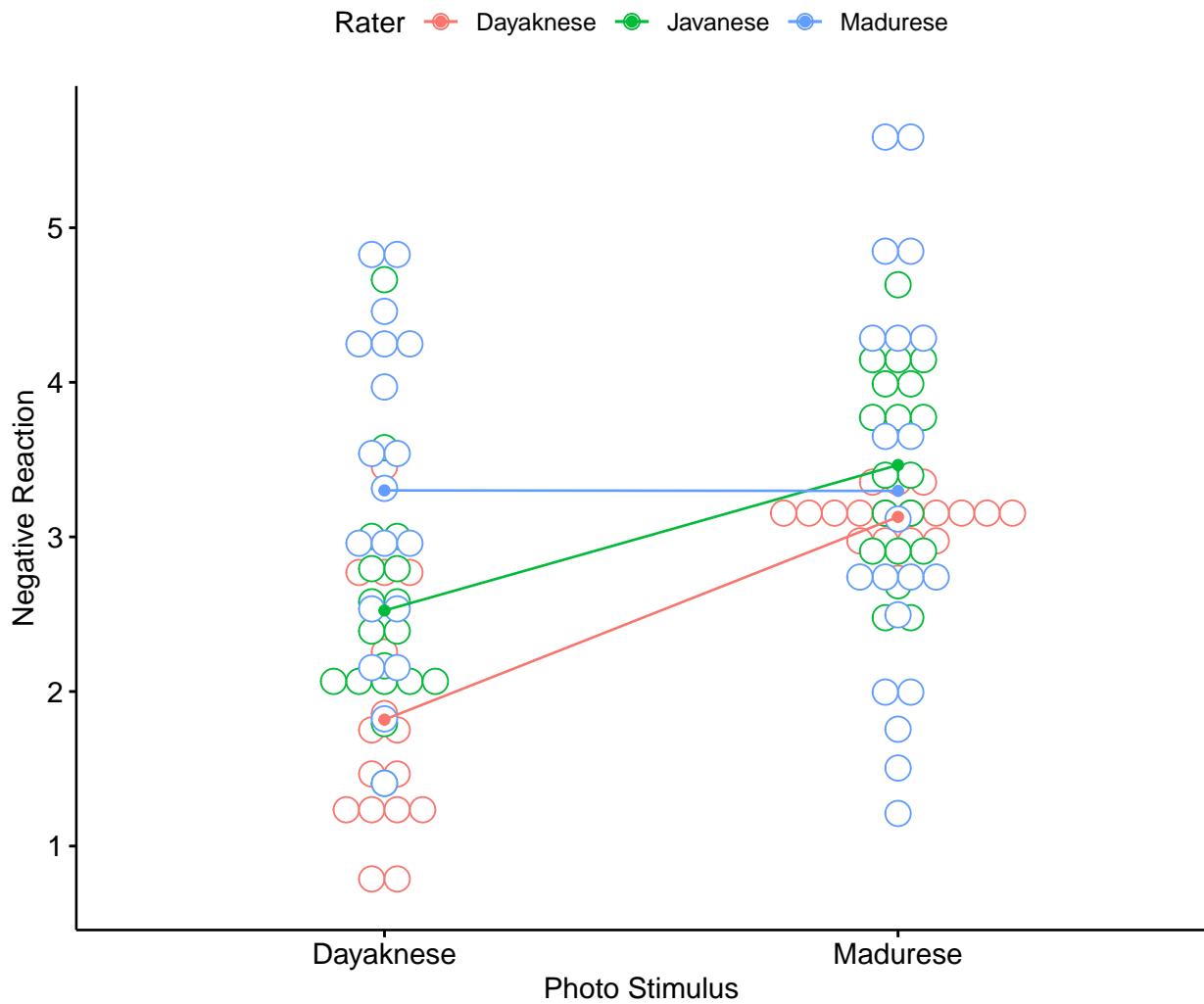
```
# add this for a different color palette: palette = c('#00AFBB',
# '#E7B800')
```

We can reverse this to see if it assists with our conceptualization.

```
ggpubr::gglime(Ramdhani_df, x = "Photo", y = "Negative", color = "Rater",
  xlab = "Photo Stimulus", ylab = "Negative Reaction", add = c("mean_se",
  "dotplot"))
```

Bin width defaults to 1/30 of the range of the data. Pick better value with `binwidth`.

```
Warning: Computation failed in `stat_summary()`:
object 'mean_se_' of mode 'function' was not found
```



5.4 Working the Factorial ANOVA (by hand)

Before we work an ANOVA let's take a moment to consider what we are doing and how it informs our decision-making. This figure (which already contains "the answers") may help conceptualize how variance becomes partitioned.

As in one-way ANOVA, we partition variance into **total**, **model**, and **residual**. However, we now further divide the SS_M into its respective factors A(column), B(row,) and their a x b product.

In this, we begin to talk about main effects and interactions.

5.4.1 Sums of Squares Total

Our formula is the same as it was for one-way ANOVA:

$$SS_T = \sum (x_i - \bar{x}_{grand})^2$$

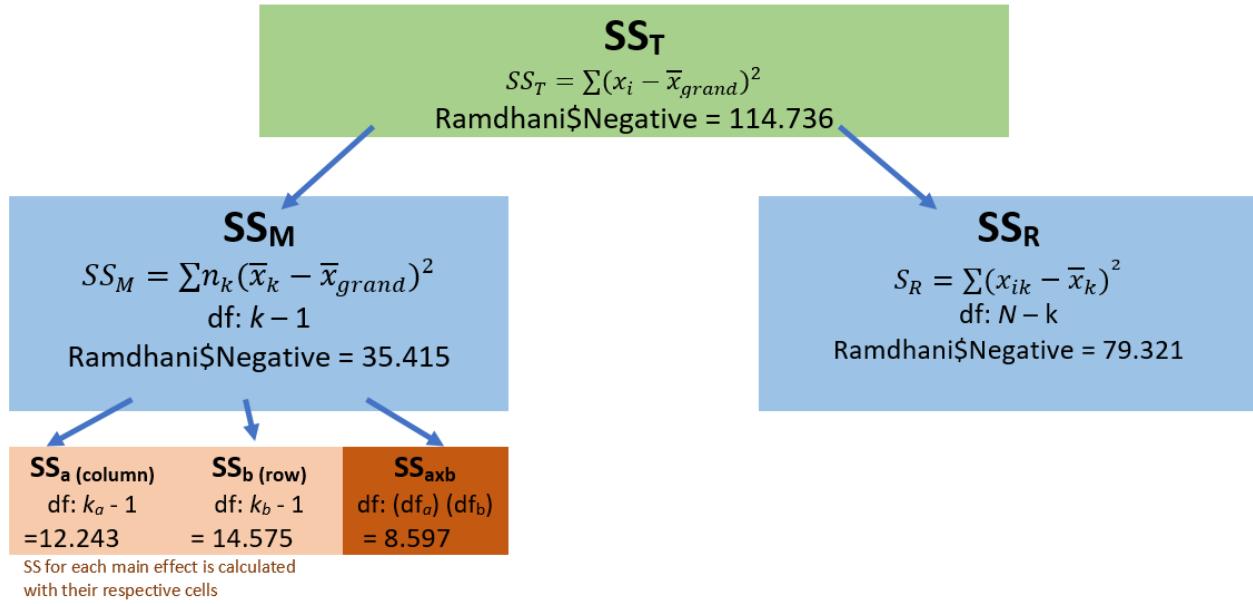


Figure 5.3: Image of a flowchart that partitions variance from sums of squares totals to its component pieces

Let's calculate it for the Ramdhani et al. [2018] data. Our grand (i.e., overall) mean is

```
mean(Ramdhani_df$Negative)
```

```
[1] 2.947369
```

Subtracting the grand mean from each Negative rating yields a mean difference.

```
library(tidyverse)
Ramdhani_df <- Ramdhani_df %>%
  mutate(m_dev = Negative - mean(Negative))
```

Pop quiz: What's the sum of our new *m_dev* variable?

Let's find out!

```
sum(Ramdhani_df$m_dev)
```

```
[1] -0.000000000000007549517
```

Of course! The sum of squared deviations around the mean is zero. Next we square those mean deviations.

```
Ramdhani_df <- Ramdhani_df %>%
  mutate(m_devSQ = m_dev^2)
```

Then we sum the squared mean deviations.

```
sum(Ramdhani_df$m_devSQ)
```

```
[1] 114.7746
```

This value, 114.775, the sum of squared deviations around the grand mean, is our SS_T ; the associated *degrees of freedom* is $N - 1$.

In factorial ANOVA, we divide SS_T into **model/between** sums of squares and **residual/within** sums of squares.

5.4.2 Sums of Squares for the Model

$$SS_M = \sum n_k (\bar{x}_k - \bar{x}_{grand})^2$$

The *model* generally represents the notion that the means are different than each other. We want the variation between our means to be greater than the variation within each of the groups from which our means are calculated.

In factorial, we need to obtain means for each of the combinations of the factors. We have a 3 x 2:

- Rater with three levels: Dayaknese, Madurese, Javanese
- Photo with two levels: Dayaknese, Madurese

Let's repeat some code we used before to obtain the cell-level means and cell sizes.

```
psych::describeBy(Negative ~ Rater + Photo, mat = TRUE, data = Ramdhani_df,
  digits = 3)
```

	item	group1	group2	vars	n	mean	sd	median	trimmed	mad	min
X11	1	Dayaknese	Dayaknese		1 17	1.818	0.768	1.692	1.783	0.694	0.706
X12	2	Javanese	Dayaknese		1 18	2.524	0.742	2.391	2.460	0.569	1.406
X13	3	Madurese	Dayaknese		1 19	3.301	1.030	3.314	3.321	1.294	1.406
X14	4	Dayaknese	Madurese		1 18	3.129	0.156	3.160	3.136	0.104	2.732
X15	5	Javanese	Madurese		1 19	3.465	0.637	3.430	3.456	0.767	2.456
X16	6	Madurese	Madurese		1 20	3.297	1.332	2.958	3.254	1.615	1.211
	max	range	skew	kurtosis	se						
X11	3.453	2.747	0.513	-0.881	0.186						
X12	4.664	3.258	1.205	1.475	0.175						
X13	4.854	3.448	-0.126	-1.267	0.236						
X14	3.423	0.691	-0.623	0.481	0.037						
X15	4.631	2.175	-0.010	-1.307	0.146						
X16	5.641	4.430	0.215	-1.238	0.298						

```
# Note. Recently my students and I have been having intermittent
# struggles with the describeBy function in the psych package. We
# have noticed that it is problematic when using .rds files and when
# using data directly imported from Qualtrics. If you are having
# similar difficulties, try uploading the .csv file and making the
# appropriate formatting changes.
```

We also need the grand mean (i.e., the mean that disregards [or “collapses across”] the factors).

```
mean(Ramdhani_df$Negative)
```

```
[1] 2.947369
```

This formula occurs in six chunks, representing the six cells of our designed. In each of the chunks we have the n , group mean, and grand mean.

```
17 * (1.818 - 2.947)^2 + 18 * (2.524 - 2.947)^2 + 19 * (3.301 - 2.947)^2 +
  18 * (3.129 - 2.947)^2 + 19 * (3.465 - 2.947)^2 + 20 * (3.297 - 2.947)^2
```

```
[1] 35.41501
```

This value, 35.415, SS_M is the value accounted for by the model. That is, the amount of variance accounted for by the grouping variable/factors, Rater and Photo.

5.4.3 Sums of Squares Residual (or within)

SS_R is error associated with within group variability. If people are randomly assigned to treatment group there should be no other covariate (confounding variable) so that all SS_R variability is *uninteresting* for the research and treated as noise.

$$SS_R = \sum (x_{ik} - \bar{x}_k)^2$$

Here's another configuration of the same:

$$SS_R = s_{group1}^2(n-1) + s_{group2}^2(n-1) + s_{group3}^2(n-1) + s_{group4}^2(n-1) + s_{group5}^2(n-1) + s_{group6}^2(n-1)$$

Again, the formula is in six chunks – but this time the calculations are *within-group*. We need the variance (the standard deviation squared) for the calculation. Let's take another look at our descriptives.

```
psych::describeBy(Negative ~ Rater + Photo, mat = TRUE, data = Ramdhani_df,
  digits = 3)
```

	item	group1	group2	vars	n	mean	sd	median	trimmed	mad	min
X11	1	Dayaknese	Dayaknese		1 17	1.818	0.768	1.692	1.783	0.694	0.706
X12	2	Javanese	Dayaknese		1 18	2.524	0.742	2.391	2.460	0.569	1.406
X13	3	Madurese	Dayaknese		1 19	3.301	1.030	3.314	3.321	1.294	1.406
X14	4	Dayaknese	Madurese		1 18	3.129	0.156	3.160	3.136	0.104	2.732
X15	5	Javanese	Madurese		1 19	3.465	0.637	3.430	3.456	0.767	2.456
X16	6	Madurese	Madurese		1 20	3.297	1.332	2.958	3.254	1.615	1.211
					max	range	skew	kurtosis	se		
X11						3.453	2.747	0.513	-0.881	0.186	
X12						4.664	3.258	1.205	1.475	0.175	
X13						4.854	3.448	-0.126	-1.267	0.236	
X14						3.423	0.691	-0.623	0.481	0.037	
X15						4.631	2.175	-0.010	-1.307	0.146	
X16						5.641	4.430	0.215	-1.238	0.298	

Calculating SS_R

$$((0.768^2) * (17 - 1)) + ((0.742^2) * (18 - 1)) + ((1.03^2) * (19 - 1)) + \\ ((0.156^2) * (18 - 1)) + ((0.637^2) * (19 - 1)) + ((1.332^2) * (20 - 1))$$

[1] 79.32078

The value for our SS_R is 79.321. Its degrees of freedom is $N - k$. That is, the total N minus the number of groups:

111 - 6

[1] 105

5.4.4 A Recap on the Relationship between SS_T , SS_M , and SS_R

$SS_T = SS_M + SS_R$ In our case:

- SS_T was 114.775
- SS_M was 35.415
- SS_R was 79.321

Considering rounding error, we were successful!

35.415 + 79.321

[1] 114.736

5.4.5 Calculating SS for Each Factor and Their Products

5.4.5.1 Rater Main Effect

SS_a : Rater is calculated the same way as SS_M for one-way ANOVA. Simply collapse across Photo and calculate the *marginal means* for Negative as a function of the Rater's ethnicity.

Reminder of the formula: $SS_{a:Rater} = \sum n_k (\bar{x}_k - \bar{x}_{grand})^2$

There are three cells involved in the calculation of SS_a : Rater.

```
psych::describeBy(Negative ~ Rater, mat = TRUE, data = Ramdhani_df, digits = 3)
```

	item	group1	vars	n	mean	sd	median	trimmed	mad	min	max	range
X11	1	Dayaknese		1	35	2.492	0.856	2.900	2.561	0.480	0.706	3.453 2.747
X12	2	Javanese		1	37	3.007	0.831	2.913	2.986	0.984	1.406	4.664 3.258
X13	3	Madurese		1	39	3.299	1.179	3.116	3.288	1.588	1.211	5.641 4.430
		skew	kurtosis			se						
X11	-0.682		-1.132	0.145								
X12	0.239		-0.923	0.137								
X13	0.117		-1.036	0.189								

Again, we need the grand mean.

```
mean(Ramdhani_df$Negative)
```

```
[1] 2.947369
```

Now to calculate the Rater main effect.

```
35 * (2.491 - 2.947)^2 + 37 * (3.007 - 2.947)^2 + 39 * (3.299 - 2.947)^2
```

```
[1] 12.24322
```

5.4.5.2 Photo Main Effect

SS_b : Photo is calculated the same way as SS_M for one-way ANOVA. Simply collapse across Rater and calculate the *marginal means* for Negative as a function of the ethnicity reflected in the Photo stimulus:

Reminder of the formula: $SS_{a:Photo} = \sum n_k (\bar{x}_k - \bar{x}_{grand})^2$.

With Photo, we have only two cells.

```
psych::describeBy(Negative ~ Photo, mat = TRUE, data = Ramdhani_df, digits = 3)
```

	item	group1	vars	n	mean	sd	median	trimmed	mad	min	max	range
X11	1	Dayaknese		1	2.575	1.043	2.449	2.516	0.921	0.706	4.854	4.148
X12	2	Madurese		1	3.300	0.871	3.166	3.280	0.667	1.211	5.641	4.430
		skew	kurtosis	se								
X11	0.47		-0.555	0.142								
X12	0.35		0.581	0.115								

Again, we need the grand mean.

```
mean(Ramdhani_df$Negative)
```

```
[1] 2.947369
```

```
54 * (2.575 - 2.947)^2 + 57 * (3.3 - 2.947)^2
```

```
[1] 14.57545
```

5.4.5.3 Interaction effect

The interaction term is simply the SS_M remaining after subtracting the SS from the main effects.

$$SS_{axb} = SS_M - (SS_a + SS_b)$$

```
35.415 - (12.243 + 14.575)
```

```
[1] 8.597
```

Let's revisit the figure I showed at the beginning of this section to see, again, how variance is partitioned.

5.4.6 Source Table Games!

As in the lesson for one-way ANOVA, we can use the hints in this source table to determine if we have statistically significance in the model. The formulas in the table provide some hints.

Summary ANOVA for Negative Reaction

Source	SS	df	$MS = \frac{SS}{df}$	$F = \frac{MS_{source}}{MS_{resid}}$	F_{CV}
Model		$k - 1$			
a		$k_a - 1$			
b		$k_b - 1$			

Source	SS	df	$MS = \frac{SS}{df}$	$F = \frac{MS_{source}}{MS_{resid}}$	F_{CV}
aXb		$(df_a)(df_b)$			
Residual		$n - k$			
Total					

```
# hand-calculating the MS values
35.415/5 #Model
```

[1] 7.083

```
12.243/2 #a: Rater
```

[1] 6.1215

```
14.575/1 #b: Photo
```

[1] 14.575

```
8.597/2 #axb interaction term
```

[1] 4.2985

```
79.321/105 #residual
```

[1] 0.7554381

```
# hand-calculating the F values
7.083/0.755 #Model
```

[1] 9.381457

```
6.122/0.755 #a: Rater
```

[1] 8.108609

```
14.575/0.755 #b: Photo
```

[1] 19.30464

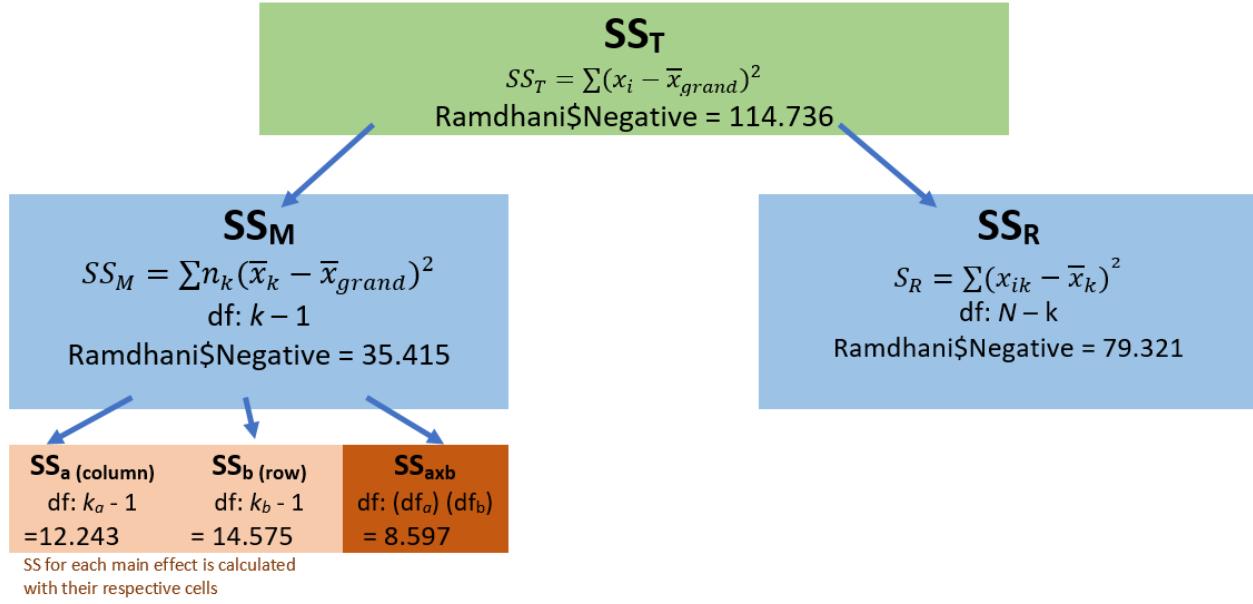


Figure 5.4: Image of a flowchart that partitions variance from sums of squares totals to its component pieces

4.299/0.755 #axb interaction term

[1] 5.69404

To find the F_{CV} we can use an [F distribution table](#).

Or use a look-up function, which follows this general form: `qf(p, df1, df2, lower.tail=FALSE)`

```
# looking up the F critical values
qf(0.05, 5, 105, lower.tail = FALSE) #Model F critical value
```

[1] 2.300888

```
qf(0.05, 2, 105, lower.tail = FALSE) #a and axb F critical value
```

[1] 3.082852

```
qf(0.05, 1, 105, lower.tail = FALSE) #b F critical value
```

[1] 3.931556

When the F value exceeds the F_{CV} , the effect is statistically significant.

 Summary ANOVA for Negative Reaction

Source	SS	df	$MS = \frac{SS}{df}$	$F = \frac{MS_{source}}{MS_{resid}}$	F_{CV}
Model	35.415	5	7.083	9.381	2.301
a	12.243	2	6.122	8.109	3.083
b	14.575	1	14.575	19.305	3.932
aXb	8.597	2	4.299	5.694	3.083
Residual	79.321	105	0.755		
Total	114.775				

5.4.7 Interpreting the results

What have we learned?

- there is a main effect for Rater
- there is a main effect for Photo
- there is a significant interaction effect

In the face of this significant interaction effect, we would follow-up by investigating the interaction effect. Why? The significant interaction effect means that findings (e.g., the story of the results) are more complex than group identity or photo stimulus, alone, can explain.

5.5 Working the Factorial ANOVA with R packages

5.5.1 Evaluating the statistical assumptions

All statistical tests have some assumptions about the data. This particular ANOVA has four:

Assumptions

- Cases represent random samples from the populations
 - This is an issue of research design
 - Although we see ANOVA used (often incorrectly) in other settings, ANOVA was really designed for the random clinical trial (RCT).
- Scores on the DV are independent of each other.
 - With correlated observations, there is a dramatic increase of Type I error
 - There are options designed for analyzing data that has dependencies (e.g., repeated measures ANOVA, dyadic data analysis, multilevel modeling)
- The DV is normally distributed for each of the populations

- that is, data for each cell (representing the combinations of each factor) is normally distributed
- Population variances of the DV are the same for all cells
 - When cell sizes are not equal, ANOVA not robust to this violation and cannot trust F ratio

Even though we position the evaluation of assumptions first – some of the best tests of the assumptions use the resulting ANOVA model. Because of this, I will quickly run the model now. I will not explain the results until after we evaluate the assumptions.

I have marked our Two-Way ANOVA Workflow with a yellow box outlined in red to let us know that we are just beginning the process of analyzing our data.

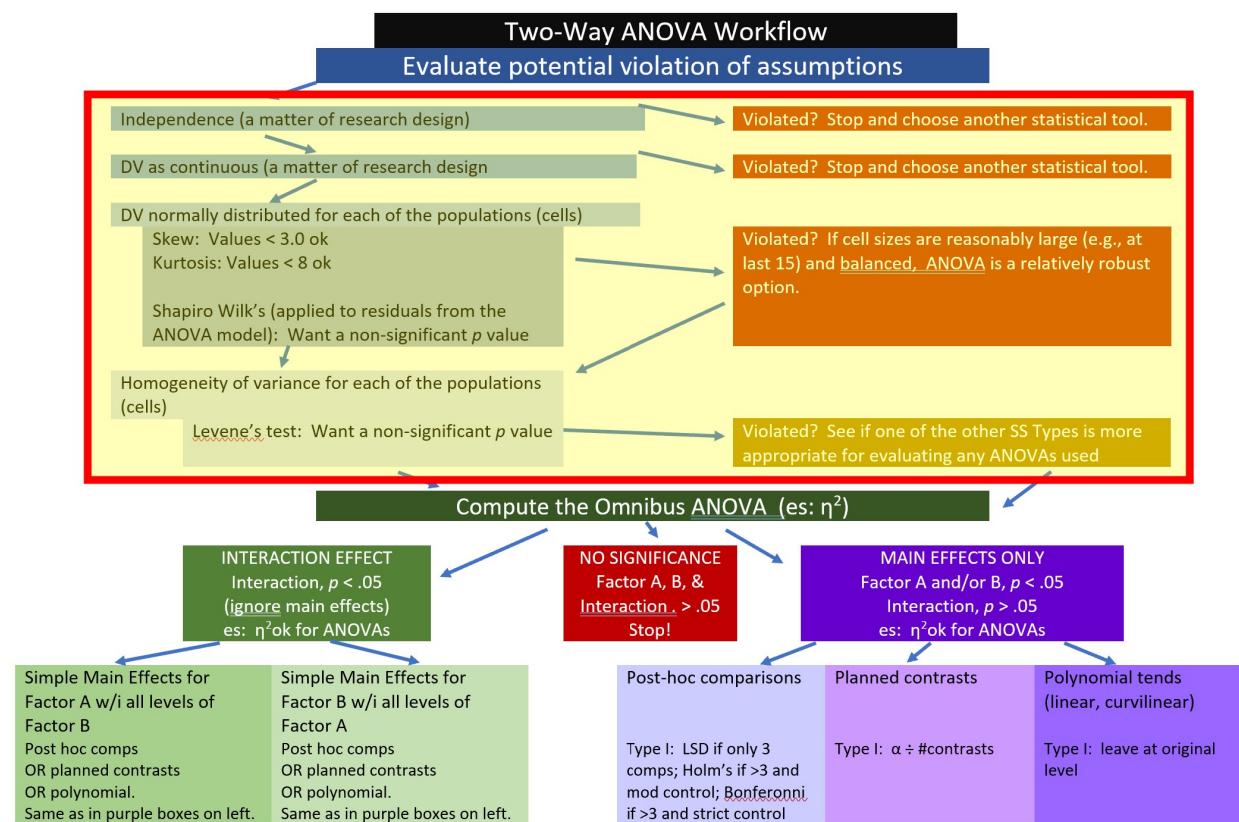


Figure 5.5: Image of a flowchart showing that we are on the “Evaluating assumptions” portion of the workflow

```
TwoWay_neg <- aov(Negative ~ Rater * Photo, Ramdhani_df)
summary(TwoWay_neg)
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Rater	2	12.21	6.103	8.077	0.000546 ***
Photo	1	14.62	14.619	19.346	0.0000262 ***

```
Rater:Photo    2   8.61   4.304   5.696  0.004480 **  
Residuals   105  79.34   0.756  
---  
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1  
  
model.tables(TwoWay_neg, "means")
```

Tables of means
Grand mean

2.947369

Rater
Dayaknese Javanese Madurese
2.492 3.007 3.299
rep 35.000 37.000 39.000

Photo
Dayaknese Madurese
2.575 3.301
rep 54.000 57.000

Rater:Photo
Photo
Rater Dayaknese Madurese
Dayaknese 1.818 3.129
rep 17.000 18.000
Javanese 2.524 3.465
rep 18.000 19.000
Madurese 3.301 3.298
rep 19.000 20.000

5.5.1.1 DV is normally distributed

Let's start by analyzing skew and kurtosis.

```
psych::describeBy(Negative ~ Rater + Photo, mat = TRUE, data = Ramdhani_df,  
digits = 3)
```

item	group1	group2	vars	n	mean	sd	median	trimmed	mad	min	
X11	1	Dayaknese	Dayaknese	1	17	1.818	0.768	1.692	1.783	0.694	0.706
X12	2	Javanese	Dayaknese	1	18	2.524	0.742	2.391	2.460	0.569	1.406
X13	3	Madurese	Dayaknese	1	19	3.301	1.030	3.314	3.321	1.294	1.406
X14	4	Dayaknese	Madurese	1	18	3.129	0.156	3.160	3.136	0.104	2.732
X15	5	Javanese	Madurese	1	19	3.465	0.637	3.430	3.456	0.767	2.456
X16	6	Madurese	Madurese	1	20	3.297	1.332	2.958	3.254	1.615	1.211

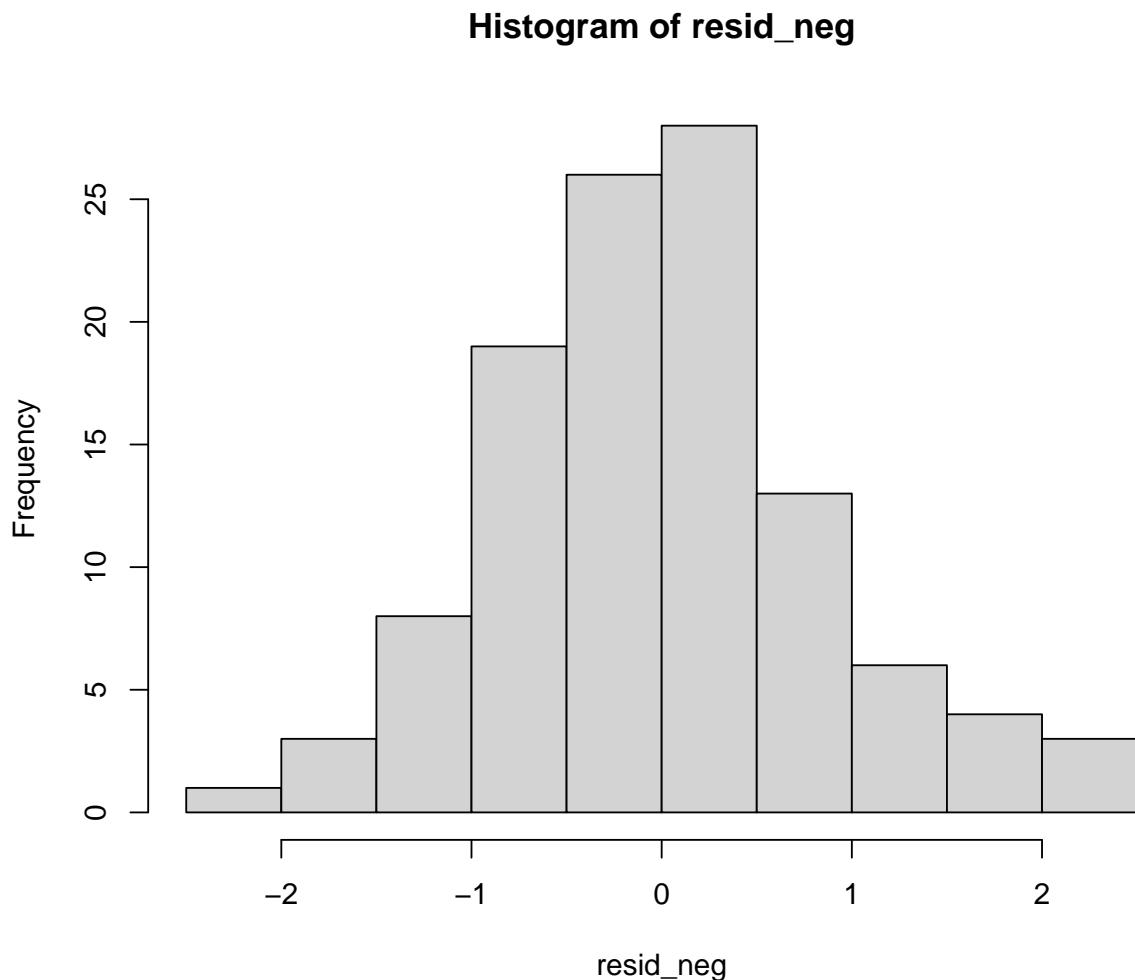
	max	range	skew	kurtosis	se
X11	3.453	2.747	0.513	-0.881	0.186
X12	4.664	3.258	1.205	1.475	0.175
X13	4.854	3.448	-0.126	-1.267	0.236
X14	3.423	0.691	-0.623	0.481	0.037
X15	4.631	2.175	-0.010	-1.307	0.146
X16	5.641	4.430	0.215	-1.238	0.298

Using guidelines from Kline [2016] our values for skewness should fall below 3.0 (they do) and all values for kurtosis should fall below 8 to 20 (ours do).

In a factorial design, the Shapiro-Wilk test is applied to residuals from the model itself. Examination of those residuals can give us a good indication of normality.

First, we extract the residuals (i.e., that which is left-over/unexplained) from the model. We can examine their distribution with a plot.

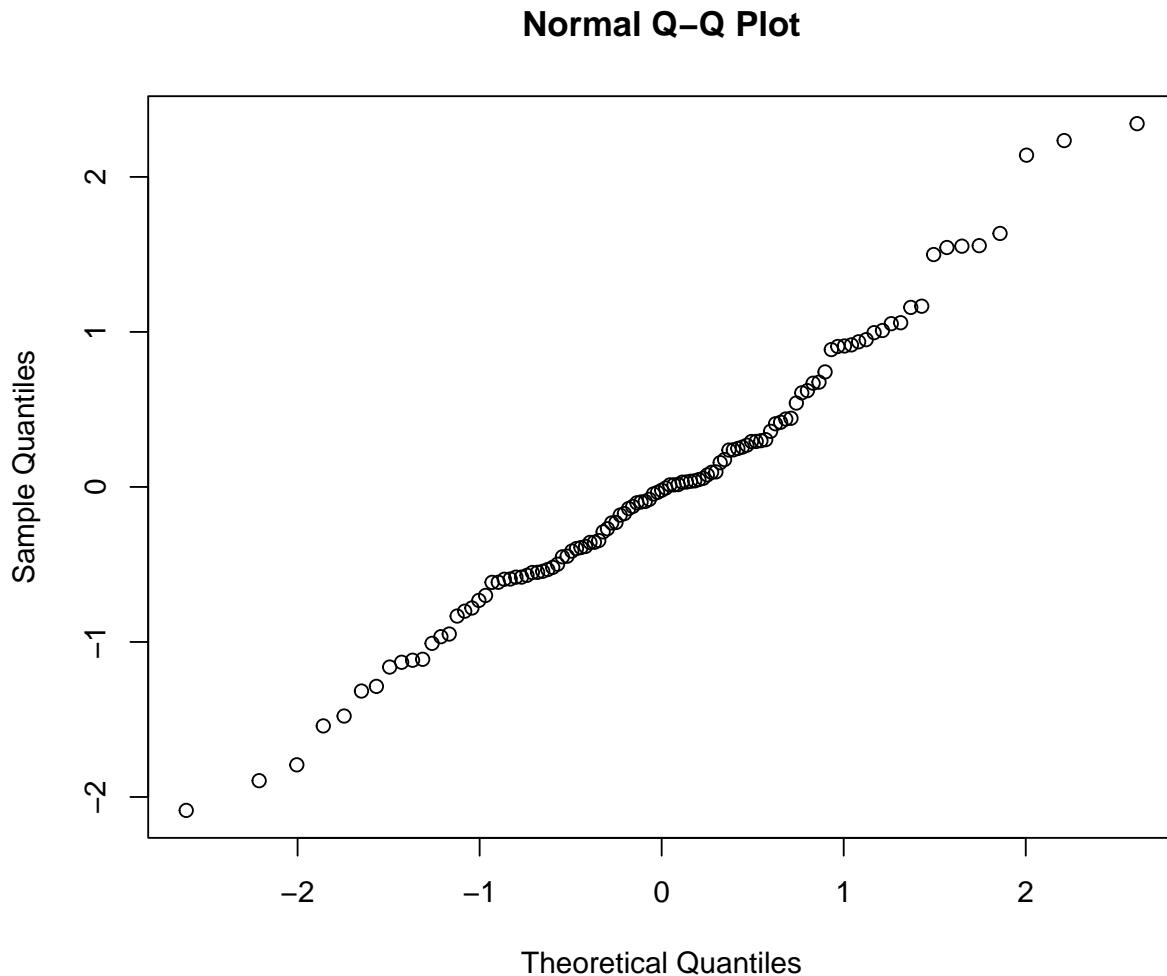
```
# creates object of residuals
resid_neg <- residuals(TwoWay_neg)
hist(resid_neg)
```



So far so good – these look normal. Let's examine a QQ plot. We want the dots (the Our distribution of *residuals* (i.e., what is leftover after the model is applied) resembles a normal distribution.

The Q-Q plot provides another view. If the residuals are normally distributed, they will line up on the diagonal line (within reason).

```
qqnorm(resid_neg)
```



We can formally test the distribution of the residuals with a Shapiro test. We want the We can formally test the distribution of the residuals with a Shapiro test. We want the associated p value to be greater than 0.05.

```
shapiro.test(resid_neg)
```

```
Shapiro-Wilk normality test

data: resid_neg
W = 0.98464, p-value = 0.2344
```

Whooo hoo! $p > 0.05$

Here's how I would summarize our data in terms of normality:

Factorial ANOVA assumes that the dependent variable is normally distributed for all cells in the design. Our analysis suggested skew and kurtosis were within the bounds considered to be normally distributed. Further, the Shapiro-Wilk normality test (applied to the residuals from the

factorial ANOVA model) suggested that the plotting of the residuals did not differ significantly from a normal distribution ($W = 0.9846, p = 0.234$).

5.5.1.2 Homogeneity of variance

We can evaluate the homogeneity of variance test with the Levene's test for the equality of error variances. Levene's requires a *fully saturated model*. This means that the prediction model requires an interaction effect (not just two, non-interacting predictors). We can use the *leveneTest()* function from the *car* package. Within the function we specify the model we will be testing in the factorial ANOVA. That is, predicting Negative from the Rater and Photo factors. The asterisk indicates that they will also be added as an interaction term.

```
car:::leveneTest(Negative ~ Rater * Photo, data = Ramdhani_df)
```

```
Levene's Test for Homogeneity of Variance (center = median)
  Df F value    Pr(>F)
group   5  8.6342 0.0000007002 ***
105
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Levene's test has indicated a violation of the homogeneity of variance assumption ($F[5, 105] = 8.634, p < .001$). This is not surprising. The boxplots shows some widely varying variances.

5.5.2 Evaluating the Omnibus ANOVA

The F -tests associated with the two-way ANOVA are the *omnibus* – providing the result for the main and interaction effects.

Here's where we are in the workflow.

When we run the two-way ANOVA we will be looking for several effects:

- main effects for each predictor, and
- the interaction effect.

It is possible that all effects will be significant, none will be significant, or some will be significant. The interaction effect always takes precedence over the main effect because it let's us know there is a more nuanced/complex story.

In specifying the ANOVA, order of entry matters. If you have a distinction between IV and moderator, put the IV first.

```
TwoWay_neg <- aov(Negative ~ Rater * Photo, Ramdhani_df)
summary(TwoWay_neg)
```

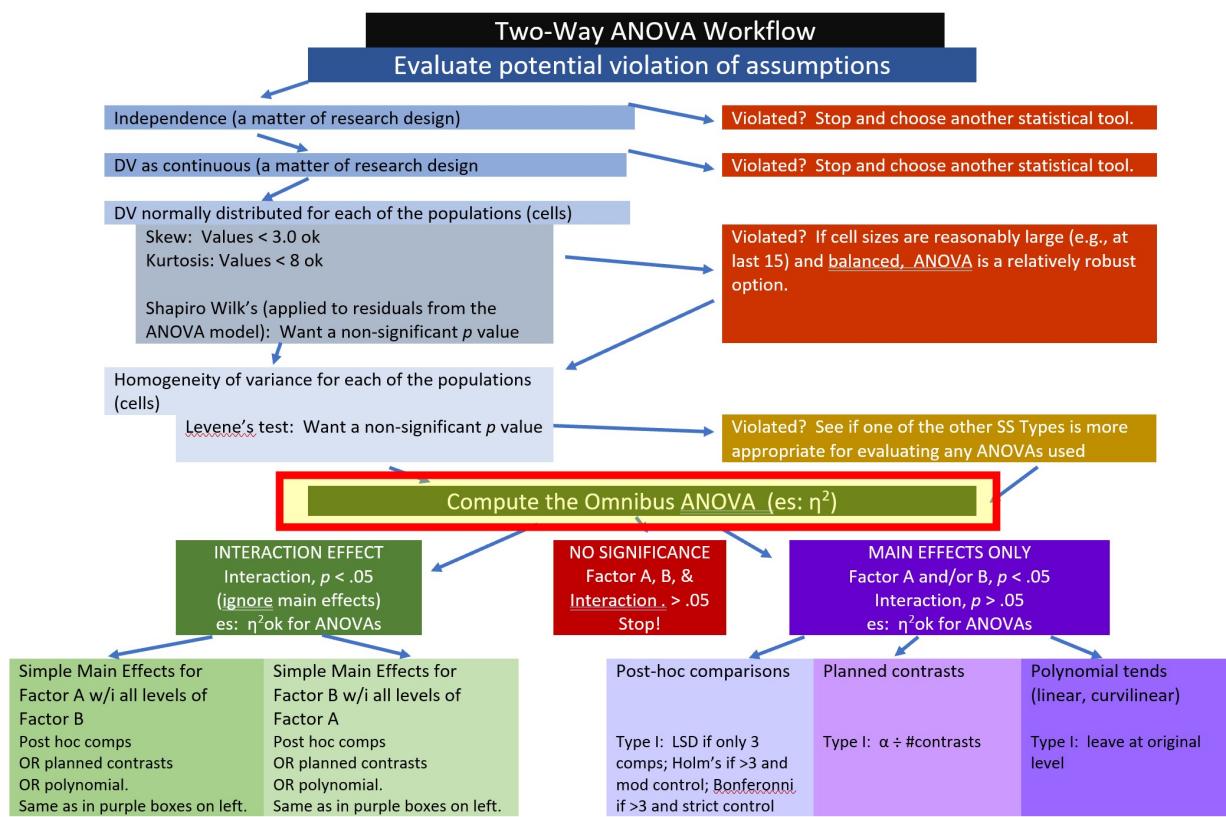


Figure 5.6: Image our place in the Two-Way ANOVA Workflow.

```
Df  Sum Sq Mean Sq F value    Pr(>F)
Rater       2   12.21   6.103   8.077  0.000546 ***
Photo        1   14.62  14.619  19.346  0.0000262 ***
Rater:Photo  2    8.61   4.304   5.696  0.004480 **
Residuals   105   79.34   0.756
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
model.tables(TwoWay_neg, "means")
```

Tables of means

Grand mean

2.947369

Rater

	Dayaknese	Javanese	Madurese
2.492	3.007	3.299	
rep	35.000	37.000	39.000

Photo

	Dayaknese	Madurese
2.575	3.301	
rep	54.000	57.000

Rater:Photo

Photo

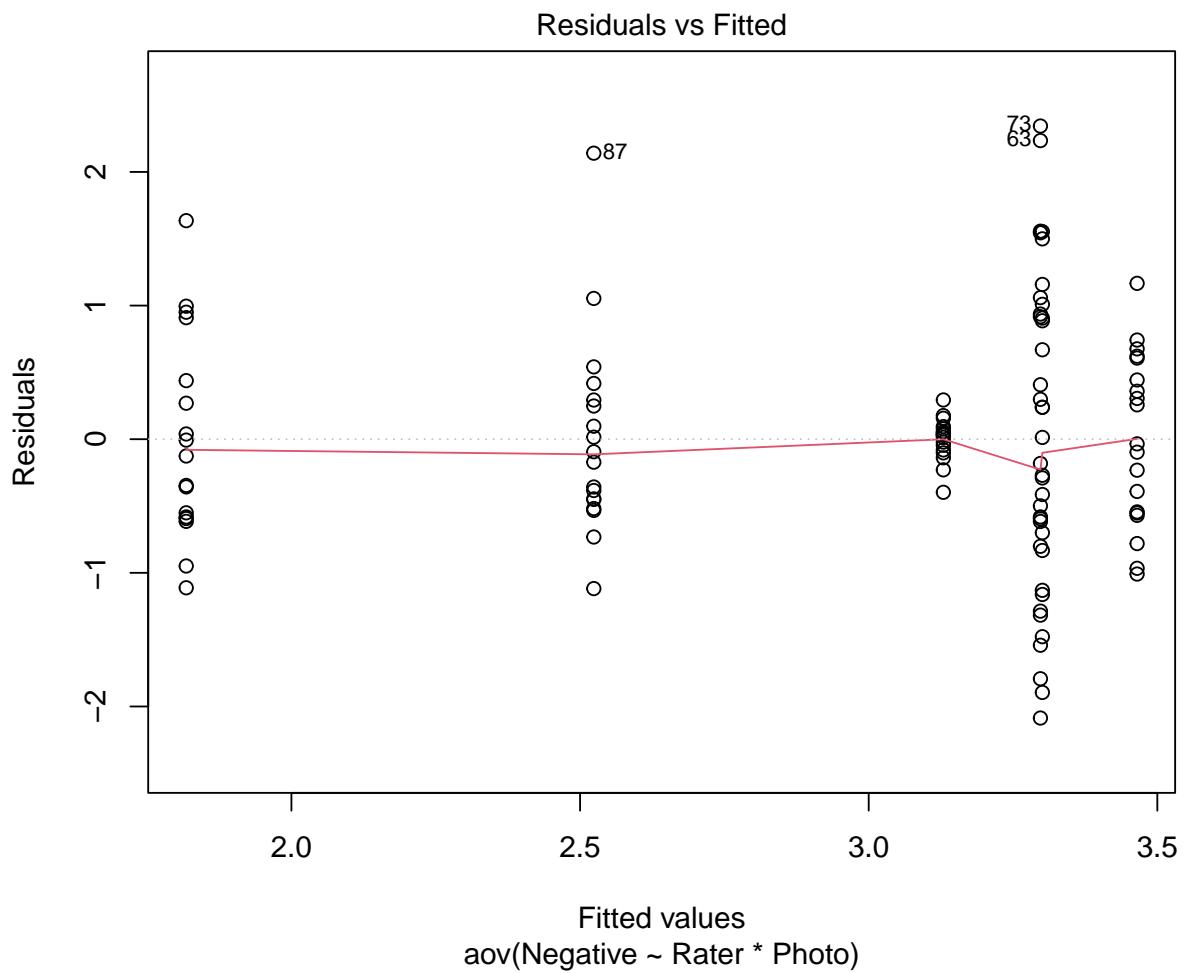
Rater	Dayaknese	Madurese
Dayaknese	1.818	3.129
rep	17.000	18.000
Javanese	2.524	3.465
rep	18.000	19.000
Madurese	3.301	3.298
rep	19.000	20.000

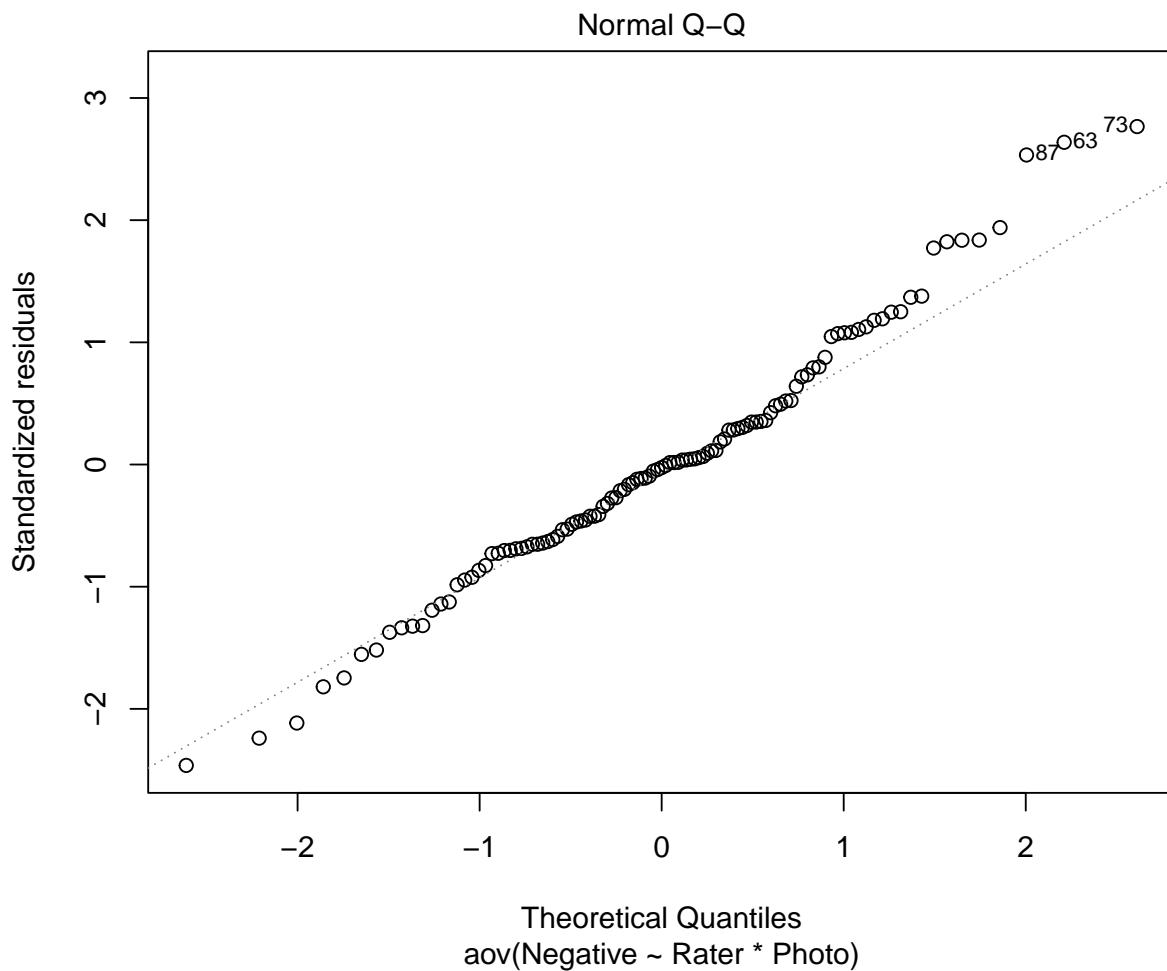
Let's write the *F strings** from the above table.

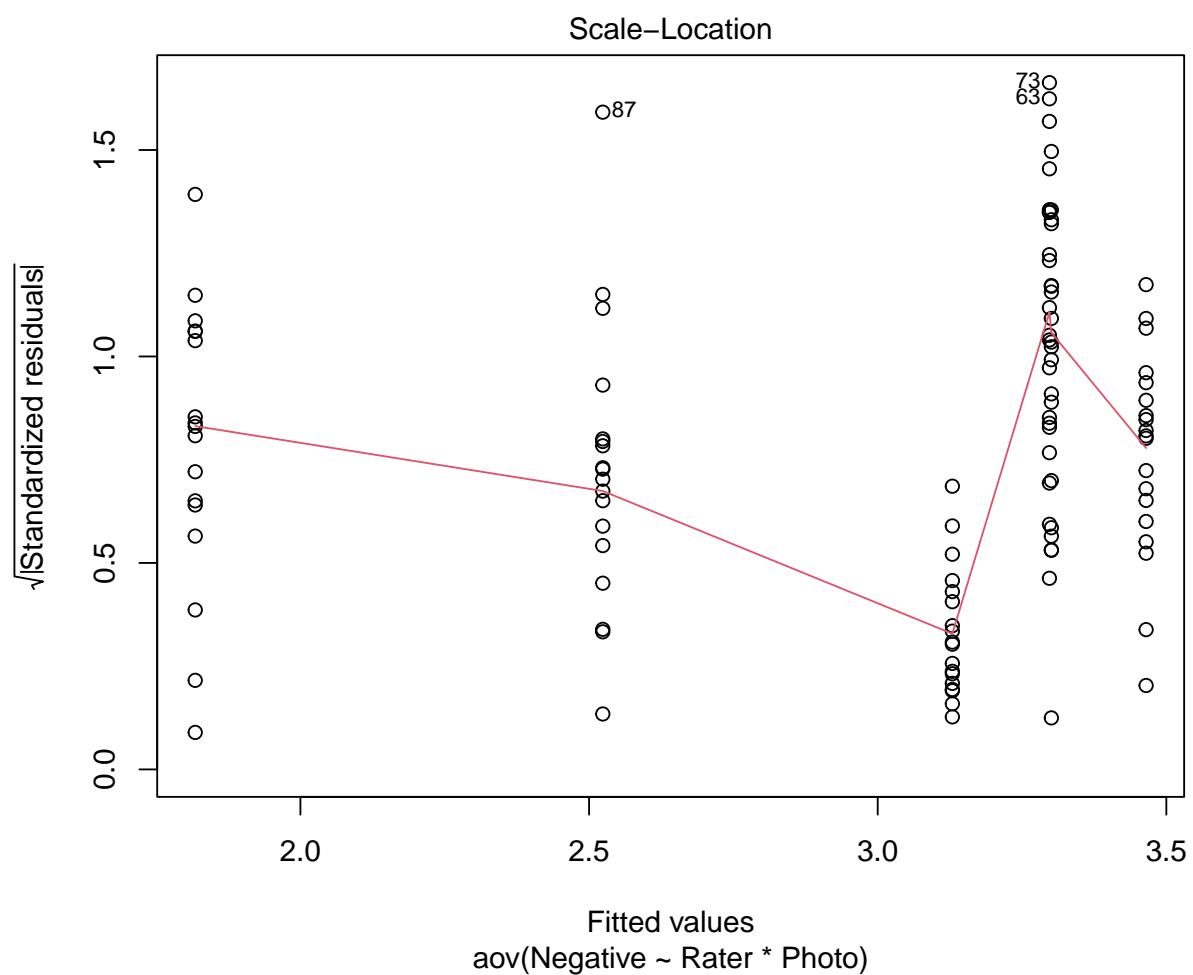
- Rater main effect: ($F[2, 105] = 8.077, p < .001$).
- Photo stimulus main effect: ($F[1, 105] = 19.346, p < .001$).
- Interaction effect: ($F[2, 105] = 5.696, p = .004$).

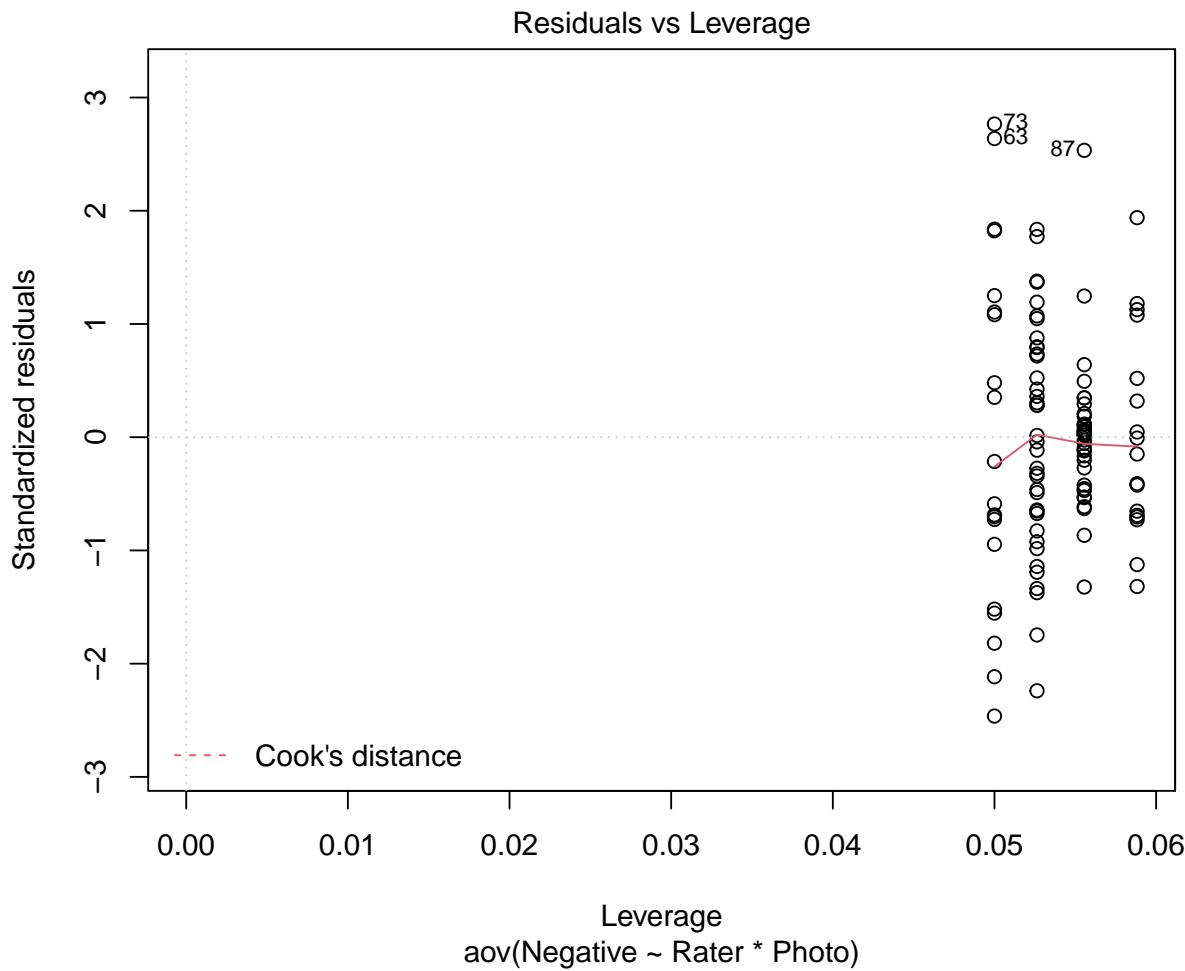
The *plot()* function provides some quick plots from the object created from the ANOVA.

```
plot(TwoWay_neg)
```









5.5.2.1 Effect sizes

Eta squared is one of the most commonly used measures of effect. It refers to the proportion of variability in the DV/outcome variable that can be explained in terms of the IVs/predictors. Traditional interpretive values are similar to the Pearson's r :

- 0 = no relationship
- .02 = small
- .13 = medium
- .26 = large
- 1.0 = a perfect (one-to-one) correspondence

The formula for η^2 is straightforward:

$$\eta^2 = \frac{SS_M}{SS_T}$$

We can apply the `etaSquared()` function from the `lsr` package to our ANOVA object to retrieve η^2 .

```
lsr::etaSquared(TwoWay_neg)
```

	eta.sq	eta.sq.part
Rater	0.10662441	0.13363091
Photo	0.12736755	0.15558329
Rater:Photo	0.07500609	0.09788289

We can update our F strings to include the effect size:

- Rater main effect: ($F[2, 105] = 8.077, p < .001, \eta^2 = 0.107$).
- Photo stimulus main effect: ($F[1, 105] = 19.346, p < .001, \eta^2 = 0.127$).
- Interaction effect: ($F[2, 105] = 5.696, p = .004, \eta^2 = 0.075$).

Before moving to follow-up, let's capture an APA style write-up so far.

5.5.2.2 APA Write-up of the omnibus results

A 3 X 2 ANOVA was conducted to evaluate the effects of rater ethnicity (3 levels, Dayaknese, Madurese, Javanese) and photo stimulus (2 levels, Dayaknese on Madurese,) on negative reactions to the photo stimuli. Results of Levene's Test for Equality of Error Variances indicated violation of the homogeneity of variance assumption, ($F[5, 105] = 8.834, p < .001$). Our analysis of the individual cell means (see Table 1 for means and standard deviations) suggested skew and kurtosis were within the bounds considered to be normally distributed [Kline, 2016]. A non-significant Shapiro-Wilk normality test (applied to the residuals from the factorial ANOVA model) provided further evidence that the assumption of normality was not violated ($W = 0.9846, p = 0.234$).

Computing sums of squares with a Type II approach, the results for the ANOVA indicated a significant main effect for ethnicity of the rater ($F[2, 105] = 8.077, p < .001, \eta^2 = 0.107$), a significant main effect for photo stimulus, ($F[1, 105] = 19.346, p < .001, \eta^2 = 0.127$), and a significant interaction effect ($F[2, 105] = 5.696, p = .004, \eta^2 = 0.075$).

Note. The next paragraph will have one of the follow-up options. We will add it later in the lesson

5.5.3 Follow-up a significant interaction effect

In factorial ANOVA we are interested in main effects and interaction effects. When the result is explained by a main effect, then there is a consistent trend as a function of a factor (e.g., Madurese raters had consistently higher Negative evaluations, irrespective of stimulus). In an interaction effect, the results are more complex (e.g., the ratings across the stimulus differed for the three groups of raters).

There are a variety of strategies to follow-up a significant interaction effect. I will demonstrate the four I believe to be the most useful in the context of psychologists operating within the scientist-practitioner-advocacy context.

When an interaction effect is significant (irrespective of the significance of one or more main effects), examination of **simple main effects** is a common statistical/explanatory approached that is used. The Two-Way ANOVA Workflow shows where we are in this process.

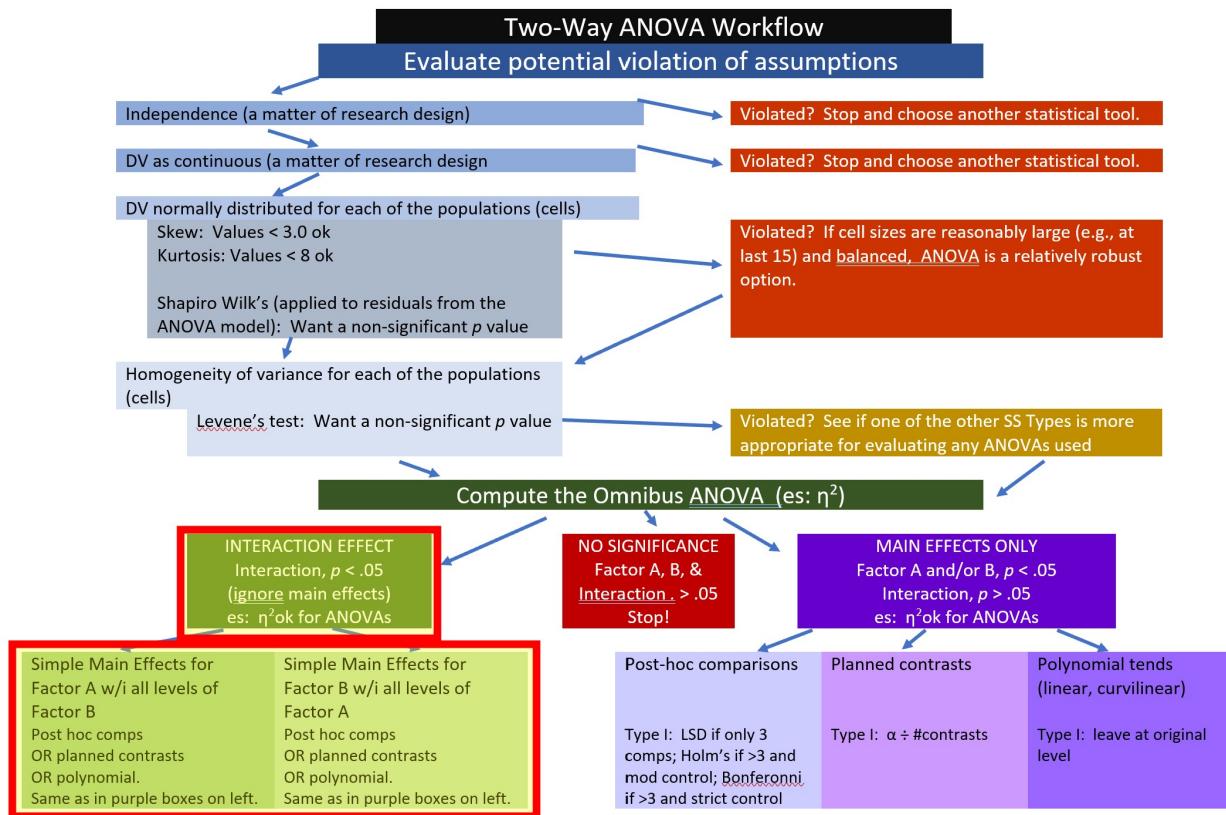


Figure 5.7: Image our place in the Two-Way ANOVA Workflow.

5.5.3.1 Option #1 the simple main effect of photo stimulus within ethnicity of the rater

Here we subset each of the three ethnic groups and then compare their ratings of the two photos. Essentially, we are conducting a one-way ANOVA for the Dyaknese ratings of the Dayaknese and Madurese photos.

```
# subset data
Dayaknese <- subset(Ramdhani_df, Rater == "Dayaknese")
# change df to subset, new model name
Dayaknese_simple <- aov(Negative ~ Photo, data = Dayaknese)
# output for simple main effect
summary(Dayaknese_simple)
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Photo	1	15.040	15.040	50.4	0.0000000395 ***
Residuals	33	9.847	0.298		

```
---
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
# effect size for simple main effect can add 'type = 1,2,3,4' to
# correspond with the ANOVA that was run
lsr::etaSquared(Dayaknese_simple, anova = FALSE)
```

```
eta.sq eta.sq.part
Photo 0.6043362 0.6043362
```

Within the Dayaknese ethnic group, there is a statistically significant difference in negative reactions to Dayaknese and Madurese photos: $F(1, 33) = 50.4$, $p < .001$, $\eta^2 = 0.60$.

Next we evaluate photo rating within the Madurese ethnic group.

```
# subset data
Madurese <- subset(Ramdhani_df, Rater == "Madurese")
# change df to subset, new model name
Madurese_simple <- aov(Negative ~ Photo, data = Madurese)
# output for simple main effect
summary(Madurese_simple)
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Photo	1	0.00	0.0001	0	0.993
Residuals	37	52.82	1.4275		

```
# effect size for simple main effect can add 'type = 1,2,3,4' to
# correspond with the ANOVA that was run
lsr::etaSquared(Madurese_simple, anova = FALSE)
```

```
eta.sq     eta.sq.part
Photo 0.000002060568 0.000002060568
```

Within the Madurese ethnic group, there was a nonsignificant difference in negative reactions to Dayaknese and Madurese photos: $F(1, 37) = 0.00$, $p = .993$, $\eta^2 < .001$.

```
# subset data
Javanese <- subset(Ramdhani_df, Rater == "Javanese")
# change df to subset, new model name
Javanese_simple <- aov(Negative ~ Photo, data = Javanese)
# output for simple main effect
summary(Javanese_simple)
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Photo	1	8.188	8.188	17.18	0.000205 ***
Residuals	35	16.678	0.477		

```
---
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
# effect size for simple main effect can add 'type = 1,2,3,4' to
# correspond with the ANOVA that was run
lsr::etaSquared(Javanese_simple, anova = FALSE)
```

```
eta.sq eta.sq.part
Photo 0.3292776 0.3292776
```

Within the Javanese ethnic group, there was a significant difference in negative reactions to Dayaknese and Madurese photos: $F(1, 35) = 17.18, p < .001, \eta^2 = 0.33$.

If I were using this approach in a 3 X 2 ANOVA, I would probably not control for Type I error. Why? I only conducted follow-up comparisons to evaluate the simple main effect of photo stimulus within rater ethnicity; that is, I would hold it at alpha = 0.05.

- Photo stimulus (Dayaknese or Madurese) within the Dayaknese ethnic group.
- Photo stimulus (Dayaknese or Madurese) within the Madurese ethnic group.
- Photo stimulus (Dayaknese or Madurese) within the Javanese ethnic group.

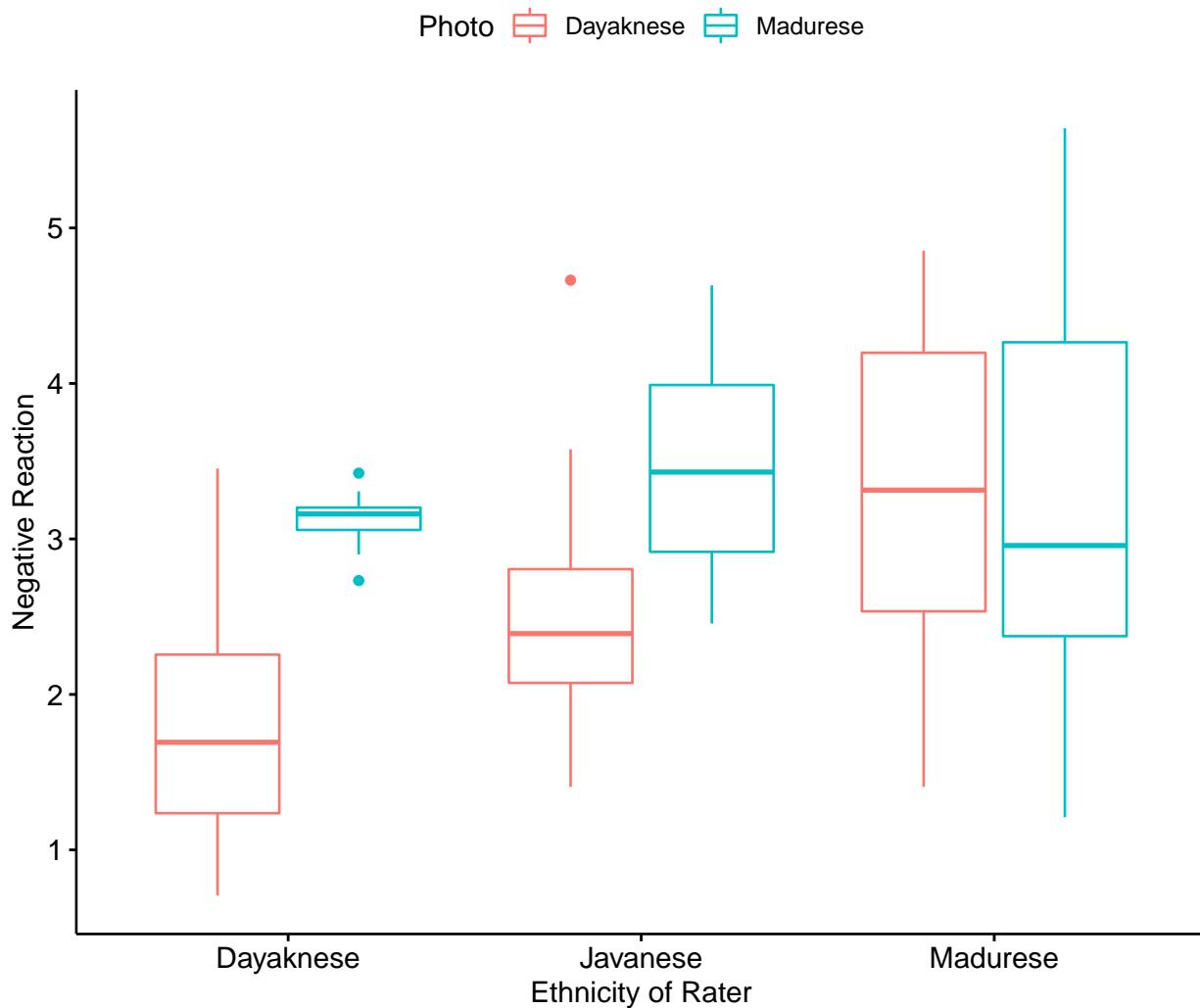
However, because it is good for instruction, it would be equally fine to use a traditional Bonferroni, dividing $.05/3 = 0.017$ and testing each at 0.017. I will use this approach in the write-up.

FAQ: Could we do the reverse simple effect, ethnicity of rater within the photo stimulus? Absolutely! The choice is yours (and sometimes the results will differ). I usually run both and then report ONE – the one that conveys the story the data has to tell. You *could* report both sets, but then you would really want to control Type I error and your repetitive contrasts are far from independent/orthogonal.

APA Style Results for Option #1 follow-up. *This would be added to the results of the omnibus two-way ANOVA.*

Option #1: To explore the interaction effect, we followed with a test of the simple main effect of photo stimulus within the ethnicity of the rater. That is, we looked at the effect of the photo stimulus within the Dayaknese, Madurese, and Javanese groups, separately. To control for Type I error across the three simple main effects, we set alpha at .017 (.05/3). Results indicated significant differences for Dayaknese ($F[1, 33] = 50.4, p < .001, \eta^2 = 0.60$) and Javanese ethnic groups ($F[1, 35] = 17.18, p < .001, \eta^2 = 0.33$), but not for the Madurese ethnic group ($F[1, 37] = 0.000, p = .993, \eta^2 < .001$). As illustrated in Figure 1, the Dayaknese and Javanese raters both reported stronger negative reactions to the Madurese. The differences in ratings for the Madurese were not statistically significantly different. In this way, the rater's ethnic group moderated the relationship between the photo stimulus and negative reactions.

```
ggpubr::ggboxplot(Ramdhani_df, x = "Rater", y = "Negative", color = "Photo",
  xlab = "Ethnicity of Rater", ylab = "Negative Reaction")
```



5.5.3.2 Option #2 the simple main effect of ethnicity of rater within photo stimulus.

In this simple main effect of ethnicity of rater (3 levels) within photo stimulus (2 levels), we will conduct two one-way ANOVAs for the Dayaknese and Madurese photos, separately. However, we will want to do orthogonal contrast-coding for rater ethnicity for the follow-up (to the follow-up).

It helps to know what the default contrast codes are; we can get that information with the `contrasts()` function.

```
contrasts(Ramdhani_df$Rater)
```

	Javanese	Madurese
Dayaknese	0	0
Javanese	1	0
Madurese	0	1

Let's create custom contrasts. Recall that an orthogonal contrast requires that there be one less contrast than the number of groups and that once a group is singled out, it cannot be compared again.

Thus, I want to compare the

- Javanese to the Dayaknese and Madurese combined, then
- Dayaknese to Madurese

```
# tell R which groups to compare
c1 <- c(1, -2, 1)
c2 <- c(-1, 0, 1)
mat <- cbind(c1, c2) #combine the above bits
contrasts(Ramdhani_df$Rater) <- mat # attach the contrasts to the variable
```

This allows us to recheck the contrasts.

```
contrasts(Ramdhani_df$Rater)
```

	c1	c2
Dayaknese	1	-1
Javanese	-2	0
Madurese	1	1

Yes, in contrast 1 we are comparing the Javanese to the combined Dayaknese and Madurese. In contrast 2 we are comparing the Dayaknese to the Madureses.

```
# subset data
Dayaknese_Ph <- subset(Ramdhani_df, Photo == "Dayaknese")
# change df to subset, new model name
Dykn_simple <- aov(Negative ~ Rater, data = Dayaknese_Ph)
# output for simple main effect
summary(Dykn_simple)
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)						
Rater	2	19.81	9.903	13.32	0.0000221 ***						
Residuals	51	37.90	0.743								

Signif. codes:	0	'***'	0.001	'**'	0.01	'*'	0.05	'..'	0.1	' '	1

```
# effect size for simple main effect can add 'type = 1,2,3,4' to
# correspond with the ANOVA that was run
lsr::etaSquared(Dykn_simple, anova = FALSE)
```

```
eta.sq eta.sq.part
Rater 0.3432006 0.3432006
```

We can capture the F string from this output: $F [2, 51] = 13.32, p < .001, \eta^2 = 0.343$.

This code produces the contrasts we specified. Note that in our code we can improve the interpretability of the output by adding labels. We know the specific contrasts from our prior work.

```
summary.aov(Dykn_simple, split = list(Rater = list(`Javanese v Dayaknese and Madurese` = 1,
                                             `Dayaknese Madurese` = 2)))
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Rater	2	19.81	9.903	13.325	0.00002211
Rater: Javanese v Dayaknese and Madurese	1	0.07	0.071	0.095	0.759
Rater: Dayaknese Madurese	1	19.73	19.735	26.554	0.00000419
Residuals	51	37.90	0.743		

Rater ***
 Rater: Javanese v Dayaknese and Madurese ***
 Rater: Dayaknese Madurese ***
 Residuals

 Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

The simple main effect of ethnicity of the rater within the reaction to the photos of members of the Dayaknese ethnic group was statistically significant: $F [2, 51] = 13.32, p < .001, \eta^2 = 0.343$. Follow-up testing indicated non-significant differences when the ratings from members of the Javanese ethnic group were compared to the Dayaknese and Madurese, combined ($F [1, 51] = 0.095, p = .759$). There was a statistically significant difference when Dayaknese and Madurese raters were compared ($F [1, 51] = 26.554, p < .001$)

We repeat the simple main effect process when the Madurese photos were the stimulus.

```
# subset data
Madurese_Ph <- subset(Ramdhani_df, Photo == "Madurese")
# change df to subset, new model name
Mdrs_simple <- aov(Negative ~ Rater, data = Madurese_Ph)
# output for simple main effect
summary(Mdrs_simple)
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Rater	2	1.04	0.5207	0.679	0.512
Residuals	54	41.44	0.7674		

```
# effect size for simple main effect can add 'type = 1,2,3,4' to
# correspond with the ANOVA that was run
lsr::etaSquared(Mdrs_simple, anova = FALSE)
```

```
eta.sq eta.sq.part
Rater 0.02451385 0.02451385
```

Let's capture the F string for ratings of the Madurese photos: $F [2, 54] = 0.679, p = .512, \eta^2 = 0.024$.

We can use the procedure described above to obtain our orthogonal contrasts.

```
summary.aov(Mdrs_simple, split = list(Rater = list(`Javanese v Dayaknese and Madurese` = 1,
                                              `Dayaknese Madurese` = 2)))
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Rater	2	1.04	0.5207	0.679	0.512
Rater: Javanese v Dayaknese and Madurese	1	0.77	0.7734	1.008	0.320
Rater: Dayaknese Madurese	1	0.27	0.2679	0.349	0.557
Residuals	54	41.44	0.7674		

Here's a write-up of this portion of the result.

The simple main effect of ethnicity of the rater within rating the photos of Madurese people was not statistically significant: ($F [2, 54] = 0.679, p = .512, \eta^2 = 0.024$). Correspondingly, follow-up testing indicated non-significant differences when the ratings of the Javanese were compared to Dayaknese and Madurese, combined ($F [1, 54] = 1.008, p = .320$) and when the ratings of the Dayaknese and Madurese were compared ($F [1, 54] = 0.349, p = .557$)

To control for Type I error, we have 4 follow-up contrasts (2 for Dayaknese, 2 for Madurese). We'll control Type I error with $.05/4 = .0125$

0.05/4

[1] 0.0125

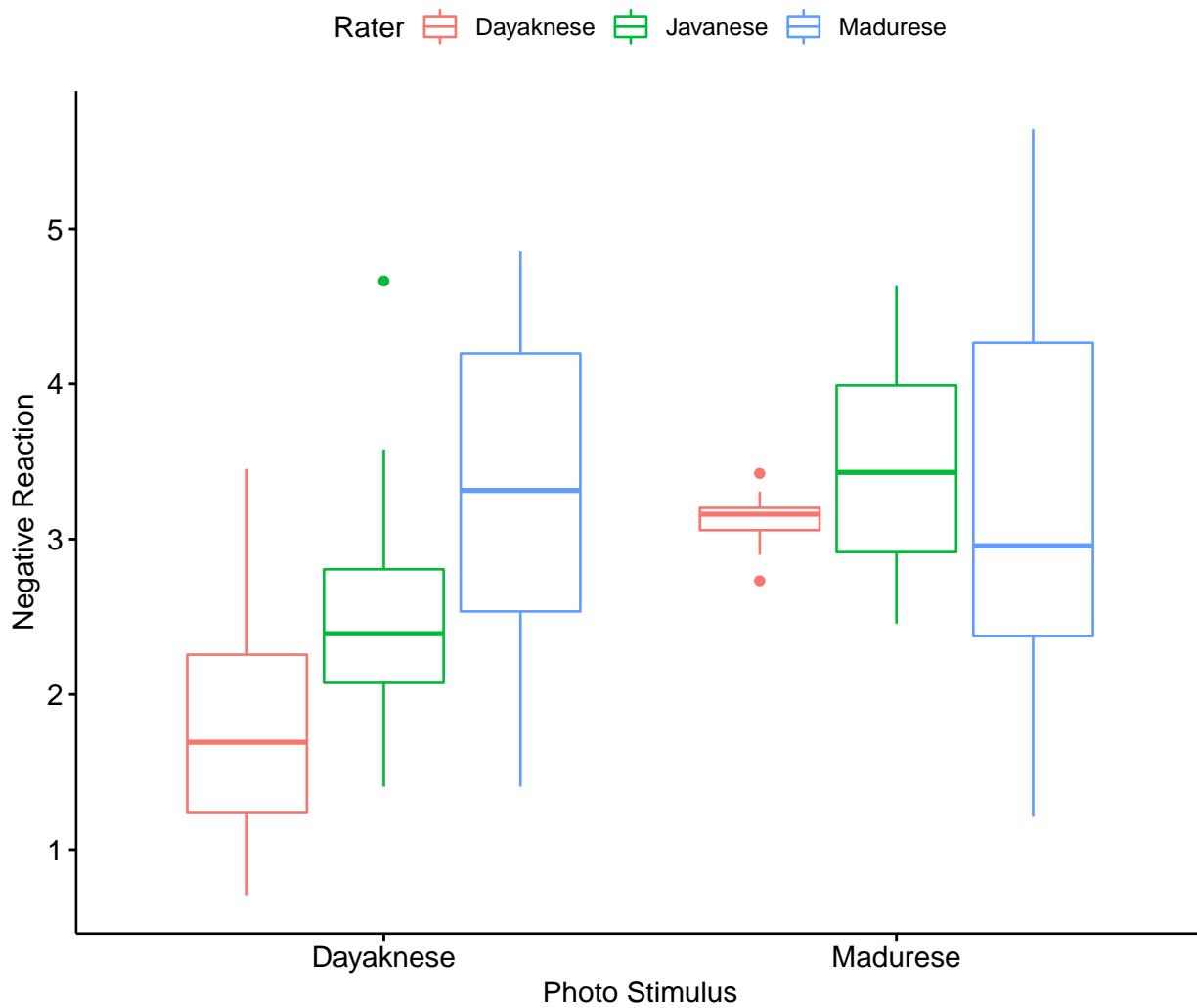
APA Write-up of the simple main effect of photo stimulus within rater ethnicity.

This would be added to the write-up of the omnibus two-way ANOVA test.

Option #2: To explore the interaction effect, we followed with tests of simple effect of rater ethnicity within the photo stimulus. That is, we looked at the effect of each rater's ethnicity within the Madurese and Dayaknese photo stimulus, separately. Our first analysis evaluated the effect of the rater's ethnicity when evaluating the Dayaknese photo; our second analysis evaluated effect of the rater's ethnicity when evaluating the Madurese photo. To control for Type I error across the two simple main effects, we set alpha at $.0125 (.05/4)$. The simple main effect of ethnicity of the rater within the reaction to the photos of members of the Dayaknese ethnic group was statistically significant: $F [2, 51] = 13.32, p < .001, \eta^2 = 0.343$. Follow-up testing indicated non-significant differences when the ratings from members of the Javanese ethnic group were compared to the Dayaknese and Madurese, combined ($F [1, 51] = 0.095, p = .759$). There was a statistically significant difference when Dayaknese and Madurese raters were compared ($F [1, 51] = 26.554, p < .001$). The simple main effect of ethnicity of the rater within when rating the photos of Madurese people was not statistically significant: ($F [2, 54] = 0.679, p = .512, \eta^2 = 0.024$). Correspondingly, follow-up testing indicated non-significant differences when the ratings of the Javanese were compared to Dayaknese and Madurese, combined ($F [1, 54] = 1.008, p = .320$) and when the ratings of the Dayaknese and Madurese were compared ($F [1, 54] = 0.349, p = .557$).

This moderating effect of ethnicity of the rater on the negative reaction to the photo stimulus is illustrated in Figure 1.

```
ggpubr::ggboxplot(Ramdhani_df, x = "Photo", y = "Negative", color = "Rater",
  xlab = "Photo Stimulus", ylab = "Negative Reaction")
```



5.5.3.3 Option #3 post hoc comparisons

Another option is compare all possible cells. These are termed *post hoc comparisons*. They are an alternative to simple main effects; you would not report both. The figure shows our place on the Two-Way ANOVA Workflow.

As the numbers of levels increase, post hoc comparisons become somewhat unwieldy. Even though this procedure produces them all, you can select which sensible number you want to compare and control for Type I error according to the number in that set.

With rater ethnicity (3 levels) and photo stimulus (2 levels), we have 6 groupings. When k is the number of groups, the total number of paired comparisons is: $k(k-1)/2$

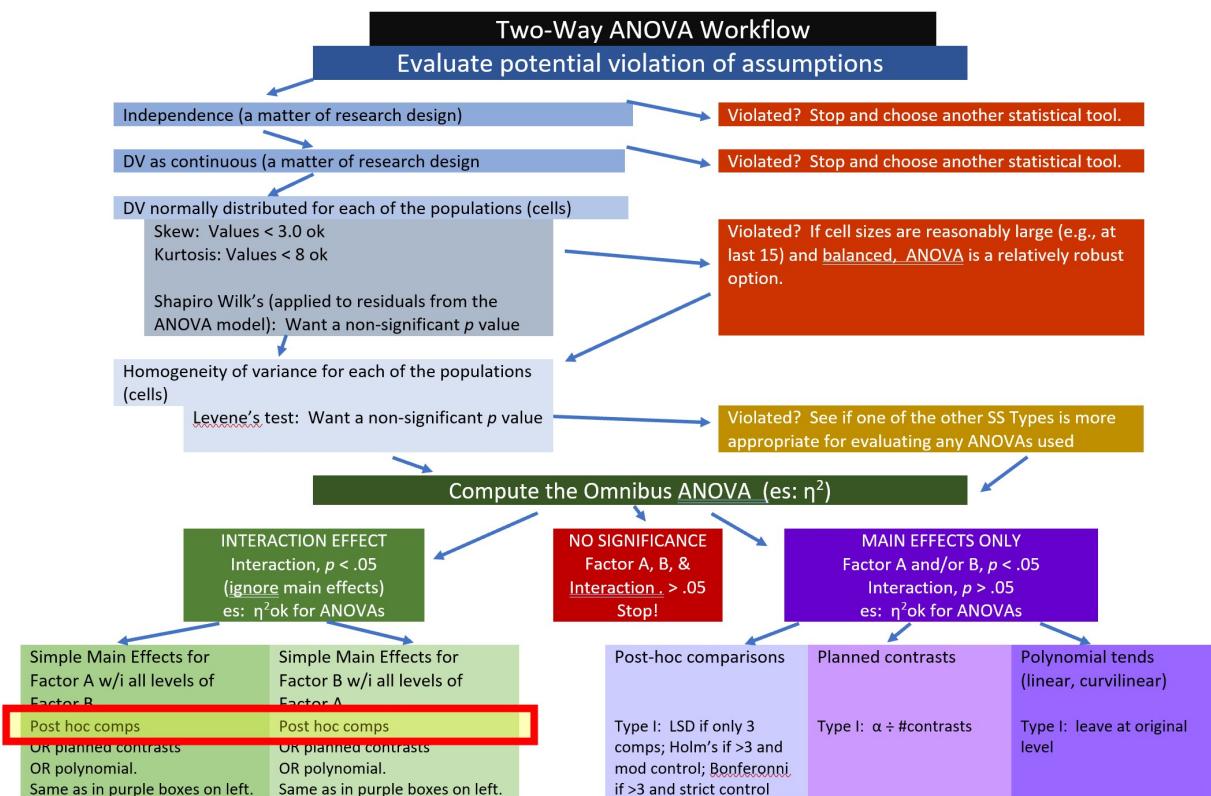


Figure 5.8: Image our place in the Two-Way ANOVA Workflow.

```
6 * (6 - 1)/2
```

```
[1] 15
```

Obtain the 15 post-hoc paired comparisons with the *TukeyHSD()* function.

```
posthocs <- TukeyHSD(TwoWay_neg, ordered = TRUE)
posthocs
```

```
Tukey multiple comparisons of means
 95% family-wise confidence level
 factor levels have been ordered
```

```
Fit: aov(formula = Negative ~ Rater * Photo, data = Ramdhani_df)
```

```
$Rater
```

	diff	lwr	upr	p	adj
Javanese-Dayaknese	0.5147954	0.02750358	1.0020872	0.0358235	
Madurese-Dayaknese	0.8068425	0.32566283	1.2880222	0.0003629	
Madurese-Javanese	0.2920471	-0.18222911	0.7663234	0.3124227	

```
$Photo
```

	diff	lwr	upr	p	adj
Madurese-Dayaknese	0.726071	0.3987575	1.053385	0.0000262	

```
$`Rater:Photo`
```

	diff	lwr	upr		
Javanese:Dayaknese-Dayaknese:Dayaknese	0.706013072	-0.14743916	1.5594653		
Dayaknese:Madurese-Dayaknese:Dayaknese	1.311568627	0.45811640	2.1650209		
Madurese:Madurese-Dayaknese:Dayaknese	1.479735294	0.64726775	2.3122028		
Madurese:Dayaknese-Dayaknese:Dayaknese	1.483077399	0.64060458	2.3255502		
Javanese:Madurese-Dayaknese:Dayaknese	1.647182663	0.80470985	2.4896555		
Dayaknese:Madurese-Javanese:Dayaknese	0.605555556	-0.23561614	1.4467273		
Madurese:Madurese-Javanese:Dayaknese	0.773722222	-0.04615053	1.5935950		
Madurese:Dayaknese-Javanese:Dayaknese	0.777064327	-0.05296553	1.6070942		
Javanese:Madurese-Javanese:Dayaknese	0.941169591	0.11113973	1.7711995		
Madurese:Madurese-Dayaknese:Madurese	0.168166667	-0.65170609	0.9880394		
Madurese:Dayaknese-Dayaknese:Madurese	0.171508772	-0.65852109	1.0015386		
Javanese:Madurese-Dayaknese:Madurese	0.335614035	-0.49441582	1.1656439		
Madurese:Dayaknese-Madurese:Madurese	0.003342105	-0.80509532	0.8117795		
Javanese:Madurese-Madurese:Madurese	0.167447368	-0.64099006	0.9758848		
Javanese:Madurese-Madurese:Dayaknese	0.164105263	-0.65463115	0.9828417		
				p	adj
Javanese:Dayaknese-Dayaknese:Dayaknese	0.1652148				
Dayaknese:Madurese-Dayaknese:Dayaknese	0.0002907				
Madurese:Madurese-Dayaknese:Dayaknese	0.0000171				

```
Madurese:Dayaknese-Dayaknese:Dayaknese 0.0000211
Javanese:Madurese-Dayaknese:Dayaknese 0.0000018
Dayaknese:Madurese-Javanese:Dayaknese 0.3005963
Madurese:Madurese-Javanese:Dayaknese 0.0760131
Madurese:Dayaknese-Javanese:Dayaknese 0.0802217
Javanese:Madurese-Javanese:Dayaknese 0.0166363
Madurese:Madurese-Dayaknese:Madurese 0.9911395
Madurese:Dayaknese-Dayaknese:Madurese 0.9908344
Javanese:Madurese-Dayaknese:Madurese 0.8482970
Madurese:Dayaknese-Madurese:Madurese 1.0000000
Javanese:Madurese-Madurese:Madurese 0.9907331
Javanese:Madurese-Madurese:Dayaknese 0.9920328
```

If we want to consider all 15 pairwise comparisons and also control for Type I error, a Holm's sequential Bonerroni [Green and Salkind, 2014b] will help us take a middle-of-the-road approach (not as strict as .05/15 with the traditional Bonferroni; not as lenient as "none") to managing Type I error.

With the Holms, we rank order the p values associated with the 15 comparisons in order from lowest (e.g., .0000018) to highest (e.g., 1.000). The first p value is evaluated with the most strict criterion (.05/15; the traditional Bonferonni approach). Then, each successive comparison calculates the p value by using the number of *remaining* comparisons as the denominator (e.g., .05/14, .05/13, .05/12). As the p values rise and the alpha levels relax, there will be a cut-point where remaining comparisons are not statistically significant.

0.05/15

```
[1] 0.003333333
```

0.05/14

```
[1] 0.003571429
```

To facilitate this contrast, let's extract the 15 TukeyHSD tests and work with them in Excel.

First, obtain the structure of the *posthoc* object

`str(posthocs)`

List of 3

```
$ Rater      : num [1:3, 1:4] 0.5148 0.8068 0.292 0.0275 0.3257 ...
..- attr(*, "dimnames")=List of 2
... ..$ : chr [1:3] "Javanese-Dayaknese" "Madurese-Dayaknese" "Madurese-Javanese"
... ..$ : chr [1:4] "diff" "lwr" "upr" "p adj"
$ Photo     : num [1, 1:4] 0.726071 0.3987575 1.0533845 0.0000262
..- attr(*, "dimnames")=List of 2
```

```

... .$. : chr "Madurese-Dayaknese"
... .$. : chr [1:4] "diff" "lwr" "upr" "p adj"
$ Rater:Photo: num [1:15, 1:4] 0.706 1.312 1.48 1.483 1.647 ...
..- attr(*, "dimnames")=List of 2
... $. : chr [1:15] "Javanese:Dayaknese-Dayaknese:Dayaknese" "Dayaknese:Madurese-Dayaknese:Da
... $. : chr [1:4] "diff" "lwr" "upr" "p adj"
- attr(*, "class")= chr [1:2] "TukeyHSD" "multicomp"
- attr(*, "orig.call")= language aov(formula = Negative ~ Rater * Photo, data = Ramdhani_df)
- attr(*, "conf.level")= num 0.95
- attr(*, "ordered")= logi TRUE

write.csv(posthocs$"Rater:Photo", "posthocsOUT.csv")

```

In Excel, I would sort my results by their p values (low to high) and consider my threshold ($p < .0033$) to determine which effects were statistically significant. Using the strictest criteria of $p < .0033$, we would have four statistically significant values.

	diff	lwr	upr	p adj
Javanese:Madurese-Dayaknese:Dayaknese	1.647182663	0.80470985	2.489655478	0.00000182
Madurese:Madurese-Dayaknese:Dayaknese	1.479735294	0.64726775	2.312202839	0.00001714
Madurese:Dayaknese-Dayaknese:Dayaknese	1.483077399	0.64060458	2.325550215	0.00002114
Dayaknese:Madurese-Dayaknese:Dayaknese	1.311568627	0.4581164	2.165020855	0.00029074
Javanese:Madurese-Javanese:Dayaknese	0.941169591	0.11113973	1.77119945	0.01663633
Madurese:Madurese-Javanese:Dayaknese	0.773722222	-0.0461505	1.593594978	0.07601309
Madurese:Dayaknese-Javanese:Dayaknese	0.777064327	-0.0529655	1.607094187	0.08022174
Javanese:Dayaknese-Dayaknese:Dayaknese	0.706013072	-0.1474392	1.559465299	0.16521479
Dayaknese:Madurese-Javanese:Dayaknese	0.605555556	-0.2356161	1.446727254	0.30059630
Javanese:Madurese-Dayaknese:Madurese	0.335614035	-0.4944158	1.165643895	0.84829701
Javanese:Madurese-Madurese:Madurese	0.167447368	-0.6409901	0.975884798	0.99073306
Madurese:Dayaknese-Dayaknese:Madurese	0.171508772	-0.6585211	1.001538632	0.99083435
Madurese:Madurese-Dayaknese:Madurese	0.168166667	-0.6517061	0.988039422	0.99113951
Javanese:Madurese-Madurese:Dayaknese	0.164105263	-0.6546311	0.982841674	0.99203282
Madurese:Dayaknese-Madurese:Madurese	0.003342105	-0.8050953	0.811779534	1.00000000

Figure 5.9: Image of the results of the Holms sequential Bonferroni.

I would ask, “Is this what we want?” Similar to the simple main effects we just tested, I am interested in two sets of comparisons:

First, how are the two sets of photos (Madurese and Dayaknese) rated within each set of raters.

- Javanese:Madurese - Javanese:Dayaknese
- Dayaknese:Madurese - Dayaknese:Dayaknese
- Madurese:Madurese - Madurese:Dayaknese

Second, focused on each photo, what are the relative ratings.

- Javanese:Madurese - Dayaknese:Madurese

- Madurese: Madurese - Dayaknese:Madurese
- Javanese:Dayaknese - Dayaknese:Dayaknese
- Madurese: Dayaknese - Dayaknese:Dayaknese

This is only seven sets of comparisons and would considerably reduce the alpha:

0.05/7

[1] 0.007142857

Below I have greyed-out the comparisons that are less interesting to me and left the seven that are my focal interest. I have highlighted in green the two comparisons that are statistically significant based on the Holms' sequential criteria. In this case, it does not make any difference in our interpretation of these focal predictors.

	diff	lwr	upr	p adj
Javanese:Madurese-Dayaknese:Dayaknese	1.647182663	0.80470985	2.489655478	0.00000182
Madurese:Madurese-Dayaknese:Dayaknese	1.479735294	0.64726775	2.312202839	0.00001714
Madurese:Dayaknese-Dayaknese:Dayaknese	1.483077399	0.64060458	2.325550215	0.00002114
Dayaknese:Madurese-Dayaknese:Dayaknese	1.311568627	0.4581164	2.165020855	0.00029074
Javanese:Madurese-Javanese:Dayaknese	0.941169591	0.11113973	1.77119945	0.01663633
Madurese:Madurese-Javanese:Dayaknese	0.773722222	-0.0461505	1.593594978	0.07601309
Madurese:Dayaknese-Javanese:Dayaknese	0.777064327	-0.0529655	1.607094187	0.08022174
Javanese:Dayaknese-Dayaknese:Dayaknese	0.706013072	-0.1474392	1.559465299	0.16521479
Dayaknese:Madurese-Javanese:Dayaknese	0.605555556	-0.2356161	1.446727254	0.30059630
Javanese:Madurese-Dayaknese:Madurese	0.335614035	-0.4944158	1.165643895	0.84829701
Javanese:Madurese-Madurese:Madurese	0.167447368	-0.6409901	0.975884798	0.99073306
Madurese:Dayaknese-Dayaknese:Madurese	0.171508772	-0.6585211	1.001538632	0.99083435
Madurese:Madurese-Dayaknese:Madurese	0.168166667	-0.6517061	0.988039422	0.99113951
Javanese:Madurese-Madurese:Dayaknese	0.164105263	-0.6546311	0.982841674	0.99203282
Madurese:Dayaknese-Madurese:Madurese	0.003342105	-0.8050953	0.811779534	1.00000000

Figure 5.10: Image of the results of the Holms sequential Bonferroni.

5.5.3.4 Option #4 polynomial trends

In the context of the significant interaction effect, we might also be interested in polynomial trends for any simple main effects where 3 or more cells are compared.

Why? If there are only two cells being compared, then the significance of that has already been tested and if significant, it is also a significant linear effect (because the shape between any two points is a line). Below is a figure of where the polynomial test of an interaction effect may fall in the process.

At the outset, let me acknowledge that this is not the best example to demonstrate a polynomial trend. Why? We do not necessarily have an ordered prediction across categories for this vignette.

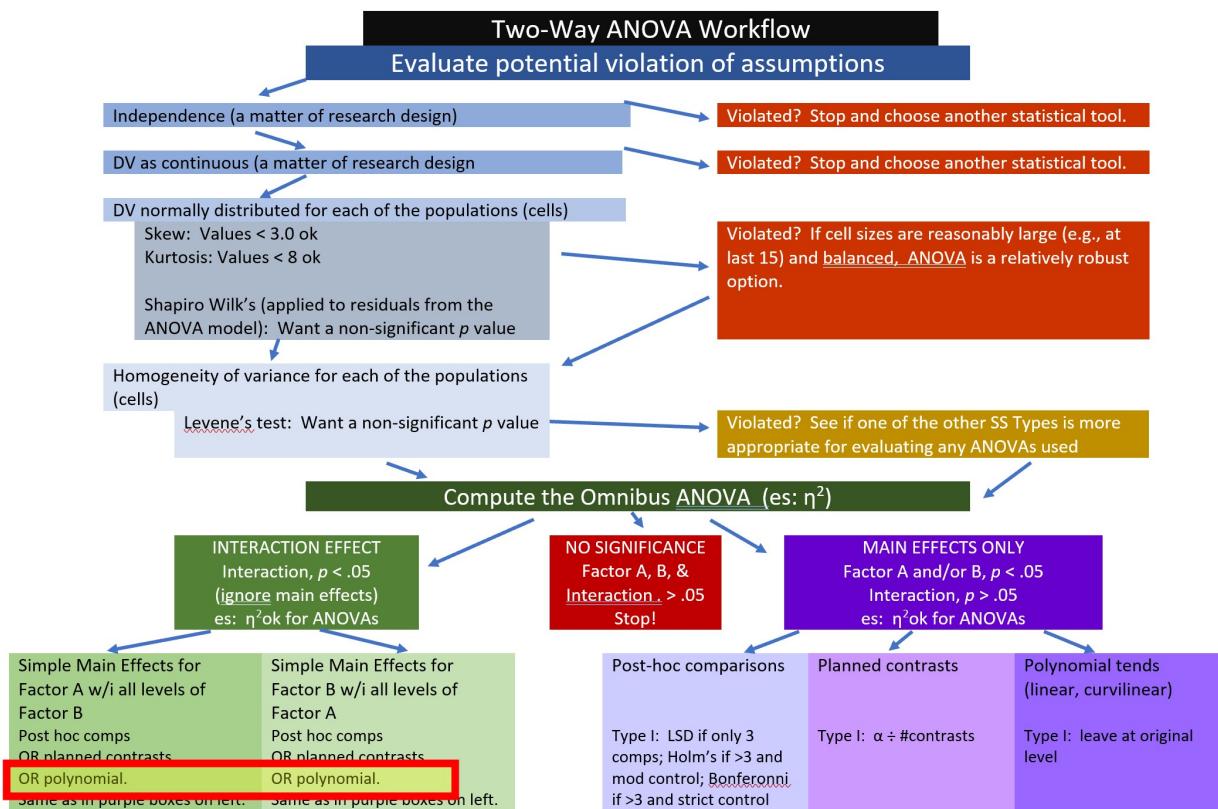


Figure 5.11: Image our place in the Two-Way ANOVA Workflow.

Other research scenarios (e.g., when dosage is none, low, high) are more readily suited for this approach.

In our example, Rater has three groups. Thus, we could evaluate a polynomial for the simple main effect of ethnicity of the rater within photo stimulus (separately for the photos of the Dayaknese and Madurese). We conduct these separately for Dayaknese, Madurese, and Javanese groups.

In the event that more than one polynomial trend select the higher one. For example, if both linear and quadratic are selected, interpret the quadratic trend

```
contrasts(Dayaknese_Ph$Rater) <- contr.poly(3)
poly_Dy <- aov(Negative ~ Rater, data = Dayaknese_Ph)
summary.lm(poly_Dy)
```

Call:

```
aov(formula = Negative ~ Rater, data = Dayaknese_Ph)
```

Residuals:

Min	1Q	Median	3Q	Max
-1.8948	-0.5463	-0.1098	0.5155	2.1402

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	2.54746	0.11744	21.693 < 0.0000000000000002	***
Rater.L	1.04869	0.20351	5.153	0.00000419 ***
Rater.Q	0.02901	0.20330	0.143	0.887

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.8621 on 51 degrees of freedom

Multiple R-squared: 0.3432, Adjusted R-squared: 0.3174

F-statistic: 13.32 on 2 and 51 DF, p-value: 0.00002211

Results of a polynomial trend analysis indicated a statistically significant linear trend for evaluation of the Dayaknese photos across the three raters $t(51) = 5.153$, $p < .001$.

```
contrasts(Madurese_Ph$Rater) <- contr.poly(3)
poly_Md <- aov(Negative ~ Rater, data = Madurese_Ph)
summary.lm(poly_Md)
```

Call:

```
aov(formula = Negative ~ Rater, data = Madurese_Ph)
```

Residuals:

Min	1Q	Median	3Q	Max
-2.08650	-0.54395	0.01367	0.35905	2.34350

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	3.2973	0.1161	28.391 <0.0000000000000002 ***	
Rater.L	0.1189	0.2012	0.591	0.557
Rater.Q	-0.2054	0.2011	-1.021	0.312
<hr/>				
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1				

Residual standard error: 0.876 on 54 degrees of freedom
 Multiple R-squared: 0.02451, Adjusted R-squared: -0.01162
 F-statistic: 0.6785 on 2 and 54 DF, p-value: 0.5116

Results of a polynomial trend analysis were non-significant when ethnicity of the rater was evaluated when rating Madurese photos.

5.6 Investigating Main Effects

We now focus on the possibility that there might be significant main effects, but a non-significant interaction effect. We only interpret main effects when there is a non-significant interaction effect. Why? Because in the presence of a significant interaction effect, the main effect will not tell a complete story. (*And, if we didn't specify a correct model, we still might have an incomplete story. But that's another issue.*) Here's where we are on the workflow.

Recall that main effects are the *marginal means* – that is the effects of Factor A *collapsed across* all the levels of Factor B.

If the main effect has only two levels (e.g., the ratings of the Dayaknese and Madurese photos):

- the comparison was already ignoring/including all levels of the rater ethnicity factor (Dayaknese, Madurese, Javanese),
- it was only a comparison of two cells (Dayaknese rater, Madurese rater), therefore
- there is no need for further follow-up.

If the main effect has three or more levels (e.g., ethnicity of rater with Dayaknese, Madurese, Javanese levels), then you would follow-up with one or more of the myriad of options. In this class we have focused on three:

- planned contrasts
- posthoc comparisons (all possible cells)
- polynomial

I will demonstrate how to do each as follow-up to a *pretend* scenario where a main effect (but not an interaction) had been significant. I will write up the portion that would be inserted in an APA style results section.

Essentially, we treat these main effect analyses as the follow-up to a significant one-way ANOVA evaluating, in our case, the ethnicity of the Rater.

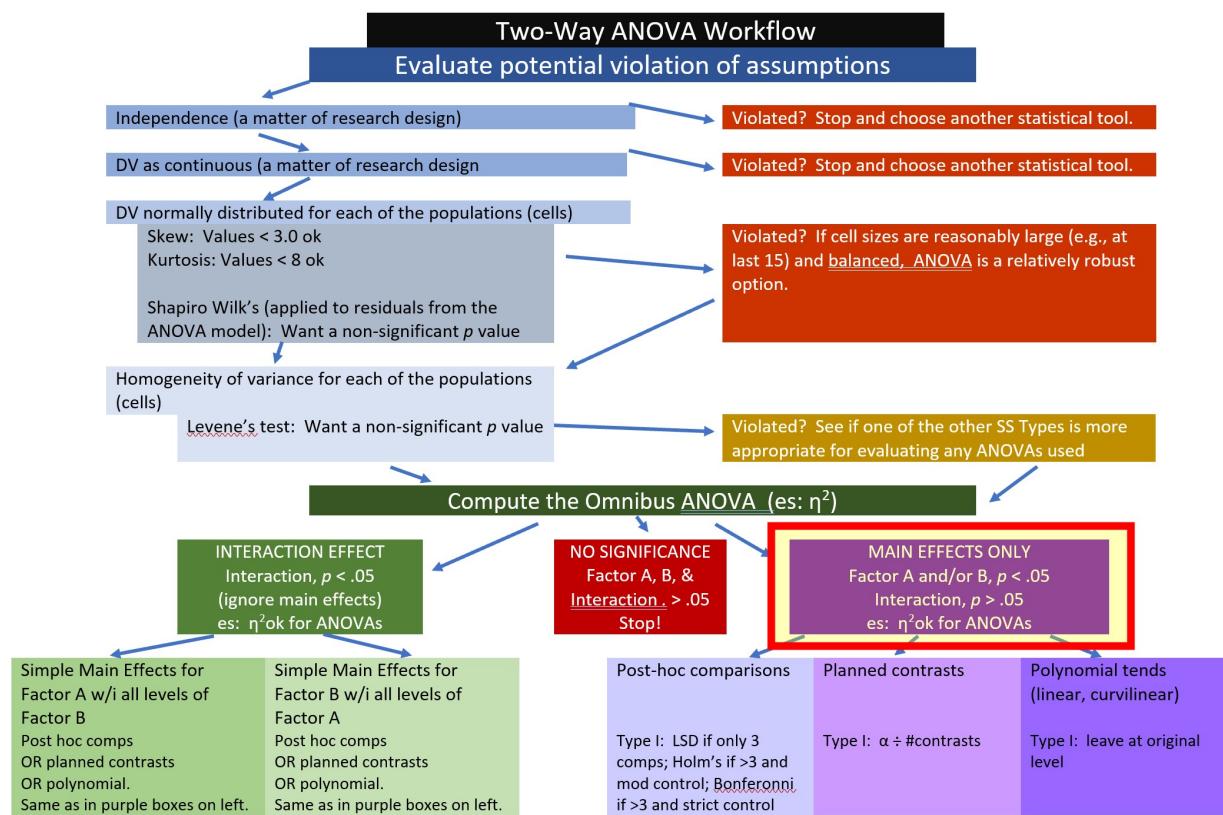


Figure 5.12: Image our place in the Two-Way ANOVA Workflow.

```
RaterMain <- aov(Negative ~ Rater, data = Ramdhani_df) #DV ~ IV I say, 'DV by IV'
model.tables(RaterMain) #ANOVA output
```

Tables of effects

	Dayaknese	Javanese	Madurese
rep	-0.4551	0.05971	0.3518
rep	35.0000	37.00000	39.0000

```
summary(RaterMain)
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)						
Rater	2	12.21	6.103	6.426	0.00231 **						
Residuals	108	102.57	0.950								

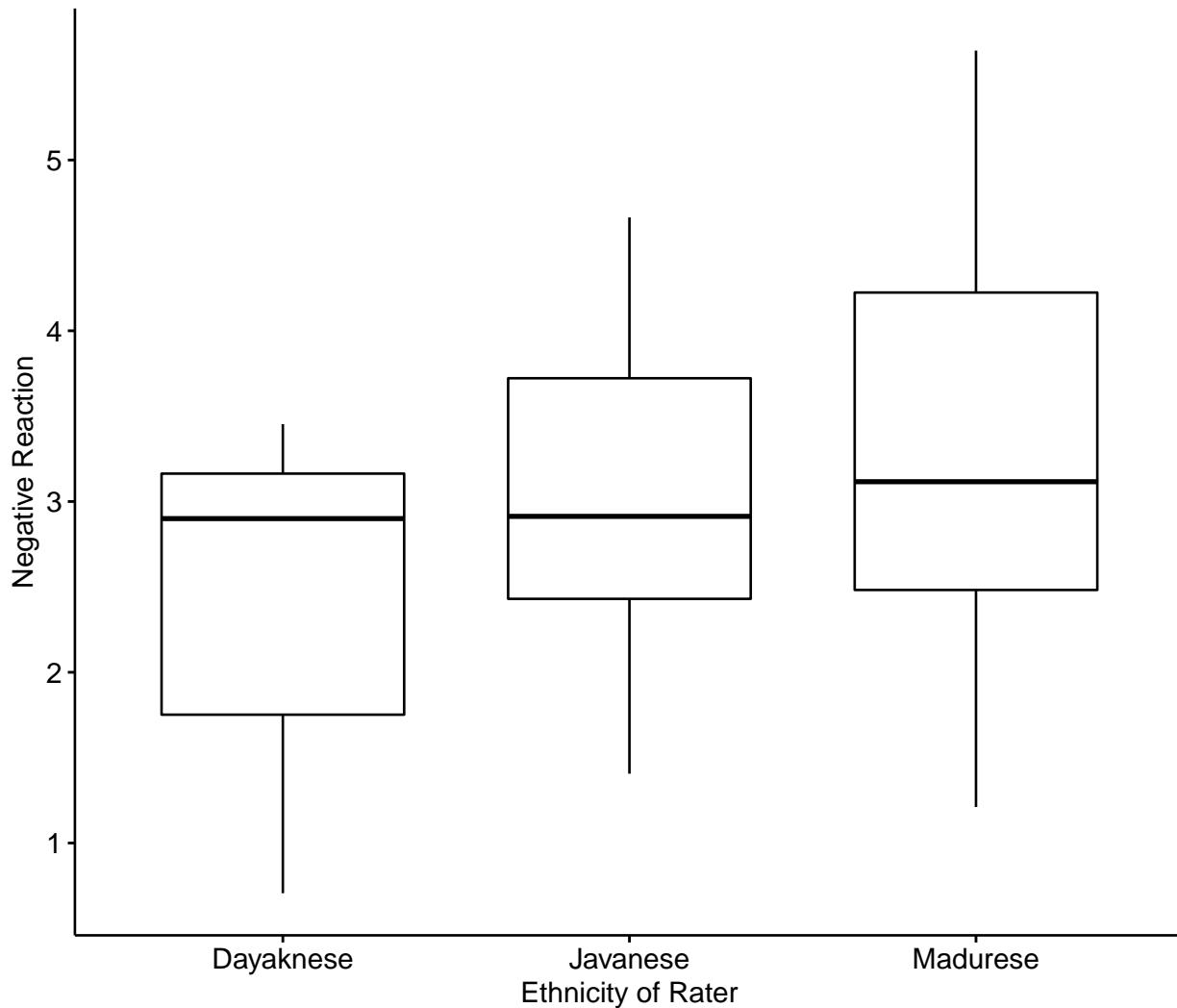
Signif. codes:	0	'***'	0.001	'**'	0.01	'*'	0.05	'. '	0.1	' '	1

```
lsr::etaSquared(RaterMain)
```

	eta.sq	eta.sq.part
Rater	0.1063485	0.1063485

A boxplot representing this main effect may help convey how the main effect of Rater (collapsed across Photo) is different than an interaction effect.

```
ggpubr::ggboxplot(Ramdhani_df, x = "Rater", y = "Negative", xlab = "Ethnicity of Rater",
ylab = "Negative Reaction")
```



5.6.1 Follow-up with all Post-Hocs

An easy possibility is to follow-up with all possible post-hocs. In the main effect case, these are far simpler than where we conducted all possible posthoocs for the interaction effect (remember the Holms sequential Bonferroni?).

Here is a reminder of our location on the workflow.

The *TukeyHSD()* function produces posthoc comparisons by providing the mean difference, a 95% confidence interval of those differences, and the associated p value.

```
TukeyHSD(RaterMain, ordered = TRUE)
```

```
Tukey multiple comparisons of means
 95% family-wise confidence level
 factor levels have been ordered
```

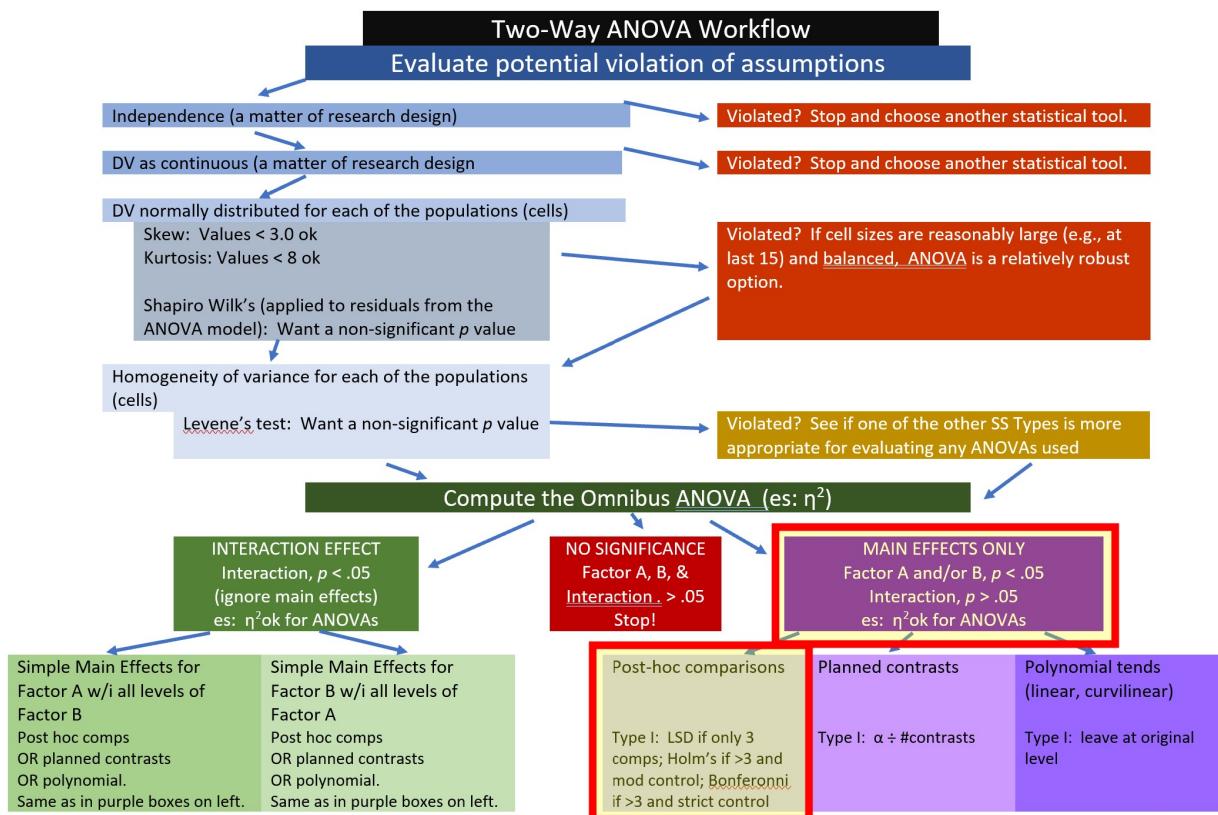


Figure 5.13: Image our place in the Two-Way ANOVA Workflow.

```
Fit: aov(formula = Negative ~ Rater, data = Ramdhani_df)
```

\$Rater

	diff	lwr	upr	p	adj
Javanese-Dayaknese	0.5147954	-0.03128503	1.0608758	0.0690316	
Madurese-Dayaknese	0.8068425	0.26761161	1.3460734	0.0016135	
Madurese-Javanese	0.2920471	-0.23944747	0.8235417	0.3950430	

Results suggest there were statistically significant differences ($p < .05$) between the Madurese and Dayaknese. These differences, though, would have been when rating *all* photos. This analysis disregards the ethnicity portrayed in the photo.

5.6.2 Follow-up with planned contrasts

We generally try for *orthogonal* contrasts so that the partitioning of variance is independent (clean, not overlapping). Planned contrasts are a great way to do this. Here's where we are in the workflow.

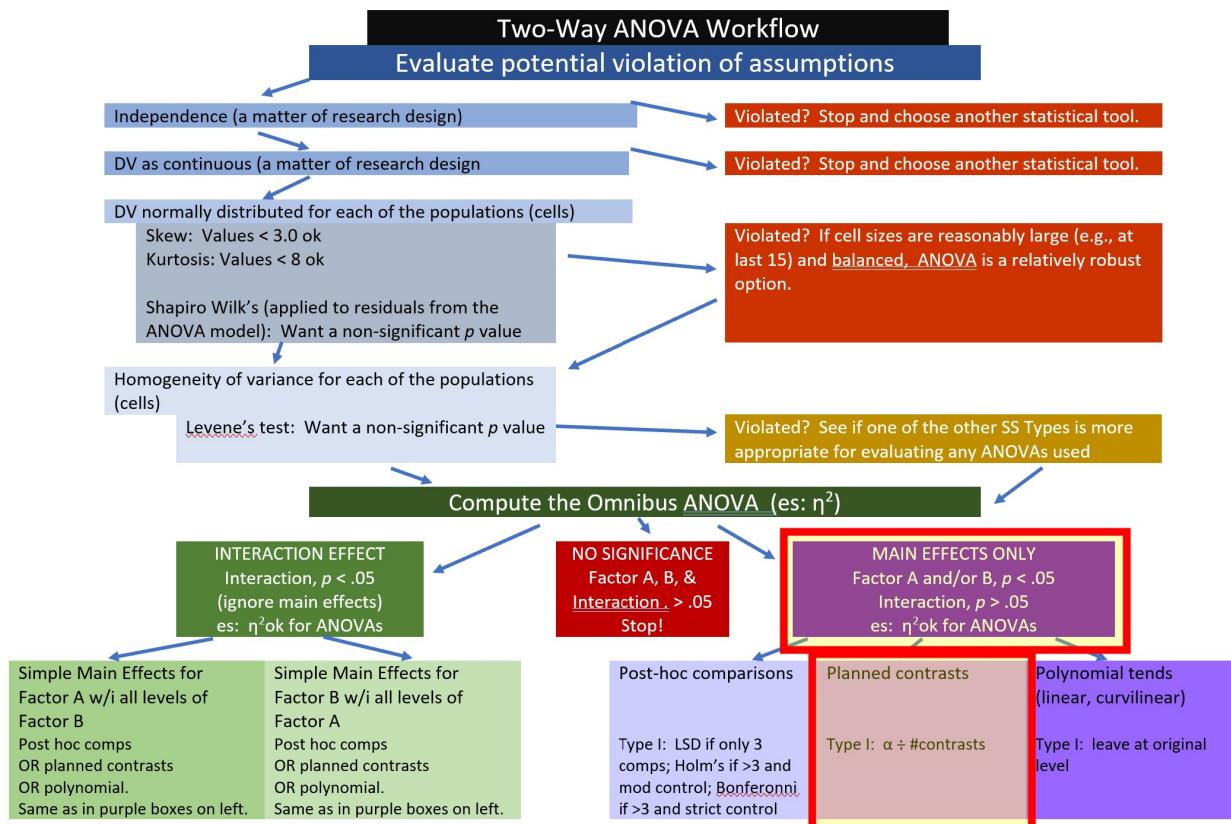


Figure 5.14: Image our place in the Two-Way ANOVA Workflow.

If you aren't extremely careful about your order-of-operations in R, it can confuse objects, so I have named these contrasts *main_c1* and *main_c2* to remind myself that they refer to the main effect of ethnicity of the rater.

In this hypothetical scenario (remember we are pretending we are in the circumstance of a non-significant interaction effect but a significant main effect), I am:

- Contrast #1: comparing the DV for the Javanese rater to the combined Dayaknese and Madurese raters.
- Contrast #2: comparing the DV for the Dayaknese and Madurese raters.

These are orthogonal because:

- there are $k - 1$ comparisons, and
- once a contrast is isolated (i.e., the Javanese rater in contrast #1) it cannot be used again
 - Recall the piece of cake analogy: once you take out a piece of the cake, you really can't put it back in

```
# Contrast1 compares Control against the combined effects of Low and
# High.
main_c1 <- c(1, -2, 1)

# Contrast2 excludes Control; compares Low to High.
main_c2 <- c(-1, 0, 1)
contrasts(Ramdhani_df$Rater) <- cbind(main_c1, main_c2)
contrasts(Ramdhani_df$Rater)
```

	main_c1	main_c2
Dayaknese	1	-1
Javanese	-2	0
Madurese	1	1

Then we run the contrast

```
mainPlanned <- aov(Negative ~ Rater, data = Ramdhani_df)
summary.lm(mainPlanned)
```

Call:

```
aov(formula = Negative ~ Rater, data = Ramdhani_df)
```

Residuals:

Min	1Q	Median	3Q	Max
-2.08813	-0.74921	0.05792	0.71482	2.34187

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	2.93283	0.09259	31.676 < 0.0000000000000002 ***	
Ratermain_c1	-0.03712	0.06544	-0.567	0.571670
Ratermain_c2	0.40342	0.11345	3.556	0.000561 ***

```
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

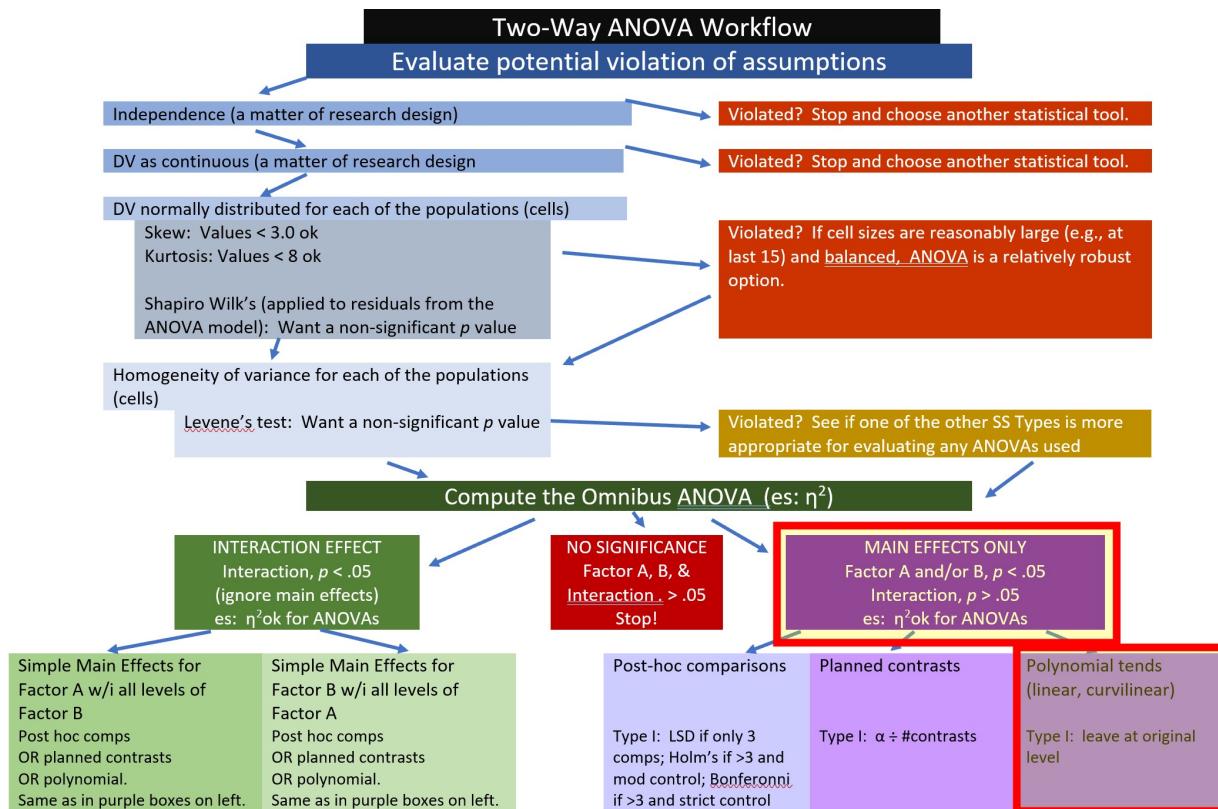
```
Residual standard error: 0.9745 on 108 degrees of freedom
Multiple R-squared: 0.1063, Adjusted R-squared: 0.0898
F-statistic: 6.426 on 2 and 108 DF, p-value: 0.002307
```

```
contrasts(Ramdhani_df$Rater) <- cbind(c(1, -2, 1), c(-1, 0, 1))
```

These planned contrasts show that when the Javanese raters are compared to the combined Dayaknese and Madurese raters, there was a non significant difference, $t(108) = -0.567$, $p = .572$. However, there were significant differences between Dayaknese and Javanese raters, $t(108) = 3.556$, $p < .001$.

5.6.3 Polynomial Trends

Polynomial contrasts let us see if there is a linear (or curvilinear) pattern to the data. To detect a trend, the data must be coded in an ascending order...and it needs to be a sensible comparison. Here's where this would fall in our workflow.



Because these three ethnic groups are not *ordered* in the same way as would an experiment involving dosage (e.g., placebo, lo dose, hi dose), evaluation of the polynomial trend is not really justified (even though it is statistically possible). None-the-less, I will demonstrate how it is conducted.

```
contrasts(Ramdhani_df$Rater) <- contr.poly(3)
mainTrend <- aov(Negative ~ Rater, data = Ramdhani_df)
summary.lm(mainTrend)
```

Call:

```
aov(formula = Negative ~ Rater, data = Ramdhani_df)
```

Residuals:

Min	1Q	Median	3Q	Max
-2.08813	-0.74921	0.05792	0.71482	2.34187

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	2.93283	0.09259	31.676	< 0.0000000000000002 ***
Rater.L	0.57052	0.16045	3.556	0.000561 ***
Rater.Q	-0.09094	0.16029	-0.567	0.571670

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

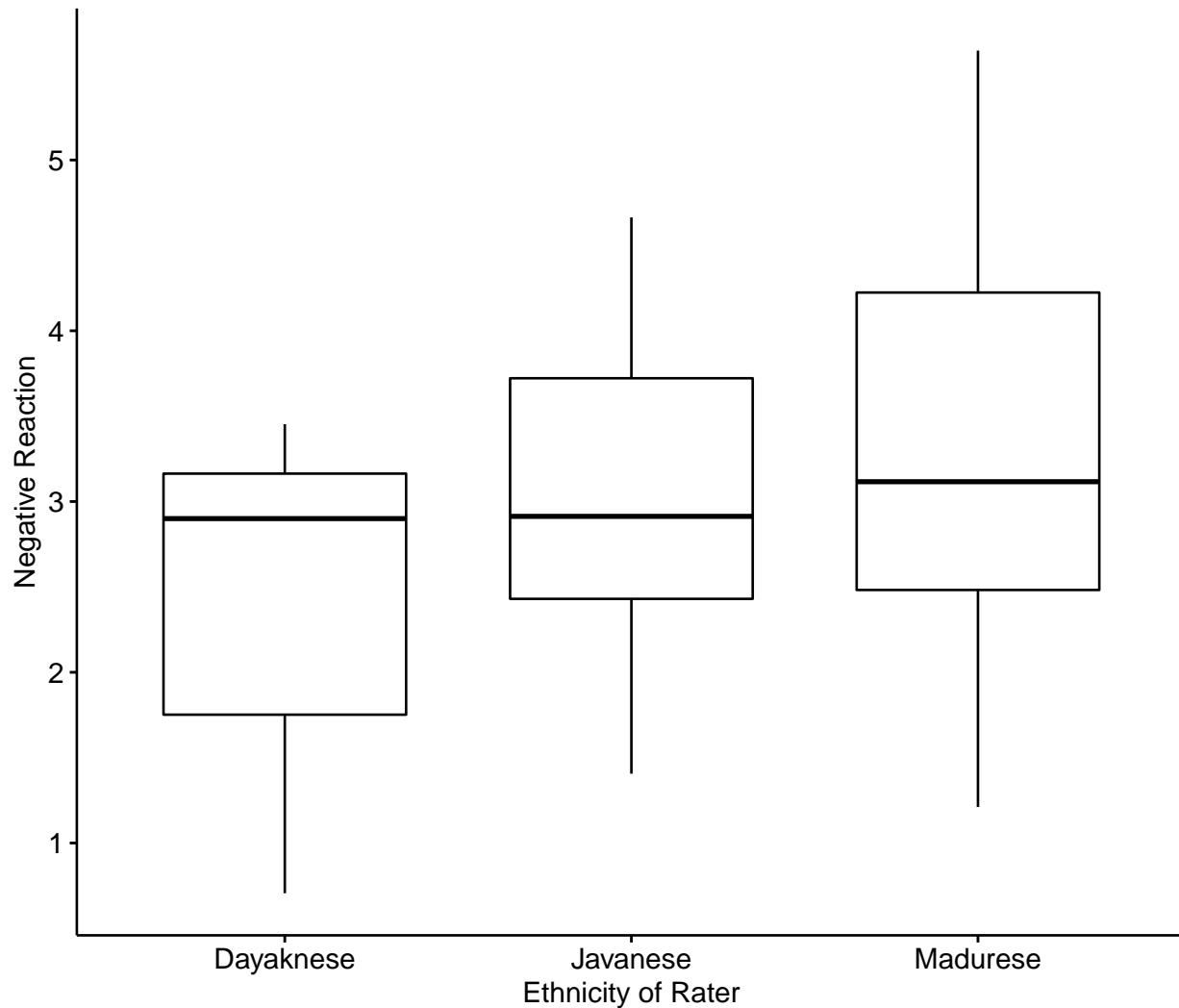
Residual standard error: 0.9745 on 108 degrees of freedom
Multiple R-squared: 0.1063, Adjusted R-squared: 0.0898
F-statistic: 6.426 on 2 and 108 DF, p-value: 0.002307

Rater.L tests the data to see if there is a significant linear trend. There is: $t = 3.556$, $p < .001$.

Rater.Q tests to see if there is a significant quadratic (curvilinear, one hump) trend. There is not: $t = -0.567$, $p = .572$.

Results supported a significant linear trend ($t = 3.556$, $p < .001$) such that negative reactions increased in a linear reaction across the three rating groups.

```
ggpubr::ggboxplot(Ramdhani_df, x = "Rater", y = "Negative", xlab = "Ethnicity of Rater",
ylab = "Negative Reaction")
```



5.7 My APA Style Results Section

First, I am reluctant to term anything “final.” It seems like there is always the possibility or revision. Given that I demonstrated a number of options, let me first show the workflow with the particular path I took:

In light of that, here’s the final write-up:

Results

A 3 X 2 ANOVA was conducted to evaluate the effects of rater ethnicity (3 levels, Dayaknese, Madurese, Javanese) and photo stimulus (2 levels, Dayaknese on Madurese,) on negative reactions to the photo stimuli. Results of Levene’s test for equality of error variances indicated violation of the assumption, ($F[5, 105] = 8.834, p < .001$). Our analysis of the individual cell means (see Table 1 for means and standard deviations) suggested skew and kurtosis were within the bounds considered to be normally distributed [Kline, 2016]. A non-significant Shapiro-Wilk normality test (applied to the residuals from the factorial ANOVA model) provided further evidence that the assumption of normality was not violated ($W = 0.9846, p = 0.234$).

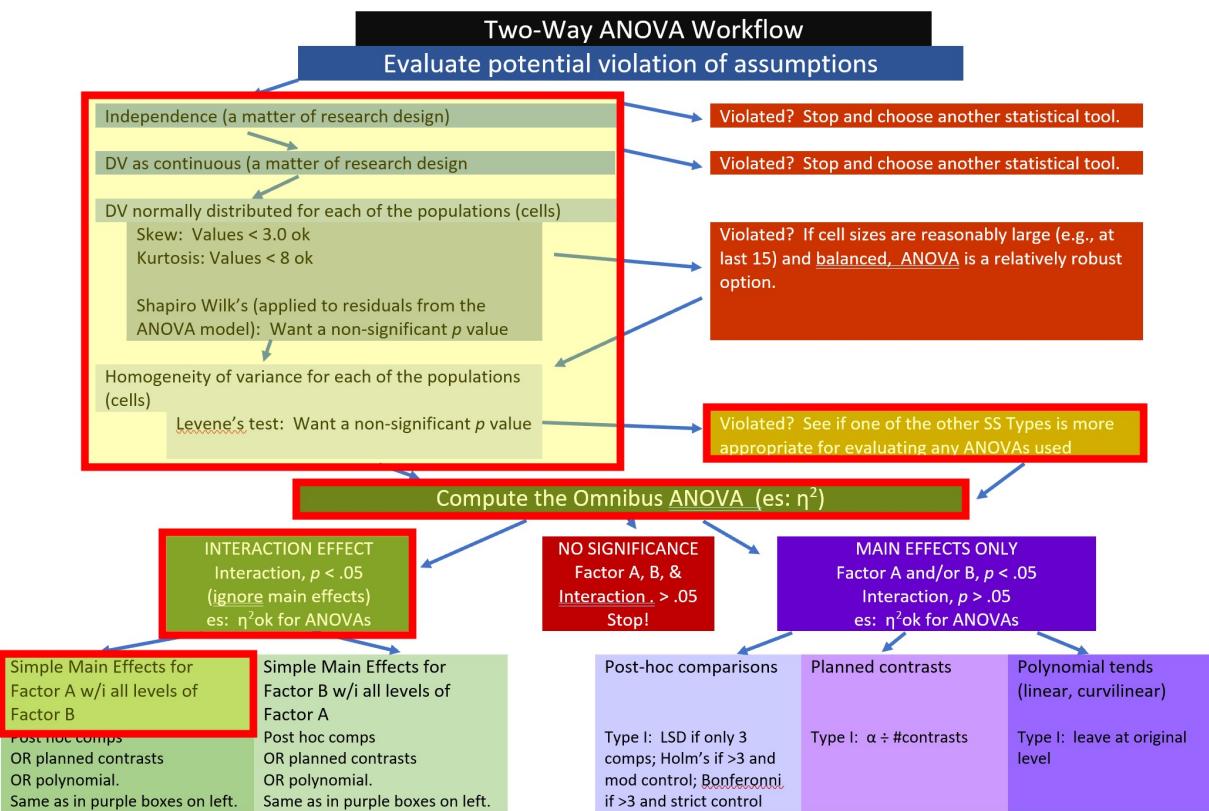
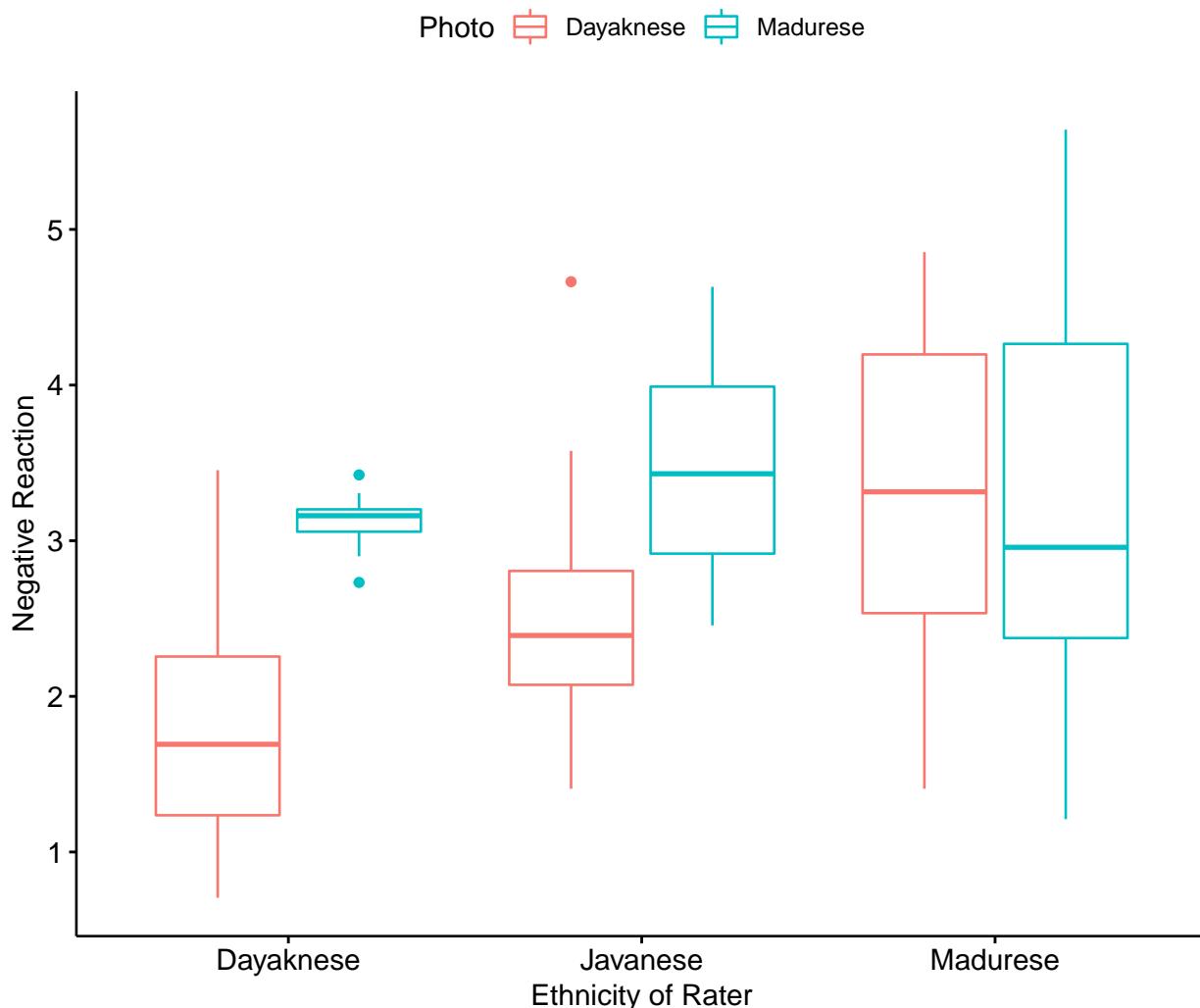


Figure 5.15: Image our place in the Two-Way ANOVA Workflow.

Computing sums of squares with a Type II approach, the results for the ANOVA indicated a significant main effect for ethnicity of the rater ($F[2, 105] = 8.077, p < .001, \eta^2 = 0.107$), a significant main effect for photo stimulus, ($F[1, 105] = 19.346, p < .001, \eta^2 = 0.127$), and a significant interaction effect ($F[2, 105] = 5.696, p = .004, \eta^2 = 0.075$).

To explore the interaction effect, we followed with a test of the simple main effect of photo stimulus within the ethnicity of the rater. That is, we looked at the effect of the photo stimulus within the Dayaknese, Madurese, and Javanese groups, separately. To control for Type I error across the three simple main effects, we set alpha at .017 (.05/3). Results indicated significant differences for Dayaknese ($F [1, 33] = 50.4, p < .001, \eta^2 = 0.60.$) and Javanese ethnic groups ($F [1, 35]= 17.18, p < .001, \eta^2 = 0.33$), but not for the Madurese ethnic group ($F [1, 37] = 0.000, p = .993, \eta^2 < .001$). As illustrated in Figure 1, the Dayaknese and Javanese rather both reported stronger negative reactions to the Madurese. The differences in ratings for the Madurese were not statistically significantly different. In this way, the rater's ethnic group moderated the relationship between the photo stimulus and negative reactions.

```
ggpubr::ggboxplot(Ramdhani_df, x = "Rater", y = "Negative", color = "Photo",
  xlab = "Ethnicity of Rater", ylab = "Negative Reaction")
```



```
apaTables::apa.2way.table(iv1 = Rater, iv2 = Photo, dv = Negative, data = Ramdhani_df,
  landscape = TRUE, table.number = 1, filename = "Table_1_MeansSDs.doc")
```

Table 1

Means and standard deviations for Negative as a function of a 3(Rater) X 2(Photo) design

	Photo			
	Dayaknese		Madurese	
Rater	M	SD	M	SD
Dayaknese	1.82	0.77	3.13	0.16
Javanese	2.52	0.74	3.46	0.64
Madurese	3.30	1.03	3.30	1.33

Note. M and SD represent mean and standard deviation, respectively.

```
apaTables::apa.aov.table(TwoWay_neg, filename = "Table_2_effects.doc",
  table.number = 2, type = "II")
```

Table 2

ANOVA results using Negative as the dependent variable

Predictor	SS	df	MS	F	p	partial_eta2	CI_90_partial_eta2
Rater	12.24	2	6.12	8.10	.001	.13	
Photo	14.62	1	14.62	19.35	.000	.16	[.06, .26]
Rater x Photo	8.61	2	4.30	5.70	.004	.10	[.02, .18]
Error	79.34	105	0.76				

Note: Values in square brackets indicate the bounds of the 90% confidence interval for partial

5.7.1 Comparing Our Results to Rhamdani et al. [2018]

As is common in simulations, our results approximate the findings reported in the manuscript, but does not replicate them exactly. Our main and interaction effects map on very closely. However, in the follow-up tests, while our findings that Dayaknese rated the Madurese photos more negatively, the findings related to the Javanese' and Madurese' ratings wiggled around some. A close look at the figures can explain that with varying variability and means with similar values, this is probable. I find it to be a useful lesson in “what it takes” to get stable, meaningful results.

5.8 Options for Assumption Violations

In one-way ANOVA we could simply apply the Welch's alternative. It's not so easy in factorial ANOVA. One alternative, though, is to change the sums of squares type used in the ANOVA calculations.

In ANOVA models sums of squares can be calculated four different ways: Type I, II, III, and IV. This matters.

SS Type II is the *aov()* default. It may be a best practice to go ahead and specify the SS Type in both the *aov()*, eta-squared, and apaTables script so that they are consistent.

Type I sums of squares is similar to hierarchical linear regression in that the first predictor in the model claims as much variance as it can and the leftovers are claimed by the variable entered next – each claiming as much as possible leaving the leftovers for what follows. Unless the variables are completely independent of each other (unlikely), Type I sums of squares cannot evaluate the true main effect of each variable. Type I should not be used to evaluate main effects and interactions because the order of predictors will affect the results.

Type II (the R default) is appropriate if you are interested main effects because it ignores the effect of any interactions involving the main effect. Thus, variance from a main effect is not “lost” to any interaction terms containing that effect. Type II is appropriate for main effects analyses only, but should not be used when evaluating interaction effects. Type II sums of squares is not affected by the type of contrast coding used to specify the predictor variables.

Type III is the default in many stats packages – but not R. In Type III all effects (main effects and interactions) are evaluated (simultaneously) taking into consideration all other effects in the model (not just the ones entered before). Type III is more robust to unequal samples sizes (e.g., unbalanced designs). Type III is best when predictors are encoded with orthogonal contrasts.

***Type IV** is identical to Type III except it requires no missing cells.

Field [2012] suggested that it is safest to stick with Type III sums of squares. We apply the type to the model we create in the initial run. For more information, check out this explanation on [r-bloggers](#).

Many researchers automatically use Type III as the SS type. Today I went with the R default because

- Type II sums of squares was used in hand-calculations,
- Our example was reasonably balanced (equal cell sizes), and
- We had only violated the homogeneity of variance assumption.

For demonstration purposes, let's run the Type III alternative to see the differences:

```
# this is what we did
car::Anova(TwoWay_neg)
```

Anova Table (Type II tests)

Response: Negative

```

      Sum Sq Df F value    Pr(>F)
Rater      12.238  2  8.0977  0.0005363 ***
Photo       14.619  1 19.3462  0.00002623 ***
Rater:Photo  8.609  2  5.6964  0.0044803 **
Residuals   79.341 105
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

```
# We change the SS type by applying it to our model.
car::Anova(TwoWay_neg, type = "III")
```

Anova Table (Type III tests)

```

Response: Negative
      Sum Sq Df F value    Pr(>F)
(Intercept) 56.173  1 74.3388 0.00000000000007422 ***
Rater       19.805  2 13.1051 0.00000830116650983 ***
Photo        15.040  1 19.9034 0.00002051829053731 ***
Rater:Photo  8.609  2  5.6964           0.00448 **
Residuals   79.341 105
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

Note that the sums of squares are somewhat different between models – and that the Type III tests includes an intercept. In today's example, the statistical significance remains the same across the models.

Now let's compare the effect sizes across models.

```
lsr::etaSquared(TwoWay_neg)
```

	eta.sq	eta.sq.part
Rater	0.10662441	0.13363091
Photo	0.12736755	0.15558329
Rater:Photo	0.07500609	0.09788289

```
lsr::etaSquared(TwoWay_neg, type = 3)
```

	eta.sq	eta.sq.part
Rater	0.17255745	0.19975734
Photo	0.13103575	0.15935009
Rater:Photo	0.07500609	0.09788289

The Type III effect size for Rater is higher; the others are quite similar.

5.9 Power

Asking about *power* can be a euphemistic way of asking, “How large should my sample size be?”

Power is defined as the ability of the test to detect statistical significance when there is such. It’s represented formulaically as $(1 - P)$ (Type II error). Power is traditionally set at 80% (or .8)

We will do both – evaluate the power of our current example and then work backwards to estimate the sample size needed (which is our usual question for MRPs and dissertations).

We’ll use the *pwr.2way()* function from the *pwr2* package. Helpful resources are found here:

- <https://cran.r-project.org/web/packages/pwr2/pwr2.pdf>
- <https://rdrr.io/cran/pwr2/man/ss.2way.html>

The *pwr.2way()* and *ss.2way()* functions require the following:

- **a** number of groups in Factor A
- **b** number of groups in Factor B
- **alpha** significant level (Type I error probability)
- **beta** Type II error probability (Power = $1 - \beta$; traditionally set at .1 or .2)
- **f.A** the *f* effect size of Factor A
- **f.B** the *f* effect size of Factor B
- **B** Iteration times, default is 100

Hints for calculating the *f.A* and *f.B* values:

- In this case, we will rerun the statistic, grab both effect sizes, and convert them to the *f* (not the f^2)
 - calculation can be straightforward, either use an online calculator, a hand-calculated formula, or the *eta2_to_f* function from the *effectsize*
- When an effect size is unknown, you can substitute what you expect using Cohen’s guidelines of .10, .25, and .40 as small, medium, and large (for the *f*, not F^2)

Let’s quickly rerun our model to get both the df and calculate the *f* effect value

```
lsr::etaSquared(TwoWay_neg, anova = TRUE)
```

	eta.sq	eta.sq.part	SS	df	MS	F
Rater	0.10662441	0.13363091	12.237770	2	6.1188848	8.097730
Photo	0.12736755	0.15558329	14.618555	1	14.6185546	19.346190
Rater:Photo	0.07500609	0.09788289	8.608791	2	4.3043955	5.696435
Residuals	0.69127788		NA	79.341113	105	0.7556296
		p				NA
Rater	0.00053629628					
Photo	0.00002622821					
Rater:Photo	0.00448026007					
Residuals	NA					

```
# get the partial eta-square (second number) and dfs
```

If we want to understand power in our analysis, we need to convert our effect size for the *interaction* to *f* effect size (this is not the same as the *F* test). The *effectsize* package has a series of converters. We can use the *eta2_to_f()* function.

```
effectsize::eta2_to_f(0.1066) #FactorA -- Rater
```

```
[1] 0.3454265
```

```
effectsize::eta2_to_f(0.1274) #Factor B -- Photo
```

```
[1] 0.3821001
```

5.9.1 Post Hoc Power Analysis

Now we calculate power for our existing model. We'll use the package *pwr2* and the function *pwr.2way()*. To specify this we identify:

- a: number of groups for Factor A (Rater)
- b: number of groups for Factor B (Photo)
- size.A: sample size per group in Factor A (because ours differ slightly, I divided the N by the number of groups)
- size.B: sample size per group in Factor B (because ours differ slightly, I divided the N by the number of groups)
- f.A: Effect size of Factor A
- f.A.: Effect size of Factor B

```
pwr2::pwr.2way(a = 3, b = 2, alpha = 0.05, size.A = 37, size.B = 55, f.A = 0.345,
f.B = 0.382)
```

```
Balanced two-way analysis of variance power calculation
```

```
a = 3
b = 2
n.A = 37
n.B = 55
sig.level = 0.05
power.A = 0.9974259
power.B = 0.9999996
power = 0.9974259
```

NOTE: power is the minimum power among two factors

At 0.997 (Rater), 0.999 (Photo), and 0.997 (interaction), our power to detect a significant effect for Factor A/Rater and Factor B/Photo was huge!

5.9.2 Estimating Sample Size Requirements

If we want to replicate this study we could use its results to estimate what would be needed for the replication.

In this specification:

- a: number of groups for Factor A (Rater)
- b: number of groups for Factor B (Photo)
- alpha: significance level (Type I error probability); usually .05
- beta: Type II error probability (Power = 1-beta); usually .80
- f.A: Effect size (f) of Factor A (this time we know; other times we can guess from previously published literature)
- f.B: Effect size (f) of Factor B
- B: iteration times, default number is 100

```
pwr2::ss.2way(a = 3, b = 2, alpha = 0.05, beta = 0.8, f.A = 0.345, f.B = 0.382,
B = 100)
```

Balanced two-way analysis of variance sample size adjustment

```
a = 3
b = 2
sig.level = 0.05
power = 0.2
n = 3
```

NOTE: n is number in each group, total sample = 18

Curiously, that's just about the number that was in each of the six cells!

Often times researchers will play around with the f values. Remember Cohen's indication of small (.10), medium (.25), and large (.40). Let's see what happens when we enter different values. Specifically, what if we only had a medium effect?

```
pwr2::ss.2way(a = 3, b = 2, alpha = 0.05, beta = 0.8, f.A = 0.25, f.B = 0.25,
B = 100) #if we expected a medium effect
```

Balanced two-way analysis of variance sample size adjustment

```
a = 3
b = 2
sig.level = 0.05
power = 0.2
n = 6
```

NOTE: n is number in each group, total sample = 36

And what would happen if we only had a small effect?

```
pwr2::ss.2way(a = 3, b = 2, alpha = 0.05, beta = 0.8, f.A = 0.1, f.B = 0.1,
  B = 100) #if we expected a small effect
```

Balanced two-way analysis of variance sample size adjustment

```
a = 3
b = 2
sig.level = 0.05
power = 0.2
n = 30
```

NOTE: n is number in each group, total sample = 180

5.10 Practice Problems

In each of these lessons I provide suggestions for practice that allow you to select one or more problems that offer differing levels of difficulty. Whichever you choose, you will focus on these larger steps in factorial-way ANOVA, including:

- test the statistical assumptions
- conduct a two-way ANOVA, including
 - omnibus test and effect size
 - report main and interaction effects
 - conduct follow-up testing of simple main effects
- write a results section to include a figure and tables

5.10.1 Problem #1: Play around with this simulation.

Copy the script for the simulation and then change (at least) one thing in the simulation to see how it impacts the results.

- If two-way ANOVA is new to you, perhaps you just change the number in “set.seed(210731)” from 210731 to something else. Your results should parallel those obtained in the lecture, making it easier for you to check your work as you go.
- If you are interested in power, change the sample size to something larger or smaller.
- If you are interested in variability (i.e., the homogeneity of variance assumption), perhaps you change the standard deviations in a way that violates the assumption.

5.10.2 Problem #2: Conduct a factorial ANOVA with the *positive evaluation* dependent variable.

The Ramdhani et al. [2018] article has two dependent variables (negative and positive evaluation). Each is suitable for two-way ANOVA. I used *negative evaluation* as the dependent variable; you are welcome to conduct the analysis with *positive evaluation* as the dependent variable.

5.10.3 Problem #3: Try something entirely new.

Using data for which you have permission and access (e.g., IRB approved data you have collected or from your lab; data you simulate from a published article; data from an open science repository; data from other chapters in this OER), complete a two-way, factorial ANOVA. Please have at least 3 levels for one predictor and at least 2 levels for the second predictor.

5.10.4 Grading Rubric

Regardless which option(s) you chose, use the elements in the grading rubric to guide you through the practice.

Assignment Component	Points Possible	Points Earned
1. Narrate the research vignette, describing the IV and DV	5	_____
2. Simulate (or import) and format data	5	_____
3. Evaluate statistical assumptions	5	_____
4. Conduct omnibus ANOVA (w effect size)	5	_____
5. Conduct one set of follow-up tests; narrate your choice	5	_____
6. Describe approach for managing Type I error	5	_____
7. APA style results with table(s) and figure	5	_____
8 Explanation to grader	5	_____
Totals	40	_____

Chapter 6

One-Way Repeated Measures ANOVA

[Screencasted Lecture Link](#)

In the prior lessons, a critical assumption is that the observations must be “independent.” That is, related people (partners, parent/child, manager/employee) cannot comprise the data and there cannot be multiple waves of data for the same person. Repeated measures ANOVA is created specifically for this *dependent* purpose. This lesson focuses on the one-way repeated measures ANOVA, where we measure changes across time.

6.1 Navigating this Lesson

There is just over one hour of lecture. If you work through the materials with me plan for an additional two hours

While the majority of R objects and data you will need are created within the R script that sources the chapter, occasionally there are some that cannot be created from within the R framework. Additionally, sometimes links fail. All original materials are provided at the [Github site](#) that hosts the book. More detailed guidelines for ways to access all these materials are provided in the OER’s [introduction](#)

6.1.1 Learning Objectives

Learning objectives from this lecture include the following:

- Evaluate the suitability of a research design/question and dataset for conducting a one-way repeated measures ANOVA; identify alternatives if the data is not suitable.
- Hand-calculate a one-way repeated measures ANOVAs
 - describing the partitioning of variance as it relates to model/residual; within/between.
- Test the assumptions for one-way repeated measures ANOVA.
- Conduct a one-way repeated measures ANOVA (omnibus and follow-up) in R.

- Interpret output from the one-way repeated measures ANOVA (and follow-up).
- Prepare an APA style results section of the one-way repeated measures ANOVA output.
- Demonstrate how an increased sample size increases the power of a statistical test.

6.1.2 Planning for Practice

The suggestions for homework are graded in complexity with more complete descriptions at the end of the chapter follow these suggestions.

- Rework the problem in the chapter by changing the random seed in the code that simulates the data. This should provide minor changes to the data, but the results will likely be very similar.
- There were no additional variables in this example. However, you'll see we do have an issue with statistical power. Perhaps change the sample size to see if it changes (maybe stabilizes?) the results.
- Conduct a one-way repeated measures ANOVA with data to which you have access. This could include data you simulate on your own or from a published article.

6.1.3 Readings & Resources

In preparing this chapter, I drew heavily from the following resource(s). Other resources are cited (when possible, linked) in the text with complete citations in the reference list.

- *Repeated Measures ANOVA in R: The Ultimate Guide.* (n.d.). Datanovia. Retrieved October 19, 2020, from <https://www.datanovia.com/en/lessons/repeated-measures-anova-in-r>
 - This website is an excellent guide for both one-way repeated measures and mixed design ANOVA. A great resource for both the conceptual and procedural. This is the guide I have used for the basis of the lecture. Working through their example would be great additional practice.
- Green, S. B., & Salkind, N. J. (2014). One-Way Repeated Measures Analysis of Variance (Lesson 29). In *Using SPSS for Windows and Macintosh: Analyzing and understanding data* (Seventh edition., pp. 209–217). Pearson.
 - For years I taught from the Green and Salkind text. Even though it was written for SPSS, the authors do a terrific job of walking the reader through the one-way repeated measures logic and process.
- Amodeo, A. L., Picariello, S., Valerio, P., & Scandurra, C. (2018). Empowering transgender youths: Promoting resilience through a group training program. *Journal of Gay & Lesbian Mental Health, 22*(1), 3–19.
 - This mixed methods (qualitative and quantitative) includes a one-way repeated measures example.

6.1.4 Packages

The packages used in this lesson are embedded in this code. When the hashtags are removed, the script below will (a) check to see if the following packages are installed on your computer and, if not (b) install them.

```
# will install the package if not already installed
# if(!require(knitr)){install.packages('knitr')}
# if(!require(tidyverse)){install.packages('tidyverse')} #manipulate
# data if(!require(psych)){install.packages('psych')}
# if(!require(ggpubr)){install.packages('ggpubr')} #easy plots
# if(!require(rstatix)){install.packages('rstatix')} #pipe-friendly R
# functions if(!require(data.table)){install.packages('data.table')}
# #pipe-friendly R functions
# if(!require(reshape2)){install.packages('reshape2')} #pipe-friendly
# R functions
# if(!require(effectsize)){install.packages('effectsize')} #converts
# effect sizes for use in power analysis
# if(!require(WebPower)){install.packages('WebPower')} #power
# analysis tools for repeated measures
# if(!require(MASS)){install.packages('MASS')} #power analysis tools
# for repeated measures
```

6.2 Introducing One-way Repeated Measures ANOVA

There are a couple of typical use cases for one-way repeated measures ANOVA. In the first, the research participant is assessed in multiple conditions – with no interested in change-over-time.

An example of a research design using this approach occurred in the Green and Salkind [2014b] statistics text, the one-way repeated measures vignette compared teachers' perception of stress when responding to parents, teachers, and school administrators.



Figure 6.1: Illustration of a research design appropriate for one-way repeated measures ANOVA

Another common use case is about time. The classic design is a pre-test, an intervention, a post-test, and a follow up. In designs like these researchers often hope that there is a positive change from pre-to-post and that that change either stays constant (from post-to-follow-up) or, perhaps, increases even further. The research vignette for this lesson is interested in change-over-time.

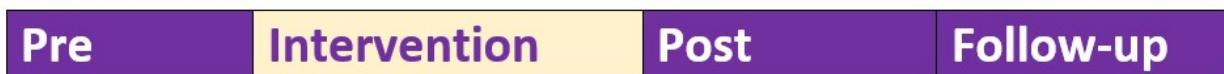


Figure 6.2: Illustration of a research design appropriate for one-way repeated measures ANOVA

6.2.1 Workflow for Oneway Repeated Measures ANOVA

The following is a proposed workflow for conducting a one-way repeated measures ANOVA.

Steps involved in analyzing the data include:

1. Preparing and importing the data.
2. Exploring the data
 - graphs
 - descriptive statistics
3. Checking distributional assumptions
 - assessing normality via skew, kurtosis, Shapiro Wilks
 - checking or violation of the *sphericity* assumption with Mauchly's test; if violated interpret the corrected output or use a multivariate approach for the analysis
4. Computing the omnibus ANOVA
5. Computing post-hoc comparisons, planned contrasts, or polynomial trends
6. Managing Type I error
7. Sample size/power analysis (which you should think about first – but in the context of teaching ANOVA, it's more pedagogically sensible, here)

6.3 Research Vignette

Amodeo [Amodeo et al., 2018] and colleagues conducted a mixed methods study (qualitative and quantitative) to evaluate the effectiveness of an empowerment, peer-group-based, intervention with participants ($N = 8$) who experienced transphobic episodes. Focus groups used qualitative methods to summarize emergent themes from the program (identity affirmation, self-acceptance, group as support) and a one-way, repeated measures ANOVA provided evidence of increased resilience from pre to three-month followup.

Eight participants (seven transgender women and one genderqueer person) participated in the intervention. The mean age was 28.5 ($SD = 5.85$). All participants were located in Italy.

The within-subjects condition was wave, represented by T1, T2, and T3:

- T1, beginning of training
- Training, three 8-hour days,
 - content included identity and heterosexism, sociopolitical issues and minority stress, resilience and empowerment
- T2, at the conclusion of the 3-day training
- Follow-up session 3 months later
- T3, at the conclusion of the +3 month follow-up session

The dependent variable (assessed at each wave) was a 14-item resilience scale [Wagnild and Young, 1993]. Items were assessed on a 7-point scale ranging from *strongly disagree* to *strongly agree* with higher scores indicating higher levels of resilience. An example items was, “I usually manage one way or another.”

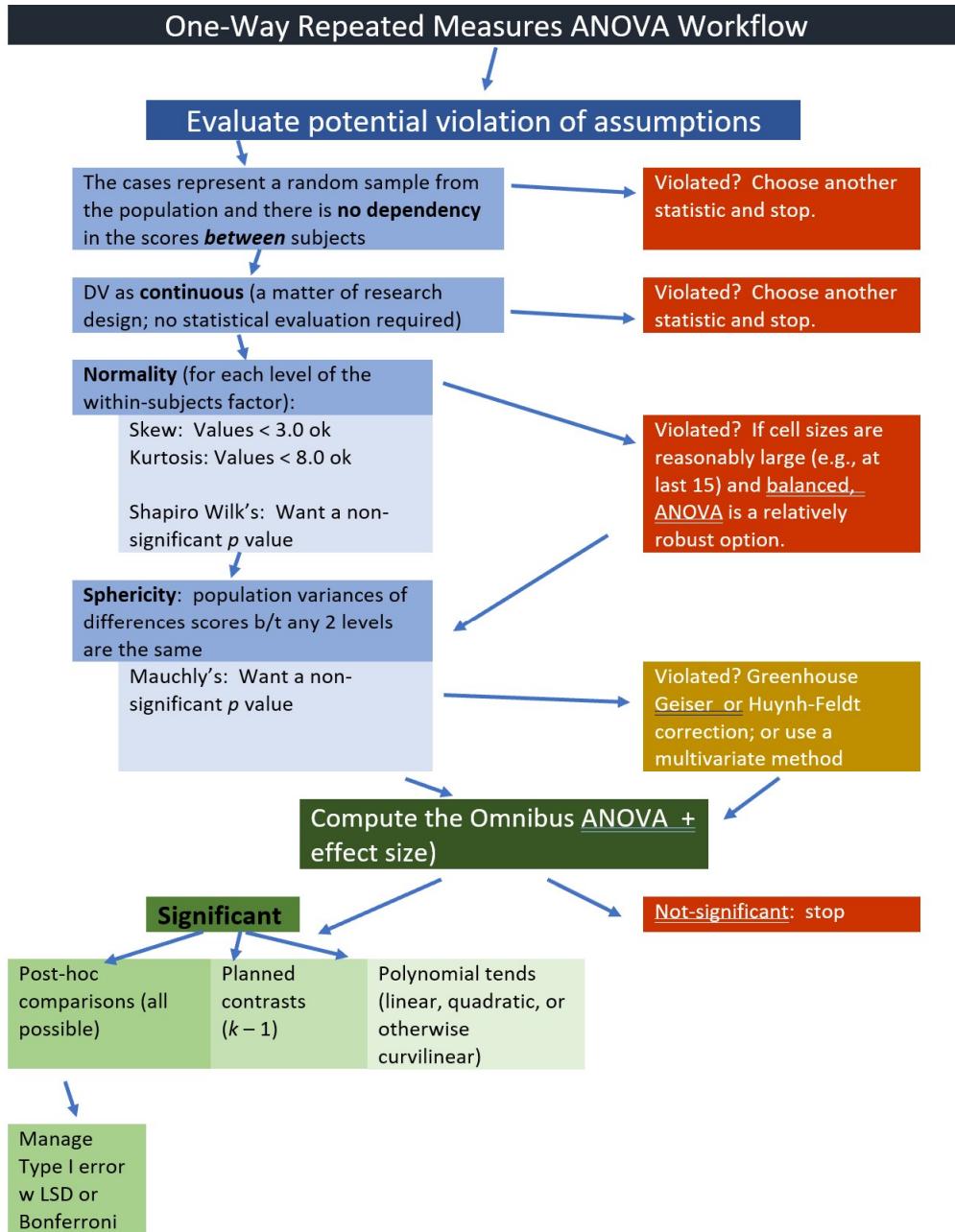


Figure 6.3: Image of a workflow for the one-way repeated measures ANOVA



Figure 6.4: Diagram of the research design for the Amodeo et al study

6.3.1 Code for simulating the data used today.

Below is the code I used to simulate data. The following code assumes 8 participants who each participated in 3 waves (pre, post, followup).

```
set.seed(2022)
# gives me 8 numbers, assigning each number 3 consecutive spots, in
# sequence
ID <- factor(rep(seq(1, 8), each = 3))
# gives me a column of 24 numbers with the specified Ms and SD
Resilience <- rnorm(24, mean = c(5.7, 6.21, 6.26), sd = c(0.88, 0.79, 0.37))
# repeats pre, post, follow-up once each, 8 times
Wave <- rep(c("Pre", "Post", "FollowUp"), each = 1, 8)
Amodeo_long <- data.frame(ID, Wave, Resilience)
```

Let's take a look at the structure of our variables. We want ID to be a factor, Resilience to be numeric, and Wave to be an ordered factor (Pre, Post, FollowUp).

```
str(Amodeo_long)
```

```
'data.frame': 24 obs. of 3 variables:
 $ ID       : Factor w/ 8 levels "1","2","3","4",...: 1 1 1 2 2 2 3 3 3 4 ...
 $ Wave     : chr  "Pre" "Post" "FollowUp" "Pre" ...
 $ Resilience: num  6.49 5.28 5.93 4.43 5.95 ...
```

We just need to change Wave to be an ordered factor. Because R's default is to order factors alphabetically, we can use the levels command and identify our preferred order.

```
Amodeo_long$Wave <- factor(Amodeo_long$Wave, levels = c("Pre", "Post",
"FollowUp"))
```

We check the structure again.

```
str(Amodeo_long)
```

```
'data.frame': 24 obs. of 3 variables:
 $ ID       : Factor w/ 8 levels "1","2","3","4",...: 1 1 1 2 2 2 3 3 3 4 ...
 $ Wave     : Factor w/ 3 levels "Pre","Post","FollowUp": 1 2 3 1 2 3 1 2 3 1 ...
 $ Resilience: num  6.49 5.28 5.93 4.43 5.95 ...
```

Shape Shifters

The form of our data matters. The simulation created a *long* form (formally called the *person-period* form) of data. That is, each observation for each person is listed in its own row. In this dataset where we have 8 people with 3 observation (pre, post, follow-up) each, we have 24 rows. This is convenient, because this is the form we need for repeated measures ANOVA.

However, for some of the calculations (particularly those we will do by hand), we need the data to be in its more familiar wide form (formally called the *person level* form). We can do this with the *data.table* and *reshape2*()* packages.

```
# Create a new df (Amodeo_wide) Identify the original df We are
# telling it to connect the values of the Resilience variable its
# respective Wave designation
Amodeo_wide <- reshape2::dcast(data = Amodeo_long, formula = ID ~ Wave,
  value.var = "Resilience")
# doublecheck to see if they did what you think
str(Amodeo_wide)
```

```
'data.frame': 8 obs. of 4 variables:
$ ID      : Factor w/ 8 levels "1","2","3","4",...: 1 2 3 4 5 6 7 8
$ Pre     : num  6.49 4.43 4.77 5.91 4.84 ...
$ Post    : num  5.28 5.95 6.43 7 6.28 ...
$ FollowUp: num  5.93 5.19 6.54 6.19 6.24 ...
```

```
Amodeo_wide$ID <- factor(Amodeo_wide$ID)
```

In this reshape script, I asked for a quick structure check. The format of the variables looks correct. If you want to export these data as files to your computer, remove the hashtags to save (and re-import) them as .rds (R object) or .csv (“Excel lite”) files. This is not a necessary step to continue working the problem in this lesson.

The code for the .rds file will retain the formatting of the variables, but is not easy to view outside of R. I would choose this option.

```
# to save the df as an .rds (think 'R object') file on your computer;
# it should save in the same file as the .rmd file you are working
# with saveRDS(Amodeo_long, 'Amodeo_longRDS.rds')
# saveRDS(Amodeo_wide, 'Amodeo_wideRDS.rds') bring back the simulated
# dat from an .rds file Amodeo_long <- readRDS('Amodeo_longRDS.rds')
# Amodeo_wide <- readRDS('Amodeo_wideRDS.rds')
```

Another option is to write them as .csv files. The code for .csv will likely lose any variable formatting, but the .csv file is easy to view and manipulate in Excel. If you choose this option, you will probably need to re-run the prior code to reformat Wave as an ordered factor

```
# write the simulated data as a .csv write.table(Amodeo_long,
# file='Amodeo_longCSV.csv', sep=',', col.names=TRUE,
# row.names=FALSE) write.table(Amodeo_wide,
# file='Amodeo_wideCSV.csv', sep=',', col.names=TRUE,
# row.names=FALSE) bring back the simulated dat from a .csv file
# Amodeo_long <- read.csv ('Amodeo_longCSV.csv', header = TRUE)
# Amodeo_wide <- read.csv ('Amodeo_wideCSV.csv', header = TRUE)
```

6.3.2 Quick peek at the data

As we work the problem we will switch between long and wide formats. Before we get into the statistic let's inspect our data. We can start with the long form.

```
str(Amdeo_long)
```

```
'data.frame': 24 obs. of 3 variables:  
 $ ID       : Factor w/ 8 levels "1","2","3","4",...: 1 1 1 2 2 2 3 3 3 4 ...  
 $ Wave     : Factor w/ 3 levels "Pre","Post","FollowUp": 1 2 3 1 2 3 1 2 3 1 ...  
 $ Resilience: num 6.49 5.28 5.93 4.43 5.95 ...
```

In the following output, note the order of presentation of the grouping variable (i.e., FollowUp, Post, Pre). Even though we have ordered our factor so that “Pre” is first, the *describeBy()* function seems to be ordering them alphabetically.

```
psych::describeBy(Amodeo_long$Resilience, Wave, mat = TRUE, data = Amodeo_long,  
  digits = 3)
```

```
# Note. Recently my students and I have been having intermittent  
# struggles with the describeBy function in the psych package. We  
# have noticed that it is problematic when using .rds files and when  
# using data directly imported from Qualtrics. If you are having  
# similar difficulties, try uploading the .csv file and making the  
# appropriate formatting changes.
```

Another view (if we use the wide file).

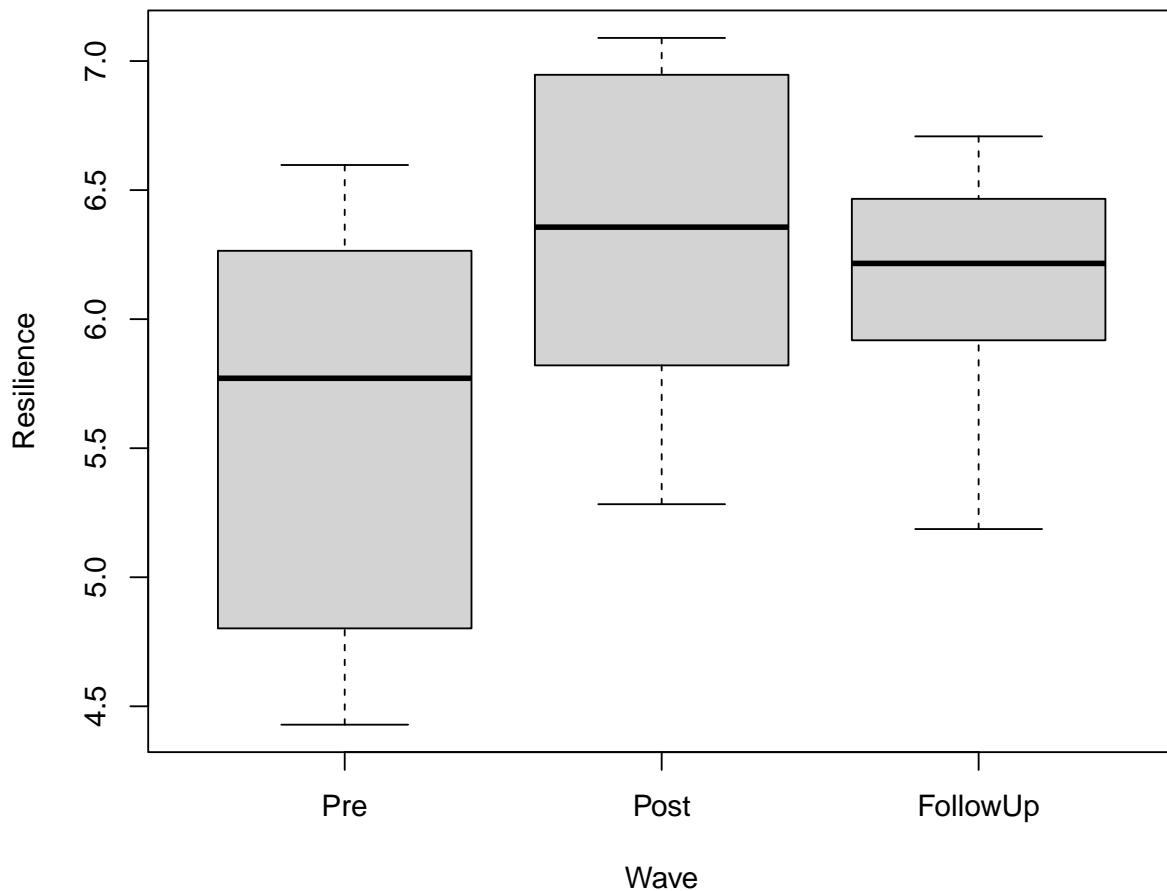
```
psych::describe(Amdeo_wide)
```

ID*	0.87
Pre	0.29
Post	0.23
FollowUp	0.17

Our means suggest that resilience increases from pre to post, then declines a bit. We use one-way repeated measures ANOVA to learn if there are statistically significant differences between the pairs of means and over time.

Let's also take a quick look at a boxplot of our data.

```
boxplot(Resilience ~ Wave, data = Amodeo_long, xlab = "Wave", ylab = "Resilience",
       n.label = TRUE)
```



6.4 Working the One-Way Repeated Measures ANOVA (by hand)

Before working our problem in R, let's gain a conceptual understanding by partitioning the variance by hand.

In repeated measures ANOVA: $SS_T = SS_B + SS_W$, where

- B = between-subjects variance
 - W = within-subjects variance
- $SS_W = SS_M + SS_R$

What differs is that SS_M and SS_R (model and residual) are located in SS_W

- $SS_T = SS_B + (SS_M + SS_R)$

		SS Within = 6.64 df (N - k) = 16		
Total	Model df	Residual	Between	
df formula	#cells-1	#levels-1	df _W - df _M	#people-1
SS	11.66	2.36	4.27	5.03
df	23	2	14	7
MS = SS/df		1.18	0.305	
F = MS _M /MS _R =		3.87		
*where N is number of cells				
F critical value = 3.73				
Because F > F _{cv} , we can reject the null hypothesis: F (2, 14) = 3.87, p < .05				

Figure 6.5: Demonstration of partitioning variance

6.4.1 Sums of Squares Total

Our formulas for SS_T are the same as they were for one-way and factorial ANOVA:

$$SS_T = \sum (x_i - \bar{x}_{grand})^2$$

$$SS_T = s_{grand}^2(n - 1)$$

Degrees of freedom for SS_T is $N - 1$, where N is the total number of cells.

Let's take a moment to *hand-calculate* SS_T (but using R).

Our grand (i.e., overall) mean is

```
mean(Amdeo_long$Resilience)
```

```
[1] 6.017408
```

Subtracting the grand mean from each resilience score yields a mean difference.

```
library(tidyverse)

Amodeo_long <- Amodeo_long %>%
  mutate(m_dev = Resilience - mean(Resilience))

head(Amodeo_long)
```

	ID	Wave	Resilience	m_dev
1	1	Pre	6.492125	0.47471697
2	1	Post	5.283057	-0.73435114
3	1	FollowUp	5.927930	-0.08947756
4	2	Pre	4.428839	-1.58856921
5	2	Post	5.948499	-0.06890871
6	2	FollowUp	5.186767	-0.83064071

Pop quiz: What's the sum of our new *m_dev* variable?

```
sum(Amodeo_long$m_dev)
```

```
[1] 0.00000000000007993606
```

If we square those mean deviations:

```
Amodeo_long <- Amodeo_long %>%
  mutate(m_devSQ = m_dev^2)

head(Amodeo_long)



|   | ID | Wave     | Resilience | m_dev       | m_devSQ     |
|---|----|----------|------------|-------------|-------------|
| 1 | 1  | Pre      | 6.492125   | 0.47471697  | 0.225356199 |
| 2 | 1  | Post     | 5.283057   | -0.73435114 | 0.539271599 |
| 3 | 1  | FollowUp | 5.927930   | -0.08947756 | 0.008006235 |
| 4 | 2  | Pre      | 4.428839   | -1.58856921 | 2.523552145 |
| 5 | 2  | Post     | 5.948499   | -0.06890871 | 0.004748410 |
| 6 | 2  | FollowUp | 5.186767   | -0.83064071 | 0.689963983 |


```

If we sum the squared mean deviations:

```
sum(Amodeo_long$m_devSQ)
```

```
[1] 11.65769
```

This value, the sum of squared deviations around the grand mean, is our SS_T ; the associated *degrees of freedom* is 23 (24 - 1; N - 1).

6.4.2 Sums of Squares Within for Repeated Measures ANOVA

The format of the formula is parallel to the formulae for SS we have seen before. In this case each person serves as its own grouping factor.

$$SS_W = s_{person1}^2(n_1 - 1) + s_{person2}^2(n_2 - 1) + s_{person3}^2(n_3 - 1) + \dots + s_{personk}^2(n_k - 1)$$

The degrees of freedom (df) within is $N - k$; or the summation of the df for each of the persons.

```
psych::describeBy(Resilience ~ ID, data = Amodeo_long, mat = TRUE, digits = 3)
```

	item	group1	vars	n	mean	sd	median	trimmed	mad	min	max	range
X11	1	1	1 3	5.901	0.605	5.928	5.901	0.836	5.283	6.492	1.209	
X12	2	2	1 3	5.188	0.760	5.187	5.188	1.124	4.429	5.948	1.520	
X13	3	3	1 3	5.912	0.992	6.430	5.912	0.160	4.768	6.537	1.769	
X14	4	4	1 3	6.370	0.568	6.191	6.370	0.414	5.913	7.005	1.092	
X15	5	5	1 3	5.787	0.824	6.240	5.787	0.064	4.836	6.283	1.447	
X16	6	6	1 3	5.744	0.146	5.693	5.744	0.095	5.629	5.908	0.279	
X17	7	7	1 3	6.627	0.248	6.597	6.627	0.300	6.395	6.889	0.494	
X18	8	8	1 3	6.612	0.533	6.708	6.612	0.565	6.038	7.090	1.052	
			skew	kurtosis	se							
X11	-0.044	-2.333	0.349									
X12	0.002	-2.333	0.439									
X13	-0.380	-2.333	0.573									
X14	0.283	-2.333	0.328									
X15	-0.384	-2.333	0.475									
X16	0.304	-2.333	0.084									
X17	0.118	-2.333	0.143									
X18	-0.175	-2.333	0.307									

With 8 people, there will be 8 chunks of the analysis, in each:

- SD squared (to get the variance)
- multiplied by the number of observations less 1

```
(0.605^2 * (3 - 1)) + (0.76^2 * (3 - 1)) + (0.992^2 * (3 - 1)) + (0.568^2 *
(3 - 1)) + (0.824^2 * (3 - 1)) + (0.146^2 * (3 - 1)) + (0.248^2 * (3 -
1)) + (0.553^2 * (3 - 1))
```

```
[1] 6.635836
```

6.4.3 Sums of Squares Model – Effect of Time

The SS_M conceptualizes the within-persons (or repeated measures) element as the grouping factor. In our case these are the pre, post, and follow-up clusters.

$$SS_M = \sum n_k (\bar{x}_k - \bar{x}_{grand})^2$$

The degrees of freedom will be $k - 1$ (number of levels, minus one).

```
psych::describe(Amodeo_wide)
```

	vars	n	mean	sd	median	trimmed	mad	min	max	range	skew	kurtosis
ID*		1	8	4.50	2.45	4.50	4.50	2.97	1.00	8.00	7.00	0.00
Pre		2	8	5.59	0.82	5.77	5.59	1.15	4.43	6.60	2.17	-0.14
Post		3	8	6.33	0.66	6.36	6.33	0.88	5.28	7.09	1.81	-0.23
FollowUp		4	8	6.14	0.47	6.22	6.14	0.44	5.19	6.71	1.52	-0.72
	se											
ID*			0.87									
Pre			0.29									
Post			0.23									
FollowUp			0.17									

In this case, we are interested in change in resilience over time. Hence, *time* is our factor. In our equation, we have three chunks representing the pre, post, and follow-up *conditions* (waves). From each, we subtract the grand mean, square it, and multiply by the n observed in each wave.

The degrees of freedom (df) for SS_M is $k - 1$

Let's calculate grand mean; that is the resilience score for all participants across all waves.

```
mean(Amodeo_long$Resilience)
```

```
[1] 6.017408
```

Now we can calculate the SS_M .

```
(8 * (6.14 - 6.017)^2) + (8 * (6.33 - 6.017)^2) + (8 * (5.59 - 6.017)^2)
```

```
[1] 2.363416
```

```
# df is 3-1 = 2
```

6.4.4 Sums of Squares Residual

Because $SS_W = SS_M + SS_R$ we can calculate SS_R with simple subtraction:

- $SS_w = 6.636$
- $SS_M = 2.363$

6.636 – 2.363

[1] 4.273

Correspondingly, the degrees of freedom (also taking the easy way out) is calculated by subtracting (the associated degrees of freedom) SS_M from SS_W .

16–2

[1] 14

6.4.5 Sums of Squares Between

The SS_B is not used in our calculations today, but also calculated easily. Given that $SS_T = SS_W + SS_B$:

- $SS_T = 11.66; df = 23$
- $SS_W = 6.64; df = 16$

11.66 – 6.64

[1] 5.02

23–16

[1] 7

$SS_B = 5.02, df = 7$

		SS Within =6.64		
		df (N - k) = 16		
<i>df formula</i>	Total	Model df	Residual	Between
	#cells-1	#levels-1	$df_w - df_m$	#people-1
SS	11.66		2.36	4.27
df	23		2	14
$MS = SS/df$			1.18	0.305
$F = MS_M/MS_R =$			3.87	
*where N is number of cells				
F critical value = 3.73				

Because $F > F_{cv}$, we can reject the null hypothesis: $F (2, 14) = 3.87, p < .05$

sourcetable, we can move through the steps to calculate our F statistic.

Looking again at our

6.4.6 Mean Squares Model & Residual

Now that we have the Sums of Squares, we can calculate the mean squares (we need these for our F statistic). Here is the formula for the mean square model.

$$MS_M = \frac{SS_M}{df_M}$$

```
#mean squares for the model
2.36/2
```

[1] 1.18

Here is the formula for mean square residual.

And $MS_R =$

$$MS_R = \frac{SS_R}{df_R}$$

Recall, degrees of freedom for the residual is $N - k$. In our case that is 90 - 3.

```
#mean squares for the residual
4.27 / 14
```

[1] 0.305

6.4.7 F ratio

The F ratio is calculated with MS_M and $MS_R =$.

$$F = \frac{MS_M}{MS_R}$$

```
1.18 / .305
```

[1] 3.868852

To find the F_{CV} we can use an [F distribution table](#). Or use a look-up function in R, which follows this general form: `qf(p, df1, df2, lower.tail=FALSE)`

```
qf(.05, 2, 14, lower.tail=FALSE)
```

[1] 3.738892

Our example has 2 (numerator) and 14 (denominator) degrees of freedom. If we use a table we find the corresponding degrees of freedom combinations for the column where $\alpha = .05$. We observe that any F value > 3.73 will be statistically significant. Our $F = 3.87$, so we have (just barely) exceeded the threshold. This is our *omnibus F*. We know there is at least 1 statistically significant difference between our pre, post, and follow-up conditions.

6.5 Working the One-Way ANOVA with R packages

6.5.1 Testing the assumptions

We will start by testing the assumptions. Highlighting in the figure notes our place in the one-way ANOVA workflow:

There are several different ways to conduct a repeated measures ANOVA. Each has different assumptions/requirements. These include:

- univariate statistics
 - This is what we will use today.
- multivariate statistics
 - Functionally similar to univariate, except the underlying algorithm does not require the sphericity assumption.
- multi-level modeling/hierarchical linear modeling
 - This is a different statistic altogether and is addressed in the [multilevel modeling OER](#).

6.5.1.1 Univariate assumptions for repeated measures ANOVA

- The cases represent a random sample from the population.
- There is no dependency in the scores *between* participants.
 - Of course there is intentional dependency in the repeated measures (or within-subjects) factor.
- There are no significant outliers in any cell of the design
 - Check by visualizing the data using box plots. The *identify_outliers()* function in the *rstatix* package identifies extreme outliers.
- The dependent variable is normally distributed in the population for each level of the within-subjects factor.
 - Conduct a Shapiro-Wilk test of normality for each of the levels of the DV.
 - Visually examine Q-Q plots.
- The population variance of difference scores computed between any two levels of a within-subjects factor is the same value regardless of which two levels are chosen; termed the **sphericity assumption**. This assumption is
 - akin to compound symmetry (both variances across conditions are equal).
 - akin to the homogeneity of variance assumption in between-group designs.
 - sometimes called the homogeneity-of-variance-of-differences assumption.
 - statistically evaluated with *Mauchly's test*. If Mauchly's $p < .05$, there are statistically significant differences. The *anova_test()* function in the *rstatix* package reports Mauchly's and two alternatives to the traditional F that correct the values by the degree to which the sphericity assumption is violated.

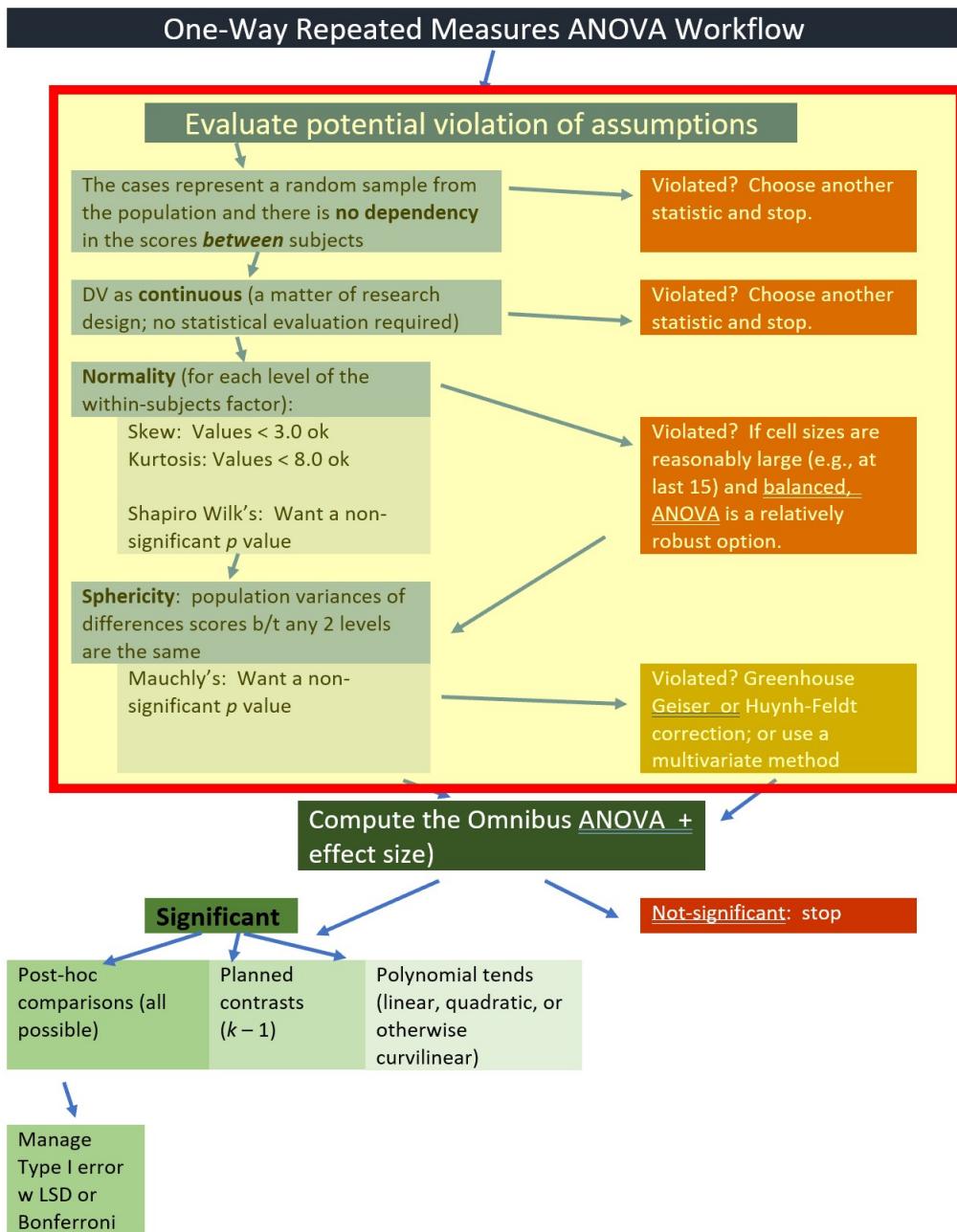


Figure 6.6: Image of our position in the workflow for the one-way repeated measures ANOVA

6.5.1.2 Demonstrating sphericity

Using the data from our motivating example, I calculated differences for each of the time variables. These are the three columns (in green shading) on the right. The variance for each is reported at the bottom of the column.

When we get into the analysis, we will use *Mauchly's test* in hopes that there are non-significant differences in variances between all three of the comparisons.

We are only concerned with the sphericity assumption if there are three or more groups.

$$\text{variance}_{A-B} \approx \text{variance}_{A-C} \approx \text{variance}_{B-C}$$

ID	Pre	Post	FollowUp	Pre-Pos	Pre-Fup	Pos-Fup
1	6.49	5.28	5.93	1.21	0.56	-0.64
2	4.43	5.95	5.19	-1.52	-0.76	0.76
3	4.77	6.43	6.54	-1.66	-1.77	-0.11
4	5.91	7.00	6.19	-1.09	-0.28	0.81
5	4.84	6.28	6.24	-1.45	-1.40	0.04
6	5.63	5.69	5.91	-0.06	-0.28	-0.21
7	6.60	6.89	6.39	-0.29	0.20	0.49
8	6.04	7.09	6.71	-1.05	-0.67	0.38
				Variance	0.95	0.60
					0.26	

Figure 6.7: Demonstration of unequal variances

6.5.1.3 Any outliers?

The boxplot is one common way for identifying outliers. The boxplot uses the median and the lower (25th percentile) and upper (75th percentile) quartiles. The difference bewteen Q3 and Q1 is the *interquartile range* (IQR). Outliers are generally identified when values fall outside these lower and upper boundaries:

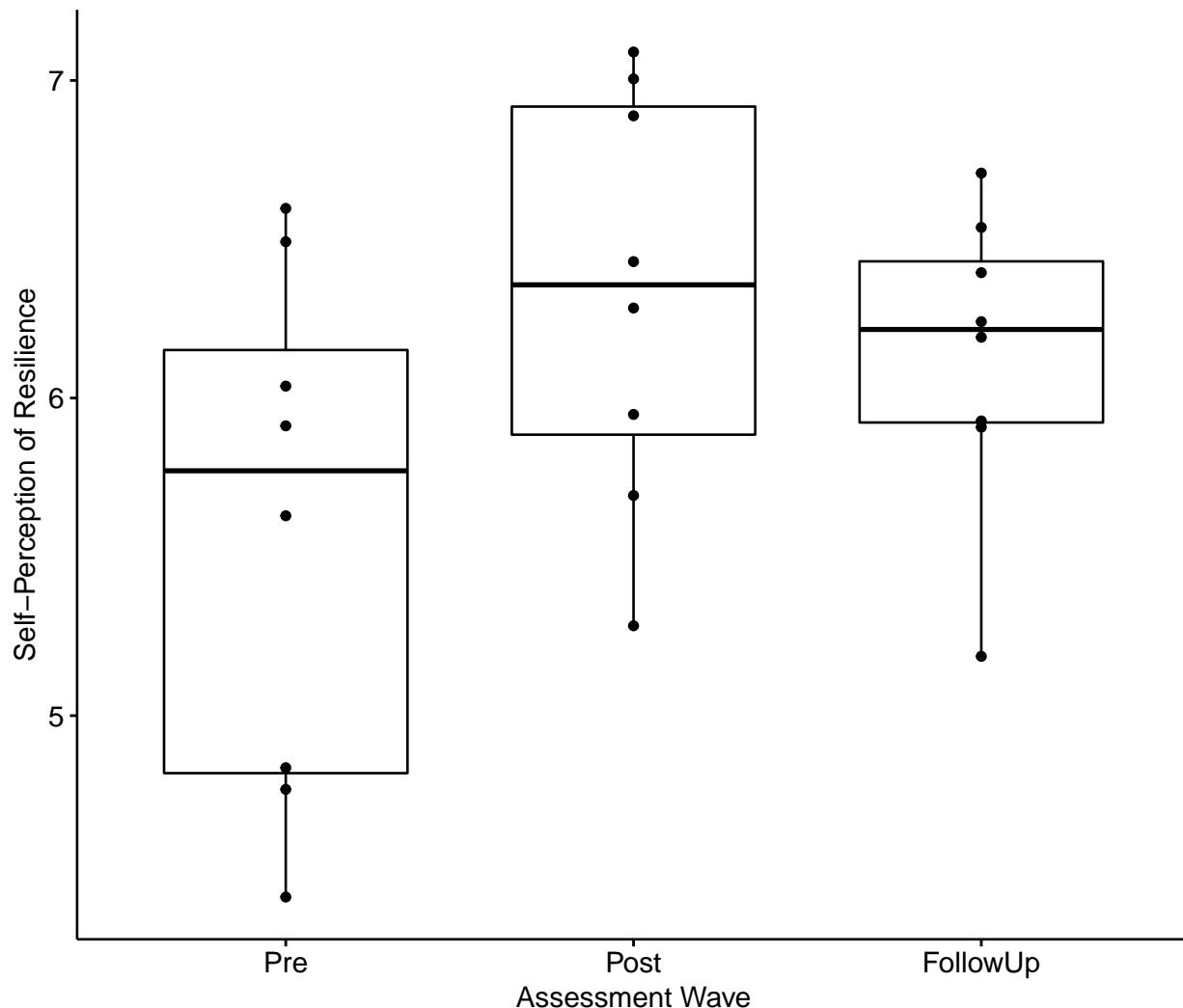
- $\text{Q1} - 1.5 \times \text{IQR}$
- $\text{Q3} + 1.5 \times \text{IQR}$

Extreme values occur when values fall outside these boundaries:

- $\text{Q1} - 3 \times \text{IQR}$
- $\text{Q3} + 3 \times \text{IQR}$

Let's take a look at a boxplot.

```
# Note that we are creating an object (bxp) from our work. This
# script creates the basic boxplot, we will add to it (by using the
# object) later.
bxp <- ggpubr::ggboxplot(Amdeo_long, x = "Wave", y = "Resilience", add = "point",
                           xlab = "Assessment Wave", ylab = "Self-Perception of Resilience")
bxp
```



The package *rstatix* has features that allow us to identify outliers.

```
Amdeo_long %>%
  group_by(Wave)%>%
  rstatix::identify_outliers(Resilience)
```

```
[1] Wave      ID       Resilience m_dev      m_devSQ    is.outlier is.extreme
<0 rows> (or 0-length row.names)
```

```
#?identify_outliers
```

The output, “0 rows” indicates there are no outliers.

This is consistent with the visual inspection of boxplots (above), where all observed scores fell within the 1.5x the interquartile range.

6.5.1.4 Assessing normality

We can obtain skew and kurtosis values for each cell in our model with the *psych::describeBy()* function.

```
psych::describeBy(Resilience ~ Wave, mat = TRUE, data = Amodeo_long)
```

item	group1	vars	n	mean	sd	median	trimmed	mad
X11	1	Pre	1 8	5.587693	0.8217561	5.770952	5.587693	1.1471137
X12	2	Post	1 8	6.327615	0.6550520	6.356491	6.327615	0.8751431
X13	3	FollowUp	1 8	6.136916	0.4729432	6.215983	6.136916	0.4416578
			min	max	range	skew	kurtosis	se
X11			4.428839	6.597214	2.168376	-0.1437061	-1.8118551	0.2905347
X12			5.283057	7.089591	1.806534	-0.2307393	-1.6287654	0.2315959
X13			5.186767	6.708259	1.521491	-0.7204842	-0.6102953	0.1672107

Our skew and kurtosis values fall below the thresholds of concern [Kline, 2016]:

- < 3 for skew
- 8 - 20 indicates extreme skew for kurtosis

We can use the Shapiro-Wilk test for a formal detection of normality. When $p < .05$, it indicates that the distribution is statistically significantly different than a normal one. Therefore, $p > .05$ indicates we did not violate the normal distribution assumption. The code below groups the DV by wave so that we can test normality for each cell in the model.

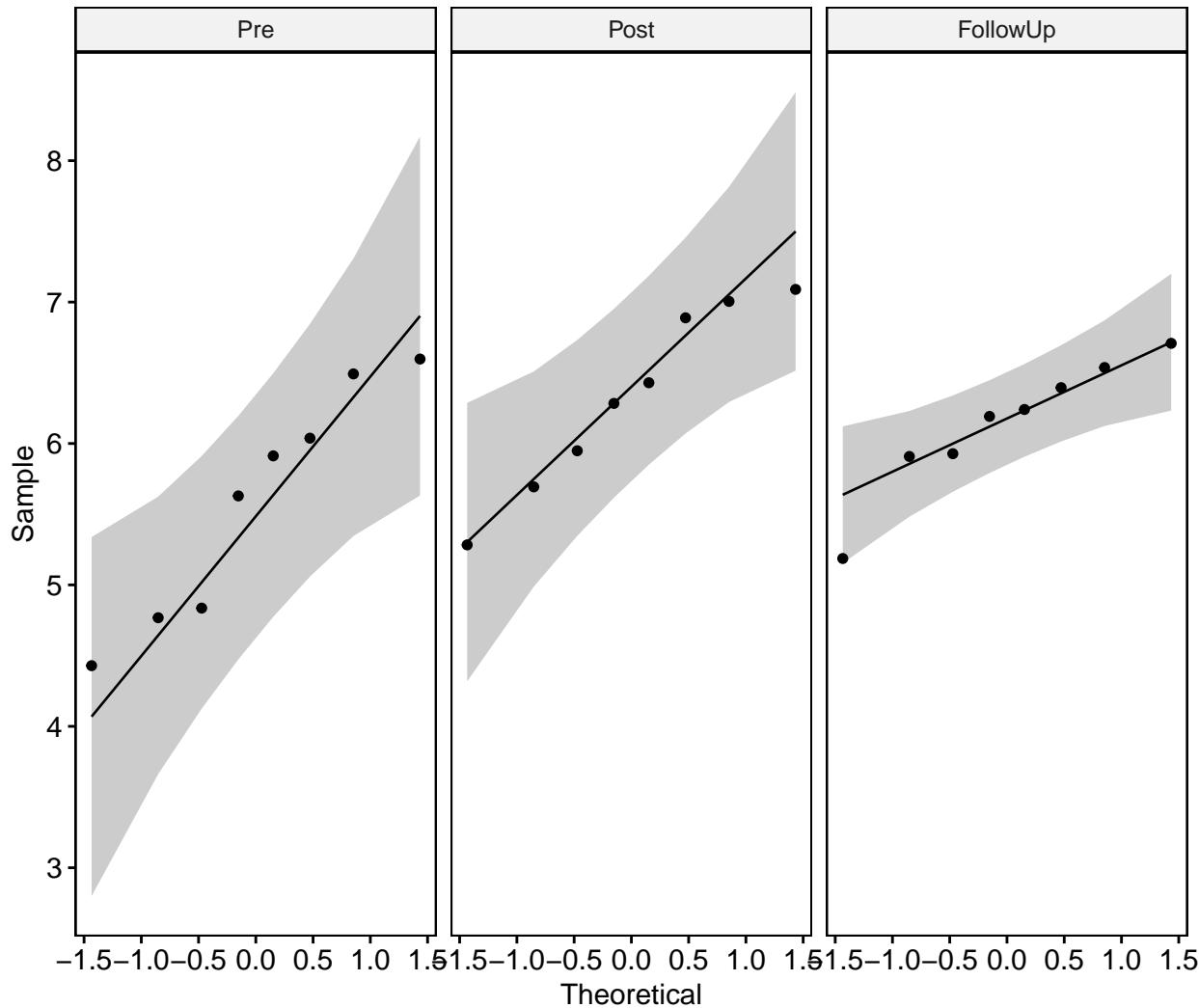
```
Amodeo_long %>%
  group_by(Wave) %>%
  rstatix::shapiro_test(Resilience)
```

```
# A tibble: 3 x 4
  Wave     variable   statistic     p
  <fct>    <chr>       <dbl> <dbl>
1 Pre      Resilience  0.919  0.419
2 Post     Resilience  0.941  0.617
3 FollowUp Resilience  0.926  0.480
```

The p value is $> .05$ for each of the cells. This provides some assurance that we have not violated the assumption of normality at any level of the design.

The Shapiro-Wilk test is sensitive to sample size [DataNovia, b]. Samples > 50 may lead to p values that are $< .05$, even when non-normality is not problematic. Therefore a second check with a Q-Q plot can be helpful. In a normal distribution the residuals will align along the line for each of the cells in the model.

```
ggpubr::ggqqplot(Amodeo_long, "Resilience", facet.by = "Wave")
```



APA Assumption Write-up So Far

Repeated measures ANOVA has several assumptions regarding outliers, normality, and sphericity. Visual inspection of boxplots for each wave of the design, assisted by the `identify_outliers()` function in the `rstatix` package (which reports values above $Q3 + 1.5 \times IQR$ or below $Q1 - 1.5 \times IQR$, where IQR is the interquartile range) indicated no outliers. Regarding normality, no values of skew and kurtosis (at each wave of assessment) fell within cautionary ranges for skew and kurtosis [Kline, 2016]. Additionally, the Shapiro-Wilk tests applied at each wave of the design were non-significant.

6.5.1.5 Assumption of Sphericity

The sphericity assumption is automatically checked with Mauchley's test during the computation of the ANOVA when the `rstatix::anova_test()` function is used. When the `rstatix::get_anova_table()` function is used, the Greenhouse-Geisser sphericity correction is automatically applied to factors violating the sphericity assumption.

The effect size, η^2 is reported in the column labeled “ges.”

Earlier in the lesson I noted that the evaluation of the sphericity assumption occurs at the same time that we evaluate the omnibus ANOVA. We are at that point in the analyses. The workflow points to our stage in the process.

6.5.2 Omnibus Repeated Measures ANOVA

As we prepare to run the omnibus ANOVA it may be helpful to think again about our variables. Our DV, Resilience, should be a continuous variable. In R, its structure should be “num.” Our predictor, Wave, should be categorical. In R case, Wave should be an ordered factor that is consistent with its timing: pre, post, follow-up.

The repeated measures ANOVA must be run with a long form of the data. This means that there needs to be a within-subjects identifier. In our case, it is the “ID” variable which is also formatted as a factor.

We can verify the format of our variables by examining the structure of our dataframe. Recall that we created the “m_dev” and “m_devSQ” variables earlier in the demonstration. We will not use them in this analysis; it does not harm anything for them to “ride along” in the dataframe.

```
str(Amodeo_long)
```

```
'data.frame': 24 obs. of 5 variables:
 $ ID       : Factor w/ 8 levels "1","2","3","4",...: 1 1 1 2 2 2 3 3 3 4 ...
 $ Wave     : Factor w/ 3 levels "Pre","Post","FollowUp": 1 2 3 1 2 3 1 2 3 1 ...
 $ Resilience: num  6.49 5.28 5.93 4.43 5.95 ...
 $ m_dev    : num  0.4747 -0.7344 -0.0895 -1.5886 -0.0689 ...
 $ m_devSQ  : num  0.22536 0.53927 0.00801 2.52355 0.00475 ...
```

We can use the `rstatix::anova_test()` function to calculate the omnibus ANOVA. Notice where our variables are included in the script:

- Resilience is the dv
- ID is the wid
- Wave is assigned to within

```
RM_AOV <- rstatix::anova_test(data = Amodeo_long, dv = Resilience, wid = ID,
                                within = Wave)
RM_AOV
```

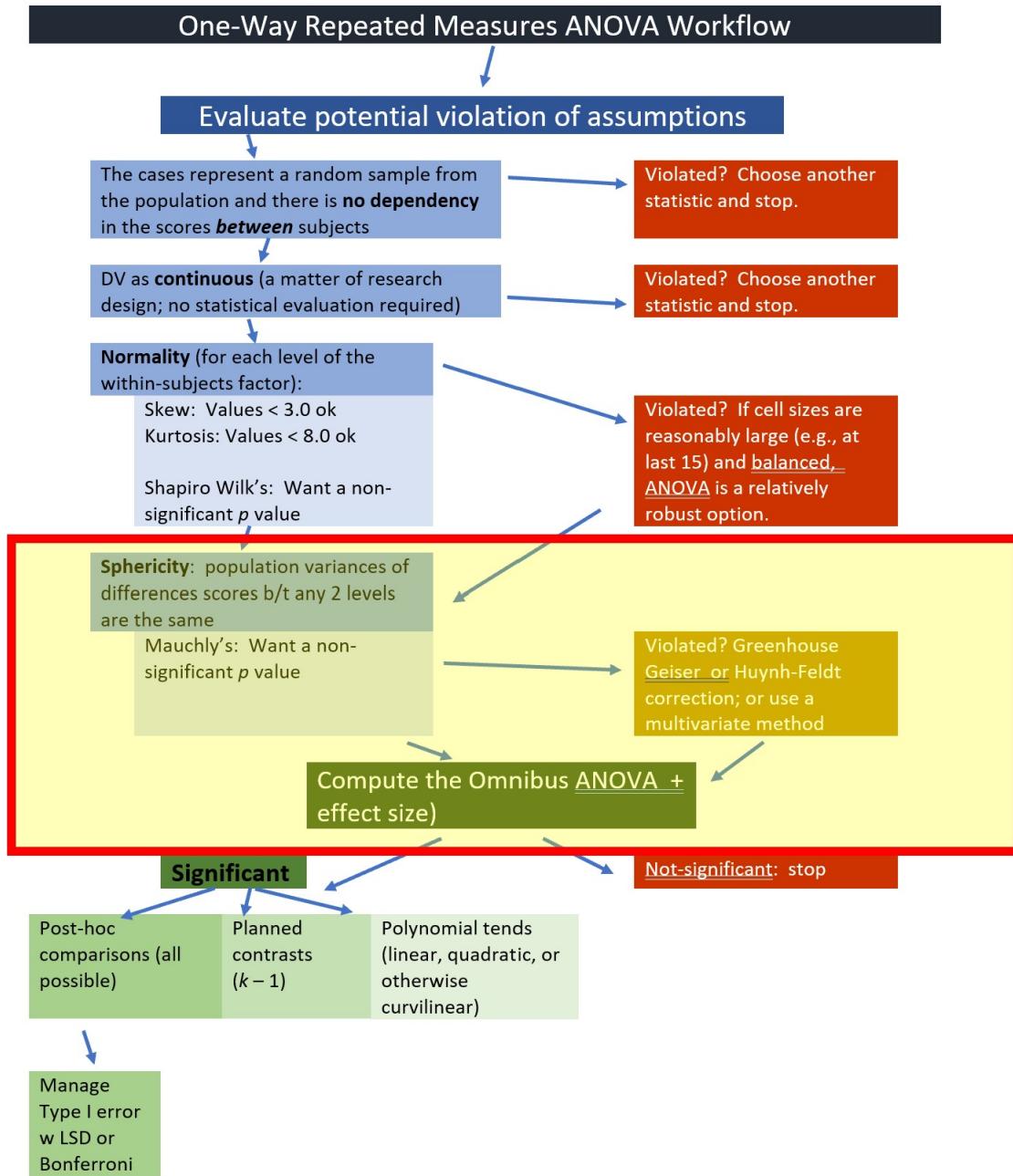


Figure 6.8: Image of our position in the workflow for the one-way repeated measures ANOVA

ANOVA Table (type III tests)

```
$ANOVA
Effect DFn DFD      F      p p<.05    ges
1   Wave    2  14 3.91 0.045      * 0.203

$`Mauchly's Test for Sphericity`
Effect      W      p p<.05
1   Wave 0.566 0.182

$`Sphericity Corrections`
Effect    GGe     DF[GG] p[GG] p[GG]<.05    HFe       DF[HF] p[HF] p[HF]<.05
1   Wave 0.698 1.4, 9.77 0.068           0.817 1.63, 11.44 0.057
```

We can assemble our F string from the ANOVA object: $F(2, 14) = 3.91, p = 0.045, \eta^2 = 0.203$

From the Mauchly's Test for Sphericity object we learn that we did not violate the sphericity assumption: $W = 0.566, p = .182$

From the Sphericity Corrections object are output for two alternative corrections to the F statistic, the Greenhouse-Geiser epsilon (GGe), and Huynh-Feldt epsilon (HFe). Because we did not violate the sphericity assumption we do not need to use them. Notice that these two tests adjust our df (both numerator and denominator) to have a more conservative p value.

If we needed to write an F string that is corrected for violation of the sphericity assumption, it might look like this:

The Greenhouse Geiser estimate was 0.698 the correct omnibus was $F(1.4, 9.77) = 3.91, p = .068$. The Huyhn Feldt estimate was 0.817 and the corrected omnibus was $F(1.63, 11.44) = 3.91 p = .057$.

You might be surprised that we are at follow-up already.

6.5.3 Follow-up

Given the simplicity of our design, it makes sense to me to follow-up with post hoc, pairwise, comparisons. Note that when I am calculating these pairwise t tests, I am creating an object (named “pwc”). The object will be a helpful tool in creating a Figure and an APA Style table.

```
pwc <- Amodeo_long %>%
  rstatix::pairwise_t_test(Resilience ~ Wave, paired = TRUE, p.adjust.method = "bonferroni")
pwc
```

```
# A tibble: 3 x 10
  .y.      group1 group2     n1     n2 statistic     df      p p.adj p.adj.signif
* <chr>    <chr>  <chr>    <int>  <int>     <dbl> <dbl> <dbl> <dbl> <chr>
1 Resilience Pre    Post      8      8     -2.15      7 0.069 0.206 ns
2 Resilience Pre    Follow~    8      8     -2.00      7 0.086 0.257 ns
3 Resilience Post   Follow~    8      8      1.06      7 0.325 0.975 ns
```

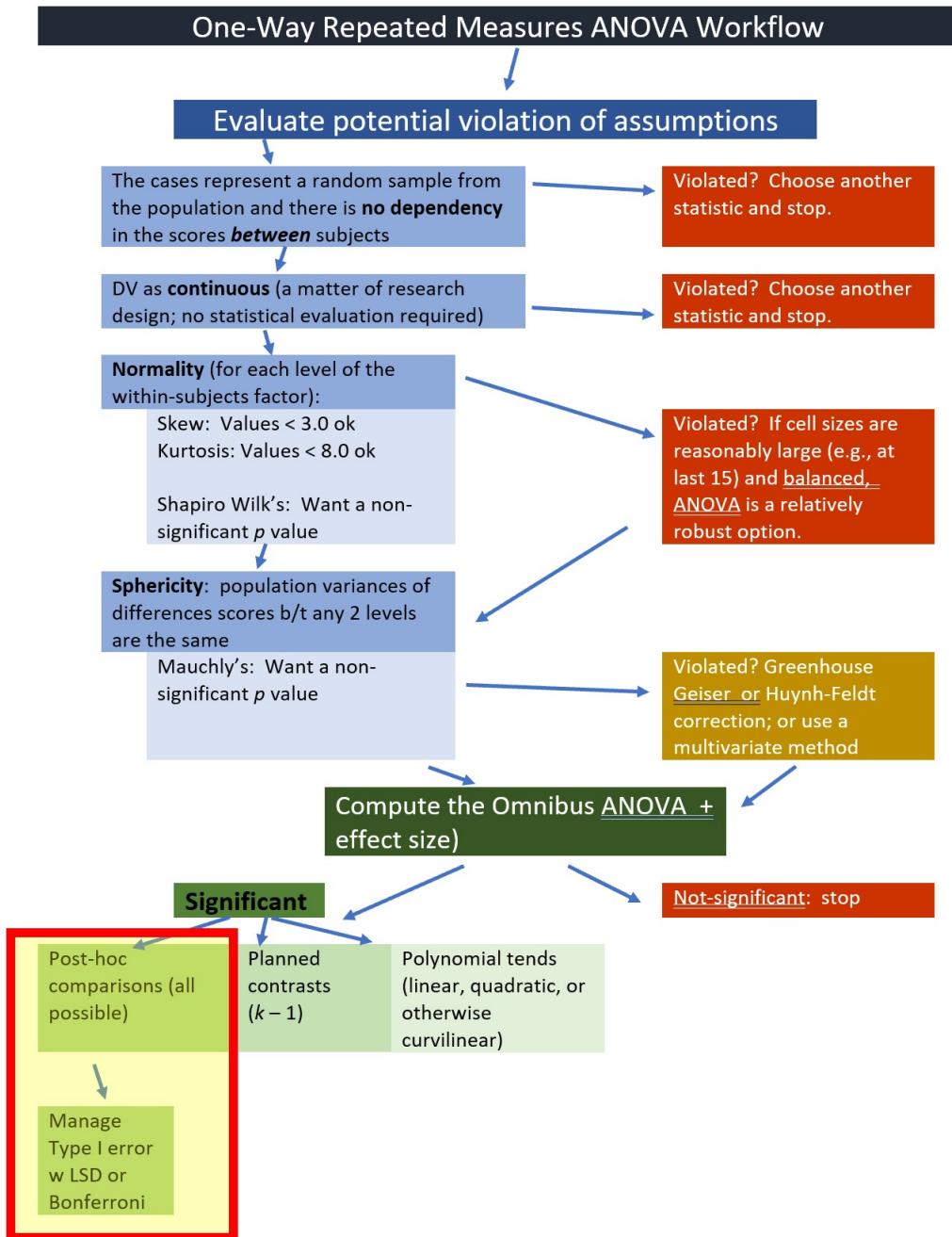


Figure 6.9: Image of our position in the workflow for the one-way repeated measures ANOVA

Although we had a statistically significant omnibus test, we did not obtain statistically significant results in any of the posthoc pairwise comparisons. Why not?

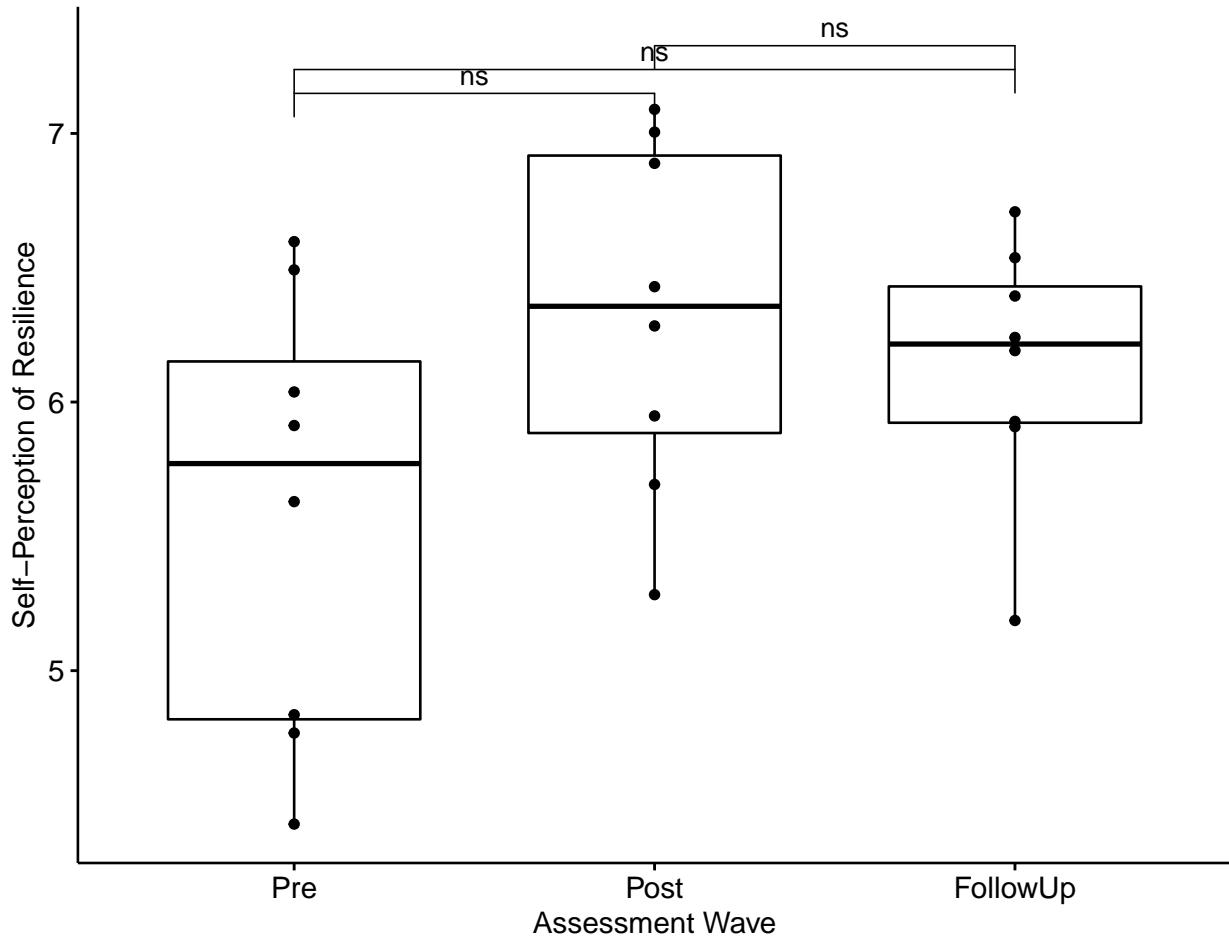
- Our omnibus F was right at the margins
 - a larger sample size (assuming that the effects would hold) would have been more powerful.
 - there could be significance if we compared pre to the combined effects of post and follow-up.

How would we manage Type I error? With only three possible post-omnibus comparisons, I would cite the Tukey LSD approach and not adjust the alpha to a more conservative level [[Green and Salkind, 2014b](#)].

We can combine information from the object we created (“bxp”) from an earlier boxplot with the object we saved from the posthoc pairwise comparisons (“pwc”) to enhance our boxplot with the F string and indications of pairwise significant (or, in our case, non-significance).

```
pwc <- pwc %>%
  rstatix::add_xy_position(x = "Wave")
bxp + ggpubr::stat_pvalue_manual(pwc) + labs(subtitle = rstatix::get_test_label(RM_AOV,
  detailed = TRUE), caption = rstatix::get_pwc_label(pwc))
```

Anova, $F(2,14) = 3.91, p = 0.045, \eta^2_g = 0.2$



Unfortunately, the *apaTables* package does not work with the *rstatix* package, so a table would need to be crafted by hand.

6.5.4 Results Section

Repeated measures ANOVA has several assumptions regarding outliers, normality, and sphericity. Visual inspection of boxplots for each wave of the design, assisted by the *rstatix::identify_outliers()* function (which reports values above $Q3 + 1.5 \times IQR$ or below $Q1 - 1.5 \times IQR$, where IQR is the interquartile range) indicated no outliers. Regarding normality, no values of skew and kurtosis (at each wave of assessment) fell within cautionary ranges for skew and kurtosis [Kline, 2016]. Additionally, the Shapiro-Wilk tests applied at each wave of the design were non-significant. A non-significant Mauchley's test ($W = 0.566, p = .182$) indicated that the sphericity assumption was not violated.

The omnibus ANOVA was significant: $F(2,14) = 3.91, p = 0.045, \eta^2 = 0.203$. We followed up with all pairwise comparisons. Curiously, and in spite of a significant omnibus test, there were not statistically significant differences between any of the pairs. Regarding pre versus post, $t = -2.15, p = .069$. Regarding pre versus follow-up, $t = -2.00, p = .068$. Regarding post versus follow-up, t

$\eta^2 = 1.059$, $p = .325$. Because there were only three pairwise comparisons subsequent to the omnibus test, alpha was retained at .05 [Green and Salkind, 2014b]. While the trajectories from pre-to-post and pre-to-follow-up were in the expected direction, the small sample size likely contributed to a Type II error. Descriptive statistics are reported in Table 1 and the differences are illustrated in Figure 1.

6.5.4.1 Creating an APA Style Table**

While I have not located a package that will take *rstatix* output to make an APA style table, we can use the *MASS* package to write the pwc object to a .csv file, then manually make our own table.

```
MASS:::write.matrix(pwc, sep = ",", file = "PWC.csv")
```

6.5.4.2 Comparison with Amodeo et al.[2018]

How do our findings and our write-up from the simulated data compare with the original article?

The F string is presented in the Table 1 note ($F[1.612, 11.283] = 6.390$, $p = 0.18$, η_p^2). We can tell from the fractional degrees of freedom that the p value has been had a correction for violation of the sphericity assumption.

Table 1 also reports all of the post-hoc, pairwise comparisons. There is no mention of control for Type I error. Had they used a traditional Bonferroni, they would have needed to reduce the alpha to $(k*(k-1)/2)$ and then divide .05 by that number.

```
(3 * (3-1))/2
```

```
[1] 3
```

```
.05/3
```

```
[1] 0.01666667
```

Although they report 6 comparisons; 3 are repeated because they are merely in reverse. Yet, the revised alpha would be .016 and the one, lone, comparison would not have been statistically significant. That said, we can use the Tukey LSD because there are only 3 comparisons and holding alpha at .05 can be defended [Green and Salkind, 2014b].

- Regarding the presentation of the results
 - there is no figure
 - there is no data presented in the text; all data is presented in Table 1
- Regarding the research design and its limitations
 - the authors note that a control condition would have better supported the conclusions

- the authors note the limited sample size and argue that this is a difficult group to recruit for intervention and evaluation
- the article is centered around the qualitative aspect of the design; the quantitative portion is, appropriately, secondary



Figure 6.10: Another peek at the research design for the Amodeo et al study

6.6 Power in Repeated Measures ANOVA

The package *wp.rmanova* was designed for power analysis in repeated measures ANOVA.

Power analysis allows us to determine the probability of detecting an effect of a given size with a given level of confidence. Especially when we don't achieve significance, we may want to stop.

In the *WebPower* package, we specify 6 of 7 interrelated elements; the package computes the missing one.

- n = sample size (number of individuals in the whole study)
- ng = number of groups
- nm = number of measurements/conditions/waves
- f = Cohen's f (an effect size; we can use a conversion calculator)
- $nscor$ = the Greenhouse Geiser correction from our output; 1.0 means no correction was needed and is the package's default; < 1 means some correction was applied.
- $alpha$ = is the probability of Type I error; we traditionally set this at .05
- $power$ = $1 - P(\text{Type II error})$ we traditionally set this at .80 (so anything less is less than what we want)
- $type$ = 0 is for between-subjects, 1 is for repeated measures, 2 is for interaction effect.

I used *effectsize* packages converter to transform our η^2 to Cohen's f .

```
effectsize::eta2_to_f(.203)
```

```
[1] 0.5046832
```

```
WebPower::wp.rmanova(n = 8, ng = 1, nm = 3, f = 0.5047, nscor = 0.689,
alpha = 0.05, power = NULL, type = 1)
```

Repeated-measures ANOVA analysis

```
n      f ng nm nscor alpha      power
8 0.5047  1  3 0.689  0.05 0.1619613
```

NOTE: Power analysis for within-effect test

URL: <http://psychstat.org/rmanova>

Here we learned that we were only powered at .16. That is, we had a 16% chance of finding a statistically significant effect if, in fact, it existed. This is low!

In reverse, setting *power* at .80 (the traditional value) and changing *n* to *NULL* yields a recommended sample size.

In many cases we won't know some of the values in advance. We can make best guesses based on our review of the literature. In the script below:

- *nscor* is the degree of violation of the sphericity assumption. If we think we won't violate it, we can enter 1.0 or leave it out (the *wp.ranova* default is 1.0)
- *f* is the effect size estimate; Cohen suggests that *f* values of 0.1, 0.25, and 0.4 represent small, medium, and large effect sizes, respectively.

```
WebPower::wp.ranova(n = NULL, ng = 1, nm = 3, f = 0.5047, nscor = 0.689,
alpha = 0.05, power = 0.8, type = 1)
```

Repeated-measures ANOVA analysis

```
n      f ng nm nscor alpha power
50.87736 0.5047  1  3 0.689  0.05    0.8
```

NOTE: Power analysis for within-effect test

URL: <http://psychstat.org/rmanova>

With these new values, we learn that we would need 50 individuals in order to feel confident in our ability to get a statistically significant result if, in fact, it existed.

6.7 Practice Problems

In each of these lessons I provide suggestions for practice that allow you to select one or more problems that offer differing levels of difficulty. Whichever you choose, you will focus on these larger steps in one-way repeated measures/within-subjects ANOVA, including:

- test the statistical assumptions
- conduct a one-way, including
 - omnibus test and effect size
 - conduct follow-up testing
- write a results section to include a figure and tables

6.7.1 Problem #1: Change the Random Seed

If repeated measures ANOVA is new to you, perhaps change the random seed and follow-along with the lesson.

6.7.2 Problem #2: Increase N

Our analysis of the Amodeo et al. [Amodeo et al., 2018] data failed to find significant increases in resilience from pre-to-post through follow-up. Our power analysis suggested that a sample size of 50 would be sufficient to garner statistical significance. The script below resimulates the data by increasing the sample size to 50 (from 8). All else remains the same.

```
set.seed(2022)
ID <- factor(c(rep(seq(1, 50), each = 3))) #gives me 8 numbers, assigning each number 3 consecutive
Resilience <- rnorm(150, mean = c(5.7, 6.21, 6.26), sd = c(0.88, 0.79,
  0.37)) #gives me a column of 24 numbers with the specified Ms and SD
Wave <- rep(c("Pre", "Post", "FollowUp"), each = 1, 50) #repeats pre, post, follow-up once each
Amodeo50_long <- data.frame(ID, Wave, Resilience)
```

6.7.3 Problem #3: Try Something Entirely New

Using data for which you have permission and access (e.g., IRB approved data you have collected or from your lab; data you simulate from a published article; data from an open science repository; data from other chapters in this OER), complete a one-way repeated measures ANOVA. Please have at least 3 levels for the predictor variable.

6.7.4 Grading Rubric

Regardless which option(s) you chose, use the elements in the grading rubric to guide you through the practice.

Assignment Component	Points Possible	Points Earned
1. Check and, if needed, format data	5	_____
2. Evaluate statistical assumptions	5	_____
3. Conduct omnibus ANOVA (w effect size)	5	_____
4. Conduct all possible pairwise comparisons (like in the lecture)	5	_____
5. Describe approach for managing Type I error	5	_____
6. APA style results with table(s) and figure	5	_____
7. Explanation to grader	5	_____
Totals	35	_____

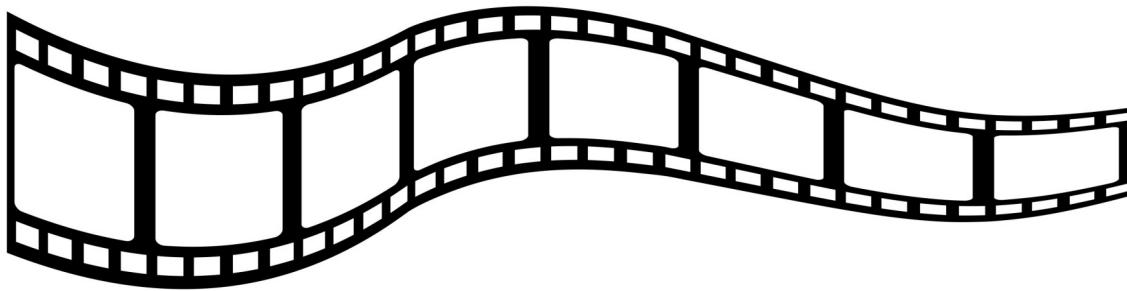


Figure 6.11: Image of a filmstrip

6.8 Bonus Reel:

Without the *rstatix* helper package, here is how the analysis would be run in the package, *car*. Note that this package results in the multivariate output. The *p* value of the omnibus *F* was non-significant from the start (*p* = .213).

```
library(car)

waveLevels <- c(1, 2, 3)
waveFactor <- as.factor(waveLevels)
waveFrame <- data.frame(waveFactor)
waveBind <- cbind(Amdeo_wide$Pre, Amdeo_wide$Post, Amdeo_wide$FollowUp)
waveModel <- lm(waveBind ~ 1)
analysis <- Anova(waveModel, idata = waveFrame, idesign = ~waveFactor)
summary(analysis)
```

Type III Repeated Measures MANOVA Tests:

Term: (Intercept)

Response transformation matrix:

(Intercept)	
[1,]	1
[2,]	1
[3,]	1

Sum of squares and products for the hypothesis:

(Intercept)	
(Intercept)	2607.062

Multivariate Tests: (Intercept)

	Df	test	stat	approx F	num Df	den Df	Pr(>F)
Pillai	1		0.9942	1200.028	1	7	0.0000000043326 ***
Wilks	1		0.0058	1200.028	1	7	0.0000000043326 ***
Hotelling-Lawley	1		171.4325	1200.028	1	7	0.0000000043326 ***
Roy	1		171.4325	1200.028	1	7	0.0000000043326 ***

Signif. codes:	0	'***'	0.001	'**'	0.01	'*'	0.05
	0.1	'	'
							1

Term: waveFactor

Response transformation matrix:

	waveFactor1	waveFactor2
[1,]	1	0
[2,]	0	1
[3,]	-1	-1

Sum of squares and products for the hypothesis:

	waveFactor1	waveFactor2
waveFactor1	2.4131705	-0.8378898
waveFactor2	-0.8378898	0.2909282

Multivariate Tests: waveFactor

	Df	test	stat	approx F	num Df	den Df	Pr(>F)
Pillai	1		0.4026101	2.021846	2	6	0.21319
Wilks	1		0.5973899	2.021846	2	6	0.21319
Hotelling-Lawley	1		0.6739486	2.021846	2	6	0.21319
Roy	1		0.6739486	2.021846	2	6	0.21319

Univariate Type III Repeated-Measures ANOVA Assuming Sphericity

	Sum Sq	num Df	Error SS	den Df	F value	Pr(>F)
(Intercept)	869.02	1	5.0692	7	1200.0279	0.000000004333 ***
waveFactor	2.36	2	4.2272	14	3.9102	0.04476 *

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Mauchly Tests for Sphericity

	Test statistic	p-value
waveFactor	0.56648	0.18179

Greenhouse-Geisser and Huynh-Feldt Corrections
for Departure from Sphericity

```
GG  eps  Pr(>F[GG])
waveFactor 0.69759    0.06754 .
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

HF  eps  Pr(>F[HF])
waveFactor 0.8172743 0.05734876
```

Chapter 7

Mixed Design ANOVA

[Screencasted Lecture Link](#)

The focus of this lecture is mixed design ANOVA. That is, we are conducting a two-way ANOVA where one of the factors is repeated measures and one of the factors is between groups. The mixed design ANOVA is often associated with the random clinical trial (RCT) where the researcher hopes for a significant interaction effect. Specifically, the researcher hopes that the individuals who were randomly assigned to the treatment condition improve from pre-test to post-test and maintain (or continue to improve) after post-test, while the people assigned to the no-treatment control are not statistically significantly different from treatment group at pre-test, and do not improve over time.

7.1 Navigating this Lesson

There is just over one hour of lecture. If you work through the materials with me it would be plan for an additional two hours.

While the majority of R objects and data you will need are created within the R script that sources the chapter, occasionally there are some that cannot be created from within the R framework. Additionally, sometimes links fail. All original materials are provided at the [Github site](#) that hosts the book. More detailed guidelines for ways to access all these materials are provided in the OER's [introduction](#)

7.1.1 Learning Objectives

Learning objectives from this lecture include the following:

- Evaluate the suitability of a research design/question and dataset for conducting a mixed design ANOVA; identify alternatives if the data is not suitable.
- Test the assumptions for mixed design ANOVA.
- Conduct a mixed design ANOVA (omnibus and follow-up) in R.
- Interpret output from the mixed design ANOVA (and follow-up).
- Prepare an APA style results section of the mixed design ANOVA output.
- Conduct a power analysis for mixed design ANOVA.

7.1.2 Planning for Practice

In each of these lessons I provide suggestions for practice that allow you to select one or more problems that are graded in difficulty. The least complex is to change the random seed and rework the problem demonstrated in the lesson. The results *should* map onto the ones obtained in the lecture.

The second option comes from the research vignette. The Murrar and Brauer [2018] article has three variables (attitudes toward Arabs, attitudes toward Whites, and a difference score) which are suitable for mixed design ANOVAs. I will demonstrate a mixed design ANOVA with the difference score. I'll leave the other two variables for opportunities for practice.

As a third option, you are welcome to use data to which you have access and is suitable for two-way ANOVA. In either case the practice options suggest that you:

- test the statistical assumptions
- conduct a mixed design ANOVA, including
 - omnibus test and effect size
 - report main and interaction effects
 - conduct follow-up testing of simple main effects
- write a results section to include a figure and tables

7.1.3 Readings & Resources

In preparing this chapter, I drew heavily from the following resource(s). Other resources are cited (when possible, linked) in the text with complete citations in the reference list.

- Repeated Measures ANOVA in R: The Ultimate Guide. (n.d.). Datanovia. Retrieved October 19, 2020, from <https://www.datanovia.com/en/lessons/repeated-measures-anova-in-r/>
 - This website is an excellent guide for both one-way repeated measures and mixed design ANOVA. It is a great resource for both the conceptual and procedural. This is the guide I have used for the basis of the lecture. Working through their example would be provide an additional, excellent, opportunity for practice.
- Murrar, S., & Brauer, M. (2018). Entertainment-education effectively reduces prejudice. *Group Processes & Intergroup Relations*, 21(7), 1053–1077. <https://doi.org/10.1177/1368430216682350>
 - This article is the source of our research vignette. Our vignette is simulated from the first of their two experiments. The authors did not conduct mixed design ANOVA. Instead, they ran independent-samples *t* tests to test the differences between the sitcom conditions for each of the three waves. This is comparable to conducting the simple-main effect analysis of condition within wave subsequent to a significant interaction.
 - Full-text of the article is available at the [authors' ResearchGate](#).

7.1.4 Packages

The packages used in this lesson are embedded in this code. When the hashtags are removed, the script below will (a) check to see if the following packages are installed on your computer and, if not (b) install them.

```
# will install the package if not already installed
# if(!require(knitr)){install.packages('knitr')}
# if(!require(tidyverse)){install.packages('tidyverse')}
# if(!require(psych)){install.packages('psych')}
# if(!require(ggpubr)){install.packages('ggpubr')}
# if(!require(rstatix)){install.packages('rstatix')}
# if(!require(MASS)){install.packages('MASS')}
# if(!require(effectsize)){install.packages('effectsize')}
# if(!require(WebPower)){install.packages('WebPower')}
```

7.2 Introducing Mixed Design ANOVA

Mixed design ANOVA is characterized by the following:

- at least two independent variables.
- Termed “mixed” because
 - one is a between-subjects factor, and
 - one is a repeated-measures (i.e., within-subjects) factor.
- In essence, we are simultaneously conducting
 - a one-way independent ANOVA and a
 - a one-way repeated-measures ANOVA.

Especially when there is a significant interaction there can be numerous ways to follow up. We will work one set of analyses: simple main effects (condition within wave; wave within condition) and, when needed, conduct posthoc pairwise comparisons as follow-up. Other good options include identifying a priori contrasts and conducting polynomials (not demonstrated in this lecture).

The steps in working the mixed design generally include,

1. Exploring the data/evaluating the assumptions
2. Evaluating the omnibus test
3. Follow-up to the omnibus
 - if significant interaction effect: simple main effects and further follow-up to those
 - if significant main effect (but no significant interaction effect), identify source of significance in the main effect
 - if no significance, stop
4. Write it up with tables, figure(s)

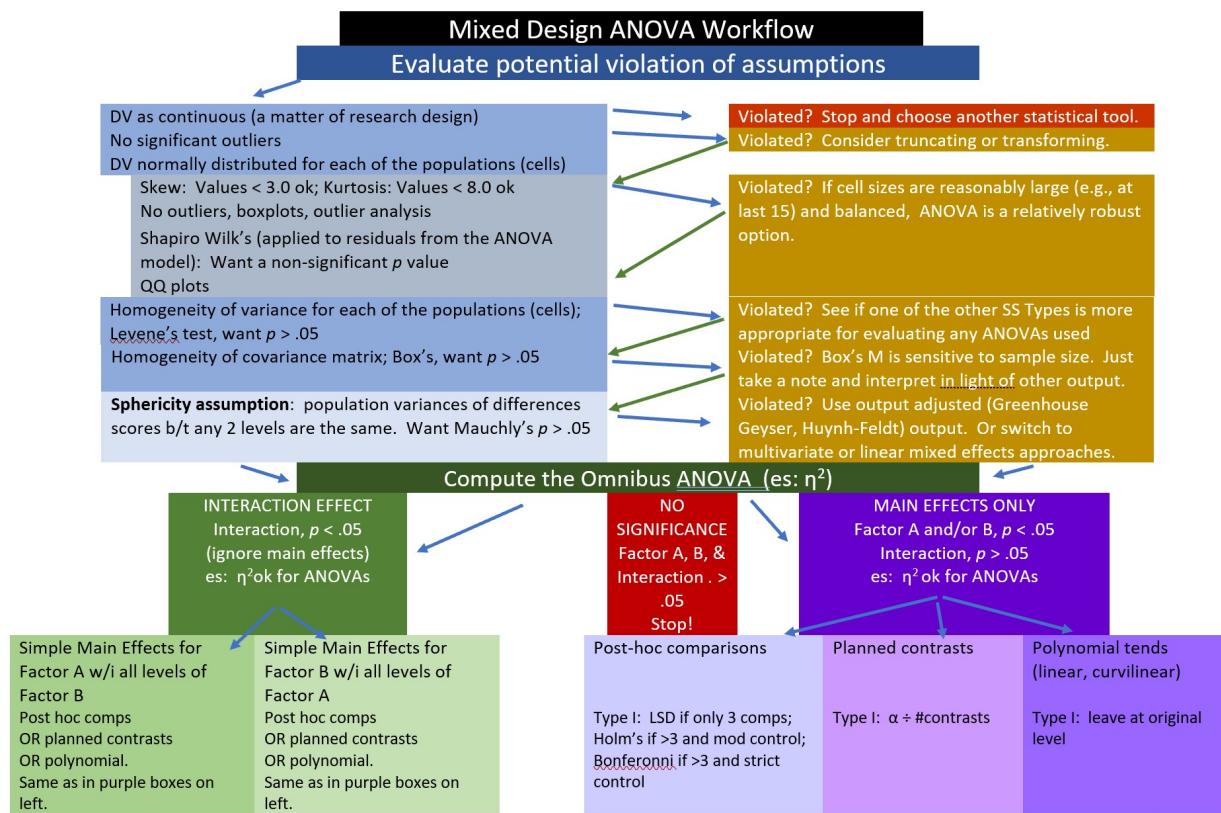


Figure 7.1: Image of a workflow for mixed design ANOVA

Assumptions for the mixed design ANOVA include the following:

- The dependent variable should be continuous with no significant outliers in any cell of the design
 - Check by visualizing the data using box plots and by using the `rstatix::identify_outliers()` function
- The DV should be approximately normally distributed in each cell of the design
 - Check with Shapiro-Wilk normality test `rstatix::shapiro_test()` function and with visual inspection by creating Q-Q plots. The `ggpubr::ggqqplot()` function is a great tool.
- The variances of the differences between groups should be equal. This is termed the **sphericity assumption**. This can be checked with Mauchly's test of sphericity, which is reported automatically in the `rstatix::anova_test()` output.

The best way to address violations of these assumptions is not always clear. Possible solutions include:

- For 2- and 3- way ANOVAs, violations of the normality assumption might be addressed by removing extreme outliers or considering transformations of the data. Transformations, though, introduce their own complexities regarding interpretation. Kline's text [2016] provides excellent coverage of options.
- A robust ANOVA option is available in the `WRS2` package
- If there are three or more waves/conditions and the sample is large, it may be possible to run a multi-level, model.
- In the absence of alternatives, it may be necessary to run the mixed design with the violated assumptions, but report them.
-and more. Internet searches continue to offer new approaches and alternatives.

7.3 Research Vignette

This lesson's research vignette is from Murrar and Brauer's [2018] article that describes the results of two studies that evaluated interventions designed to reduce prejudice against Arabs/Muslims. We are working only a portion of the first study reported in the article. Participants ($N = 193$), all who were White, were randomly assigned to one of two conditions where they watched six episodes of the sitcom *Friends* or *Little Mosque on the Prairie*. The sitcoms and specific episodes were selected after significant pilot testing. The selection was based on the tension selecting stimuli that were as similar as possible, yet the intervention-oriented sitcom needed to invoke psychological processes known to reduce prejudice. The authors felt that both series had characters that were likable and relatable and were engaged in regular activities of daily living. The Friends series featured characters who were predominantly White, cis-gendered, and straight. The Little Mosque series portrayed the experience of Western Muslims and Arabs as they lived in a small Canadian town. This study involved assessment across three waves: baseline (before watching the assigned episodes), post1 (immediately after watching the episodes), and post2 (completed 4-6 weeks after watching the episodes).

The study used *feelings and liking thermometers*, rating their feelings and liking toward 10 different groups of people on a 0 to 100 sliding scale (with higher scores reflecting greater liking and positive feelings). For the purpose of this analysis, the ratings of attitudes toward White people and attitudes toward Arabs/Muslims were used. A third metric was introduced by subtracting the attitudes towards Arabs/Muslims from the attitudes toward Whites. Higher scores indicated more positive attitudes toward Whites where as low scores indicated no difference in attitudes. To recap, there were three potential dependent variables, all continuously scaled:

- *AttWhite*: attitudes toward White people; higher scores reflect greater liking
- *AttArab*: attitudes toward Arab people; higher scores reflect greater liking
- *Diff*: the difference between AttWhite and AttArab; higher scores reflect a greater liking for White people

With random assignment, nearly equal cell sizes, a condition with two levels (Friends, Little Mosque), and three waves (baseline, post1, post2), this is perfect for mixed design ANOVA.

COND	Baseline At start of study (prior to viewing sitcoms)	Intervention 6 episodes of the sitcom	Post1 Toward end of viewing the sitcoms	Post2 4-6 weeks after viewing the final sitcom
Friends	X		X	X
Little Mosque on the Prairie	X	Selected for potential for prejudice reduction	X	X

Figure 7.2: Image of the design for the Murrar and Brauer (2018) study

7.3.1 Simulating the data from the journal article

Below is the code I have used to simulate the data. The simulation includes two dependent variables (AttWhite, AttArab), Wave (baseline, post1, post2), and COND (condition; Friends, Little_Mosque). There is also a caseID (repeated three times across the three waves) and rowID (giving each observation within each case an ID). This creates the long-file, where each person has 3 rows of data representing baseline, post1, and post2. You can use this simulation for two of the three practice suggestions.

```
library(tidyverse)
# change this to any different number (and rerun the simulation) to
# rework the chapter problem
set.seed(210813)
AttWhite <- round(c(rnorm(98, mean = 76.79, sd = 18.55), rnorm(95, mean = 75.37,
  sd = 18.99), rnorm(98, mean = 77.47, sd = 18.95), rnorm(95, mean = 75.81,
  sd = 19.29), rnorm(98, mean = 77.79, sd = 17.25), rnorm(95, mean = 75.89,
  sd = 19.44)), 3) #sample size, M and SD for each cell; this will put it in a long file
# set upper bound for variable
```

```

AttWhite[AttWhite > 100] <- 100
# set lower bound for variable
AttWhite[AttWhite < 0] <- 0
AttArab <- round(c(rnorm(98, mean = 64.11, sd = 20.97), rnorm(95, mean = 64.37,
  sd = 20.03), rnorm(98, mean = 64.16, sd = 21.64), rnorm(95, mean = 70.52,
  sd = 18.55), rnorm(98, mean = 65.29, sd = 19.76), rnorm(95, mean = 70.3,
  sd = 17.98)), 3)
# set upper bound for variable
AttArab[AttArab > 100] <- 100
# set lower bound for variable
AttArab[AttArab < 0] <- 0
rowID <- factor(seq(1, 579))
caseID <- rep((1:193), 3)
Wave <- c(rep("Baseline", 193), rep("Post1", 193), rep("Post2", 193))
COND <- c(rep("Friends", 98), rep("LittleMosque", 95), rep("Friends", 98),
  rep("LittleMosque", 95), rep("Friends", 98), rep("LittleMosque", 95))
# groups the 3 variables into a single df: ID#, DV, condition
Murrar_df <- data.frame(rowID, caseID, Wave, COND, AttArab, AttWhite)

```

Let's check the structure. We want

- rowID and caseID to be unordered factors
- Wave and COND to be ordered factors
- AttArab and AttWhite to be numerical

```
str(Murrar_df)
```

```
'data.frame': 579 obs. of 6 variables:
 $ rowID   : Factor w/ 579 levels "1","2","3","4",...: 1 2 3 4 5 6 7 8 9 10 ...
 $ caseID  : int  1 2 3 4 5 6 7 8 9 10 ...
 $ Wave     : chr  "Baseline" "Baseline" "Baseline" "Baseline" ...
 $ COND     : chr  "Friends" "Friends" "Friends" "Friends" ...
 $ AttArab : num  74.3 55.8 33.3 66.3 71 ...
 $ AttWhite: num  100 79 75.9 68.2 100 ...
```

The script below changes

- caseID from integer to factor
- Wave and COND from factor to ordered factors
 - It makes sense to order Friends and LittleMosque, since we believe that LittleMosque contains prejudice-reducing properties

```
# make caseID a factor
Murrar_df[, "caseID"] <- as.factor(Murrar_df[, "caseID"])
# make Wave an ordered factor
Murrar_df$Wave <- factor(Murrar_df$Wave, levels = c("Baseline", "Post1",
"Post2"))
# make COND an ordered factor
Murrar_df$COND <- factor(Murrar_df$COND, levels = c("Friends", "LittleMosque"))
```

Let's check the structure again.

```
str(Murrar_df)
```

```
'data.frame': 579 obs. of 6 variables:
 $ rowID   : Factor w/ 579 levels "1","2","3","4",...: 1 2 3 4 5 6 7 8 9 10 ...
 $ caseID  : Factor w/ 193 levels "1","2","3","4",...: 1 2 3 4 5 6 7 8 9 10 ...
 $ Wave    : Factor w/ 3 levels "Baseline","Post1",...: 1 1 1 1 1 1 1 1 1 1 ...
 $ COND    : Factor w/ 2 levels "Friends","LittleMosque": 1 1 1 1 1 1 1 1 1 1 ...
 $ AttArab : num  74.3 55.8 33.3 66.3 71 ...
 $ AttWhite: num  100 79 75.9 68.2 100 ...
```

A key dependent variable in the Murrar and Brauer [Murrar and Brauer, 2018] article is *attitude difference*. Specifically, the attitudes toward Arabs score was subtracted from the attitudes toward Whites scores. Higher attitude difference indicate a greater preference for Whites. Let's create that variable, here.

```
Murrar_df$Diff <- Murrar_df$AttWhite - Murrar_df$AttArab
```

The code for the .rds file will retain the formatting of the variables, but is not easy to view outside of R. This is what I would do.

```
# to save the df as an .rds (think 'R object') file on your computer;
# it should save in the same file as the .rmd file you are working
# with saveRDS(Murrar_df, 'Murrar_RDS.rds') bring back the simulated
# dat from an .rds file Murrar_df <- readRDS('Murrar_RDS.rds')
```

If you want to export this data as a file to your computer, remove the hashtags to save it (and re-import it) as a .csv (“Excel lite”) or .rds (R object) file. This is not a necessary step.

The code for .csv will likely lose the formatting (i.e., stripping Wave and COND of their ordered factors), but it is easy to view in Excel.

```
# write the simulated data as a .csv write.table(Murrar_df,
# file='DiffCSV.csv', sep=',', col.names=TRUE, row.names=FALSE) bring
# back the simulated dat from a .csv file Murrar_df <- read.csv
# ('DiffCSV.csv', header = TRUE)
```

7.4 Working the Mixed Design ANOVA with R packages

7.4.1 Exploring data and testing assumptions

We begin the 2x3 mixed design ANOVA with a preliminary exploration of the data and testing of the assumptions. Here's where we are on the workflow:

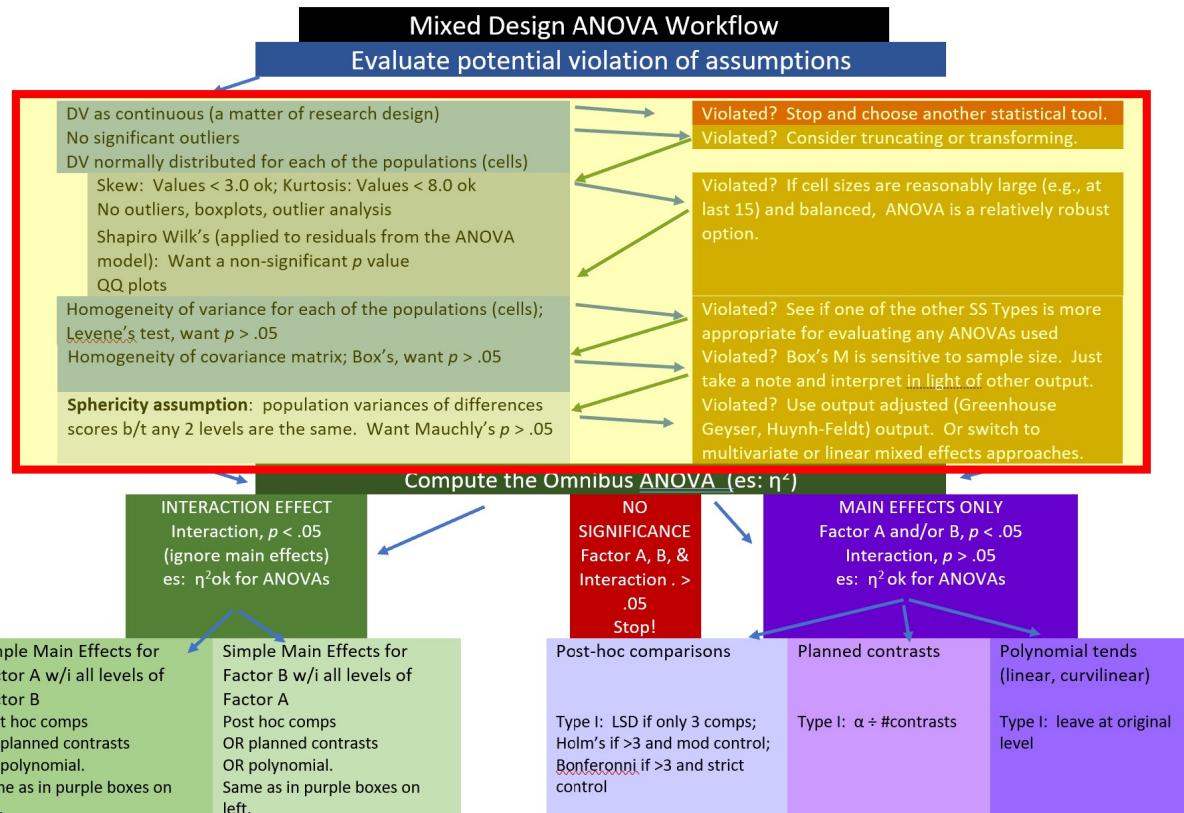


Figure 7.3: Image of the workflow showing that we are on the “Evaluating assumptions” portion

First, let's examine the overall descriptive statistics.

```
psych::describe(Murrar_df)
```

	vars	n	mean	sd	median	trimmed	mad	min	max	range
rowID*	1	579	290.00	167.29	290.00	290.00	214.98	1.00	579.00	578.00
caseID*	2	579	97.00	55.76	97.00	97.00	71.16	1.00	193.00	192.00
Wave*	3	579	2.00	0.82	2.00	2.00	1.48	1.00	3.00	2.00
COND*	4	579	1.49	0.50	1.00	1.49	0.00	1.00	2.00	1.00
AttArab	5	579	66.84	19.75	68.04	67.64	20.38	6.14	100.00	93.86
AttWhite	6	579	75.31	17.02	76.72	76.19	18.50	23.03	100.00	76.97
Diff	7	579	8.47	26.33	8.65	8.50	25.73	-71.51	90.74	162.25
			skew	kurtosis	se					
rowID*		0.00	-1.21	6.95						

caseID*	0.00	-1.21	2.32
Wave*	0.00	-1.51	0.03
COND*	0.03	-2.00	0.02
AttArab	-0.39	-0.16	0.82
AttWhite	-0.37	-0.51	0.71
Diff	0.03	0.00	1.09

Our analysis will use the difference score (Diff) as the dependent variable. Let's look at this variable in its combinations of wave and condition.

```
psych::describeBy(Diff ~ Wave + COND, data = Murrar_df, mat = TRUE)
```

item	group1	group2	vars	n	mean		sd	median	trimmed
					Friends	LittleMosque			
X11	1	Baseline	Friends	1	98	9.3064898	23.90867	8.804	8.9906625
X12	2	Post1	Friends	1	98	15.9261327	26.41789	16.191	16.2309375
X13	3	Post2	Friends	1	98	11.9540102	23.33602	10.882	11.8340000
X14	4	Baseline	LittleMosque	1	95	9.7331158	30.51895	10.797	10.5544156
X15	5	Post1	LittleMosque	1	95	-0.1486632	26.96858	-1.280	-0.9402727
X16	6	Post2	LittleMosque	1	95	3.6704737	23.66524	1.860	3.7857403
			mad	min	max	range	skew	kurtosis	se
X11	24.48143	-47.342	72.565	119.907	0.17600603	-0.35619536	2.415140		
X12	29.77135	-42.598	82.288	124.886	-0.04613379	-0.57456759	2.668609		
X13	24.59189	-46.528	75.014	121.542	0.13240007	0.06485450	2.357294		
X14	30.90480	-71.510	90.737	162.247	-0.20135358	-0.03033805	3.131178		
X15	23.93213	-65.259	83.367	148.626	0.32819576	0.54919109	2.766918		
X16	25.26054	-53.856	55.264	109.120	-0.06475209	-0.42366384	2.428002		

```
# Note. Recently my students and I have been having intermittent
# struggles with the describeBy function in the psych package. We
# have noticed that it is problematic when using .rds files and when
# using data directly imported from Qualtrics. If you are having
# similar difficulties, try uploading the .csv file and making the
# appropriate formatting changes.
```

First we inspect the means. We see that the baseline scores for the Friends and Little Mosque conditions are similar. However, the post1 and post2 difference scores (i.e., difference in attitudes toward White and Arab individuals, where higher scores indicate more favorable ratings of White individuals) are higher in the Friends condition than in the Little Mosque condition.

7.4.1.1 Assumption of Normality

We can use this output to evaluate the distributional characteristics of the dependent variable. Recall that mixed design ANOVA assumes a normal distribution.

Our values of skew and kurtosis are well within the limits [Kline, 2016] of a normal distribution.

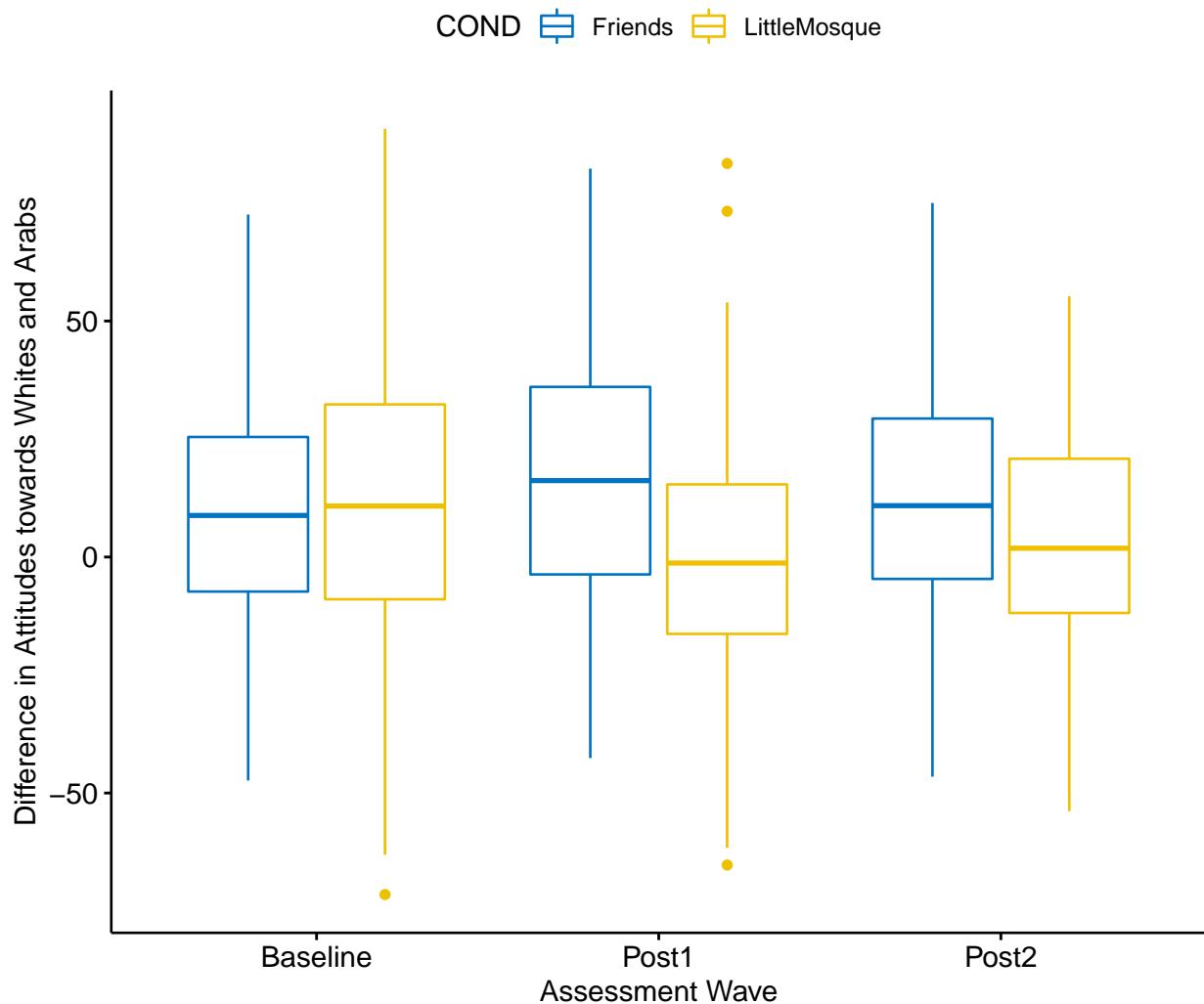
- skew: < 3; the highest skew value in our data is 0.32

- kurtosis: extreme values are between 8 and 20; the highest kurtosis value in our data is .55

The boxplot is one common way for identifying outliers. The boxplot uses the median and the lower (25th percentile) and upper (75th percentile) quartiles. The difference between Q3 and Q1 is the *interquartile range* (IQR).

You'll notice that as we are creating these boxplots we are saving them as objects. This is not necessary to produce the graph. However, we will combine the object with other data, later, to embed results if the analysis in our figures.

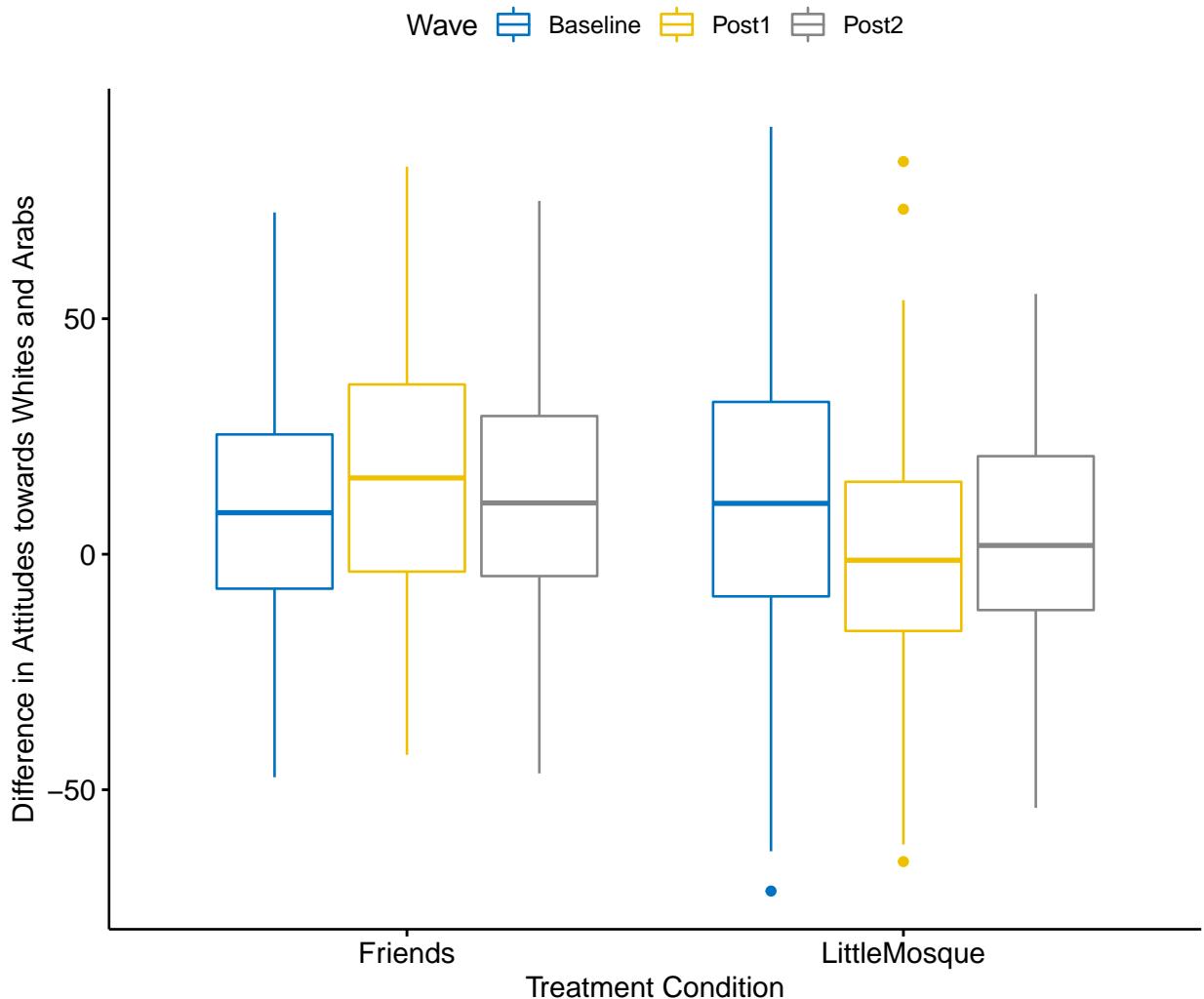
```
CNDwiWV <- ggpubr::ggboxplot(Murrar_df, x = "Wave", y = "Diff", color = "COND",
  palette = "jco", xlab = "Assessment Wave", ylab = "Difference in Attitudes towards Whites and Arabs")
CNDwiWV
```



The distributions look relatively normal with the mean well-centered. Given that we simulated the data from means and standard deviations, this is somewhat expected. This boxplot also provides a glimpse of the patterns in our data. That is, the means are quite similar at baseline; in the post intervention waves we see greater difference scores for the Friends condition.

Let's reconfigure the data by putting the wave on the X axis. Plotting it both ways (i.e., swapping roles of predictor and moderator) can help us get a sense of what is happening.

```
WVwiCND <- ggpubr::ggboxplot(Murrar_df, x = "COND", y = "Diff", color = "Wave",
  palette = "jco", xlab = "Treatment Condition", ylab = "Difference in Attitudes towards White and Arabs")
```



Outliers are generally identified when values fall outside these lower and upper boundaries. In the short formulas below, IQR is the *interquartile range* (i.e., the middle 50%, the distance of the box):

- $Q1 - 1.5 \times IQR$
- $Q3 + 1.5 \times IQR$

Extreme values occur when values fall outside these boundaries:

- $Q1 - 3 \times IQR$
- $Q3 + 3 \times IQR$

Using the `rstatix::identify_outliers` function we can look for outliers in the dependent variable, doubly grouped by our predictor variables.

```
Murrar_df %>%
  group_by(Wave, COND) %>%
  rstatix::identify_outliers(Diff)
```

	Wave	COND	rowID	caseID	AttArab	AttWhite	Diff	is.outlier	is.extreme
	<fct>	<fct>	<fct>	<fct>	<dbl>	<dbl>	<dbl>	<lgl>	<lgl>
1	Baseline	LittleMosq~	107	107	100	28.5	-71.5	TRUE	FALSE
2	Post1	LittleMosq~	297	104	16.6	100	83.4	TRUE	FALSE
3	Post1	LittleMosq~	315	122	26.8	100	73.2	TRUE	FALSE
4	Post1	LittleMosq~	337	144	97.4	32.2	-65.3	TRUE	FALSE

While we have some outliers (where “is.outlier” = “TRUE”), none are extreme (where “is.outlier” = “FALSE”). We’ll keep these in mind as we continue to evaluate the data.

If I had extreme outliers, I would individually inspect them. Especially if something looked awry (e.g., erratic responding extreme scores across variables) I might consider deleting them.

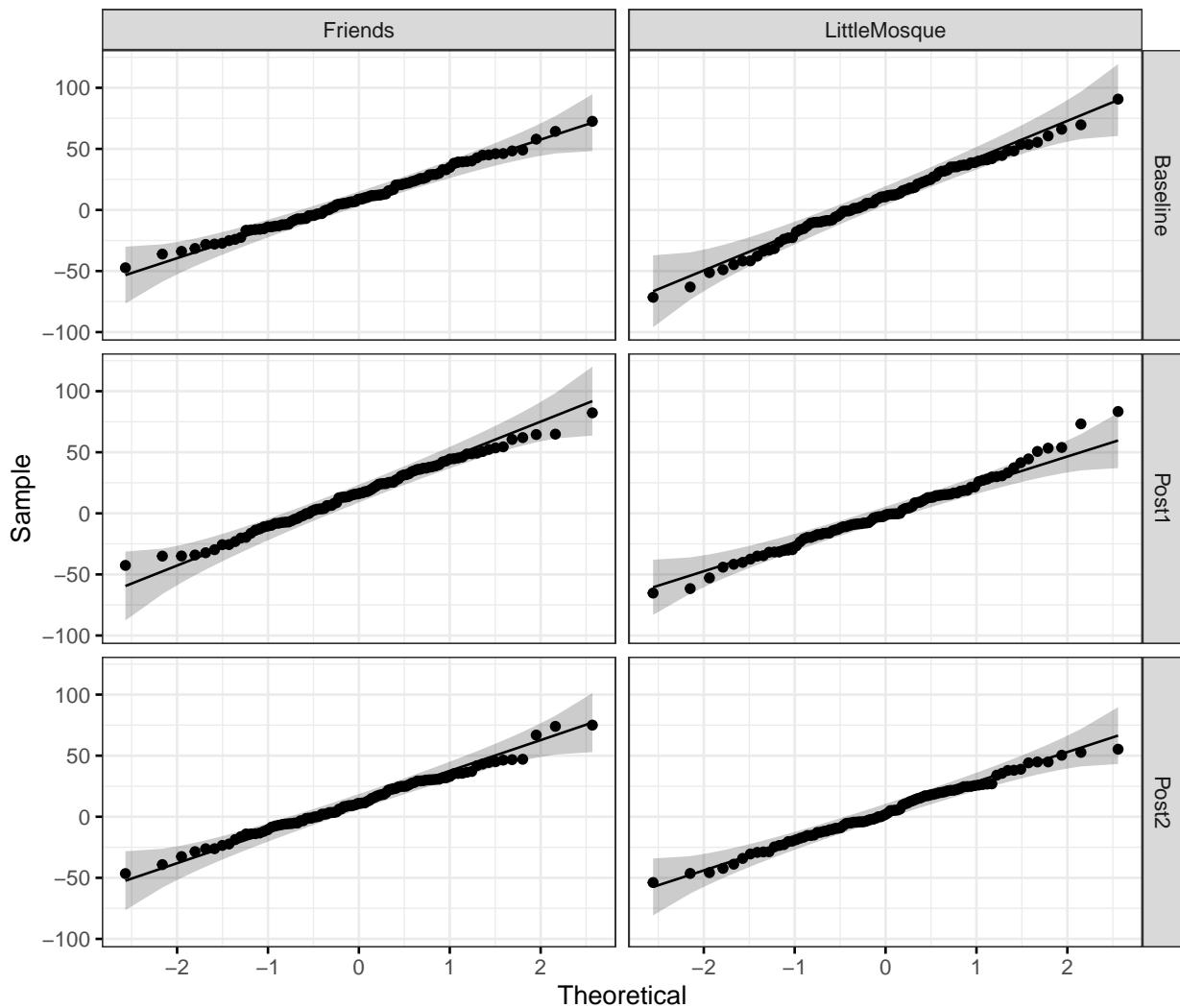
Next we can use the `rstatix::shapiro_test()` to see if any of the distributions of the dependent variable (Diff) within each wave-by-condition combinations differs significantly from a normal distribution.

```
Murrar_df %>%
  group_by(Wave, COND) %>%
  rstatix::shapiro_test(Diff)
```

	Wave	COND	variable	statistic	p
	<fct>	<fct>	<chr>	<dbl>	<dbl>
1	Baseline	Friends	Diff	0.993	0.915
2	Baseline	LittleMosque	Diff	0.993	0.923
3	Post1	Friends	Diff	0.992	0.798
4	Post1	LittleMosque	Diff	0.986	0.437
5	Post2	Friends	Diff	0.990	0.708
6	Post2	LittleMosque	Diff	0.991	0.762

The Shapiro Wilks test suggests that distribution in each of our cells is not significantly different than normal. We can further visualize this with QQ plots.

```
ggpubr::ggqqplot(Murrar_df, "Diff", ggtheme = theme_bw()) + facet_grid(Wave ~ COND)
```



7.4.1.2 Homogeneity of variance assumption

Because there is a between-subjects variable, we need to evaluate the homogeneity of variance assumption. As before, we can use the Levene's test with the `rstatix::levene_test()` function. Considering each of the comparisons of condition within wave, there is no instance where we violate the assumption.

```
Murrar_df %>%
  group_by(Wave) %>%
  rstatix::levene_test(Diff ~ COND)
```

```
# A tibble: 3 x 5
  Wave      df1    df2 statistic     p
  <fct>    <int> <int>     <dbl>   <dbl>
1 Baseline     1    191      3.97 0.0477
2 Post1        1    191      0.141 0.708
```

```
3 Post2      1   191     0.107 0.744
```

Levene's test indicated a violation of this assumption between the Friends and Little Mosque conditions at baseline ($F [1, 191] = 3.973, p = .047$). However, there was no indication of assumption violation at post1 ($F [1, 191] = 0.141, p = .708$), and post2 ($F [1, 191] = 0.107, p = .743$) waves of the design.

7.4.1.3 Assumption of homogeneity of covariance matrices

In this multivariate sample, the Box's M test evaluates if two or more covariance matrices are homogeneous. Like other tests of assumptions, we want a non-significant test result (i.e., where $p > .05$). Box's M has some disadvantages. Box's M has low power in small sample sizes and is overly sensitive in large sample sizes. We would unlikely make a decision about our data with Box's M alone. Rather, we consider it along with our dashboard of diagnostic screeners.

```
rstatix::box_m(Murrar_df[, "Diff", drop = FALSE], Murrar_df$COND)
```

```
# A tibble: 1 x 4
  statistic p.value parameter method
  <dbl>    <dbl>     <dbl> <chr>
1       3.21  0.0732      1 Box's M-test for Homogeneity of Covariance Matric~
```

None-the-less, Box's M indicated no violation of the homogeneity of covariance matrices assumption ($M = 3.209, p = .073$)

7.4.1.4 APA style writeup of assumptions

At this stage we are ready to draft the portion of the APA style writeup that evaluates the assumptions.

Mixed design ANOVA has a number of assumptions related to both the within-subjects and between-subjects elements. Data are expected to be normally distributed at each level of design. Visual inspection of boxplots for each wave of the design, assisted by the `rstatix::identify_outliers()` function (which reports values above $Q3 + 1.5 \times IQR$ or below $Q1 - 1.5 \times IQR$, where IQR is the interquartile range) indicated some outliers, but none at the extreme level. There was no evidence of skew (all values were at or below the absolute value of 0.32) or kurtosis (all values were below the absolute value of .57; [Kline, 2016]). Additionally, the Shapiro-Wilk tests applied at each level of the design were non-significant. Because of the between-subjects aspect of the design, the homogeneity of variance assumption was evaluated. Levene's test indicated a violation of this assumption between the Friends and Little Mosque conditions at baseline $F [1, 191] = 3.973, p = .047$. However, there was no indication of assumption violation at post1 ($F [1, 191] = 0.141, p = .708$), and post2 ($F [1, 191] = 0.107, p = .743$) waves of the design. Further, Box's M-test ($M = 3.209, p = .073$) indicated no violation of the homogeneity of covariance matrices. *LATER WE WILL ADD INFORMATION ABOUT THE SPHERICITY ASSUMPTION.*

7.4.2 Omnibus ANOVA

Having evaluated the assumptions (excepting sphericity) we are ready to move to the evaluation of the omnibus ANOVA. This next step produces both the omnibus test as well as testing the sphericity assumption. Conceptually, evaluating the sphericity assumption precedes the omnibus; procedurally these are evaluated simultaneously. The figure also reflects that decisions related to follow-up are dependent upon the significance of the main and omnibus effects.

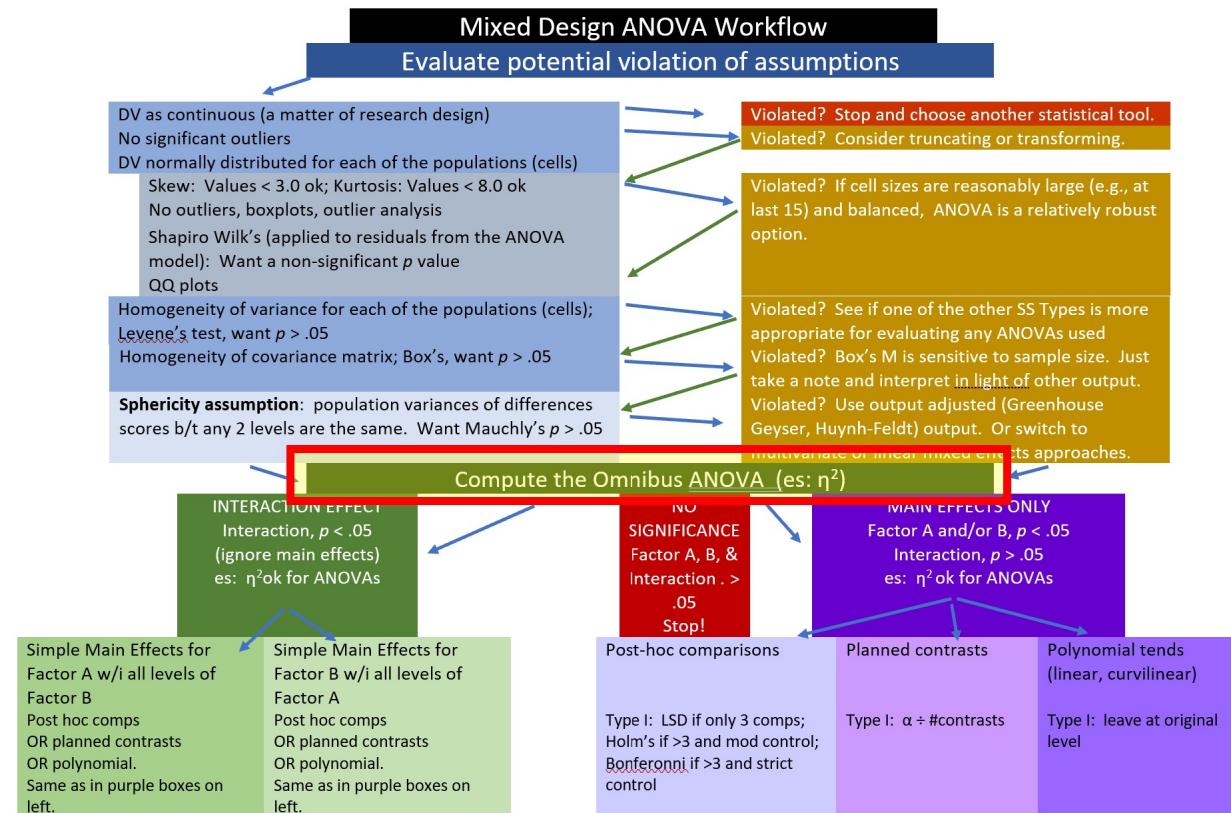


Figure 7.4: Image of the workflow showing that we are at the “Compute the Omnibus ANOVA” step

The *rstatix* package is a wrapper for the *car* package. Authors of *wrappers* attempt to streamline a more complex program to simplify the input needed and maximize the output produced for the typical use-cases.

If we are ever confused about a function, we can place a question mark in front of it. It will summons information and, if the package is in our library, let us know to which package it belongs and open the instructions that are embedded in R/R Studio.

```
#?anova_test
```

In the code below the identification of the data, DV, between, and within variables are likely to be intuitive. The within-subjects identifier (*wid*) is the person-level ID that assists the statistic in controlling for the dependency introduced by the repeated-measures factor.

```
# Murrar_df is our df, Diff is our df, wid is the caseID between is
# the between-subjects variable, within is the within subjects
# variable
Diff_2way <- rstatix::anova_test(data = Murrar_df, dv = Diff, wid = caseID,
  between = COND, within = Wave)
Diff_2way
```

ANOVA Table (type III tests)

\$ANOVA

	Effect	DFn	DFd	F	p	p<.05	ges
1	COND	1	191	13.149	0.000369	*	0.023000
2	Wave	2	382	0.273	0.761000		0.000933
3	COND:Wave	2	382	5.008	0.007000	*	0.017000

\$`Mauchly's Test for Sphericity`

	Effect	W	p	p<.05
1	Wave	0.99	0.369	
2	COND:Wave	0.99	0.369	

\$`Sphericity Corrections`

	Effect	GGe	DF[GG]	p[GG]	p[GG]<.05	HFe	DF[HF]	p[HF]	p[HF]<.05
1	Wave	0.99	1.98,	378.06	0.759		1	2,	382 0.761
2	COND:Wave	0.99	1.98,	378.06	0.007	*	1	2,	382 0.007 *

7.4.2.1 Checking the sphericity assumption

First, we check Mauchly's test for the main and interaction effects that involve the repeated measures variable.

- main effect for Wave: $W = .99$, $p = .369$
- main effect for Wave: $W = .99$, $p = .369$

We will be able to add this statement to our assumptions write-up: Mauchly's test indicated no violation of the sphericity assumption for the main effect ($W = 0.99$, $p = .369$) and interaction effect ($W = 0.99$, $p = .369$).

If the p value associated with Mauchly's test had been less than .05, we could have used one of the two options (Greenhouse Geyser/GGe or Huynh-Feldt/HFe). In each of these an epsilon value provides an adjustment to the degrees of freedom used in the estimation of the p value. There is also an option to use a multivariate approach when ANOVA designs include a repeated measures factor.

Omnibus Results

Results of the omnibus ANOVA indicated a significant main effect for condition ($F[1, 191] = 13.149$, $p < .001$, $\eta^2 = 0.023$), a non-significant main effect for wave ($F[2, 382] = 0.273$, $p = .761$, $\eta^2 = 0.001$), and a significant interaction effect ($F[2, 382] = 5.008$, $p = 0.007$, $\eta^2 = 0.017$). We note that

according to Cohen et al.'s [Cohen et al., 2003] guidelines, the effect size for the interaction term is small.

In the output, the column labeled “ges” provides the value for the effect size, η^2 . Recall that *eta-squared* is one of the most commonly used measures of effect. It refers to the proportion of variability in the dependent variable/outcome that can be explained in terms of the independent variable/predictor. Traditional interpretive values are similar to the Pearson's *r*:

- 0 = no relationship
- .02 = small
- .13 = medium
- .26 = large
- 1 = a perfect (one-to-one) correspondence

The effect size for our interaction effect (0.017) is small.

With a significant interaction effect, we would focus on interpreting one or both of the simple main effects. Let's first look at the simple main effect of condition within wave option.

7.4.3 Simple main effect of condition within wave

The figure reflects our path in the workflow. In the presence of a significant interaction effect we could choose from a variety of follow-up tests.

If we take this option we follow up with 3 one-way ANOVAs. When we look at condition within wave, our ANOVAs will look like this:

- comparison of Friends and Little Mosque within the baseline wave
- comparison of Friends and Little Mosque within the post1 wave
- comparison of Friends and Little Mosque within the post2 wave

```
# create an object to hold the output the group_by function is what
# results in three, one-way ANOVAs for each of the waves, separately
# the between = Cond means that each level of cond will be compared
# method - 'bonferroni' gets us both the standard and adjusted p
# values
SimpleWave <- Murrar_df %>%
  group_by(Wave) %>%
  rstatix::anova_test(dv = Diff, wid = caseID, between = COND) %>%
  rstatix::get_anova_table() %>%
  rstatix::adjust_pvalue(method = "bonferroni")
```

```
Coefficient covariances computed by hccm()
Coefficient covariances computed by hccm()
Coefficient covariances computed by hccm()
```

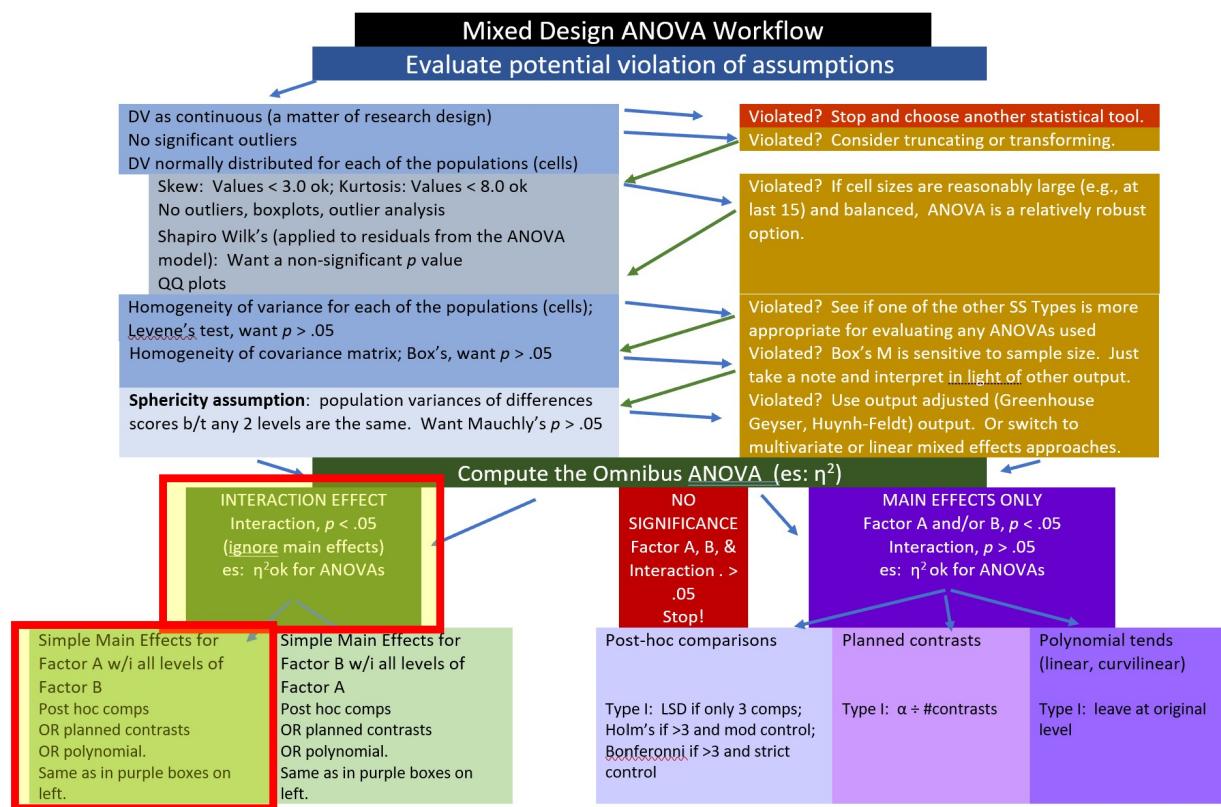


Figure 7.5: Image of the workflow showing that we are at the “Simple Main Effects for Factor A within all levels of Factor B” step

SimpleWave

	Wave	Effect	DFn	DFd	F	p `p < .05`	ges	p.adj
*	<fct>	<chr>	<dbl>	<dbl>	<dbl>	<chr>	<dbl>	<dbl>
1	Baseline	COND	1	191	0.012	0.914	""	0.0000614
2	Post1	COND	1	191	17.5	0.0000438	"*"	0.084
3	Post2	COND	1	191	5.99	0.015	"*"	0.03

In prior lectures we have adjusted the p value against which we compare the resulting p value. When we specify “bonferroni” on the `adjust_pvalue()` command, the algorithm adjusts the reported p value for us. We can see the unadjusted p value in the “ $p < .05$ ” column and the Bonferroni adjustment in the “ $p.adj$ ” column.

I think that it will be easiest for us to interpret this simple main effect as the traditional $p < .05$ and then apply the restrictions to the alpha at the next level of analysis. In this particular instance, we would have statistically significant differences somewhere between the Friends and Little Mosque conditions for both the Post ($p = .027$) and Post2 ($p = .010$) waves.

F strings:

- Pre: $F(1, 191) = 0.012, p = .914, \eta^2 = 0.000$ (the effect size is zero)
- Post: $F(1, 191) = 17.497, p < .001, \eta^2 = 0.084$ (approaching a moderate effect size)
- Post2: $F(1, 191) = 5.994, p = .015, \eta^2 = 0.030$ (a small effect size)

Recall, interpretation for the eta-squared are $.02 \sim$ small, $.13 \sim$ medium, and $>.26 \sim$ large

Because there are only two levels (Friends, Little Mosque) within each wave (baseline, post1, post2), this simple effects analysis is complete with the three pairwise comparisons.

As always, we have several choices about how to manage Type I error. In a circumstance when the analysis of simple main effects (condition within wave) includes only three pairwise comparisons, we can use the LSD method [Green and Salkind, 2014b]. This means that we can leave the alpha at 0.05. If we were to use a traditional Bonferroni, we would use $\alpha = .017$ (.05/3). Although the more restrictive Bonferroni criteria comes close, in both cases we would still have one non-significant(baseline) and two significant (post1, post2) simple main effects.

.05/3

[1] 0.01666667

If we were to write up this result:

We followed the significant interaction effect with an evaluation of simple main effects of condition within wave. Because there were only three comparisons following the omnibus evaluation, we used the LSD method to control for Type I error and left the alpha at .05 [Green and Salkind, 2014b]. There was a non-statistically significant difference between conditions at baseline: $F(1, 191) = 0.012, p = .914, \eta^2 = 0.000$. However other were statistically significant differences at post1 ($F[1, 191] = 17.497, p < .001, \eta^2 = 0.084$) and post2 ($F[1, 191] = 5.994, p = .015, \eta^2 = 0.030$). We note that the effect size at post1 approached a moderate size; the effect size at post2 was small.

7.4.4 Simple main effect of wave within condition

Alternatively, we could evaluate the simple main effect of wave within condition. The figure reflects our path along the workflow.

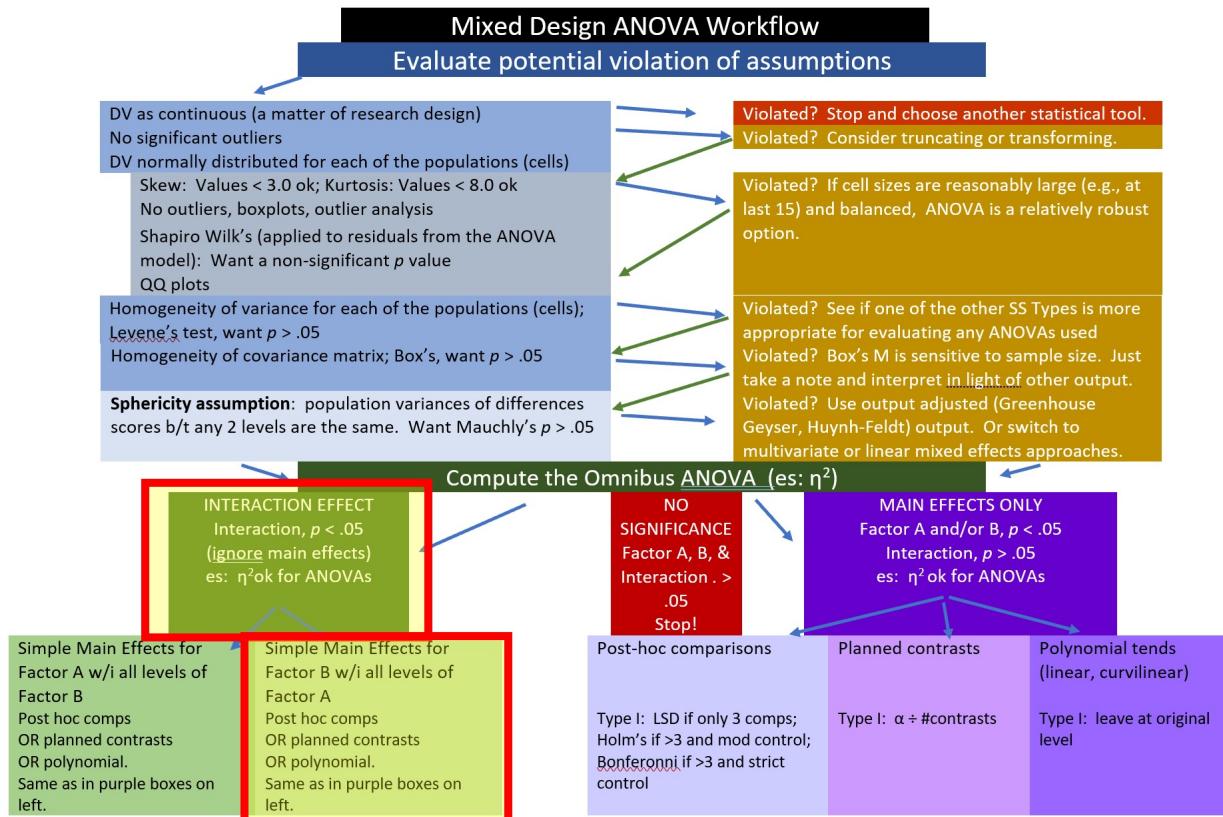


Figure 7.6: Image of the workflow showing that we are at the “Simple Main Effects for Factor B within all levels of Factor A” step

If we conducted this alternative we would start with three one-way ANOVAs and then follow each of those with pairwise comparisons. First, the one-way repeated measures ANOVAs:

- comparison of baseline, post1, and post2 within the Friends condition
- comparison of baseline, post1, and post2 within the Little Mosque condition

```
SimpleCond <- Murrar_df %>%
  group_by(COND) %>%
  rstatix::anova_test(dv = Diff, wid = caseID, within = Wave) %>%
  rstatix::get_anova_table() %>%
  rstatix::adjust_pvalue(method = "bonferroni")
SimpleCond
```

```
# A tibble: 2 x 9
  COND      Effect    DFn     DFd      F      p `p<.05`    ges p.adj
  <fct>    <fct>    <dbl>    <dbl>    <dbl>    <dbl>    <dbl>    <dbl>
```

1	Friends	Wave	2	194	1.76	0.175	""	0.012	0.35	
2	LittleMosque	Wave	2	188	3.39	0.036	"*"	0.022	0.072	

Below are the F strings for the one-way ANOVAs the followed the omnibus, mixed design, ANOVA:

- Friends: $F(2, 194) = 1.759, p = 0.175, \eta^2 = 0.012$ (effect size indicates no relationship)
- Little Mosque: $F(2, 188) = 3.392, p = 0.036, \eta^2 = 0.072$ (a small-to-moderate effect size)

Because each of these one-way ANOVAs has three levels, we need to follow with pairwise comparisons. However, we only need to conduct them for the Little Mosque condition. As you can see we generally work our way down to comparing chunks to each other to find the source(s) of significant differences.

You will notice that we are saving the results of the pairwise comparisons as an object. This is not necessary, however we can use this object in combination with the boxplot we created earlier to embed results of our analysis in the resulting figure.

```
pwcWVwiGP <- Murrar_df %>%
  group_by(COND) %>%
  rstatix::pairwise_t_test(Diff ~ Wave, paired = TRUE, detailed = TRUE,
                           p.adjust.method = "bonferroni") # %>%
# select(-df, -statistic, -p) # Remove details
pwcWVwiGP
```

```
# A tibble: 6 x 16
  COND   estimate .y.   group1 group2     n1     n2 statistic      p     df conf.low
* <fct>    <dbl> <chr> <chr> <chr> <int> <int>    <dbl> <dbl> <dbl>    <dbl>
1 Frien~   -6.62 Diff  Basel~ Post1     98     98    -1.80  0.075    97   -13.9
2 Frien~   -2.65 Diff  Basel~ Post2     98     98    -0.793 0.43     97   -9.27
3 Frien~    3.97 Diff  Post1  Post2     98     98     1.09  0.276    97   -3.23
4 Littl~    9.88 Diff  Basel~ Post1     95     95     2.45  0.016    94    1.86
5 Littl~    6.06 Diff  Basel~ Post2     95     95     1.62  0.108    94   -1.36
6 Littl~   -3.82 Diff  Post1  Post2     95     95    -1.03  0.304    94   -11.2
# ... with 5 more variables: conf.high <dbl>, method <chr>, alternative <chr>,
#   p.adj <dbl>, p.adj.signif <chr>
```

At this point, we likely need to control for Type I error. Why? We have already conducted two one-way ANOVAs after the omnibus. Now we will conduct three more pairwise comparisons in the Little Mosque condition. I would divide $.05/3$ and interpret these pairwise comparisons with an alpha of $.017$.

We find a significant difference between baseline and post1 ($t[95] = 2.447, p = .016$), but non-significant differences between baseline and post2 ($t[95] = 1.621, p = .108$) and post1 and post2 ($t[95] = -1.034, p = .304$)

If we were to write up this result:

We followed the significant interaction effect with an evaluation of simple main effects of wave within condition. There were non-significant difference within the Friends condition ($F [2, 194] = 1.759, p = 0.175, \eta^2 = 0.012$). There were significant differences with an effect size indicating a small-to-moderate effect in the Little Mosque condition ($F [2, 188] = 3.392, p = 0.036, \eta^2 = 0.072$). We followed up the significant simple main effect for the Little Mosque condition with pairwise comparisons. At this level we controlled for Type I error by dividing alpha (.05) by the number of paired comparisons (3). We found a significant difference between baseline and post1 ($t[95] = 2.447, p = .016$), but non-significant differences between baseline and post2 ($t[95] = 1.621, p = .108$) and post1 and post2 ($t[95] = -1.034, p = .304$).

7.4.5 If we only had a main effect

When there is an interaction effect, we do not interpret main effects. This is because the solution is more complicated than a main effect could explain. It is important, though, to know how to interpret a main effect. We would do this if we had one or more significant main effects and no interaction effect.

The figure shows our place on the workflow.

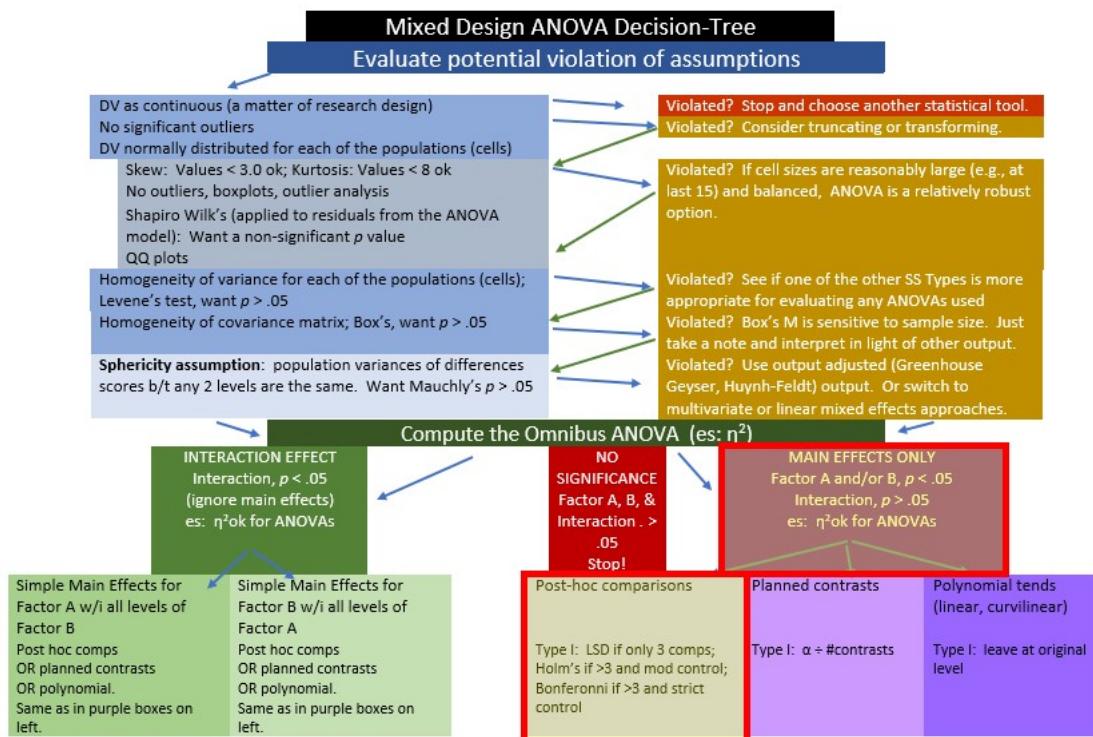


Figure 7.7: Image of a workflow showing that we are at the “Main effects only” step

If we had not had a significant interaction, but did have a significant main effect for wave, we could have conducted pairwise comparisons for pre, post1, and post2 – collapsing across condition.

```
Murrar_df %>%
  rstatix::pairwise_t_test(Diff ~ Wave, paired = TRUE, p.adjust.method = "bonferroni")
```

```
# A tibble: 3 x 10
  .y.   group1   group2     n1     n2 statistic     df      p p.adj p.adj.signif
* <chr> <chr>    <chr> <int> <int>     <dbl> <dbl> <dbl> <dbl> <chr>
1 Diff  Baseline Post1     193     193     0.539     192  0.59      1 ns
2 Diff  Baseline Post2     193     193     0.652     192  0.515     1 ns
3 Diff  Post1    Post2     193     193     0.0528    192  0.958     1 ns
```

Ignoring condition (Friends, Little Mosque), we do not see changes across time. This is not surprising since the F test for the main effect was also non-significant ($F[2, 382] = 0.273, p = .761, \eta^2 = 0.0014$),

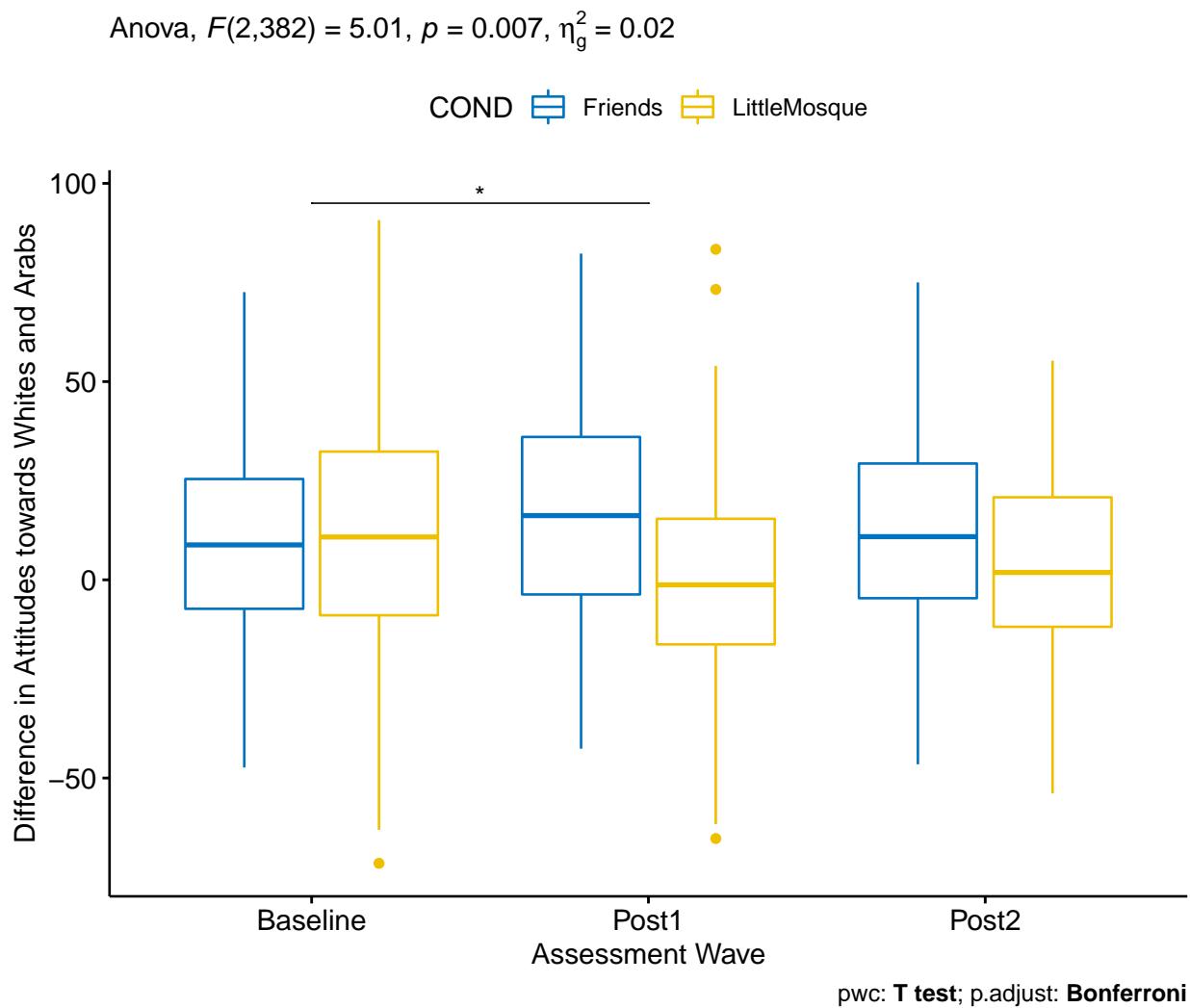
If we had had a non-significant interaction effect but a significant main effect for condition, there would have been no need for further follow-up. Why? Because there were only two levels the significant main effect already tells us there were statistically significant differences between Friends and Little Mosque ($F[1, 191] = 13.149, p < .001, \eta^2 = 0.023$).

7.4.6 APA Style Write-up of the Results

Recall that earlier in this lesson we save objects for the boxplots (e.g., CNDwiWV) and the pairwise comparisons (e.g., pwcVWwiGP). The script below use the objects created from omnibus ANOVA and the pairwise comparisons to add results to the figure. Depending on where you are presenting your results, these may be useful.

This first figure would pair well if you report the simple main effect of condition within wave.

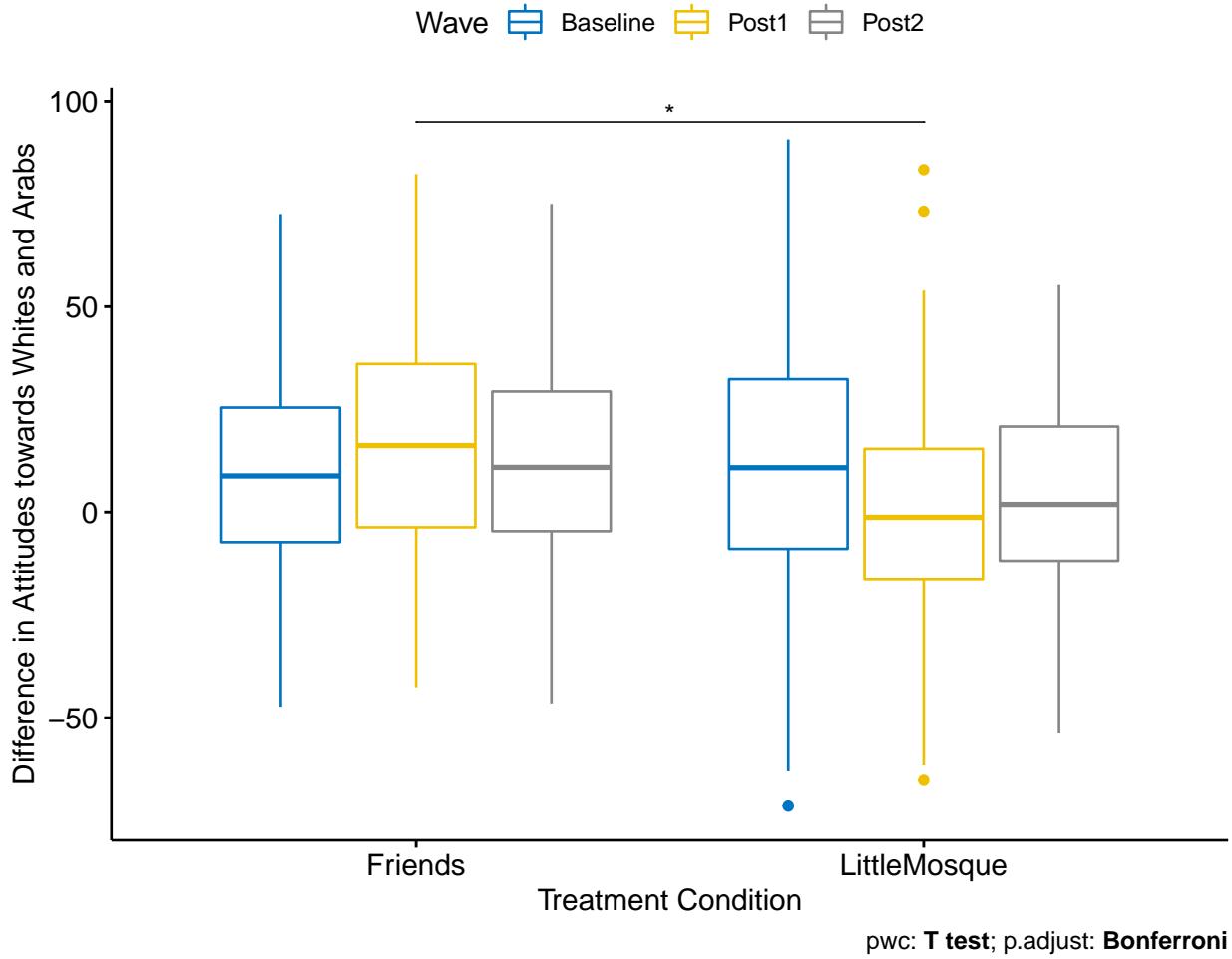
```
pwcVWwiGP <- pwcVWwiGP %>%
  rstatix::add_xy_position(x = "Wave")
CNDwiWV + ggpubr::stat_pvalue_manual(pwcVWwiGP, tip.length = 0, hide.ns = TRUE) +
  labs(subtitle = rstatix::get_test_label(Diff_2way, detailed = TRUE),
       caption = rstatix::get_pwc_label(pwcVWwiGP))
```



This second figure would pair well with the results that reported the simple main effect of wave within condition.

```
# pwcWVwiGP were my pairwise comparisons for the simple effect
# UE_2way was my omnibus ANOVA object WVwiCND was the boxplot before
# I did the ANOVA
pwcWVwiGP <- pwcWVwiGP %>%
  rstatix::add_xy_position(x = "Wave")
WVwiCND + ggpubr::stat_pvalue_manual(pwcWVwiGP, tip.length = 0, hide.ns = TRUE) +
  labs(subtitle = rstatix::get_test_label(Diff_2way, detailed = TRUE),
       caption = rstatix::get_pwc_label(pwcWVwiGP))
```

Anova, $F(2,382) = 5.01, p = 0.007, \eta^2_g = 0.02$



7.4.6.1 Results

We conducted a 2 X 3 mixed design ANOVA to evaluate the combined effects of condition (Friends and Little Mosque) and wave (baseline, post1, post2) on a difference score that compared attitudes toward White and Arab people.

Mixed design ANOVA has a number of assumptions related to both the within-subjects and between-subjects elements. Data are expected to be normally distributed at each level of design. Visual inspection of boxplots for each wave of the design, assisted by the `rstatix::identify_outliers()` function (which reports values above $Q3 + 1.5 \times IQR$ or below $Q1 - 1.5 \times IQR$, where IQR is the interquartile range) indicated some outliers, but none at the extreme level. There was no evidence of skew (all values were at or below the absolute value of 0.32) or kurtosis (all values were below the absolute value of .57; [Kline, 2016]). Additionally, the Shapiro-Wilk tests applied at each level of the design were non-significant. Because of the between-subjects aspect of the design, the homogeneity of variance assumption was evaluated. Levene's test indicated a violation of this assumption between the Friends and Little Mosque conditions at baseline $F[1, 191] = 3.973, p = .047$. However, there was no indication of assumption violation at post1 ($F[1, 191] = 0.141, p =$

.708), and post2 ($F[1, 191] = 0.107, p = .743$) waves of the design. Further, Box's M-test ($M = 3.209, p = .073$) indicated no violation of the homogeneity of covariance matrices. Mauchly's test indicated no violation of the sphericity assumption for the main effect ($W = 0.99, p = .369$) and interaction effect ($W = 0.99, p = .369$).

Results of the omnibus ANOVA indicated a significant main effect for condition ($F[1, 191] = 13.149, p < .001, \eta^2 = 0.023$), a non-significant main effect for wave ($F[2, 382] = 0.273, p = .761, \eta^2 = 0.001$), and a significant interaction effect ($F[2, 382] = 5.008, p = 0.007, \eta^2 = 0.017$).

We followed the significant interaction effect with an evaluation of simple main effects of wave within condition. There were non-significant difference within the Friends condition ($F[2, 194] = 1.759, p = 0.175, \eta^2 = 0.012$). There were significant differences with an effect size indicating a small-to-moderate effect in the Little Mosque condition ($F[2, 188] = 3.392, p = 0.036, \eta^2 = 0.072$). We followed up the significant simple main effect for the Little Mosque condition with pairwise comparisons. At this level we controlled for Type I error by dividing alpha (.05) by the number of paired comparisons (3). We found a significant difference between baseline and post1 ($t[95] = 2.447, p = .016$), but non-significant differences between baseline and post2 ($t[95] = 1.621, p = .108$) and post1 and post2 ($t[95] = -1.034, p = .304$).

As illustrated in Figure 1 difference scores were comparable at baseline. After the intervention, difference scores increased for those in the Friends condition – indicating more favorable attitudes toward White people. In contrast, those exposed to the Little Mosque condition had difference scores that were lower. Means and standard deviations are reported in Table 1.

The following code can be used to write output to .csv files. From there it is easy(er) to manipulate them into tables for use in an empirical manuscript.

```
MASS::write.matrix(pwcWVwiGP, sep = ",", file = "pwcWVwiGP.csv")
# this command can also be used to export other output
MASS::write.matrix(Diff_2way$ANOVA, sep = ",", file = "Diff_2way.csv")
MASS::write.matrix(SimpleWave, sep = ",", file = "SimpleWave.csv")
MASS::write.matrix(SimpleCond, sep = ",", file = "SimpleCond.csv")
```

7.4.6.2 Comparing our findings to Murrar and Brauer [2018]

In general, the results of our simulation mapped onto the findings. If you have access to the article I encourage you to examine it as you consider my observations.

- The authors started their primary analyses of Experiment 1 with independent t tests comparing the Friends and Little Mosque conditions within each of the baseline, post1, and post2 waves. This is equivalent to our simple main effects of condition within wave that we conducted as follow-up to the significant interaction effect. It is not clear to me why they did not precede this with a mixed design ANOVA.
 - The results of the article are presented in their Table 1
 - Our results were comparable in that we found no attitude difference at baseline
 - Similar to the results in the article we found statistically significant differences (with comparable p values and effect sizes) at post1 and post2

- With two experiments (each with a number of associated hypotheses) in a single paper there were a large number of analyses conducted by the authors. I think they designed tables and figures that provided an efficient and clear review of the study design and their findings.
- This finding is exciting to me. Anti-racism education frequently encourages individuals to expose themselves to content authored/created by individuals from groups with marginalized identities. This finding supports that approach to prejudice reduction.

7.5 Power in Mixed Design ANOVA

The package `wp.rmanova` was designed for power analysis in repeated measures ANOVA.

Power analysis allows us to determine the probability of detecting an effect of a given size with a given level of confidence. Especially when we don't achieve significance, we may want to stop.

In the *WebPower* package, we specify 6 of 7 interrelated elements; the package computes the missing element

- n = sample size (number of individuals in the whole study)
- ng = number of groups
- nm = number of repeated measurements (i.e., waves)
- f = Cohen's f (an effect size; we can use a conversion calculator); Cohen suggests that f values of 0.1, 0.25, and 0.4 represent small, medium, and large effect sizes, respectively
- $nscor$ = the Greenhouse Geiser correction from our output; 1.0 means no correction was needed and is the package's default; < 1 means some correction was applied
- $alpha$ = is the probability of Type I error; we traditionally set this at .05
- $power$ = $1 - P(\text{Type II error})$ we traditionally set this at .80 (so anything less is less than what we want)
- $type$ = 0 is for between-subjects, 1 is for repeated measures, 2 is for interaction effect; in a mixed design ANOVA we will select "2"

As in the prior lessons, we need to convert our effect size for the *interaction* to f effect size (this is not the same as the F test). The *effectsize* package has a series of converters. We can use the `eta2_to_f()` function to translate the η^2 associated with the interaction effect to Cohen's f .

```
#interaction effect
effectsize::eta2_to_f(0.017)
```

```
[1] 0.1315066
```

We can now retrieve information from our study (including the Cohen's f value we just calculated) and insert it into the script for the power analysis.

```
WebPower::wp.rmanova(n=193, ng=2, nm=3, f = .1315, nscor = .99, alpha = .05, power = NULL, type = 2)
```

Repeated-measures ANOVA analysis

n	f	ng	nm	nscor	alpha	power
193	0.1315	2	3	0.99	0.05	0.3493183

NOTE: Power analysis for interaction-effect test

URL: <http://psychstat.org/rmanova>

We are powered at .349 (we have a 35% of rejecting the null hypothesis, if it is true)

In reverse, setting *power* at .80 (the traditional value) and changing *n* to *NULL* yields a recommended sample size.

```
WebPower::wp.ranova(n = NULL, ng = 2, nm = 3, f = 0.1315, nscor = 0.99,
alpha = 0.05, power = 0.8, type = 2)
```

Repeated-measures ANOVA analysis

n	f	ng	nm	nscor	alpha	power
562.608	0.1315	2	3	0.99	0.05	0.8

NOTE: Power analysis for interaction-effect test

URL: <http://psychstat.org/rmanova>

Given our desire for strong power and our weak effect size, this power analysis suggests a sample size of 562 participants to detect a significant interaction effect.

7.6 Practice Problems

In each of these lessons I provide suggestions for practice that allow you to select one or more problems that offer differing levels of difficulty. Whichever you choose, you will focus on these larger steps in one-way ANOVA, including:

- test the statistical assumptions
- conduct a two-way (minimally a 2x3), mixed design, ANOVA, including
 - omnibus test and effect size
 - report main and interaction effects
 - conduct follow-up testing of simple main effects
- write a results section to include a figure and tables

7.6.1 Problem #1: Play around with this simulation.

Copy the script for the simulation and then change (at least) one thing in the simulation to see how it impacts the results.

- If mixed design ANOVA is new to you, perhaps you just change the number in “set.seed(210813)” from 210813 to something else. Your results should parallel those obtained in the lecture, making it easier for you to check your work as you go.
- If you are interested in power, change the sample size to something larger or smaller.
- If you are interested in variability (i.e., the homogeneity of variance assumption), perhaps you change the standard deviations in a way that violates the assumption.

7.6.2 Problem #2: Conduct a one-way ANOVA with a different dependent variable.

The Murrar et al. [2018] article has three dependent variables (attitudes toward people who are Arab, attitudes toward people who are White, and the difference score). I analyzed the difference score. Select one of the other dependent variables. If you do not get a significant interaction, play around with the simulation (changing the sample size, standard deviations, or both) until you get a significant interaction effect.

7.6.3 Problem #3: Try something entirely new.

Using data for which you have permission and access (e.g., IRB approved data you have collected or from your lab; data you simulate from a published article; data from an open science repository; data from other chapters in this OER), complete a mixed design ANOVA. Please have at least 3 levels for one predictor and at least 2 levels for the second predictor.

7.6.4 Grading Rubric

Regardless which option(s) you chose, use the elements in the grading rubric to guide you through the practice.

Assignment Component	Points Possible	Points Earned
1. Narrate the research vignette, describing the IV and DV	5	_____
2. Simulate (or import) and format data	5	_____
3. Evaluate statistical assumptions	5	_____
4. Conduct omnibus ANOVA (w effect size)	5	_____
5. Conduct one set of follow-up tests; narrate your choice	5	_____
6. Describe approach for managing Type I error	5	_____
7. APA style results with table(s) and figure	5	_____
8 Explanation to grader	5	_____
Totals	40	_____

Chapter 8

Analysis of Covariance

[Screencasted Lecture Link](#)

The focus of this lecture is analysis of covariance. Sticking with the same research vignette as we used for the mixed design ANOVA, we rearrange the variables a bit to see how they work in an ANCOVA design. The results help clarify the distinction between *moderator* and *covariate*.

8.1 Navigating this Lesson

There is about just about an hour of lecture. If you work through the materials with me, plan for an additional hour or two

While the majority of R objects and data you will need are created within the R script that sources the chapter, occasionally there are some that cannot be created from within the R framework. Additionally, sometimes links fail. All original materials are provided at the [Github site](#) that hosts the book. More detailed guidelines for ways to access all these materials are provided in the OER's [introduction](#)

8.1.1 Learning Objectives

Learning objectives from this lecture include the following:

- Define a *covariate* and distinguish it from a *moderator*.
- Recognize the case where ANCOVA is a defensible statistical approach for analyzing the data.
- Name and test the assumptions underlying ANCOVA.
- Analyze, interpret, and write up results for ANCOVA.
- List the conditions that are prerequisite for the appropriate use of a covariate or control variable.

8.1.2 Planning for Practice

In each of these lessons I provide suggestions for practice that allow you to select one or more problems that are graded in difficulty. The least complex is to change the random seed and rework

the problem demonstrated in the lesson. The results *should* map onto the ones obtained in the lecture.

The second option comes from the research vignette. For this ANCOVA article, I take a lot of liberties with the variables and research design. You could further mix and match for a different ANCOVA constellation.

As a third option, you are welcome to use data to which you have access and is suitable for ANCOVA. In either case the practice options suggest that you:

- test the statistical assumptions
- conduct an ANCOVA, including
 - omnibus test and effect size
 - report main effects and engage in any follow-up testing
 - interpret results in light of the role of the second predictor variable as a *covariate* (as opposed to the moderating role in the prior lessons)
- write a results section to include a figure and tables

8.1.3 Readings & Resources

In preparing this chapter, I drew heavily from the following resource(s). Other resources are cited (when possible, linked) in the text with complete citations in the reference list.

- Green, S. B., & Salkind, N. J. (2014). One-Way Analysis of Covariance (Lesson 27). In *Using SPSS for Windows and Macintosh: Analyzing and understanding data* (Seventh edition., pp. 151–160). Boston: Pearson. OR
 - This lesson provides an excellent review of ANCOVA with examples of APA style write-ups. The downside is that it is written for use in SPSS.
- ANCOVA in R: The Ultimate Practical Guide. (n.d.). Retrieved from <https://www.datanovia.com/en/lessons/ancova-in-r/>
 - This is the workflow we are using for the lecture and written specifically for R.
- Bernerth, J. B., & Aguinis, H. (2016). A critical review and best-practice recommendations for control variable usage. *Personnel Psychology*, 69(1), 229–283. <https://doi.org/10.1111/peps.12103>
 - An article from the industrial-organizational psychology world. Especially relevant for this lesson is the flowchart on page 273 and the discussion (pp. 270 to the end).
- Murrar, S., & Brauer, M. (2018). Entertainment-education effectively reduces prejudice. *Group Processes & Intergroup Relations*, 21(7), 1053–1077. <https://doi.org/10.1177/1368430216682350>
- This article is the source of our research vignette. I used this same article in the lesson on **mixed design ANOVA**. Swapping variable roles can be useful in demonstrating how ANCOVA is different than mixed design ANOVA.

8.1.4 Packages

The packages used in this lesson are embedded in this code. When the hashtags are removed, the script below will (a) check to see if the following packages are installed on your computer and, if not (b) install them.

```
# used to convert data from long to wide
# if(!require(reshape2)){install.packages('reshape2')}
# if(!require(broom)){install.packages('broom')}
# if(!require(tidyverse)){install.packages('tidyverse')}
# if(!require(psych)){install.packages('psych')} easy plots
# if(!require(ggpubr)){install.packages('ggpubr')} pipe-friendly R
# functions if(!require(rstatix)){install.packages('rstatix')} export
# objects for table making
# if(!require(MASS)){install.packages('MASS')}
# if(!require(knitr)){install.packages('knitr')}
# if(!require(dplyr)){install.packages('dplyr')}
# if(!require(apaTables)){install.packages('apaTables')}
```

8.2 Introducing Analysis of Covariance (ANCOVA)

Analysis of covariance (ANCOVA) evaluates the null hypothesis that

- population means on a dependent variable are equal across levels of a factor(s) adjusting for differences on a covariate(s); stated differently -
- the population adjusted means are equal across groups

This lecture introduces a distinction between **moderators** and **covariates**.

Moderator: a variable that changes the strength or direction of an effect between two variables X (predictor, independent variable) and Y (criterion, dependent variable).

Covariate: an observed, continuous variable, that (when used properly) has a relationship with the dependent variable. It is included in the analysis, as a predictor, so that the predictive relationship between the independent (IV) and dependent (DV) are adjusted.

Understanding this difference may be facilitated by understanding one of the assumptions of ANCOVA – that the slopes relating the covariate to the dependent variable are the same for all groups (i.e., the homogeneity-of-slopes assumption). If this assumption is violated then the between-group differences in adjusted means are not interpretable and the covariate should be treated as a moderator and analyses that assess the simple main effects (i.e., follow-up to a significant interaction) should be conducted.

A one-way ANCOVA requires three variables:

- IV/factor – categorical (2 or more)
- DV – continuous
- covariate – continuous

Green and Salkind [Green and Salkind, 2014a] identified common uses of ANCOVA:

- Studies with a pretest and random assignment of subjects to factor levels. Variations on this research design include:
 - assignment to factor levels based on that pretest,
 - matching based on the pretest, and random assignment to factor levels,
 - simply using the pretest as a covariate for the posttest DV.
- Studies with a potentially confounding variable (best when there is theoretical justification and prior empirical evidence for such) over which the researcher wants “control”

Although it is possible to have multi-way (e.g., 2-way, 3-way) ANCOVA, in this lecture we will only work two, one-way ANCOVAs representing these common use cases.

ANCOVA has four primary assumptions:

Linearity: The covariate is linearly related to the dependent variable within all levels of the factor (IV).

Homogeneity of regression slopes: The weights or slopes relating the covariate to the DV are equal across all levels of the factor.

Normally distributed: The DV is normally distributed in the population for any specific value of the covariate and for any one level of a factor. This assumption applies to every combination of the values of the covariate and levels ohttps://www.datanovia.com/en/lessons/ancova-in-r/f the factor and requires them all to be normally distributed. To the degree that population distributions are not normal and sample sizes are small, p values may not be trustworthy and power reduced. Evaluating this is frequently operationalized by inspecting the residuals and identifying outliers.

Homogeneity of variances: The variances of the DV for the conditional distributions (i.e., every combination of the values of the covariate and levels of the factor) are equal.

We are following the approach to analyzing ANCOVA identified in the Datanovia lesson on ANCOVA [[Datanovia, a](#)].

Our analytic process will be similar to others in the ANOVA series:

1. Prepare the data
2. Evaluate potential violation of the assumptions
3. Compute the omnibus ANCOVA, and follow-up accordingly
 - If significant: follow-up with post-hoc comparisons, planned contrasts, and/or polynomial
 - If non-significant: stopping.

An ANCOVA workflow maps this in further detail.

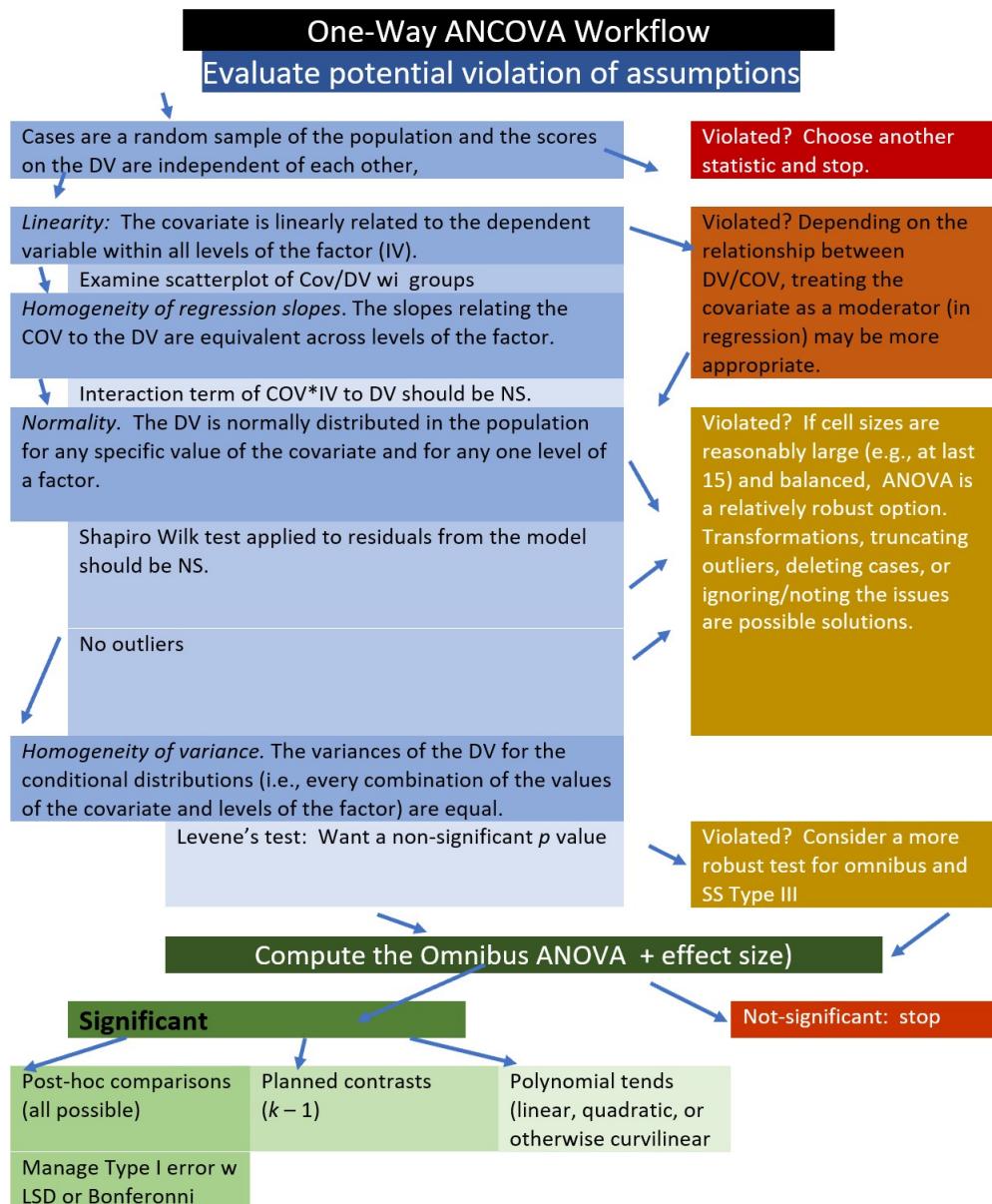


Figure 8.1: Image of the ANCOVA workflow

8.3 Research Vignette

We will continue with the example used in the [mixed design ANOVA lesson](#). The article does not contain any ANCOVA analyses, but there is enough data that I can demonstrate the two general ways (i.e., controlling for the pretest, controlling for a potentially confounding variable) that ANCOVA is used.

Here is a quick reminder of the research vignette.

Murrar and Brauer's [2018] article described the results of two studies designed to reduce prejudice against Arabs/Muslims. In the lesson on mixed design ANOVA, we only worked the first of two experiments reported in the study. Participants ($N = 193$), all who were White, were randomly assigned to one of two conditions where they watched six episodes of the sitcom *Friends* or *Little Mosque on the Prairie*. The sitcoms and specific episodes were selected after significant pilot testing. The selection was based on the tension selecting stimuli that were as similar as possible, yet the intervention-oriented sitcom needed to invoke psychological processes known to reduce prejudice. The authors felt that both series had characters that were likable and relateable who were engaged in activities of daily living. The Friends series featured characters who were predominantly White, cis-gendered, and straight. The Little Mosque series portrays the experience Western Muslims and Arabs as they live in a small Canadian town. This study involved assessment across three waves: baseline (before watching the assigned episodes), post1 (immediately after watching the episodes), and post2 (completed 4–6 weeks after watching the episodes).

The study used *feelings and liking thermometers*, rating their feelings and liking toward 10 different groups of people on a 0 to 100 sliding scale (with higher scores reflecting greater liking and positive feelings). For the purpose of this analysis, the ratings of attitudes toward White people and attitudes toward Arabs/Muslims were used. A third metric was introduced by subtracting the attitudes towards Arabs/Muslims from the attitudes toward Whites. Higher scores indicated more positive attitudes toward Whites whereas low scores indicated no difference in attitudes. To recap, there were three potential dependent variables, all continuously scaled:

- AttWhite: attitudes toward White people; higher scores reflect greater liking
- AttArab: attitudes toward Arab people; higher scores reflect greater liking
- Diff: the difference between AttWhite and AttArab; higher scores reflect a greater liking for White people

With random assignment, nearly equal cell sizes, a condition with two levels (Friends, Little Mosque), and three waves (baseline, post1, post2), this is perfect for mixed design ANOVA but suitable for an ANCOVA demonstration.

8.3.1 Simulating the data from the journal article

Below is the code I have used to simulate the data. The simulation includes two dependent variables (AttWhite, AttArab), Wave (baseline, post1, post2), and COND (condition; Friends, Little_Mosque). There is also a caseID (repeated three times across the three waves) and rowID (giving each observation within each case an ID). You can use this simulation for two of the three practice suggestions.

COND	Baseline At start of study (prior to viewing sitcoms)	Intervention 6 episodes of the sitcom	Post1 Toward end of viewing the sitcoms	Post2 4-6 weeks after viewing the final sitcom
Friends	X		X	X
Little Mosque on the Prairie	X	Selected for potential for prejudice reduction	X	X

Figure 8.2: Image of the design for the Murrar and Brauer (2018) study

```

library(tidyverse)
# change this to any different number (and rerun the simulation) to
# rework the chapter problem
set.seed(210813)
# sample size, M and SD for each cell; this will put it in a long
# file
AttWhite <- round(c(rnorm(98, mean = 76.79, sd = 18.55), rnorm(95, mean = 75.37,
  sd = 18.99), rnorm(98, mean = 77.47, sd = 18.95), rnorm(95, mean = 75.81,
  sd = 19.29), rnorm(98, mean = 77.79, sd = 17.25), rnorm(95, mean = 75.89,
  sd = 19.44)), 3)
# set upper bound for variable
AttWhite[AttWhite > 100] <- 100
# set lower bound for variable
AttWhite[AttWhite < 0] <- 0
AttArab <- round(c(rnorm(98, mean = 64.11, sd = 20.97), rnorm(95, mean = 64.37,
  sd = 20.03), rnorm(98, mean = 64.16, sd = 21.64), rnorm(95, mean = 70.52,
  sd = 18.55), rnorm(98, mean = 65.29, sd = 19.76), rnorm(95, mean = 70.3,
  sd = 17.98)), 3)
# set upper bound for variable
AttArab[AttArab > 100] <- 100
# set lower bound for variable
AttArab[AttArab < 0] <- 0
rowID <- factor(seq(1, 579))
caseID <- rep((1:193), 3)
Wave <- c(rep("Baseline", 193), rep("Post1", 193), rep("Post2", 193))
COND <- c(rep("Friends", 98), rep("LittleMosque", 95), rep("Friends", 98),
  rep("LittleMosque", 95), rep("Friends", 98), rep("LittleMosque", 95))
# groups the 3 variables into a single df: ID#, DV, condition
Murrar_df <- data.frame(rowID, caseID, Wave, COND, AttArab, AttWhite)
# make caseID a factor
Murrar_df[, "caseID"] <- as.factor(Murrar_df[, "caseID"])
# make Wave an ordered factor
Murrar_df$Wave <- factor(Murrar_df$Wave, levels = c("Baseline", "Post1",
  "Post2"))
# make COND an ordered factor

```

```
Murrar_df$COND <- factor(Murrar_df$COND, levels = c("Friends", "LittleMosque"))
# creates the difference score
Murrar_df$Diff <- Murrar_df$AttWhite - Murrar_df$AttArab
```

Let's check the structure. We want

- rowID and caseID to be unordered factors,
- Wave and COND to be ordered factors,
- AttArab, AttWhite, and Diff to be numerical

```
str(Murrar_df)
```

```
'data.frame': 579 obs. of 7 variables:
 $ rowID    : Factor w/ 579 levels "1","2","3","4",...: 1 2 3 4 5 6 7 8 9 10 ...
 $ caseID   : Factor w/ 193 levels "1","2","3","4",...: 1 2 3 4 5 6 7 8 9 10 ...
 $ Wave      : Factor w/ 3 levels "Baseline","Post1",...: 1 1 1 1 1 1 1 1 1 1 ...
 $ COND      : Factor w/ 2 levels "Friends","LittleMosque": 1 1 1 1 1 1 1 1 1 1 ...
 $ AttArab   : num  74.3 55.8 33.3 66.3 71 ...
 $ AttWhite  : num  100 79 75.9 68.2 100 ...
 $ Diff      : num  25.71 23.18 42.67 1.92 29.01 ...
```

The structure looks satisfactory. R will automatically “order” factors alphabetically or numerically. In this lesson’s example the alphabetical ordering (i.e., Baseline, Post1, Post2; Friends, LittleMosque) is consistent with the logic in our study.

If you want to export this data as a file to your computer, remove the hashtags to save it (and re-import it) as a .csv (“Excel lite”) or .rds (R object) file. This is not a necessary step.

The code for the .rds file will retain the formatting of the variables, but is not easy to view outside of R. This is what I would do. *Note: My students and I have discovered that the psych::describeBy() function seems to not work with files in the .rds format, but does work when the data are imported with .csv.*

```
# to save the df as an .rds (think 'R object') file on your computer;
# it should save in the same file as the .rmd file you are working
# with saveRDS(Murrar_df, 'Murrar_RDS.rds') bring back the simulated
# dat from an .rds file Murrar_df <- readRDS('Murrar_RDS.rds')
```

The code for .csv will likely lose the formatting (i.e., stripping Wave and COND of their ordered factors), but it is easy to view in Excel.

```
# write the simulated data as a .csv write.table(Murrar_df,
# file='DiffCSV.csv', sep=',', col.names=TRUE, row.names=FALSE) bring
# back the simulated dat from a .csv file Murrar_df <- read.csv
# ('DiffCSV.csv', header = TRUE)
```

8.4 Scenario #1: Controlling for the pretest

So that we can begin to understand how the covariate operates, we are going to predict attitudes towards Arabs at post-test (AttArabP1) by condition (COND), controlling for attitudes toward Arabs at baseline (AttArabB). You may notice that in this analysis we are ignoring the second post-test. This is because I am simply demonstrating ANCOVA. To ignore the second post test would be a significant loss of information.

8.4.1 Preparing the data

When the covariate in ANCOVA is a pretest, we need three variables:

- IV that has two or more levels; in our case it is the Friends and Little Mosque conditions
- DV that is continuous; in our case it is the attitudes toward Arabs at post1
- Covariate that is continuous; in our case it is the attitudes toward Arabs at baseline

The form of our data matters. The simulation created a *long* form (formally called the *person-period* form) of data. That is, each observation for each person is listed in its own row. In this dataset where we have 193 people with 3 observations (baseline, post1, post2) each, we have 579 rows. In ANCOVA where we use the pre-test as a covariate, we need all the data to be on a single row. This is termed the *person level* form of data. We can restructure the data with the *data.table* and *reshape2()** packages.

```
# Create a new df (Murrar_wide) Identify the original df In the
# transition from long-to-wide it seems like you can only do one
# time-varying variable at a time When there are multiple
# time-varying and time-static variables, put all the time-static
# variables on the left side of the tilde Put the name of the single
# time-varying variable in the concatenated list
Murrar1 <- reshape2::dcast(data = Murrar_df, formula = caseID + COND ~
    Wave, value.var = "AttArab")
# before restructuring a second variable, rename the first variable
Murrar1 <- rename(Murrar1, AttArabB = "Baseline", AttArabP1 = "Post1",
    AttArabP2 = "Post2")
# repeat the process for additional variables; but give the new df
# new names -- otherwise you'll overwrite your work
Murrar2 <- reshape2::dcast(data = Murrar_df, formula = caseID ~ Wave, value.var = "AttWhite")
Murrar2 <- rename(Murrar2, AttWhiteB = "Baseline", AttWhiteP1 = "Post1",
    AttWhiteP2 = "Post2")
# Now we join them
Murrar_wide <- dplyr::full_join(Murrar1, Murrar2, by = c("caseID"))

str(Murrar_wide)

'data.frame': 193 obs. of 8 variables:
 $ caseID   : Factor w/ 193 levels "1","2","3","4",...: 1 2 3 4 5 6 7 8 9 10 ...

```

```
$ COND      : Factor w/ 2 levels "Friends","LittleMosque": 1 1 1 1 1 1 1 1 1 1 ...
$ AttArabB  : num  74.3 55.8 33.3 66.3 71 ...
$ AttArabP1 : num  80.3 76.6 92 96.5 59.1 ...
$ AttArabP2 : num  64.8 43.3 40.3 69.1 74.9 ...
$ AttWhiteB : num  100 79 75.9 68.2 100 ...
$ AttWhiteP1: num  95.6 51 91.9 86.7 75.8 ...
$ AttWhiteP2: num  100 89.7 49.5 99.4 83.1 ...
```

If you want to export this data as a file to your computer, remove the hashtags to save it (and re-import it) as a .csv (“Excel lite”) or .rds (R object) file. This is not a necessary step.

The code for the .rds file will retain the formatting of the variables, but is not easy to view outside of R. This is what I would do.

```
# to save the df as an .rds (think 'R object') file on your computer;
# it should save in the same file as the .rmd file you are working
# with saveRDS(Murrar_wide, 'MurrarW_RDS.rds') bring back the
# simulated dat from an .rds file Murrar_wide <-
# readRDS('MurrarW_RDS.rds')
```

The code for .csv will likely lose the formatting (i.e., stripping Wave and COND of their ordered factors), but it is easy to view in Excel.

```
# write the simulated data as a .csv write.table(Murrar_wide,
# file='MurrarW_CSV.csv', sep=',', col.names=TRUE, row.names=FALSE)
# bring back the simulated dat from a .csv file Murrar_wide <-
# read.csv ('MurrarW_CSV.csv', header = TRUE)
```

8.4.2 Checking the assumptions

There are a number of assumptions in ANCOVA. These include:

- random sampling
- independence in the scores representing the dependent variable
 - there is, of course, intentional dependence in any repeated measures or within-subjects variable
- linearity of the relationship between the covariate and DV within all levels of the independent variable
- homogeneity of the regression slopes
- a normally distributed DV for any specific value of the covariate and for any one level of a factor
- homogeneity of variance

These are depicted in the flowchart, below.

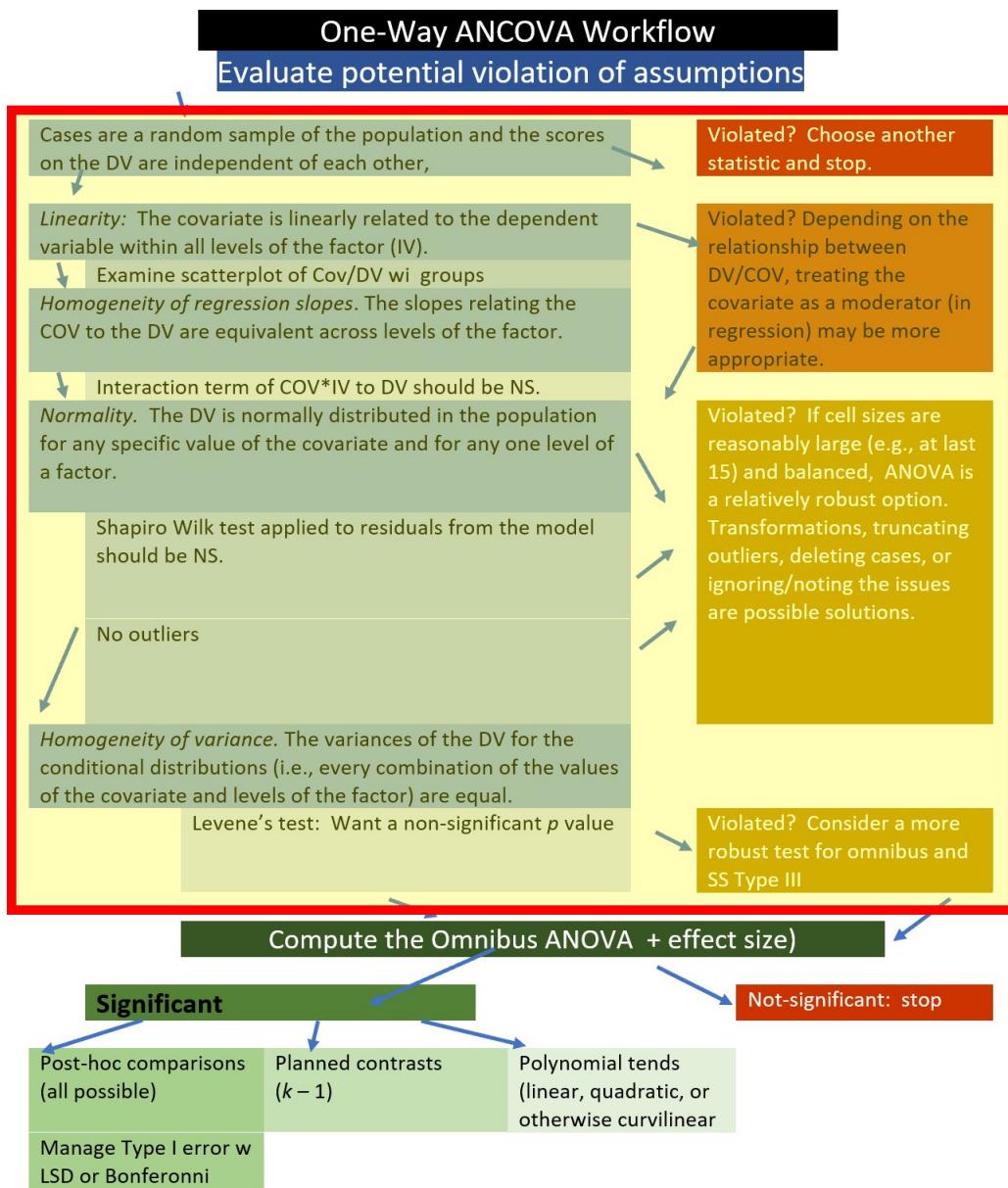


Figure 8.3: Image of the ANCOVA workflow, showing our current place in the process

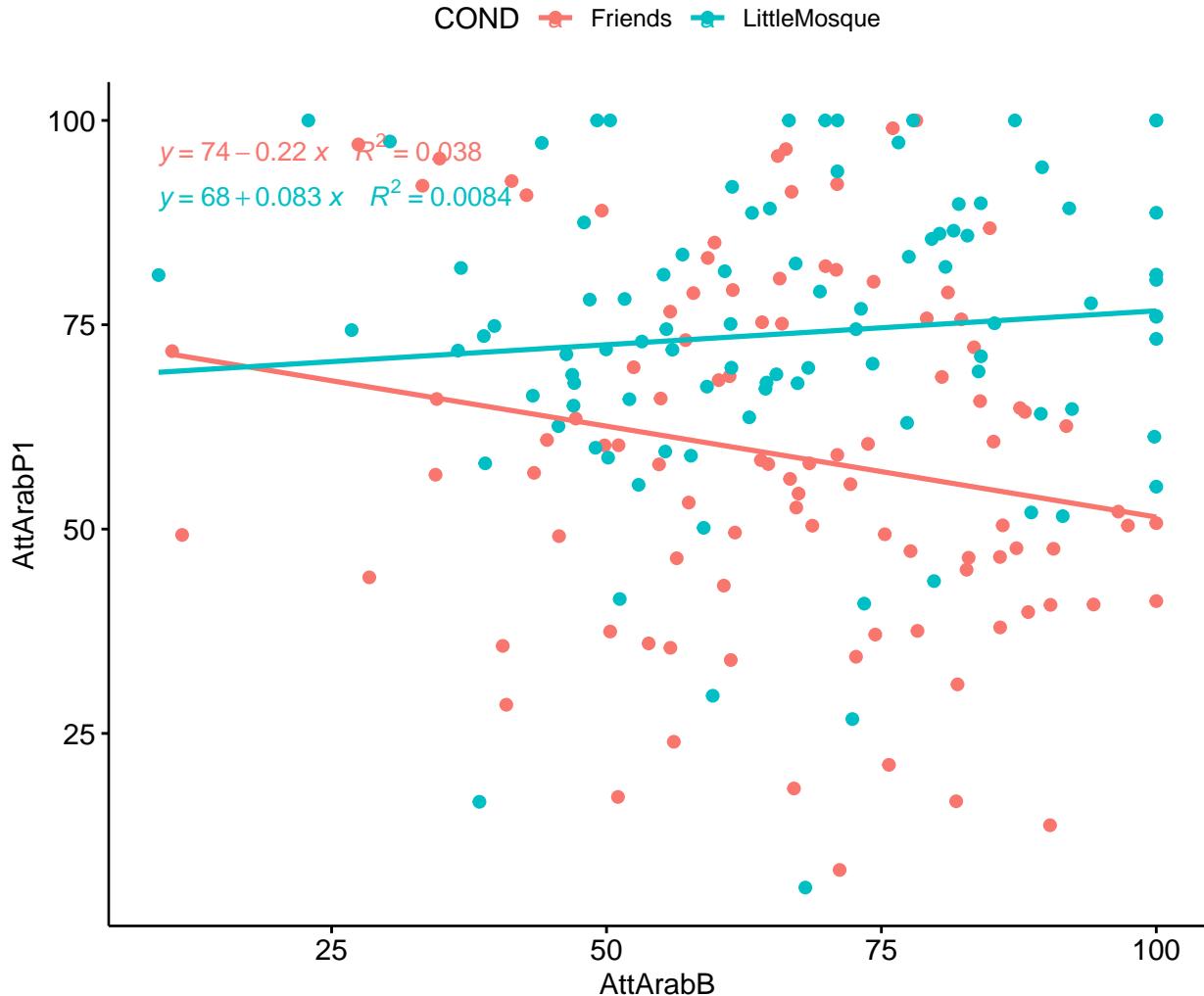
8.4.2.1 Linearity assumption

ANCOVA assumes that there is linearity between the covariate and outcome variable at each level of the grouping variable. In our case this means that there is linearity between the pre-test (covariate) and post-test (outcome variable) at each level of the intervention (Friends, Little Mosque).

We can create a scatterplot (with regression lines) between covariate (our pretest) and the outcome (post-test1).

```
ggpubr::ggscatter(Murrar_wide, x = "AttArabB", y = "AttArabP1", color = "COND",
  add = "reg.line") + ggpubr::stat_regrline_equation(aes(label = paste(..eq.label..,
  ..rr.label.., sep = "~~~~"), color = COND))
```

```
`geom_smooth()` using formula 'y ~ x'
```



As is not surprising (because we tested a similar set of variables in the mixed design chapter), this relationship look like an interaction effect. Let's continue our exploration.

8.4.2.2 Homogeneity of regression slopes

This assumption requires that the slopes of the regression lines formed by the covariate and the outcome variable are the same for each group. The assumption evaluates that there is no interaction between the outcome and covariate. The plotted regression lines should be parallel.

```
Murrar_wide %>%
  rstatix::anova_test(AttArabP1 ~ COND * AttArabB)
```

Coefficient covariances computed by hccm()

ANOVA Table (type II tests)

	Effect	DFn	DFd	F	p	p < .05	ges
1	COND	1	189	26.819	0.000000569	*	0.124
2	AttArabB	1	189	0.676	0.412000000		0.004
3	COND:AttArabB	1	189	4.297	0.040000000	*	0.022

Because the statistically significant interaction term is violation of homogeneity of regression slopes ($F [1, 189] = 4.297, p = .040, \eta^2 = 0.022$) we should not proceed with ANCOVA as a statistical option. However, for the sake of demonstration, I will continue. One of the reasons I wanted to work this example as ANCOVA is to demonstrate that covariates and moderators each have their role. We can already see how this data is best analyzed with mixed design ANOVA.

8.4.2.3 Normality of residuals

Our goal here is to specify a model and extract *residuals*: the difference between the observed value of the DV and its predicted value. Each data point has one residual. The sum and mean of residuals are equal to 0.

Once we have saved the residuals, we can treat them as data and evaluate the shape of their distribution. We hope that the distribution is not statistically significantly different from a normal one. We first compute the model with *lm()* (*lm* stands for “linear model”). This is a linear regression.

```
# Create a linear regression model predicting DV from COV & IV
AttArabB_Mod <- lm(AttArabP1 ~ AttArabB + COND, data = Murrar_wide)
AttArabB_Mod
```

Call:

```
lm(formula = AttArabP1 ~ AttArabB + COND, data = Murrar_wide)
```

Coefficients:

(Intercept)	AttArabB	CONDLittleMosque
63.01428	-0.06042	14.92165

With the `broom::augment()` function we can augment our `lm()` model object to add fitted values and residuals.

```
# new model by augmenting the lm model
AttArabB_Mod.metrics <- broom::augment(AttArabB_Mod)
# shows the first three rows of the UEmodel.metrics
head(AttArabB_Mod.metrics, 3)

# A tibble: 3 x 9
  AttArabP1 AttArabB COND     .fitted   .resid    .hat   .sigma   .cooksdi .std.resid
  <dbl>      <dbl> <fct>     <dbl>     <dbl>   <dbl>   <dbl>     <dbl>
1     80.3     74.3 Friends    58.5    21.7 0.0111   20.2 0.00440    1.08
2     76.6     55.8 Friends    59.6    17.0 0.0116   20.2 0.00280    0.845
3     92.0     33.3 Friends    61.0    31.0 0.0247   20.1 0.0204    1.56
```

From this, we can assess the normality of the residuals using the Shapiro Wilk test

```
# apply shapiro_test to that augmented model
rstatix::shapiro_test(AttArabB_Mod.metrics$.resid)
```

```
# A tibble: 1 x 3
  variable           statistic p.value
  <chr>              <dbl>    <dbl>
1 AttArabB_Mod.metrics$.resid     0.984  0.0261
```

The statistically significant Shapiro Wilk test has indicated a violation of the normality assumption ($W = 0.984$, $p = .026$).

8.4.2.4 Homogeneity of variances

ANCOVA presumes that the variance of the residuals is equal for all groups. We can check this with the Levene's test.

```
AttArabB_Mod.metrics %>%
  rstatix::levene_test(.resid ~ COND)
```

```
# A tibble: 1 x 4
  df1   df2 statistic     p
  <int> <int>    <dbl>   <dbl>
1     1    191     3.52 0.0623
```

A non-significant Levene's test indicated no violation of the homogeneity of the residual variances for all groups ($F[1, 191] = 3.515$ $p = .062$).

8.4.2.5 Outliers

We can identify outliers by examining the standardized (or studentized) residuals. This is the residual divided by its estimated standard error. Standardized residuals are interpreted as the number of standard errors away from the regression line.

```
# from our model metrics show us any standardized residuals that are
# >3
AttArabB_Mod.metrics %>%
  filter(abs(.std.resid) > 3) %>%
  as.data.frame()
```

	AttArabP1	AttArabB	COND	.fitted	.resid	.hat	.sigma
1	6.137	68.085	LittleMosque	73.82234	-67.68534	0.01056251	19.62279
			.cooksdi	.std.resid			
1	0.04044273			-3.371254			

We do have one outlier with a standardized residual that has an absolute value greater than 3. At this point I am making a mental note of this. If this were “for real” I might more closely inspect these data. I would look at the whole response. If any response seemed invalid (e.g., random, extreme, or erratic responding) I would delete it. If the responses seemed valid, I *could* truncate them to exactly 3 SEs or. I could also ignore it. Kline [2016] has a great section on some of these options.

As noted by the suggestion of an interaction effect, our preliminary analyses suggests that ANCOVA is not the best option. We know from the prior lesson that a mixed design ANOVA worked well. In the spirit of an example, here’s a preliminary write-up so far:

8.4.2.6 Write-up of Assumptions

A one-way analysis of covariance (ANCOVA) was conducted. The independent variable, condition, had two levels: Friends, Little Mosque. The dependent variable was attitudes towards Arabs expressed by the participant at post-test and covariate was the pre-test assessment of the same variable. A preliminary analysis evaluating the homogeneity-of-slopes assumption indicated that the relationship between the covariate and the dependent variable differed significantly as a function of the independent variable, $F(1, 189) = 4.297, p = .040, \eta^2 = 0.022$. Regarding the assumption that the dependent variable is normally distributed in the population for any specific value of the covariate and for any one level of a factor, results of the Shapiro-Wilk test of normality on the model residuals was also significant, $W = 0.984, p = .026$. Only one datapoint (in the Little Mosque condition) had a standardized residual (-3.37) that exceeded an absolute value of 3.0. A non-significant Levene’s test indicated no violation of the homogeneity of the residual variances for all groups, $F(1, 191) = 3.515, p = .062$.

8.4.3 Calculating the Omnibus ANOVA

We are ready to conduct the omnibus ANOVA.

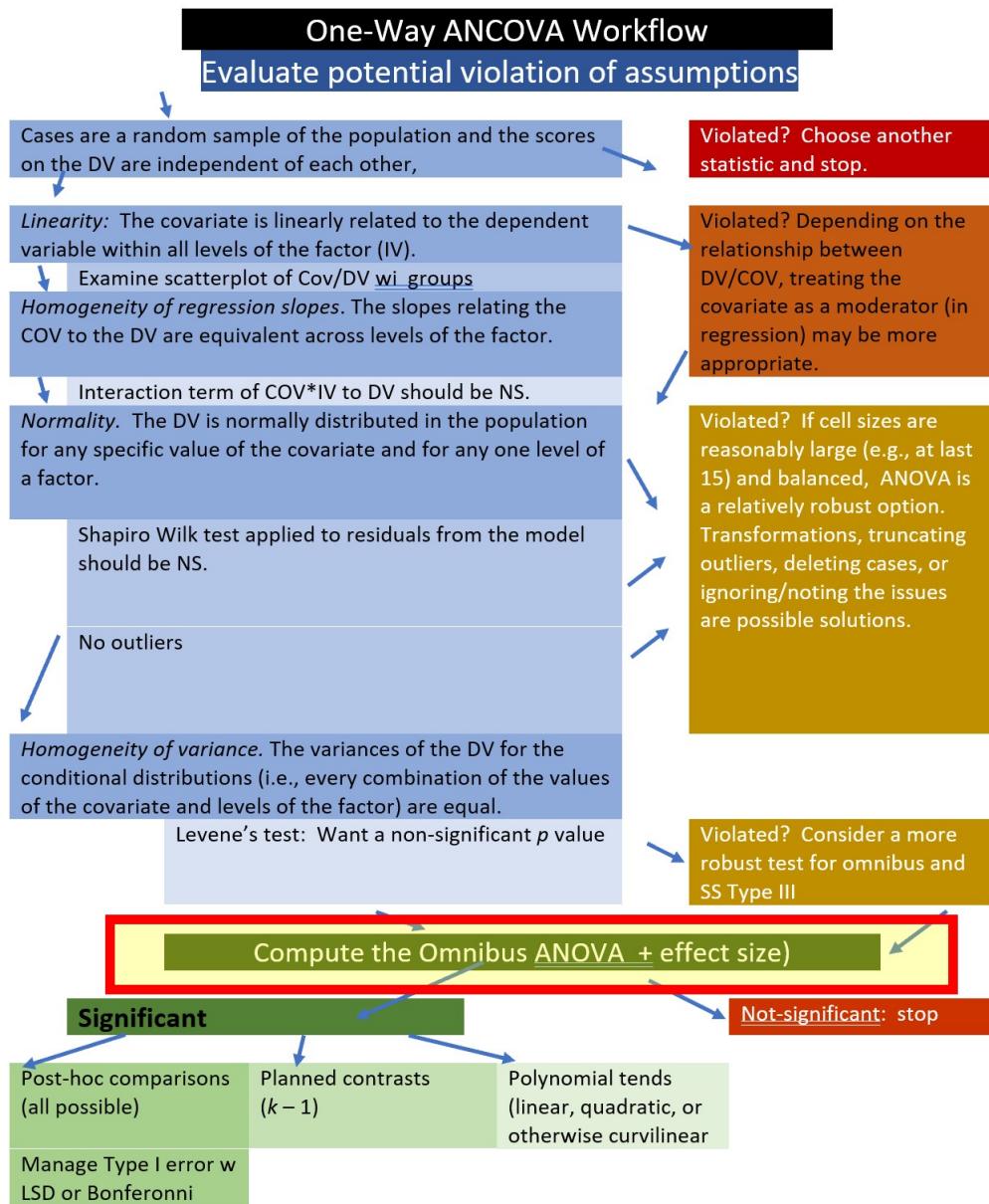


Figure 8.4: Image of the ANCOVA workflow, showing our current place in the process.

Order of variable entry matters in ANCOVA. Thinking of the *controlling for* language associate with covariates, we want to remove the effect of the covariate before we run the one-way ANOVA. With this ANCOVA we are asking the question, “Does the condition (Friends or Little Mosque) contribute to more positive attitudes toward Arabs, when controlling for the pre-test score?”

In repeated measures projects, we expect there to be dependency in the data. That is, in most cases prior waves will have significant prediction on later waves. When ANCOVA uses a prior assessment or wave as a covariate, that variable “claims” as much variance as possible and the subsequent variable can capture what is left over.

In the code below, we are predicting attitudes toward Arabs at post1 from the condition (Friends or Little Mosque), controlling for attitudes toward Arabs at baseline.

The *ges* column provides the effect size, η^2 where a general rule-of-thumb for interpretation is .02 (small), .13 (medium), and .26 (large) [Lakens, 2013].

```
MurrarB_ANCOVA <- Murrar_wide %>%
  rstatix::anova_test(AttArabP1 ~ AttArabB + COND)
```

Coefficient covariances computed by hccm()

```
rstatix::get_anova_table(MurrarB_ANCOVA)
```

ANOVA Table (type II tests)

	Effect	DFn	DFd	F	p	p<.05	ges
1	AttArabB	1	190	0.665	0.416	0.000000000	0.003
2	COND	1	190	26.361	0.000000698	*	0.122

There was a non-significant effect of the baseline covariate on the post-test ($F[1, 190] = 0.665, p = .416, \eta^2 = 0.003$). After controlling for the baseline attitudes toward Arabs, there was a statistically significant effect of condition on post-test attitudes toward Arabs, $F(1,190) = 26.361, p < .001, \eta^2 = 0.122$. This appears to be a moderately sized effect.

8.4.4 Post-hoc pairwise comparisons (controlling for the covariate)

Just like in one-way ANOVA, we follow-up the significant effect of condition. We'll use all-possible pairwise comparisons. In our case, we only have two levels of the categorical factor, so this run wouldn't be necessary. I included it to provide the code for doing so. If there were three or more variables, we would see all possible comparisons.

```
pwc_B <- Murrar_wide %>%
  rstatix::emmeans_test(AttArabP1 ~ COND, covariate = AttArabB, p.adjust.method = "none")
```

```
# A tibble: 1 x 9
  term      .y. group1 group2    df statistic      p   p.adj p.adj.signif
* <chr>     <chr> <chr> <chr> <dbl>    <dbl> <dbl> <dbl> <chr>
1 AttArabB*COND AttA~ Frien~ Littl~    190     -5.13 6.98e-7 6.98e-7 ****
```

Not surprisingly (since this single pairwise comparison is redundant with the omnibus ANCOVA), results suggest a statistically significant difference between Friends and Little Mosque at Post1.

With the script below we can obtain the covariate-adjusted marginal means. These are termed *estimated marginal means*. Take a look at these and compare them to what we would see in the regular descriptives. It is helpful to see the grand mean (AttArabB) and then the marginal means (emmmean).

```
emmmeans_B <- rstatix::get_emmeans(pwc_B)
emmmeans_B
```

	AttArabB	COND	emmmean	se	df	conf.low	conf.high	method
	<dbl>	<fct>	<dbl>	<dbl>	<dbl>	<dbl>	<dbl>	<chr>
1	66.2	Friends	59.0	2.04	190	55.0	63.0	Emmeans test
2	66.2	LittleMosque	73.9	2.07	190	69.8	78.0	Emmeans test

Note that the *emmeans* process produces slightly different means than the raw means produced with the *psych* package's *describeBy()* function. Why? Because the *get_emmeans()* function uses the model that included the covariate. That is, the *estimated* means are covariate-adjusted.

```
descripts_P1 <- psych::describeBy(AttArabP1 ~ COND, data = Murrar_wide,
    mat = TRUE)
descripts_P1
```

	item	group1	vars	n	mean	sd	median	trimmed	mad	min
X11	1	Friends	1	98	59.02351	21.65024	57.9955	59.31306	23.67045	8.297
X12	2	LittleMosque	1	95	73.92134	18.51082	74.4600	75.52858	15.98984	6.137
	max	range	skew	kurtosis	se					
X11	100	91.703	-0.0518848	-0.6252126	2.187005					
X12	100	93.863	-0.9798189	1.6335325	1.899170					

```
# Note. Recently my students and I have been having intermittent
# struggles with the describeBy function in the psych package. We
# have noticed that it is problematic when using .rds files and when
# using data directly imported from Qualtrics. If you are having
# similar difficulties, try uploading the .csv file and making the
# appropriate formatting changes.
```

$(M = 59.02, SD = 21.65)$ $(M = 73.92, SD = 18.51)$

In our case the adjustments are very minor. Why? The effect of the attitudes toward Arabs baseline test on the attitudes toward Arabs post test was nonsignificant. We can see this in the bivariate correlations, below.

```
MurP1_Rmat <- psych::corr.test(Murrar_wide[c("AttArabB", "AttArabP1")])
MurP1_Rmat
```

Call:psych::corr.test(x = Murrar_wide[c("AttArabB", "AttArabP1")])
Correlation matrix

	AttArabB	AttArabP1
AttArabB	1.00	-0.05
AttArabP1	-0.05	1.00

Sample Size
[1] 193
Probability values (Entries above the diagonal are adjusted for multiple tests.)

	AttArabB	AttArabP1
AttArabB	0.00	0.47
AttArabP1	0.47	0.00

To see confidence intervals of the correlations, print with the short=FALSE option

The correlation between attitudes toward Arabs at baseline and post test are nearly negligible ($r = -0.05$, $p = .47$)

8.4.5 Toward an APA style results section

As we assemble the elements for an APA style result sections, a table with the means, adjusted means, and correlations may be helpful.

```
apaTables::apa.cor.table(Murrar_wide[c("AttArabB", "AttArabP1")], table.number = 1)
```

Table 1

Means, standard deviations, and correlations with confidence intervals

Variable	M	SD	1
1. AttArabB	66.25	19.66	
2. AttArabP1	66.36	21.46	-.05 [-.19, .09]

Note. M and SD are used to represent mean and standard deviation, respectively.
Values in square brackets indicate the 95% confidence interval.

The confidence interval is a plausible range of population correlations that could have caused the sample correlation (Cumming, 2014).

* indicates $p < .05$. ** indicates $p < .01$.

```
# You can save this as a Microsoft word document by adding this
# statement into the command: filename = 'your_filename.doc'
```

Additionally, writing this output to excel files helped create the two tables that follow. The *MASS* package is useful to export the model objects into .csv files. They are easily opened in Excel where they can be manipulated into tables for presentations and manuscripts.

```
MASS::write.matrix(pwc_B, sep = ",", file = "pwc_B.csv")
MASS::write.matrix(emmeans_B, sep = ",", file = "emmeans_B.csv")
MASS::write.matrix(descripts_P1, sep = ",", file = "descripts_P1.csv")
```

Ultimately, I would want a table that included this information. Please refer to the APA style manual for more proper formatting for a manuscript that requires APA style.

Table 1

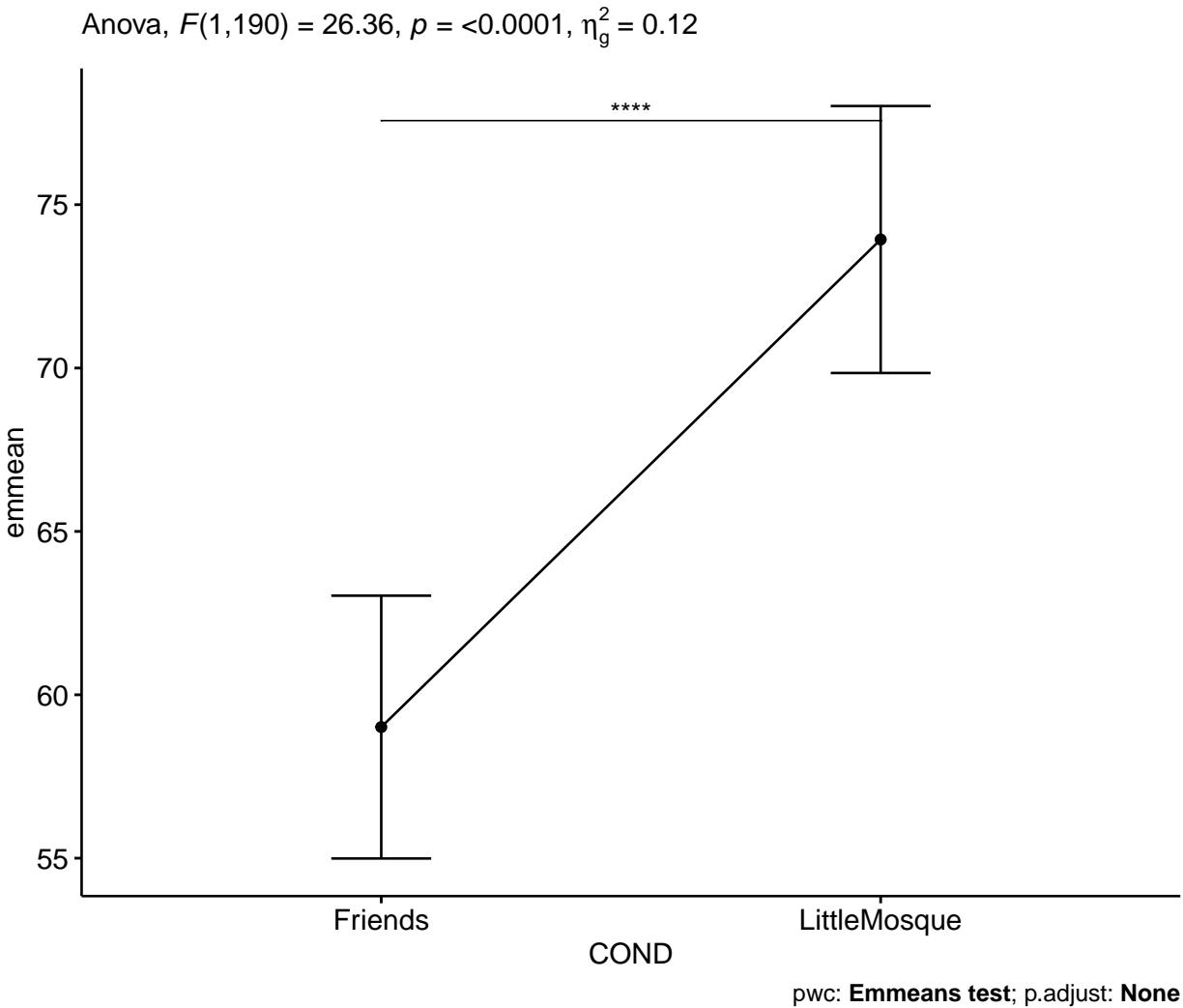
Unadjusted and Covariate-Adjusted Descriptive Statistics

Condition	Unadjusted	Covariate-Adjusted
-----------	------------	--------------------

	<i>M</i>	<i>SD</i>	<i>EMM</i>	<i>SE</i>
Friends	59.02	21.65	59.01	2.04
Little Mosque	73.92	18.51	73.93	2.07

Unlike the figure we created when we were testing assumptions, this script creates a plot from the model (which identifies AttArabB in its role as covariate). Thus, the relationship between condition and AttArabP1 controls for the effect of the AttArabB covariate.

```
pwc_B <- pwc_B %>%
  rstatix::add_xy_position(x = "COND", fun = "mean_se")
ggpubr::ggline(rstatix::get_emmeans(pwc_B), x = "COND", y = "emmean") +
  geom_errorbar(aes(ymin = conf.low, ymax = conf.high), width = 0.2) +
  ggpubr::stat_pvalue_manual(pwc_B, hide.ns = TRUE, tip.length = FALSE) +
  labs(subtitle = rstatix::get_test_label(MurrarB_ANCOVA, detailed = TRUE),
       caption = rstatix::get_pwc_label(pwc_B))
```



Results

A one-way analysis of covariance (ANCOVA) was conducted. The independent variable, condition, had two levels: Friends, Little Mosque. The dependent variable was attitudes towards Arabs expressed by the participant at post-test and covariate was the baseline assessment of the same variable. Descriptive statistics are presented in Table 1. A preliminary analysis evaluating the homogeneity-of-slopes assumption indicated that the relationship between the covariate and the dependent variable differed significantly as a function of the independent variable, $F(1, 189) = 4.297, p = .040, \eta^2 = 0.022$. Regarding the assumption that the dependent variable is normally distributed in the population for any specific value of the covariate and for any one level of a factor, results of the Shapiro-Wilk test of normality on the model residuals was also significant, $W = 0.984, p = .026$. Only one datapoint (in the Little Mosque condition) had a standardized residual (-3.37) that exceeded an absolute value of 3.0. A non-significant Levene's test indicated no violation of the homogeneity of the residual variances for all groups, $F(1, 191) = 3.515 p = .062$.

There was a non-significant effect of the baseline covariate on the post-test ($F[1, 190] = 0.665, p = .416, \eta^2 = 0.003$). After controlling for the baseline attitudes toward Arabs, there was a statistically significant effect of condition on post-test attitudes toward Arabs, $F(1, 190) = 26.361, p < .001, \eta^2 = 0.122$. This appears to be moderate in size. Given there were only two conditions, no further

follow-up was required. As illustrated in Figure 1, results suggest that those in the Little Mosque condition ($M = 73.92$, $SD = 18.51$) had more favorable attitudes toward Arabs than those in the Friends condition ($M = 59.02$, $SD = 21.65$). Means and covariate-adjusted means are presented in Table 1b.

8.5 Scenario #2: Controlling for a confounding or covarying variable

In the scenario below, I am simulating a one-way ANCOVA, predicting attitudes toward Arabs at post1 as a function of sitcom condition (Friends, Little Mosque), controlling for the participants' attitudes toward Whites. That is, the ANCOVA will compare the the means of the two groups (at post1, only), adjusted for level of attitudes toward Whites

TO BE CLEAR: This is not the best way to analyze this data. With such a strong, balanced design, the multi-way, mixed design ANOVAs were an excellent choice that provided much fuller information than this demonstration, below. The purpose of this over-simplified demonstration is merely to give another example of using a variable as a *covariate* rather than a *moderator*.

8.5.1 Preparing the data

When the covariate in ANCOVA is a potentially confounding variable, we need three variables:

- IV that has two or more levels; in our case it is the Friends and Littls Mosque sitcom conditions.
 - DV that is continuous; in our case it attitudes toward Arabs at post1 (AttArabP1).
 - Covariate that is continuous; in our case it attitudes toward Whites at post1 (AttWhiteP1).
- Note* We could have also chosen attitudes toward Whites at baseline.

We can continue using the Murrar_wide df.

8.5.2 Checking the assumptions

There are a number of assumptions in ANCOVA. These include:

- random sampling
- independence in the scores representing the dependent variable
- linearity of the relationship between the covariate and DV within all levels of the independent variable
- homogeneity of the regression slopes
- a normally distributed DV for any specific value of the covariate and for any one level of a factor
- homogeneity of variance

These are depicted in the flowchart, below.

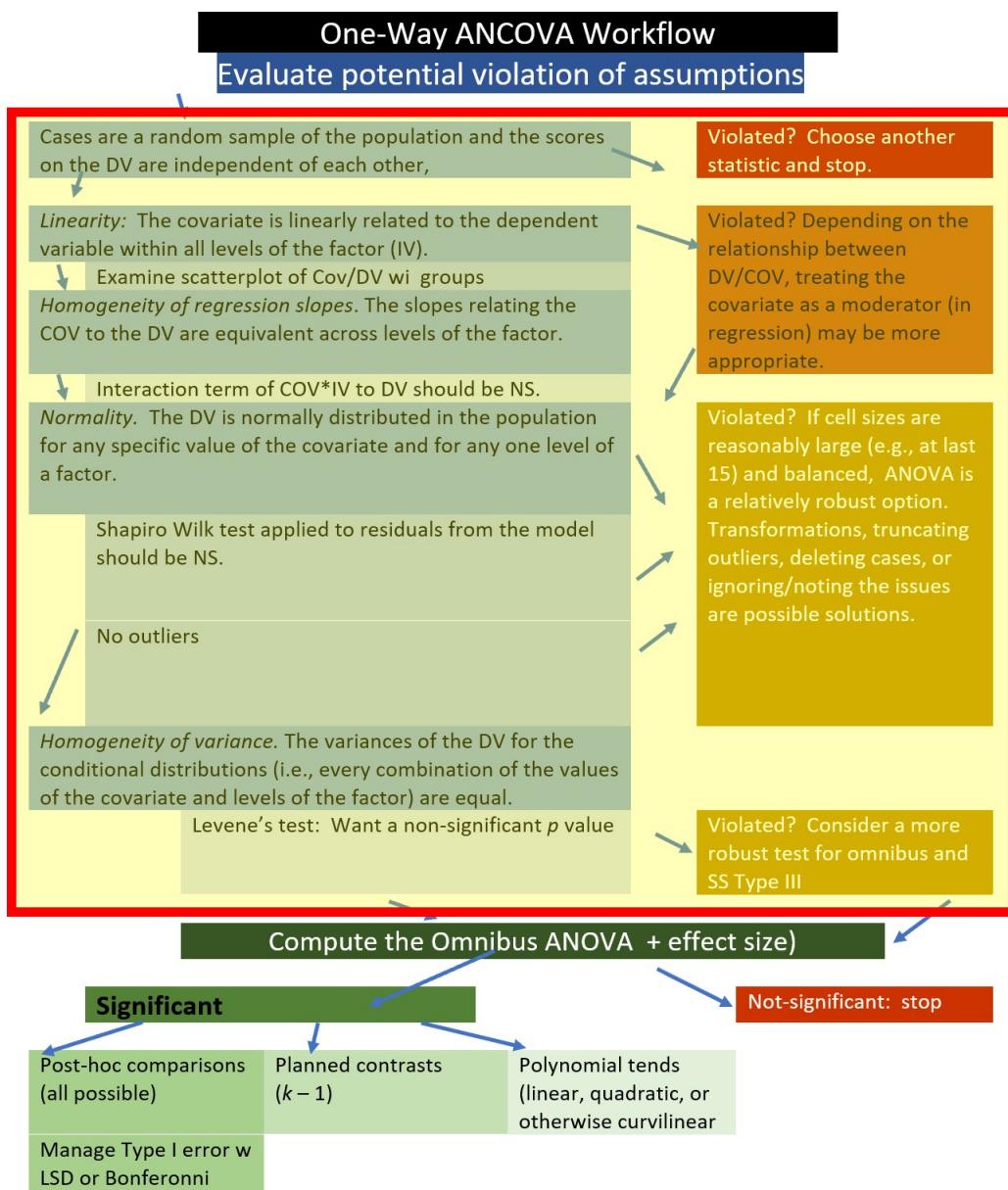


Figure 8.5: Image of the ANCOVA workflow, showing our current place in the process

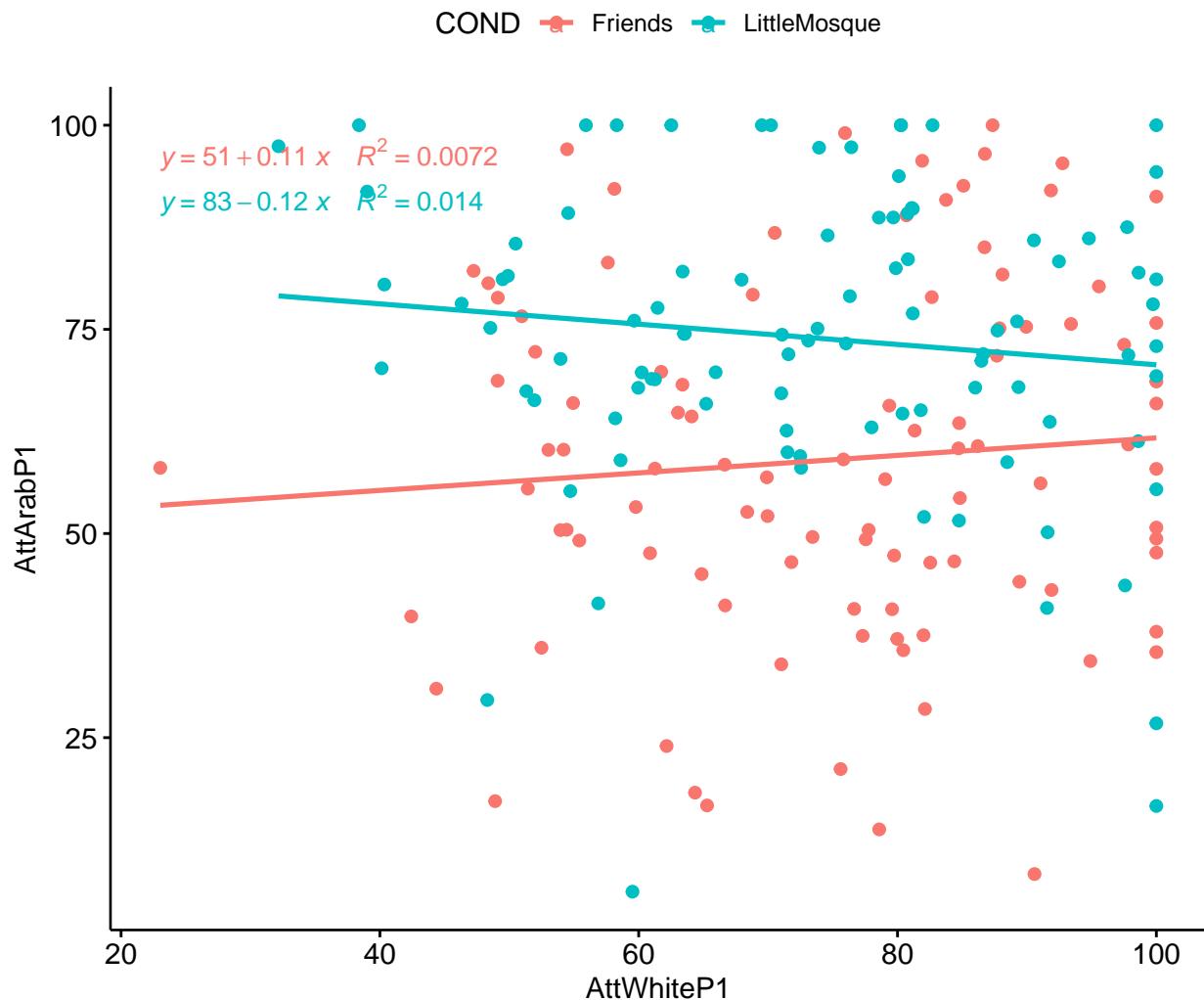
8.5.2.1 Linearity assumption

ANCOVA assumes that there is linearity between the covariate and outcome variable at each level of the grouping variable. In our case this means that there is linearity between the attitudes toward Whites (covariate) and attitudes toward Arabs (outcome variable) at each level of the intervention (Friends, Little Mosque).

We can create a scatterplot (with regression lines) between the covariate (attitudes toward Whites) and the outcome (attitudes toward Arabs).

```
ggpubr::ggscatter(Murrar_wide, x = "AttWhiteP1", y = "AttArabP1", color = "COND",
  add = "reg.line") + ggpubr::stat_regrline_equation(aes(label = paste(..eq.label..,
  ..rr.label.., sep = "~~~~"), color = COND))

`geom_smooth()` using formula 'y ~ x'
```



As we look at this scatterplot, we are trying to determine if there is an interaction effect (rather than a covarying effect). The linearity here, looks reasonable and not terribly “interacting” (to

help us decide whether empathy should be a covariate or a moderator). More testing can help us make this distinction.

8.5.2.2 Homogeneity of regression slopes

This assumption requires that the slopes of the regression lines formed by the covariate and the outcome variable are the same for each group. The assumption evaluates that there is no interaction between the outcome and covariate. The plotted regression lines should be parallel.

```
Murrar_wide %>%
  rstatix::anova_test(AttArabP1 ~ COND * AttWhiteP1)
```

Coefficient covariances computed by hccm()

ANOVA Table (type II tests)

	Effect	DFn	DFd	F	p	p<.05	ges
1	COND	1	189	26.240	0.00000074	*	0.1220000
2	AttWhiteP1	1	189	0.014	0.90700000		0.0000729
3	COND:AttWhiteP1	1	189	1.886	0.17100000		0.0100000

Preliminary analysis supported ANCOVA as a statistical option in that there was no violation of the homogeneity of regression slopes as the interaction term was not statistically significant, $F(1, 189) = 1.886$, $p = .171$, $\eta^2 = 0.010$.

8.5.2.3 Normality of residuals

Assessing the normality of residuals means running the model, capturing the unexplained portion of the model (i.e., the *residuals*), and then seeing if they are normally distributed. Proper use of ANCOVA is predicated on normally distributed residuals.

We first compute the model with *lm()*. The *lm()* function is actually testing what we want to test. However, at this early stage, we are just doing a “quick run and interpretation” to see if we are within the assumptions of ANCOVA.

```
# Create a linear regression model predicting DV from COV & IV
WhCov_mod <- lm(AttArabP1 ~ AttWhiteP1 + COND, data = Murrar_wide)
WhCov_mod
```

Call:

```
lm(formula = AttArabP1 ~ AttWhiteP1 + COND, data = Murrar_wide)
```

Coefficients:

(Intercept)	AttWhiteP1	CONDLittleMosque
59.765300	-0.009897	14.886178

We can use the `augment(model)` function from the `broom` package to add fitted values and residuals.

```
WhCov_mod.metrics <- broom::augment(WhCov_mod)
# shows the first three rows of the UEcon_model.metrics
head(WhCov_mod.metrics, 3)

# A tibble: 3 x 9
  AttArabP1 AttWhiteP1 COND     .fitted .resid   .hat .sigma .cooksdi .std.resid
  <dbl>      <dbl> <fct>     <dbl>   <dbl> <dbl>  <dbl>    <dbl>
1     80.3      95.6 Friends    58.8   21.4  0.0176  20.2  0.00685   1.07
2     76.6      51.0 Friends    59.3   17.3  0.0203  20.2  0.00518   0.867
3     92.0      91.9 Friends    58.9   33.2  0.0152  20.1  0.0140    1.65
```

Now we assess the normality of residuals using the Shapiro Wilk test. The script below captures the “.resid” column from the model.

```
rstatix::shapiro_test(WhCov_mod.metrics$.resid)
```

```
# A tibble: 1 x 3
  variable           statistic p.value
  <chr>              <dbl>    <dbl>
1 WhCov_mod.metrics$.resid    0.984  0.0294
```

The statistically significant Shapiro Wilk test indicate a violation of the normality assumption ($W = 0.984$, $p = .029$). As I mentioned before, there are better ways to analyze this research vignette. None-the-less, we will continue with this demonstration so that you will have the procedural and conceptual framework for conducting ANCOVA.

8.5.2.4 Homogeneity of variances

ANCOVA presumes that the variance of the residuals is equal for all groups. We can check this with the Levene’s test.

```
WhCov_mod.metrics %>%
  rstatix::levene_test(.resid ~ COND)

# A tibble: 1 x 4
  df1   df2 statistic      p
  <int> <int>    <dbl>  <dbl>
1     1    191     4.54  0.0344
```

Contributing more evidence that ANCOVA is not the best way to analyze this data, a statistically significant Levene’s test indicates a violation of the homogeneity of the residual variances ($F[1, 191] = 4.539$, $p = .034$).

8.5.2.5 Outliers

We can identify outliers by examining the standardized (or studentized) residual. This is the residual divided by its estimated standard error. Standardized residuals are interpreted as the number of standard errors away from the regression line.

```
WhCov_mod.metrics %>%
  filter(abs(.std.resid) > 3) %>%
  as.data.frame()

AttArabP1 AttWhiteP1      COND .fitted   .resid     .hat   .sigma
1       6.137     59.518 LittleMosque 74.06242 -67.92542 0.01407535 19.65185
       .cooks.d .std.resid
1 0.05447684 -3.383443
```

There is one outlier with a standardized residual with an absolute value greater than 3. At this point I am making a mental note of this. If this were “for real” I might more closely inspect these data. I would look at the whole response. If any response seems invalid (e.g., random, erratic, or extreme responding) I would delete it. If the response seem valid, I *could* truncate them to within 3 SEs. I could also ignore it. Kline [2016] has a great section on some of these options.

8.5.2.6 Write-up of Assumptions

A one-way analysis of covariance (ANCOVA) was conducted. The independent variable, sitcom condition, had two levels: Friends, Little Mosque. The dependent variable was attitudes towards Arabs at pre-test. Preliminary analyses which tested the assumptions of ANCOVA were mixed. Results suggesting that the relationship between the covariate and the dependent variable did not differ significantly as a function of the independent variable ($F[1, 189] = 1.886, p = .171, \eta^2 = 0.010$) provided evidence that we did not violate the homogeneity-of-slopes assumption. In contrast, the Shapiro-Wilk test of normality on the model residuals was statistically significant ($W = 0.984, p = .029$). This means that we likely violated the assumption that the dependent variable is normally distributed in the population for any specific value of the covariate and for any one level of a factor. Regarding outliers, one datapoint (-3.38) had a standardized residual that exceeded an absolute value of 3.0. Further, a statistically significant Levene’s test indicated a violation of the homogeneity of the residual variances for all groups, ($F[1, 191] = 4.539, p = .034$). *Because the intent of this analysis was to demonstrate how ANCOVA differs from mixed design ANOVA we proceeded with the analysis. Were this for “real research” we would have chosen a different analysis.*

8.5.3 Calculating the Omnibus ANOVA

We are ready to conduct the omnibus ANOVA.

Order of variable entry matters in ANCOVA. Thinking of the *controlling for* language associated with covariates, we firstly want to remove the effect of the covariate.

In the code below we are predicting attitudes toward Arabs at post1 from attitudes toward Whites at post1 (the covariate) and sitcom condition (Friends, Little Mosque).

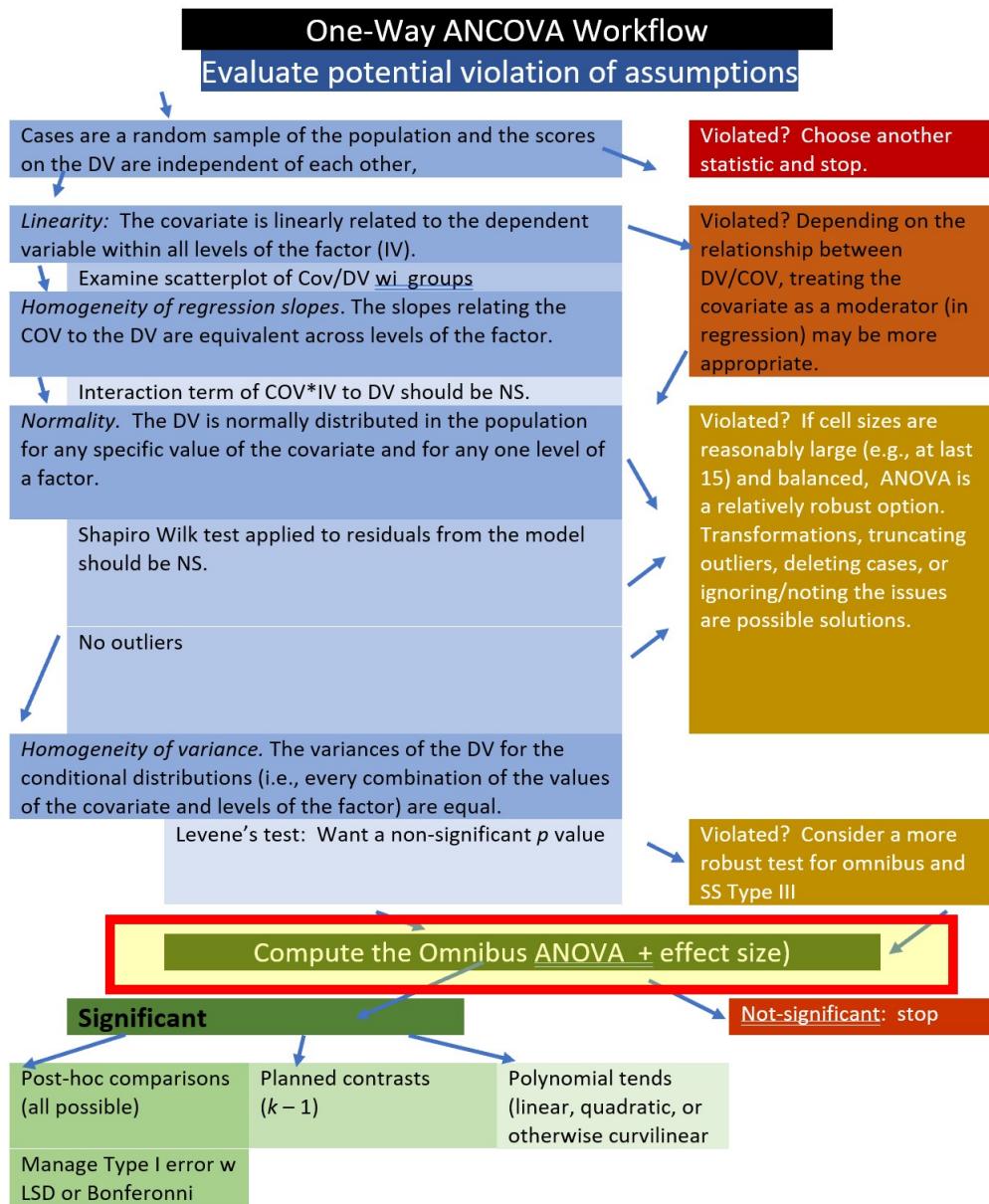


Figure 8.6: Image of the ANCOVA workflow, showing our current place in the process.

The *ges* column provides the effect size, η^2 where a general rule-of-thumb for interpretation is .01 (small), .06 (medium), and .14 (large) [Lakens, 2013].

```
WhCov_ANCOVA <- Murrar_wide %>%
  rstatix::anova_test(AttArabP1 ~ AttWhiteP1 + COND)
```

Coefficient covariances computed by hccm()

```
rstatix::get_anova_table(WhCov_ANCOVA)
```

ANOVA Table (type II tests)

	Effect	DFn	DFd	F	p	p<.05	ges
1	AttWhiteP1	1	190	0.014	0.907000000		0.0000722
2	COND	1	190	26.119	0.000000779	*	0.1210000

There was a non-significant effect of the attitudes toward Whites covariate on the attitudes toward Arabs at post-test, $F(1, 190) = 0.014$, $p = .907$, $\eta^2 < .001$. After controlling for attitudes toward Whites, there was a statistically significant effect in attitudes toward Arabs at post-test between the conditions, $F(1, 190) = 26.119$, $p < .001$, $\eta^2 = 0.121$. The effect size was moderate.

8.5.4 Post-hoc pairwise comparisons (controlling for the covariate)

With only two levels of sitcom condition (Friends, Little Mosque), we do not need to conduct post-hoc pairwise comparisons. However, because many research designs involve three or more levels, I will use code that evaluates them here.

```
pwc_cond <- Murrar_wide %>%
  rstatix::emmeans_test(AttArabP1 ~ COND, covariate = AttWhiteP1, p.adjust.method = "none")
```

```
# A tibble: 1 x 9
  term        .y. group1 group2   df statistic      p  p.adj p.adj.signif
* <chr>      <chr> <chr>  <chr> <dbl>    <dbl>  <dbl>  <dbl> <chr>
1 AttWhiteP1*C~ AttA~ Frien~ Littl~    190     -5.11 7.79e-7 7.79e-7 ****
```

Results suggest a statistically significant post-test difference between the Friends and Little Mosque sitcom conditions. With the script below we can obtain the covariate-adjusted marginal means. These are termed *estimated marginal means*.

```
emmeans_cond <- rstatix::get_emmeans(pwc_cond)
emmeans_cond
```

```
# A tibble: 2 x 8
  AttWhiteP1 COND      emmean     se     df conf.low conf.high method
  <dbl> <fct>      <dbl> <dbl> <dbl>    <dbl>    <dbl> <chr>
1    74.4 Friends    59.0  2.04   190    55.0    63.1 Emmeans test
2    74.4 LittleMosque 73.9  2.08   190    69.8    78.0 Emmeans test
```

As before, these means are usually different (even if only ever-so-slightly) than the raw means you would obtain from the descriptives.

```
descripts_cond <- psych::describeBy(AttArabP1 ~ COND, data = Murrar_wide,
  mat = TRUE)
descripts_cond
```

	item	group1	vars	n	mean	sd	median	trimmed	mad	min
X11	1	Friends	1	98	59.02351	21.65024	57.9955	59.31306	23.67045	8.297
X12	2	LittleMosque	1	95	73.92134	18.51082	74.4600	75.52858	15.98984	6.137
	max	range	skew	kurtosis	se					
X11	100	91.703	-0.0518848	-0.6252126	2.187005					
X12	100	93.863	-0.9798189	1.6335325	1.899170					

8.5.5 Toward an APA style results section

Tables with the means, adjusted means, and pairwise comparison output may be helpful. The *apa.cor.table()* function in the *apaTables* package is helpful for providing means, standarddeviations, and correlations.

```
apaTables::apa.cor.table(Murrar_wide[c("AttArabP1", "AttWhiteP1")], table.number = 2)
```

Table 2

Means, standard deviations, and correlations with confidence intervals

Variable	M	SD	1
1. AttArabP1	66.36	21.46	
2. AttWhiteP1	74.37	17.28	-.02 [-.16, .12]

Note. M and SD are used to represent mean and standard deviation, respectively.
Values in square brackets indicate the 95% confidence interval.

The confidence interval is a plausible range of population correlations that could have caused the sample correlation (Cumming, 2014).

* indicates $p < .05$. ** indicates $p < .01$.

```
# You can save this as a Microsoft word document by adding this
# statement into the command: filename = 'your_filename.doc'
```

Writing this output to excel files helped create the two tables that follow.

```
MASS::write.matrix(pwc_cond, sep = ",", file = "pwc_con.csv")
MASS::write.matrix(emmeans_cond, sep = ",", file = "emmeans_con.csv")
MASS::write.matrix(descripts_cond, sep = ",", file = "descripts_con.csv")
```

Ultimately, I would want a table that included this information. Please refer to the APA style manual for more proper formatting for a manuscript that requires APA style.

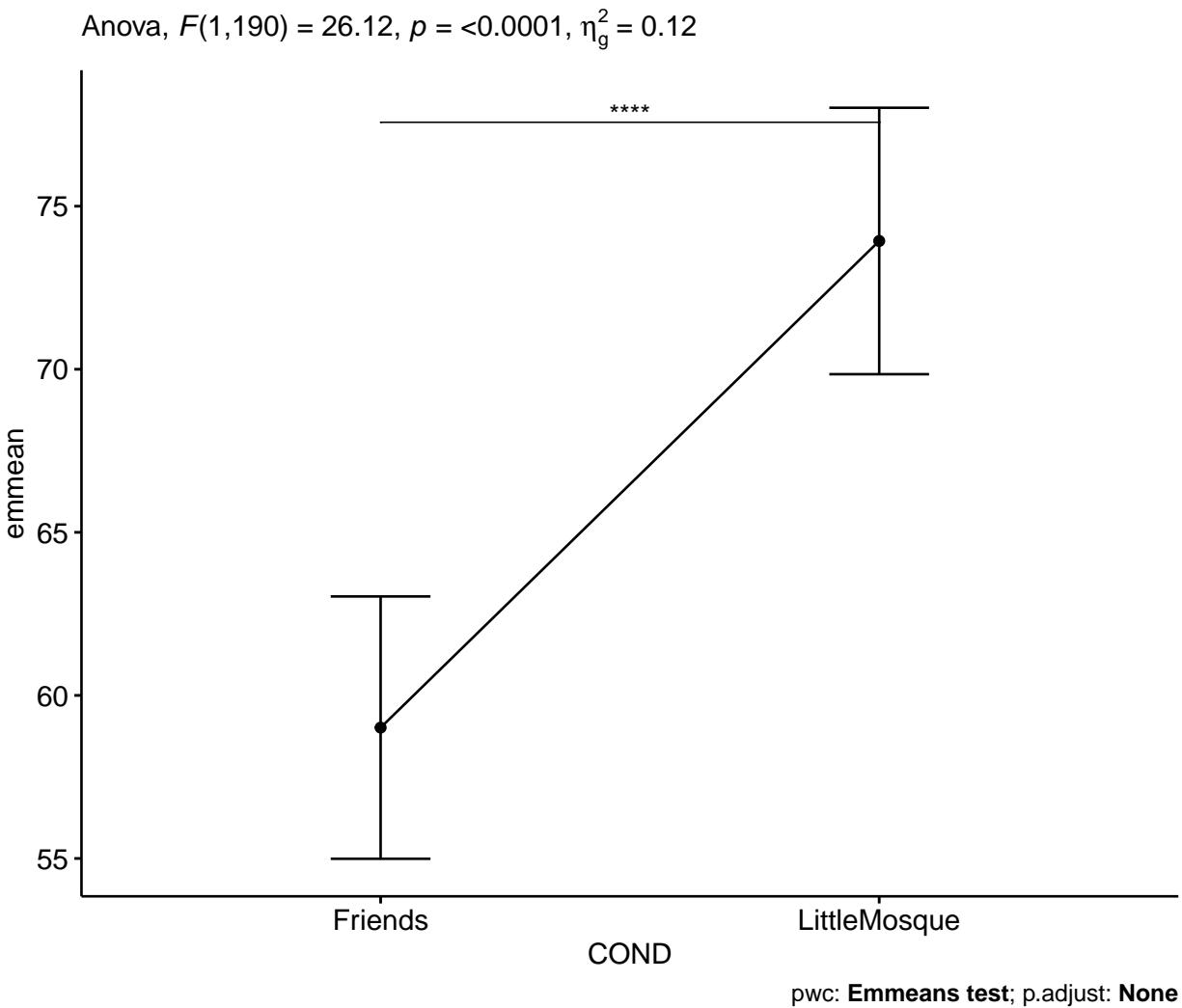
Table 1b

Unadjusted and Covariate-Adjusted Descriptive Statistics

Condition	Unadjusted	Covariate-Adjusted		
	<i>M</i>	<i>SD</i>	<i>EMM</i>	<i>SE</i>
Friends	59.02	21.65	59.03	2.04
Little Mosque	73.92	18.51	73.92	2.08

Unlike the figure we created when we were testing assumptions, this script creates a plot from the model (which identifies AttWhiteP1 in its role as covariate). Thus, the relationship between condition and AttArabP1 controls for the effect of the AttArabB covariate.

```
pwc_cond <- pwc_cond %>%
  rstatix::add_xy_position(x = "COND", fun = "mean_se")
ggpubr::ggline(rstatix::get_emmeans(pwc_B), x = "COND", y = "emmean") +
  geom_errorbar(aes(ymin = conf.low, ymax = conf.high), width = 0.2) +
  ggpubr::stat_pvalue_manual(pwc_B, hide.ns = TRUE, tip.length = FALSE) +
  labs(subtitle = rstatix::get_test_label(WhCov_ANCOVA, detailed = TRUE),
       caption = rstatix::get_pwc_label(pwc_cond))
```



Results

A one-way analysis of covariance (ANCOVA) was conducted. The independent variable, sitcom condition, had two levels: Friends, Little Mosque. The dependent variable was attitudes towards Arabs at pre-test. Preliminary analyses which tested the assumptions of ANCOVA were mixed. Results suggesting that the relationship between the covariate and the dependent variable did not differ significantly as a function of the independent variable ($F [1, 189] = 1.886, p = .171, \eta^2 = 0.010$) provided evidence that we did not violate the homogeneity-of-slopes assumption. In contrast, the Shapiro-Wilk test of normality on the model residuals was statistically significant ($W = 0.984, p = .029$). This means that we likely violated the assumption that the dependent variable is normally distributed in the population for any specific value of the covariate and for any one level of a factor. Regarding outliers, one datapoint (-3.38) had a standardized residual that exceeded an absolute value of 3.0. Further, a statistically significant Levene's test indicated a violation of the homogeneity of the residual variances for all groups, ($F[1, 191] = 4.539, p = .034$). *Because the intent of this analysis was to demonstrate how ANCOVA differs from mixed design ANOVA we proceeded with the analysis. Were this for “real research” we would have chosen a different analysis.*

There was a non-significant effect of the attitudes toward Whites covariate on the attitudes toward Arabs post-test, $F (1,190) = 0.014, p = .907, \eta^2 < .001$. After controlling for attitudes toward

Whites, there was a statistically significant effect in attitudes toward Arabs at post-test between the conditions, $F(1, 190) = 26.119, p < .001, \eta^2 = 0.121$. The effect size was moderately large. Means and covariate-adjusted means are presented in Table 1b.

8.6 More (and a recap) on covariates

Covariates, sometimes termed *controls* are often used to gain statistical control over variables that are difficult to control in a research design. That is, it may be impractical for polychotomize an otherwise continuous variable and/or it is impractical to have multiple factors and so a covariate is a more manageable approach. Common reasons for including covariates include [Bernerth and Aguinis, 2016]:

- they mathematically remove variance associated with nonfocal variables,
- the *purification principle* – removing unwanted or confusing variance,
- they remove the *noise* in the analysis to clear up the relationship between IV and DVs.

Perhaps it is an oversimplification, but we can think of three categories of variables: moderators, covariates, and mediators. Through ANOVA and ANCOVA, we distinguish between moderator and covariate.

Moderator: a variable that changes the strength or direction of an effect between two variables X (predictor, independent variable) and Y (criterion, dependent variable).

Covariate: an observed, continuous variable, that (when used properly) has a relationship with the dependent variable. It is included in the analysis, as a predictor, so that the predictive relationship between the independent (IV) and dependent (DV) are adjusted.

Bernerth and Aguinis [2016] conducted a review of how and when control variables were used in nearly 600 articles published between 2003 and 2012. Concurrently with their analysis, they provided guidance for when to use control variables (covariates). The flowchart that accompanies their article is quite helpful. Control variables (covariates) should only be used when:

1. Theory suggests that the potential covariate(s) relate(s) to variable(s) in the current study.
2. There is empirical justification for including the covariate in the study.
3. The covariate can be measured reliably.

Want more? Instructions for calculating a two-way ANCOVA are here: <https://www.datanovia.com/en/lessons/ancova-in-r/>

8.7 Practice Problems

In each of these lessons I provide suggestions for practice that allow you to select one or more problems that offer differing levels of difficulty. Whichever you choose, you will focus on these larger steps in one-way ANCOVA, including:

- test the statistical assumptions
- conduct an ANCOVA
- if the predictor variable has more than three or more levels, conduct follow-up testing
- present both means and covariate-adjusted means
- write a results section to include a figure and tables

8.7.1 Problem #1: Play around with this simulation.

Copy the script for the simulation and then change (at least) one thing in the simulation to see how it impacts the results.

- If ANCOVA is new to you, perhaps you just change the number in “set.seed(210813)” from 210813 to something else. Then rework Scenario#1, Scenario#2, or both. Your results should parallel those obtained in the lecture, making it easier for you to check your work as you go.
- If you are interested in power, change the sample size to something larger or smaller.
- If you are interested in variability (i.e., the homogeneity of variance assumption), perhaps you change the standard deviations in a way that violates the assumption.

8.7.2 Problem #2: Conduct a one-way ANCOVA with the DV and covariate at post2.

The Murra et al. [2018] article has three waves: baseline, post1, post2. In this lesson, I focused on the post1 waves. Rerun this analysis using the post2 wave data.

8.7.3 Problem #3: Try something entirely new.

Using data for which you have permission and access (e.g., IRB approved data you have collected or from your lab; data you simulate from a published article; data from an open science repository; data from other chapters in this OER), complete an ANCOVA.

8.7.4 Grading Rubric

Regardless which option(s) you chose, use the elements in the grading rubric to guide you through the practice. Using the lecture and workflow (chart) as a guide, please work through all the steps listed in the proposed assignment/grading rubric.

Assignment Component	Points Possible	Points Earned
1. Check and, if needed, format data	5	_____
2. Evaluate statistical assumptions	5	_____
3. Conduct omnibus ANCOVA (w effect size)	5	_____
4. If the IV has three or more levels, conduct follow-up tests	5	_____
5. Present means and covariate-adjusted means; interpret them	5	_____

Assignment Component	Points Possible	Points Earned
6. APA style results with table(s) and figure	5	_____
7. Explanation to grader	5	_____
Totals	35	_____

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