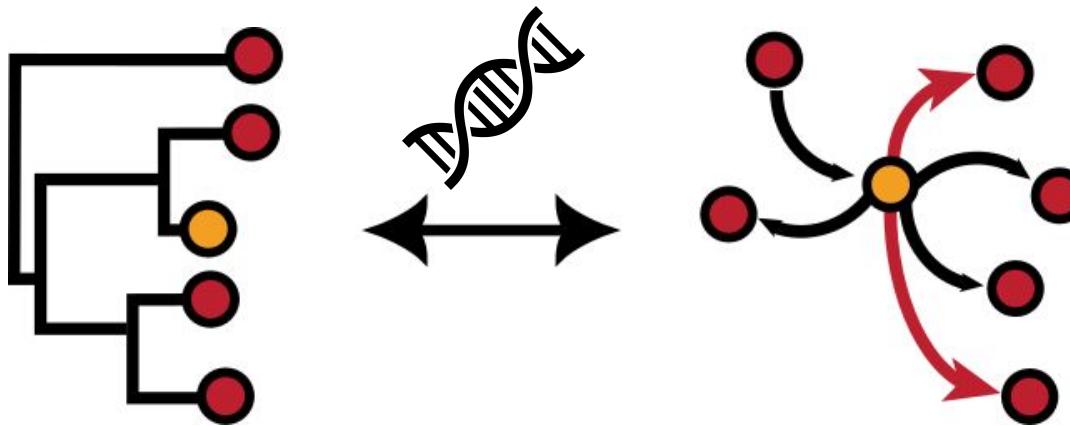
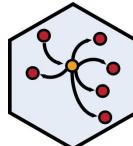


EPAH6052: Pathogen Genomic Epidemiology



DALHOUSIE
UNIVERSITY

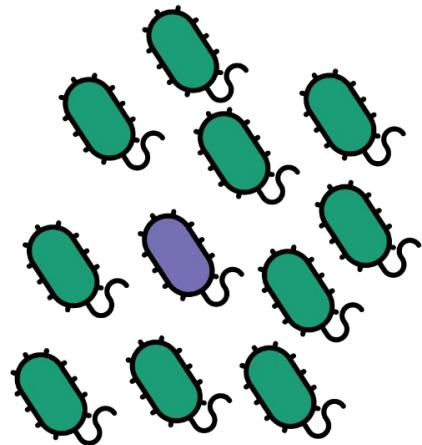
Finlay Maguire
Dalhousie University
Shared Hospital Laboratory
Public Health Alliance for Genomic Epidemiology



Outcomes

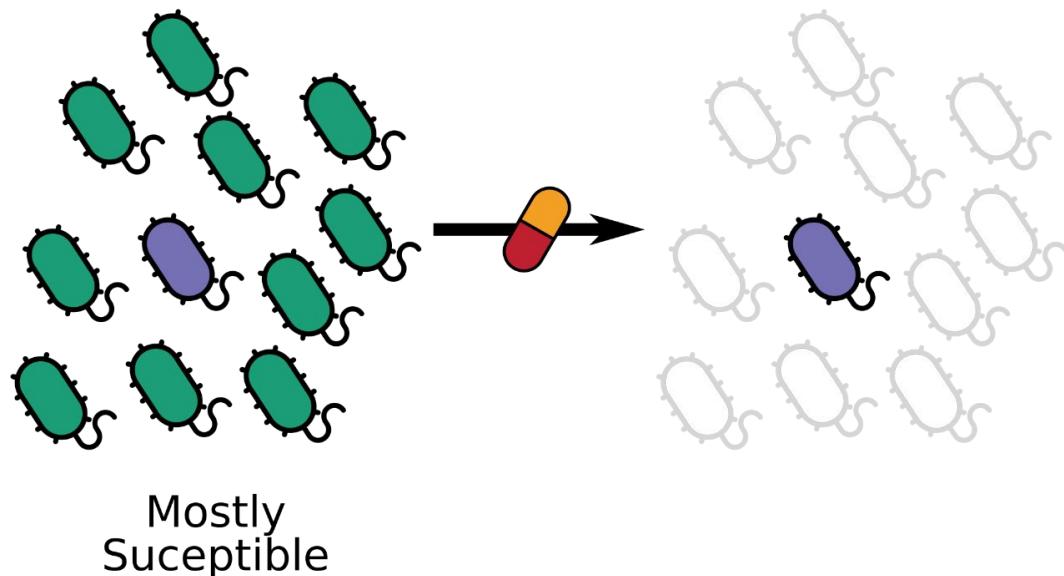
- Explain how evolution and infectious disease epidemiology are connected
- Identify what additional information genomic data can provide
- Give example of how genomics can be used for diagnostics
- Explain how you can infer an evolutionary tree (phylogeny)
- Provide examples of processes which determine the shape of a phylogeny
- Articulate the role of Bayesian models in genomic epidemiology
- List at least 3 ways in which a phylogeny can be used in epidemiology

Agents of infectious diseases undergo evolution

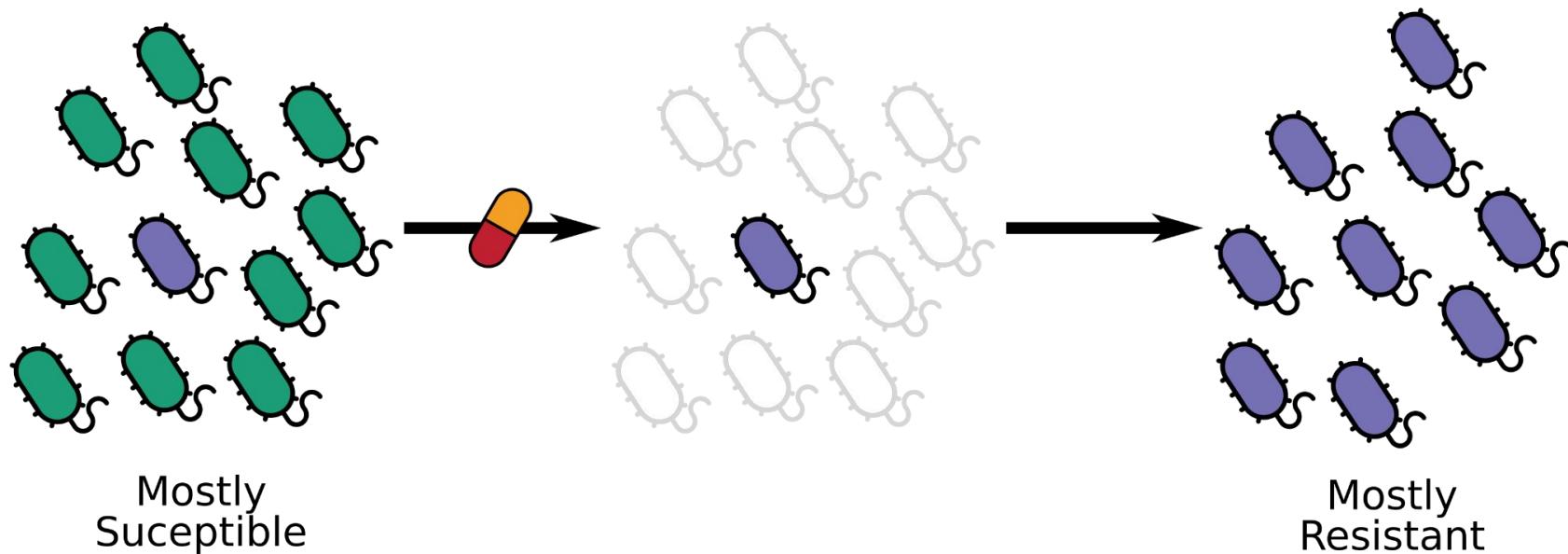


Mostly
Susceptible

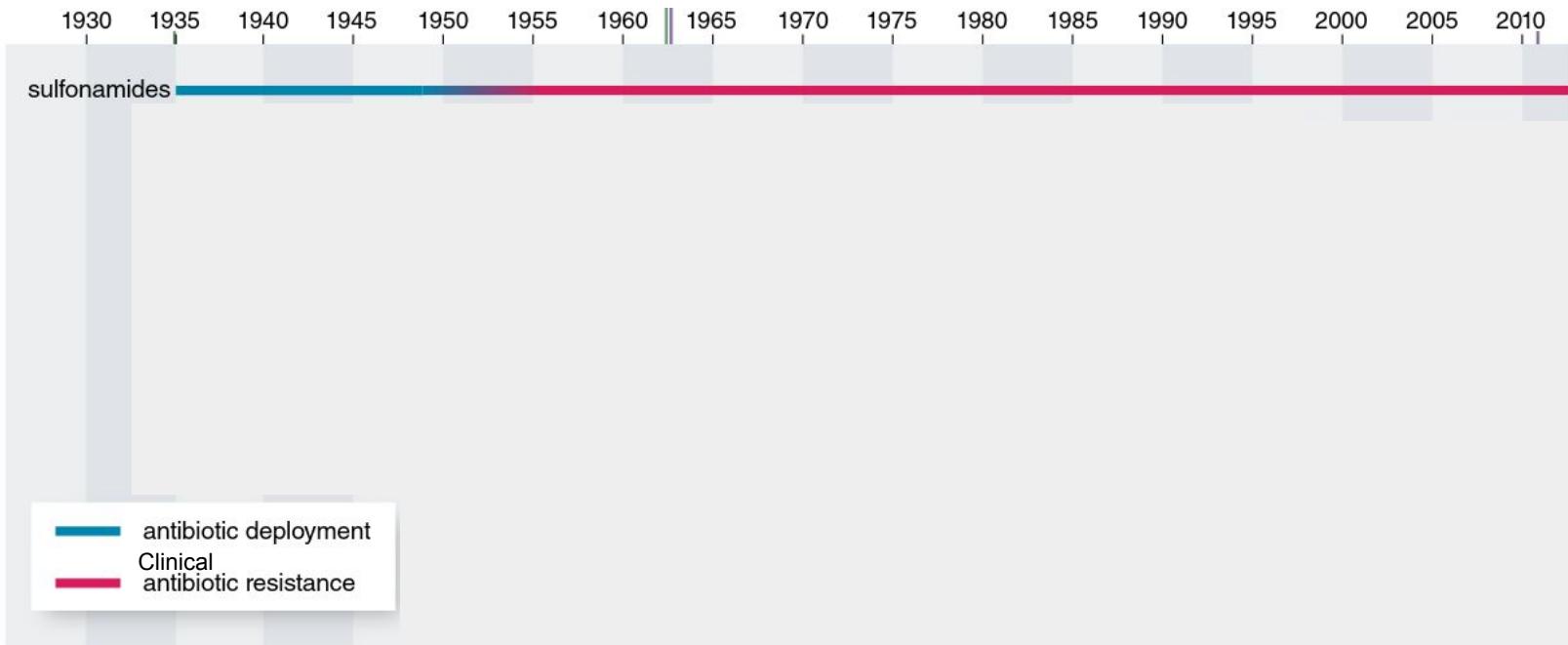
Agents of infectious diseases undergo evolution



Agents of infectious diseases undergo evolution

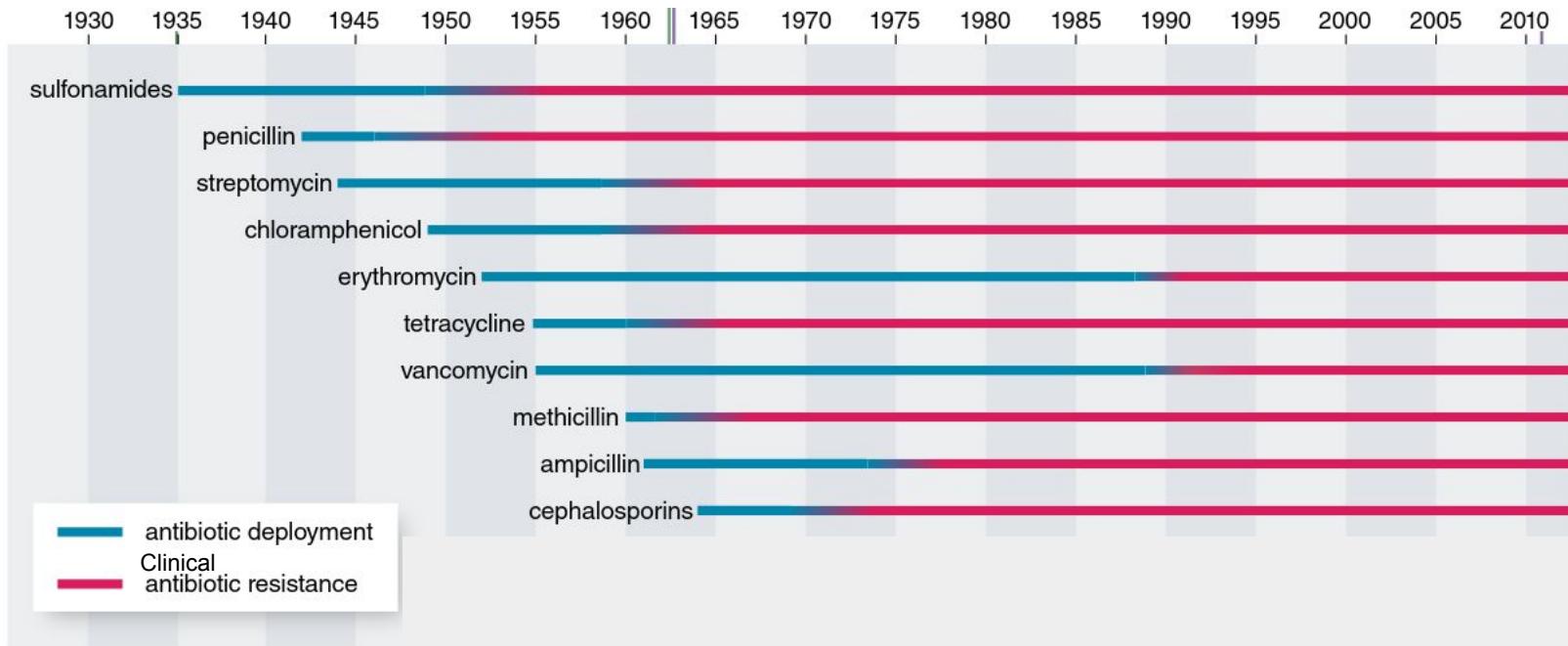


Pathogen evolution impacts treatment



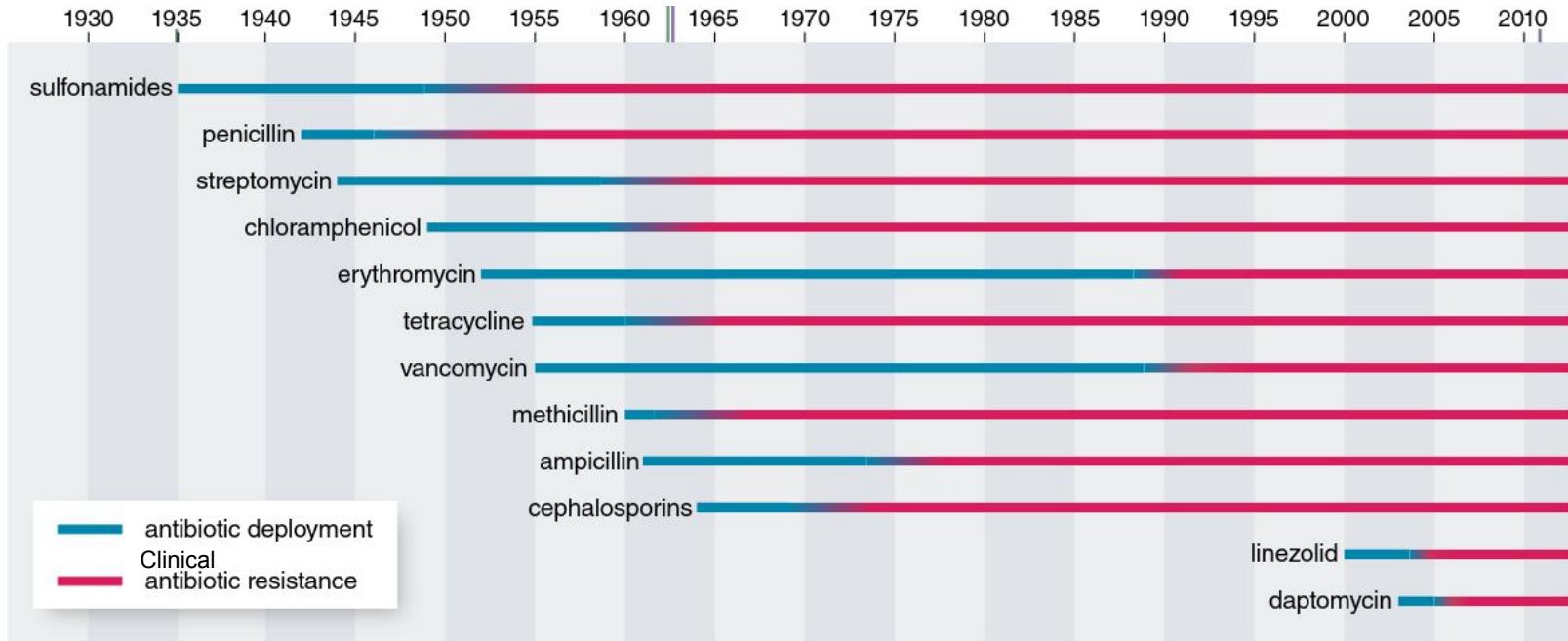
10.1511/2014.106.42

Pathogen evolution impacts treatment



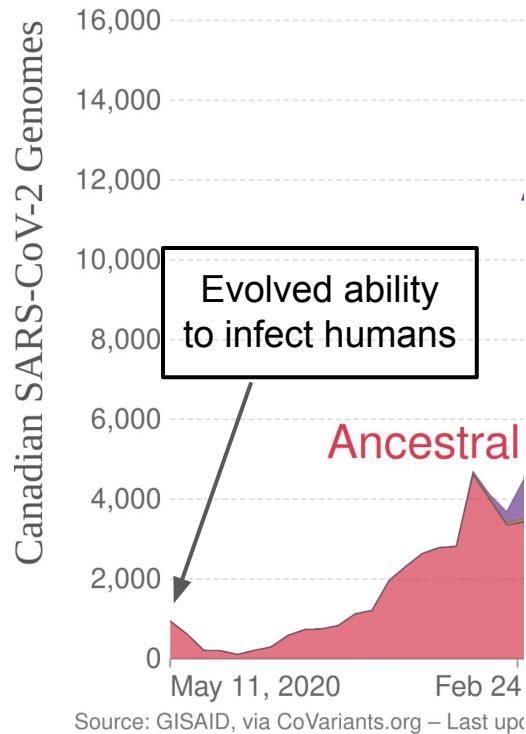
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Pathogen evolution impacts treatment

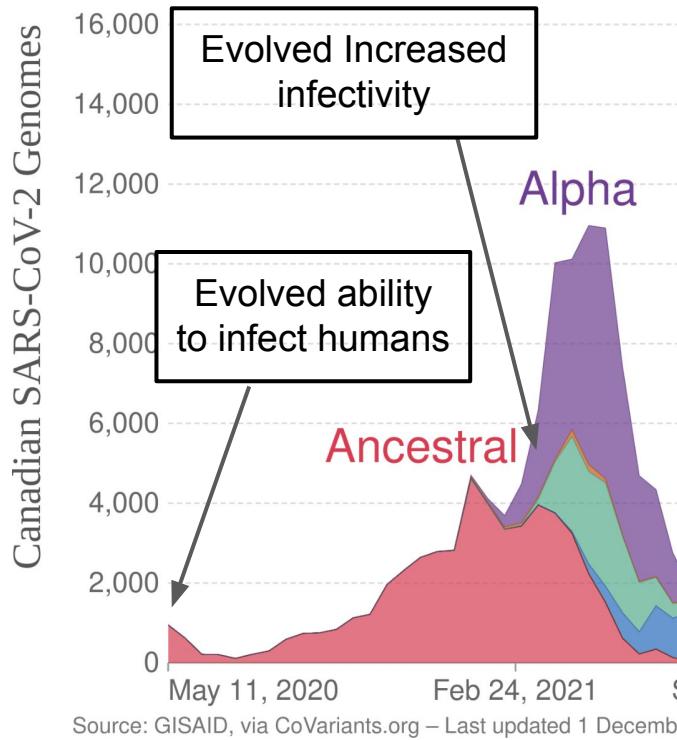


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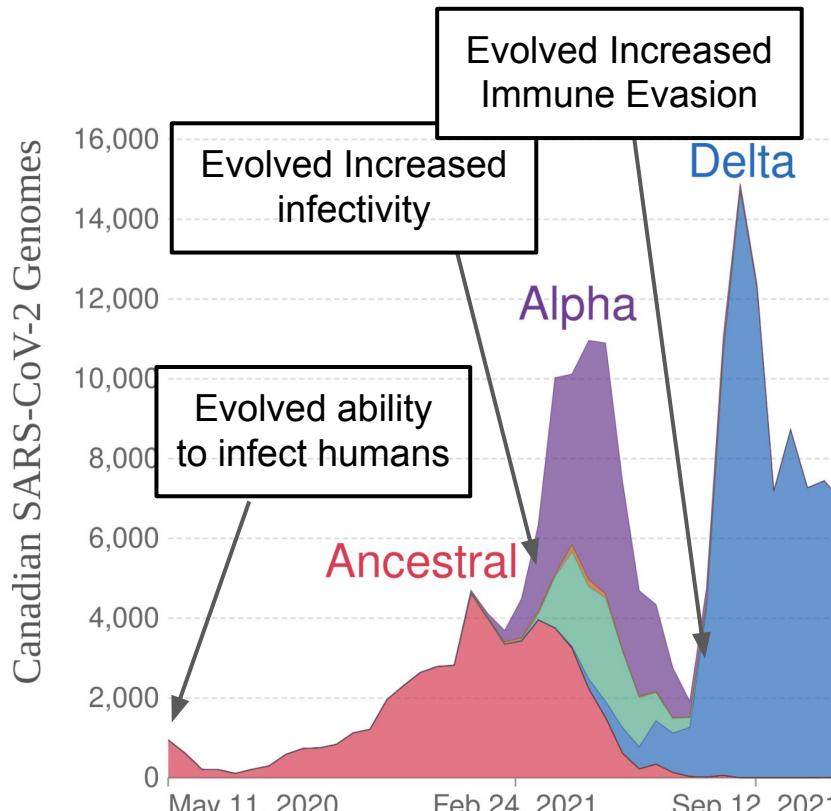
Evolution drives cases: SARS-CoV-2 waves



Evolution drives cases: SARS-CoV-2 waves

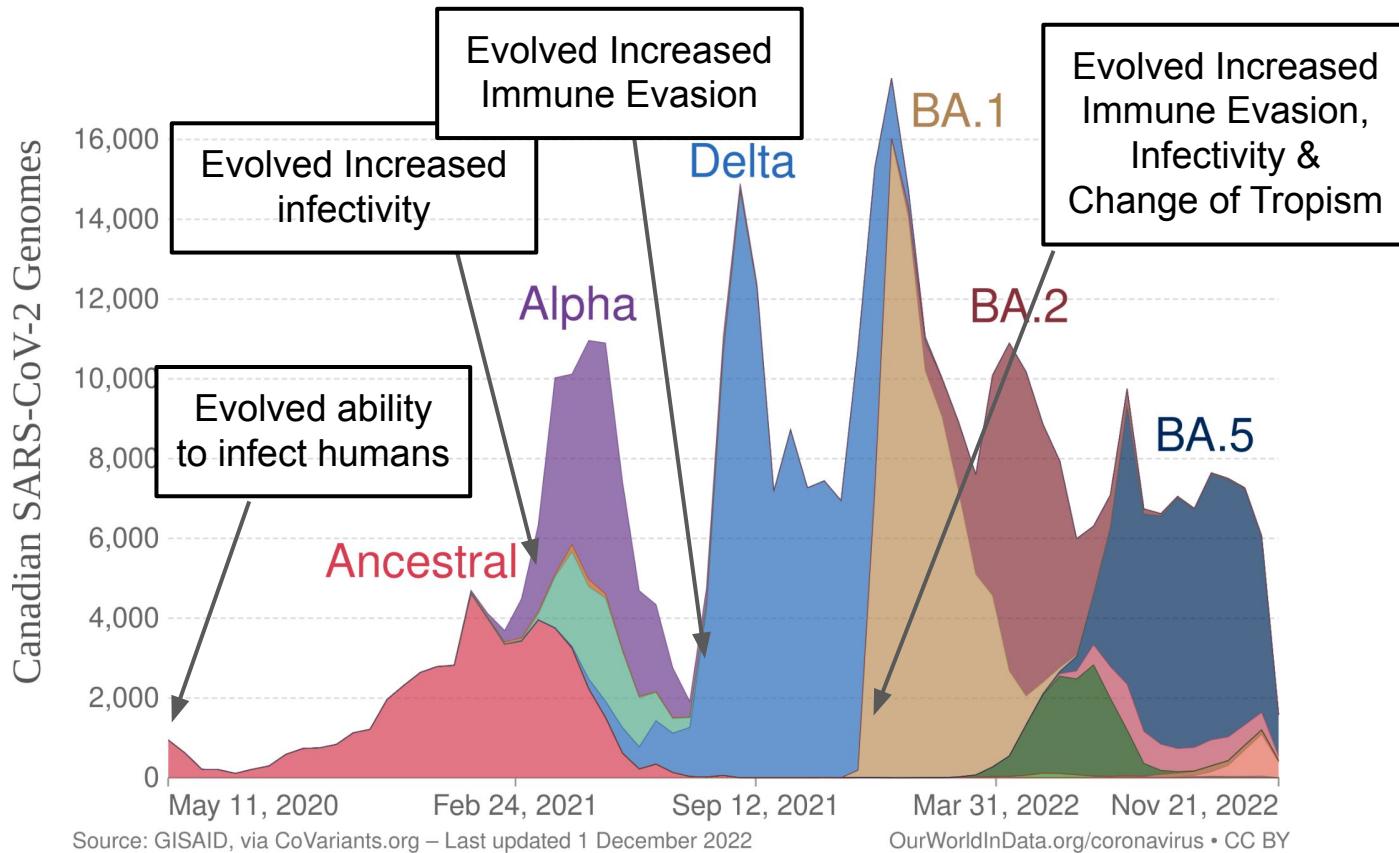


Evolution drives cases: SARS-CoV-2 waves

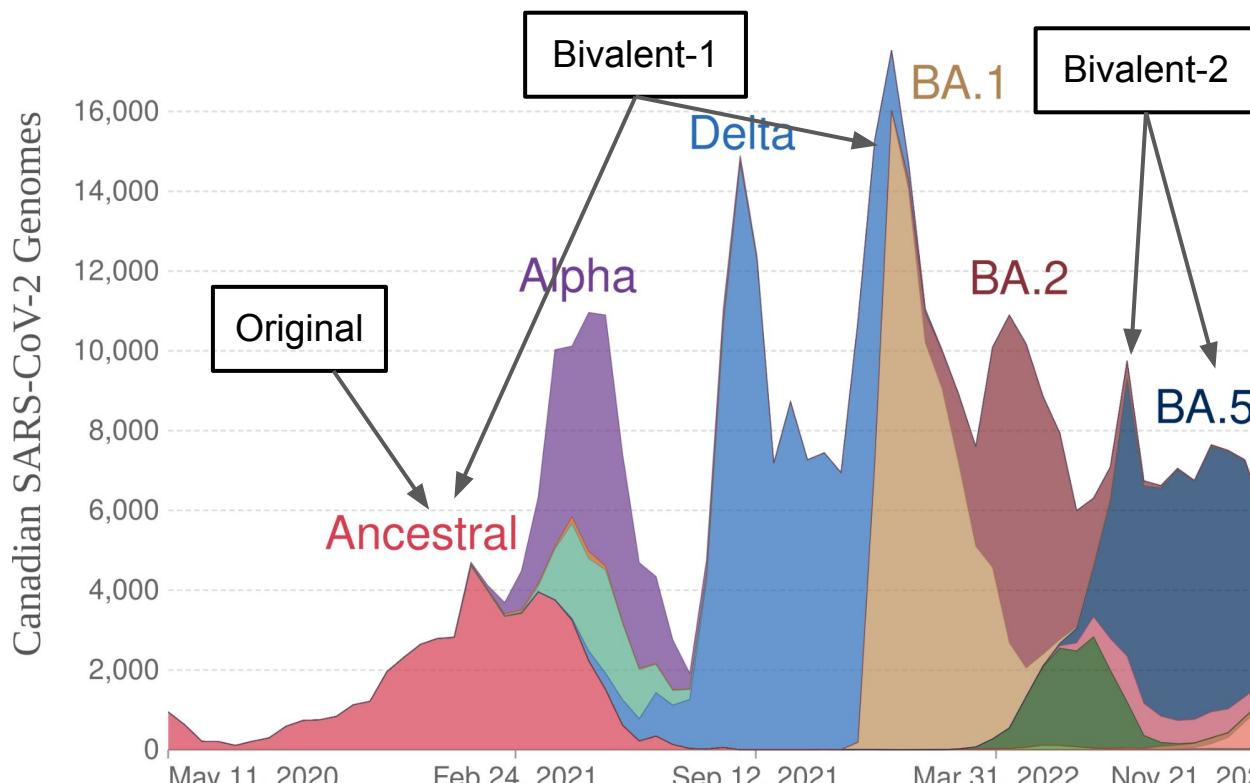


Source: GISAID, via CoVariants.org – Last updated 1 December 2022

Evolution drives cases: SARS-CoV-2 waves



Evolution drives vaccine design/effectiveness

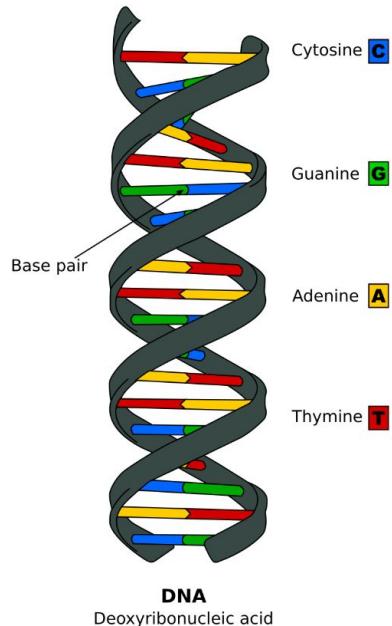


Source: GISAID, via CoVariants.org – Last updated 1 December 2022

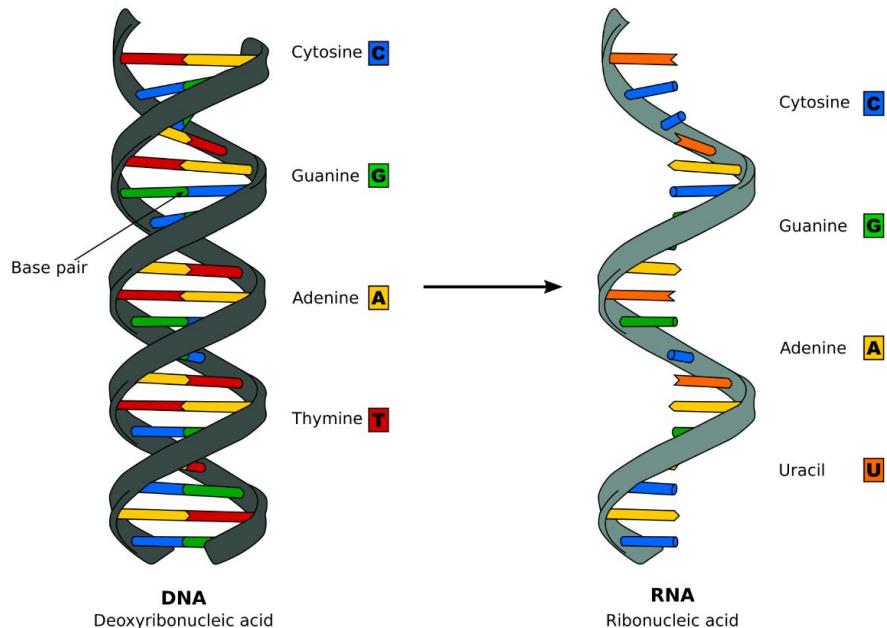
OurWorldInData.org/coronavirus • CC BY

How can we monitor evolution?

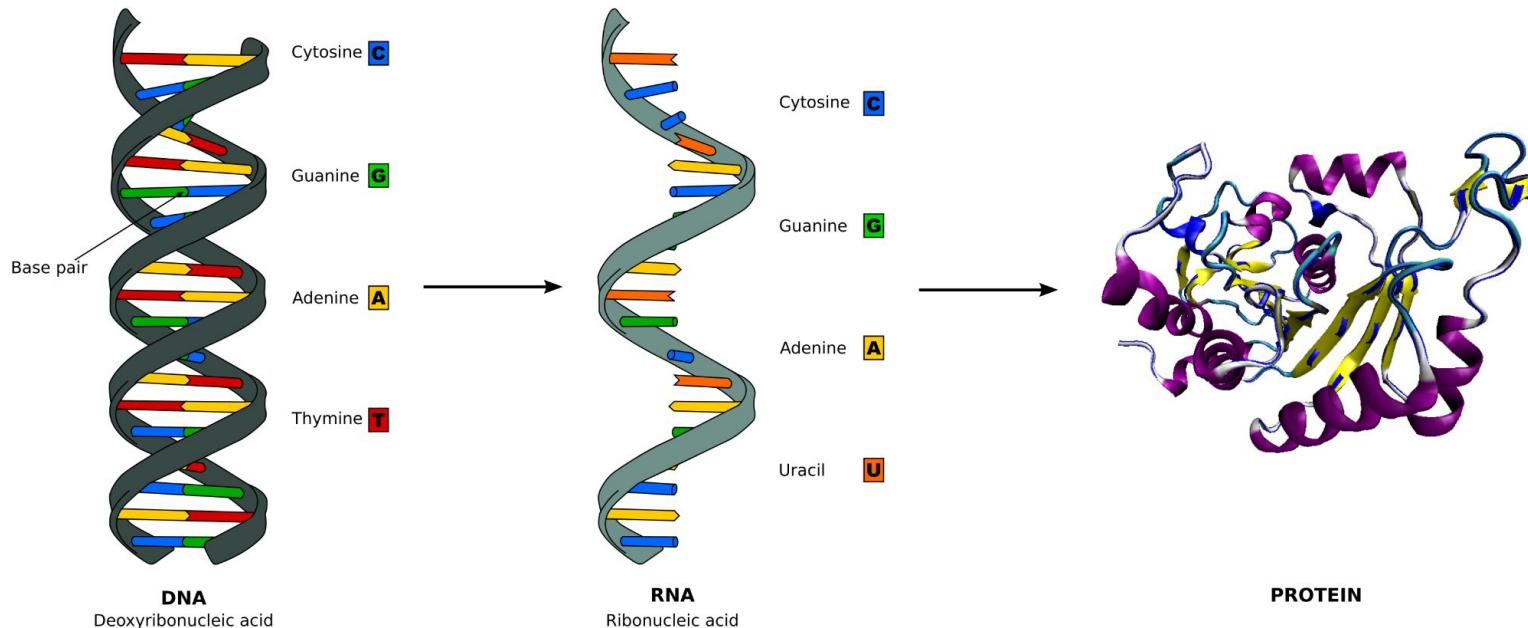
Genomes are the substrate of evolution



Genomes are the substrate of evolution

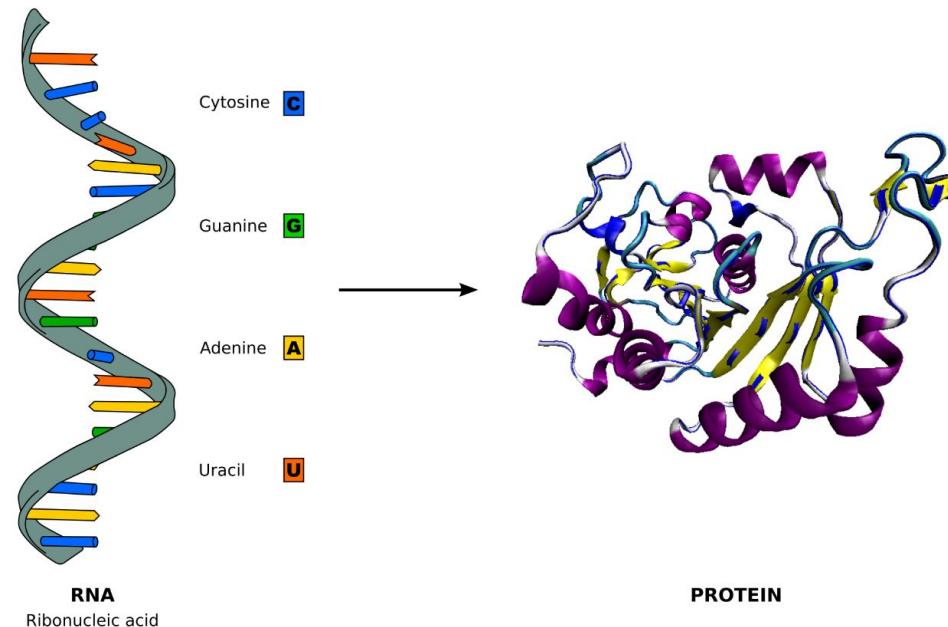


Genomes are the substrate of evolution



- DNA encodes RNA which encodes proteins

Genomes are the substrate of evolution



- DNA encodes RNA which encodes proteins
- Viruses like SARS-CoV-2 skip DNA

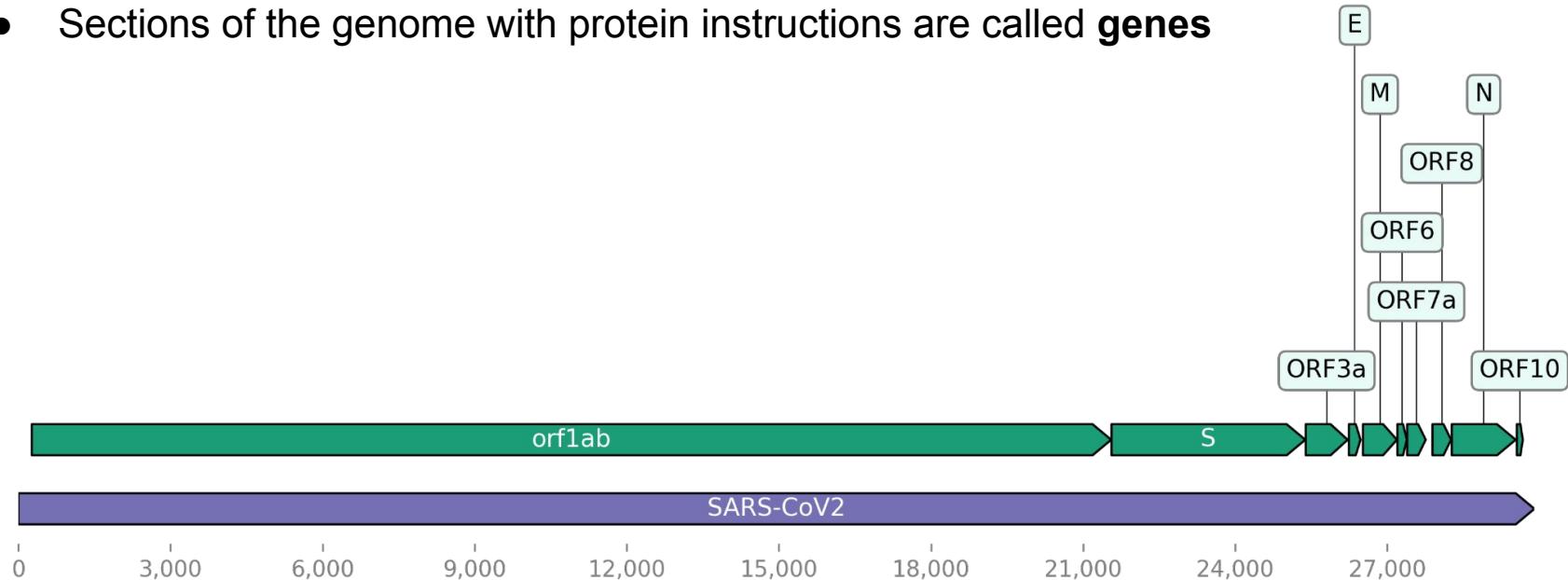
Genomes are the substrate of evolution

- Genomes are the complete collection of genetic instructions

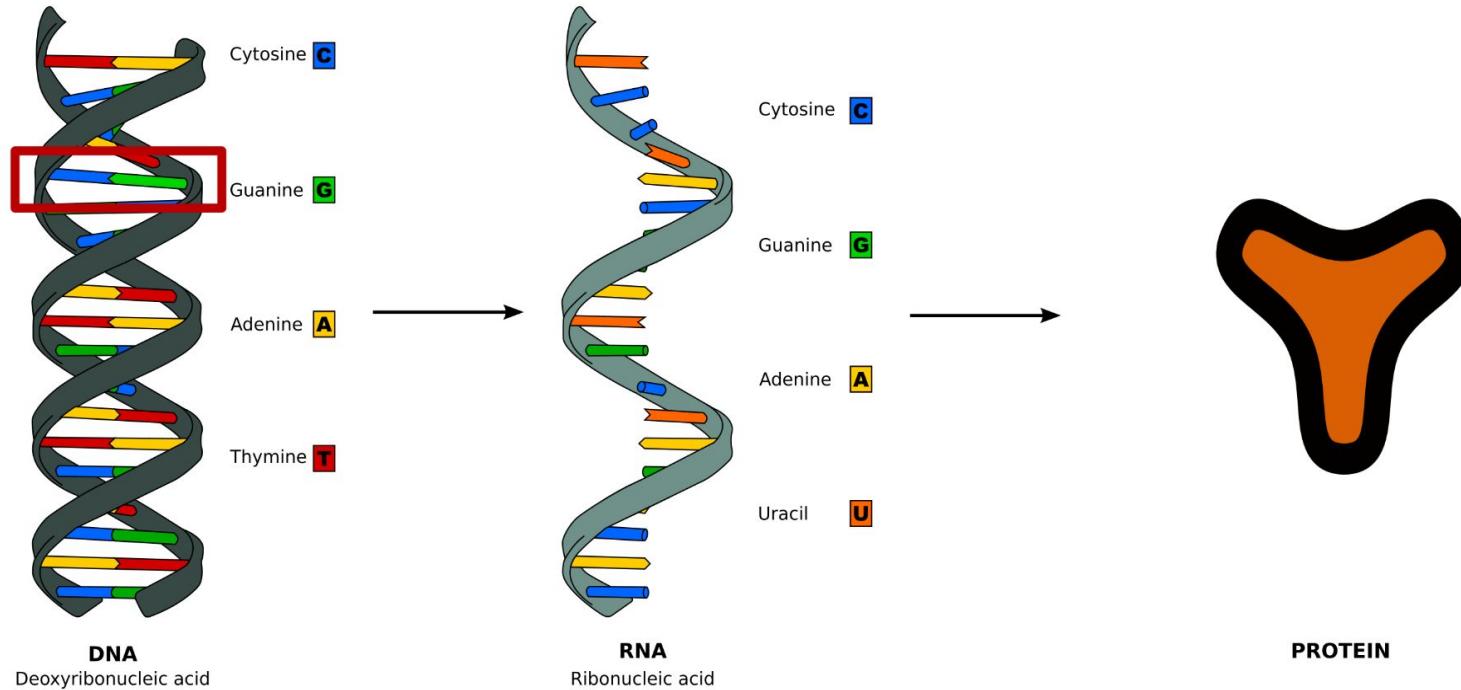


Genomes are the substrate of evolution

- Genomes are the complete collection of genetic instructions
- Sections of the genome with protein instructions are called **genes**

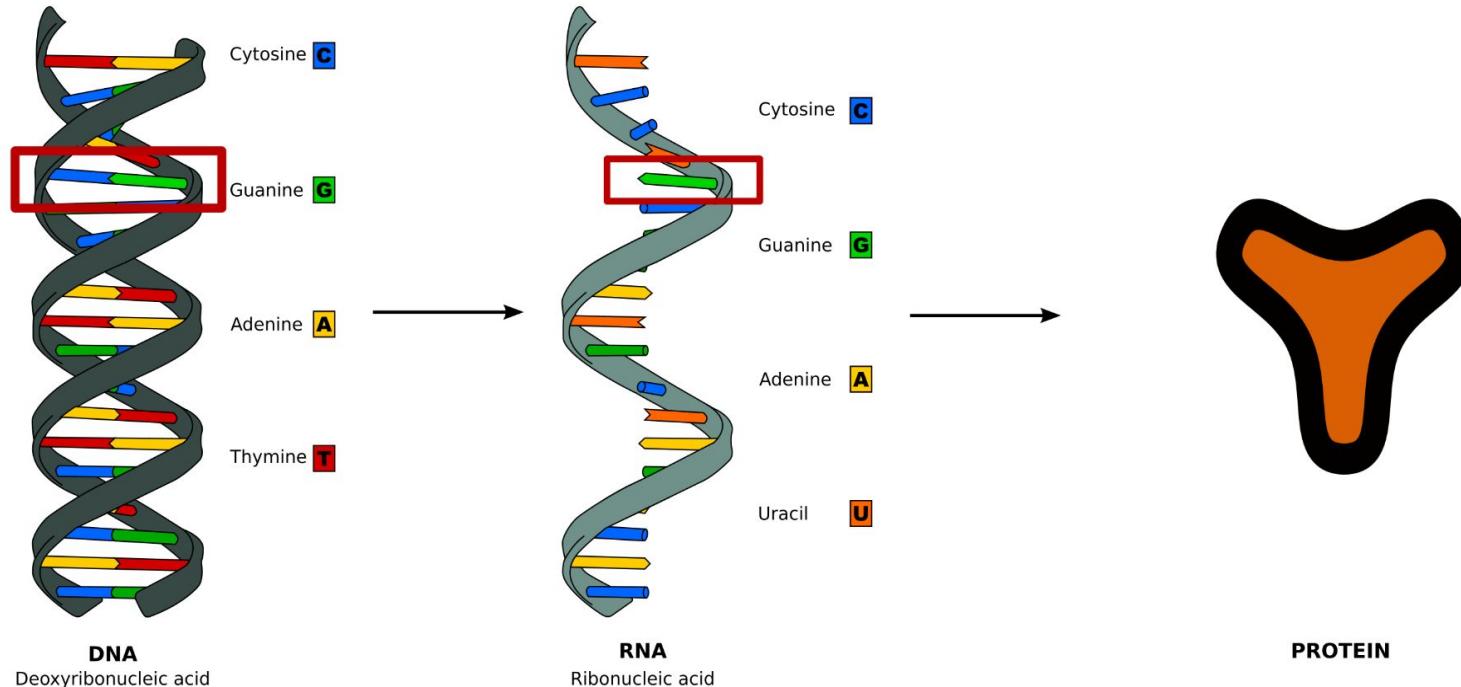


Mutations provide the variation upon which evolution acts



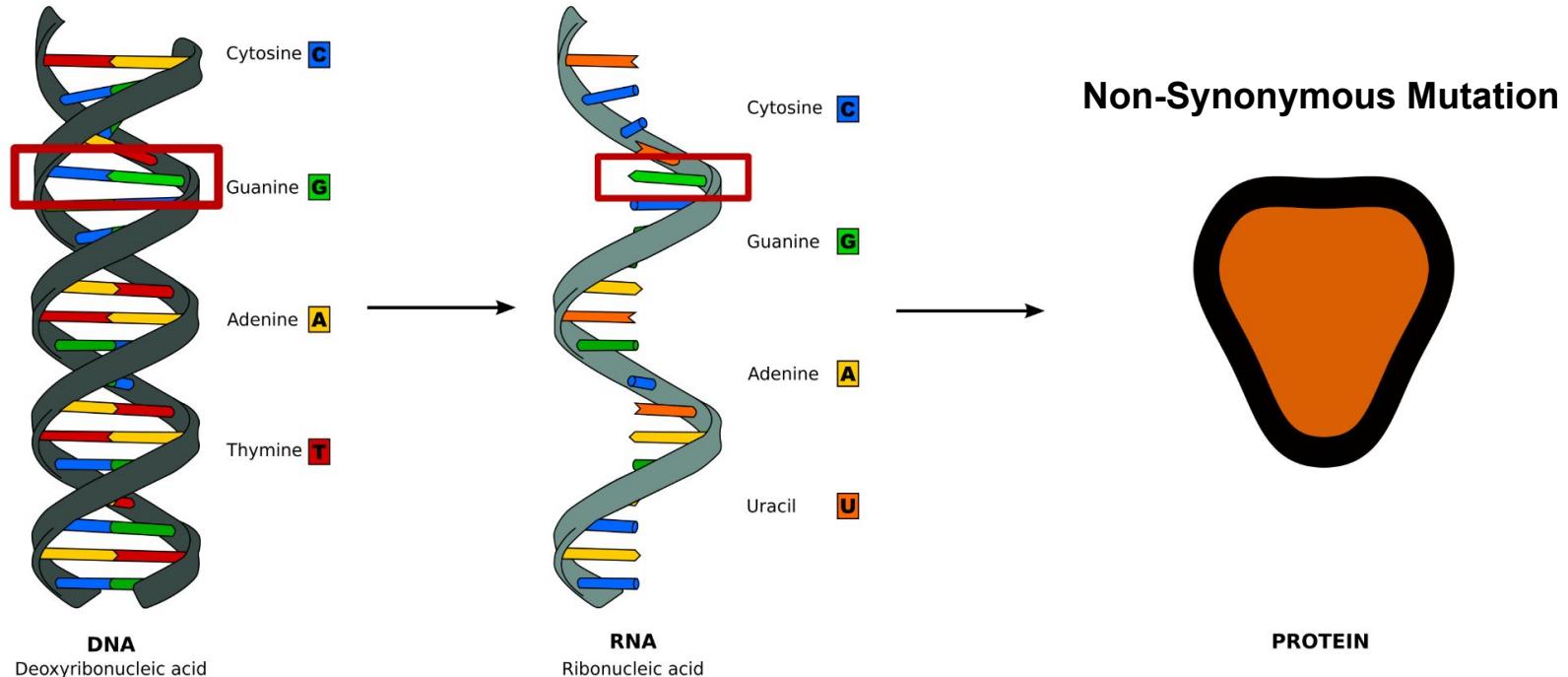
- Genome copying is error-prone

Mutations provide the variation upon which evolution acts



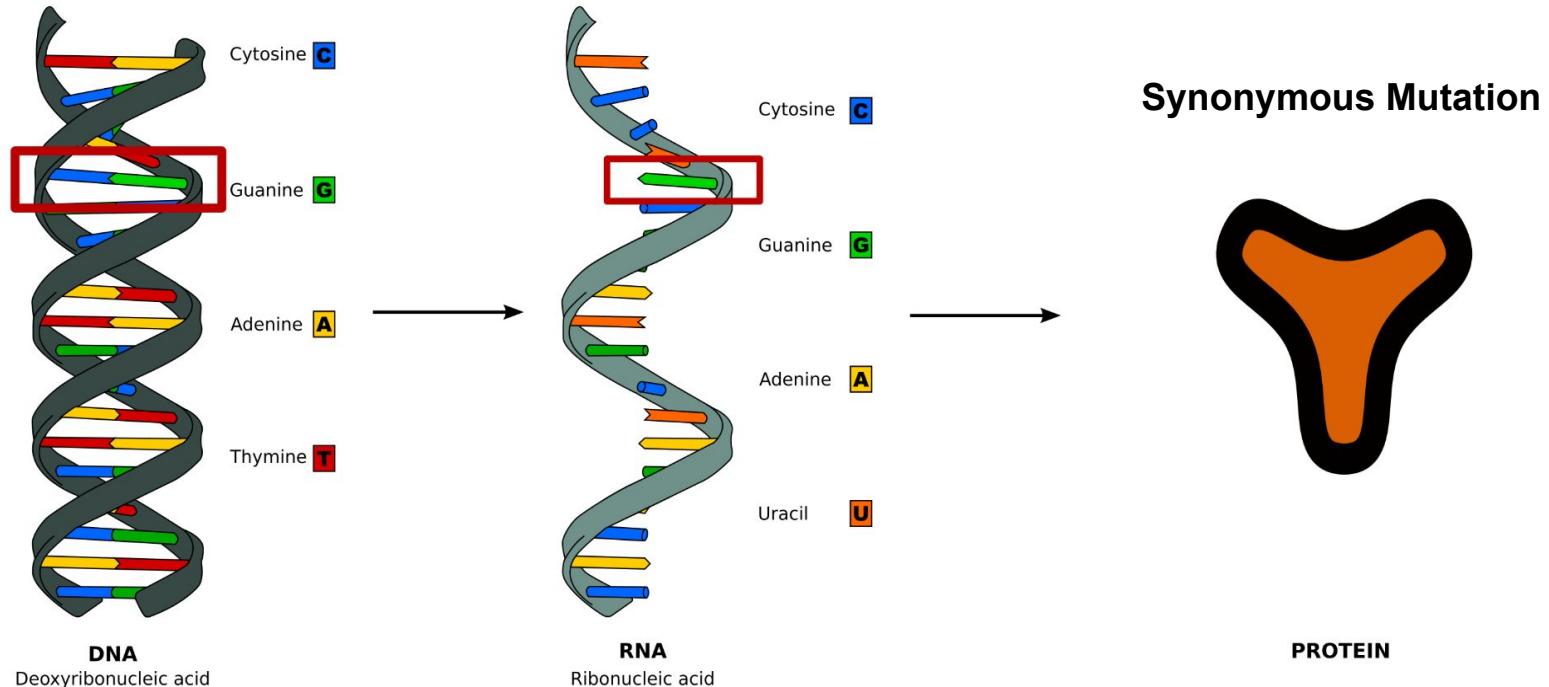
- Genome copying is error-prone
- Errors are called mutations

Mutations provide the variation upon which evolution acts



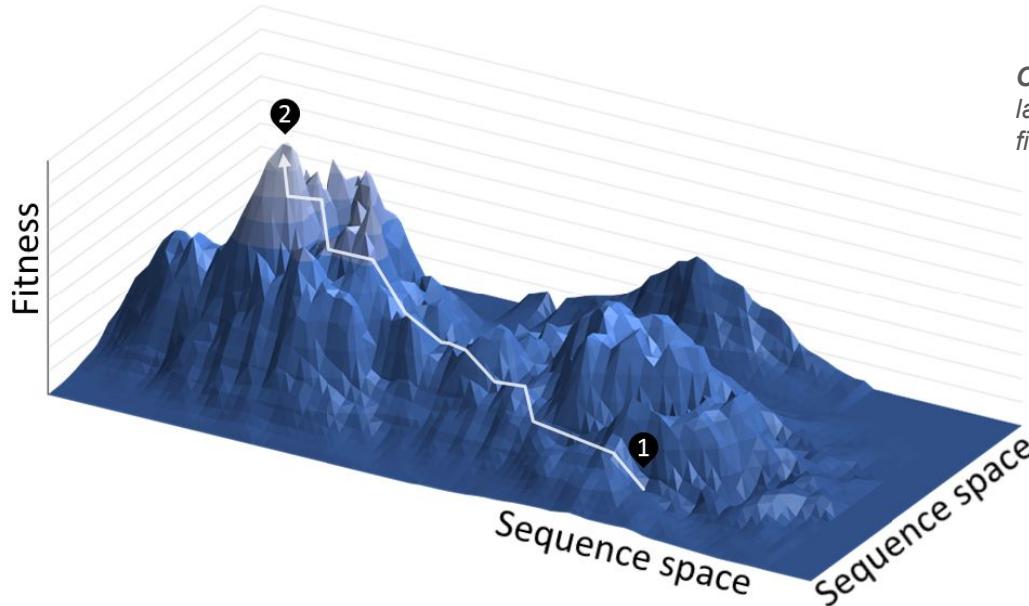
- Genome copying is error-prone
- Errors are called mutations
- Mutations can change protein sequence

Mutations provide the variation upon which evolution acts



- Genome copying is error-prone
- Errors are called mutations
- Mutations can change protein sequence - *but don't always*

Mutations are a random walk across a fitness landscape

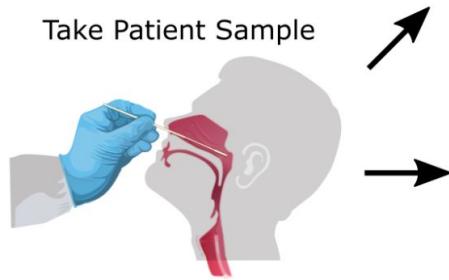


OVERSIMPLIFICATION - actual landscapes are dynamic/changing & fitness is hard to measure

- *Fitness = quantitative representation of individual reproductive success*
- *High Fitness = more descendants (larger proportion of circulating population)*
- *Low Fitness = fewer descendants (higher chance of dying out)*

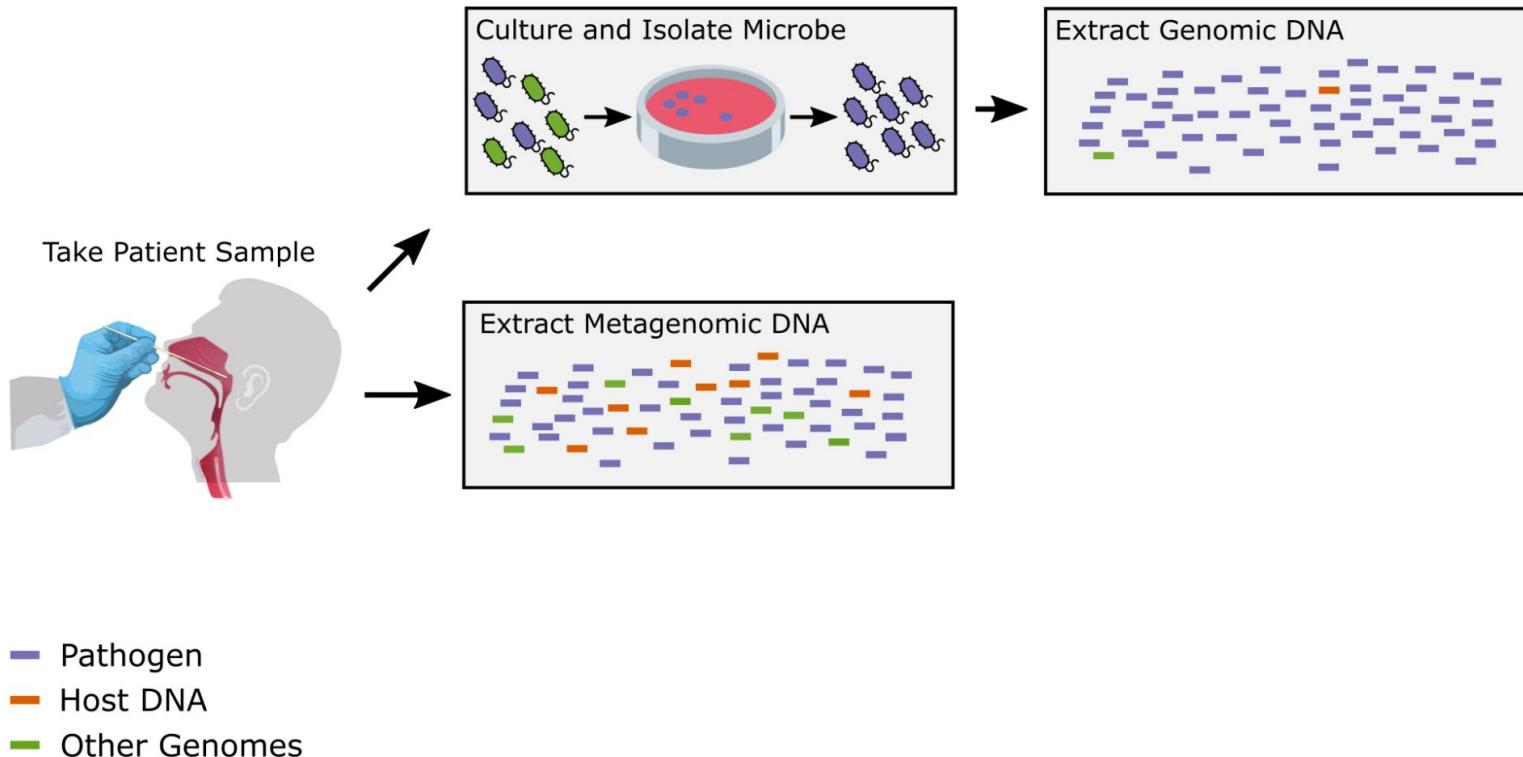
So, how do we get genomes?

Sequencing Pathogen Genomes

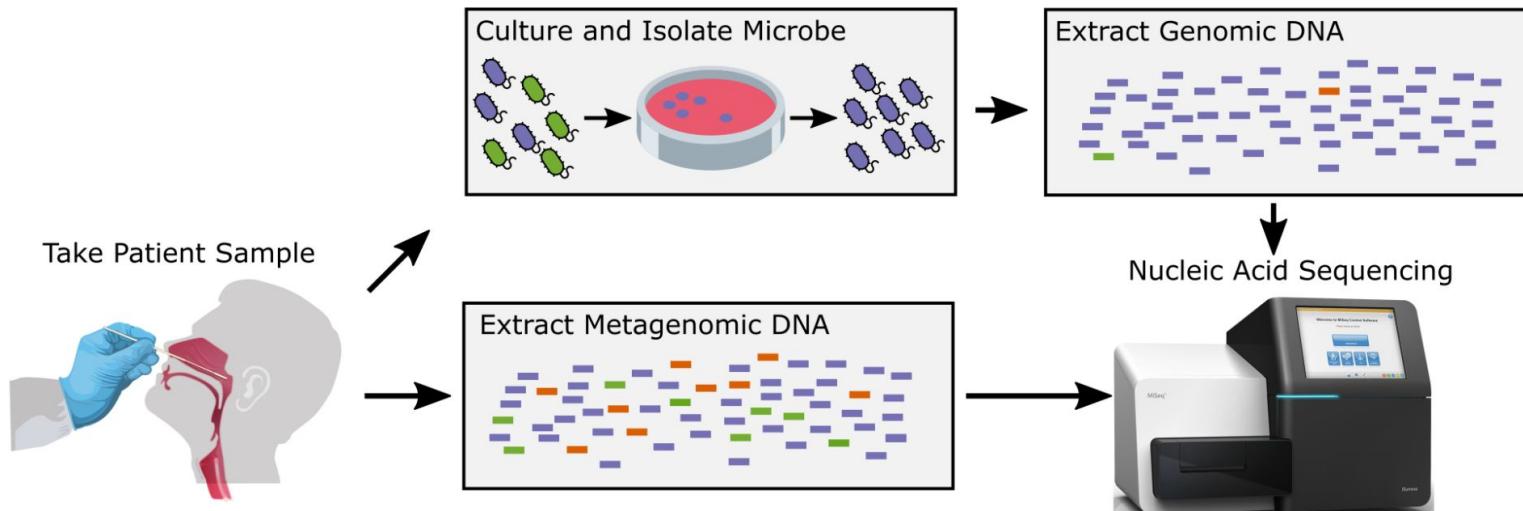


- Pathogen
- Host DNA
- Other Genomes

Sequencing Pathogen Genomes

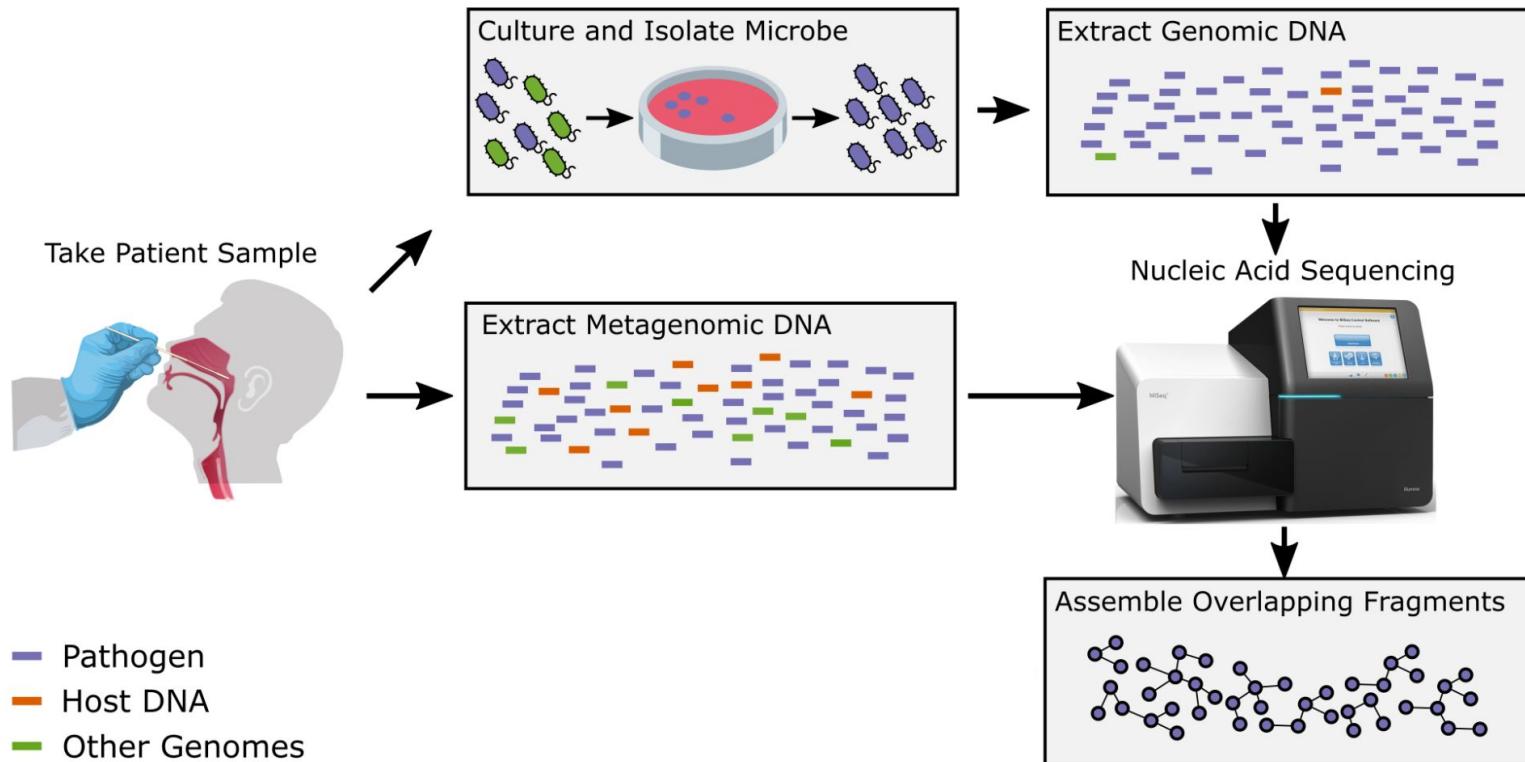


Sequencing Pathogen Genomes

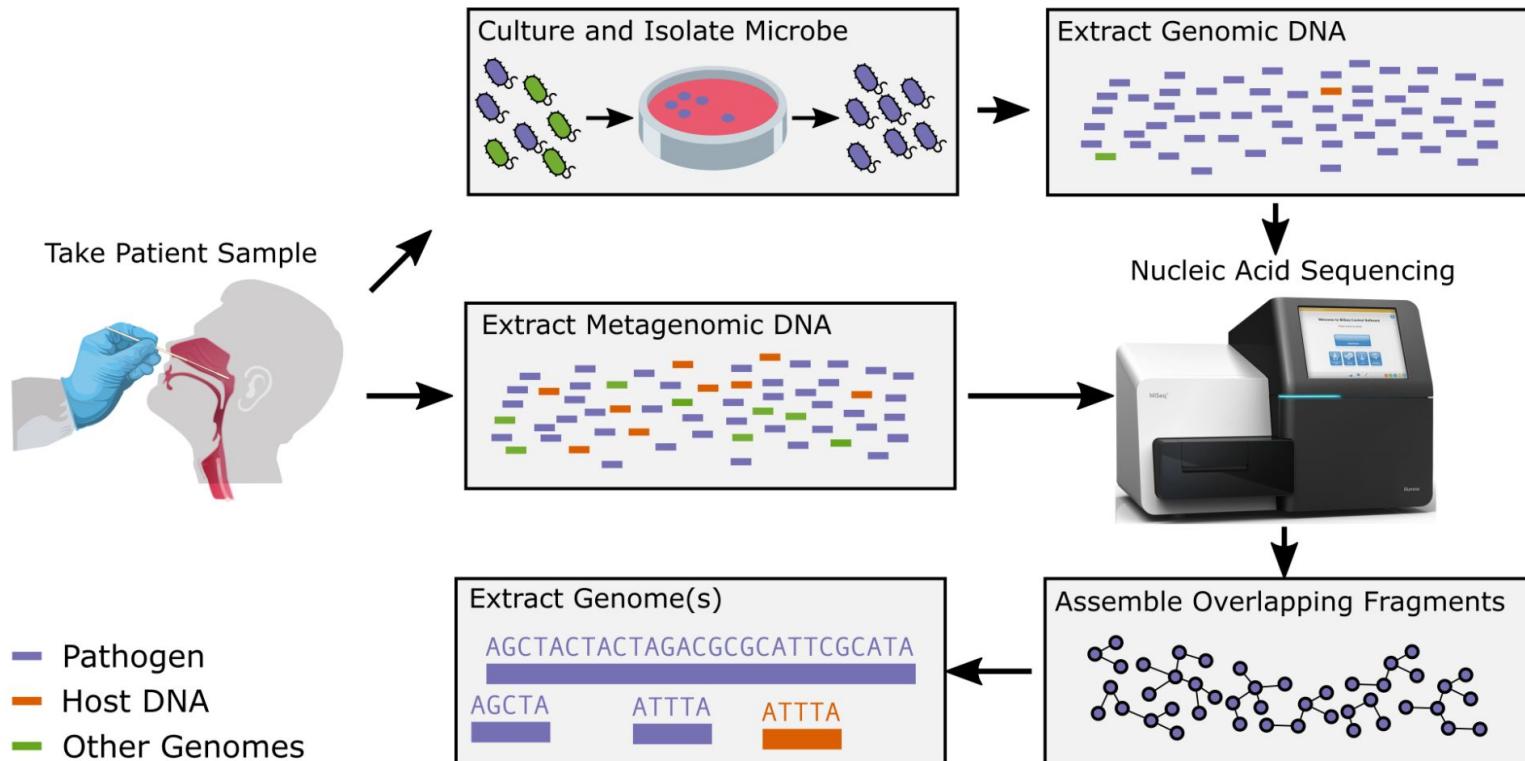


- Pathogen
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Sequencing Pathogen Genomes

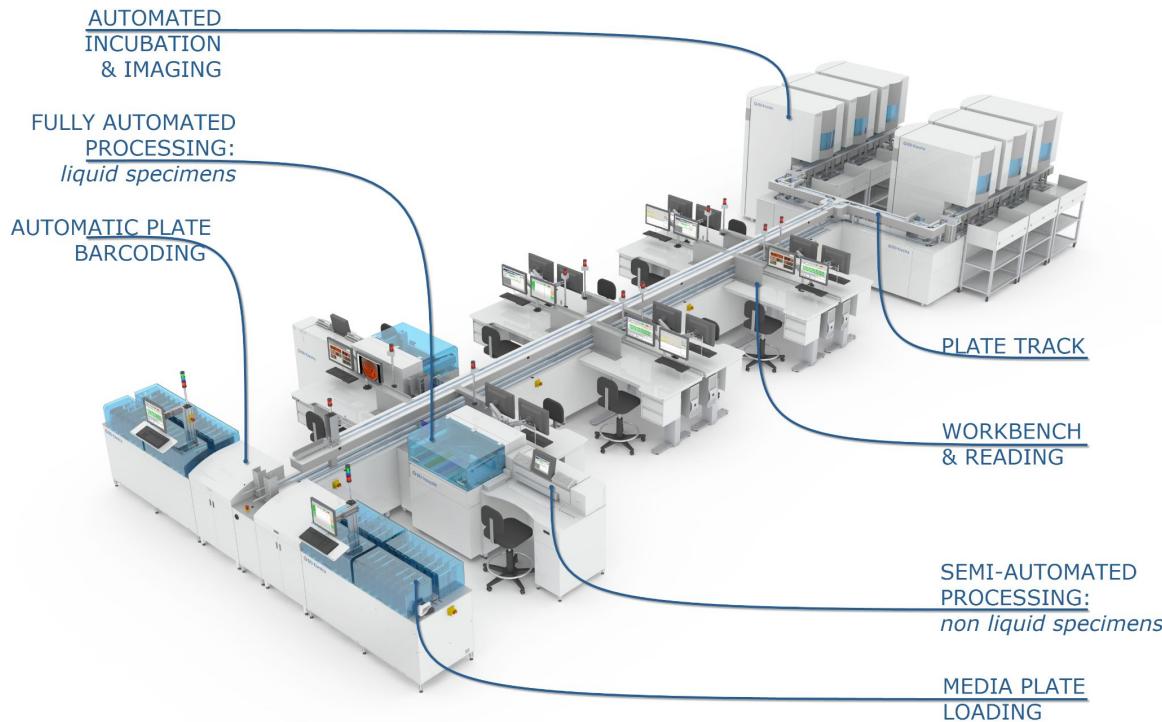


Sequencing Pathogen Genomes



How do we do this at scale?

Automation of labour intensive steps



<https://www.bd.com/scripts/europe/labautomation/productsdrilldown.asp?CatID=455&SubID=1836&siteID=20309&d=&s=europe%2Flabautomation&sTitle=Lab+Automation&metaTitle=Total+Lab+Automation&dc=europe&dcTitle=Europe>

Automation of labour intensive steps



MEDIA PLATE
LOADING

<https://www.bd.com/scripts/europe/labautomation/productsdrilldown.asp?CatID=455&SubID=1836&siteID=20309&d=&s=europe%2Flabautomation&sTitle=Lab+Automation&metaTitle=Total+Lab+Automation&dc=europe&dcTitle=Europe>

Sequencing technology has rapidly changed and improved

~1972-1977

First generation

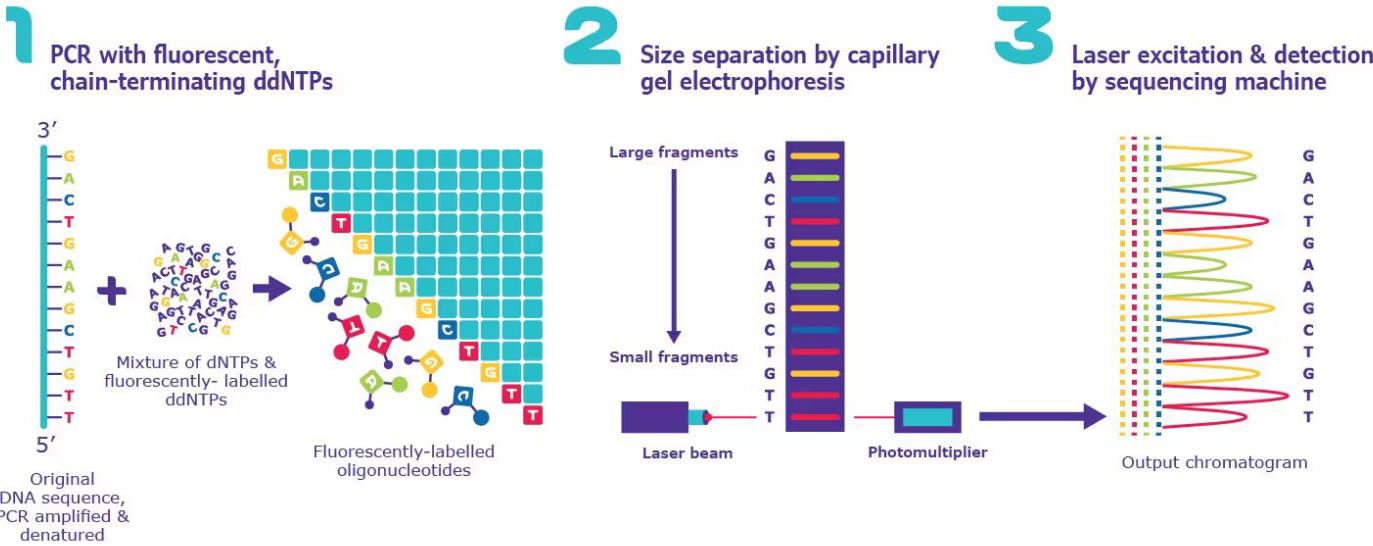


Sanger sequencing
Maxam and Gilbert
Sanger chain termination

Infer nucleotide identity using dNTPs,
then visualize with electrophoresis

500–1,000 bp fragments

Sanger Sequencing



Sequencing technology has rapidly changed and improved

~1972-1977

~2001-2004

First generation

Second generation
(next generation sequencing)



Sanger sequencing
Maxam and Gilbert
Sanger chain termination

Infer nucleotide identity using dNTPs,
then visualize with electrophoresis

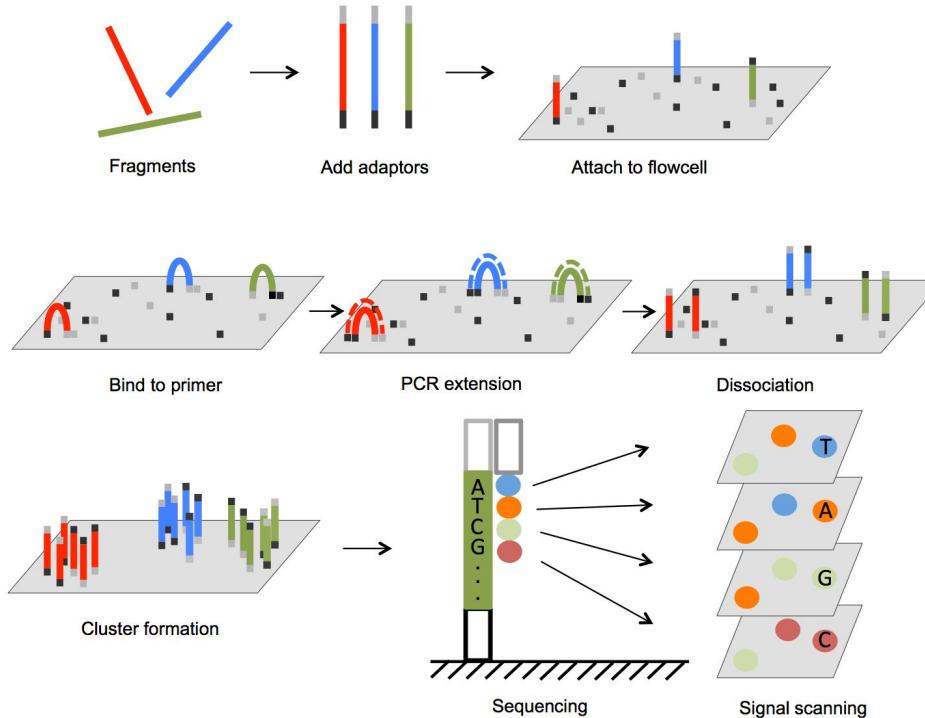
500–1,000 bp fragments

454, Solexa,
Ion Torrent,
Illumina

High throughput from the
parallelization of sequencing reactions

~50–500 bp fragments

Sequencing by Synthesis



Sequencing technology has rapidly changed and improved

~1972-1977

~2001-2004

First generation

Second generation
(next generation sequencing)



Sanger sequencing
Maxam and Gilbert
Sanger chain termination

Infer nucleotide identity using dNTPs,
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454, Solexa,
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Illumina

High throughput from the
parallelization of sequencing reactions

~50–500 bp fragments

Short-read sequencing

Sequencing technology has rapidly changed and improved

~1972-1977

~2001-2004

~2011-2015

First generation

Second generation
(next generation sequencing)

Third generation



Sanger sequencing
Maxam and Gilbert
Sanger chain termination

Infer nucleotide identity using dNTPs,
then visualize with electrophoresis

500–1,000 bp fragments

454, Solexa,
Ion Torrent,
Illumina

High throughput from the
parallelization of sequencing reactions

~50–500 bp fragments

PacBio
Oxford Nanopore

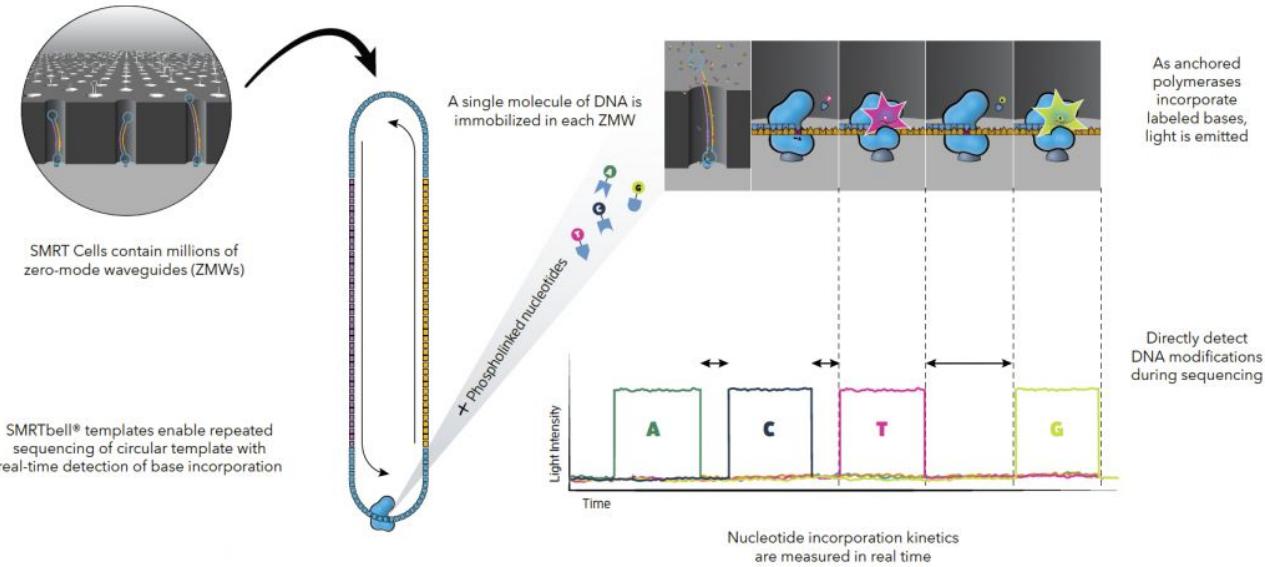
Sequence native DNA in real time
with single-molecule resolution

Tens of kb fragments, on average

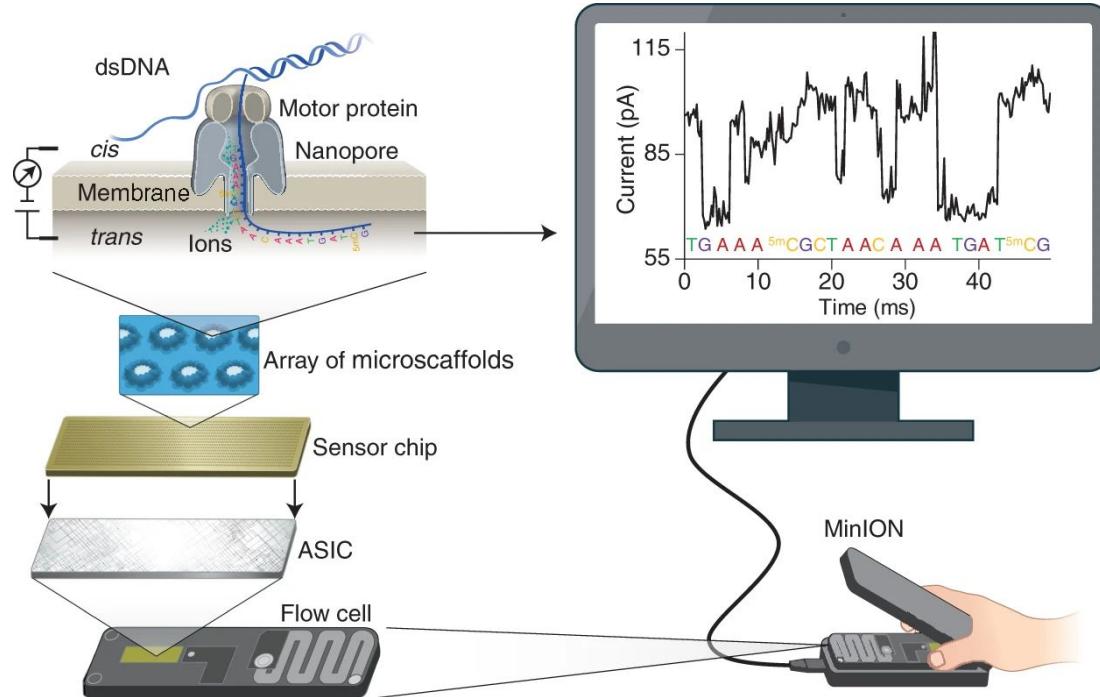
Short-read sequencing

Long-read sequencing

PacBio Sequencing



Nanopore Sequencing



Sequencing technology has rapidly changed and improved

~1972-1977

~2001-2004

~2011-2015

First generation

Second generation
(next generation sequencing)

Third generation



Sanger sequencing
Maxam and Gilbert
Sanger chain termination

Infer nucleotide identity using dNTPs,
then visualize with electrophoresis

500–1,000 bp fragments

454, Solexa,
Ion Torrent,
Illumina

High throughput from the
parallelization of sequencing reactions

Main Pathogenomics Platforms

~50–500 bp on average

PacBio
Oxford Nanopore

Sequence native DNA in real time
with single-molecule resolution

Short-read sequencing

Long-read sequencing

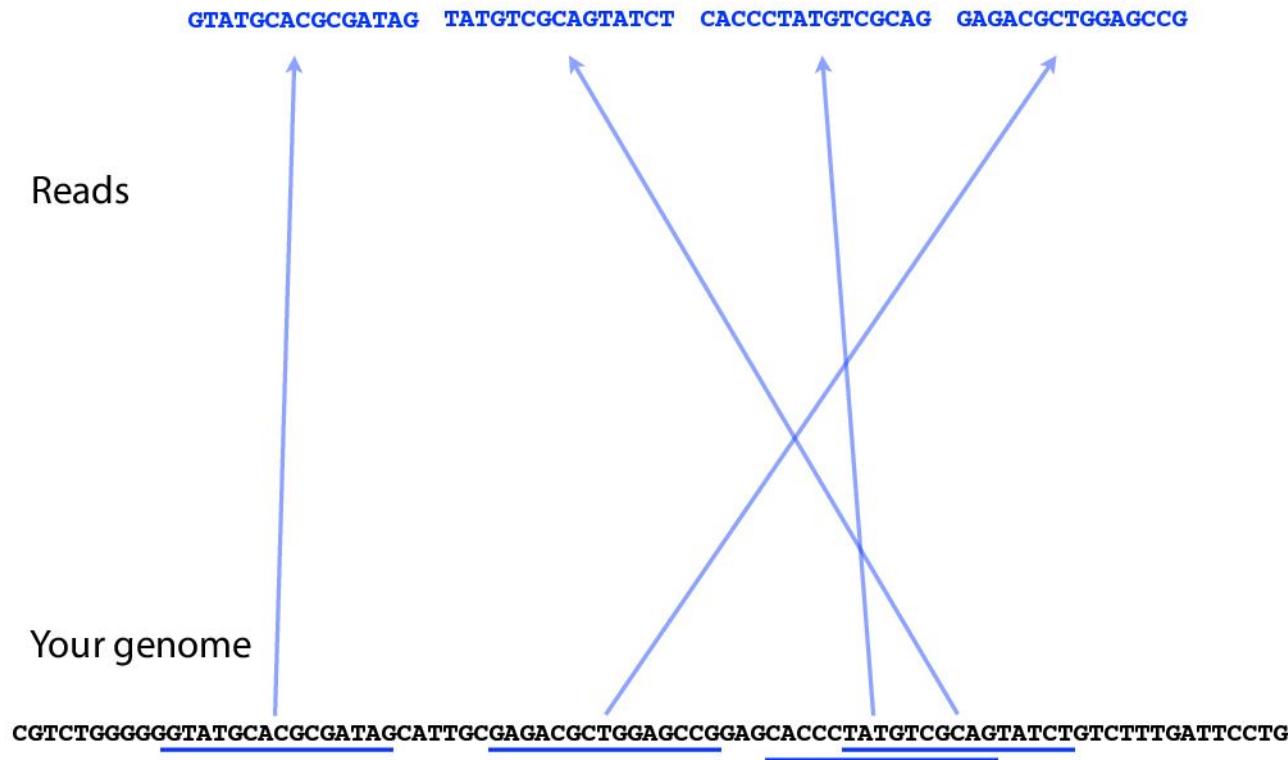
Mobile sequencing lab in a suitcase



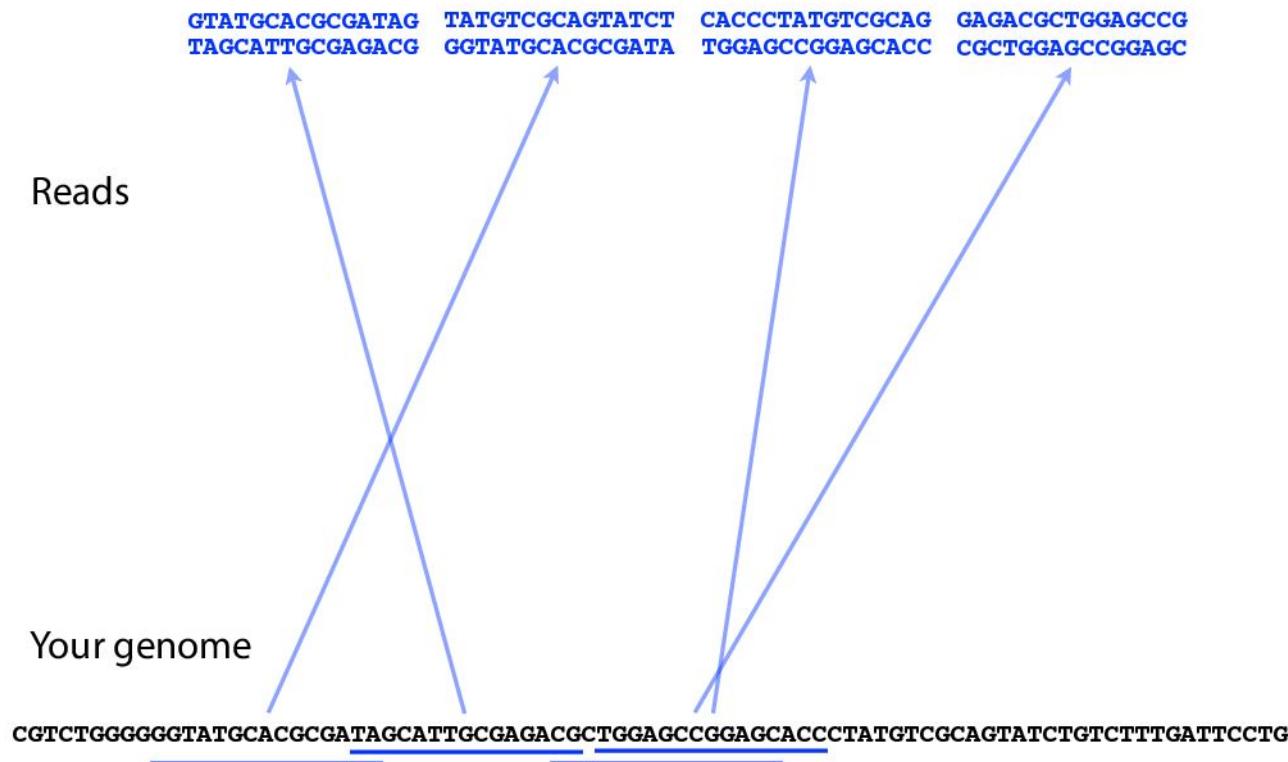
https://crain-platform-genomeweb-prod.s3.amazonaws.com/s3fs-public/styles/1200x630/public/lab_in_a_suitcase.jpeg

See lecture reading: Loman & Gardy 2017

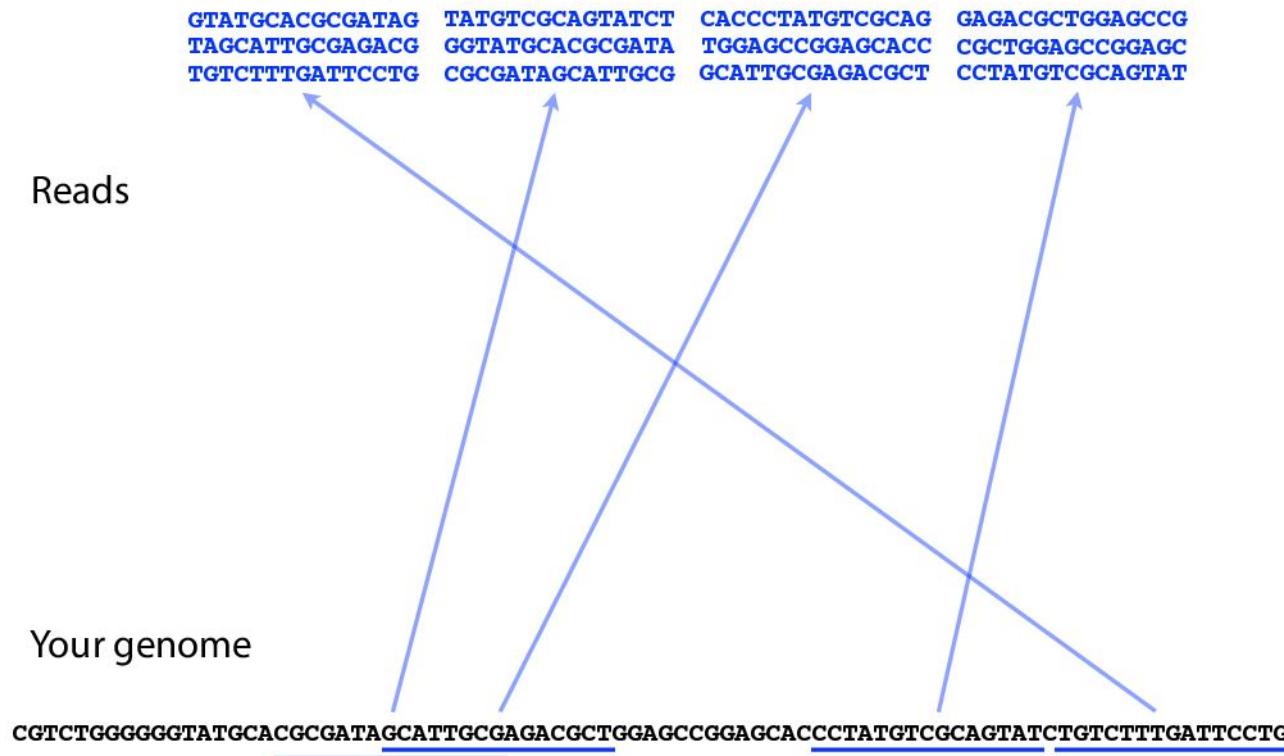
Reads are randomly(-ish) sampled from the DNA



Reads are randomly(-ish) sampled from the DNA



Reads are randomly(-ish) sampled from the DNA



Reads are randomly(-ish) sampled from the DNA

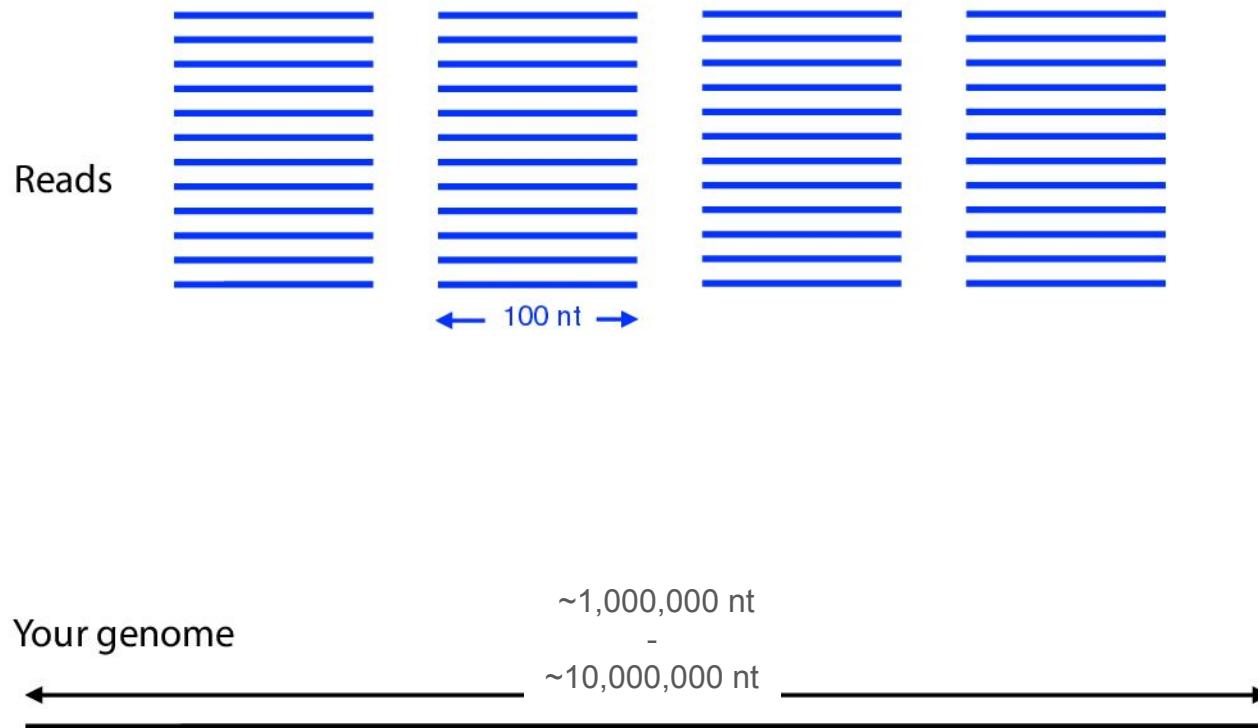
Reads

GTATGCACGCGATAG	TATGTCGCAGTATCT	CACCCCTATGTCGCAG	GAGACGCTGGAGCCG
TAGCATTGCGAGACG	GGTATGCA CGCGATA	TGGAGCCGGAGCACC	CGCTGGAGCCGGAGC
TGTCTTTGATTCTTG	CGCGATAGCATTGCG	GCATTGCGAGACGCT	CCTATGTCGCAGTAT
GACGCTGGAGCCGGA	GCACCCCTATGTCGA	GTATCTGCTTTGAT	CCTCATCCTATTATT
TATCGCACCTACGTT	CAATATTGATCATG	GATCACAGGTCTATC	ACCCCTATTAACCACT
CACGGGAGCTCTCCA	TGCATTGGTATTTT	CGTCTGGGGGTATG	CACGGATAGCATTG
GTATGCACGCGATAG	ACCTACGTTCAATAT	TATTTATGCA CCTA	CCACCTCACGGGAGCT
GCGAGACGCTGGAGC	CTATCACCCCTATTAA	CTGTCTTTGATTCT	ACTCACGGGAGCTCT
CCTACGTTCAATATT	GCACCCCTACGTTCAAT	GTCTGGGGGTATGC	AGCCGGAGCACCTA
GACGCTGGAGCCGGA	GCACCCCTATGTCGA	GTATCTGCTTTGAT	CCTCATCCTATTATT
TATCGCACCTACGTT	CAATATTGATCATG	GATCACAGGTCTATC	ACCCCTATTAACCACT
CACGGGAGCTCTCCA	TGCATTGGTATTTT	CGTCTGGGGGTATG	CACGGATAGCATTG

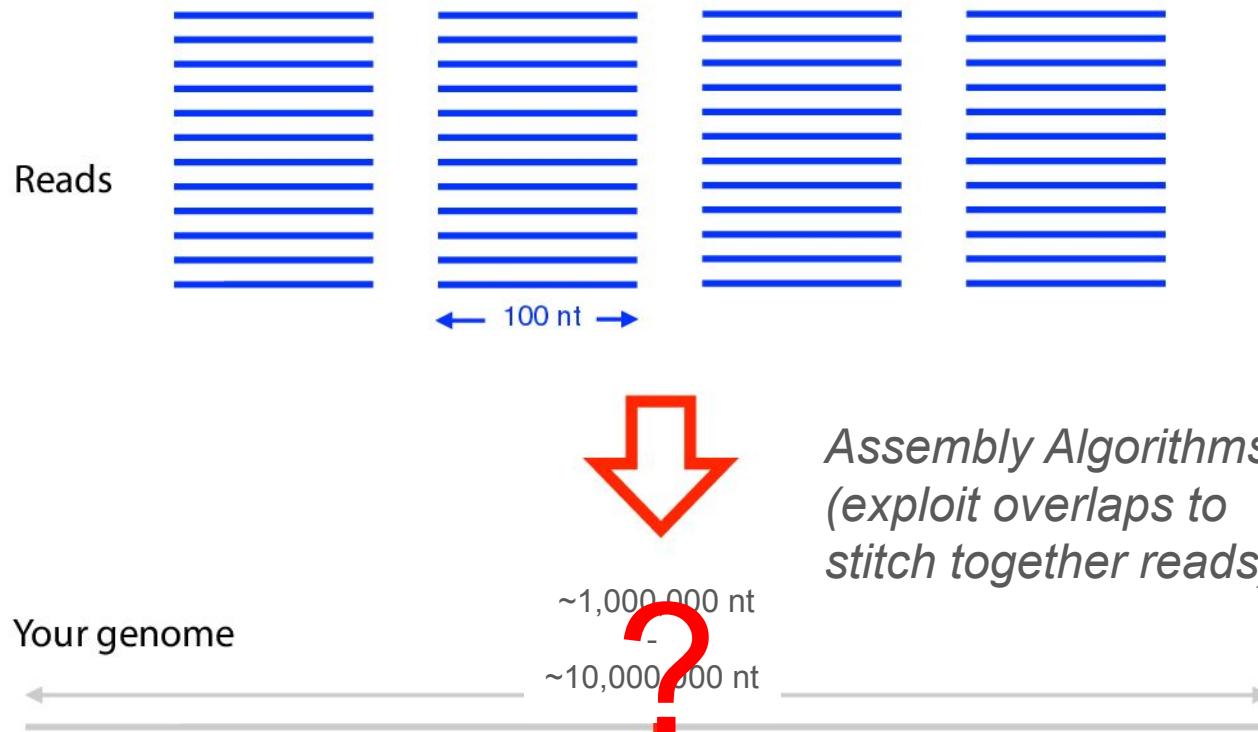
Your genome

CGTCTGGGGGTATGCACGCGATAGCATTGCGAGACGCTGGAGCCGGAGCACCCCTATGTCGCAGTATCTGCTTTGATTCTG

Reads are randomly(-ish) sampled from the DNA



Reads are randomly(-ish) sampled from the DNA



Got a genome, now what?

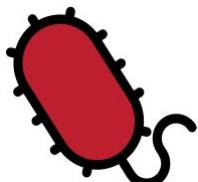
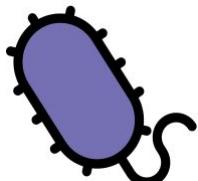
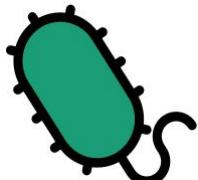
Genomic Diagnostics: What is the pathogen?



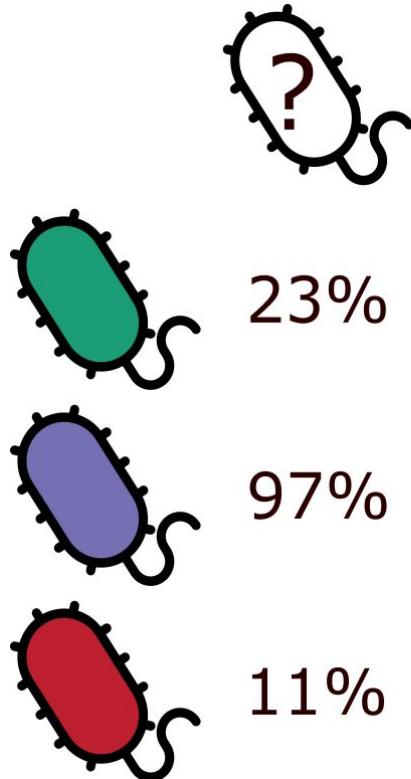
Genomic Diagnostics: What is the pathogen?



- Compare to genomes in database from known organisms



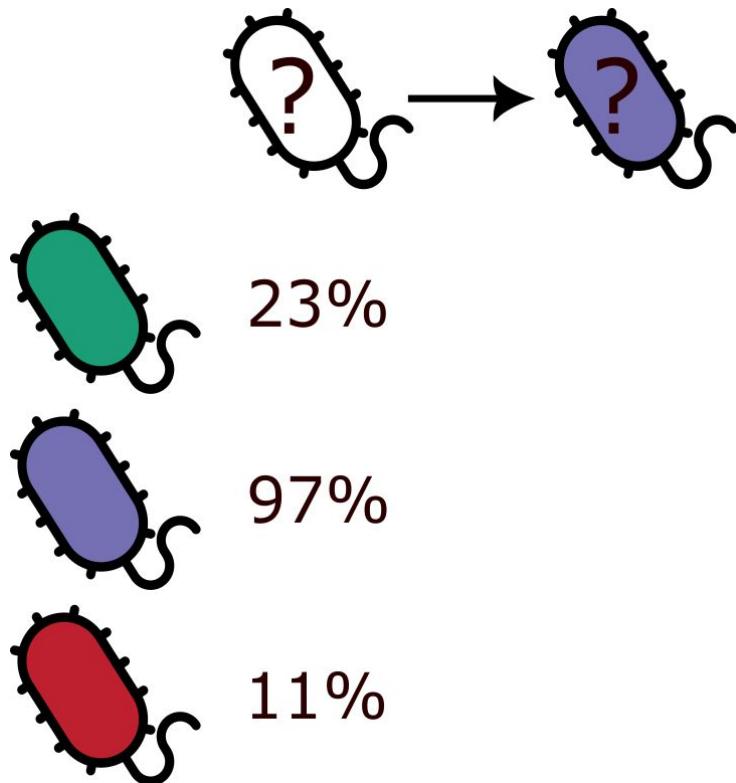
Genomic Diagnostics: What is the pathogen?



- Compare to genomes in database from known organisms
- Average Nucleotide Identity (ANI) is an example of a similarity metric

$$g\text{ANI} = \frac{\sum_{bbh} (\text{Percent Identity} * \text{Alignment length})}{\text{lengths of BBH genes}}$$

Genomic Diagnostics: What is the pathogen?

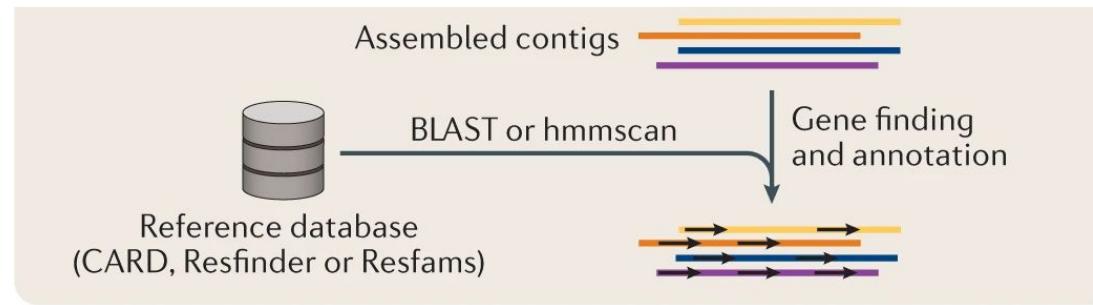


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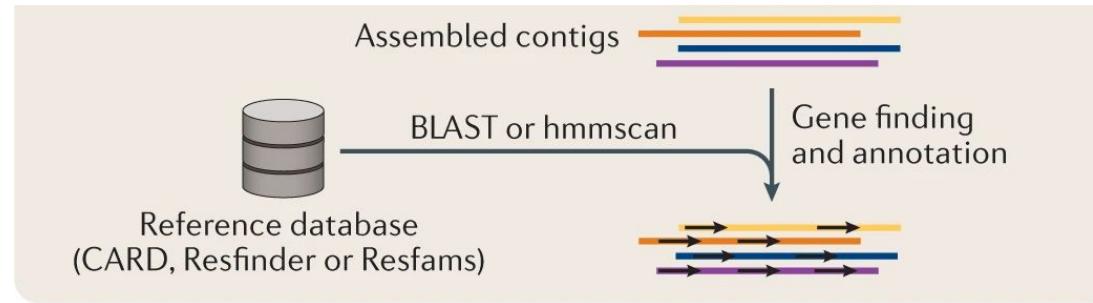
- Identify pathogen as closest reference genome taxa
- Use identity to drive treatment (if x then treat by y)
- Typing for outbreak investigation linkage

Genomic Diagnostics: What drugs will work?



10.1038/s41576-019-0108-4

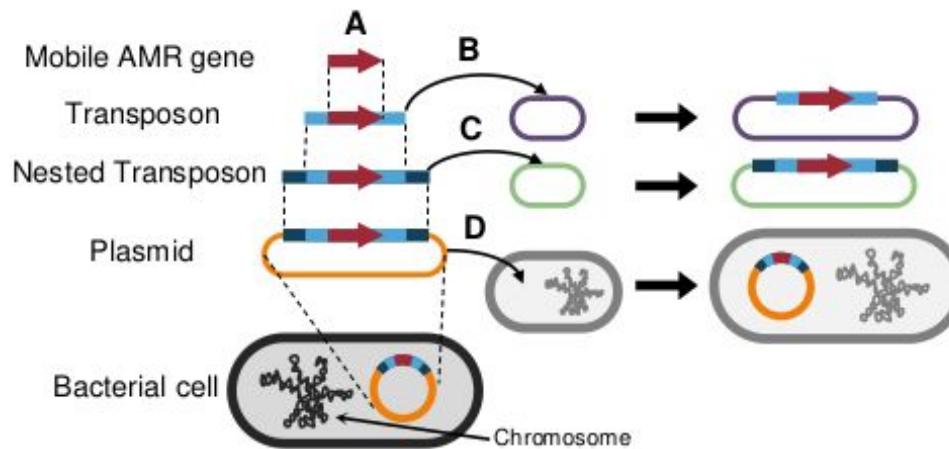
Genomic Diagnostics: What drugs will work?



10.1038/s41576-019-0108-4

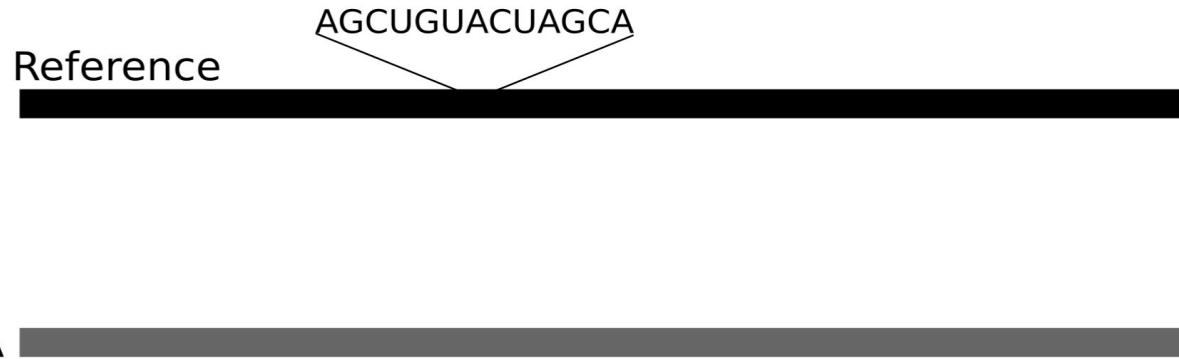
- Detect NDM-1 carbapenemase gene
- Pathogen protein that destroys many antibiotics (beta-lactams)
- => Treat with alternative class of antibiotics (e.g., colistin)
- Hours vs weeks for some pathogens (TB)

Genomic Diagnostics: identifying mobile genetic elements

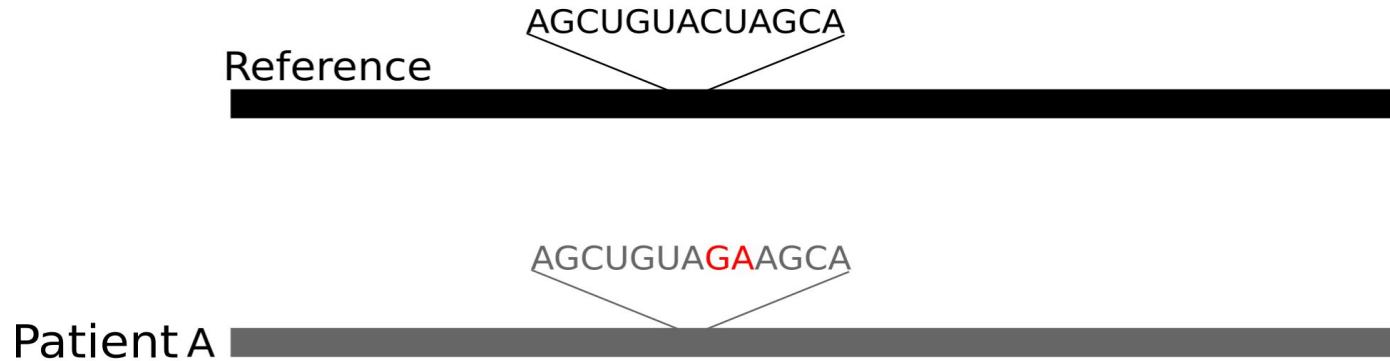


OK, but what else can we do with genomes?

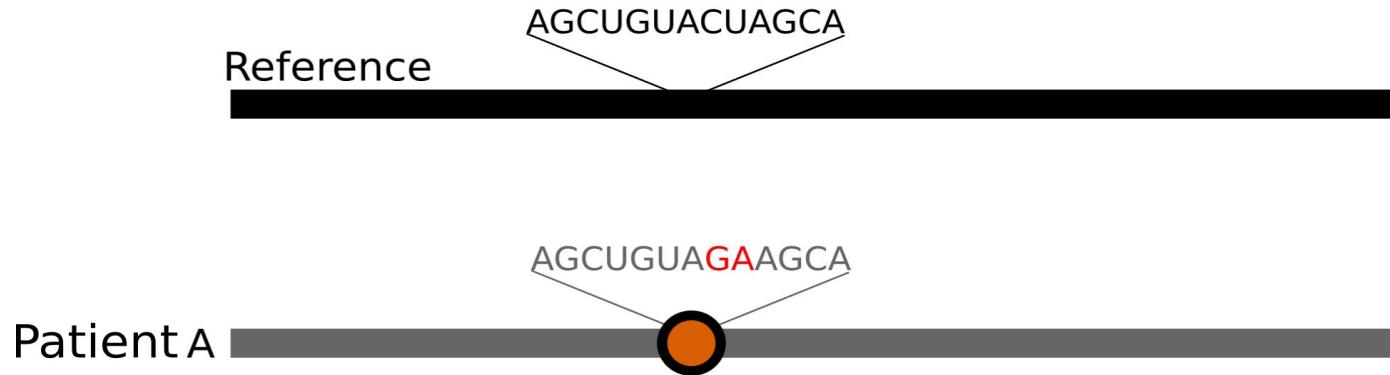
Detection mutations relative to reference



Detection mutations relative to reference



Detection mutations relative to reference



Compare mutations across patients

Reference

A

B

C

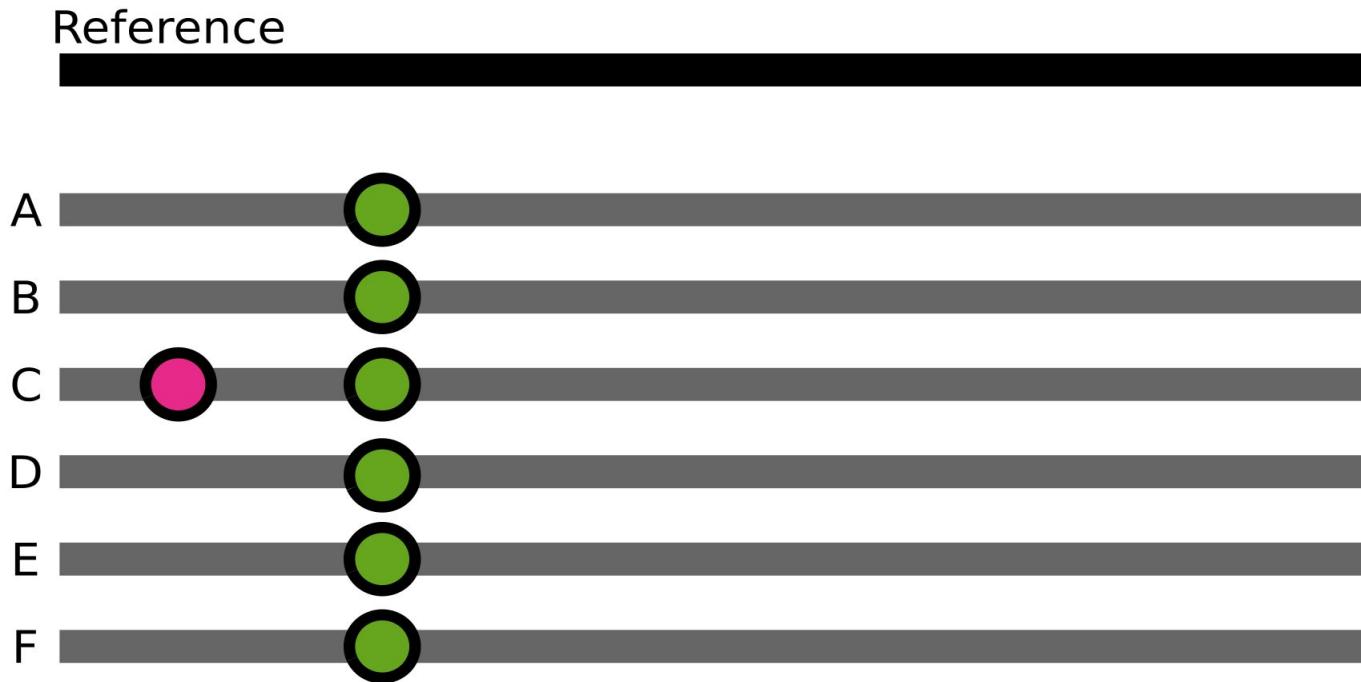
D

E

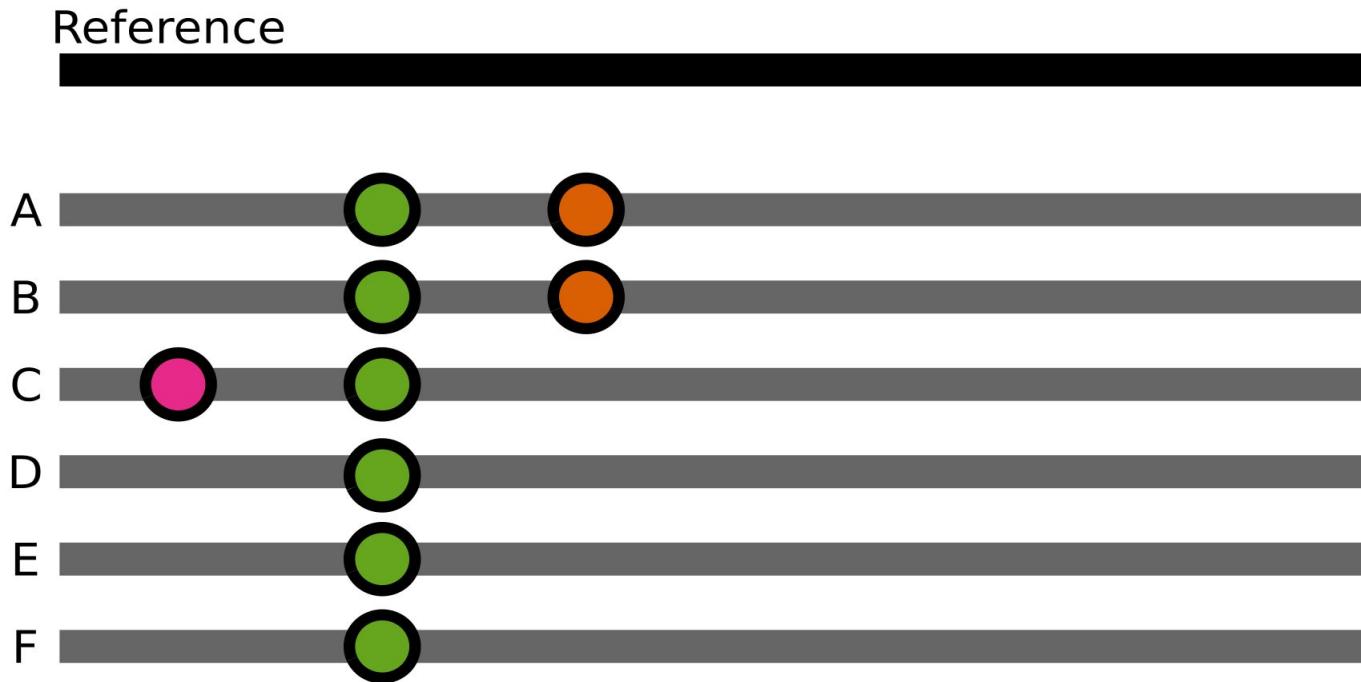
F



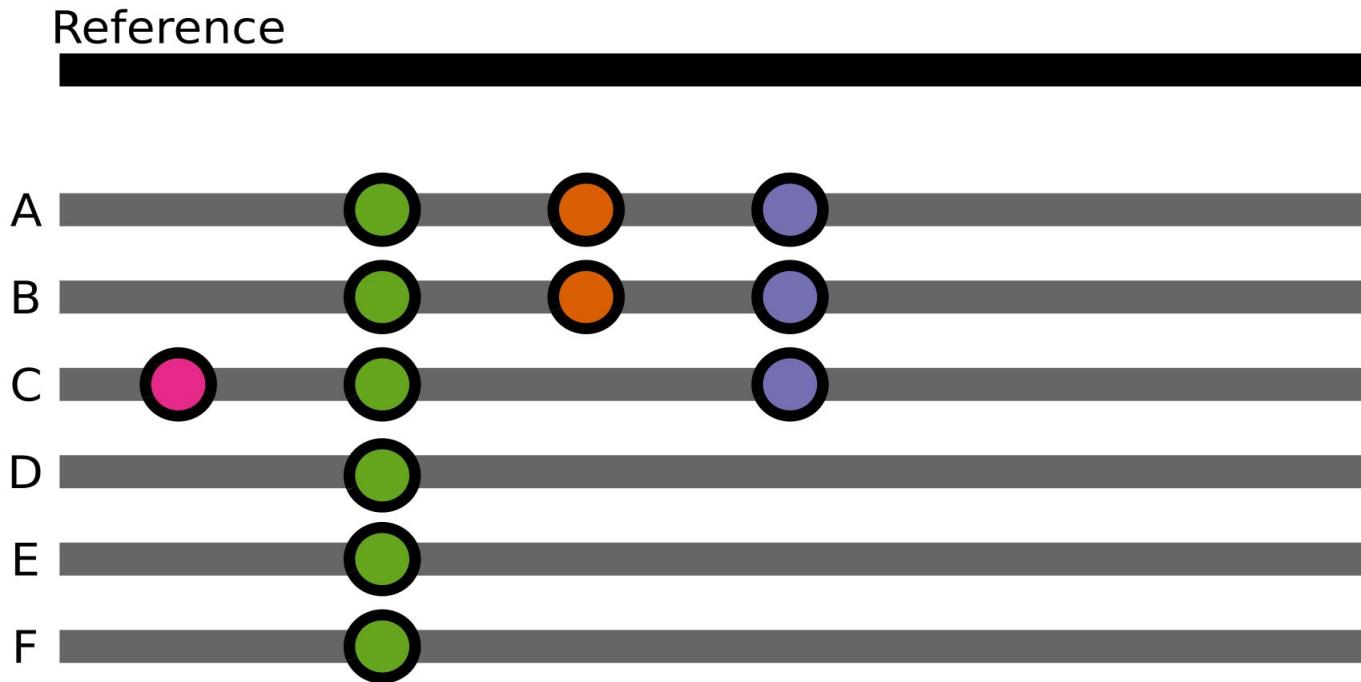
Compare mutations across patients



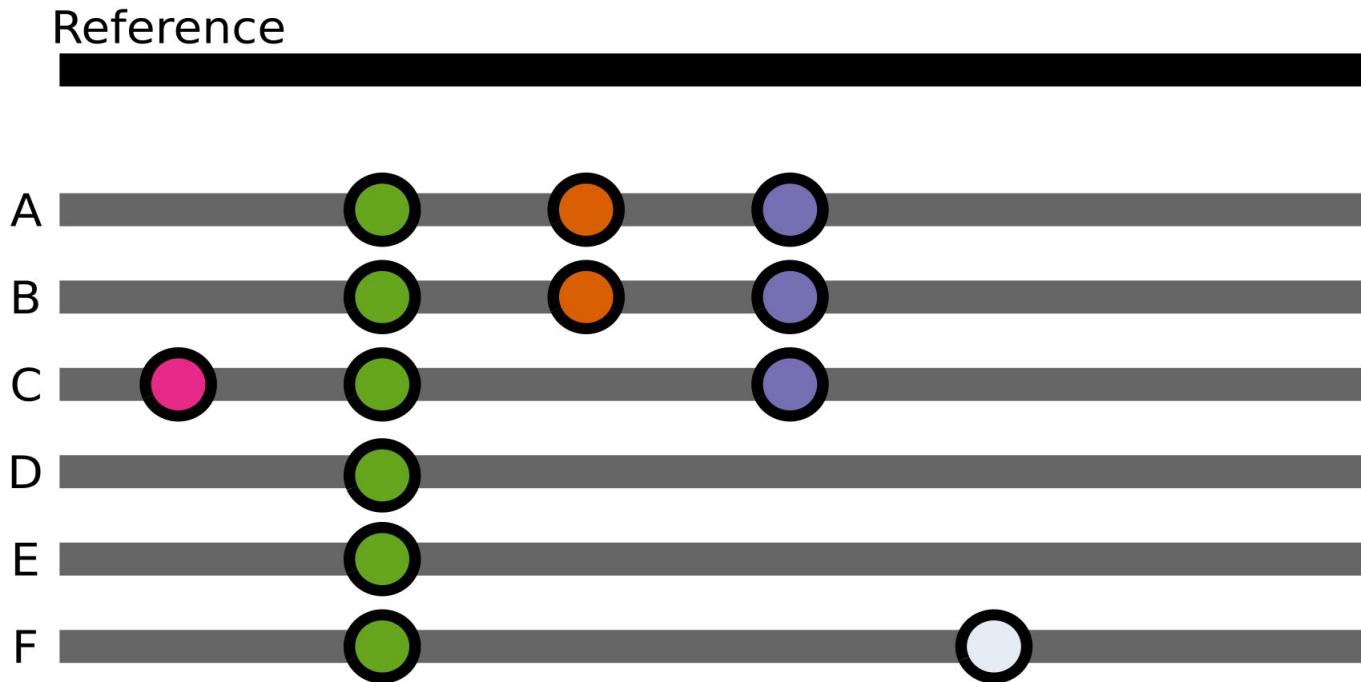
Compare mutations across patients



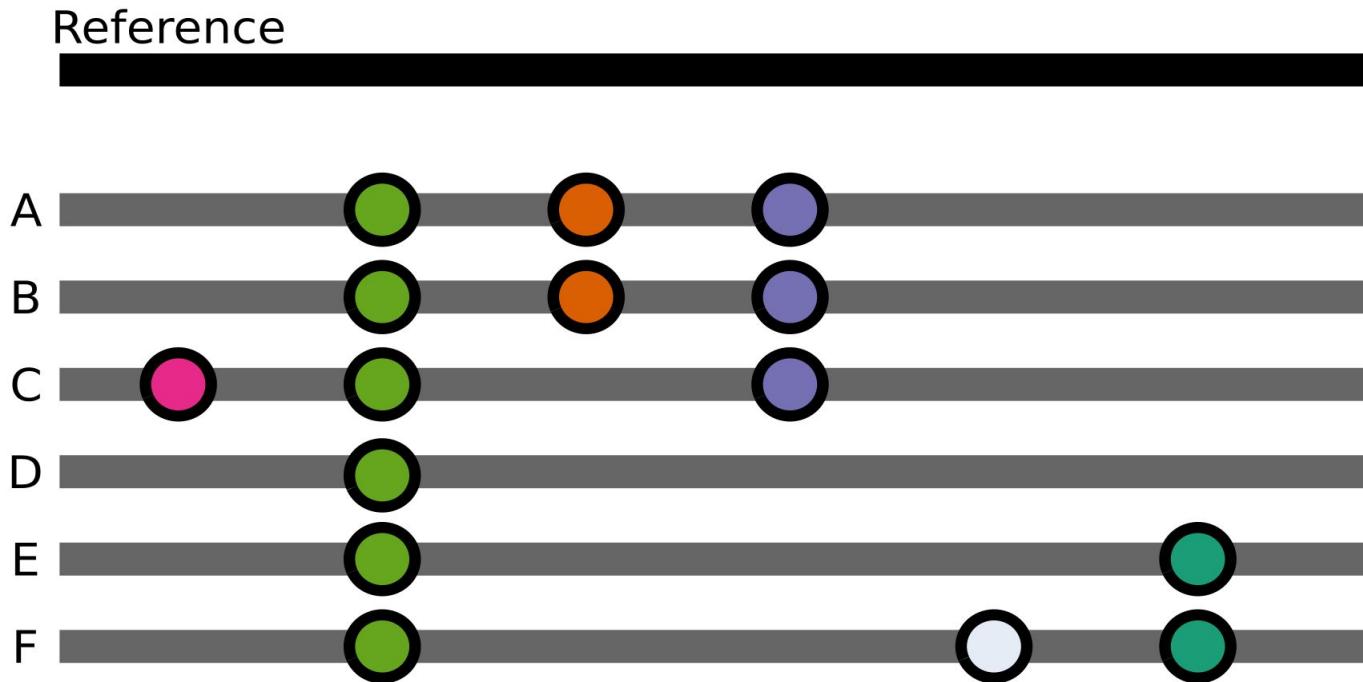
Compare mutations across patients



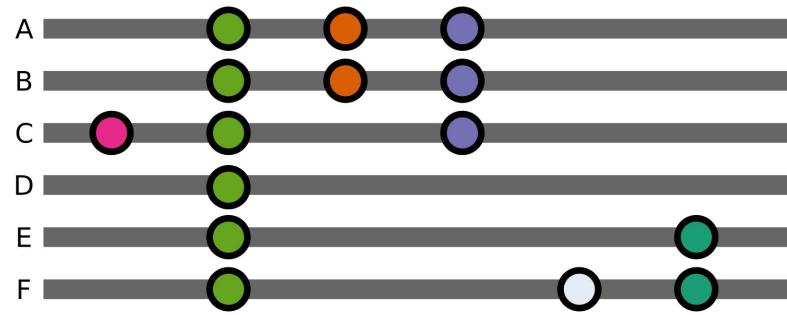
Compare mutations across patients



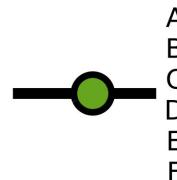
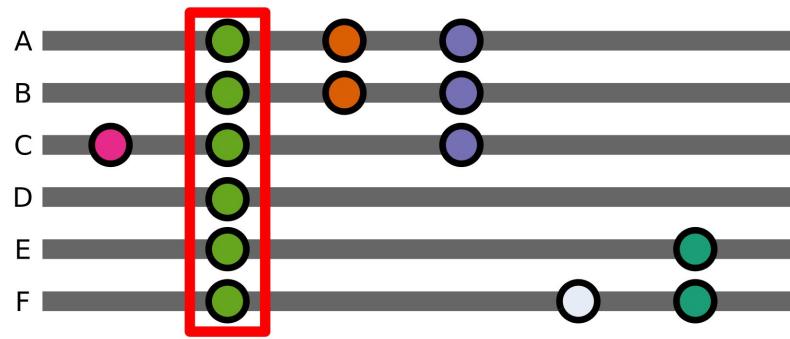
Compare mutations across patients



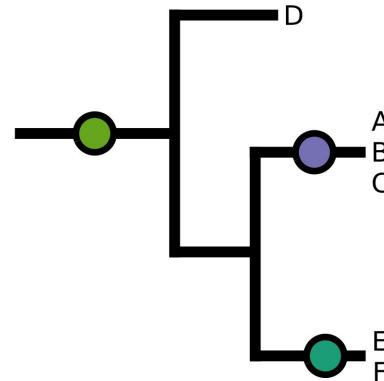
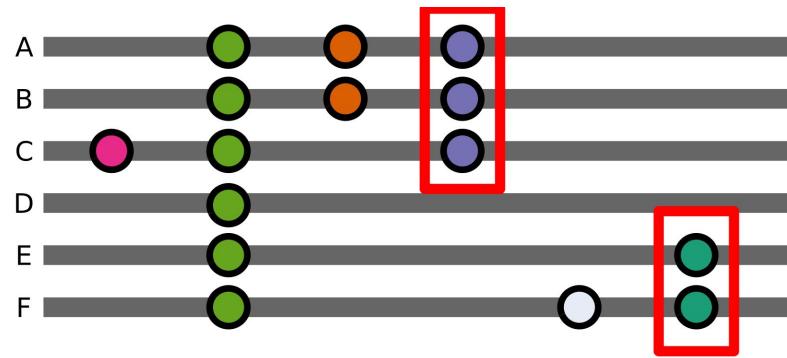
Using pattern of mutations to infer relationships



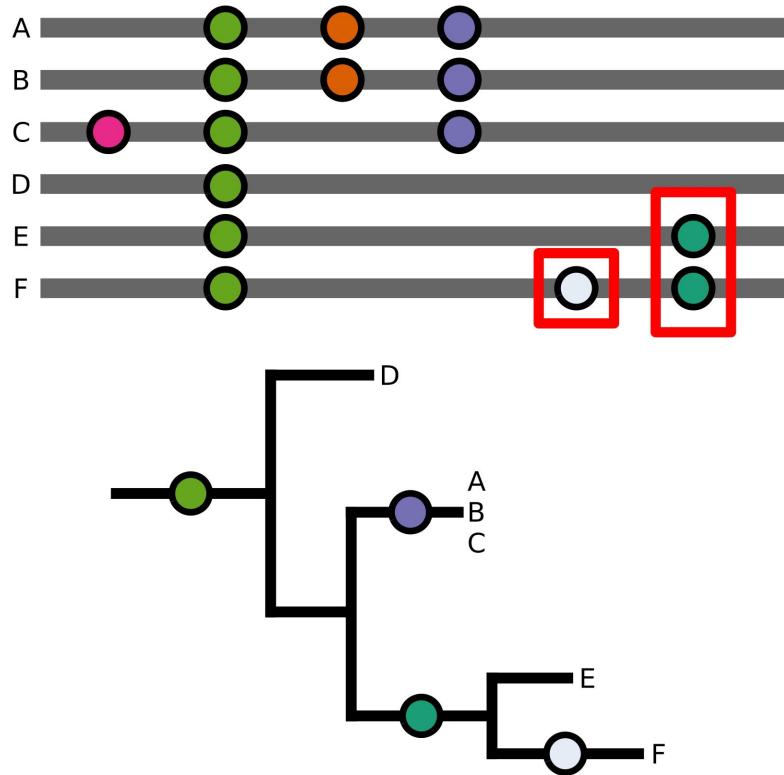
Using pattern of mutations to infer relationships



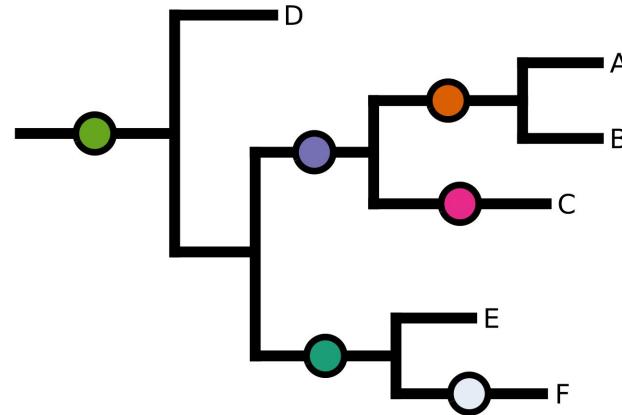
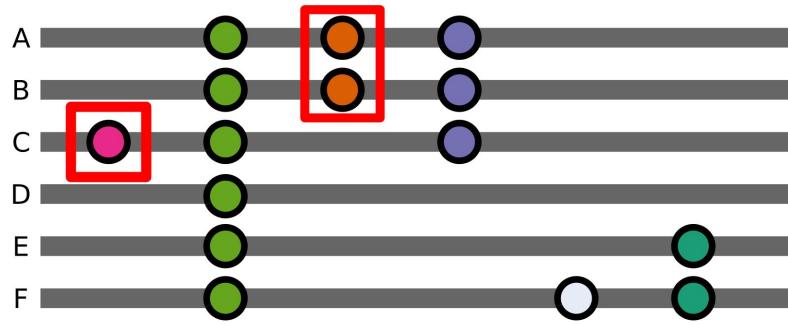
Using pattern of mutations to infer relationships



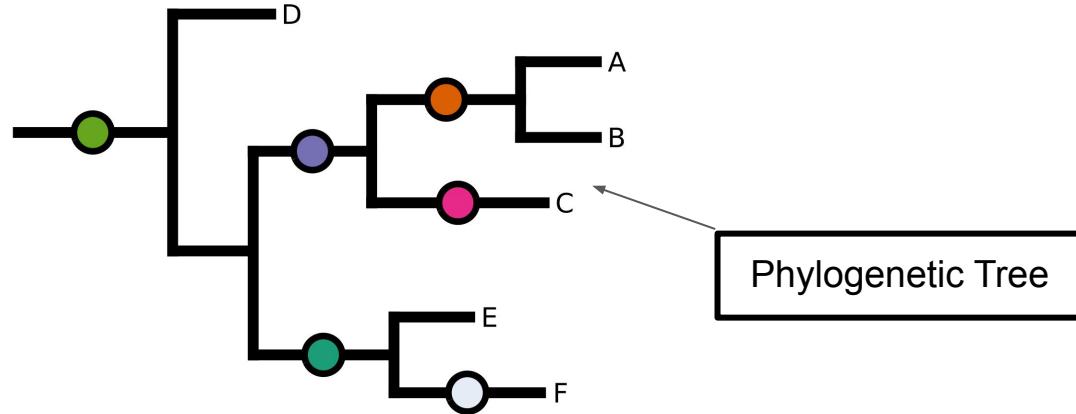
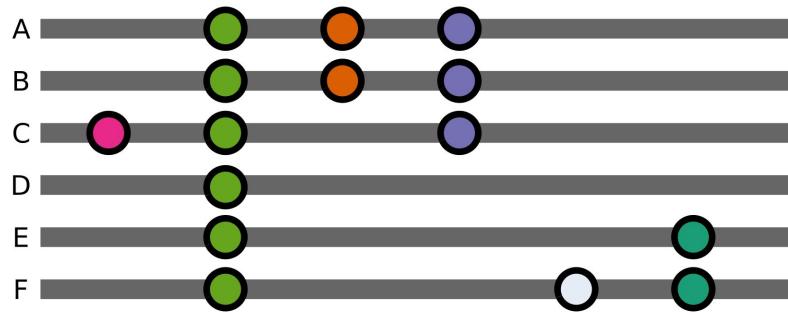
Using pattern of mutations to infer relationships



Using pattern of mutations to infer relationships

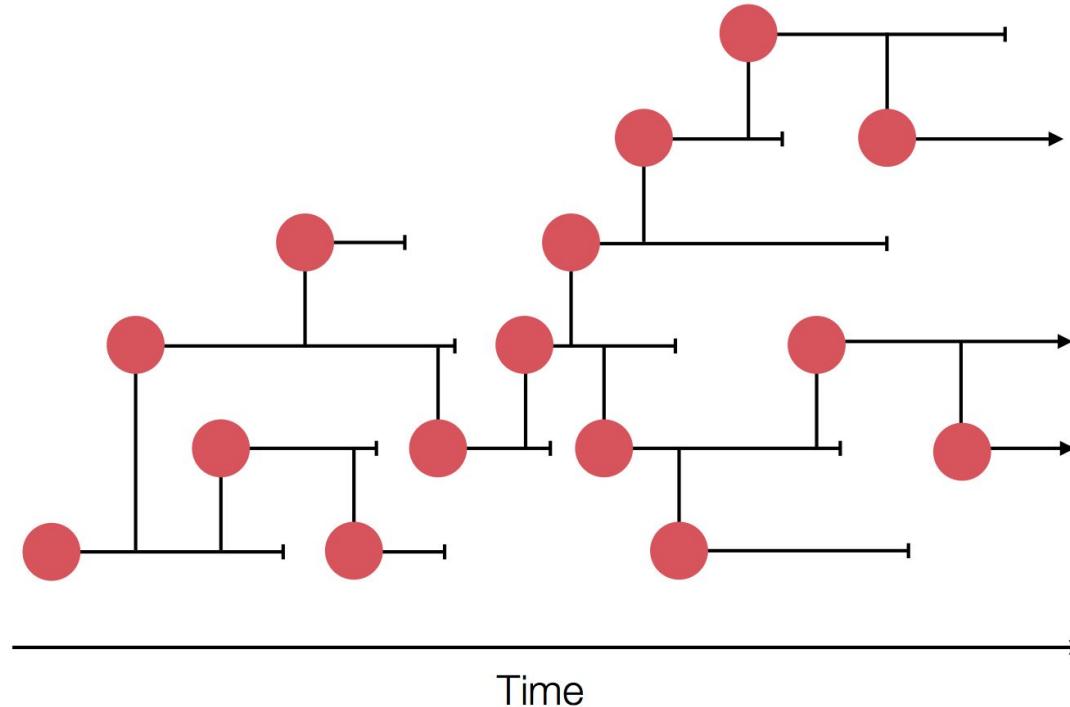


Using pattern of mutations to infer relationships

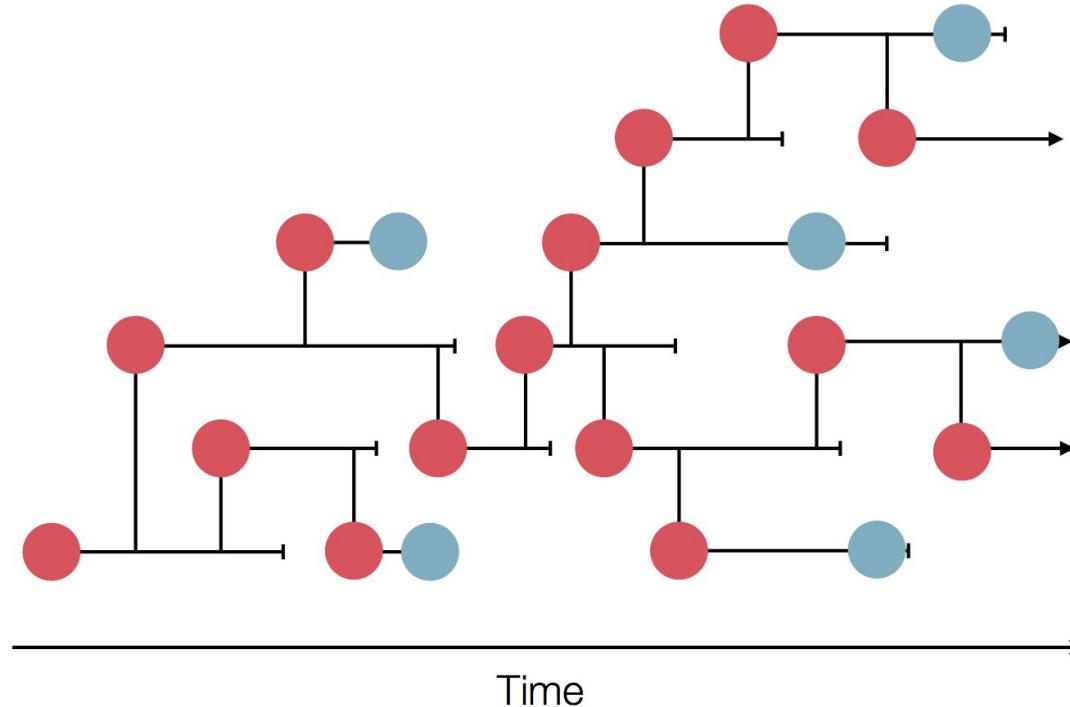


What does this tree actually represent?

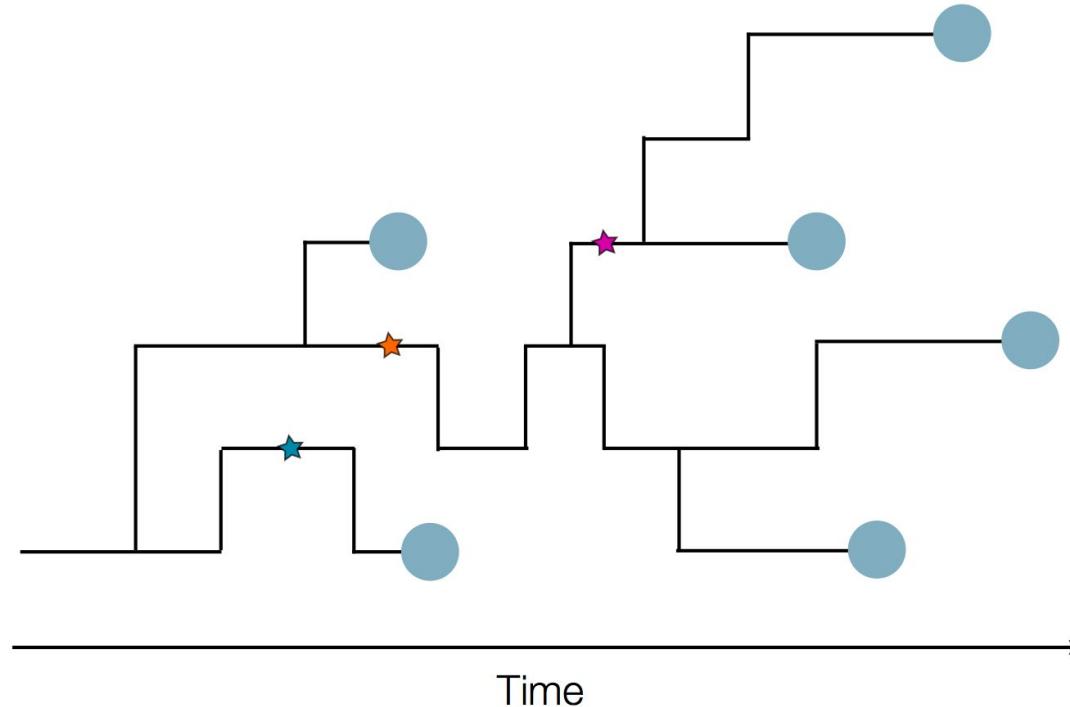
Sampling & partially reconstructing underlying epidemic process



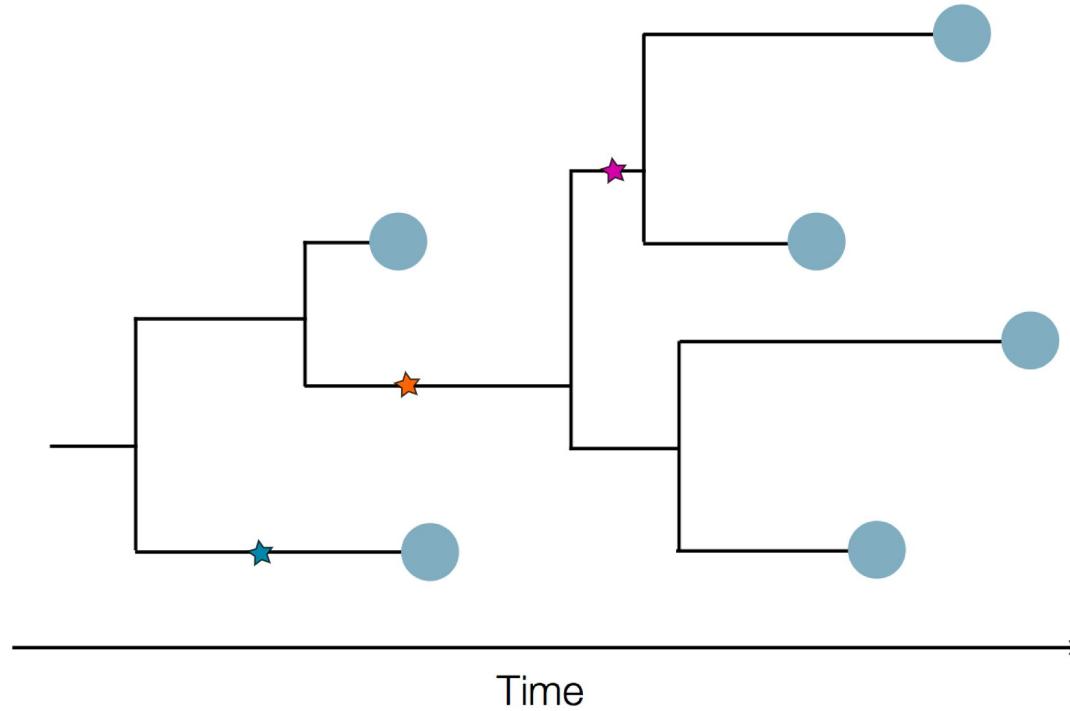
Sampling & partially reconstructing underlying epidemic process



Sampling & partially reconstructing underlying epidemic process

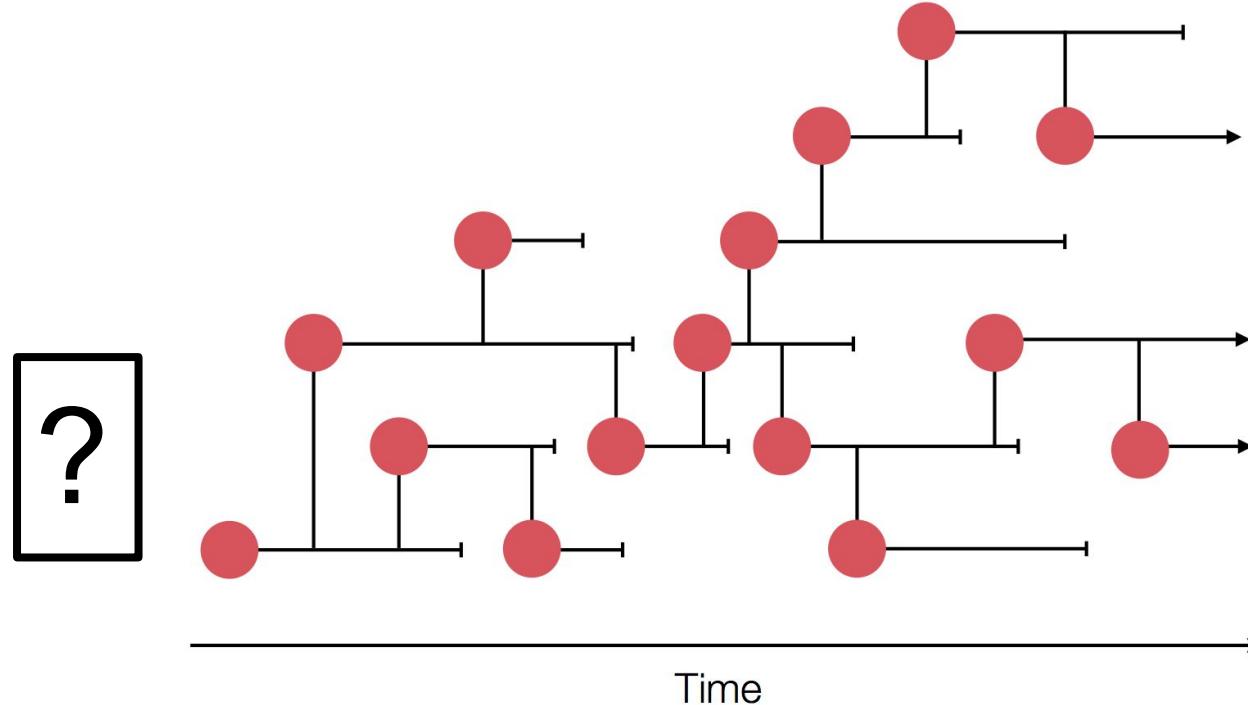


Sampling & partially reconstructing underlying epidemic process

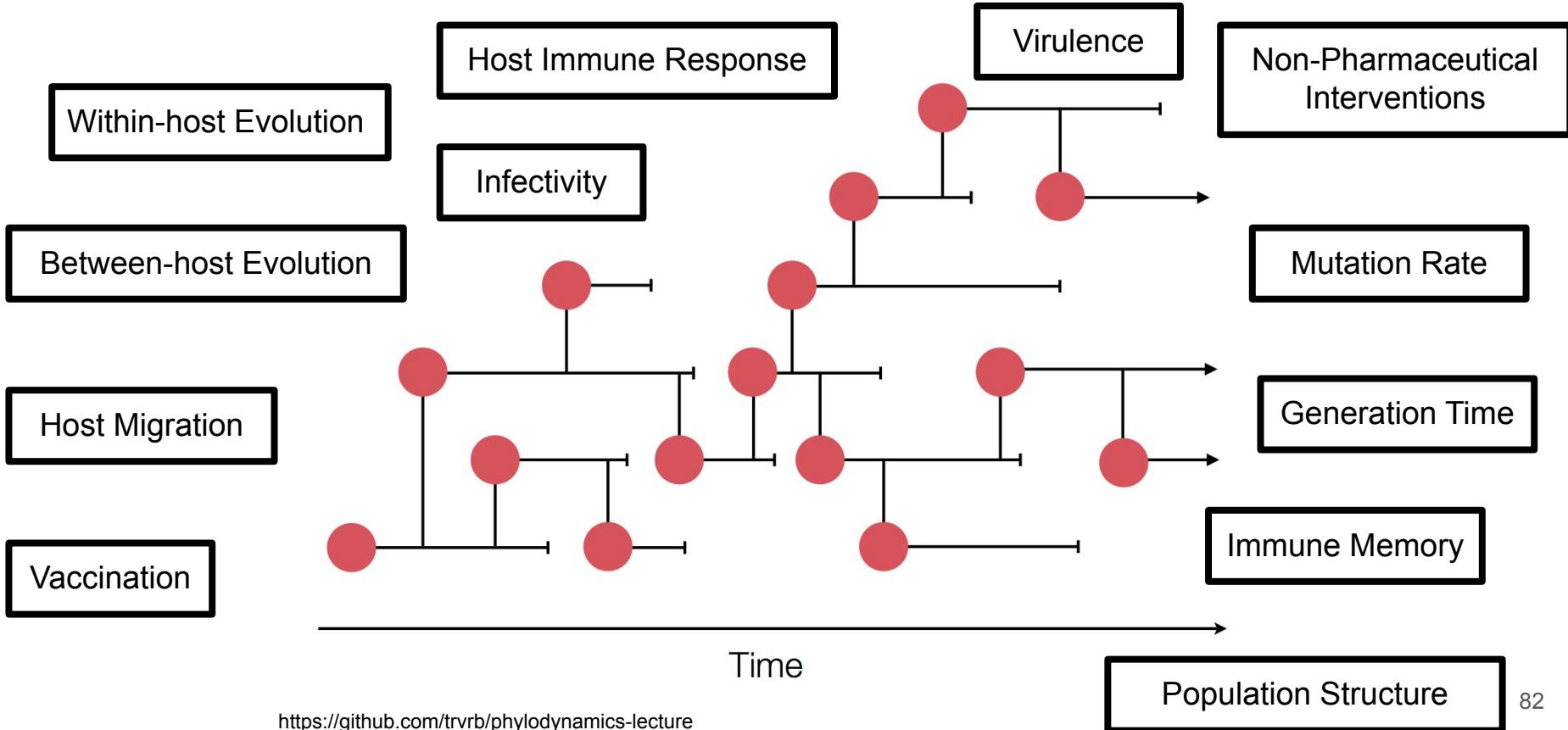


What determines underlying process?

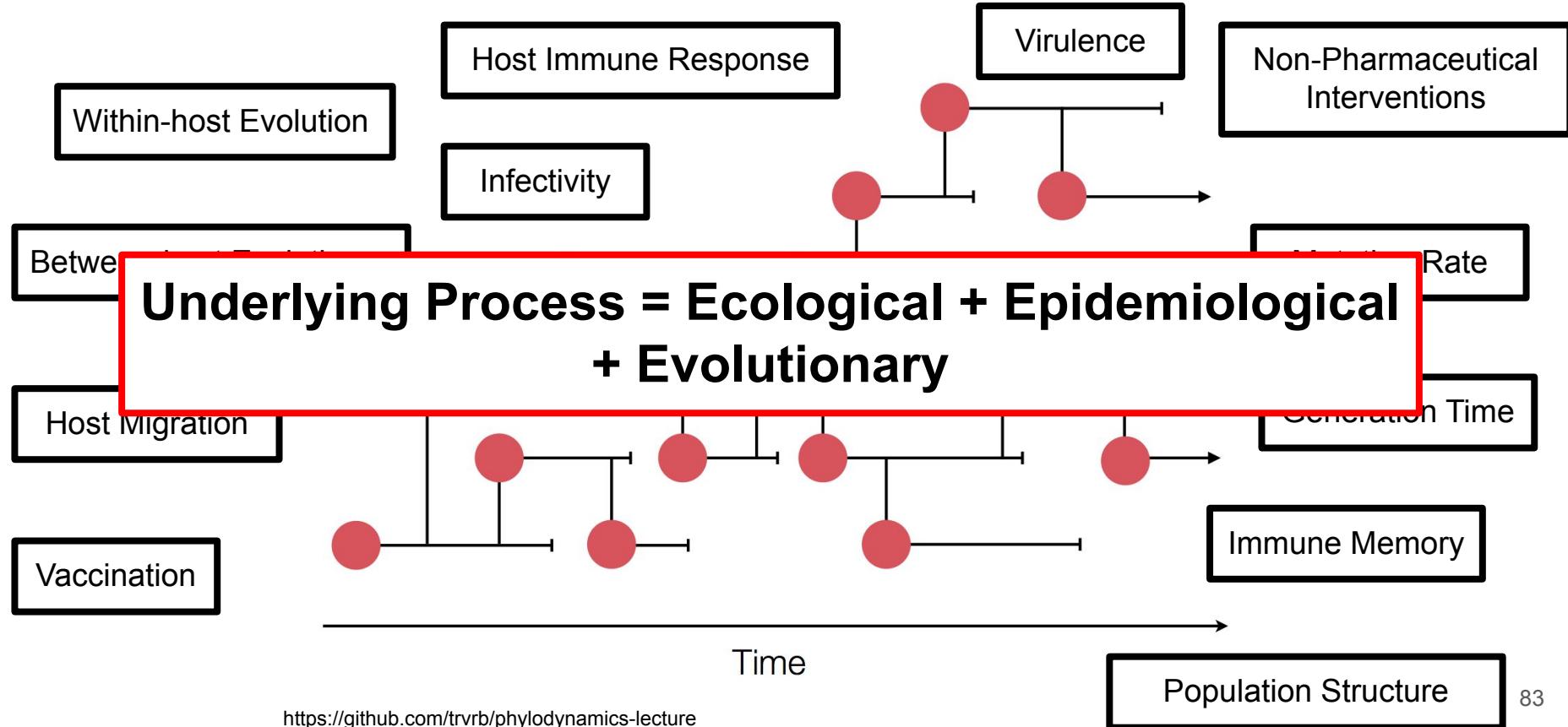
Many forces shaping underlying process



Many forces shaping underlying process

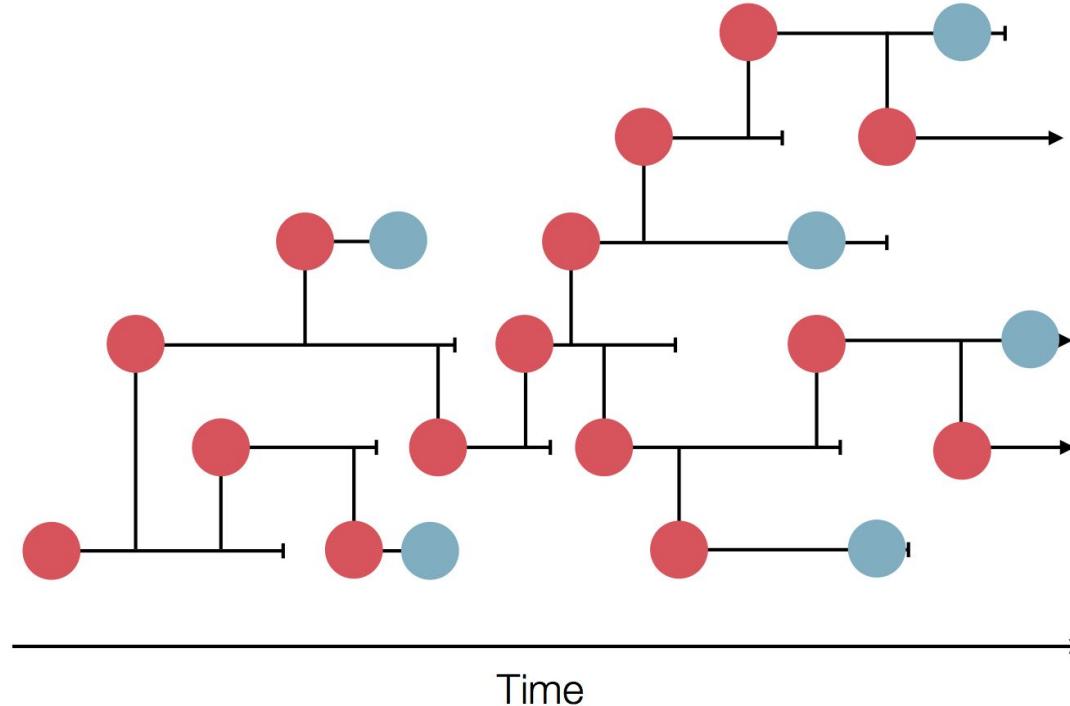


Many forces shaping underlying process

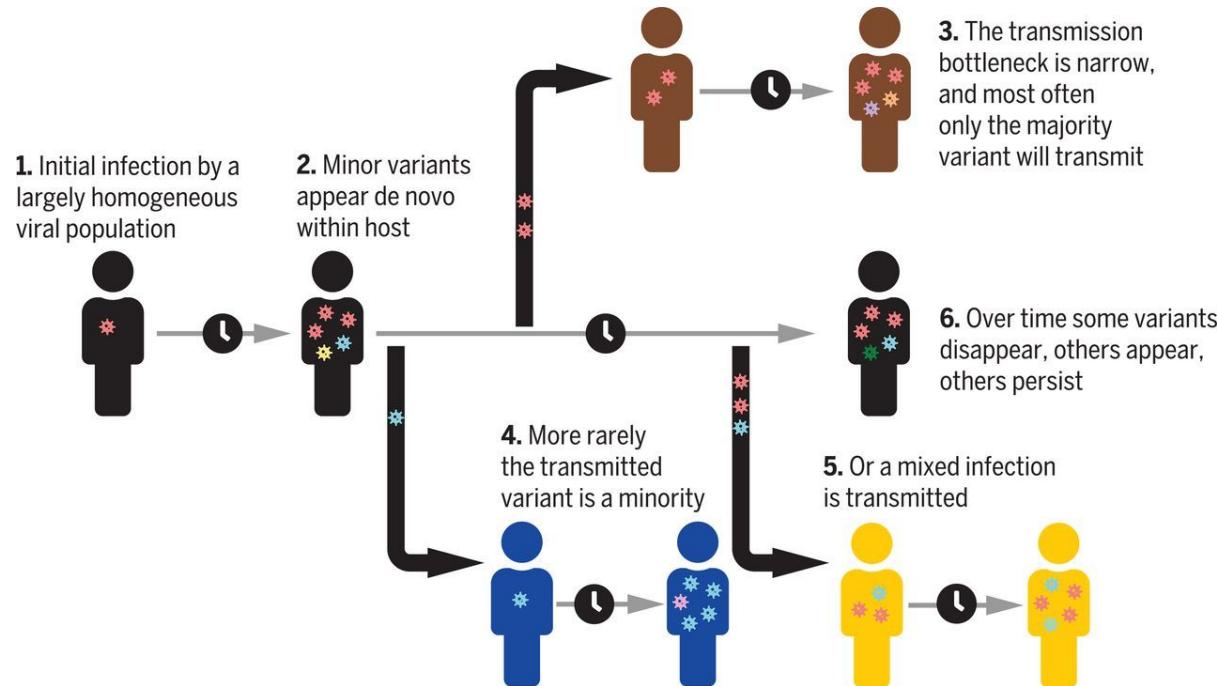


Let's start with the “simple” reconstruction of
the transmission network

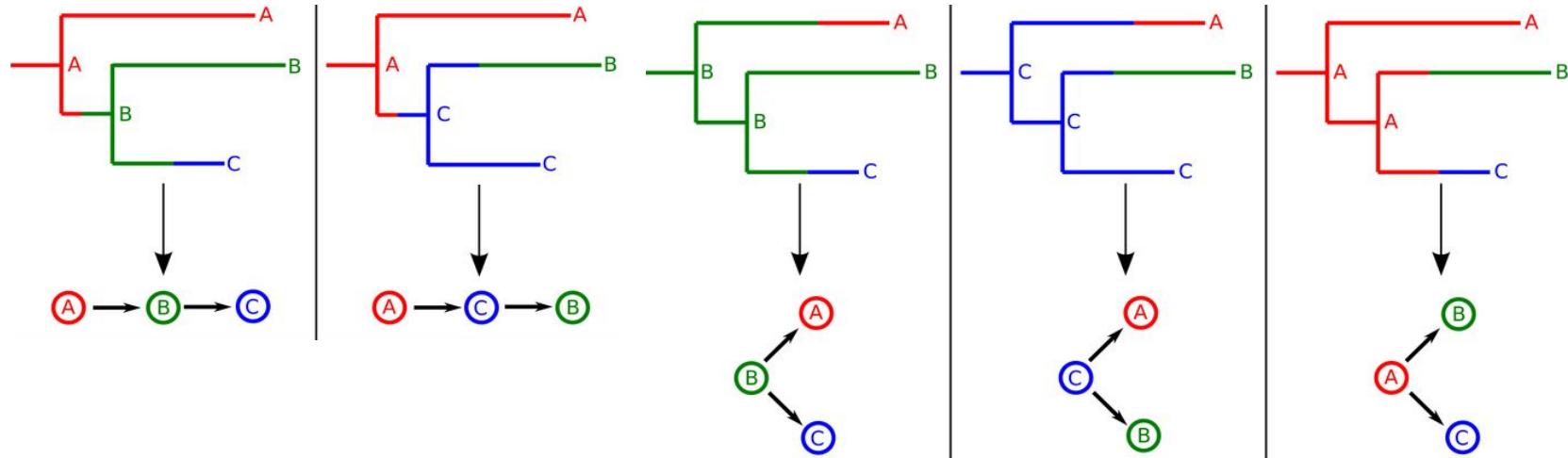
Complicated sampling of a (within-host) population of a (between host) population



Complicated sampling of a (within-host) population of a (between host) population

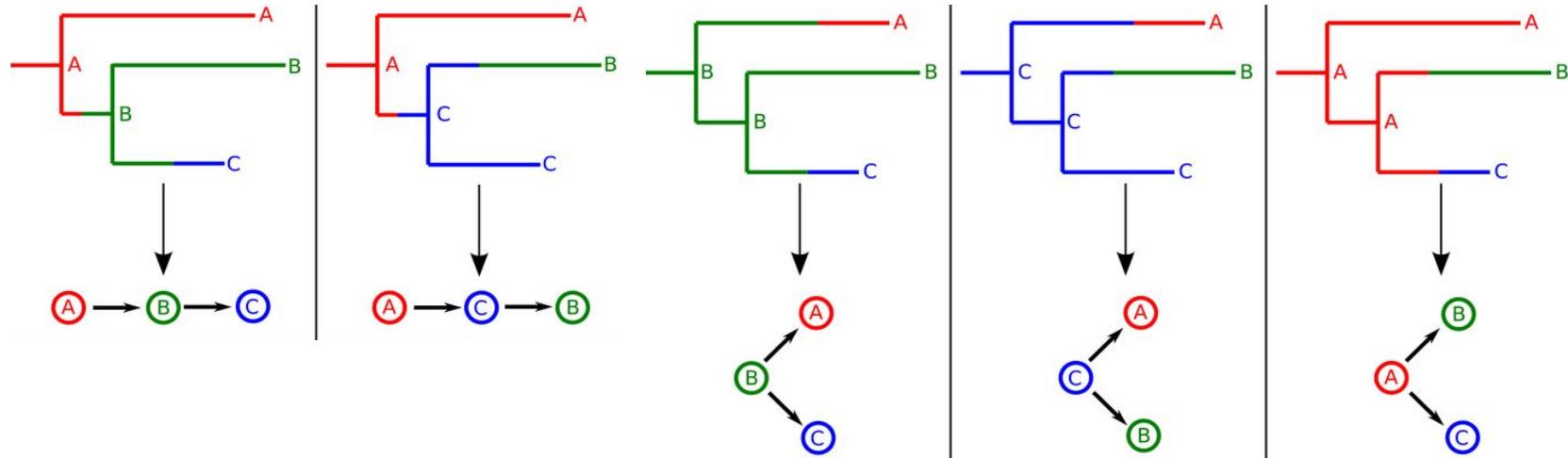


Same tree can be consistent with different scenarios



10.1371/journal.pcbi.1004613

Same tree can be consistent with different scenarios



10.1371/journal.pcbi.1004613

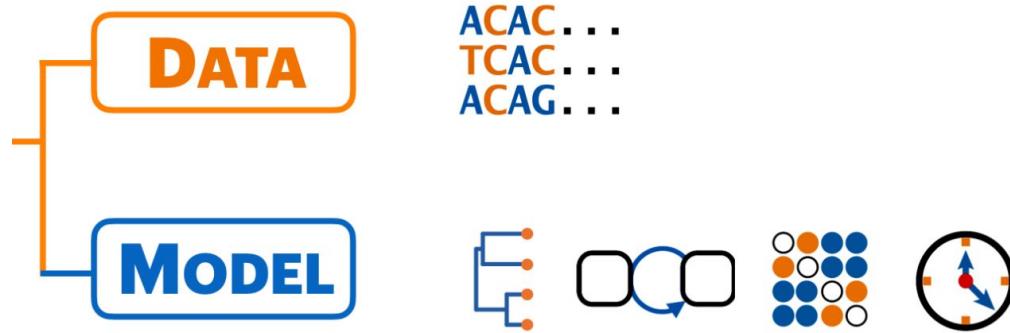
=> Probabilistic inference!

How can we model phylodynamic processes probabilistically?

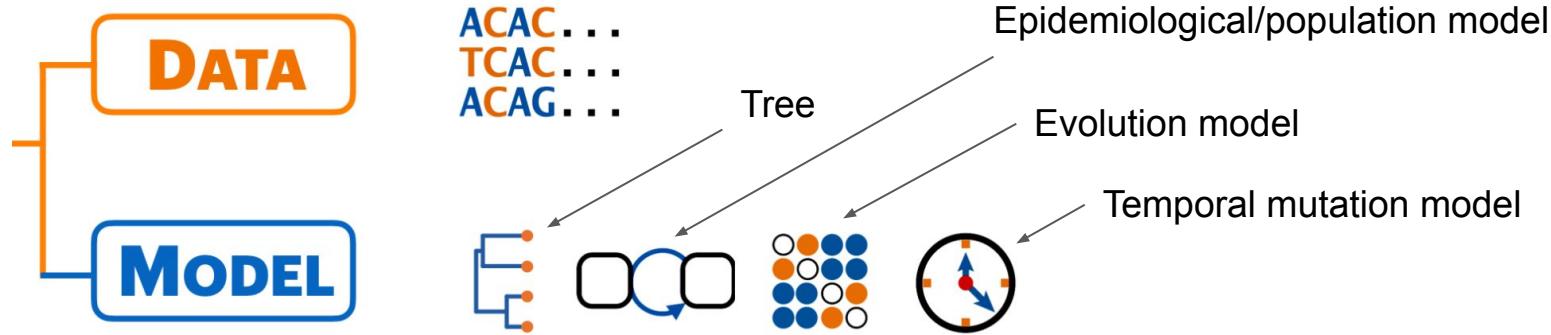
Bayesian inference is a key tool in genomic epidemiology



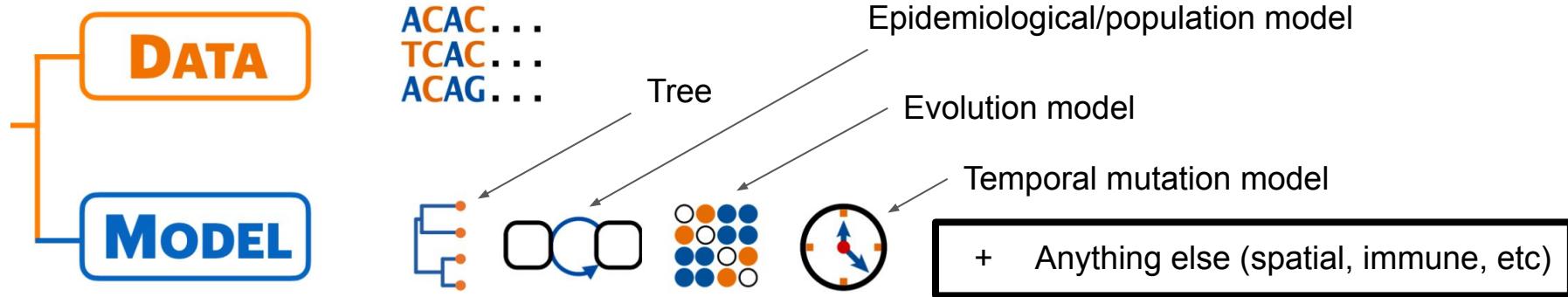
Bayesian inference is a key tool in genomic epidemiology



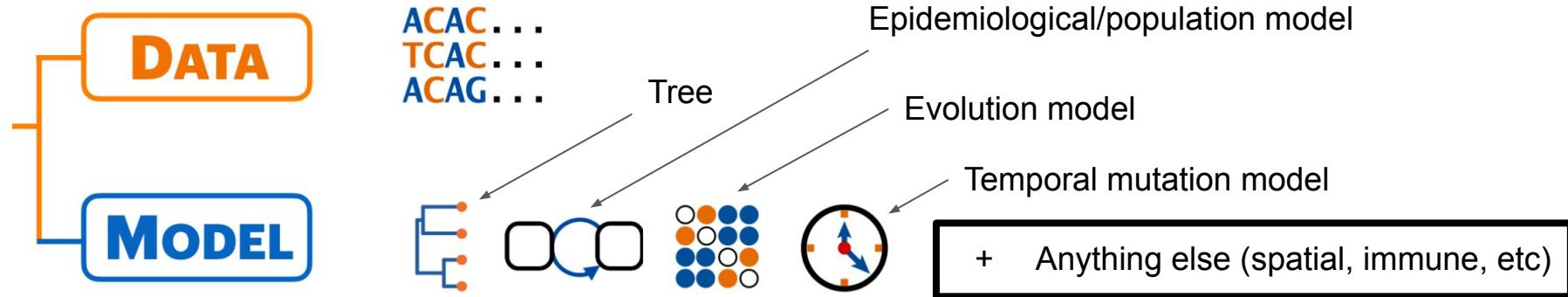
Bayesian inference is a key tool in genomic epidemiology



Bayesian inference is a key tool in genomic epidemiology

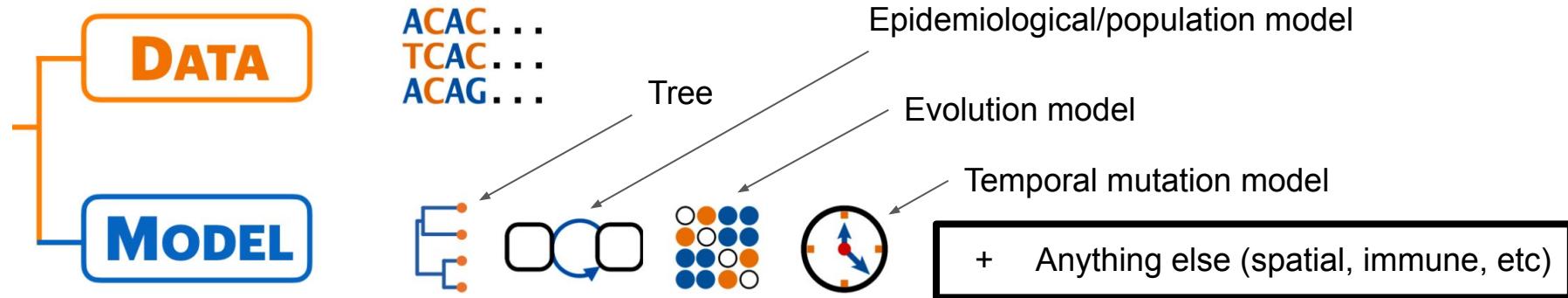


Bayesian inference is a key tool in genomic epidemiology



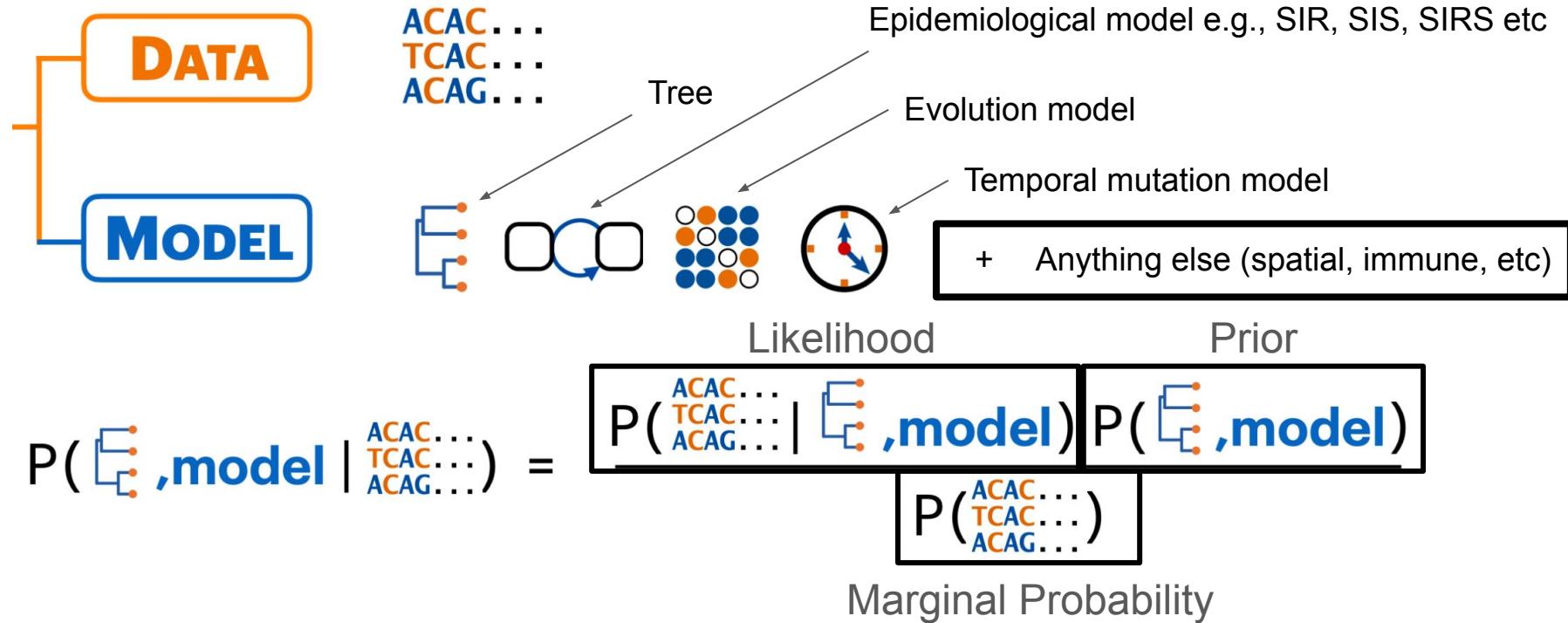
$$P(\text{E}, \text{model} | \text{ACAC...}, \text{TCAC...}, \text{ACAG...})$$

Bayesian inference is a key tool in genomic epidemiology



$$P(\text{E} \cdot, \text{model} | \text{ACAC...}, \text{TCAC...}, \text{ACAG...}) = \frac{P(\text{ACAC...}, \text{TCAC...}, \text{ACAG...} | \text{E} \cdot, \text{model}) P(\text{E} \cdot, \text{model})}{P(\text{ACAC...}, \text{TCAC...}, \text{ACAG...})}$$

Bayesian inference is a key tool in phylodynamics



Markov Chain Monte Carlo (MCMC) Sample

$$P(\text{E}^*, \text{model}^* | \text{ACAC...}) = \frac{P(\text{ACAC...} | \text{E}^*, \text{model}^*) P(\text{E}^*, \text{model}^*)}{P(\text{ACAC...})}$$

Intractable!

Markov Chain Monte Carlo (MCMC) Sample

$$P(\text{E}^*, \text{model}^* | \text{ACAC...TCAC...ACAG...}) = \frac{P(\text{ACAC...TCAC...ACAG...} | \text{E}^*, \text{model}^*) P(\text{E}^*, \text{model}^*)}{P(\text{ACAC...TCAC...ACAG...})}$$

$$P(\text{E}^*, \text{model}^* | \text{ACAC...TCAC...ACAG...}) = \frac{P(\text{ACAC...TCAC...ACAG...} | \text{E}^*, \text{model}^*) P(\text{E}^*, \text{model}^*)}{P(\text{ACAC...TCAC...ACAG...})}$$

Posterior Odds

Markov Chain Monte Carlo (MCMC) Sample

$$P(\text{E}^*, \text{model}^* | \text{ACAC...TCAC...ACAG...}) = \frac{P(\text{ACAC...TCAC...ACAG...} | \text{E}^*, \text{model}^*) P(\text{E}^*, \text{model}^*)}{P(\text{ACAC...TCAC...ACAG...})}$$

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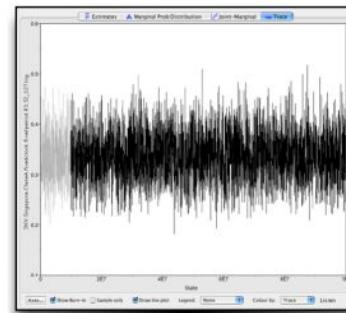
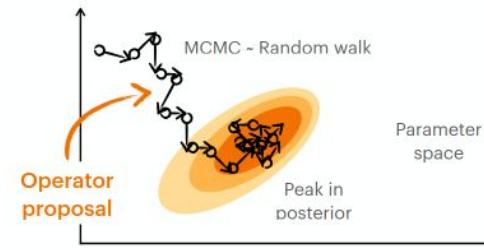
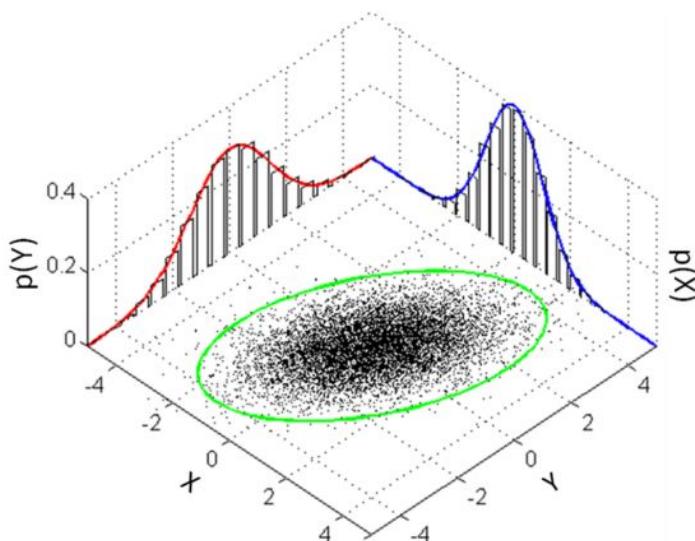
Posterior Odds

Likelihood Ratio

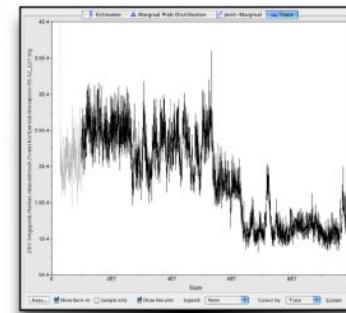
Prior Odds

Markov Chain Monte Carlo (MCMC) Sample

We sample from the joint posterior



Mixing well! 😊

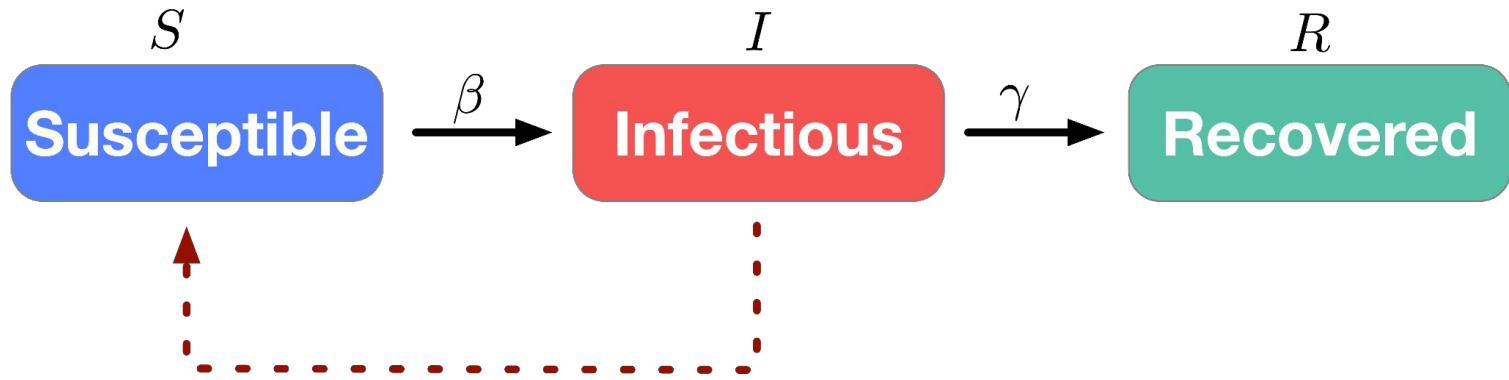


Not mixing! 😠

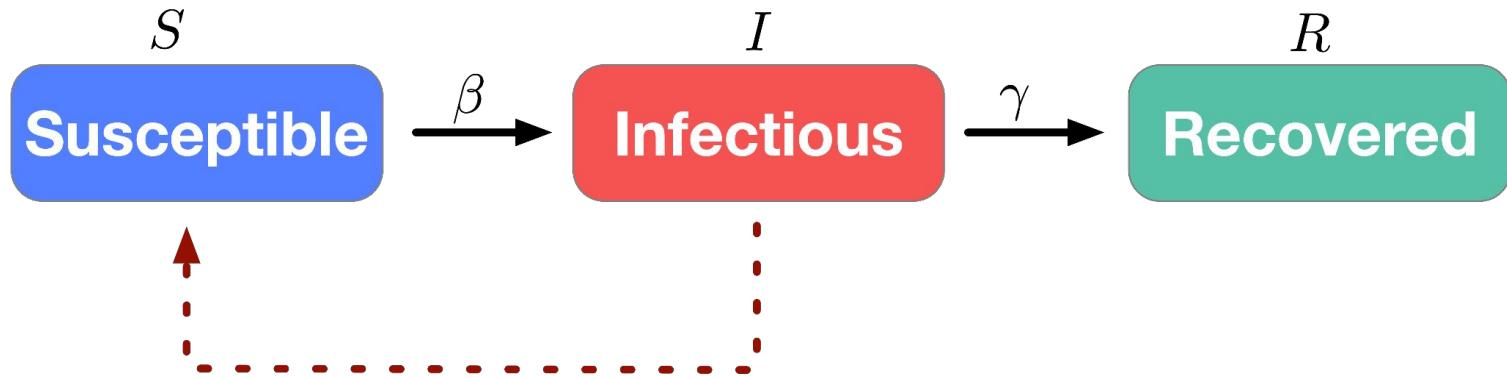
Live Demo - <https://delphy.fathom.info/>

What kind of epidemic process parameters
do we want to infer?

Compartmental models are used to model infections

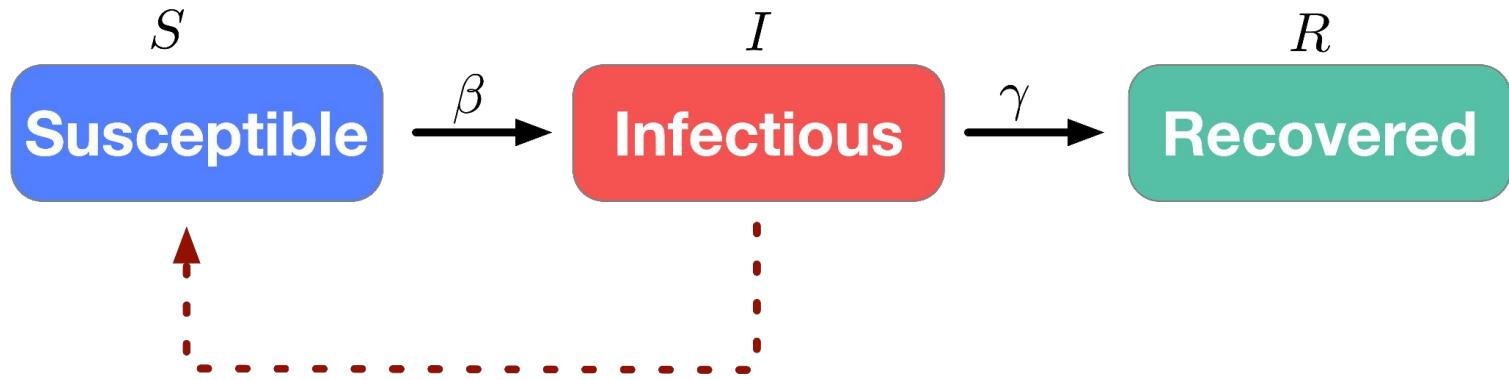


Compartmental models are used to model infections



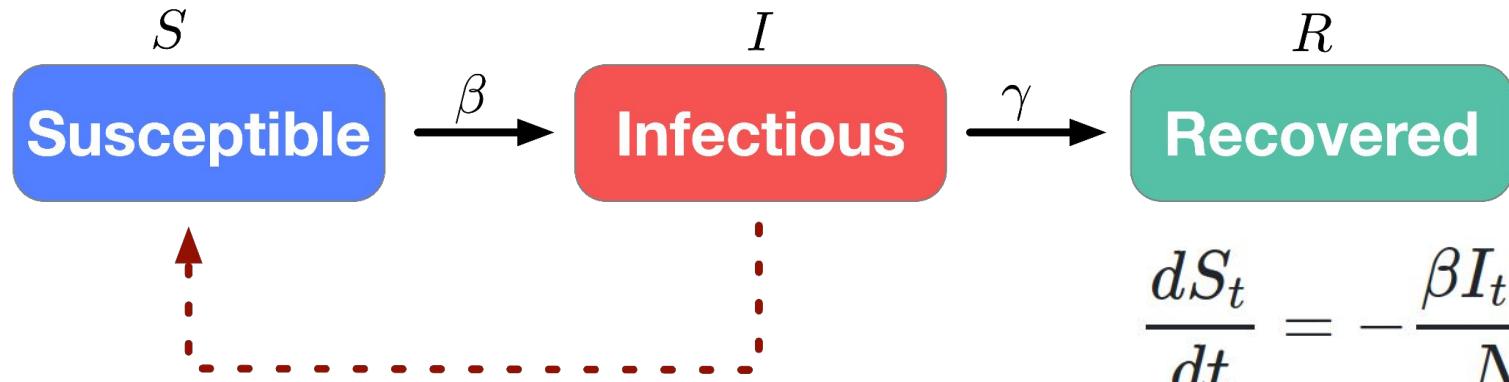
Disclaimer: many more complex models!

Compartmental models are used to model infections



- S_t : the number of susceptible individuals
- I_t : the number of infectious individuals
- R_t : the number of recovered/deceased/immune individuals

Compartmental models are used to model infections



- S_t : the number of susceptible individuals
- I_t : the number of infectious individuals
- R_t : the number of recovered/deceased/immune individuals

$$\frac{dS_t}{dt} = -\frac{\beta I_t S_t}{N}$$

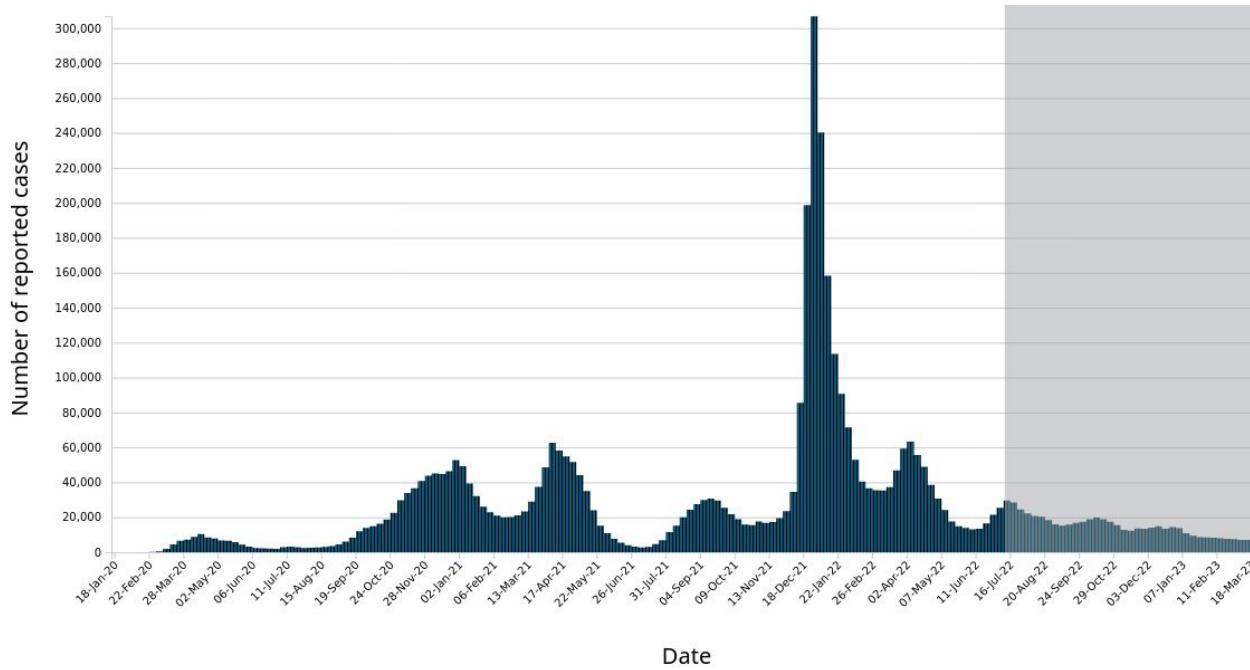
$$\frac{dI_t}{dt} = \frac{\beta I_t S_t}{N} - \gamma I_t$$

$$\frac{dR_t}{dt} = \gamma I_t$$

Assuming N = fixed pop

Can calculate $P(\text{observed case counts} \mid \beta=? , \gamma=?)$ with cases

Figure 2. Weekly number of COVID-19 (n=4,359,630) in Canada as of April 3, 2023, 9 am ET



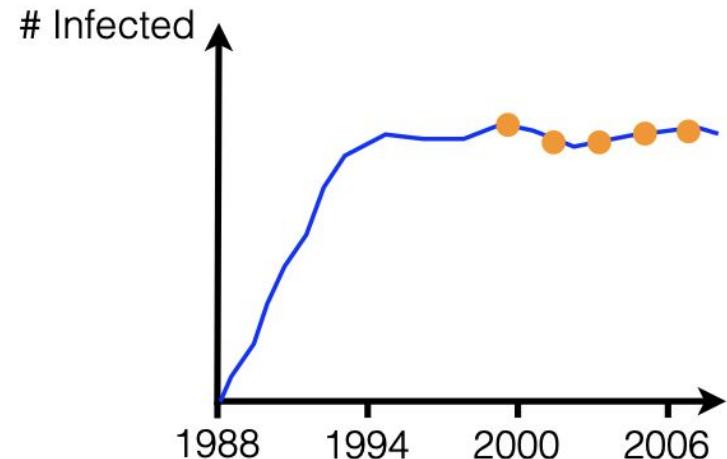
Same idea as Maximum likelihood Phylogenetics
(just without any trees)

So, if we can this with case why do we bother using phylodynamics and genomic data?

Genomics can be used to infer unobserved events

If sampling in early epidemic was missed:

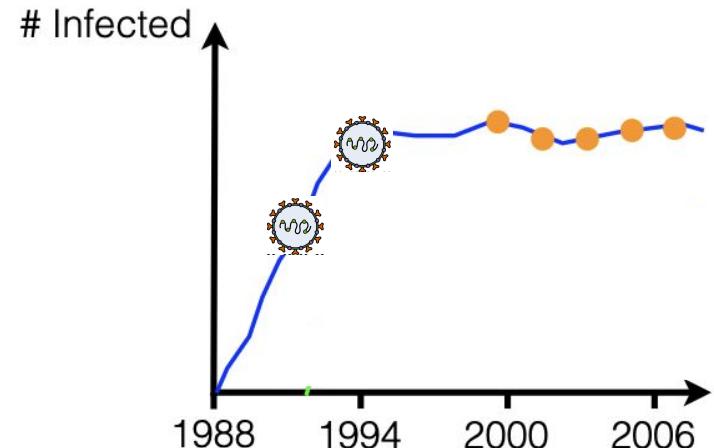
- ▶ **Time of epidemic outbreak?**
- ▶ **Basic reproductive number R_0 ?**



Genomics can be used to infer unobserved events

If sampling in early epidemic was missed:

- ▶ **Time of epidemic outbreak?**
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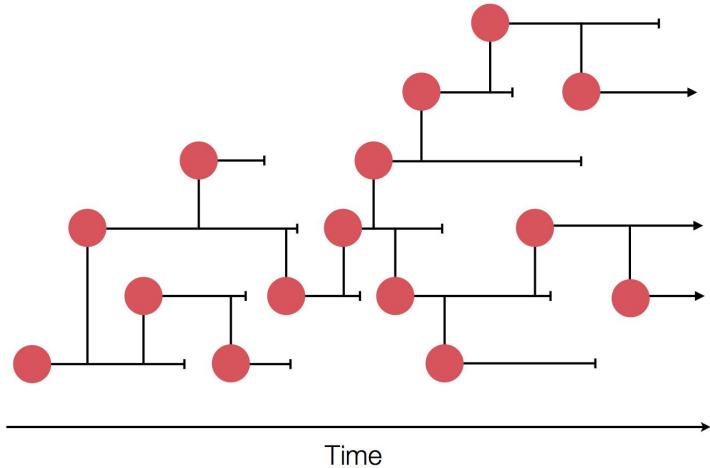
Genomics can be used to infer unobserved events

If sampling in early epidemic was missed:

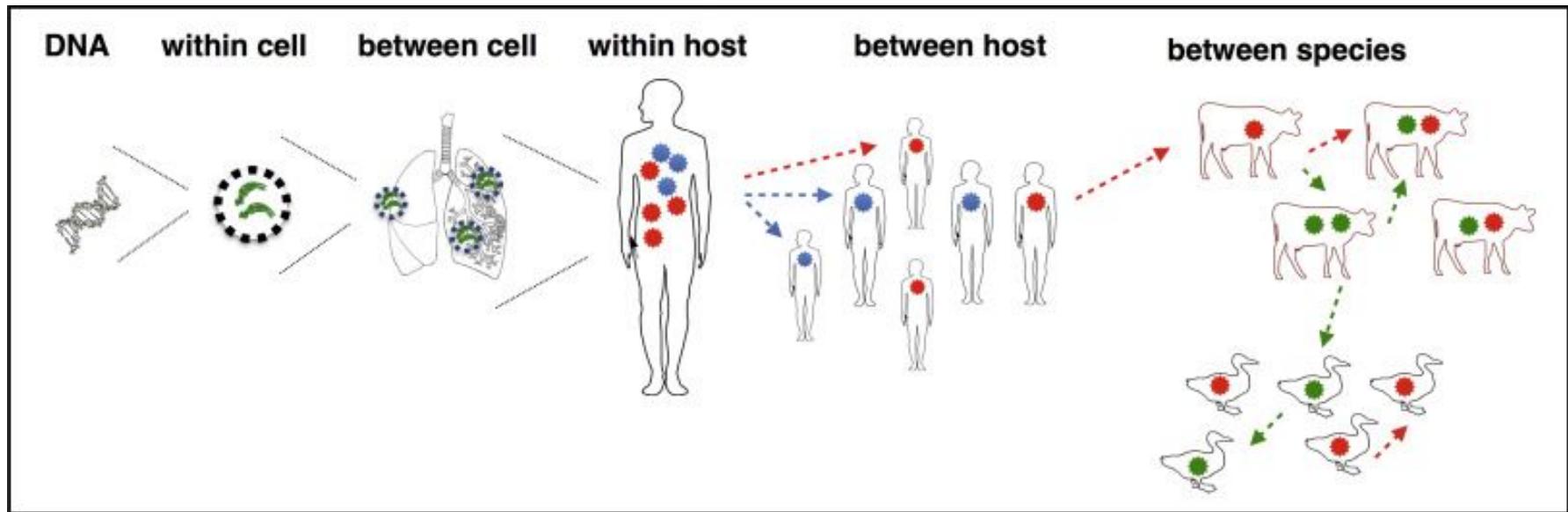
- ▶ **Time of epidemic outbreak?**
- ▶ **Basic reproductive number R_0 ?**

Data does not tell who infected whom:

- ▶ **Population structure?**



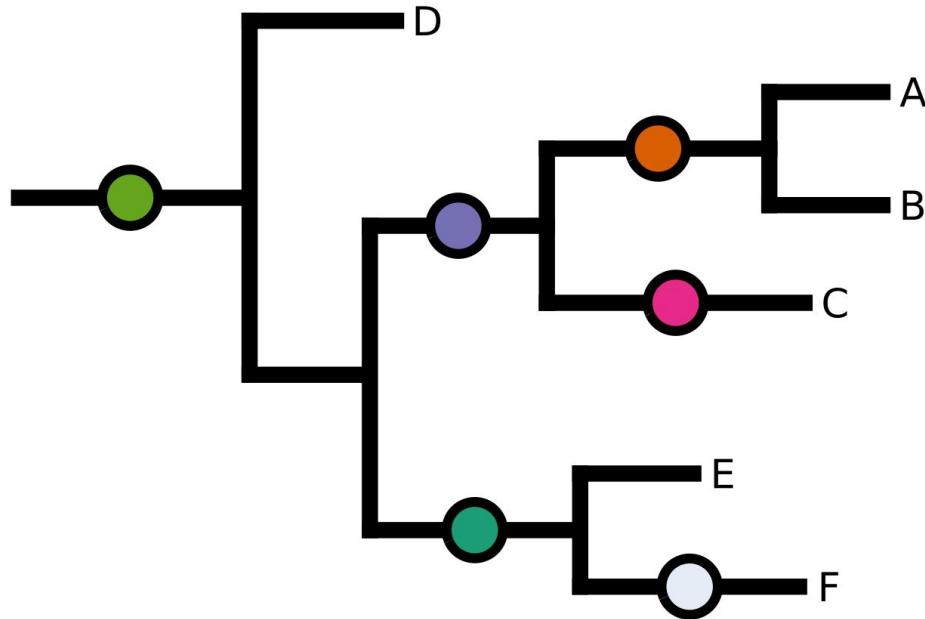
Cases don't tell you (much) about pathogen evolution



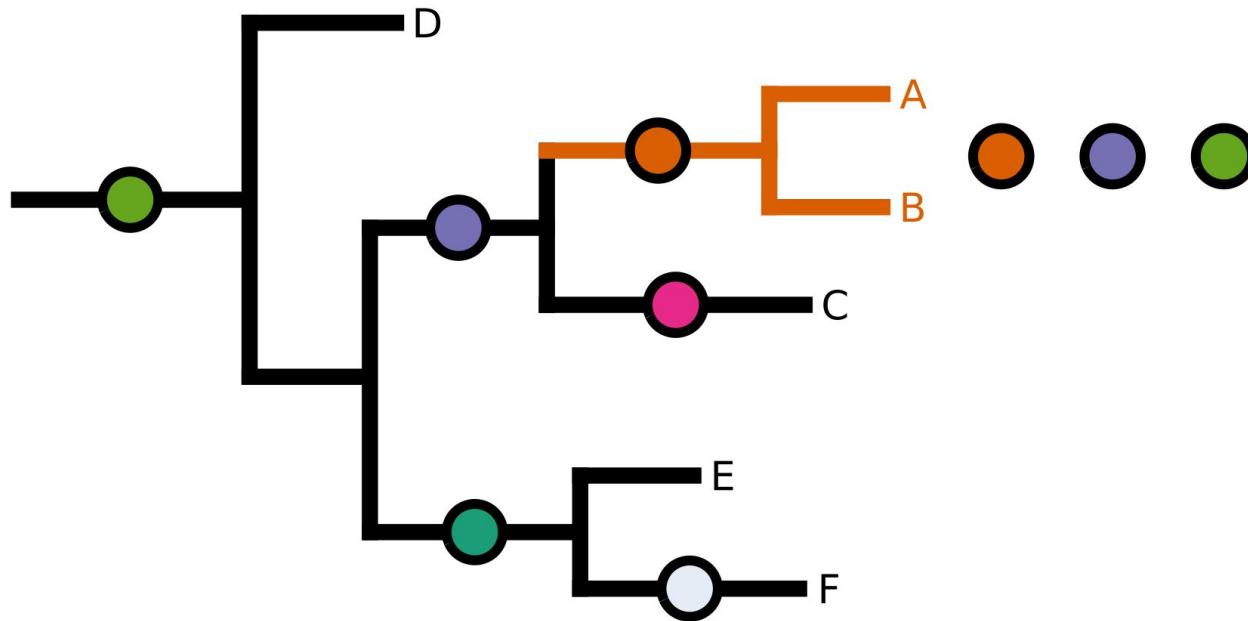
<https://www.sciencedirect.com/science/article/pii/S1755436514000723>

Let's look at some specific use-cases

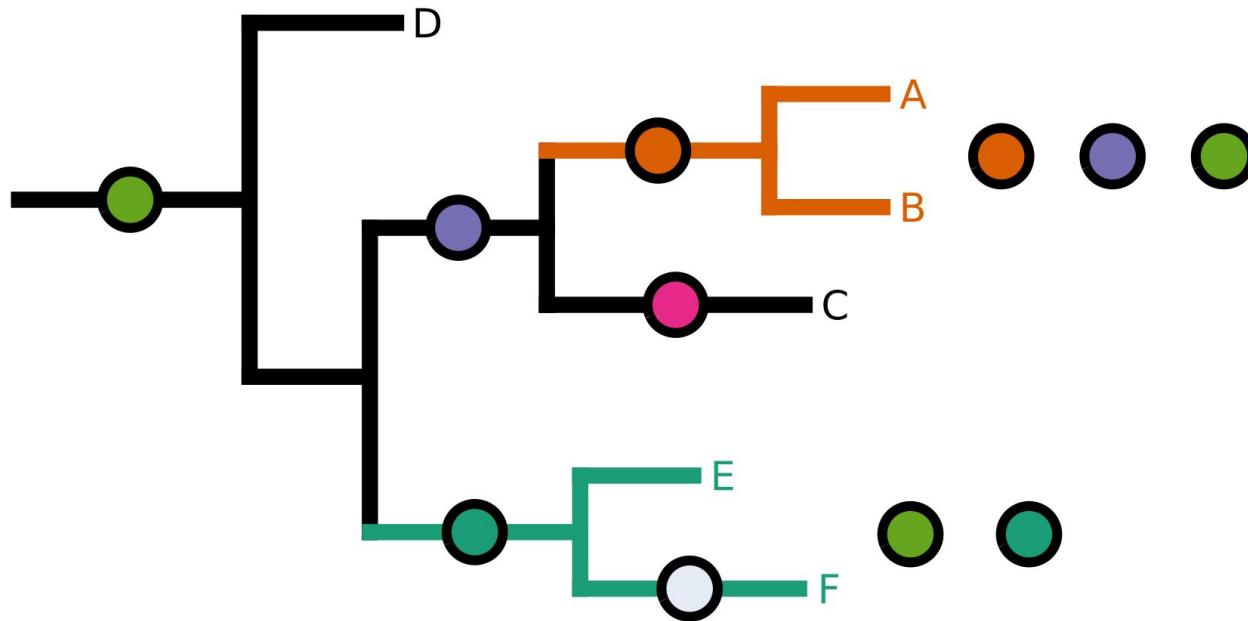
Define lineages (groups) of pathogens



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Define lineages (groups) of pathogens

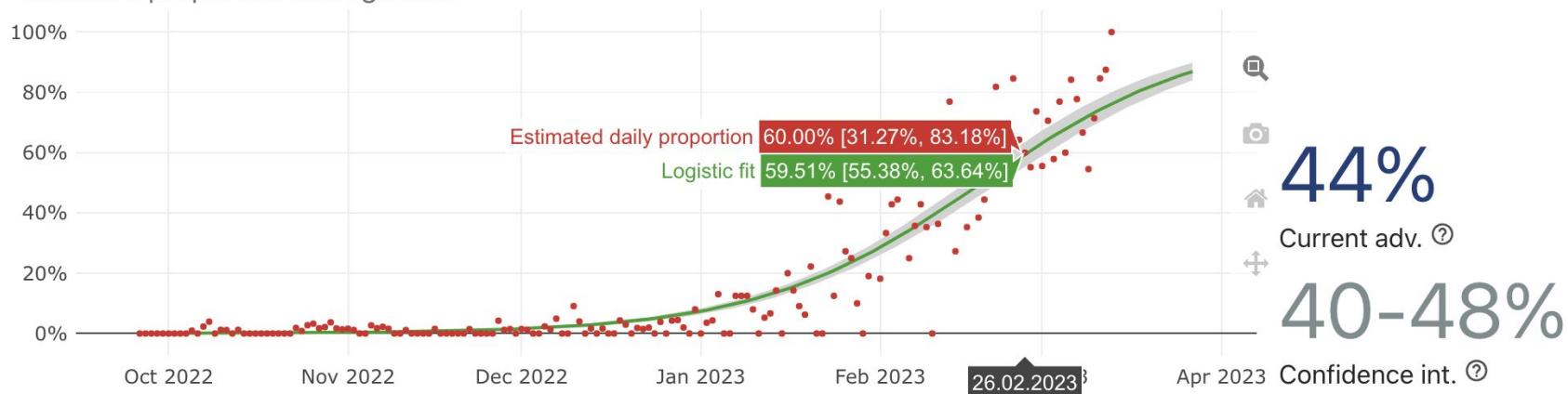


Early warning modelling lineage relative growth advantage

Relative growth advantage

If variants spread pre-dominantly by local transmission across demographic group... (show more)

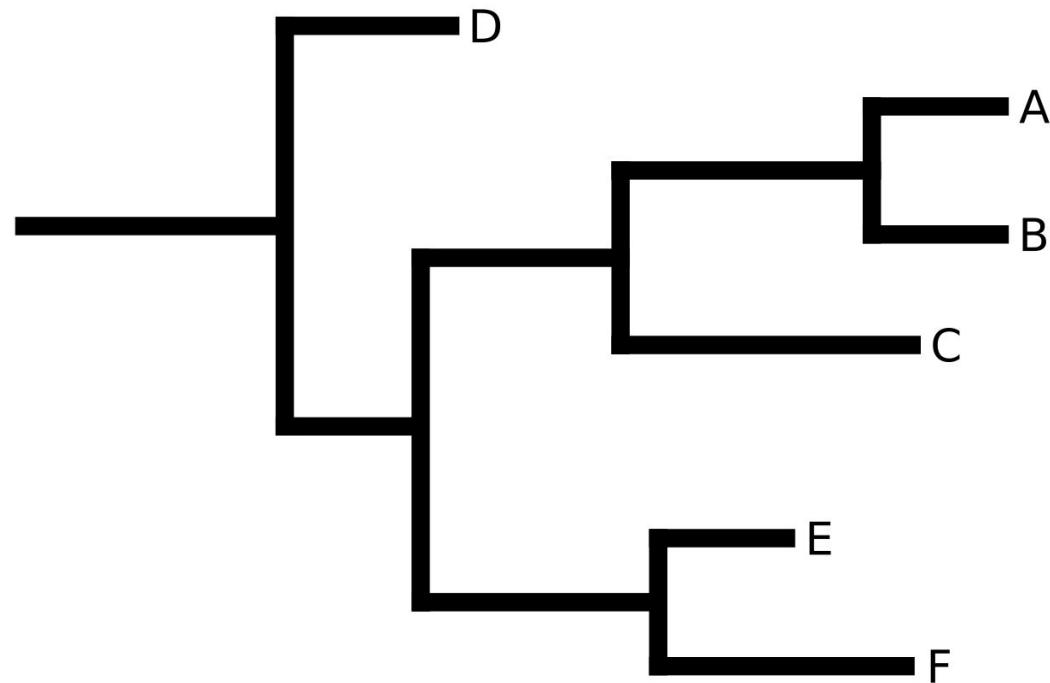
Estimated proportion through time



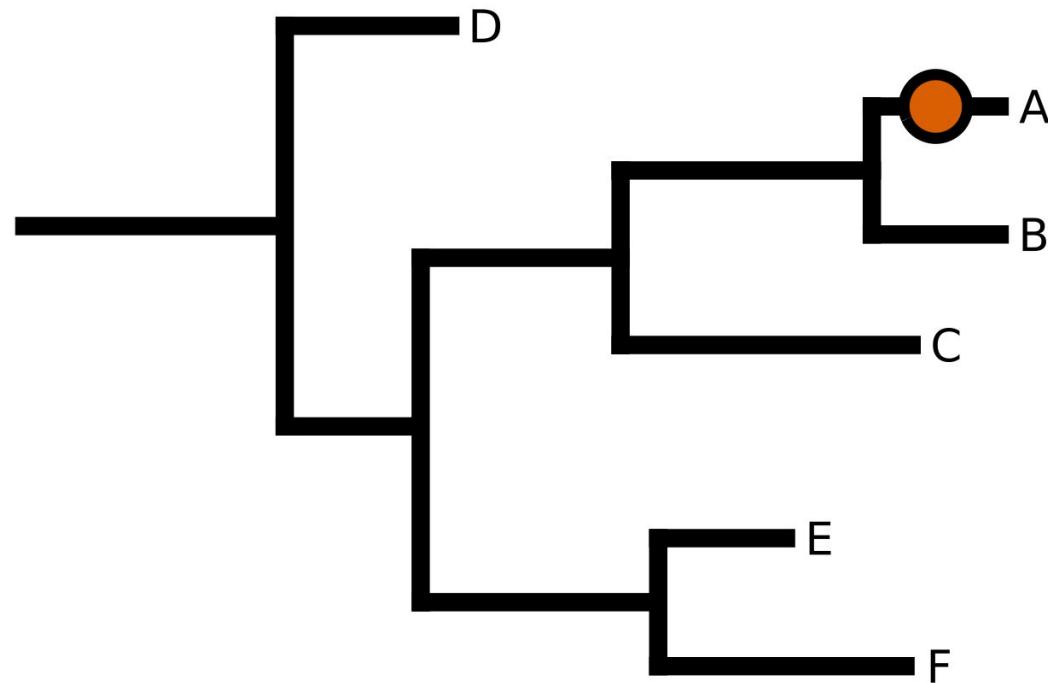
(*) Assumes that the current advantage is due to an intrinsic viral advantage (a combination of increased transmission, immune escape, and prolonged infectious period).

https://cov-spectrum.org/explore/Switzerland/Surveillance/Past6M/variants?nextcladePangoLineage=xbb*&

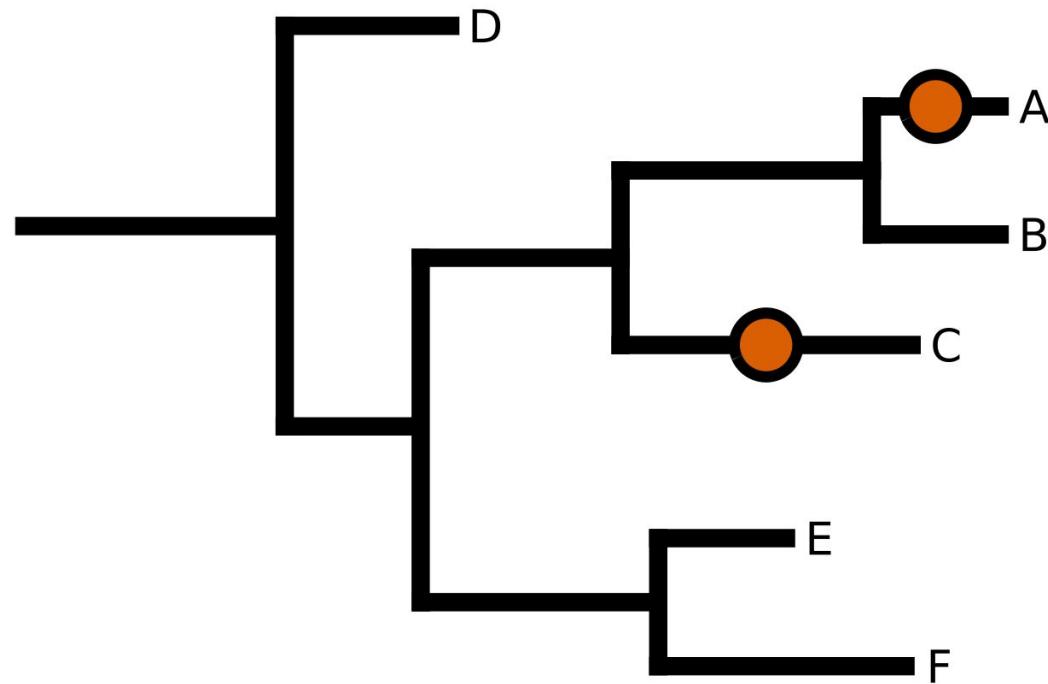
Identify clinically relevant mutations



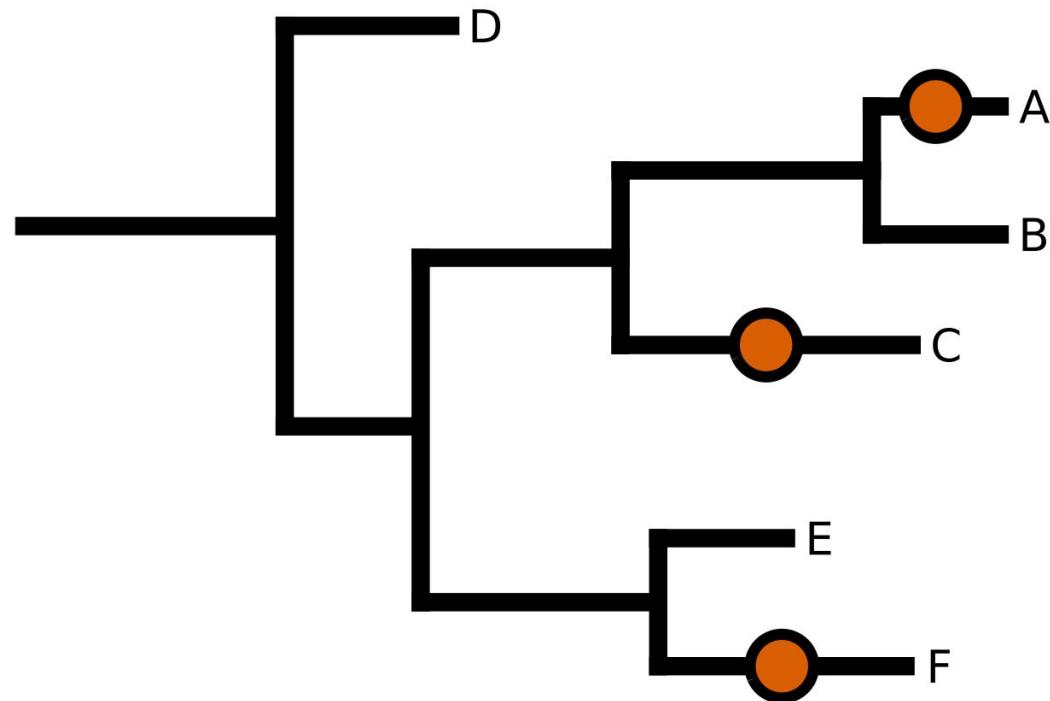
Identify clinically relevant mutations



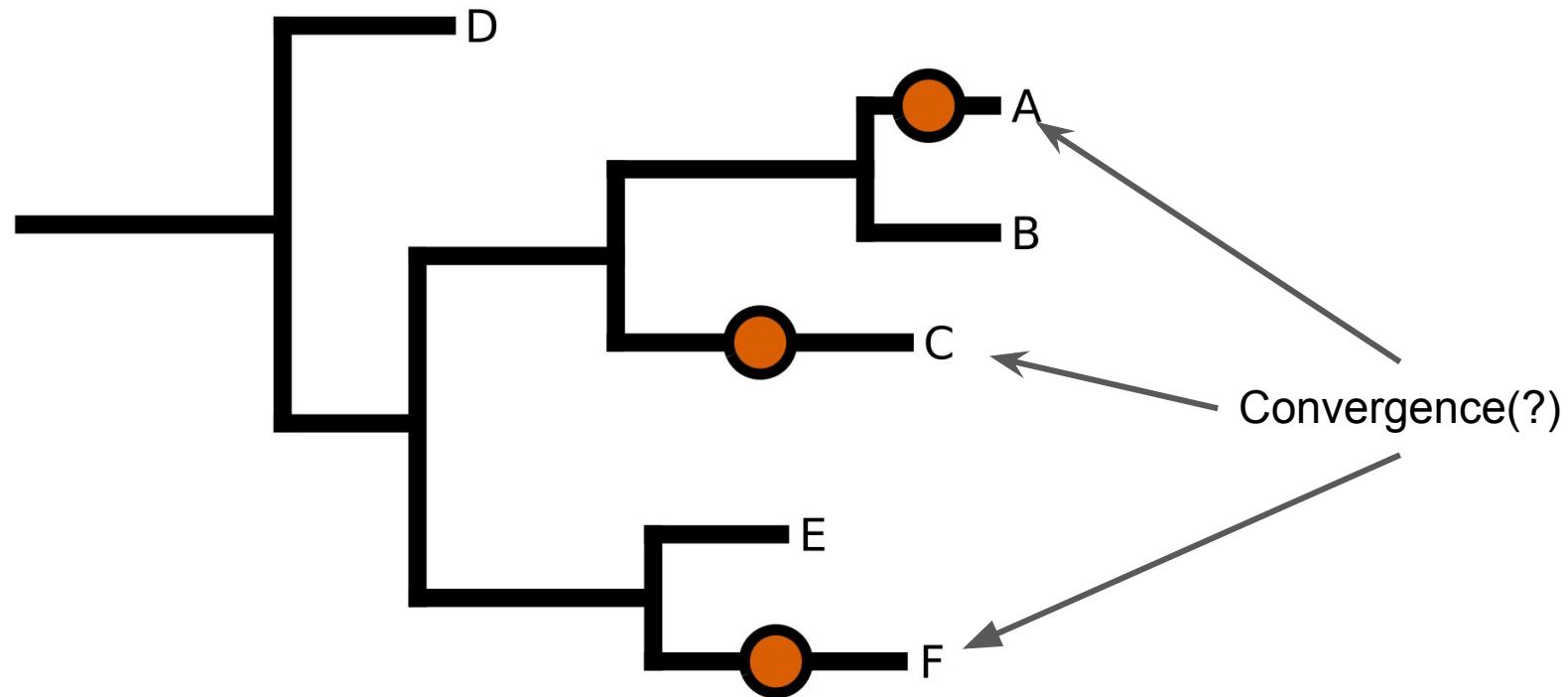
Identify clinically relevant mutations



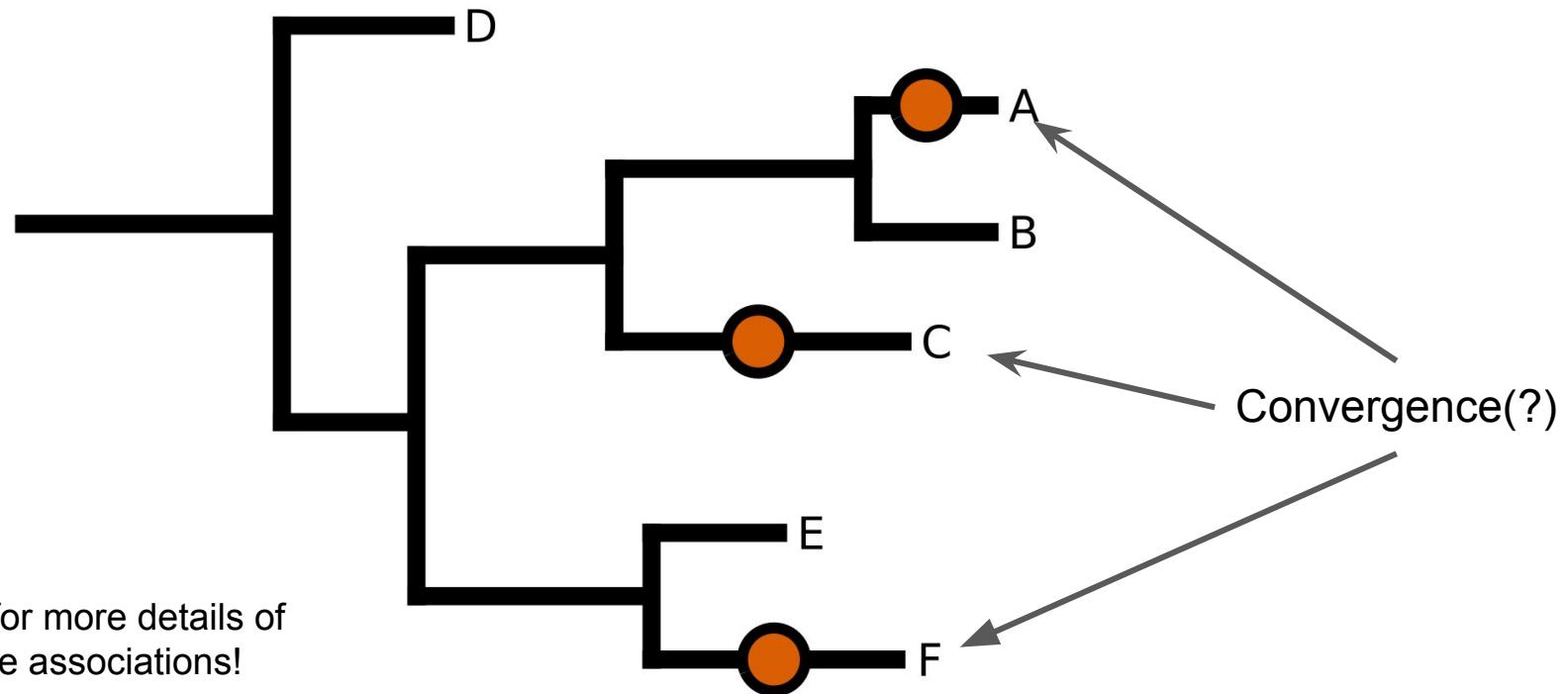
Identify clinically relevant mutations



Identify clinically relevant mutations

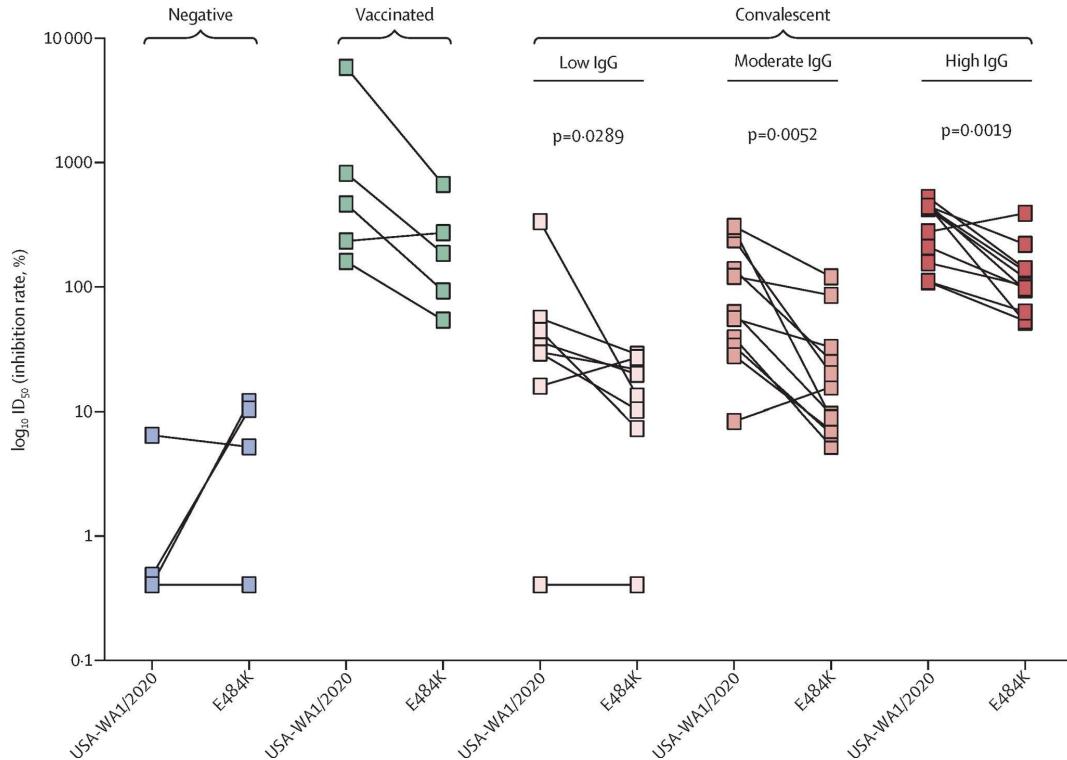


Identify clinically relevant mutations



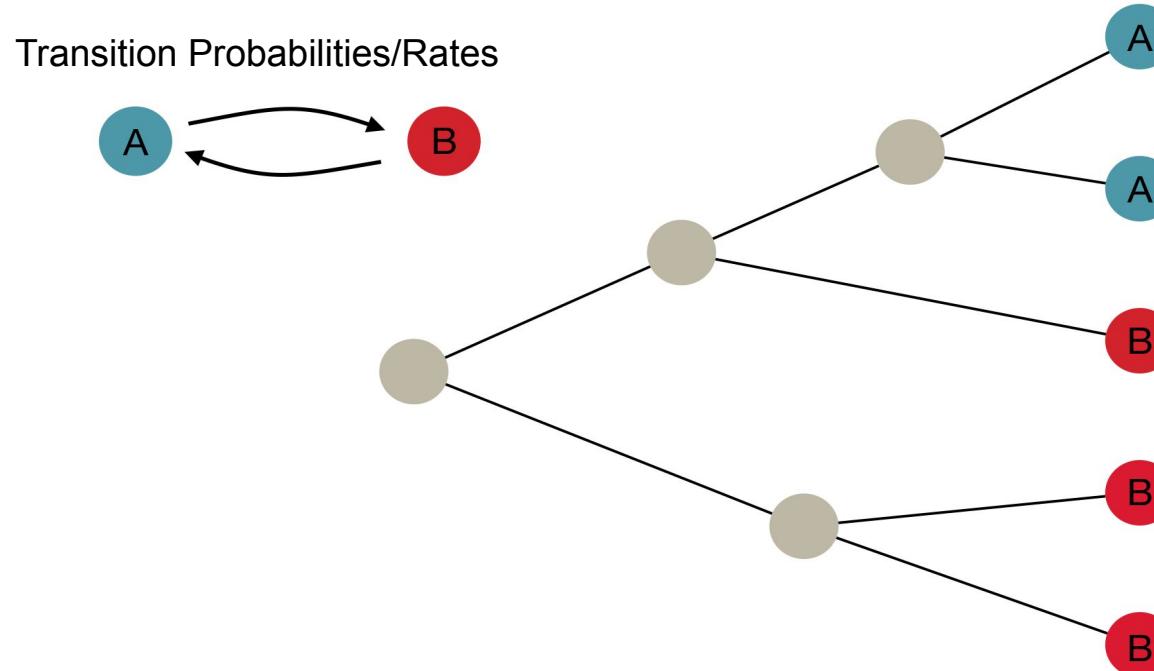
See part 2 for more details of testing these associations!

Prioritise characterisation of mutations: S:E484K

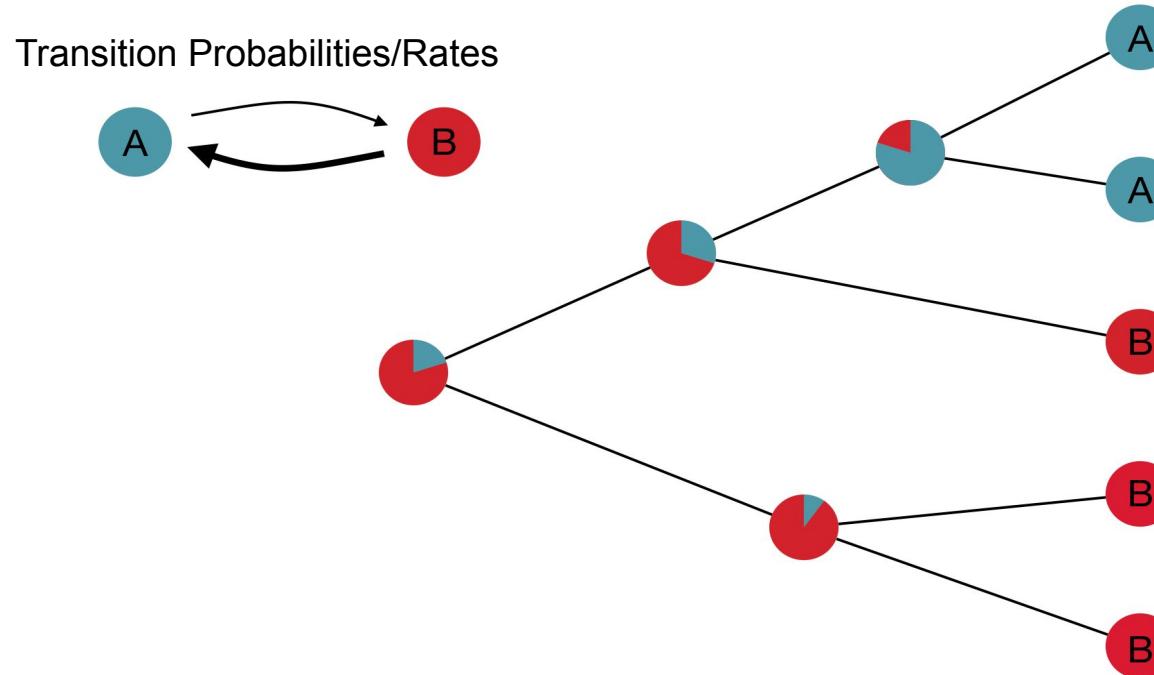


[https://www.thelancet.com/journals/lanmic/article/PIIS2666-5247\(21\)00068-9](https://www.thelancet.com/journals/lanmic/article/PIIS2666-5247(21)00068-9)

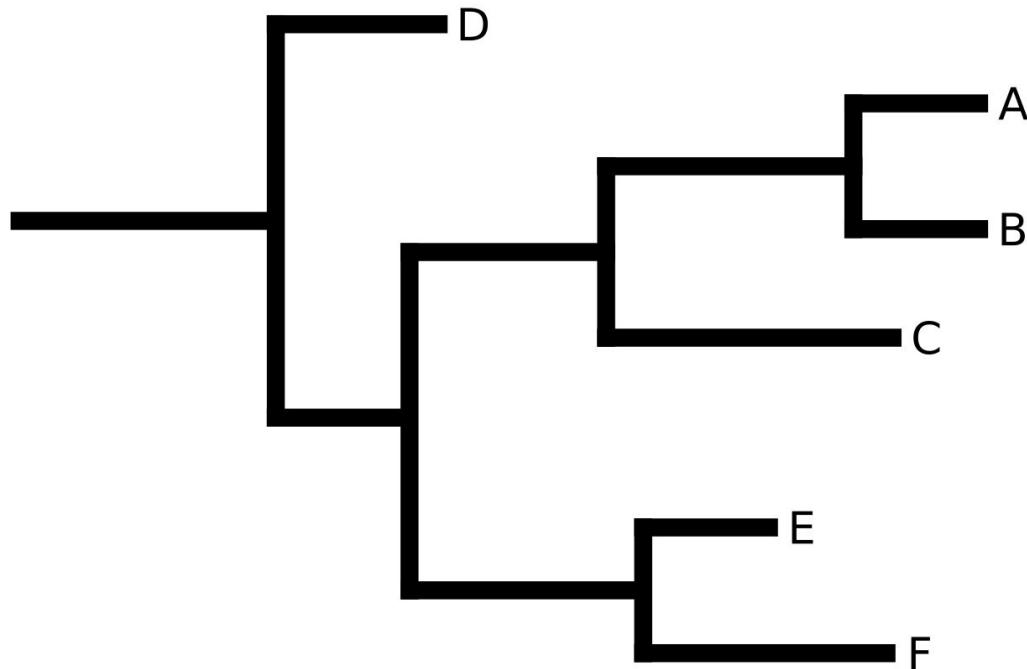
Inferring internal ancestral states from observed tips



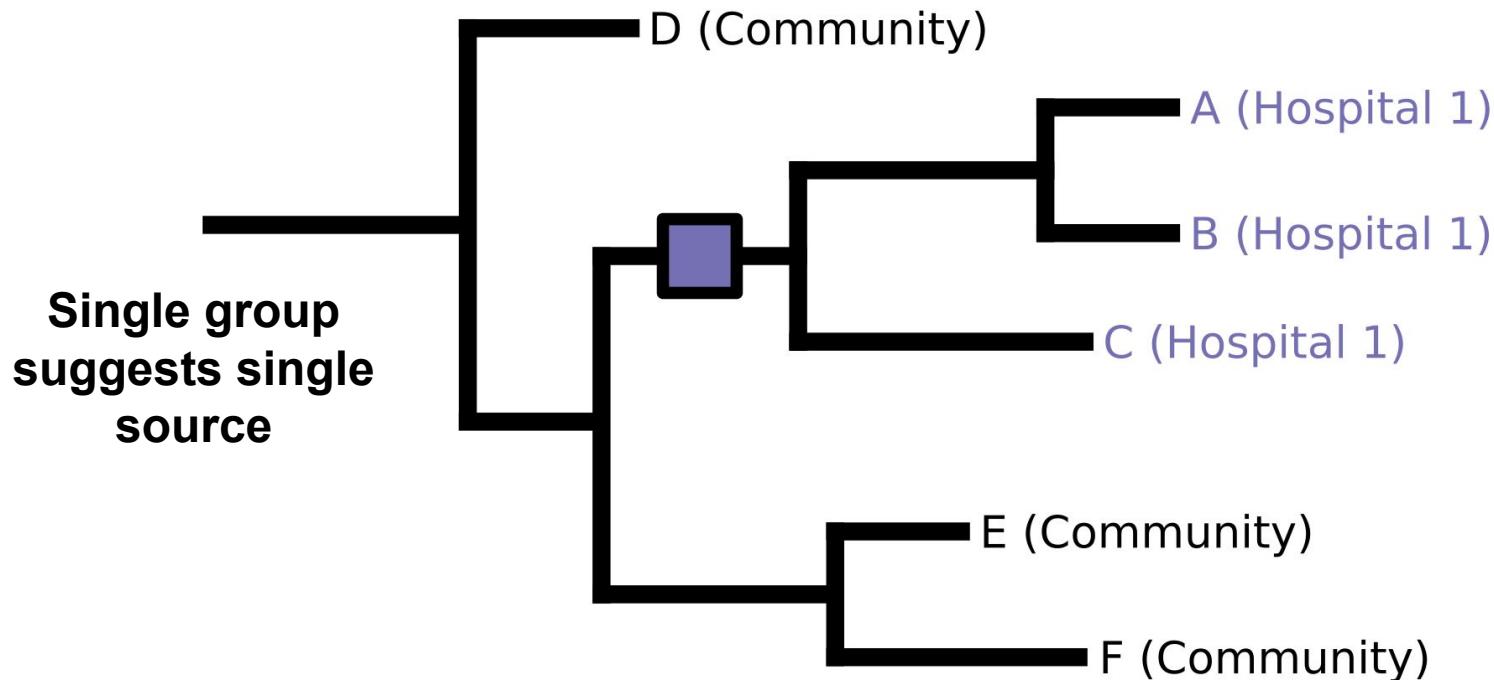
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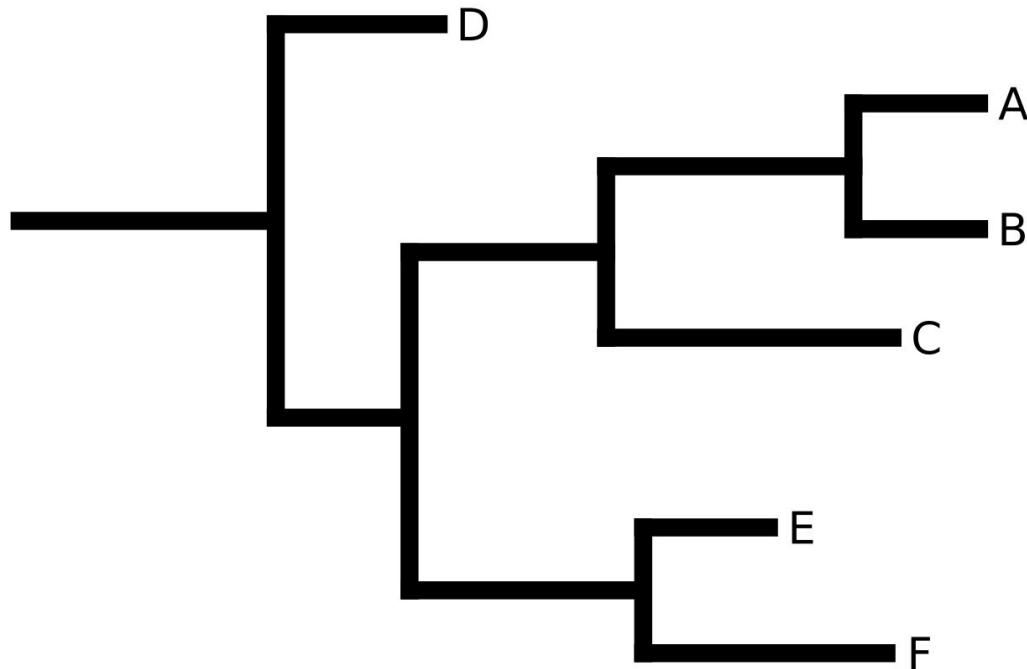
Trace sources of outbreaks



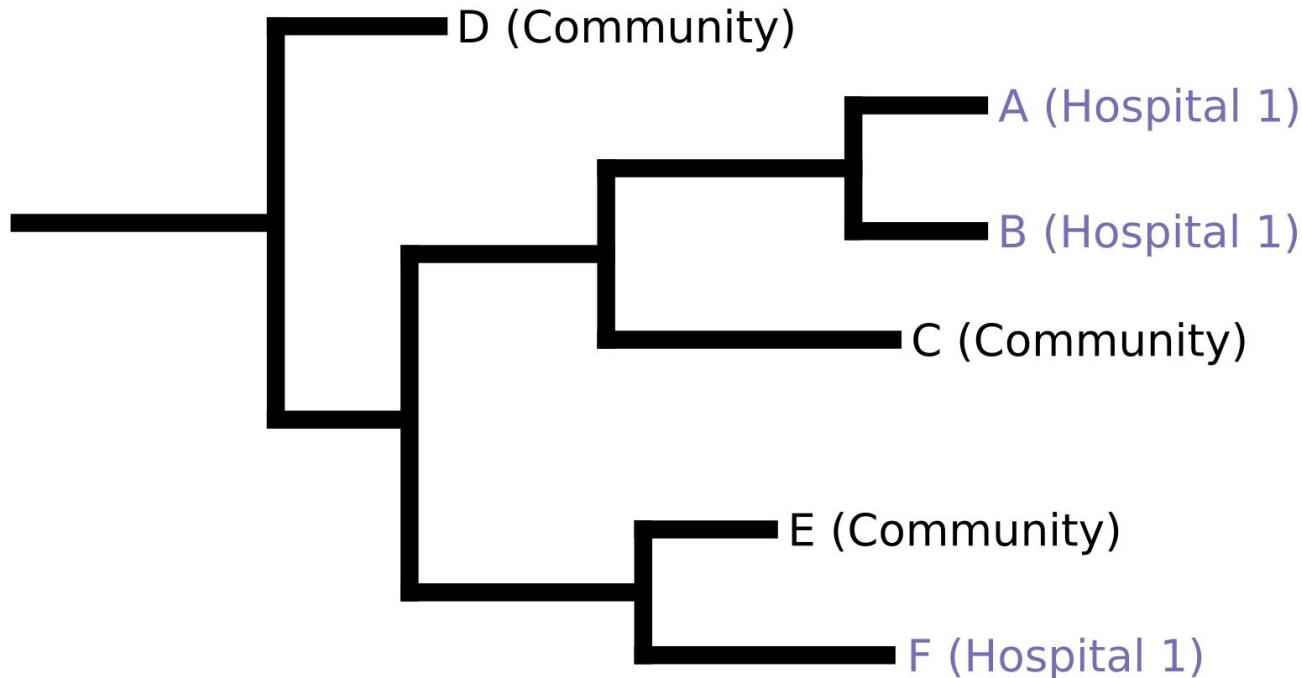
Trace sources of outbreaks



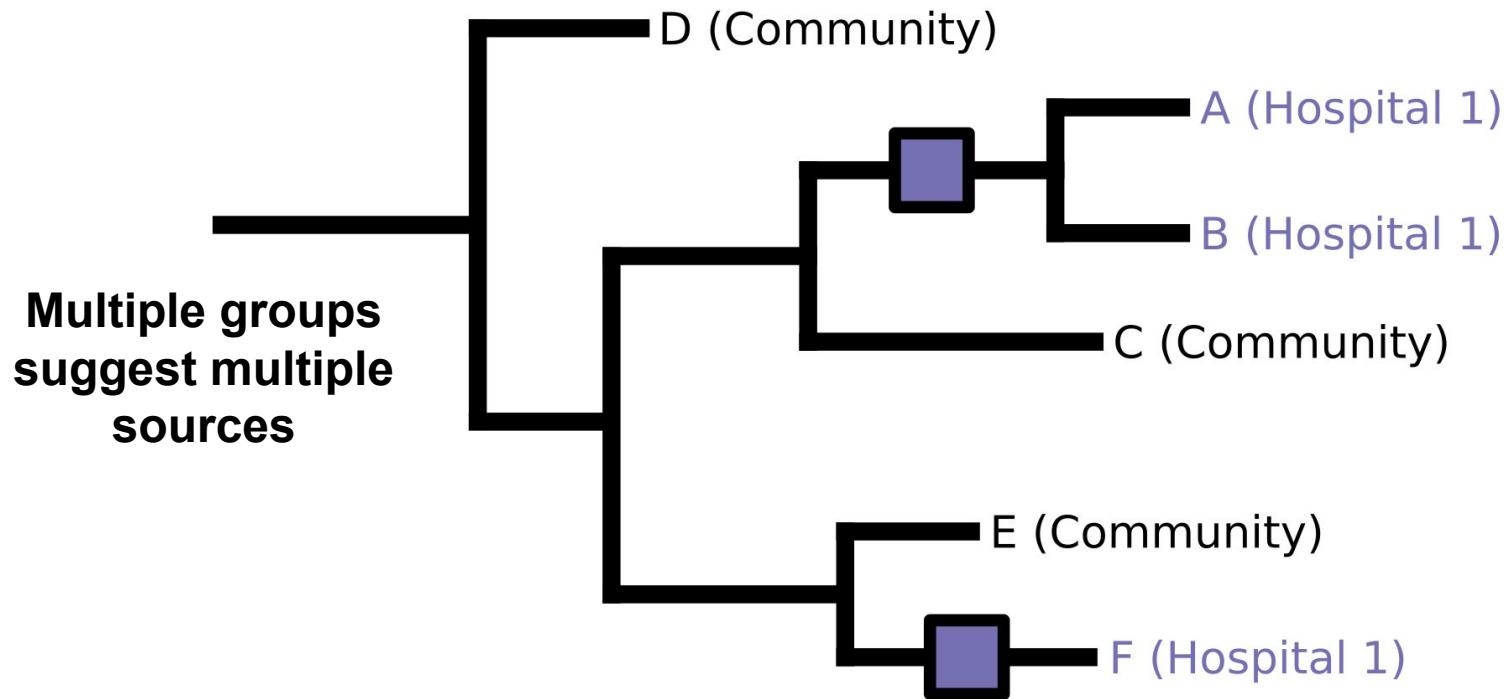
Trace sources of outbreaks



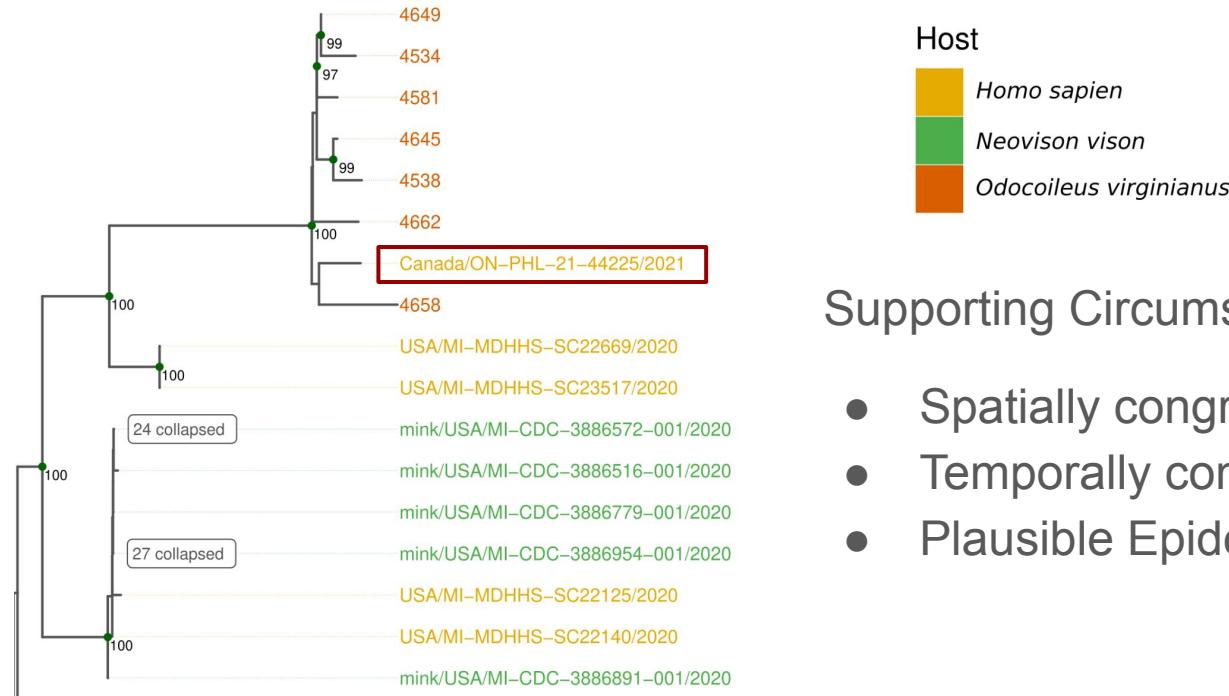
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Trace sources of outbreaks



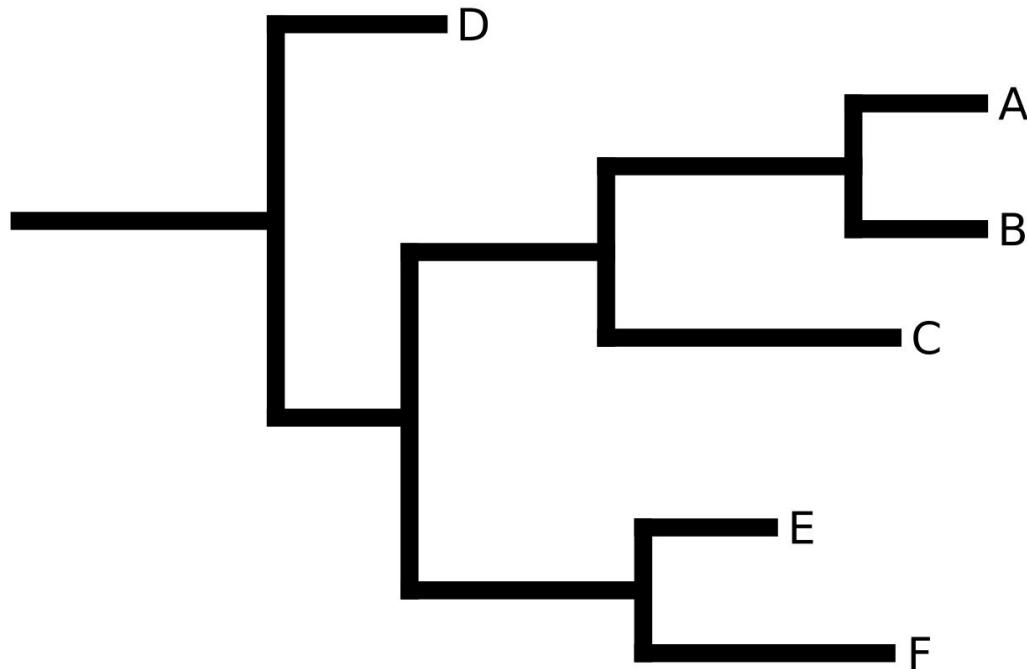
Finding Deer-to-Human transmission



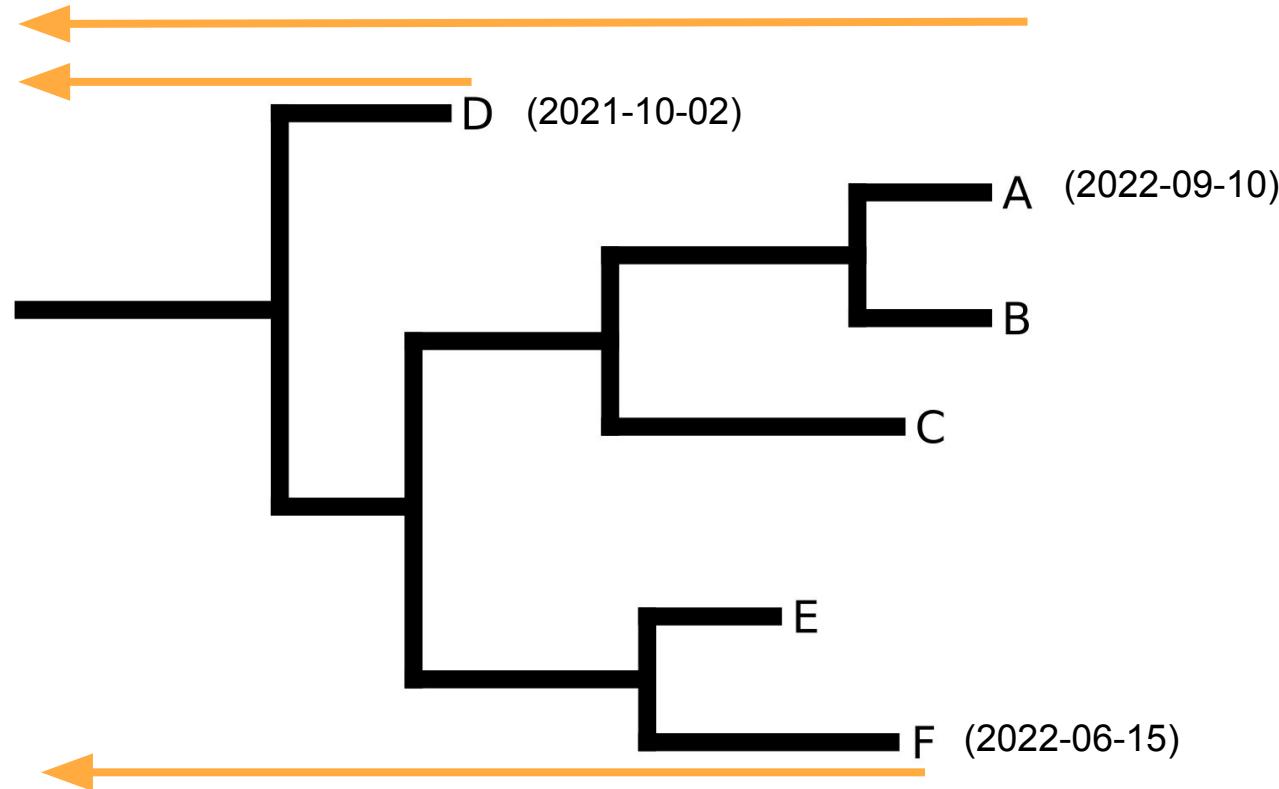
Supporting Circumstantial Evidence:

- Spatially congruent
- Temporally congruent
- Plausible Epidemiological Link

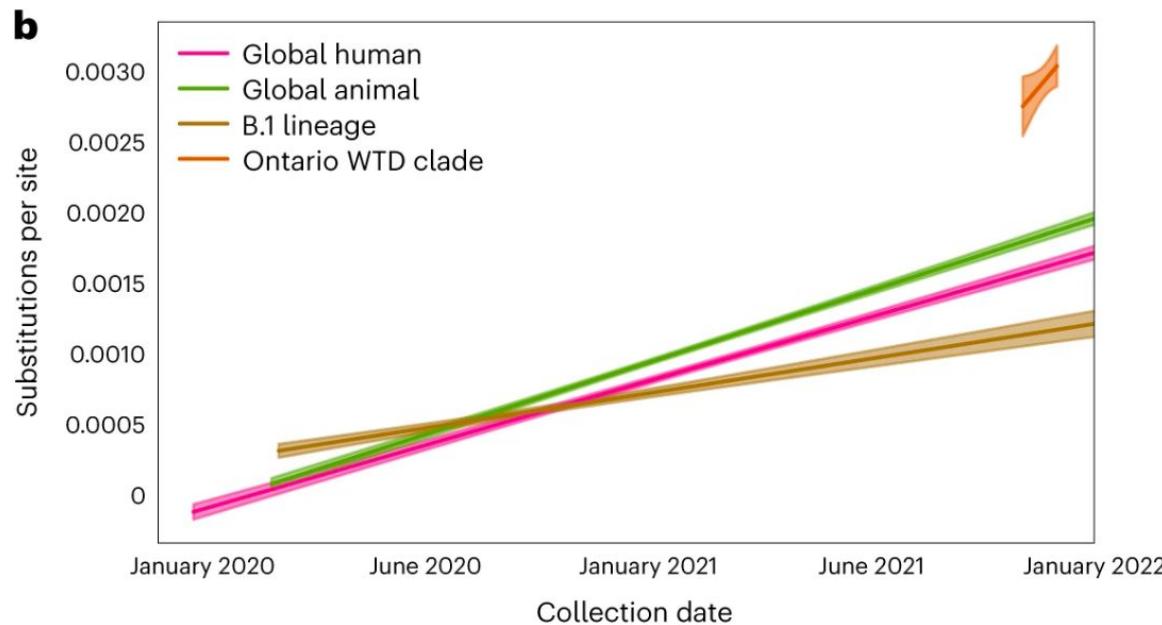
Convert genomic distance to time



Convert genomic distance to time

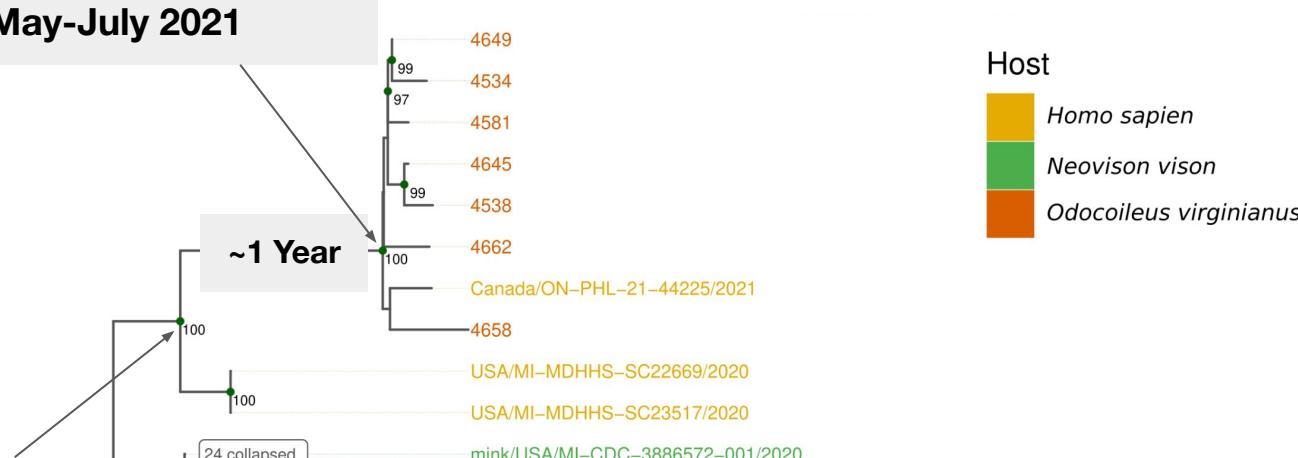


Mutation rates from Root to Tip Regression



Mutations rates let us time of unobserved events

May-July 2021



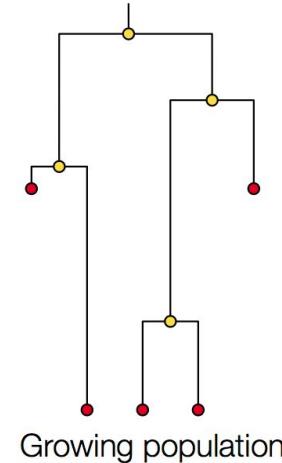
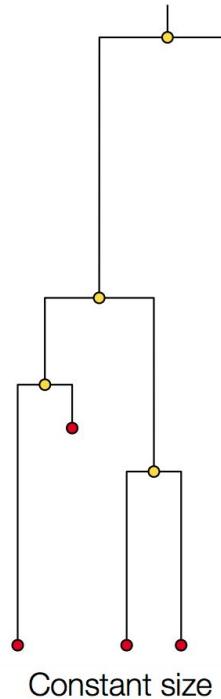
May-Aug 2020



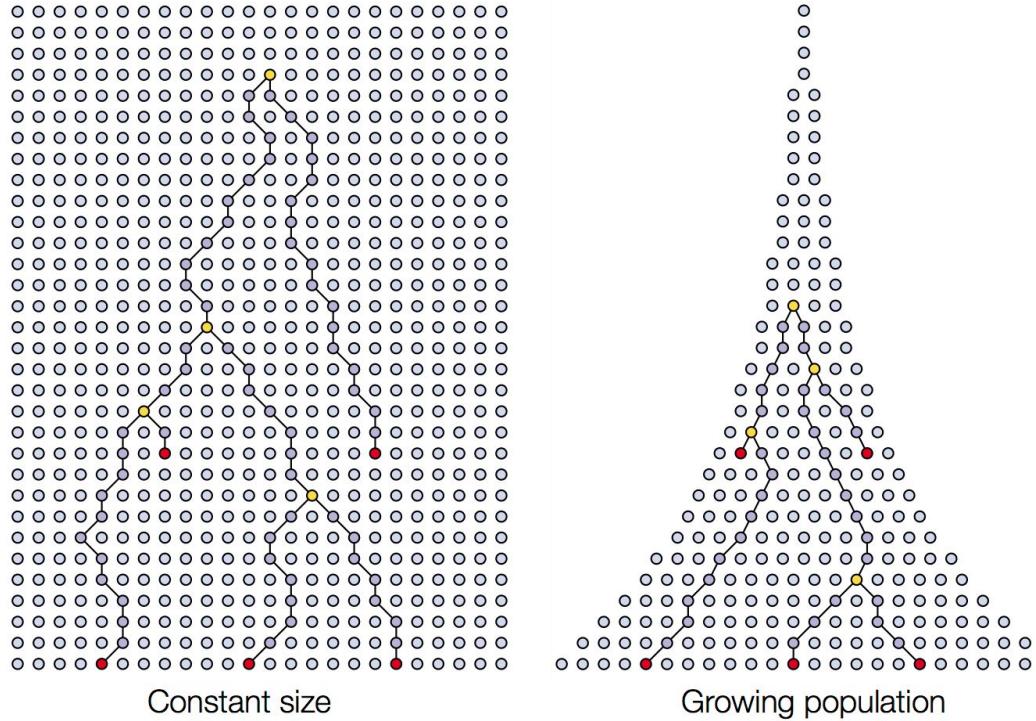
Host

- Homo sapien*
- Neovison vison*
- Odocoileus virginianus*

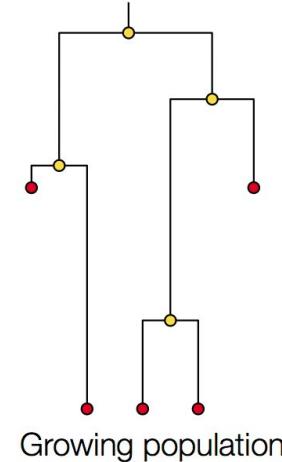
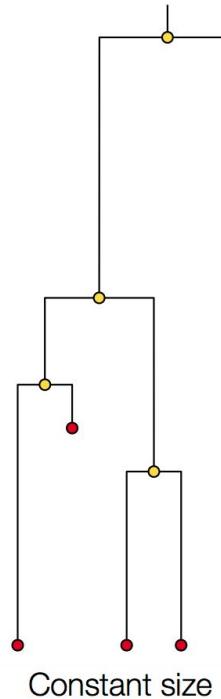
Tree shape tells us about population size



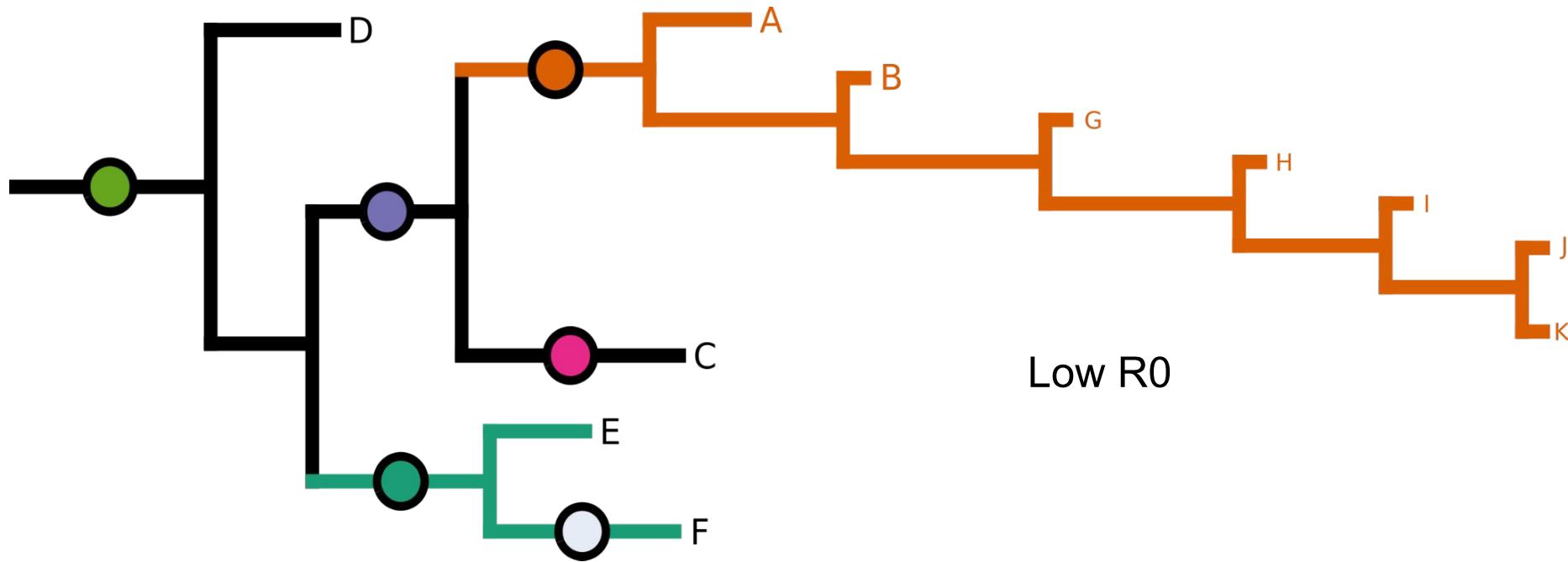
Tree shape tells us about population size



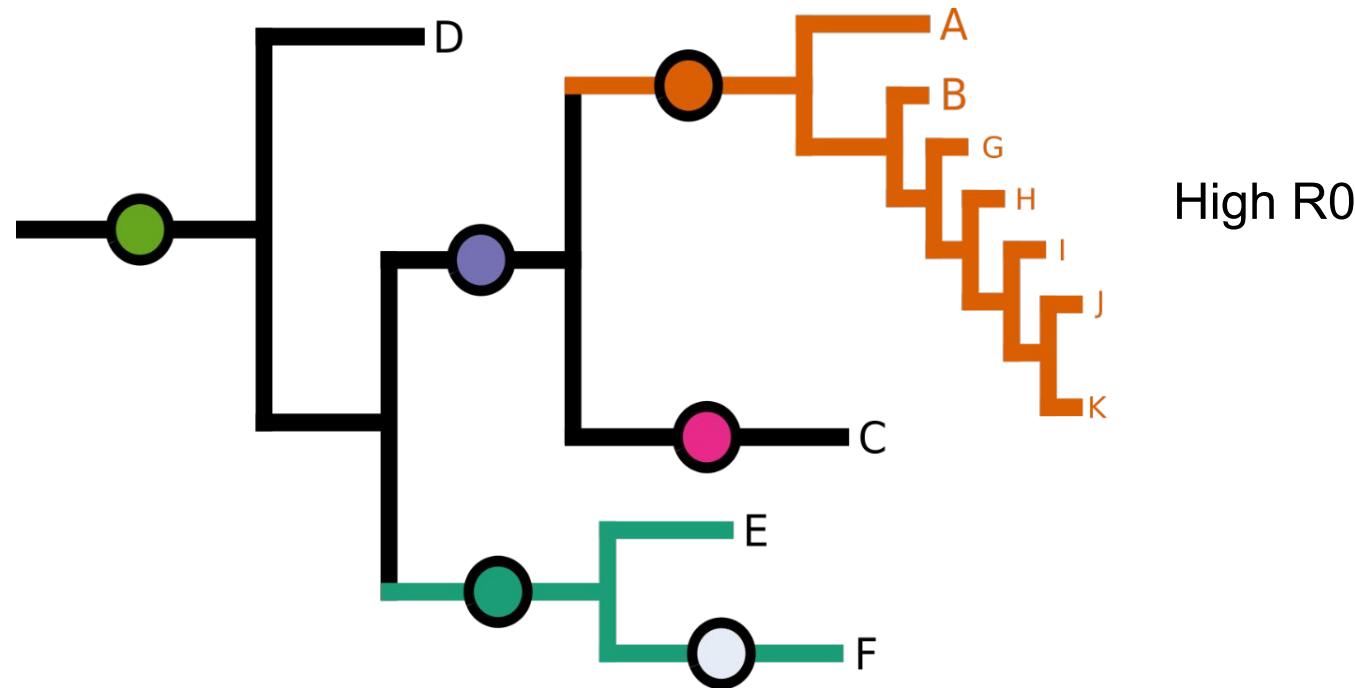
Tree shape tells us about population size



Inferring epidemiological parameters from shape



Inferring epidemiological parameters from shape



What about evolutionary forces like selection?

dN/dS is one way to detect selection

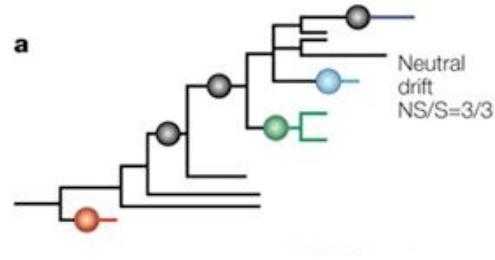
dN = non-synonymous mutations (normalised)

dS = synonymous mutations (normalised)

dN/dS is one way to detect selection

dN = non-synonymous mutations (normalised)

dS = synonymous mutations (normalised)



$dN/dS \sim 1$: drift/neutral selection

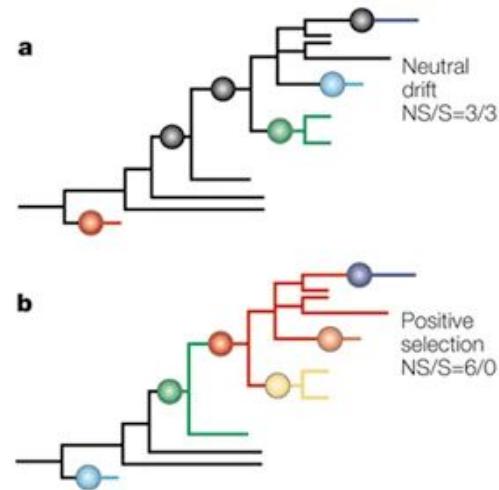
dN/dS is one way to detect selection

dN = non-synonymous mutations (normalised)

dS = synonymous mutations (normalised)

$dN/dS > 1$: adaptive/positive selection

$dN/dS \sim 1$: drift/neutral selection



dN/dS is one way to detect selection

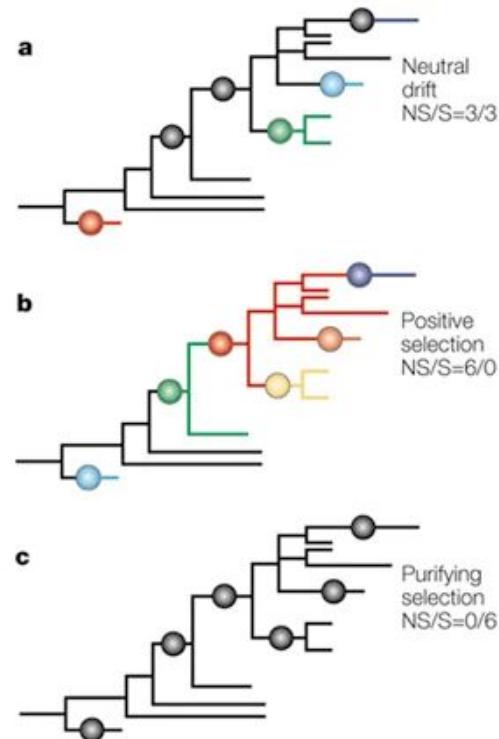
dN = non-synonymous mutations (normalised)

dS = synonymous mutations (normalised)

$dN/dS > 1$: adaptive/positive selection

$dN/dS \sim 1$: drift/neutral selection

$dN/dS < 1$: purifying/negative selection



dN/dS is one way to detect selection

dN = non-synonymous mutations (normalised)

dS = synonymous mutations (normalised)

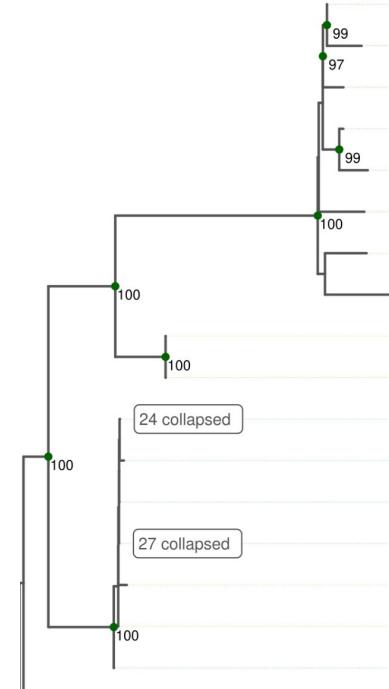
$dN/dS > 1$: adaptive/positive selection

$dN/dS \sim 1$: drift/neutral selection

$dN/dS < 1$: purifying/negative selection

Challenges:

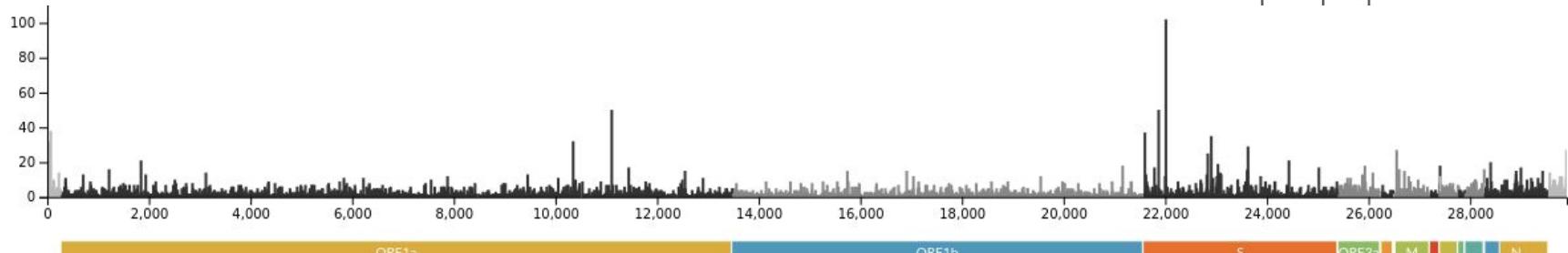
- Mutation rates vary over time/groups



dN/dS is one way to detect selection

dN = non-synonymous mutations (normalised)

dS = synonymous mutations (normalised)



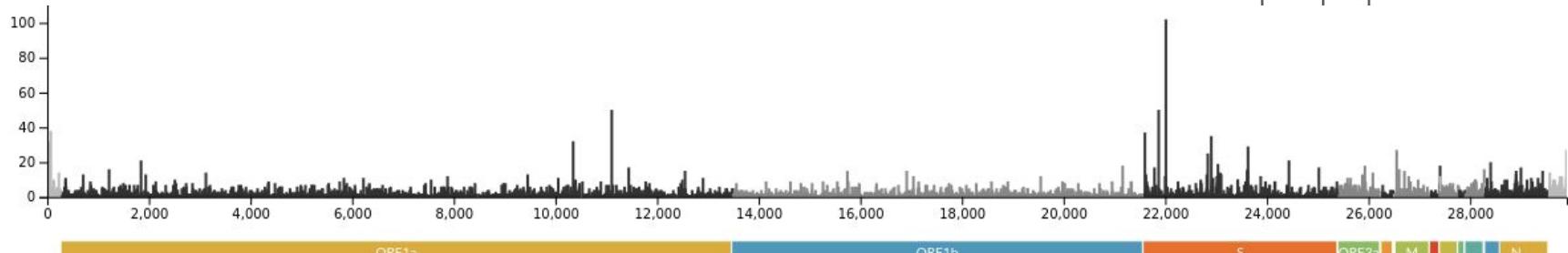
Challenges:

- Mutation rates vary over time/groups
- Mutation rates vary across genomes

dN/dS is one way to detect selection

dN = non-synonymous mutations (normalised)

dS = synonymous mutations (normalised)



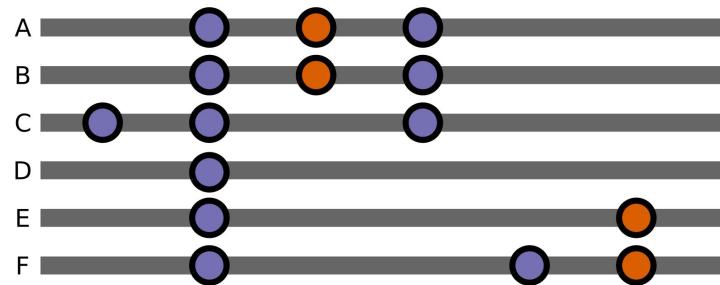
Challenges:

- Mutation rates vary over time/groups
- Mutation rates vary across genomes
- **Genomes are related** (mutations are non-independent)

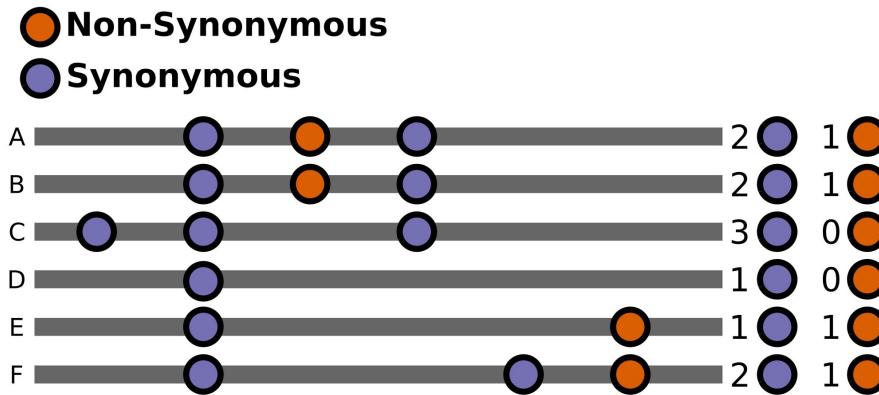
Non-independence of events in related genomes

● Non-Synonymous

● Synonymous



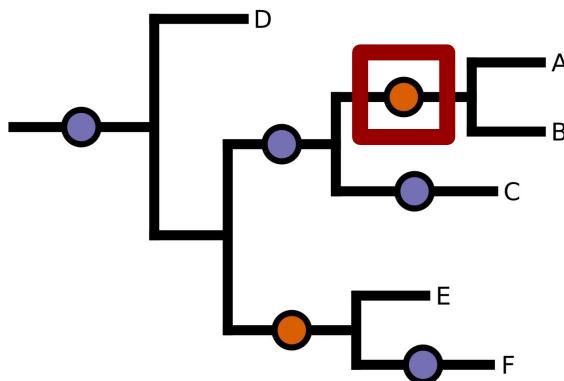
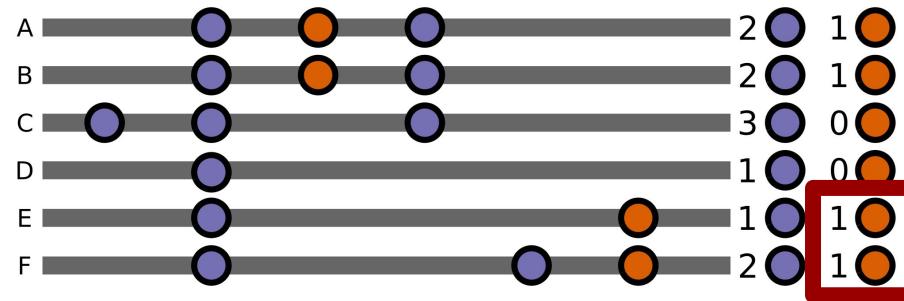
Non-independence of events in related genomes



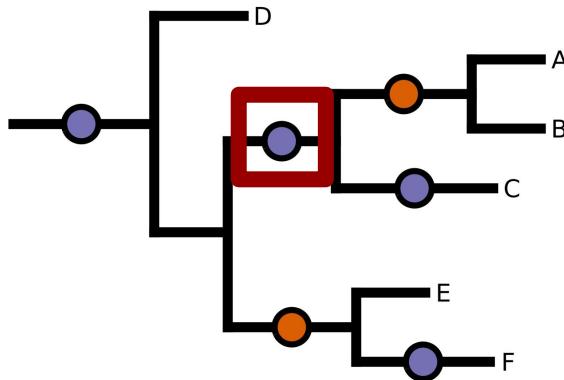
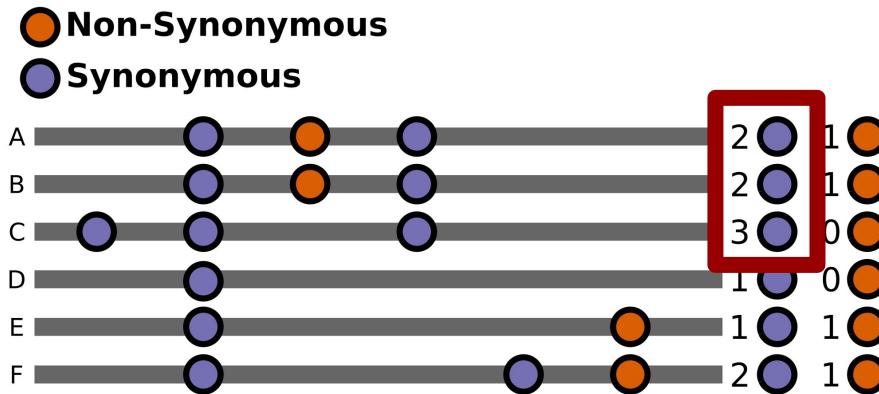
Non-independence of events in related genomes

● Non-Synonymous

● Synonymous



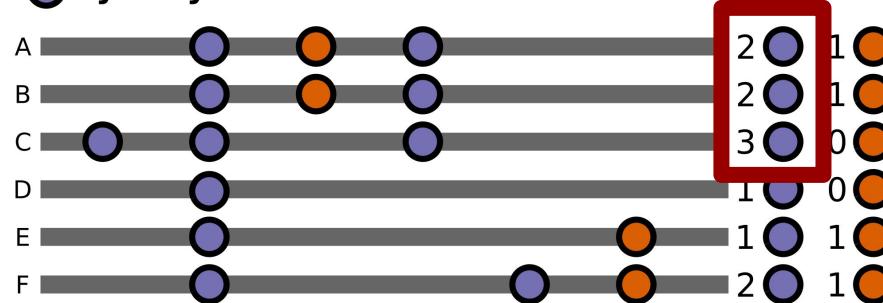
Non-independence of events in related genomes



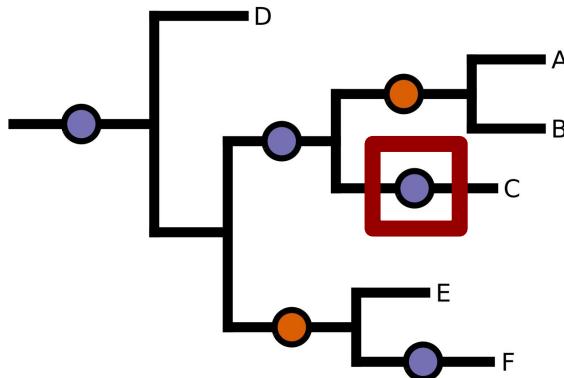
Non-independence of events in related genomes

● Non-Synonymous

● Synonymous



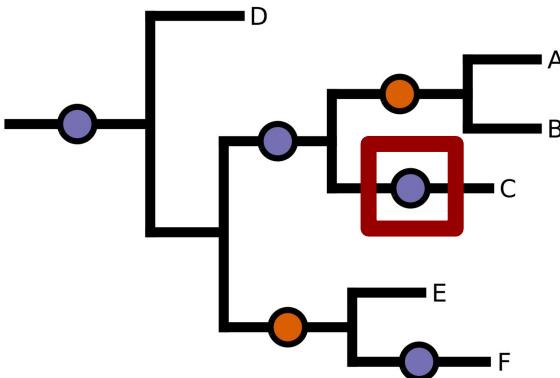
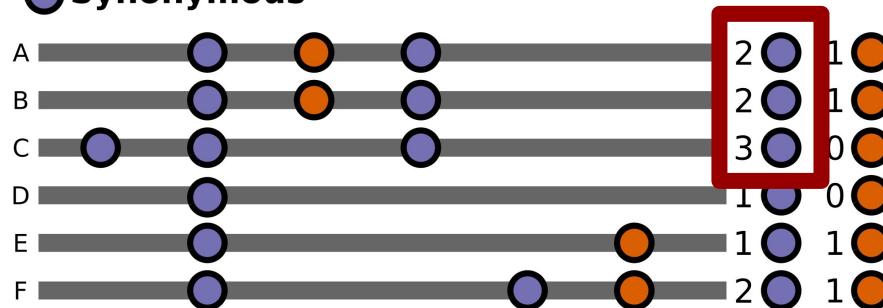
- Phylogeny captures dependency structure of genomic data



Non-independence of events in related genomes

● Non-Synonymous

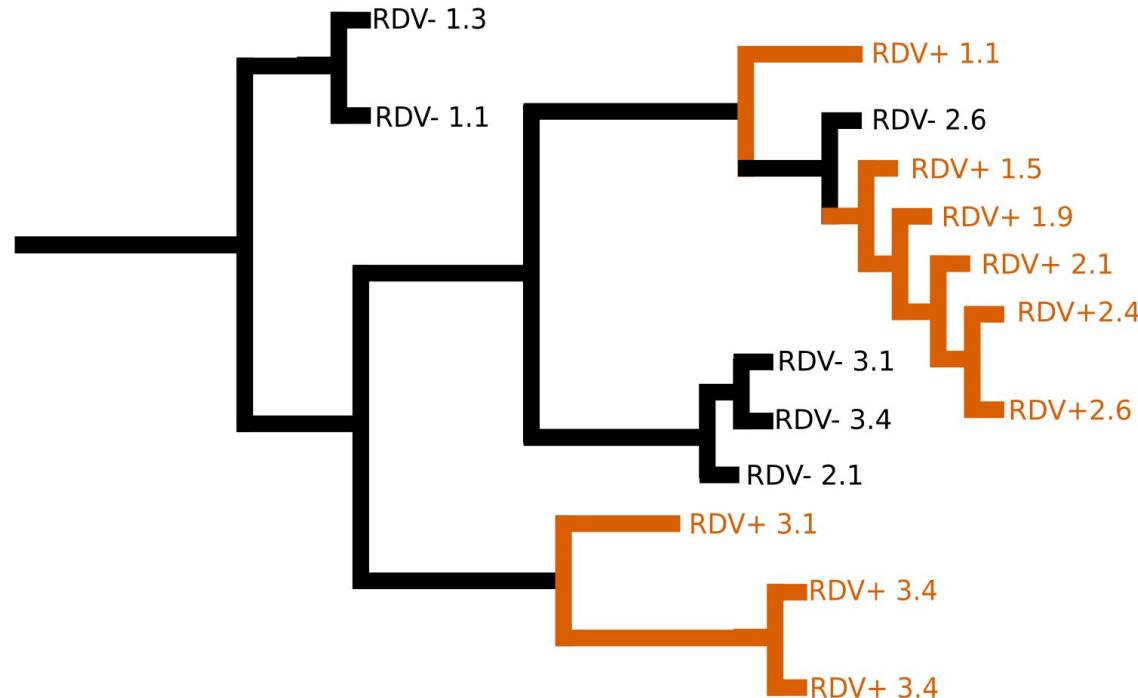
● Synonymous



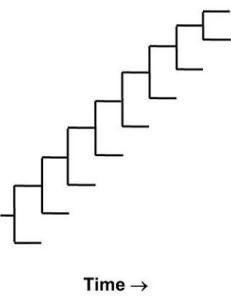
- Phylogeny captures dependency structure of genomic data
- Informs error term for models (e.g., regression)
- adaptive Branch-Site Random Effects Likelihood: Is there a significant proportion of sites within selected branches with $dN/dS > 1$

Smith, MD et al. "Less is more: an adaptive branch-site random effects model for efficient detection of episodic diversifying selection." Mol. Biol. Evol. 32, 1342–1353 (2015).

Testing for remdesevir resistance selection



Many other analyses are possible

	Continual Immune Selection	Weak or Absent Immune Selection	
Idealized Phylogeny Shapes		<p>Tree shape controlled by non-selective population dynamic processes</p>	
Examples	Human influenza A virus intra-host HIV	inter-host HIV inter-host HCV	Measles, rabies inter-host HIV
Tree Inferences	Detection of antigenic escape mutations	Estimation of population growth rates	Estimation of population migration rates

Massive area:

- Birth-death models
- Coalescent models
- Bayesian skyline/skygrid models
- Spatiotemporal models (phylogeography)
- Recombination

Summary

- Pathogen **evolution** and **epidemiology** are intrinsically linked
- Genomics provides insights into **evolution and unobserved events**
- Comparison of DNA sequences to databases can be used for **diagnostics**
- Pattern of mutations across genomes can be used to generate **phylogenies**
- Phylogenies are structured by **sampling, ecology, evolution, and epidemiology**
- Probabilistic **Bayesian phylogenetic inference** is a key tool
- Can use these approaches to do many things including:
 - Identify **lineages**
 - Monitor **evolution**
 - Infer **timing/location** of outbreaks/events
 - Determine **epidemiological parameters**
 - Characterise strength and direction of **selection**