

CORONAVIRUS

Evolution and epidemic spread of SARS-CoV-2 in Brazil

Darlan S. Candido^{1,2*}, Ingra M. Claro^{2,3*}, Jaqueline G. de Jesus^{2,3*}, William M. Souza^{4*}, Filipe R. R. Moreira^{5*}, Simon Dellicour^{6,7*}, Thomas A. Mellan^{8*}, Louis du Plessis¹, Rafael H. M. Pereira⁹, Flavia C. S. Sales^{2,3}, Erika R. Manuli^{2,3}, Julien Thézé¹⁰, Luiz Almeida¹¹, Mariane T. Menezes⁵, Carolina M. Voloch⁵, Marcilio J. Fumagalli⁴, Thaís M. Coletti^{2,3}, Camila A. M. da Silva^{2,3}, Mariana S. Ramundo^{2,3}, Mariene R. Amorim¹², Henrique H. Hoeltgebaum¹³, Swapnil Mishra⁸, Mandev S. Gill⁷, Luiz M. Carvalho¹⁴, Lewis F. Buss², Carlos A. Prete Jr.¹⁵, Jordan Ashworth¹⁶, Helder I. Nakaya¹⁷, Pedro S. Peixoto¹⁸, Oliver J. Brady^{19,20}, Samuel M. Nicholls²¹, Amílcar Tanuri⁵, Átila D. Rossi⁵, Carlos K. V. Braga⁹, Alexandra L. Gerber¹¹, Ana Paula de C. Guimarães²¹, Nelson Gaburo Jr.²², Cecília Salete Alencar²³, Alessandro C. S. Ferreira²⁴, Cristiano X. Lima^{25,26}, José Eduardo Levi²⁷, Celso Granato²⁸, Giulia M. Ferreira²⁹, Ronaldo S. Francisco Jr.¹¹, Fabiana Granja^{12,30}, Marcia T. Garcia³¹, Maria Luíza Moretti³¹, Mauricio W. Perroud Jr.³², Terezinha M. P. P. Castilheiras³³, Carolina S. Lazari³⁴, Sarah C. Hill¹³⁵, Andreza Aruska de Souza Santos³⁶, Camila L. Simeoni¹², Julia Forato¹², Andrei C. Sposito³⁷, Angelica Z. Schreiber³⁸, Magnus N. N. Santos³⁸, Camila Zolini de Sá³⁹, Renan P. Souza³⁹, Luciana C. Resende-Moreira⁴⁰, Mauro M. Teixeira⁴¹, Josy Hubner⁴², Patricia A. F. Leme⁴³, Rennan G. Moreira⁴⁴, Maurício L. Nogueira⁴⁵, Brazil-UK Centre for Arbovirus Discovery, Diagnosis, Genomics and Epidemiology (CADDE) Genomic Network, Neil M. Ferguson⁸, Silvia F. Costa^{2,3}, José Luiz Proença-Modena¹², Ana Tereza R. Vasconcelos¹¹, Samir Bhatt⁸, Philippe Lemey⁷, Chieh-Hsi Wu⁴⁶, Andrew Rambaut⁴⁷, Nick J. Loman²¹, Renato S. Aguiar³⁹, Oliver G. Pybus⁸, Ester C. Sabino^{2,3†}, Nuno Rodrigues Faria^{1,2,8†}

Brazil currently has one of the fastest-growing severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) epidemics in the world. Because of limited available data, assessments of the impact of nonpharmaceutical interventions (NPIs) on this virus spread remain challenging. Using a mobility-driven transmission model, we show that NPIs reduced the reproduction number from >3 to 1 to 1.6 in São Paulo and Rio de Janeiro. Sequencing of 427 new genomes and analysis of a geographically representative genomic dataset identified >100 international virus introductions in Brazil. We estimate that most (76%) of the Brazilian strains fell in three clades that were introduced from Europe between 22 February and 11 March 2020. During the early epidemic phase, we found that SARS-CoV-2 spread mostly locally and within state borders. After this period, despite sharp decreases in air travel, we estimated multiple exportations from large urban centers that coincided with a 25% increase in average traveled distances in national flights. This study sheds new light on the epidemic transmission and evolutionary trajectories of SARS-CoV-2 lineages in Brazil and provides evidence that current interventions remain insufficient to keep virus transmission under control in this country.

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is a novel beta-coronavirus with a 30-kb genome that was first reported in December 2019 in Wuhan, China (1, 2). SARS-CoV-2 was declared a public health emergency of international concern on 30 January 2020. As of 12 July 2020, there were >12.5 million cases of coronavirus disease 2019 (COVID-19) and 561,000 deaths globally (3). The virus can be classified into two main phylogenetic lineages, A and B, which spread from Wuhan before strict travel restrictions were enacted (4, 5) and now cocirculate around the world (6). The case fatality ratio of SARS-CoV-2 infection has been estimated at between 1.2 and 1.6% (7–9), with substantially higher ratios in those >60 years of age (8). Some estimates suggest that 18 to 56% of SARS-CoV-2 transmission is from asymptomatic or presymptomatic individuals (10–13), complicating epidemiological assessments and public health efforts to curb the pandemic.

Challenges of real-time assessment of transmission

Although the SARS-CoV-2 epidemics in several countries, including China, Italy, and Spain, have been brought under control through nonpharmaceutical interventions (NPIs) (3), the number of SARS-CoV-2 cases and deaths in Brazil continues to increase (14) (Fig. 1A). As of 12 July 2020, Brazil had reported 1,800,827 SARS-CoV-2 cases, the second-largest number in the world, and 70,398 deaths. More than one-third of the cases (34%) in Brazil are concentrated in the southeast region, which includes São Paulo city (Fig. 1B), the world's fourth-largest conurbation, where the first case in Latin America was reported on 25 February 2020 (15). Diagnostic assays for SARS-CoV-2 molecular detection were widely distributed across the regional reference centers of the national public health laboratory network from 21 February 2020 on (16, 17). However, several factors, including delays in reporting, changes in notification, and heterogeneous access to testing across populations,

obfuscate the real-time assessment of virus transmission using SARS-CoV-2 case counts (15). Consequently, a more accurate measure of SARS-CoV-2 transmission in Brazil is the number of reported deaths caused by severe acute respiratory infections (SARIs), which is provided by the Sistema Único de Saúde (SUS) (18). Changes in the opportunity for SARS-CoV-2 transmission are strongly associated with changes in average mobility (18–20) and can typically be measured by calculating the effective reproduction number, R , defined as the average number of secondary infections caused by an infected person. $R > 1$ indicates a growing epidemic, whereas $R < 1$ is needed to achieve a decrease in transmission.

We used a Bayesian semimechanistic model (21, 22) to analyze SARI mortality statistics and human mobility data to estimate daily changes in R in São Paulo city (12.2 million inhabitants) and Rio de Janeiro city (6.7 million inhabitants), the largest urban metropolises in Brazil (Fig. 1, C and D). NPIs in Brazil consisted of school closures implemented between 12 and 23 March 2020 across the country's 27 federal units/states and store closures implemented between 13 and 23 March 2020. In São Paulo city, schools started closing on 16 March 2020 and stores closed 4 days later. At the start of the epidemics, we found $R > 3$ in São Paulo and Rio de Janeiro and, concurrent with the timing of state-mandated NPIs, R values fell close to 1.

Mobility-driven changes in R

Analysis of R values after NPI implementation highlights several notable mobility-driven features. There was a period immediately after NPIs, between 21 and 31 March 2020, when R was consistently <1 in São Paulo city (Fig. 1C). However, after this initial decrease, the R value for São Paulo rose to >1 and increased through time, a trend associated with increased population mobility. This can be seen in the Google transit stations index, which rose from -60 to -52% , and by a decrease in the social isolation index from 54 to 47%. By 4 May 2020, we estimate $R = 1.3$ [95% Bayesian credible interval (BCI): 1.0 to 1.6] in both São Paulo and Rio de Janeiro cities (table S1). However, we note that there were instances in the previous 7 days when the 95% credible intervals for R included values <1 , drawing attention to the fluctuations and uncertainty in the estimated R for both cities.

Early sharing of genomic sequences, including the first SARS-CoV-2 genome, Wuhan-Hu-1, released on 10 January (23), has enabled unprecedented global levels of molecular testing for an emerging virus (24, 25). However, despite the thousands of virus genomes deposited on public access databases, there is a lack of consistent sampling structure and there are limited data from Brazil (26–28), which

hampers accurate reconstructions of virus movement and transmission using phylogenetic analyses. To investigate how SARS-CoV-2 became established in the country, and to quantify the impact of NPIs on virus

spatiotemporal spread, we tested a total of 26,732 samples from public and private laboratories using real-time quantitative polymerase chain reaction (RT-qPCR) assays and found 7944 (29%) to be positive for SARS-

CoV-2. We then focused our sequencing efforts on generating a large and spatially representative genomic dataset with curated metadata to maximize the association between the number of sequences and the

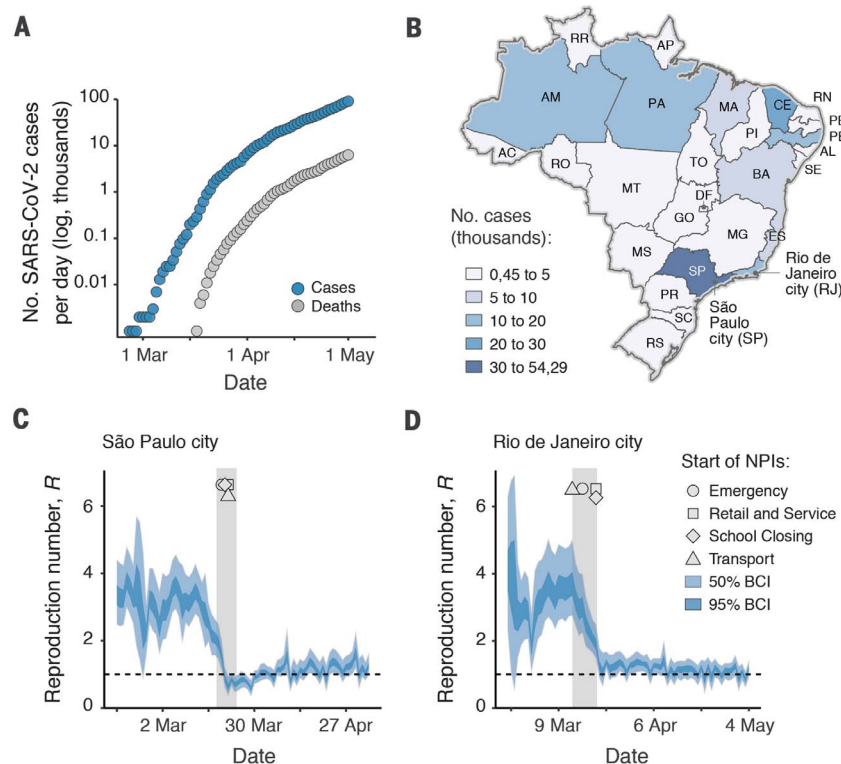


Fig. 1. SARS-CoV-2 epidemiology and epidemic spread in Brazil.

(A) Cumulative number of SARS-CoV-2 reported cases (blue) and deaths (gray) in Brazil. (B) States are colored according to the number of cumulative confirmed cases by 30 April 2020. (C and D) R over time for the cities of São Paulo (C) and Rio de Janeiro (D). R values were estimated using a Bayesian approach incorporating the daily number of deaths and four variables related to mobility data (a social isolation index from Brazilian

geolocation company *InLoco* and Google mobility indices for time spent in transit stations, parks, and the average between groceries and pharmacies, retail and recreational, and workspaces). Dashed horizontal line indicates $R = 1$. Gray area and geometric symbols show the times at which NPIs were implemented. BCI of 50 and 95% are shown as shaded areas. The two-letter ISO 3166-1 codes for the 27 federal units in Brazil are provided in the supplementary materials.

¹Department of Zoology, University of Oxford, Oxford, UK. ²Instituto de Medicina Tropical, Faculdade de Medicina da Universidade de São Paulo, São Paulo, Brazil. ³Departamento de Moléstias Infecciosas e Parasitárias, Faculdade de Medicina da Universidade de São Paulo, São Paulo, Brazil. ⁴Centro de Pesquisa em Virologia, Faculdade de Medicina de Ribeirão Preto, Ribeirão Preto, Brazil. ⁵Departamento de Genética, Instituto de Biologia, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil. ⁶Spatial Epidemiology Lab, Université Libre de Bruxelles, Brussels, Belgium. ⁷Department of Microbiology, Immunology and Transplantation, Rega Institute, KU Leuven, Leuven, Belgium. ⁸MRC Centre for Global Infectious Disease Analysis, J-IDEA, Imperial College London, London, UK. ⁹Institute for Applied Economic Research, Brasília, Brazil. ¹⁰Université Clermont Auvergne, INRAE, VetAgro Sup, UMR EPIA, Saint-Genès-Champagnelle, France. ¹¹Laboratório de Bioinformática, Laboratório Nacional de Computação Científica, Petrópolis, Brazil. ¹²Departamento de Genética, Evolução, Microbiologia e Imunologia, Instituto de Biologia, Universidade Estadual de Campinas, Campinas, Brazil. ¹³Department of Mathematics, Imperial College London, London, UK. ¹⁴Escola de Matemática Aplicada (EMAp), Fundação Getúlio Vargas, Rio de Janeiro, Brazil. ¹⁵Department of Electronic Systems Engineering, University of São Paulo, São Paulo, Brazil. ¹⁶Usher Institute, University of Edinburgh, Edinburgh, UK. ¹⁷Department of Clinical and Toxicological Analyses, School of Pharmaceutical Sciences, University of São Paulo, São Paulo, Brazil. ¹⁸Departamento de Matemática Aplicada, Instituto de Matemática e Estatística, Universidade de São Paulo, São Paulo, Brazil. ¹⁹Department of Infectious Disease Epidemiology, Faculty of Epidemiology and Population Health, London School of Hygiene & Tropical Medicine, London, UK. ²⁰Centre for the Mathematical Modelling of Infectious Diseases, London School of Hygiene & Tropical Medicine, London, UK. ²¹Institute for Microbiology and Infection, University of Birmingham, Birmingham, UK. ²²DB Diagnósticos do Brasil, São Paulo, Brazil. ²³LIM 03 Laboratório de Medicina Laboratorial, Hospital das Clínicas Faculdade de Medicina da Universidade de São Paulo, São Paulo, Brazil. ²⁴Instituto Hermes Pardini, Belo Horizonte, Brazil. ²⁵Departamento de Cirurgia, Faculdade de Medicina, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil. ²⁶Simile Instituto de Imunologia Aplicada Ltda, Belo Horizonte, Brazil. ²⁷Laboratório DASA, São Paulo, Brazil. ²⁸Laboratório Fleury, São Paulo, Brazil. ²⁹Laboratório de Virologia, Instituto de Ciências Biomédicas, Universidade Federal de Uberlândia, Uberlândia, Brazil. ³⁰Centro de Estudos da Biodiversidade, Universidade Federal de Roraima, Boa Vista, Brazil. ³¹Divisão de Doenças Infecciosas, Faculdade de Ciências Médicas, Universidade Estadual de Campinas, Campinas, Brazil. ³²Hospital Estadual Sumaré, Universidade Estadual de Campinas, Campinas, Brazil. ³³Departamento de Doenças Infecciosas e Parasitárias, Faculdade de Medicina, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil. ³⁴Divisão de Laboratório Central do Hospital das Clínicas, da Faculdade de Medicina da Universidade de São Paulo, São Paulo, Brazil. ³⁵Department of Pathobiology and Population Sciences, Royal Veterinary College, Hatfield, UK. ³⁶University of Oxford, Latin American Centre, Oxford School of Global and Area Studies, Oxford, UK. ³⁷Departamento de Clínica Médica, Faculdade de Ciências Médicas, Universidade Estadual de Campinas, Campinas, Brazil. ³⁸Departamento de Patologia Clínica, Faculdade de Ciências Médicas, Universidade Estadual de Campinas, Campinas, Brazil. ³⁹Departamento de Genética, Ecologia e Evolução, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil. ⁴⁰Departamento de Botânica, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil. ⁴¹Departamento de Bioquímica e Imunologia, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil. ⁴²Departamento de Biologia Celular, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil. ⁴³Centro de Saúde da Comunidade, Universidade Estadual de Campinas, Campinas, Brazil. ⁴⁴Centro de Laboratórios Multiusuários, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil. ⁴⁵Laboratório de Pesquisas em Virologia, Faculdade de Medicina de São José do Rio Preto, São José do Rio Preto, São Paulo, Brazil. ⁴⁶Mathematical Sciences, University of Southampton, Southampton, UK. ⁴⁷Institute of Evolutionary Biology, University of Edinburgh, Edinburgh, UK.

*These authors contributed equally to this work.

†Corresponding author. Email: sabinoec@usp.br (E.C.S.); nfaria@ic.ac.uk (N.R.F.)

number of SARS-CoV-2 confirmed cases per state.

Spatially representative sequencing efforts

We generated 427 new SARS-CoV-2 genomes with >75% genome coverage from Brazilian samples collected between 5 March and 30 April 2020 (figs. S1 to S3 and data S1). For each state, the time between the date of the first reported case and the collection date of the first sequence analyzed in that state was only 4.5 days on average (Fig. 2A). For eight federal states, genomes were obtained from samples collected up to 6 days before the first case notifications. The genomes generated here were

collected in 85 municipalities across 18 of 27 federal units spanning all regions in Brazil (Fig. 2A and fig. S2). Sequenced genomes were obtained from samples collected 4 days on average (median, range: 0 to 29 days) after the onset of symptoms and were generated in three laboratories using harmonized sequencing and bioinformatic protocols (table S2). When we include 63 additional available sequences from Brazil deposited in GISAID (29) (see data S1 and S2), we found the dataset to be representative of the spatial heterogeneity of the Brazilian epidemic. Specifically, the number of genomes per state strongly correlated with SARI SARS-CoV-2 confirmed cases and

SARI cases with unknown etiology per state ($n = 490$ sequences from 21 states, Spearman's correlation, $\rho = 0.83$; Fig. 2A). This correlation varied from 0.70 to 0.83 when considering SARI cases and deaths caused by SARS-CoV-2 and SARI cases and deaths from unknown etiology (fig. S4). Most ($n = 485/490$) Brazilian sequences belong to SARS-CoV-2 lineage B, with only five strains belonging to lineage A (two from Amazonas, one from Rio Grande do Sul, one from Minas Gerais, and one from Rio de Janeiro; data S1 and fig. S5 show detailed lineage information for each sequence). Moreover, we used an *in silico* assessment of diagnostic assay specificity for Brazilian strains ($n = 490$) to identify potential mismatches in some assays targeting these strains. We found that the forward primers of the Chinese CDC and Hong Kong University nucleoprotein-targeting RT-qPCR may be less appropriate for use in Brazil than other diagnostic assays, for which few or no mismatches were identified (fig. S6 and table S3). The impact of these mismatches on the sensitivity of these assays should be confirmed experimentally. If sensitivity is affected, then the use of duplex RT-qPCR assays that concurrently target different genomic regions may help in the detection of viruses with variants in primer- or probe-binding regions.

Phylogenetic analyses and international introductions

We estimated maximum likelihood and molecular clock phylogenies for a global dataset with a total of 1182 genomes sampled from 24 December 2019 to 30 April 2020 (root-to-tip genetic distance correlation with sampling dates, $r^2 = 0.53$; Fig. 3A and fig. S7). We inferred a median evolutionary rate of 1.13×10^{-3} (95% BCI: 1.03 to 1.23×10^{-3}) substitutions per site per year using an exponential growth coalescent model, equating to 33 changes per year on average across the virus genome. This is within the range of evolutionary rates estimated for other human coronaviruses (30–33). We estimate the date of the common ancestor (TMRCA) of the SARS-CoV-2 pandemic to around mid-November 2019 (median = 19 November 2019, 95% BCI: 26 October 2019 to 6 December 2019), which is consistent with recent findings (34, 35).

Phylogenetic analysis revealed that the majority of the Brazilian genomes (76%, $n = 370/490$) fell into three clades, hereafter referred to as Clade 1 ($n = 186/490$, 38% of Brazilian strains), Clade 2 ($n = 166$, 34%), and Clade 3 ($n = 18/490$, 4%) (Fig. 3A and figs. S8 and S9), which were largely in agreement with those identified in a phylogenetic analysis using 13,833 global genomes. The most recent common ancestors of the three main Brazilian clades (Clades 1 to 3) were dated from 28 February (21 February to 4 March 2020) (Clade 1),

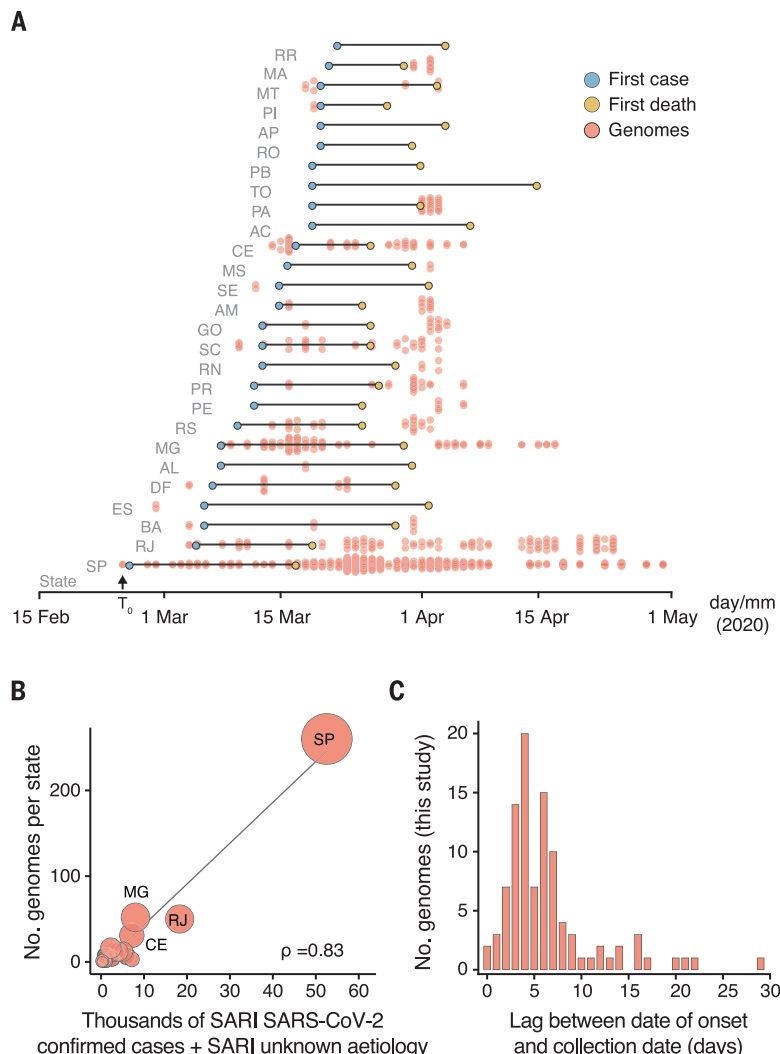


Fig. 2. Spatially representative genomic sampling. (A) Dumbbell plot showing the time intervals between date of collection of sampled genomes, notification of first cases, and first deaths in each state. Red lines indicate the lag between the date of collection of first genome sequence and first reported case. The key for the two-letter ISO 3166-1 codes for Brazilian federal units (or states) are provided in the supplementary materials. (B) Spearman's rank correlation between the number of SARI SARS-CoV-2 confirmed and SARI cases with unknown etiology against the number of sequences for each of the 21 Brazilian states included in this study (see also fig. S4). Circle sizes are proportional to the number of sequences for each federal unit. (C) Interval between the date of symptom onset and the date of sample collection for the sequences generated in this study.

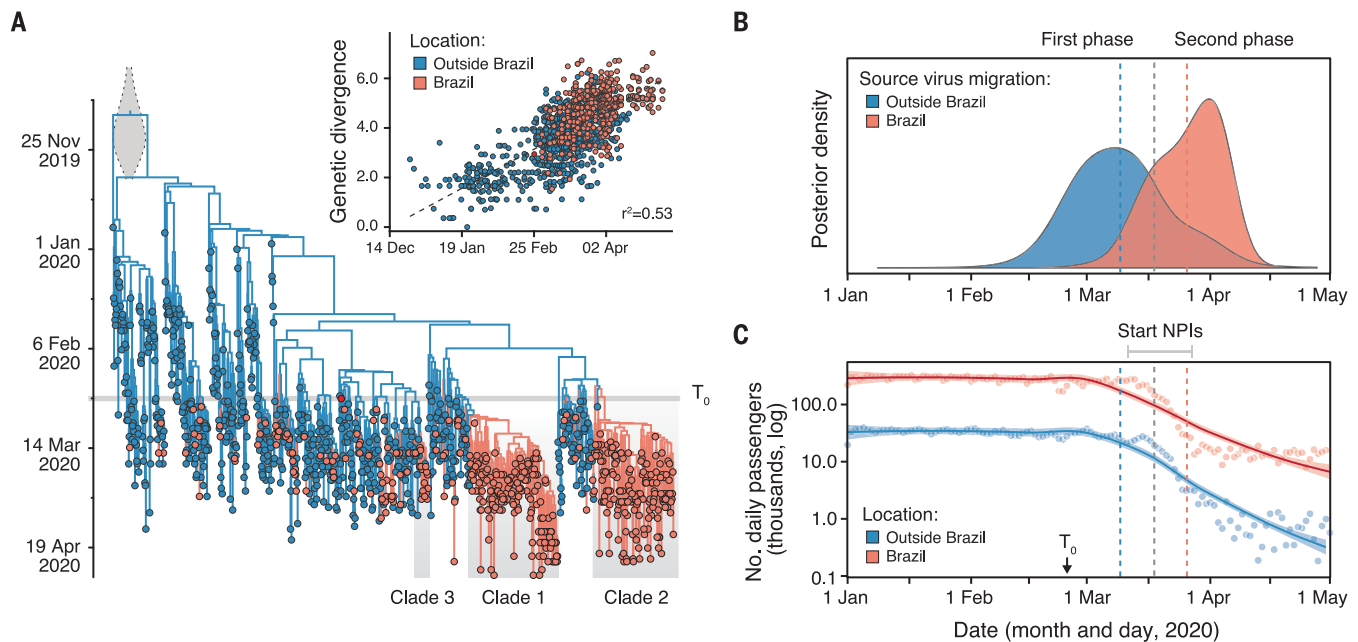


Fig. 3. Evolution and spread of SARS-CoV-2 in Brazil. (A) Time-resolved maximum clade credibility phylogeny of 1182 SARS-CoV-2 sequences, 490 of which are from Brazil (salmon) and 692 from outside of Brazil (blue). The largest Brazilian clades are highlighted by gray boxes (Clade 1, Clade 2, and Clade 3). Inset shows a root-to-tip regression of genetic divergence against dates of sample collection. Red tip corresponds to the first reported case in Brazil. (B) Dynamics of SARS-CoV-2 import events in Brazil. Dates of international and national (between federal states)

migration events were estimated from virus genomes using a phylogeographic approach. The first phase was dominated by virus migrations from outside of Brazil, whereas the second phase was marked by virus spread within Brazil. Dashed vertical lines correspond to the mean posterior estimate for migration events from outside of Brazil (blue) and within Brazil (red). (C) Locally estimated scatterplot smoothing of the daily number of international (blue) and national (red) air passengers in Brazil in 2020. T_0 , date of first reported case in Brazil (25 February 2020).

22 February (17 to 24 February 2020) (Clade 2), to 11 March (9 to 12 March 2020) (Clade 3) (Fig. 3A and fig. S10). This indicates that community-driven transmission was already established in Brazil by early March, suggesting that international travel restrictions initiated after this period would have had limited impact. Brazilian Clade 1 is characterized by a nucleotide substitution in the spike protein (G25088T, numbering relative to GenBank reference NC_045512.2) and circulates predominantly in São Paulo state ($n = 159$, 85.4%; figs. S9 and S11). Clade 2 is defined by two nucleotide substitutions in ORF6 (T27299C) and nucleoprotein (T29148C); this is the most spatially widespread lineage, with sequences from a total of 16 states in Brazil. Clade 3 is concentrated in Ceará state ($n = 16$, 89%) and falls in a global cluster with sequences mainly from Europe. In the Amazon region, where the epidemic is expanding rapidly (14, 22), we found evidence for multiple national and international introductions, with 37% ($n = 7/19$) of sequences from Pará and Amazonas states clustering in Clade 1 and 32% ($n = 6/19$) in Clade 2.

Time-measured phylogeographic analyses revealed at least 102 (95% BCI: 95 to 109) international introductions of SARS-CoV-2 in Brazil (Fig. 3A and figs. S8 and S12). This represents an underestimate of the real number of introductions because we sequenced,

on average, only one out of 200 confirmed cases. Most of these estimated introductions were directed to internationally well-connected states (36) such as São Paulo (36% of all imports), Minas Gerais (24%), Ceará (10%), and Rio de Janeiro (8%) (fig. S12). We further assessed the contribution of international versus national virus lineage movement events through time (Fig. 3B). In the first phase of the epidemic, we found an increasing number of international introductions until 10 March 2020 (Fig. 2B). Limited available travel history data (15) suggested that these early cases were predominantly acquired from Italy (26%, $n = 70$ of 266 unambiguously identified country of infection) and the United States (28%, $n = 76$ of 266). After this initial phase, we found that the estimated number of international imports decreased concomitantly with the decline in the number of international passengers traveling to Brazil (Fig. 3, B and C, and S13). By contrast, despite the declines in the number of passengers traveling on national flights (Fig. 3C), we detected an increase in virus lineage movement events between Brazilian regions at least until early April 2020.

Modeling spatiotemporal spread within Brazil

To better understand virus spread across spatiotemporal scales within Brazil, we used a continuous phylogeographic model that maps phylogenetic nodes to their inferred origin loca-

tions (37) (Fig. 4). We distinguished branches that remain within a state versus those that cross a state to infer the proportion of within-state versus between-state observed virus movement.

We estimate that during the first epidemic phase, SARS-CoV-2 spread mostly locally and within state borders. By contrast, the second phase was characterized by long-distance movement events and the ignition of the epidemic outside of the southeast region of Brazil (Fig. 4A). Throughout the epidemic, we found that within-state virus lineage movement was, on average, 5.1-fold more frequent than between-state movement. Moreover, our data suggest that within-state virus spread and, to a lesser extent, between-state virus spread decreased after the implementation of NPIs (Fig. 4B). However, the more limited sampling after 6 April 2020 (see fig. S2) decreased inferred virus lineage movement to the present (Figs. 3B and 4B).

We found that the average route length traveled by passenger increased by 25% during the second phase of the epidemic (Fig. 4C) despite a concomitant reduction in the number of passengers flying within Brazil (Fig. 3C). The increase in the average route length after NPI implementation resulted from a larger reduction in the number of air passengers flying on shorter-distance journeys compared with those flying on longer-distance journeys. For example, we found an 8.8-fold reduction in

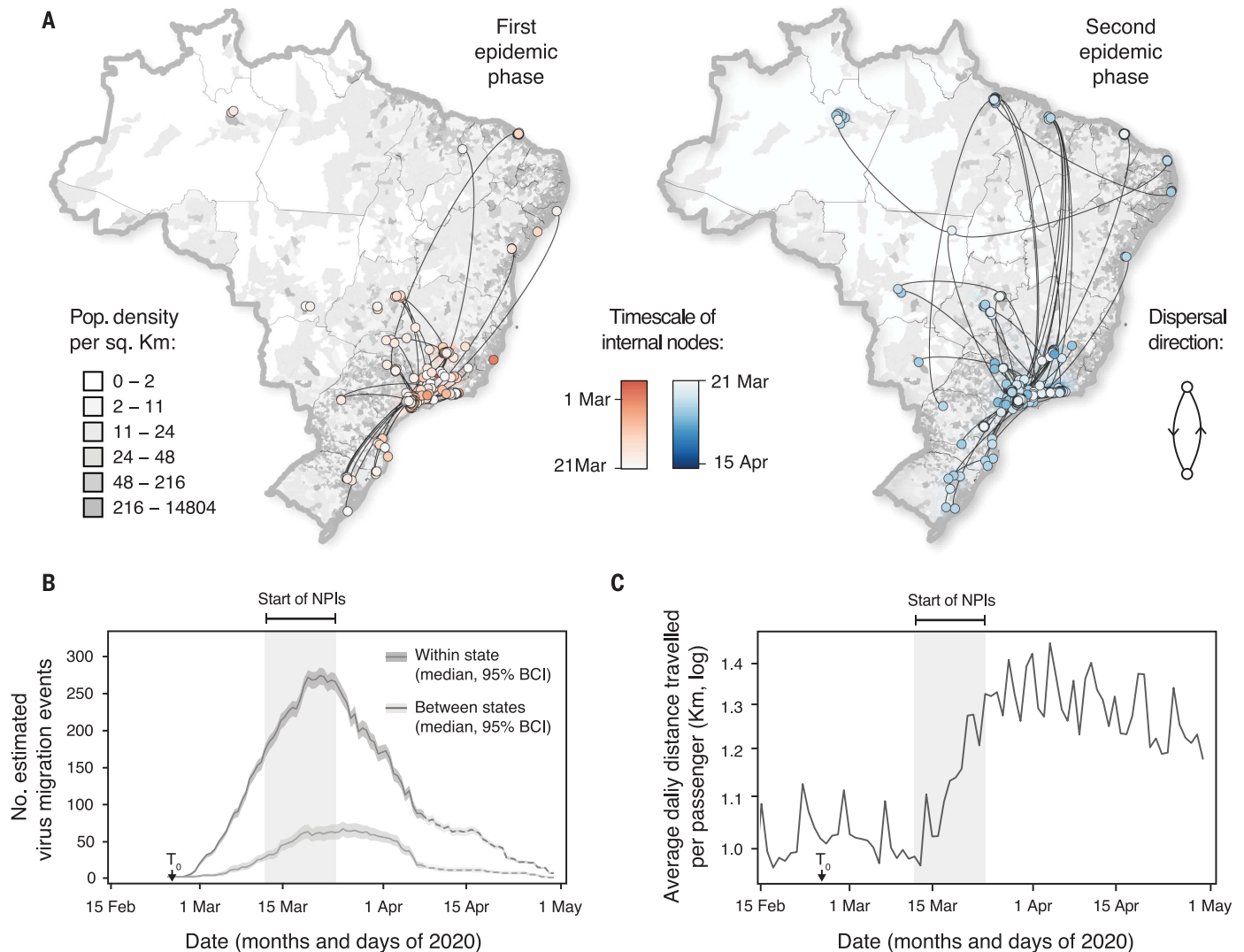


Fig. 4. Spread of SARS-CoV-2 in Brazil. (A) Spatiotemporal reconstruction of the spread of Brazilian SARS-CoV-2 clusters containing more than two sequences during the first (left) and the second (right) epidemic phase (Fig. 3B). Circles represent nodes of the maximum clade credibility phylogeny and are colored according to their inferred time of occurrence. Shaded areas represent the 80% highest posterior density interval and depict the uncertainty of the phylogeographic estimates for each node. Solid curved lines denote the links between nodes and the directionality of movement. Sequences belonging to clusters with fewer than three sequences were also plotted on the map with no

lines connecting them. Background population density for each municipality was obtained from the Brazilian Institute of Geography (<https://www.ibge.gov.br/>). See Fig. S14 for details of virus spread in the southeast region. (B) Estimated number of within-state (or within a given federal unit) and between-state (or between federal units) virus migrations over time. Dashed lines indicate estimates obtained during the period of limited sampling (fig. S2). (C) Average distance in kilometers traveled by an air passenger per day in Brazil. The number of daily air passengers is shown in Fig. 3B. Light gray boxes indicate the starting dates of NPIs across Brazil.

the number of passengers flying in flight legs <1000 km, compared with a 4.4-fold reduction in those flying >2000 km (fig. S15). These findings emphasize the roles of within- and between-state mobility as a key driver of both local and interregional virus spread, with highly populated and well-connected urban conurbations in the southeast region acting as the main sources of virus exports within the country (fig. S12).

Discussion

We provide a comprehensive analysis of SARS-CoV-2 spread in Brazil showing the importance

of community- and nation-wide measures to control the COVID-19 epidemic in Brazil. Although NPIs initially reduced virus transmission and spread, the continued increase in the number of cases and deaths in Brazil highlights the urgent need to prevent future virus transmission by implementing rapid and accessible diagnostic screening, contact tracing, quarantining of new cases, and coordinated social and physical distancing measures across the country (38). With the recent relaxation of NPIs in Brazil and elsewhere, continued molecular, immunological, and genomic surveil-

lance are required for real-time data-driven decisions. Our analysis shows how changes in mobility may affect global and local transmission of SARS-CoV-2 and demonstrates how combining genomic and mobility data can complement traditional surveillance approaches.

REFERENCES AND NOTES

1. F. Wu et al., *Nature* **579**, 265–269 (2020).
2. K. G. Andersen, A. Rambaut, W. I. Lipkin, E. C. Holmes, R. F. Garry, *Nat. Med.* **26**, 450–452 (2020).
3. World Health Organization, *Coronavirus Disease (COVID-19) Situation Reports* (2020); www.who.int/emergencies/diseases/novel-coronavirus-2019/situation-reports.

4. H. Tian *et al.*, *Science* **368**, 638–642 (2020).
5. M. U. G. Kraemer *et al.*, *Science* **368**, 493–497 (2020).
6. A. Rambaut *et al.*, *Nat. Microbiol.* (2020).
7. T. W. Russell *et al.*, *Euro Surveill.* **25**, 2000256 (2020).
8. R. Verity *et al.*, *Lancet Infect. Dis.* **20**, 669–677 (2020).
9. J. T. Wu *et al.*, *Nat. Med.* **26**, 506–510 (2020).
10. M. M. Arons *et al.*, *N. Engl. J. Med.* **382**, 2081–2090 (2020).
11. L. Ferretti *et al.*, *Science* **368**, eabb6936 (2020).
12. E. Lavezzo *et al.*, *Nature* (2020).
13. K. Mizumoto, K. Kagaya, A. Zarebski, G. Chowell, *Euro Surveill.* **25**, 2000180 (2020).
14. Brazilian Ministry of Health, *Painel de Casos de Doença Pelo Coronavírus 2019 (COVID-19) No Brasil Pelo Ministério da Saúde* (2020); <http://covid.saude.gov.br>.
15. W. M. de Souza *et al.*, *Nat. Hum. Behav.* **4**, 856–865 (2020).
16. J. Croda *et al.*, *Rev. Soc. Bras. Med. Trop.* **53**, e20200167 (2020).
17. J. Croda, L. Garcia, *Epidemiol. Ser. Saúde* **29**, e2020002 (2020).
18. S. B. Oliveira *et al.*, Monitoring social distancing and SARS-CoV-2 transmission in Brazil using cell phone mobility data. medRxiv 2020.04.30.20082172 [Preprint] (5 May 2020); <https://doi.org/10.1101/2020.04.30.20082172>.
19. S. M. Kissler, Reductions in commuting mobility predict geographic differences in SARS-CoV-2 prevalence in New York City (Harvard DASH Repository, 2020); https://dash.harvard.edu/bitstream/handle/1/42665370/Kissler_et_al_NYC_mobility.pdf?sequence=1&isAllowed=y.
20. H. J. T. Unwin *et al.*, *Report 23: State-Level Tracking of COVID-19 in the United States (21-05-2020)* (Imperial College London, 2020); <https://doi.org/10.25561/79231>.
21. S. Flaxman *et al.*, *Nature* **584**, 257–261 (2020).
22. T. A. Mellan *et al.*, *Report 21: Estimating COVID-19 Cases and Reproduction Number in Brazil* (2020); <https://doi.org/10.25561/78872>.
23. Y.-Z. Zhang, E. C. Holmes, Novel 2019 coronavirus genome, *Virological* (2020); <https://virological.org/t/novel-2019-coronavirus-genome/319>.
24. V. M. Corman *et al.*, *Euro Surveill.* **25**, 2000045 (2020).
25. T. Thi Nhu Thao *et al.*, *Nature* **582**, 561–565 (2020).
26. P. C. Resende *et al.*, Genomic surveillance of SARS-CoV-2 reveals community transmission of a major lineage during the early pandemic phase in Brazil. bioRxiv 020.06.17.158006 [Preprint] (2020); <https://doi.org/10.1101/2020.06.17.158006>.
27. J. Xavier *et al.*, *Emerg. Microbes Infect.* **9**, 1824–1834 (2020).
28. V. A. Nascimento *et al.*, *Memoirs of the Oswaldo Cruz Institute* 10.1590/0074-0276200200310 (2020).
29. Y. Shu, J. McCauley, *Euro. Surveill.* **22**, 30494 (2017).
30. M. Cotten *et al.*, *Lancet* **382**, 1993–2002 (2013).
31. M. Cotten *et al.*, *mBio* **5**, e01062-13 (2014).
32. G. Dudas, L. M. Carvalho, A. Rambaut, T. Bedford, *eLife* **7**, e31257 (2018).
33. Z. Zhao *et al.*, *BMC Evol. Biol.* **4**, 21 (2004).
34. S. Duchene *et al.*, Temporal signal and the phylodynamic threshold of SARS-CoV-2. bioRxiv 2020.05.04.077735 [Preprint] (2020); <https://doi.org/10.1101/2020.05.04.077735>.
35. J. Lu *et al.*, *Cell* **181**, 997–1003.e9 (2020).
36. D. D. S. Candido *et al.*, *J. Travel Med.* **27**, taaa042 (2020).
37. S. Dellicour *et al.*, A phylodynamic workflow to rapidly gain insights into the dispersal history and dynamics of SARS-CoV-2 lineages. bioRxiv 2020.05.05.078758 [Preprint] (2020); <https://doi.org/10.1101/2020.05.05.078758>.
38. World Health Organization, Coronavirus disease 2019 (COVID-19): Situation report –72 (WHO, 2020); https://www.who.int/docs/default-source/coronaviruse/situation-reports/20200401-sitrep-72-covid-19.pdf?sfvrsn=3dd8971b_2.
39. Centre for Genomic Pathogen Surveillance, Imperial College London, Report of 427 novel genomes from Brazil and the associated metadata, Microreact (2020); <https://microreact.org/project/rKjKLMrjdPVHKR1erUzKy>.
40. Data and code for: D. S. Candido *et al.*, Evolution and epidemic spread of SARS-CoV-2 in Brazil, Dryad (2020); <https://doi.org/10.5061/dryad.rxdbrv5z>.

ACKNOWLEDGMENTS

A full list acknowledging those involved in the diagnostics and generation of new sequences as part of the CADDE-Genomic-Network can be found in the supplementary materials. We thank the administrators of the GISAID database for supporting rapid and transparent sharing of genomic data during the COVID-19 pandemic. A full list acknowledging the authors submitting data used in this study can be found in data S2. We thank P. Resende (FIOCRUZ), T. Adelino (FUNED), C. Sacchi (IAL), V. Nascimento (FIOCRUZ Amazonia), and their colleagues for submitting Brazilian data to GISAID; A. Pinter (SUCEN), N. Gouveia (USP), and I. Marçilio de Souza (HCFM-USP) for fruitful discussions; L. Matkin and J. Quick for logistic support; and the UNICAMP Task Force against Covid-19 for support in generating genomes from Campinas. The analysis of openly available epidemiological data from <https://covid.saude.gov.br/> has benefited from the COVID-19 surveillance efforts by the Secretaria de Vigilância em Saúde, Ministry of Health, Brazil. **Funding:** This project was supported by a Medical Research Council-São Paulo Research Foundation (FAPESP) CADDE partnership award (MR/S0195/1 and FAPESP 18/14389-0) (<http://caddecentre.org/>). FAPESP further supports I.M.C. (2018/17176-8 and 2019/12000-1), J.G.J. (2018/17176-8 and 2019/12000-1, 18/14389-0), F.C.S.S. (2018/25468-9), W.M.S. (2017/13981-0, 2019/24251-9), M.F. (2018/09383-3), T.M.C. (2019/07544-2), C.A.M.S. (2019/21301-5), H.I.N. (2018/14933-2), P.S.P. (16/18445-7), M.L.N. (20/04836-0), and J.L.M. (2020/04558-0 and 2016/00194-8). N.R.F. is supported by a Wellcome Trust and Royal Society Sir Henry Dale Fellowship (204311/Z/16/Z). D.S.C. is supported by the Clarendon Fund and by the Department of Zoology, University of Oxford. S.D. is supported by the Fonds National de la Recherche Scientifique (FNRS, Belgium). J.T. and P.L. are supported by European Union's Horizon 2020 project MOOD (874850). This project was supported by CNPq (M.T.M., M.L.N., and A.T.R.V.: 303170/2017-4; R.S.A.: 312688/2017-2 and 439119/2018-9; R.P.S.: 310627/2018-4; and W.M.S.: 408338/2018-0), FAPERJ (A.T.R.V.: E-26/202.826/2018 and R.S.A.: 202.922/2018). M.S.R. is supported by FMUSP. C.A.P., G.M.F., J.H., and M.R.A. are supported by CAPES. O.J.B. is supported by a Sir Henry Wellcome Fellowship funded by the Wellcome Trust (206471/Z/17/Z). R.P.S. is supported by FAPEMIG (APQ-00475-20). M.M.T. is supported by Instituto Nacional de Ciência e Tecnologia em Dengue (INCT Dengue 465425/2014-3). A.T.R.V. is supported by FINEP

(0116.0078.00). P.L. and N.J.L. are supported by the Wellcome Trust ARTIC network (collaborators award no. 206298/Z/17/Z). P.L. and A.R. are supported by the European Research Council (grant no. 725422-ReservoirDOCS). O.G.P., N.R.F., and L.D.P. are supported by the Oxford Martin School. This work received funding from the U.K. Medical Research Council under a concordat with the U.K. Department for International Development. We additionally acknowledge support from Community Jameel and the NIHR Health Protection Research Unit in Modelling Methodology. **Author contributions:** Conceptualization: D.S.C., I.M.C., J.G.J., E.C.S., N.R.F.; Formal analysis: D.S.C., I.M.C., J.G.J., W.M.S., F.R.R.M., S.D., T.A.M., L.P., R.H.M.P., J.T., L.A., C.M.V., H.H., S.M., M.S.G., L.M.C., L.F.B., C.A.P., O.J.B., S.M.N., S.C.H., J.L.P.M., A.T.R.V., S.B., O.G.P., P.L., C.H.W., R.S.A., N.R.F.; Investigation: D.S.C., I.M.C., J.G.J., W.M.S., F.R.R.M., R.H.M.P., F.C.S.S., E.R.M., M.T.M., C.M.V., M.J.F., T.M.C., C.A.M.S., M.S.R., M.R.A., J.A., H.N., P.S.P., A.T., A.D.R., C.K.V.B., A.L.G., A.P.G., N.G., C.S.A., A.C.S.F., C.X.L., J.E.L., C.G., G.M.F., R.S.F., F.G., M.T.G., M.L.M., M.W.P., T.M.P.P.C., C.S.L., A.A.S.S., C.L.S., J.F., A.C.S., A.Z.S., M.N.N.S., C.Z.S., R.P.S., L.C.R.M., M.M.T., J.H., P.A.F.L., R.G.M., M.L.N., S.F.C., J.L.P.M., A.T.R.V., R.S.A., E.C.S., N.R.F.; Interpretation: D.S.C., I.M.C., J.G.J., W.M.S., F.R.R.M., S.D., T.A.M., L.P., R.H.M.P., S.C.H., A.A.S.S., N.M.F., A.T.R.V., S.B., P.L., C.H.W., A.R., R.S.A., O.G.P., E.C.S., N.R.F.; Writing – original draft: D.S.C., I.M.C., J.G.J., W.M.S., F.R.R.M., S.D., T.A.M., R.S.A., O.G.P., E.C.S., N.R.F.; Writing – review & editing: All authors have read and approved the final version of the manuscript. Funding acquisition: W.M.S., M.L.N., N.M.F., J.L.P.M., A.T.R.V., N.J.L., R.S.A., O.G.P., E.C.S., N.R.F. **Competing interests:** The authors declare no competing interests. **Data and materials availability:** The 427 SARS-CoV newly generated genomes from this study can be found on GISAID under the accession IDs: EPI_ISL_470568-470655 and EPI_ISL_476152-476490. An interactive visualization of the temporal, geographic and mutational patterns in our data can be found at <https://microreact.org/project/rKjKLMrjdPVHKR1erUzKy> (39). Reads have been deposited to accession numbers PRJEB39487 (IMT-USP and UNICAMP) and PRJNA640656 (UFRJ-LNCC). All data, code, and materials used in the analysis are available on DRYAD (40). The IRB protocol number is CAAE 30127020.0.0000.0068 as described in the materials and methods. This work is licensed under a Creative Commons Attribution 4.0 International (CC BY 4.0) license, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. To view a copy of this license, visit <https://creativecommons.org/licenses/by/4.0/>. This license does not apply to figures/photos/artwork or other content included in the article that is credited to a third party; obtain authorization from the rights holder before using such material.

SUPPLEMENTARY MATERIALS

science.sciencemag.org/content/369/6508/1255/suppl/DC1
Materials and Methods
Figs. S1 to S15
Tables S1 to S3
List of Members of the CADDE Genomic Network
References (41–77)
Data S1 and S2
MDAR Reproducibility Checklist

10 June 2020; accepted 16 July 2020
Published online 23 July 2020
10.1126/science.abd2161

Evolution and epidemic spread of SARS-CoV-2 in Brazil

Darlan S. Candido, Ingra M. Claro, Jaqueline G. de Jesus, William M. Souza, Filipe R. R. Moreira, Simon Dellicour, Thomas A. Mellan, Louis du Plessis, Rafael H. M. Pereira, Flavia C. S. Sales, Erika R. Manuli, Julien Théze, Luiz Almeida, Mariane T. Menezes, Carolina M. Voloch, Marcilio J. Fumagalli, Thaís M. Coletti, Camila A. M. da Silva, Mariana S. Ramundo, Mariene R. Amorim, Henrique H. Hoeltgebaum, Swapnil Mishra, Mandev S. Gill, Luiz M. Carvalho, Lewis F. Buss, Carlos A. Prete Jr., Jordan Ashworth, Helder I. Nakaya, Pedro S. Peixoto, Oliver J. Brady, Samuel M. Nicholls, Amílcar Tanuri, Átila D. Rossi, Carlos K. V. Braga, Alexandra L. Gerber, Ana Paula de C. Guimarães, Nelson Gaburo Jr., Cecília Salete Alencar, Alessandro C. S. Ferreira, Cristiano X. Lima, José Eduardo Levi, Celso Granato, Giulia M. Ferreira, Ronaldo S. Francisco Jr., Fabiana Granja, Marcia T. Garcia, Maria Luíza Moretti, Mauricio W. Perroud Jr., Terezinha M. P. P. Castiñeiras, Carolina S. Lazari, Sarah C. Hill, Andreza Aruska de Souza Santos, Camila L. Simeoni, Julia Forato, Andrei C. Sposito, Angelica Z. Schreiber, Magnun N. N. Santos, Camila Zolini de Sá, Renan P. Souza, Luciana C. Resende-Moreira, Mauro M. Teixeira, Josy Hubner, Patricia A. F. Leme, Rennan G. Moreira, Maurício L. Nogueira, Brazil-UK Centre for Arbovirus Discovery, Diagnosis, Genomics and Epidemiology (CADDE) Genomic Network, Neil M. Ferguson, Silvia F. Costa, José Luiz Proença-Modena, Ana Tereza R. Vasconcelos, Samir Bhatt, Philippe Lemey, Chieh-Hsi Wu, Andrew Rambaut, Nick J. Loman, Renato S. Aguiar, Oliver G. Pybus, Ester C. Sabino and Nuno Rodrigues Faria

Science **369** (6508), 1255-1260.
DOI: 10.1126/science.abd2161 originally published online July 23, 2020

The spread of SARS-CoV-2 in Brazil

Brazil has been hard-hit by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) pandemic. Candido *et al.* combined genomic and epidemiological analyses to investigate the impact of nonpharmaceutical interventions (NPIs) in the country. By setting up a network of genomic laboratories using harmonized protocols, the researchers found a 29% positive rate for SARS-CoV-2 among collected samples. More than 100 international introductions of SARS-CoV-2 into Brazil were identified, including three clades introduced from Europe that were already well established before the implementation of NPIs and travel bans. The virus spread from urban centers to the rest of the country, along with a 25% increase in the average distance traveled by air passengers before travel bans, despite an overall drop in short-haul travel. Unfortunately, the evidence confirms that current interventions remain insufficient to keep virus transmission under control in Brazil.

Science, this issue p. 1255

ARTICLE TOOLS

<http://science.sciencemag.org/content/369/6508/1255>

SUPPLEMENTARY MATERIALS

<http://science.sciencemag.org/content/suppl/2020/07/22/science.abd2161.DC1>

RELATED CONTENT

<http://stm.sciencemag.org/content/scitransmed/12/546/eabc1931.full>
<http://stm.sciencemag.org/content/scitransmed/12/554/eabc1126.full>
<http://stm.sciencemag.org/content/scitransmed/12/550/eabc3539.full>
<http://stm.sciencemag.org/content/scitransmed/12/549/eabb9401.full>

REFERENCES

This article cites 58 articles, 9 of which you can access for free
<http://science.sciencemag.org/content/369/6508/1255#BIBL>

Use of this article is subject to the [Terms of Service](#)

Science (print ISSN 0036-8075; online ISSN 1095-9203) is published by the American Association for the Advancement of Science, 1200 New York Avenue NW, Washington, DC 20005. The title *Science* is a registered trademark of AAAS.

Copyright © 2020 The Authors, some rights reserved; exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works

PERMISSIONS

<http://www.sciencemag.org/help/reprints-and-permissions>

Use of this article is subject to the [Terms of Service](#)

Science (print ISSN 0036-8075; online ISSN 1095-9203) is published by the American Association for the Advancement of Science, 1200 New York Avenue NW, Washington, DC 20005. The title *Science* is a registered trademark of AAAS.

Copyright © 2020 The Authors, some rights reserved; exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works