

# Pituitary prolactin drives transcriptional changes in the reproductive axis of parenting pigeons

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**Abstract** In species that provide parental care, successful rearing of offspring involves a shift from aggressive and sexual behaviors to more caring and nurturing ones. We hypothesize that these shifts in behavior are related to changes in gene expression that are controlled by internal mechanisms rather than by the external cues. We found that gene expression in the hypothalamus is fairly insensitive to the natural transition between egg and nestling stages; however, in the pituitary and gonads, gene expression showed much more plasticity over the course of reproduction. We show evidence that natural transitions in reproductive and parental behavior by mediated internal clock controlling gene expression in the pituitary and gonads but that the hypothalamus is very sensitive to external cues that signal a disruption in the natural parental care cycle. This research provides a deeper understanding of the interplay of genes, hormones, and parental care and has important implications for understanding molecular processes that underlie behavior transitions.

## Introduction

Reproduction is a critical period in an organism's life cycle. Parents must balance their own needs with the energetic costs and time-constraints required to successfully reproduce and rear their offspring to independence (cite). A complex series of evolutionarily conserved physiological processes underlie behaviors typically associated with reproduction in vertebrates, including courtship, nesting, and parental care. These processes are primarily mediated by the reproductive axis, which consists of the hypothalamus in the brain, the pituitary, and the gonads (i.e., the HPG). Our understanding of the physiological changes that adults navigate as they transition through reproductive states are primarily limited to a number of circulating hormones and a small set of well-known genes (cite). We have yet to come close to conceptualizing the larger symphony of reproductive events that occur to yield reproductive and parental care behaviors.

Using the model of a biparental vertebrate species, the rock dove (*Columba livia*), our team has previously demonstrated the dynamic nature of gene activity within the reproductive or HPG axis. MacManes et al.<sup>(5)</sup> reported that HPG gene activity varied by sex, with females exhibiting higher levels of overall gene expression than males. Calisi et al.<sup>(1)</sup> reported that HPG gene activity varied response to restraint-stress, and Austin et al. identify genes that responds to both corticosterone treatment and restraint stress. While each of these studies advanced our understanding of sex-differences and transcriptional plasticity in the reproductive axis, none of these studies examined activity of the reproductive axis of bird that are actively reproducing or caring for offspring. Studying parental care in both sexes of mammalian species is challenging because only the females are able to use lactation to feed their young, and the physiological mechanisms regulating this process are hard to control for. The rock dove is an excellent system for studying reproduction and parental care in both sexes because both males and females feed their young an energy-rich secretion produced from the epithelial lining of their crops, known as crop milk.

In this study, we investigate the activity of the HPG axis over the course of reproduction and parental care by characterizing gene expression as birds transition from non-breeding to nest-building, as they lay and incubate their clutches, and finally as they become parents and raise their chicks. Then, we conducted a series of nest manipulations to ascertain constraints associated with biological timing versus input from external stimuli. We conducted a comprehensive series of analyses examine broad patterns of variation as well as the activity of specific genes in specific tissues. We have also made our data and scripts available so that others can reuse the data to ask different questions or reproduce our results. This research provides a deeper understanding of the interplay of genes, hormones, and parental care and has important implications for understanding molecular processes that underlie behavioral transitions. reviewed in<sup>(4)</sup> say this, but other say that reviewed in<sup>(2)</sup>.

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## Results

### *Characterization of parental care*

To characterize transcriptional activity, we first examined broad patterns of variation in gene expression in the hypothalamus, pituitary, and gonads of male and female rock doves across a series of timepoints that encapsulate important transitions in reproduction and parental care (Fig. 1a). t-Distributed Stochastic Neighbor Embedding (t-SNE) is a nonlinear, dimension reduction technique used to visualize high-dimensional data by giving each datapoint a location in a two-dimensional map (van der Maaten and Hinton 2008). t-SNE is widely used in single-cell data process for identify related cell types. Here, we use t-SNE to identify variation associated with tissue, sex, and treatment (Fig. 1j).

Our results suggest that tissue is the greatest source of variation in the dataset, with hypothalamic, pituitary, and gonadal samples all forming distinct clusters, with only a few pituitary and gonad samples falling outside the 95% confidence interval (Fig. 1b). Sex-differences are most pronounced in the gonads with male and female samples form two discrete clusters (Fig. 1c). Treatment accounts for much less of the variation than tissue and sex, and even after removing sex as a variable, treatment effects in all three tissues are hard to visualize simultaneously (Fig. 1d,e), so we next looked at each tissue and sex separately. In the hypothalamus, control samples form tight clusters that stands out from all other timepoints that are largely overlapping in space (Fig. 1h). The pituitary shows the strongest treatment effect with one cluster of the “earlier” parental timepoints (nest-building, lay, incubation days 3 and 9), a second cluster of “later” parental timepoints (incubation day 17, hatch, and nestling care days 5 and 9; Fig. 1i). Like the hypothalamus, in the gonads, the control timepoints appear to be much less variable than the parental timepoints, but there is no clear separation of gonadal samples by parental stage (Fig. 1j). While the tSNE analysis reveals broad patterns across all levels of the experiment, additional tools are needed to hone in on the specific differences associated with parental care in each tissue. Differential gene expression analysis can shed insight into specific differences between treatment groups and thus deepen our understanding of the transitions between reproduction and parental care behaviors. We used DESeq2 to identify genes whose expression changed significantly between sequential timepoints in each tissue and sex separately (Fig. 2a).

In the hypothalamus only two sequential stage comparisons results in a sizable change in expression; in females, 1927 genes changes their expression between hatch to nestling care day 5 (with 1076 decreasing and 851 increased their expression), and in the male hypothalamus, 87 genes were differentially expressed between incubation days 9 and 17 (with 38 decreasing and 49 increasing their expression (Fig. 2b)). In the pituitary, differential gene expression differences are greatest when comparing incubation days 9 and 17, and this difference is more pronounced in females (Fig. 2c). The next largest difference in the pituitary is between hatch and n5. The female pituitary shows moderate changes from nest-building to lay to incubation day 3 (Fig. 2c). The male pituitary changes more than the female pituitary changes between nestling care days 5 and 9 (Fig. 2c). About 2000 genes also change their expression in the female pituitary and about 425 in the male pituitary when comparing incubation days 9 and 17 (Fig. 2). In the female gonads, 270 genes are decreased while only 56 are increased when comparing incubation days 9 and 17 (Fig. 2). Many more significant changes in gene expression in the female gonad occur between nest-building and lay and between lay and clutch completion (incubation day 3) than in any other comparison (Fig. 2).

### *Internal physiology versus external stimuli hypothesis*

We show that samples from the pituitary on incubation day 17 cluster more closely with hatch and nestling care day 5 samples than with incubation days 3 and 9. This suggests that, in the pituitary, internal mechanisms that prepare for chick arrival have the greatest effect on gene expression rather than suggesting that gene expression responds to changes in the external environment.

Blah 3, blah, 4 blah.

### *Potential implications for cancer research*

These results confirmed prolactin (PRL) gene expression fluctuates throughout parental care in a manner that is consistent with its role in promoting lactation and maintaining parental behaviors. Interestingly, we identified about 100 genes whose expression was correlated with PRL, including (BRCA1, which is associated with breast cancer). Additionally, we identified thousands of genes whose expression changed over the course of parental care. Exploring the relationship between PRL and other differentially expressed genes could provide important insights into the symphony of gene expression that regulates behavior at the organismal and cellular level.

### *Software for data exploration and hypothesis testing*

Data accessibility and reproducibility are key challenges for today’s researchers. Using Binder and Shiny, we have provided a means for scientists, physicians, and the general public to explore the data, code, and results from this project at <https://raynamharris.shinyapps.io/musicalgenes/>.

Like Hipposeq (Cembrowski et al *elife* 2016, 2018), this tool allows quick and easy data exploration of RNAseq data from multiple cell types that relate to brain function.

## Discussion

This RNA-sequencing project was designed to characterize the reproductive axis of bi-parental pigeons across a characteristic reproductive cycle and through the parental main parental stages of egg incubation and parental care using data- and hypotheses driven approaches. We hypothesized that differences in gene expression would be driven by either internal physiology or external stimuli. In the pituitary, we found that the most explanatory source of variation in gene expression was the expression level of prolactin (PRL). Circulating levels of prolactin in the blood in this species are known to rise around day nine and peak on or the day before hatch. Our results confirm this and also identified one hundred genes whose expression is highly correlated with prolactin. Today, this variation, which changes more within a parental stage (e.g. egg incubation or nestling care) suggests that internal physiology drives changes in gene expression that precede changes in external stimuli that lead to behavioral changes (e.g. crop milk or regurgitation). We predicted that either sex steroid peptide hormones would play a role in governing the reproductive transitions, but we did not anticipate discovering a link between PRL expression to genes that are involved with DNA repair and regulation of the cycle. Finally, we aimed to make these data and analyses FAIR (findable, accessible, interoperable, and reproducible), open and reproducible to encourage others to reuse our data to test new hypotheses or explore the data with a different perspective or toolset. The scripts that made the figures and tables are browseable online thanks to RStudio, GitHub and Binder. The data can be visualized and sonified (turned into sound) interactively thanks to RStudio, GitHub, and Shiny.

## Materials and Methods

### *Animal Care*

All animal care and use comply with the Public Health Service Policy on Humane Care and Use of Laboratory Animals and were approved by the University of California, Davis IACUC permit #18895. Birds were housed at the University of California, Davis in enclosed aviaries with ~8 sexually reproductive adult pairs per aviary. All adult birds are uniquely associated with nests by their unique color band combination. Food and water were provided *ad libitum*, and nests were monitored daily.

### *Experimental design and tissue processing*

To characterize reproduction and parental care, birds were sampled at 8 timepoints across the parental care cycle (Fig. 1a) as previously described in Austin et al 2020. Briefly, nest-building pairs where at least one individual was seen carrying nesting material or shaping a nest (bldg); the day the 1st egg was laid (lay), clutch completion (also known as incubation day 3 (inc.d3)), mid-incubation (inc.d9), late incubation (inc.d17), the day the first chick hatched (where hatch), nestling care days 5 and 9 (n5 and n9). Additionally, a non-breeding (control) timepoint was added using control birds from previously published studies (2,3).

To test our hypothesis that gene expression is governed more by internal mechanisms than external cues, we conducted multiple offspring replacement or removal manipulations... Add details here.

To control for circadian rhythm confounds, each member of the pair was collected between 0900-1200 (PST). We sampled a total of 331 birds (16 treatments with ~10 pairs per treatment). Pairs were captured and collected simultaneously within 5 min of entering their cage, were anesthetized using isoflurane until unresponsive (<2 min), and then decapitated. Trunk blood, brains, pituitaries, and gonads were collected. Processing and analysis of trunk blood is described in Austin et al. 2020.

### *RNA sequencing analysis*

Processing of brains, pituitaries, and gonads for RNA sequencing is described in detail in Lang et al. 2020. Briefly, RNA from the hypothalamus, pituitary, and gonads was processed for Illumina sequencing using the NEB Next Ultra Directional RNA Library Prep Kit. Reads were pseudomapped (insert Kalisto citation) to the Rock Dove transcriptome v1.1.0 whose transcripts were annotated with genes from *Gallus gallus* genome v5.

Statistical analyses and modeling were performed using R<sup>(6;8)</sup>. Limma was used to process all samples using the model  $\sim \text{tissue} * \text{sex} * \text{treatment}$  (citation). t-Distributed Stochastic Neighbor Embedding (t-SNE) analysis was conducted using a perplexity of 10<sup>(7)</sup>. Principal Component Analysis (PCA) was conducted using the top 500 most varying genes (citation). DESeq2 was used to model gene expression for sex and tissue combination separately using the model  $\sim \text{treatment}$  (Love2014). Weighted correlation network analysis (WGCNA) was used to identify sets of genes with similar patterns of gene expression over the course of reproduction and parental care (citation). ANOVAs were used to test whether specific genes were differentially expressed between sequential

timepoints.

#### Data availability

RNA sequencing data for available through the European Nucleotide Archive (ENA) project ID PRJEB16136 and XXXXX. Code and data for reproducing the results described in the manuscript are available at <https://github.com/macmanes-lab/DoveParentsRNAseq> (upload to zenodo and cite). Data can be explored in the cloud using Binder<sup>(3)</sup>.

## Acknowledgements

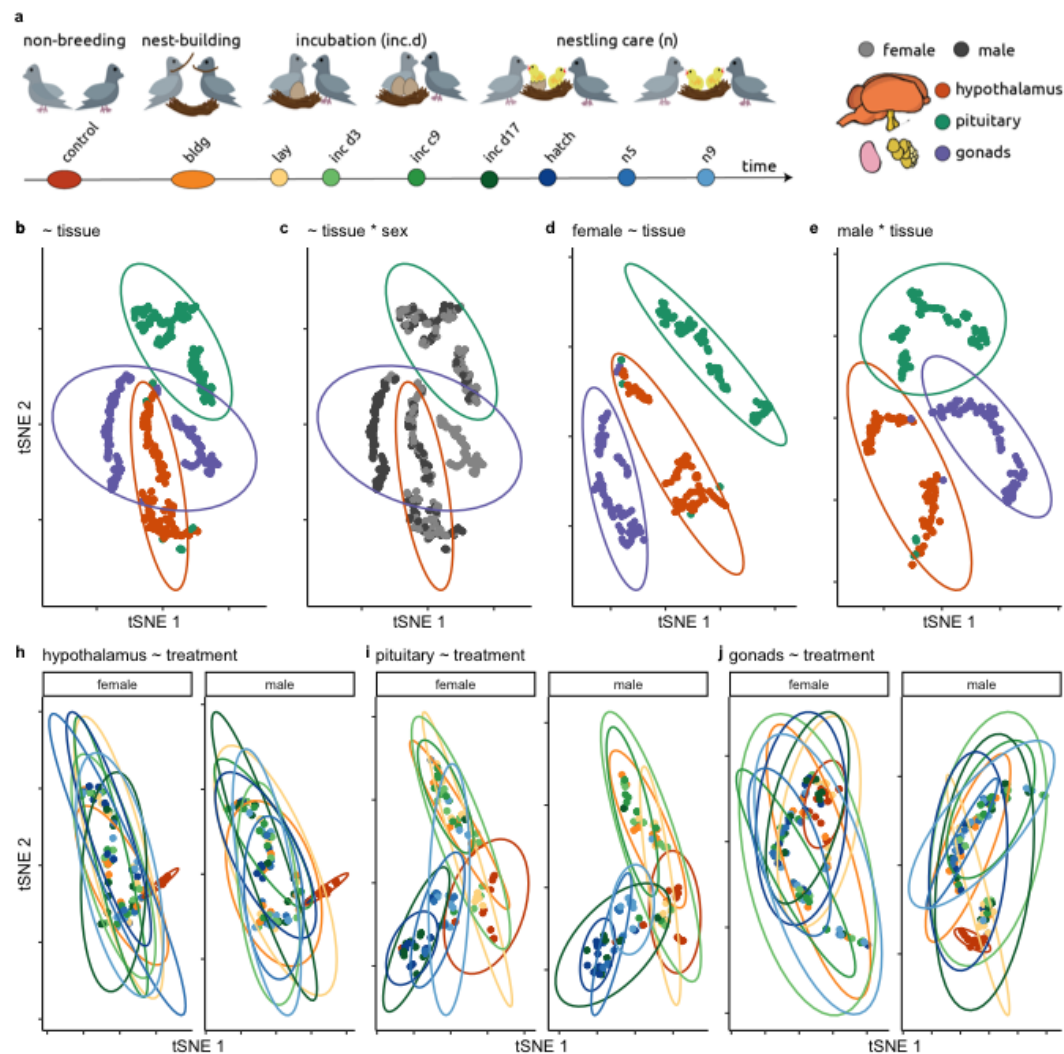
Thanks to Owen for discussion of data sonification. Thanks to DIB lab members for software and methods discussion (Titus, Taylor, Luiz, Amanda, Alicia, Picasso). We thank Pacha for help with database and Shiny.

We thank Irene, Erica, Beth Krestoff and Tiffany Chen for assistance in managing the aviary and collecting nest stage data. We thank Candice Lee, Annie Bond, and Tanner Feustel and others?? for assistance with tissue collections. We thank Dr. Fred Angelier for performing the prolactin RIA. This research was supported by NSF IOS 1455960 to R.M.C. and M.D.M. Author contributions: RMC designed the experiment. RMC and MDM acquired funding. SHA contributed to the experimental design by adding several time points. SHA, with the assistance of VF, AB, and RMC, conducted the data collection. SHA and a team of trained undergraduate students biopsy punched the hypothalamic and lateral septum. ASL extracted the RNA, created the libraries, and conducted all early bioinformatic work. RMH conducted all later analyses of the genomic data. RMH and SHA drafted the manuscript and all other authors provided edits.

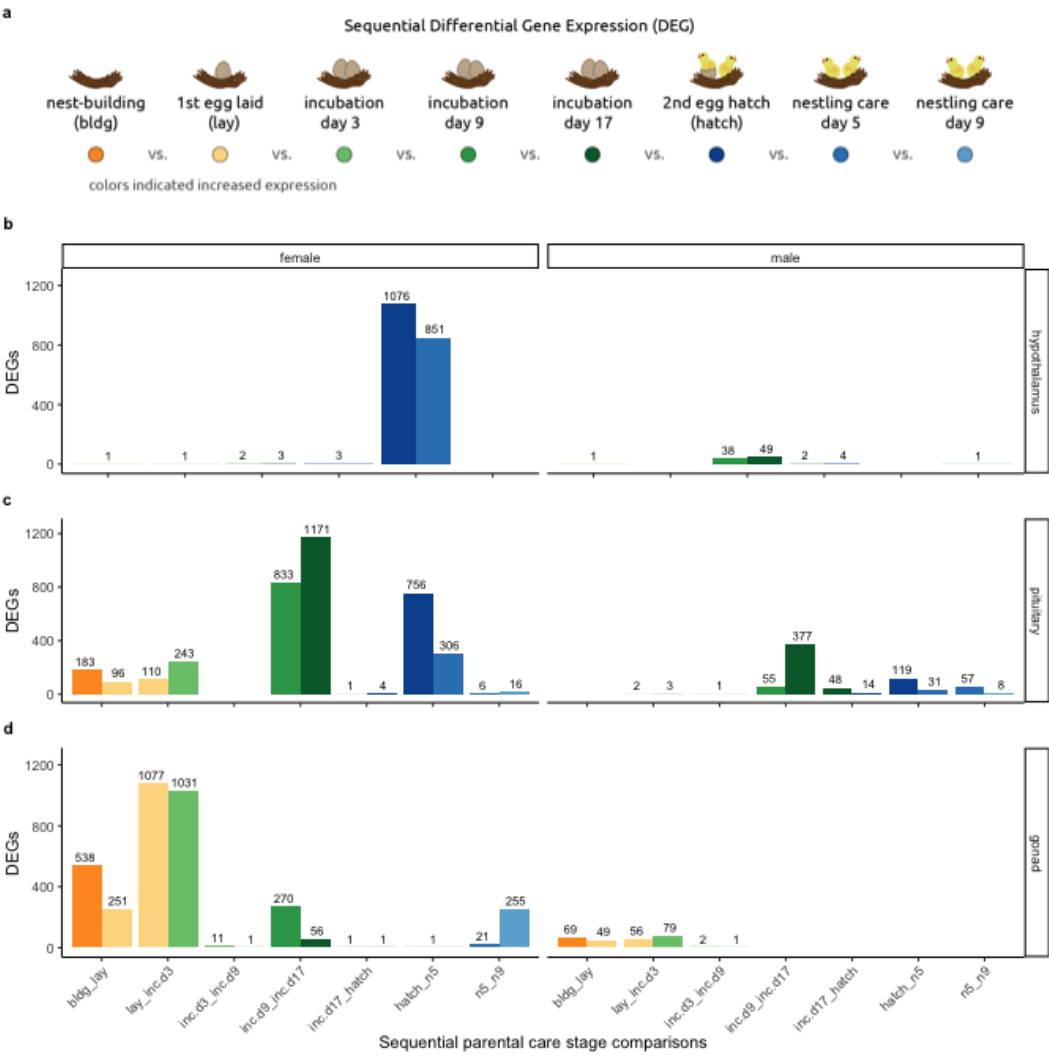
## References

## Bibliography

- [1] R. M. Calisi, S. H. Austin, A. S. Lang, and M. D. MacManes. Sex-biased transcriptomic response of the reproductive axis to stress. *Hormones and Behavior*, 100:56–68, apr 2018. ISSN 0018-506X. doi: 10.1016/J.YHBEH.2017.11.011. URL <https://www.sciencedirect.com/science/article/pii/S0018506X17302696?via=ihub>. [p1]
- [2] C. Dulac, L. A. O’Connell, and Z. Wu. Neural control of maternal and paternal behaviors. *Science*, 345(6198):765–770, aug 2014. ISSN 10959203. doi: 10.1126/science.1253291. [p1]
- [3] P. Jupyter, M. Bussonnier, J. Forde, J. Freeman, B. Granger, T. Head, C. Holdgraf, K. Kelley, G. Nalvarte, A. Osheroff, M. Pacer, Y. Panda, F. Perez, B. Ragan-Kelley, and C. Willing. Binder 2.0 - Reproducible, interactive, sharable environments for science at scale. In F. Akici, D. Lippa, D. Niederhut, and M. Pacer, editors, *Proceedings of the 17th Python in Science Conference*, pages 113–120, 2018. doi: 10.25080/Majora-4af1f417-011. [p4]
- [4] R. W. Lea and P. J. Sharp. Concentrations of plasma prolactin and luteinizing hormone following nest deprivation and renesting in ring doves (*Streptopelia risoria*). *Hormones and Behavior*, 23(2): 279–289, jun 1989. ISSN 10956867. doi: 10.1016/0018-506X(89)90067-6. [p1]
- [5] M. D. MacManes, S. H. Austin, A. S. Lang, A. Booth, V. Farrar, and R. M. Calisi. Widespread patterns of sexually dimorphic gene expression in an avian hypothalamic-pituitary-gonadal (HPG) axis. *Scientific Reports*, 7(1):1–13, apr 2017. ISSN 20452322. doi: 10.1038/srep45125. [p1]
- [6] R Development Core Team. R: A language and environment for statistical computing, 2013. URL <http://www.r-project.org/>. [p3]
- [7] L. Van Der Maaten and G. Hinton. Visualizing Data using t-SNE. Technical report, 2008. [p3]
- [8] H. Wickham and G. Grolemund. *R for Data Science: Import, Tidy, Transform, Visualize, and Model Data*. 2016. ISBN 978-1491910399. [p3]



**Figure 1:** Broad patterns of variation in hypothalamic, pituitary, and gonadal gene expression in male and female rock doves across reproductive and parental transitions. a) Experimental design. b) t-SNE plot highlighting tissue differences in gene expression. Each dot represents 1 sample and is colored by the source tissue (hypothalamus: orange, pituitary: teal, gonads: purple). c) t-SNE plot highlighting sex differences in tissue and sex, as shown in light grey (females) and dark grey (males). t-SNE plots highlight treatment effects in females (d), males (e), the hypothalamus (h), pituitary (i), and gonads (j), with samples colored by parental stage (control: red or , nest-building: orange, lay: yellow; inc.d3: light green, inc.d9: green, inc.d17: dark green, hatch: dark blue, n5: blue, and n9: light blue).



**Figure 2:** Differential gene expression differences between sequential parental stages. a) We calculated the differential gene expression between each consecutive parental stage samples, for a total of 8 pair-wise comparisons for each sex in each tissue. The y-axis of these bar plots displays the total number of differentially expressed genes for each comparison in the hypothalamus (b), pituitary (c), gonads (d). Bars are colored by the direction of the change in gene expression. Control: control: red or , nest-building (bldg): orange, lay: yellow; incubation day 3 (inc.d3): light green, inc.d9: green, inc.d17: dark green, hatch: dark blue, nestling care day 5 (n5): blue, and n9: light blue.



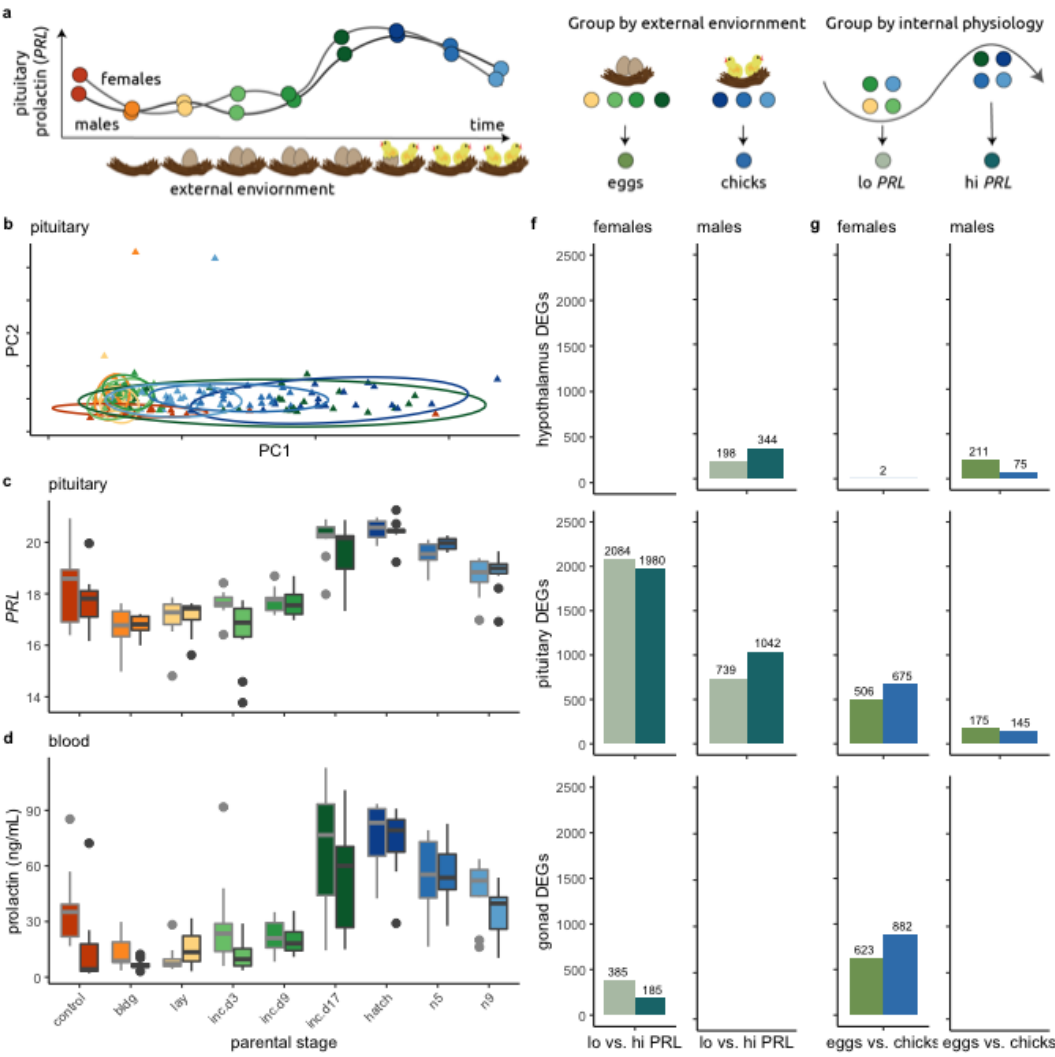


Figure 3: Another figure

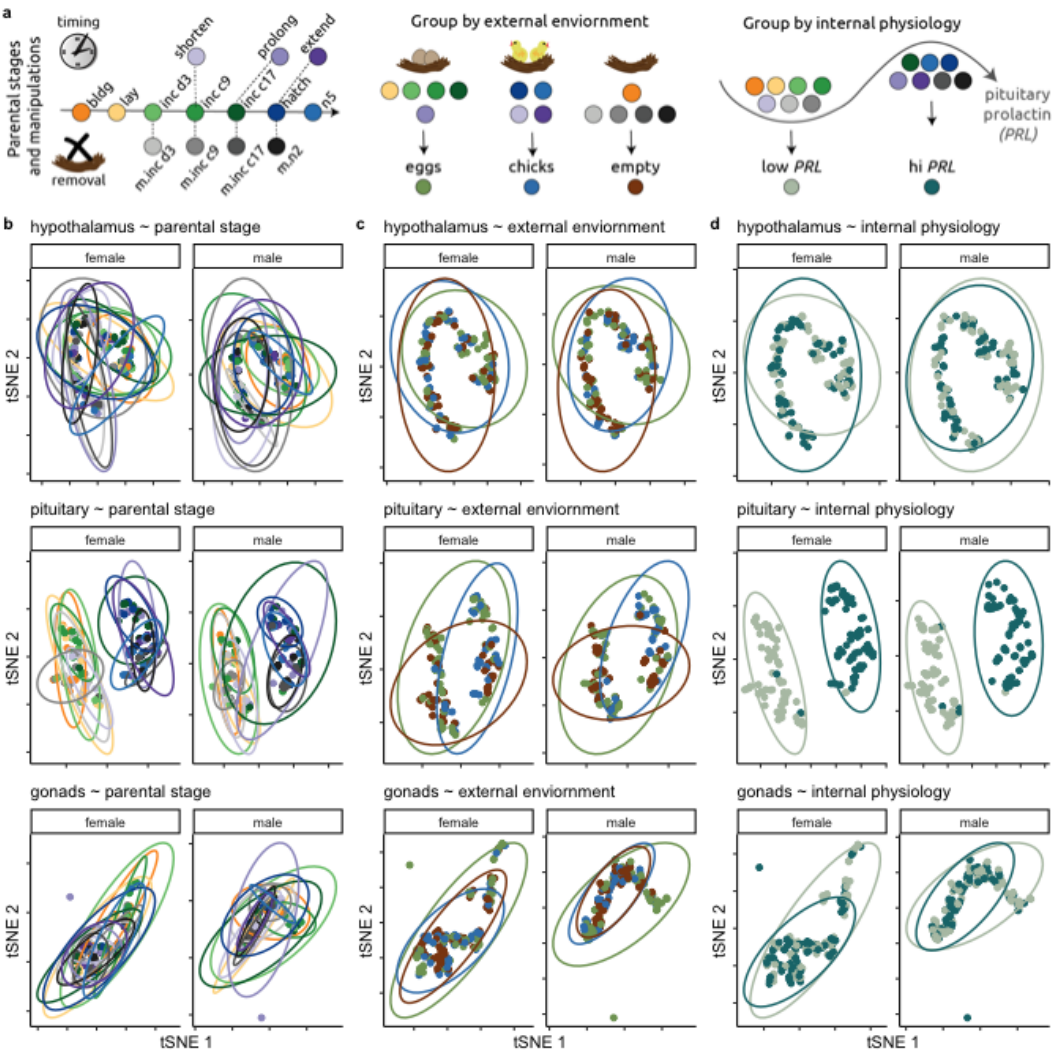


Figure 4: Another another figure



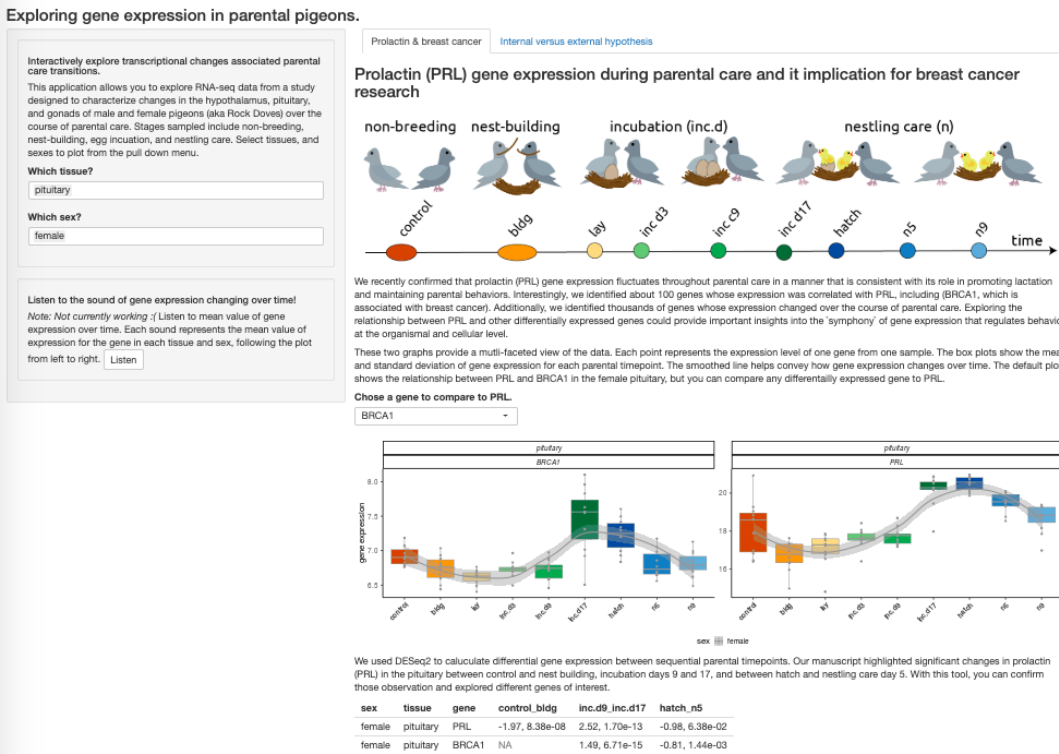


Figure 5: R shiny, image 1

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